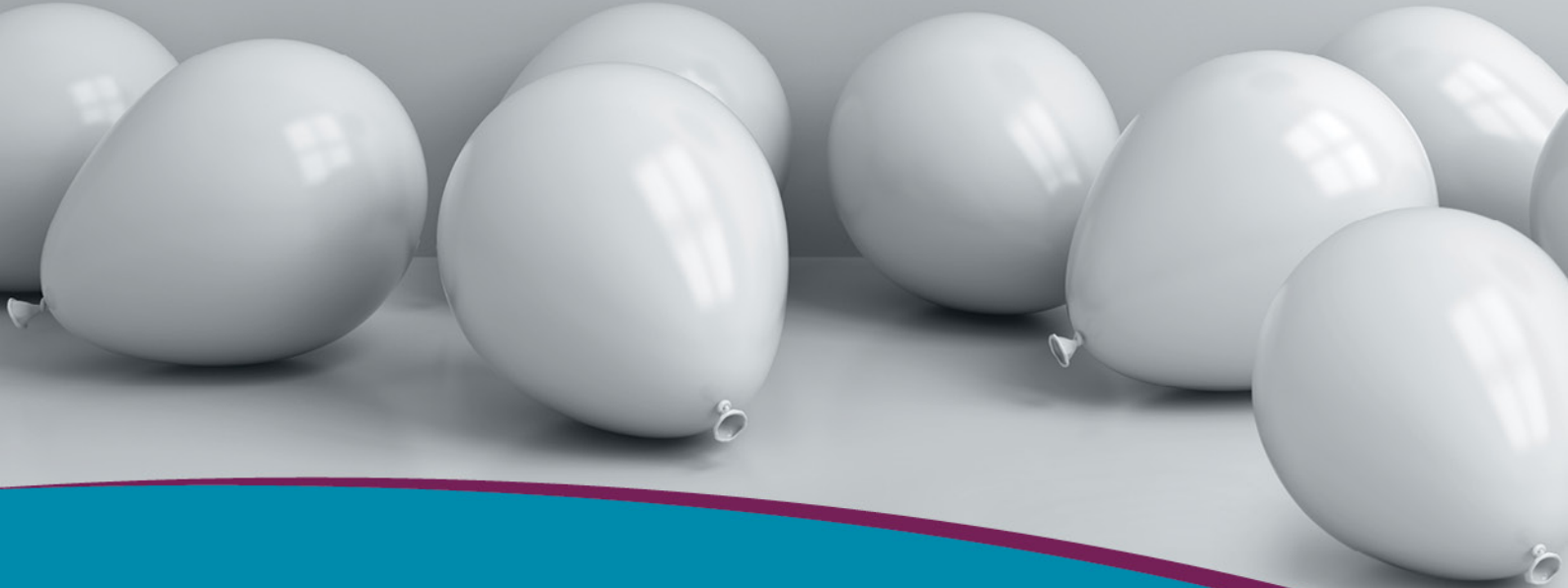


Helium crisis getting you

down?



Zero-Helium (ZHM) 1.5T high-field MRI could be the answer

Did you know that helium is a finite resource?

The world's first veterinary-specific small animal MRI is helium-free, giving you improved access to advanced diagnostics.







As part of Q-Care, our unique solution reduces financial, environmental and supply-chain risks.

We speak vet at hallmarq.net

Hallmarq
Advanced Veterinary Imaging



Medical causes of poor performance and their associations with fitness in Standardbred racehorses

Chiara M. Lo Feudo¹  | Luca Stucchi²  | Bianca Conturba²  |
 Giovanni Stancari²  | Enrica Zucca¹  | Francesco Ferrucci¹ 

¹Equine Sports Medicine Laboratory “Franco Tradati”, Department of Veterinary Medicine and Animal Sciences, Università degli Studi di Milano, Lodi, Italy

²Equine Unit, Veterinary Teaching Hospital, Department of Veterinary Medicine and Animal Sciences, Università degli Studi di Milano, Lodi, Italy

Correspondence

Luca Stucchi, Department of Veterinary Medicine and Animal Sciences, Università degli Studi di Milano, Via dell'Università 6, 26900 Lodi, Italy.

Email: luca.stucchi@unimi.it

Abstract

Background: Poor performance is a multifactorial syndrome of racehorses, commonly associated with subclinical disorders, which can be diagnosed by exercise testing.

Objectives: Describe the prevalence of medical causes of poor performance in Standardbreds unassociated with lameness, and evaluate their relationships with fitness variables measured by exercise treadmill test.

Animals: Hospital population of 259 nonlame Standardbred trotters referred for poor performance.

Methods: The horses' medical records were retrospectively reviewed. Horses underwent a diagnostic protocol including resting examination, plasma lactate concentration, treadmill test with continuous ECG and assessment of fitness variables, creatine kinase activity, treadmill endoscopy, postexercise tracheobronchoscopy, bronchoalveolar lavage (BAL), and gastroscopy. The prevalence of different disorders was evaluated, including cardiac arrhythmias, exertional myopathies, dynamic upper airway obstructions (DUAOs), exercise-induced pulmonary hemorrhage (EIPH), moderate equine asthma (MEA), and gastric ulcers (EGUS). The associations of these disorders with fitness variables were investigated individually and using multivariable models.

Results: Moderate equine asthma and EGUS were the most common disorders, followed by EIPH, DUAOs, cardiac arrhythmias, and exertional myopathies. Hemosiderin score was positively correlated with BAL neutrophils, eosinophils, and mast cells; increased creatine kinase activity was associated with BAL neutrophilia, DUAOs, premature complexes, and squamous gastric disease. Treadmill velocity at a plasma lactate concentration of 4 mmol/L and at heart rate of 200 beats per minute was negatively affected by BAL neutrophilia, multiple DUAOs, exertional myopathies, and squamous gastric disease.

Conclusions: The multifactorial nature of poor performance was confirmed, with MEA, DUAOs, myopathies and EGUS representing the main diseases involved in fitness impairment.

Abbreviations: BAL, bronchoalveolar lavage; CK, creatine-kinase; DUAO, dynamic upper airway obstruction; ECG, electrocardiogram; EGGD, equine glandular gastric disease; EGUS, equine gastric ulcer syndrome; EIPH, exercise-induced pulmonary hemorrhage; ER, exertional rhabdomyolysis; ESGD, equine squamous gastric disease; HSTE, high-speed treadmill endoscopy; MEA, mild-moderate equine asthma; PC, premature complex; PLH, pharyngeal lymphoid hyperplasia; TB, tracheal bifurcation blunting; THS, total hemosiderin score; TM, tracheal mucus.

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial](https://creativecommons.org/licenses/by-nc/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

© 2023 The Authors. *Journal of Veterinary Internal Medicine* published by Wiley Periodicals LLC on behalf of American College of Veterinary Internal Medicine.

KEYWORDS

equine exercise physiology, equine performance, equine sports medicine, poor performance, sport horses, treadmill test

1 | INTRODUCTION

Poor performance is a common and complex condition of racehorses, usually associated with subclinical disorders.¹ In addition to musculoskeletal conditions, medical disorders are common, and their identification can be challenging for the clinician. In fact, poorly performing horses are mostly normal on physical examination, and the identification of underlying medical diseases often requires dynamic tests.² Moreover, multiple disorders can concomitantly affect horses,^{1,3} and it can be difficult to determine the contribution of each to the impaired racing performance. To investigate poor performance in sport horses, investigators rely either on the subjective expectations of trainers and owners⁴⁻⁶ or on quantitative data, such as racing placements and earnings.⁷⁻¹⁰ Many biases however can interfere with competition results, such as the number of other participants, level of the race and possible handicaps, ground surface, distance, driver interventions, and other factors.^{11,12} Objective and measurable information on the fitness of Standardbred racehorses can be obtained by standardized exercise tests, which can be performed either in the field or using a treadmill.^{1,13-18} The treadmill allows for control of environment, speed, inclination, and ensures higher repeatability of the results and reproduction of racing conditions. In particular, for nonmusculoskeletal conditions, the treadmill velocity at which a blood lactate concentration of 4 mmol/L (VL4) is reached and the treadmill velocity at a heart rate of 200 beats per minute (bpm; V200) are considered good indicators of the level of performance.¹⁹⁻²²

Musculoskeletal disorders are the most common cause of decreased performance in Standardbred racehorses,² followed by medical disorders such as respiratory diseases affecting the upper or lower airways,^{1,3} including dynamic upper airway obstructions (DUAO),²³⁻²⁵ mild to moderate equine asthma (MEA),^{5,26-28} and exercise-induced pulmonary hemorrhage (EIPH).²⁹⁻³² In fact, the respiratory system is considered the main limiting factor to performance even in healthy racehorses (both Thoroughbreds and Standardbreds), and related disorders can substantially impact the aerobic and athletic capacity of the horse.³³ Other reported medical causes of poor performance include exertional rhabdomyolysis (ER) and other myopathies,^{18,34} equine gastric ulcer syndrome (EGUS),³⁵⁻³⁷ and cardiac arrhythmias.³⁸⁻⁴⁰ Numerous studies have investigated the effects of these conditions on the fitness of Standardbred racehorses, reporting contrasting results, probably because of different inclusion criteria, study design, and the variables chosen to define performance. Moreover, poor performance is rarely caused by a single disorder.^{1,3} Therefore, the effects of different diseases should not be considered individually but rather with the perspective of a multifactorial condition.

We aimed to describe the prevalence and distribution of the medical disorders diagnosed in a population of poorly performing Standardbred trotters and their associations with fitness variables measured

during a standardized exercise test on treadmill. Moreover, the associations between concomitantly encountered diseases was evaluated, and the contribution of each disorder on changes in VL4 and V200 in a multifactorial setting was determined.

2 | MATERIALS AND METHODS

2.1 | Study population

In our retrospective study, the medical records of the Standardbred trotters referred to the Equine Sports Medicine Unit of the Veterinary Teaching Hospital, University of Milan (Italy), for poor performance between 2002 and 2021 were reviewed. All horses ($n = 259$) were in full training upon admission and underwent a diagnostic evaluation for poor performance with identification of subclinical causes. Horses presenting with signs of systemic illness, lameness, clinically relevant arrhythmias at rest, or clinically relevant valvular regurgitations were excluded from the study. The age of the horses varied from 2 to 9 years (median, 3; interquartile range [IQR], 3-4 years). The population included 93 females (35.91%) and 166 males (146 stallions, 20 geldings; 64.09%).

Ethical approval was waived because only clinical patients undergoing standard diagnostic procedures were included. Informed consent for the use of clinical data was obtained from all owners or holders.

2.2 | Poor performance diagnostic protocol

Because the study included a period of time over 20 years, the poor performance diagnostic protocol varied slightly throughout the years; the number of horses undergoing each clinical procedure is specified in parentheses. The protocol included:

- Collection of history, clinical examination and laboratory analyses ($n = 259$);
- ECG at rest ($n = 259$);
- Resting endoscopy of the upper airway ($n = 259$);
- Incremental exercise test on high-speed treadmill ($n = 259$), during which a continuous ECG was obtained by Holter recorder ($n = 238$);
- Assessment of serum creatine kinase (CK) activity 6 hours after exercise ($n = 255$);
- High-speed treadmill endoscopy (HSTE; $n = 248$);
- Postexercise tracheobronchoscopy (30 minutes after HSTE; $n = 246$);
- Lower airway endoscopy, bronchoalveolar lavage (BAL) collection and cytological examination ($n = 215$); and,
- Gastroscopy ($n = 173$).

2.3 | Fitness variables

Fitness variables were obtained during an incremental exercise test on a high-speed treadmill, performed as described previously.²⁸ Before performing the test, horses were trained to the use of treadmill by 2 daily sessions; during the test, horses wore the equipment normally used during racing and a heart rate monitor (Polar, Equine Inzone FT1, Steinhausen, Switzerland). A venous catheter was placed into the left jugular vein and connected to an extension tube to allow blood collection during the test. The belt was inclined with a 5% slope, and horses were warmed up at walk (1.5 m/s) for 4 minutes and trot (6 m/s) for 3 minutes. At the end of the warm-up phase, a first sample of blood was collected. Then, 1-minute-long phases followed, and at each phase the velocity of the treadmill was increased by 1 m/s until the onset of fatigue. At the end of each phase, blood was collected. Horses were cooled down at walk with 0% inclination for 30 minutes. At 1, 5, 15, and 30 minutes after the end of maximal exercise, blood samples were collected and used for the measurement of plasma lactate concentration, blood pH, and hematocrit, as described previously.²⁵

The obtained variables of fitness included:

- V200 (m/s): velocity at a heart rate of 200 bpm;
- VLa4 (m/s): velocity at a plasma lactate concentration of 4 mmol/L;
- HRLa4 (bpm): heart rate at a plasma lactate concentration of 4 mmol/L;
- Lac_{max} (mmol/L): maximum peak of plasma lactate concentration;
- V_{max} (m/s): maximum velocity until the onset of fatigue;
- Lac₃₀ (mmol/L): plasma lactate concentration at 30 min postexercise;
- HR₃₀ (bpm): heart rate at 30 min postexercise;
- pH_{min}: minimum pH;
- Hct_{max}: maximum hematocrit.

The exact values of VLa4 and HRLa4 were calculated using specific software (Lactate-E 1.0, Dr. David Higgins), providing precise lactate threshold markers using inverse prediction.⁴¹

2.4 | Arrhythmias

During the incremental treadmill test and recovery, horses wore a Holter recorder (Cardioline Click Holter, Trento, Italy), which recorded 3 unipolar leads. A modified base-apex configuration was used, and electrodes were placed and fixed as described previously.⁴⁰ Obtained data were analyzed using dedicated software (Click Holter Cardioline Prima Manager, Trento, Italy). Because the software is specifically designed for human patients, all ECG recordings were reviewed and analyzed by an experienced operator. Arrhythmias considered clinically relevant in affecting performance were defined either as the presence of at least 2 isolated premature complexes (PCs) during peak exercise (from V200 to the end of maximal exercise) or as the presence of at least 5 PCs or pairs of paroxysms of PCs detected during peak exercise or

immediately after.³ For statistical purposes, horses were divided into affected or nonaffected by PCs.

2.5 | Exertional myopathies

Six hours after the end of the treadmill test, blood samples were collected in plain tubes from the jugular vein, and immediately centrifuged. Serum was isolated and creatine kinase (CK) activity was measured as described elsewhere.⁴² Serum CK activity was considered within normal ranges from 44 to 735 IU/L; higher values were considered diagnostic of ER.⁴² Moreover, history and postexercise clinical signs of exertional myopathies were recorded.

2.6 | Dynamic upper airway obstructions

After the incremental treadmill test, horses were given 1 day of active rest, during which they were hand-walked twice a day. On the next day, HSTE was performed during a maximum intensity test at constant speed, as described previously.²⁵ The presence of any DUAO was recorded and classified as mild (medial deviation of aryepiglottic folds, epiglottic entrapment) or severe (dorsal displacement of the soft palate, nasopharyngeal collapse, epiglottic retroversion, dynamic laryngeal collapse).²⁵ For statistical purposes, horses were divided into 4 groups: no DUAO, mild DUAO, severe DUAO, and multiple DUAOs (if concomitantly affected by >1 type of DUAO).

2.7 | Exercise-induced pulmonary hemorrhage and asthma

Thirty minutes after the end of HSTE, tracheobronchoscopy was performed to assess EIPH, as described previously³²; the presence of blood in the trachea and bronchi was graded from 0 to 4 based on a recognized scoring system.⁴³

Twenty-four hours after the HSTE, horses were restrained in a stock and sedated using 0.01 mg/kg detomidine hydrochloride IV, and a second lower airway endoscopy was performed. Endoscopic scores were assigned to pharyngeal lymphoid hyperplasia (PLH; from 0 to 4),⁴⁴ tracheal mucus accumulation (TM; from 0 to 5),⁴⁵ and tracheal bifurcation blunting (TB; from 0 to 4).⁴⁶ The endoscope was advanced into the bronchial tree until it was wedged within a segmental bronchus, BAL was performed as described previously²⁸ and the fluid was collected in sterile EDTA tubes. Within 90 minutes of collection, 300 µL of pooled BAL were centrifuged at 26g for 5 min (Rotofix 32, Hettich Cyto System, Tuttlingen, Germany). The slides were air-dried, stained with May-Grünwald-Giemsa (for leukocyte differential cell count) and Perl's Prussian Blue (for hemosiderophage count), and observed under light microscopy at 400× and 1000×.

Based on the presence and subtype of lower airway inflammation,¹⁵ horses were classified as:

- Healthy: neutrophils $\leq 5\%$, eosinophils $\leq 1\%$, and mast cells $\leq 2\%$;
- Neutrophilic MEA: neutrophils $> 5\%$, eosinophils $\leq 1\%$, and mast cells $\leq 2\%$;
- Eosinophilic-mastocytic MEA: neutrophils $\leq 5\%$, eosinophils $> 1\%$, mast cells $> 2\%$; and,
- Mixed MEA: neutrophils $> 5\%$, eosinophils $> 1\%$, and/or mast cells $> 2\%$.

For the evaluation of EIPH, 100 macrophages were assessed; a total hemosiderin score (THS) was calculated by multiplying the percentage of hemosiderophages of the total of macrophages by the median 0-4 grade of hemosiderin, as described previously.³² A score between 0 and 400 was obtained; horses were considered as positive for EIPH when THS was > 75 .⁴⁷

2.8 | Equine gastric ulcer syndrome

At the end of the diagnostic protocol, gastroscopy was performed as described previously.³⁷ As recommended by the European College of Equine Internal Medicine consensus statement,³⁶ the squamous mucosa was scored for equine squamous gastric disease (ESGD) from 0 to 4, according to the Equine Gastric Ulcer Council scoring system,⁴⁸ and the glandular mucosa was evaluated for the presence or absence of equine glandular gastric disease (EGGD).

2.9 | Statistical analysis

The data were analyzed using commercially available statistical software (GraphPad Prism 9.5.0 for MacOS; GraphPad Software, San Diego, California). Normality of all data was evaluated using the Shapiro-Wilk test and descriptive statistics were determined. Statistical tests used to evaluate the associations among signalment, fitness variables, and disease-related data are presented in Table 1.

Normally-distributed data are presented as mean \pm SD, whereas nonnormally distributed data are expressed as median and IQR. Statistical significance was set at $P < .05$. For the prediction of V200 and VLa4, multiple linear regression models were designed, including only variables that showed statistical association in the previous tests (Spearman's correlation, unpaired *t* test, Mann-Whitney test, Kruskal-Wallis test, 1-way analysis of variance); variables showing collinearity and covariance with other variables were excluded. Moreover, for V200 and VLa4, multiple logistic regression models were designed, with outcomes defined as ≥ 7.5 and < 7.5 m/s for V200, and ≥ 8 and < 8 m/s for VLa4 (cut-offs were chosen based on median values in the study population). Variables were preliminarily screened using univariable logistic regression, and those with a likelihood ratio test $P < .10$ were selected for inclusion in the multivariable model. The model was built using a manual backwards method of elimination of variables based on the likelihood ratio test *P* value and by comparing the corrected Akaike information criteria (AICc).

TABLE 1 Statistical tests used for data analysis in the present study.

Statistical test	Analyzed data
Spearman's correlation	<ul style="list-style-type: none"> • Age vs fitness variables, BAL leukocyte counts, THS, endoscopic scores, CK values, ESGD grade • Fitness variables vs BAL leukocyte counts, THS, endoscopic scores, CK values, ESGD grade
Kruskal-Wallis and Dunn's multiple comparisons test	<ul style="list-style-type: none"> • Age vs DUAO groups, MEA groups • Fitness variables vs DUAO groups, MEA groups • DUAO groups vs BAL neutrophils, BAL eosinophils, BAL mast cells, THS, endoscopic scores, CK values, ESGD grade • MEA groups vs THS, endoscopic scores, CK values, ESGD grade
One-way ANOVA and Tukey's multiple comparisons test	<ul style="list-style-type: none"> • DUAO groups vs BAL macrophages, BAL lymphocytes
Unpaired <i>t</i> test	<ul style="list-style-type: none"> • Sex vs BAL macrophages, BAL lymphocytes • Rhabdomyolysis vs BAL macrophages • Cardiac arrhythmias vs BAL macrophages, BAL lymphocytes • EGGD vs BAL macrophages, BAL lymphocytes
Mann-Whitney test	<ul style="list-style-type: none"> • Age vs cardiac arrhythmias, rhabdomyolysis, EGGD • Sex vs fitness variables, BAL neutrophils, BAL eosinophils, BAL mast cells, THS, endoscopic scores, CK values, ESGD grade • Rhabdomyolysis vs fitness variables, BAL lymphocytes, BAL neutrophils, BAL eosinophils, BAL mast cells, THS, endoscopic scores, CK values, ESGD grade • Cardiac arrhythmias vs fitness variables, BAL neutrophils, BAL eosinophils, BAL mast cells, THS, endoscopic scores, CK values, ESGD grade • EGGD vs fitness variables, BAL neutrophils, BAL eosinophils, BAL mast cells, THS, endoscopic scores, CK values, ESGD grade
Chi-square test	<ul style="list-style-type: none"> • DUAO groups vs sex, cardiac arrhythmias, rhabdomyolysis, EGGD, MEA groups • MEA groups vs sex, cardiac arrhythmias, rhabdomyolysis, EGGD
Fisher's exact test	<ul style="list-style-type: none"> • Sex vs cardiac arrhythmias, rhabdomyolysis, EGGD • Cardiac arrhythmias vs rhabdomyolysis, EGGD • EGGD vs rhabdomyolysis

Abbreviations: BAL, bronchoalveolar lavage; CK, creatine kinase; DUAO, dynamic upper airway obstruction; EGGD, equine glandular gastric disease; ESGD, equine squamous gastric disease; MEA, mild-moderate equine asthma; THS, total hemosiderin score.

TABLE 2 Associations between variables related to different medical disorders in the study population.

	Macrophages	Lymphocytes	Neutrophils	Eosinophils	Mast Cells	MEA group
Premature complexes	ns	ns	ns	ns	$P = .03$	$P = .02$
Serum CK	$P = .01$ $r = .17$	$P < .0001$ $r = -.31$	$P < .0001$ $r = .35$	ns	ns	$P = .0004$
Clinical myopathies	ns	ns	$P = .0001$	ns	ns	ns
DUAO group	ns	ns	ns	ns	ns	ns
EIPH grade	$P = .03$ $r = -.15$	ns	ns	ns	$P = .04$ $r = .14$	ns
THS	ns	$P = .0004$ $r = -.20$	$P = .03$ $r = .15$	$P = .001$ $r = .18$	$P < .0001$ $r = .24$	ns
PLH	ns	ns	ns	ns	ns	ns
TM	ns	ns	ns	ns	ns	ns
TB	ns	ns	ns	ns	ns	ns
Macrophages	-	$P < .0001$ $r = -.65$	$P = .01$ $r = .17$	$P < .0001$ $r = -.26$	$P = .01$ $r = -.18$	$P = .03$
Lymphocytes	$P < .0001$ $r = -.65$	-	$P < .0001$ $r = -.72$	ns	ns	$P < .0001$
Neutrophils	$P = .01$ $r = .17$	$P < .0001$ $r = -.72$	-	ns	$P = .001$ $r = -.22$	$P < .0001$
Eosinophils	$P < .0001$ $r = -.26$	ns	ns	-	ns	$P = .01$
Mast cells	$P = .01$ $r = -.18$	ns	$P = .001$ $r = -.22$	ns	-	$P < .0001$
MEA group	$P = .03$	$P < .0001$	$P < .0001$	$P = .01$	$P < .0001$	-
ESGD grade	ns	ns	ns	ns	ns	ns
EGGD	ns	ns	ns	ns	ns	ns
	THS	EIPH grade	PLH	TM	TB	DUAO group
Premature complexes	ns	ns	ns	ns	ns	ns
Serum CK	ns	ns	ns	$P = .03$ $r = .14$	ns	$P = .02$
Clinical myopathies	ns	ns	ns	ns	ns	ns
DUAO group	$P = .02$	ns	ns	ns	ns	-
EIPH grade	$P < .0001$ $r = .32$	-	$P = .04$ $r = -.13$	ns	ns	ns
THS	-	$P < .0001$ $r = .32$	ns	ns	ns	$P = .02$
PLH	ns	$P = .04$ $r = -.13$	-	ns	$P = .04$ $r = .13$	ns
TM	ns	ns	ns	-	$P < .0001$ $r = .22$	ns
TB	ns	ns	$P = .04$ $r = .13$	$P < .0001$ $r = .22$	-	ns
Macrophages	ns	$P = .03$ $r = -.15$	ns	ns	ns	ns
Lymphocytes	$P = .0004$ $r = -.20$	ns	ns	ns	ns	ns
Neutrophils	$P = .03$ $r = .15$	ns	ns	ns	ns	ns
Eosinophils	$P = .001$ $r = .18$	ns	ns	ns	ns	ns
Mast cells	$P < .0001$ $r = .24$	$P = .04$ $r = .14$	ns	ns	ns	ns

(Continues)

TABLE 2 (Continued)

	THS	EIPH grade	PLH	TM	TB	DUAO group
MEA group	ns	ns	ns	ns	ns	ns
ESGD grade	ns	ns	ns	$P = .01$ $r = .19$	$P = .02$ $r = .18$	ns
EGGD	ns	ns	ns	ns	ns	ns
	Premature complexes	Serum CK	Clinical myopathies	ESGD grade	EGGD	
Premature complexes	-	$P = .01$	ns	ns	ns	ns
Serum CK	$P = .01$	-	$P < .0001$	$P = .001$ $r = .20$	ns	ns
Clinical myopathies	ns	$P < .0001$	-	ns	ns	ns
DUAO group	ns	$P = .02$	ns	ns	ns	ns
EIPH grade	ns	ns	ns	ns	ns	ns
THS	ns	ns	ns	ns	ns	ns
PLH	ns	ns	ns	ns	ns	ns
TM	ns	$P = .03$ $r = .14$	ns	$P = .01$ $r = .19$	ns	ns
TB	ns	ns	ns	$P = .02$ $r = .18$	ns	ns
Macrophages	ns	$P = .01$ $r = .17$	ns	ns	ns	ns
Lymphocytes	ns	$P < .0001$ $r = -.31$	ns	ns	ns	ns
Neutrophils	ns	$P < .0001$ $r = .35$	$P = .0001$	ns	ns	ns
Eosinophils	ns	ns	ns	ns	ns	ns
Mast cells	$P = .03$	ns	ns	ns	ns	ns
MEA group	$P = .02$	$P = .0004$	ns	ns	ns	ns
ESGD grade	ns	$P = .001$ $r = .20$	ns	-	$P = .01$	-
EGGD	ns	ns	ns	$P = .01$	-	-

Abbreviations: CK, creatine kinase; DUAO, dynamic upper airway obstruction; EGGD, equine glandular gastric disease; EIPH, exercise-induced pulmonary hemorrhage; ESGD, equine squamous gastric disease; MEA, mild-moderate equine asthma; ns, nonsignificant; PLH, pharyngeal lymphoid hyperplasia; TB, tracheal blunting; THS, total hemosiderin score; TM, tracheal mucus.

3 | RESULTS

3.1 | Diagnosis

In the study population, a single disorder was diagnosed in 30 horses (11.6%), 2 concomitant disorders in 67 horses (25.9%), 3 disorders in 97 horses (37.4%), 4 disorders in 48 horses (18.5%), 5 disorders in 15 horses (5.8%), and 6 disorders in 1 horse (0.4%); in 1 horse (0.4%), no definitive diagnosis was obtained. The associations among all disorder variables are presented in Table 2.

3.2 | Arrhythmias

Among the horses for which continuous ECG was obtained during and after incremental treadmill exercise, 194 showed no or nonclinically relevant arrhythmias (81.5%), whereas 44 experienced clinically

relevant PCs (18.5%). No significant difference between groups was detected for age and sex distribution.

3.3 | Exertional myopathies

In the study population, median CK activity was 204 (104-407) UI/L; it fell within normal limits in 227 horses (89%), and was >735 UI/L in 28 horses (11%). Neither positive nor negative association was detected between CK and age. Females showed significantly higher postexercise CK activity (median, 307.5 IU/L; IQR, 172.3-572 IU/L) than males (median, 184 IU/L; IQR, 88-332 IU/L; $P < .0001$; Figure 1).

Clinical signs of exertional myopathies were inferred by CK activity or observed in 42 horses (16.5%), 22 females (52.4%) and 20 males (47.6%), with a median age of 3.5 (IQR, 3-5) years. Conversely, 213 horses showed no clinical signs, and included 143 males (67.1%) and 70 females (32.9%), with a median age of 3 (IQR, 3-4) years.

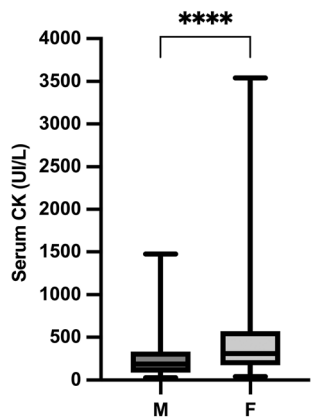


FIGURE 1 Box and whiskers graph representing postexercise serum creatine kinase (CK) activity in males (M) and females (F). The statistical significance is shown as **** $P < .0001$.

TABLE 3 Fitness variables, expressed as median (IQR), in the study population.

Fitness variable	Value
V200	7.5 (6.5-8.5) m/s
VLa4	8 (6.4-9.1) m/s
HRLa4	203.6 (193.1-213.9) bpm
LaC _{max}	22.31 (16.76-27.20) mmol/L
HR _{max}	231 (226-236) bpm
V _{max}	11 (11-11) m/s
HR ₃₀	75 (64-88) bpm
LaC ₃₀	11.88 (6.97-16.69) mmol/L
pH _{min}	7.14 (7.08-7.21)
Hct _{max}	66 (63-69)%

Abbreviations: Hct_{max}, maximum hematocrit; HR₃₀, heart rate at 30 minutes postexercise; HRLa4, heart rate at a plasma lactate concentration of 4 mmol/L; HR_{max}, maximum heart rate; LaC₃₀, plasma lactate concentration at 30 minutes postexercise; LaC_{max}, peak of plasma lactate concentration; pH_{min}, minimum pH; V200, velocity at a heart rate of 200 bpm; VLa4, velocity at a plasma lactate concentration of 4 mmol/L; V_{max}, maximum velocity.

Age did not differ between groups, whereas females were significantly more affected by clinical myopathies compared to males ($P = .02$).

3.4 | Dynamic upper airway obstructions

At HSTE, no DUAO was observed in 140 horses (56.5%), mild DUAOs in 11 horses (4.4%), severe DUAOs in 64 horses (25.8%), and multiple concomitant DUAOs in 33 horses (13.3%). No significant difference in age and sex distribution was observed between groups.

3.5 | Exercise-induced pulmonary hemorrhage

At postexercise tracheobronchoscopy, median EIPH score was 1 (IQR, 0-2). Specifically, no blood was detected in 98 horses (39.8%), and

TABLE 4 Results of Spearman's correlations between different disorder-related variables and fitness variables in the study population.

Fitness variable	Associated disorder-related variables
V200	<ul style="list-style-type: none"> CK ($P < .0001$, $r = -.46$) TM score ($P = .03$, $r = -.14$) ESGD grade ($P = .03$, $r = -.16$)
VLa4	<ul style="list-style-type: none"> CK ($P < .0001$, $r = -.55$) TM score ($P = .02$, $r = -.15$) BAL macrophages ($P = .03$, $r = -.15$) BAL lymphocytes ($P = .001$, $r = .22$) BAL neutrophils ($P = .001$, $r = -.22$) ESGD grade ($P = .04$, $r = -.16$)
HRLa4	<ul style="list-style-type: none"> CK ($P < .0001$, $r = -.29$) BAL macrophages ($P = .02$, $r = -.17$) BAL lymphocytes ($P = .004$, $r = .20$) BAL neutrophils ($P = .03$, $r = -.15$)
LaC _{max}	<ul style="list-style-type: none"> CK ($P < .0001$, $r = .42$) BAL lymphocytes ($P = .03$, $r = -.15$) BAL neutrophils ($P = .004$, $r = .20$)
V _{max}	<ul style="list-style-type: none"> CK ($P < .0001$, $r = -.25$) THS ($P = .03$, $r = .15$) PLH score ($P = .01$, $r = -.15$) BAL mast cells ($P = .002$, $r = .21$)
HR ₃₀	<ul style="list-style-type: none"> PLH score ($P = .02$, $r = -.15$)
LaC ₃₀	<ul style="list-style-type: none"> CK ($P < .0001$, $r = .42$) BAL macrophages ($P = .02$, $r = .16$) BAL lymphocytes ($P < .0001$, $r = -.26$) BAL neutrophils ($P = .001$, $r = .24$)
pH _{min}	<ul style="list-style-type: none"> CK ($P < .0001$, $r = -.30$)
Hct _{max}	<ul style="list-style-type: none"> EIPH grade ($P = .01$, $r = .24$) PLH score ($P < .0001$, $r = -.39$) TM score ($P = .004$, $r = -.26$) TB score ($P = .02$, $r = -.21$)

EIPH score was grade 1 in 57 horses (23.2%), grade 2 in 56 horses (22.8%), grade 3 in 26 horses (10.6%), and grade 4 in 9 horses (3.7%). The median value of THS was 28 (IQR, 8-61), with 37 horses (17.4%) having a THS score >75 and being considered positive for EIPH.

Age was positively correlated with EIPH grade ($P = .002$, $r = .20$) and THS ($P = .002$, $r = .22$). Although median EIPH grade was 1 (IQR, 0-2) in both males and females, higher scores were detected in males ($P = .04$); no differences for THS were observed between males and females.

3.6 | Asthma

At airway endoscopy, median PLH score was 2 (IQR, 1-2), whereas median TM and TB scores were 1 (IQR, 1-2). Age was inversely correlated with PLH ($P < .0001$, $r = -.52$) and with TM ($P = .01$, $r = -.16$), although it was not associated with TB. Females showed higher PLH scores (median, 2; IQR, 2-2.5) compared to males (median, 2; IQR, 1-2; $P = .01$), whereas no associations were observed between sex and TM or TB scores.

The BAL leukocyte differential cell count consisted of means of $44.9 \pm 8.6\%$ macrophages, $36.7 \pm 11.6\%$ lymphocytes, and medians of

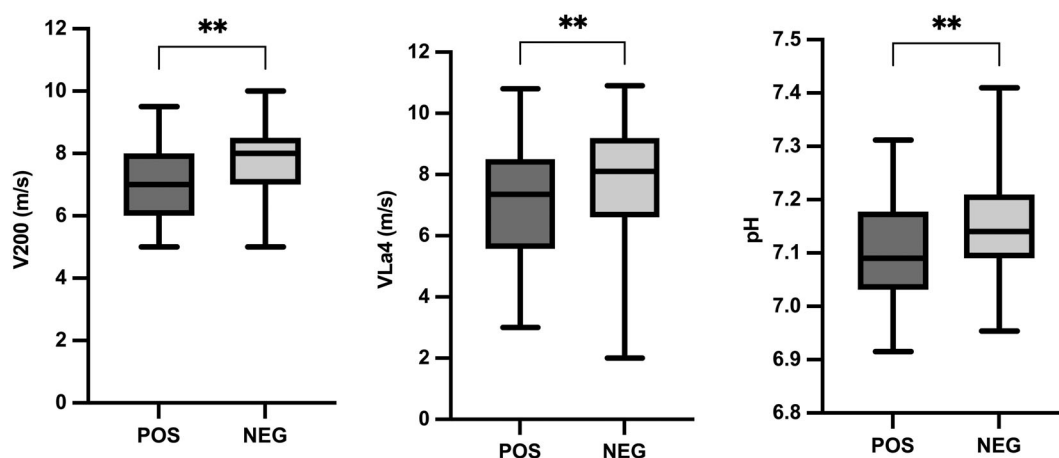


FIGURE 2 Box and whiskers graph representing V200, VLa4 and pHmin in horses with (pos) or without (neg) clinical signs of exertional myopathies. The statistical significance is shown as $**P < .01$.

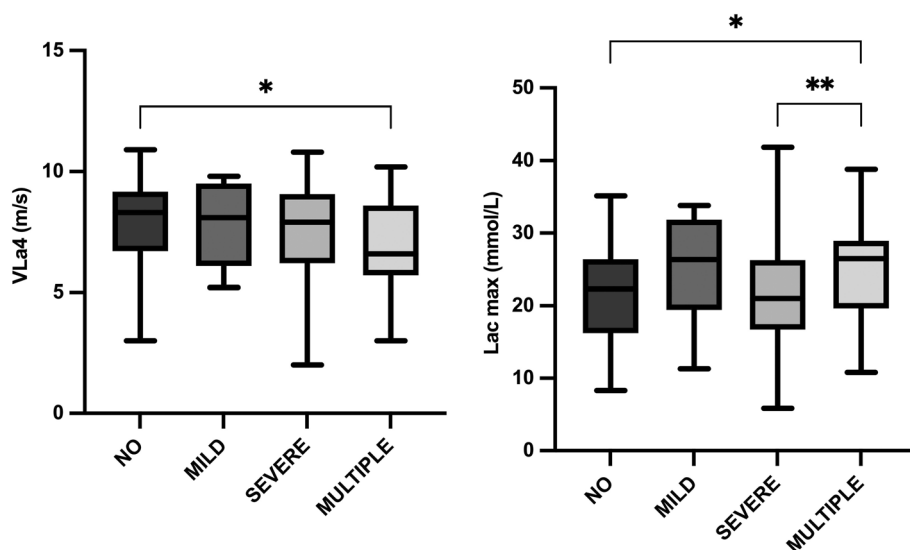


FIGURE 3 Box and whiskers graph representing VLa4 and Lacmax in horses with no DUAO, mild DUAO, severe DUAO, and multiple DUAOs. The statistical significance is shown as $*P < .05$ and $**P < .01$.

8 (IQR, 4.2-17)% neutrophils, 1 (IQR, 0-3)% eosinophils, and 4 (IQR, 3-6)% mast cells. In the population, 6 horses were healthy (2.8%), 23 had neutrophilic MEA (10.7%), 66 had eosinophilic-mastocytic MEA (30.7%), and 120 had mixed MEA (55.8%).

Age was inversely correlated with macrophages ($P = .02$, $r = -.16$) and eosinophils ($P = .02$, $r = -.16$) and positively correlated with mast cells ($P = .01$, $r = .19$). Conversely, age did not differ between healthy and different MEA subtypes. Females had slightly higher macrophages percentages ($46.5 \pm 8.5\%$) than males ($44.1 \pm 8.6\%$; $P = .05$), whereas no other differences for sex were observed for lymphocytes, neutrophils, eosinophils, and mast cells. No difference in sex distribution was detected between groups.

3.7 | Equine gastric ulcer syndrome

At gastroscopy, the squamous mucosa was healthy in 2 horses (1.2%), grade 1 ESGD was observed in 2 horses (1.2%), grade 2 in 17 horses

(9.8%), grade 3 in 32 horses (18.5%), and grade 4 in 120 horses (69.4%). The glandular mucosa was affected by EGGD in 100 horses (57.8%) and healthy in 73 horses (42.2%). Age was not associated with ESGD grade and did not differ between horses with and without EGGD. Median grade of ESGD was equal in females and males and sex distribution did not differ between horses affected and not affected by EGGD.

3.8 | Fitness variables

Medians and IQRs of the fitness variables in the study population are shown in Table 3. Age was inversely correlated with HR_{max} ($P = .02$, $r = -.14$), and positively correlated with V_{max} ($P < .0001$, $r = .35$) and Hct_{max} ($P < .0001$, $r = .48$). The values of V200 and VLa4 were higher in males (V200 median, 8 m/s; IQR, 7-8.5 m/s; VLa4 median, 8.35 m/s; IQR, 6.68-9.3) than in females (V200 median, 7 m/s; IQR, 6-8 m/s; VLa4 median, 7.6 m/s; IQR, 6.2-8.6; V200 $P = .02$;

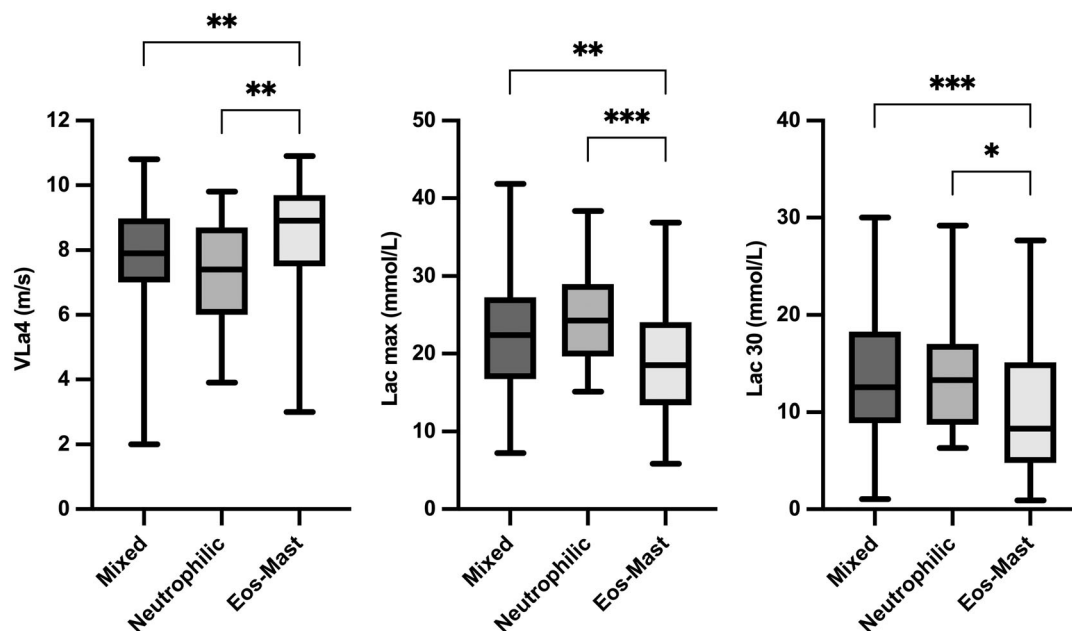


FIGURE 4 Box and whiskers graph representing VL44, Lacmax and Lac30 in horses with mixed MEA, neutrophilic MEA, and eosinophilic-mastocytic (Eos-Mast) MEA. The statistical significance is shown as * $P < .05$, ** $P < .01$, and *** $P < .001$.

TABLE 5 Multiple linear regression model for V200, including only variables which showed a significant association.

Variable	Estimate	95% confidence interval	P value
Sex (male)	.15	−0.21 to 0.50	.42
TM score	−.14	−0.32 to 0.05	.14
ESGD grade	−.12	−0.33 to 0.09	.25
Serum CK	−.0008	−0.001 to 0.0005	<.0001
Model			<.0001

Abbreviations: CK, creatine kinase; ESGD, equine squamous gastric disease; TM, tracheal mucus.

VL44 $P = .01$). Moreover, males reached a higher Hct_{max} ($66.5 \pm 3.8\%$) than females ($64.3 \pm 4.6\%$; $P = .004$). No other associations among age, sex and fitness variables were detected.

3.9 | Associations between disorders and fitness variables

The results of Spearman's correlations between different disorder-related parameters and fitness variables are shown in Table 4. Horses showing clinical signs of myopathies had lower values of V200 ($P = .001$), VL44 ($P = .004$), and pH_{min} ($P = .004$; Figure 2). Fitness variables that differed among DUAO groups included VL44, which was lower in horses with multiple DUAOs compared to horses with no DUAO ($P = .02$), and Lac_{max}, which was higher in horses with multiple DUAOs compared to horses with no DUAO ($P = .05$) and severe DUAO ($P = .01$; Figure 3). When dividing horses based on MEA subtype, VL44 was higher in the eosinophilic-mastocytic MEA group

TABLE 6 Multiple linear regression model for VL44, including only variables which showed a significant association.

Variable	Estimate	95% confidence interval	P value
Sex (male)	0.04	−0.61 to 0.54	0.90
BAL neutrophils	−0.01	−0.04 to 0.02	0.40
TM score	−0.22	−0.52 to 0.08	0.15
Mild DUAO	0.42	−0.85 to 1.68	0.52
Severe DUAO	−0.52	−1.16 to 0.11	0.10
Multiple DUAOs	−0.87	−1.73 to −0.02	0.05
ESGD grade	−0.13	−0.45 to 0.20	0.44
Serum CK	−0.002	−0.002 to 0.001	<.0001
Model			0.0001

Abbreviations: BAL, bronchoalveolar lavage; CK, creatine kinase; DUAO, dynamic upper airway obstruction; ESGD, equine squamous gastric disease; TM, tracheal mucus.

compared to the neutrophilic MEA group ($P = .02$), whereas Lac_{max} and Lac₃₀ were lower in the eosinophilic-mastocytic MEA group compared to the neutrophilic (Lac_{max} $P = .003$; Lac₃₀ $P = .02$) and mixed (Lac_{max} $P = .01$; Lac₃₀ $P = .001$) MEA groups (Figure 4). Horses with EGGD reached a higher Hct_{max} ($66.8 \pm 4.3\%$) than nonaffected horses ($64.6 \pm 3.5\%$; $P = .01$). The percentage of eosinophils in the BAL and the presence of clinically relevant PCs were not associated with any fitness variable.

Models of multiple linear regression designed for prediction of V200 ($r^2 = .17$) and VL44 ($r^2 = .21$) are presented, respectively, in Tables 5 and 6. Models of multivariable logistic regression for the prediction of V200 and VL44 values are shown, respectively, in Tables 7 and 8.

Variable	Odds ratio	95% confidence interval	Likelihood ratio P value
Sex (female)	0.70	0.36-1.34	0.07
MEA group (mixed)	0.84	0.33-2.12	0.07
MEA group (neutrophilic)	0.80	0.38-1.68	0.07
EIPH score	0.79	0.62-1.03	0.07
TM score	0.77	0.55-1.08	0.03
Severity of DUAO	0.72	0.55-0.94	0.005
Serum CK	0.998	0.997-0.999	<0.0001

	Likelihood ratio P value	AUC (95% confidence interval)	ROC curve p value	PPV (%)	NPV (%)
Model	.0001	0.73 (0.66-0.80)	<.0001	69.35	68.75

Abbreviations: AUC, area under the curve; CK, creatine kinase; DUAO, dynamic upper airway obstruction; EIPH, exercise-induced pulmonary hemorrhage; MEA, mild-moderate equine asthma; NPV, negative predictive value; PPV, positive predictive value; ROC, receiver operating characteristic; TM, tracheal mucus.

Variable	Odds ratio	95% confidence interval	Likelihood ratio P value
Sex (female)	0.77	0.34-1.74	.02
BAL neutrophils	0.99	0.95-1.04	.09
EIPH grade	0.74	0.52-1.02	.10
Severity of DUAO	0.80	0.57-1.12	.03
ESGD grade	0.68	0.39-1.11	.03
Serum CK	0.996	0.994-0.998	<.0001

	Likelihood ratio P value	AUC (95% confidence interval)	ROC curve P value	PPV (%)	NPV (%)
Model	<.0001	0.78 (0.70-0.86)	<.0001	68.75	75.41

Abbreviations: AUC, area under the curve; BAL, bronchoalveolar lavage; CK, creatine kinase; DUAO, dynamic upper airway obstruction; EIPH, exercise-induced pulmonary hemorrhage; ESGD, equine squamous gastric disease; NPV, negative predictive value; PPV, positive predictive value; ROC, receiver operating characteristic.

4 | DISCUSSION

Our study confirms the multifactorial medical nature of decreased athletic capacity, with the majority of poorly-performing Standardbred trotters being concomitantly affected by multiple disorders. The most commonly diagnosed disorders were ESGD and MEA, followed by EIPH, EGGD and DUAOs. All disorders, except for PCs, were associated with ≥ 1 fitness variables. In addition to evaluating the effects of each disorder on the athletic capacity of the horses, we investigated the contribution of different disorders to impaired performance in a multifactorial model. In particular, the greatest impairment of fitness, represented by lower values of V200 and VLa4, was related to the diagnosis of exertional myopathies, severe and multiple DUAOs, neutrophilic inflammation of the lower airway, and severe grades of ESGD.

A definitive diagnosis was reached for 254 of 255 horses, which suggests that the diagnostic protocol performed in our study was comprehensive enough to evaluate the most common subclinical non-orthopedic causes of racing poor performance. For the remaining horse, the cause of decreased performance presumably could be

inadequate conditioning, training, or management, or nonspecific musculoskeletal pain. The main limitation of our study is the fact that some horses did not undergo the complete diagnostic protocol, and therefore some disorders may have been underdiagnosed. However, our study was retrospective and included horses evaluated over a period of 20 years, during which time the clinical approach and equipment availability varied slightly. Nevertheless, we believe the study's population size may overcome this limitation.

The highest prevalence was observed for EGUS, which was diagnosed in almost all horses. In particular, severe ESGD (grade 3 or 4) was detected in 87% of the horses, whereas the prevalence of EGGD was approximately 58%, similar to that previously reported.⁴⁹⁻⁵¹ In fact, intensity and duration of exercise are predisposing factors to the development of gastric ulcers in either the squamous and glandular mucosa, and racehorses are most commonly affected.^{49,52,53}

Respiratory diseases represented an equally common cause of poor performance in our population. A diagnosis of MEA was made in 97% of the horses, with the majority of them affected by a mixed inflammatory form. Similarly, previously reported MEA prevalence in

TABLE 7 Multivariate logistic regression model for V200 (equal or higher than 8 m/s vs lower than 8 m/s).

TABLE 8 Multivariate logistic regression model for VLa4 (equal or higher than 8 m/s vs lower than 8 m/s).

Standardbred racehorses in active training ranged from 58% to 97%,^{1,15,54} which may be higher in a selected poorly-performing population. In fact, MEA is universally recognized as 1 of the most common causes of impaired athletic capacity in horses.^{1,26,27} Another common disorder of the lower airway is EIPH, which was assessed in our study by postexercise tracheobronchoscopy and calculation of THS in the BAL fluid. Based on endoscopy, approximately 60% of the horses had blood in the trachea, but only 17% were EIPH-positive based on THS and could therefore be considered as bleeders. In the literature, the prevalence of EIPH varies based on the diagnostic method used. Hemosiderophages in the BAL fluid are found in up to 100% of racehorses,⁵⁵ and tracheal blood is reported in 87% of Standardbreds when examined postrace on at least 3 occasions⁵⁶; when diagnosed by a single endoscopic examination, the prevalence is 43%-75%.⁵⁷ Severity of EIPH seemed to increase with age. Several previous studies have reported such findings,⁵⁸ probably because EIPH is a cumulative condition associated with the number of racing starts.^{59,60} The presence of DUAOs was observed at HSTE in 44% of the horses, which agrees with previous studies.^{3,61,62} In 30% of affected horses, multiple concomitant DUAOs were detected, confirming the findings of others.^{63,64}

In our study, clinically relevant PCs were detected in 18% of the horses during treadmill exercise. The prevalence of PCs in Standardbred racehorses ranges from 54% to 78%^{39,65} but, similar to our results, it decreases to 16%-19% in studies including only clinically relevant arrhythmias.³

Finally, postexercise CK activity was higher than normal in 11% of the horses, whereas clinical myopathies (observed during hospitalization or inferred by history) affected 17% of the horses. Exertional rhabdomyolysis is reported to have a prevalence of 6% among Standardbred racehorses,⁶⁶ which may be higher in our study because of the inclusion of only poorly-performing horses. In fact, similar studies reported prevalences of increased CK activity of 15%-18%,^{1,3} similar to our findings. Females were more prone to develop ER in our population, and such predisposition has been widely recognized.^{3,66-68}

Two or more disorders were simultaneously observed in almost 90% of the horses, similar to what has been reported previously.^{1,3} Among observed associations, those between upper and lower airway disorders are of particular interest. It has been long debated, without consensus, about the possible predisposition of DUAO-affected horses to EIPH or simultaneously to MEA. Some authors, reporting an association between EIPH and DUAO,^{23,69} hypothesized that DUAO may cause an increase in transmural pulmonary pressure gradient, leading to pulmonary capillaries rupture.^{70,71} Other studies, however, found no association between DUAO and EIPH.^{24,72} Similarly, many studies reported associations between DUAO and MEA, hypothesizing that DUAO may predispose to lower airway inflammation or vice versa.^{20,23,69,73-75} Conversely, other authors did not find any relationship between MEA and DUAO.^{7,72,76} Surprisingly, in our study, the severity of DUAO was associated with lower values of THS, whereas no association with MEA subtype or BAL cell populations was observed. Another widely discussed topic concerns a possible cause and effect relationship between MEA and EIPH.^{16,60,77,78} Our results support this hypothesis, because the THS was positively correlated with the BAL percentages of neutrophils,

eosinophils, and mast cells, suggesting that horses with EIPH were more likely to suffer from lower airway inflammation. Moreover, tracheal EIPH score was positively correlated with BAL mastocytosis, which has been reported previously in other studies,^{32,79} although the underlying mechanisms are still unknown. In our study, high postexercise CK activity was positively correlated with detection of clinically relevant PCs, DUAO severity, BAL neutrophilia, TM accumulation, and ESGD severity. The cause and effect relationships between increased CK activity and these disorders are unclear, but some hypotheses may be proposed. First, high CK activity suggests generalized muscular damage and fatigue, which also could involve the myocardium and muscles responsible of the stability of upper airway structures, with the consequent onset of arrhythmias and DUAOs. However, severe DUAOs and lower airway inflammation may result in decreased oxygenation of the muscles, an early transition to anaerobic metabolism, and the onset of muscular fatigue. Therefore, it is not clear whether muscular damage represents the cause or consequence of other disorders. The association observed between ER and severity of ESGD may be explained by the fact that both conditions seem to be associated with a nervous temperament of horses,^{34,36,66,80} although the underlying pathogenetic mechanisms have not yet been determined.

In our study, age was positively correlated with V_{max} and Hct_{max} and inversely correlated with HR_{max} . All of these associations are recognized consequences of conditioning and increased training.^{32,81,82} Moreover, similar to previous reports,^{32,83} males had higher values of $V200$, $VLa4$, and Hct_{max} . Among the diseases affecting fitness, ER was strongly associated with lower values of $V200$, $VLa4$, $HRLa4$, and V_{max} , suggesting a substantial athletic impairment. Moreover, affected horses reached higher peak lactate concentrations and higher lactate concentrations 30 minutes postexercise, and consequently lower values of pH_{min} . Also, neutrophilic inflammation of the lower airways was correlated with lower values of $VLa4$ and $HRLa4$, and higher peak lactate concentration and lactate concentration 30 minutes postexercise. Similarly, TM accumulation was associated with lower $V200$ and $VLa4$. Another respiratory condition affecting fitness was the presence of multiple DUAOs, which were associated with lower $VLa4$ and higher peak lactate concentration. Finally, severe forms of ESGD were correlated with lower $V200$ and $VLa4$. These results suggest that, as previously reported,^{1,7,25,28,37,63,84} these conditions may affect performance by impairing the aerobic capacity of Standardbred trotters, with an early transition to anaerobic metabolism and prolonged lactate accumulation. In the case of ER and DUAOs, it is possible that their occurrence may be a consequence, rather than the cause, of the early switch to anaerobic metabolism and onset of fatigue. Interestingly, cardiac arrhythmias were not associated with any fitness variables, suggesting that they do not play an important role in performance impairment. Similarly, previous studies also found a high prevalence of arrhythmias detected during exercise in clinically healthy well-performing horses.^{39,65} Finally, in our study, higher grades of EIPH were associated with higher V_{max} and Hct_{max} . These findings may be related to the pathogenetic mechanisms of this condition, rather than fitness status.³²

The multivariable models highlighted that ER was the condition most likely associated with unsatisfactory values of $V200$ and $VLa4$,

followed by severe and multiple DUAOs, BAL neutrophilia, ESGD, and TM accumulation. Moreover, horses with high grades of EIPH were less likely to have satisfactory VLa4. However, the coefficient of determination of the multivariable models was relatively low, suggesting that the values of V200 and VLa4, and therefore the level of fitness, only partially depended on the presence of subclinical diseases. In fact, these values can be influenced by conditioning, trainer, preferred ground, stress, and other factors. Horses included in our study came from different training centers over a period of 20 years. Therefore, different training techniques may have influenced our results. Consequently, the proposed statistical models are not necessarily predictive of V200 and VLa4, but rather may indicate which subclinical disorders play a major role in fitness impairment in a complex and multifactorial condition such as poor performance. Finally, because our study was performed retrospectively and only included poorly-performing Standardbred racehorses, future studies should verify whether or not the identified associations are found in a mixed population, including a control group of well-performing horses.

ACKNOWLEDGMENT

No funding was received for this study. The authors acknowledge the University of Milan for the support through the article publication charge (APC) initiative, and all the colleagues, technicians, students, owners and trainers, whose contribution throughout the years made this study possible.

CONFLICT OF INTEREST DECLARATION

Authors declare no conflict of interest.

OFF-LABEL ANTIMICROBIAL DECLARATION

Authors declare no off-label use of antimicrobials.

INSTITUTIONAL ANIMAL CARE AND USE COMMITTEE (IACUC) OR OTHER APPROVAL DECLARATION

Because the present study was performed retrospectively on clinical records, and the procedures were performed on clinical horses for diagnostic purposes, ethical review and approval were waived for this study. All of the procedures were performed according to relevant guidelines and informed owner consent for the use of clinical data was obtained from the owners or holders of the included horses.

HUMAN ETHICS APPROVAL DECLARATION

Authors declare human ethics approval was not needed for this study.

ORCID

Chiara M. Lo Feudo  <https://orcid.org/0000-0002-9964-2753>

Luca Stucchi  <https://orcid.org/0000-0002-2056-5426>

Bianca Conturba  <https://orcid.org/0000-0002-1302-3274>

Giovanni Stancari  <https://orcid.org/0000-0002-8668-1171>

Enrica Zucca  <https://orcid.org/0000-0003-0494-1329>

Francesco Ferrucci  <https://orcid.org/0000-0002-7285-9880>

REFERENCES

- Richard EA, Fortier GD, Pitel PH, et al. Sub-clinical diseases affecting performance in Standardbred trotters: diagnostic methods and predictive parameters. *Vet J.* 2010;185:282-289.
- Morris EA, Seeherman HJ. Clinical evaluation of poor performance in the racehorse: the results of 275 evaluations. *Equine Vet J.* 1991;23:169-174.
- Martin BJ, Reef VB, Parente EG, et al. Causes of poor performance of horses during training, racing, or showing: 348 cases (1992-1996). *J Am Vet Med Assoc.* 2000;216:554-558.
- Widmer A, Doherr MG, Tessier C, et al. Association of increased tracheal mucus accumulation with poor willingness to perform in showjumpers and dressage horses. *Vet J.* 2009;182:430-435.
- Bedenice D, Mazan MR, Hoffman AM. Association between cough and cytology of bronchoalveolar lavage fluid and pulmonary function in horses diagnosed with inflammatory airway disease. *J Vet Intern Med.* 2008;22:1022-1028.
- Fogarty U, Buckley T. Bronchoalveolar lavage findings in horses with exercise intolerance. *Equine Vet J.* 1991;23:434-437.
- Holcombe SJ, Robinson NE, Derksen FJ, et al. Effect of tracheal mucus and tracheal cytology on racing performance in thoroughbred racehorses. *Equine Vet J.* 2006;38:300-304.
- Ivester KM, Couetil LL, Moore GE. An observational study of environmental exposures, airway cytology, and performance in racing thoroughbreds. *J Vet Intern Med.* 2018;32:1754-1762.
- Salz RO, Alhern BJ, Boston R, et al. Association of tracheal mucus or blood and airway neutrophilia with racing performance in thoroughbred horses in an Australian racing yard. *Aust Vet J.* 2016;94:96-100.
- Sauze MN, Gummow B. Prevalence of pharyngeal, laryngeal and tracheal disorders in thoroughbred racehorses, and effect on performance. *Vet Rec.* 2009;165:431-435.
- Wylie CE, Newton JR. A systematic literature search to identify performance measure outcomes used in clinical studies of racehorses. *Equine Vet J.* 2018;50:304-311.
- Egenvall A, Byström A, Pökelmann M, et al. Rein tension in harness trotters during on-track exercise. *Front Vet Sci.* 2022;9:987852.
- Couetil LL, Denicola DB. Blood gas, plasma lactate and bronchoalveolar lavage cytology analyses in racehorses with respiratory disease. *Equine Vet J.* 1999;30:77-82.
- Mazan MR, Hoffman AM. Effects of aerosolized albuterol on physiologic responses to exercise in Standardbreds. *Am J Vet Res.* 2001;62:1812-1817.
- Nolen-Walston RD, Harris M, Agnew ME, et al. Clinical and diagnostic features of inflammatory airway disease subtypes in horses examined because of poor performance: 98 cases (2004-2010). *J Am Vet Med Assoc.* 2013;242:1138-1145.
- Sanchez A, Couetil LL, Ward MP, et al. Effect of airway disease on blood gas exchange in racehorses. *J Vet Intern Med.* 2005;19:87-92.
- Courouce A, Chatard JC, Auvinet B. Estimation of performance potential of Standardbred trotters from blood lactate concentrations measured in field conditions. *Equine Vet J.* 1997;29:365-369.
- McGowan C. Clinical pathology in the racing horse: the role of clinical pathology in assessing fitness and performance in the racehorse. *Vet Clin Equine.* 2008;24:405-421.
- Leleu C, Cotrel C, Courouce-Malblanc A. Relationships between physiological variables and race performance in French Standardbred trotters. *Vet Rec.* 2005;156:339-342.
- Courouce-Malblanc A, Pronost S, Fortier G, et al. Physiological measurements and upper and lower respiratory tract evaluation in French Standardbred trotters during a standardised exercise test on the treadmill. *Equine Vet J Suppl.* 2002;34:402-407.
- Lindner A. Relationships between racing times of Standardbreds and V4 and V200. *J Anim Sci.* 2010;88:950-954.
- Davie AJ, Priddle TL, Evans DL. Metabolic responses to submaximal field exercise tests and relationships with racing performance in pacing Standardbreds. *Equine Exerc Physiol.* 2002;34:112-115.

23. Courouce-Malblanc A, Deniau V, Rossignol F, et al. Physiological measurements and prevalence of lower airway diseases in trotters with dorsal displacement of the soft palate. *Equine Vet J Suppl.* 2010;42:246-255.
24. Davidson EJ, Harris M, Martin BB, Nolen-Walston R, Boston RC, Reef V. Exercising blood gas analysis, dynamic upper respiratory tract obstruction, and postexercising bronchoalveolar lavage cytology. A comparative study in poor performing horses. *J Equine Vet Sci.* 2011;31:475-480.
25. Lo Feudo CM, Stucchi L, Cavicchioli P, et al. Association between dynamic upper airway obstructions and fitness parameters in Standardbred racehorses during high-speed treadmill exercise. *J Am Vet Med Assoc.* 2022;260:1343-1350.
26. Couetil LL, Cardwell JM, Gerber V, et al. Inflammatory airway disease of horses—revised consensus statement. *J Vet Intern Med.* 2016;30:503-515.
27. Kinnison T, McGilvray TA, Couetil LL, et al. Mild-moderate equine asthma: a scoping review of evidence supporting the consensus definition. *Vet J.* 2022;286:105865.
28. Lo Feudo CM, Stucchi L, Conturba B, Stancari G, Ferrucci F. Impact of lower airway inflammation on fitness parameters in Standardbred racehorses. *Animals.* 2022;12:3228.
29. Hinchcliff KW, Couetil LL, Knight PK, et al. Exercise induced pulmonary hemorrhage in horses: American College of Veterinary Internal Medicine consensus statement. *J Vet Intern Med.* 2015;29:743-758.
30. Birks EK, Shuler KM, Soma LR, et al. EIPH: posttrace endoscopic evaluation of Standardbreds and thoroughbreds. *Equine Vet J Suppl.* 2002;34:375-378.
31. MacNamara B, Bauer S, Iafe J. Endoscopic evaluation of exercise-induced pulmonary hemorrhage and chronic obstructive pulmonary disease in association with poor performance in racing Standardbreds. *J Am Vet Med Assoc.* 1990;196:443-445.
32. Lo Feudo CM, Stucchi L, Stancari G, et al. Associations between exercise-induced pulmonary hemorrhage (EIPH) and fitness parameters measured by incremental treadmill test in Standardbred racehorses. *Animals.* 2022;12:449.
33. Franklin SH, Van Erck-Westergren E, Bayly WM. Respiratory responses to exercise in the horse. *Equine Vet J.* 2012;44:726-732.
34. Valberg SJ. Muscle conditions affecting sport horses. *Vet Clin Equine.* 2018;34:253-276.
35. Andrews FM. Poor performance: can heartburn slow a horse? *Equine Vet Educ.* 2008;20:125-126.
36. Sykes BW, Hewetson M, Hepburn RJ, Luthersson N, Tamzali Y. European college of equine internal medicine consensus statement—equine gastric ulcer syndrome in adult horses. *J Vet Intern Med.* 2015;29:1288-1299.
37. Lo Feudo CM, Stucchi L, Conturba B, Stancari G, Zucca E, Ferrucci F. Equine gastric ulcer syndrome affects fitness parameters in poorly performing Standardbred racehorses. *Front Vet Sci.* 2022;9:1014619.
38. Reef VB, Davidson EJ, Slack J, Stefanovski D. Hypercapnia and hyperlactatemia were positively associated with higher-grade arrhythmias during peak exercise in horses during poor performance evaluation on a high-speed treadmill. *Vet J.* 2020;266:105572.
39. Slack J, Stefanovski D, Madsen TF, Fjordbakk CT, Strand E, Fintl C. Cardiac arrhythmias in poorly performing Standardbred and Norwegian-Swedish coldblooded trotters undergoing high-speed treadmill testing. *Vet J.* 2021;267:105574.
40. Alberti E, Stucchi L, Lo Feudo CM, et al. Evaluation of cardiac arrhythmias before, during, and after treadmill exercise testing in poorly performing Standardbred racehorses. *Animals.* 2021;11:2413.
41. Newell J, Higgins D, Madden N, et al. Software for calculating blood lactate endurance markers. *J Sports Sci.* 2007;25:1403-1409.
42. Stucchi L, Valli C, Stancari G, Zucca E, Ferrucci F. Creatine-kinase reference intervals at rest and after maximal exercise in Standardbred racehorses. *Comp Exerc Physiol.* 2019;15:319-325.
43. Hinchcliff KW, Jackson MA, Brown JA, et al. Tracheobronchoscopic assessment of exercise-induced pulmonary hemorrhage in horses. *Am J Vet Res.* 2005;66:596-598.
44. Burrell MH. Endoscopic and virological observations on respiratory disease in a group of young thoroughbred horses in training. *Equine Vet J.* 1985;17:99-103.
45. Gerber V, Straub R, Marti E, et al. Endoscopic scoring of mucus quantity and quality: observer and horse variance and relationship to inflammation, mucus viscoelasticity and volume. *Equine Vet J.* 2004;36:576-582.
46. Lo Feudo CM, Stucchi L, Alberti E, et al. The role of thoracic ultrasonography and airway endoscopy in the diagnosis of equine asthma and exercise-induced pulmonary hemorrhage. *Vet Sci.* 2021;8:276.
47. Doucet Y, Viel L. Alveolar macrophage graded hemosiderin score from bronchoalveolar lavage in horses with exercise-induced pulmonary hemorrhage and controls. *J Vet Intern Med.* 2002;16:281-286.
48. Andrews FM, Bernard W, Byars D, et al. Recommendations for the diagnosis and treatment of equine gastric ulcer syndrome (EGUS). *Equine Vet Educ.* 1999;11:262-272.
49. Sykes BW, Bowen M, Habershon-Butcher JL, Green M, Hallowell GD. Management factors and clinical implications of glandular and squamous gastric disease in horse. *J Vet Intern Med.* 2019;33:233-240.
50. Begg LM, O'Sullivan CB. The prevalence and distribution of gastric ulceration in 345 racehorses. *Aust Vet J.* 2003;81:199-201.
51. Rabuffo TS, Orsini JA, Sullivan E, Engiles J, Norman T, Boston R. Associations between age or sex and prevalence of gastric ulceration in Standardbred racehorses in training. *J Am Vet Med Assoc.* 2002;221:1156-1159.
52. Roy MA, Vrins A, Beauchamp G, Doucet MY. Prevalence of ulcers of the squamous gastric mucosa in Standardbred horses. *J Vet Intern Med.* 2005;19:744-750.
53. Pedersen S, Cribb A, Windeyer M, et al. Risk factors for equine glandular and squamous gastric disease in show jumping warmbloods. *Equine Vet J.* 2018;50:747-751.
54. Depecker M, Richard EA, Pitel PH, Fortier G, Leleu C, Couroucé-Malblanc A. Bronchoalveolar lavage fluid in Standardbred racehorses: influence of unilateral/bilateral profiles and cut-off values on lower airway disease diagnosis. *Vet J.* 2014;199:150-156.
55. Meyer T, Fedde M, Gaughan E, et al. Quantification of exercise-induced pulmonary haemorrhage with bronchoalveolar lavage. *Equine Vet J.* 1998;30:284-288.
56. Lapointe J, Vrins A, McCarville E. A survey of exercise-induced pulmonary haemorrhage in Quebec Standardbred racehorses. *Equine Vet J.* 1994;26:482-485.
57. Crispe EJ, Lester GD. Exercise-induced pulmonary hemorrhage—is it important and can it be prevented? *Vet Clin Quine.* 2019;35:339-350.
58. McGilvray TA, Cardwell JM. Training related risk factors for exercise induced pulmonary haemorrhage in British National Hunt racehorses. *Equine Vet J.* 2021;54:283-289.
59. Costa MFM, Thomassian A. Evaluation of race distance, track surface and season of the year on exercise-induced pulmonary haemorrhage in flat racing thoroughbreds in Brazil. *Equine Vet J Suppl.* 2006;36:487-489.
60. Hinchcliff KW, Morley PS, Jackson MA, et al. Risk factors for exercise-induced pulmonary haemorrhage in thoroughbred racehorses. *Equine Vet J.* 2010;42:228-234.
61. Priest DT, Cheetham J, Regner AL, et al. Dynamic respiratory endoscopy of Standardbred racehorses during qualifying races. *Equine Vet J.* 2012;44:529-534.

62. Kannegieter NJ, Dore ML. Endoscopy of the upper respiratory tract during treadmill exercise: a clinical study of 100 horses. *Aust Vet J*. 1995;72:101-107.
63. Franklin SH, Allen KJ. Assessment of dynamic upper respiratory tract function in the equine athlete. *Equine Vet Educ*. 2017;29:92-103.
64. Tan RHH, Dowling BA, Dart AJ. High-speed treadmill videoendoscopic examination of the upper respiratory tract in the horse: the results of 291 clinical cases. *Vet J*. 2005;170:243-248.
65. Buhl R, Petersen EE, Lindholm M, Bak L, Nostell K. Cardiac arrhythmias in Standardbreds during and after racing—possible association between heart size, valvular regurgitations, and arrhythmias. *J Equine Vet Sci*. 2013;33:590-596.
66. Isgren CM, Upjohn MM, Fernandez-Fuente M, et al. Epidemiology of exertional rhabdomyolysis susceptibility in Standardbred horses reveals associated risk factors and underlying enhanced performance. *PLoS One*. 2010;5:e11594.
67. MacLeay JM, Sorum SA, Valberg SJ, Marsh WE, Sorum MD. Epidemiologic analysis of factors influencing exertional rhabdomyolysis in thoroughbreds. *Am J Vet Res*. 1999;60:1562-1566.
68. Upjohn MM, Archer RM, Christley RM, McGowan CM. Incidence and risk factors associated with exertional rhabdomyolysis syndrome in National Hunt racehorses in Great Britain. *Vet Rec*. 2005;156:763-766.
69. Trope G. Dynamic endoscopy of the equine upper airway—what is significant? *Vet Rec*. 2013;172:499-500.
70. Van Erck-Westergren E, Franklin SH, Bayly WM. Respiratory diseases and their effects on respiratory function and exercise capacity. *Equine Vet J*. 2013;45:376-387.
71. Ducharme NG, Hackett RP, Glead RD, et al. Pulmonary capillary 586 pressure in horses undergoing alteration of pleural pressure by imposition of various upper airway resistive loads. *Equine Vet J*. 2010;31:27-33.
72. Lo Feudo CM, Stancari G, Collavo F, et al. Upper and lower airways evaluation in 360 racehorses with dynamic upper airway obstruction. *Animals*. 2022;12:1563.
73. Van Erck E. Dynamic respiratory videoendoscopy in ridden sport horses: effect of head flexion, riding and airway inflammation in 129 cases. *Equine Vet J*. 2011;43:18-24.
74. Joo K, Povaszai A, Nyerges-Bohak Z, et al. Asthmatic disease as an underlying cause of dorsal displacement of the soft palate in horses. *J Equine Vet Sci*. 2021;96:103308.
75. Wysocka B, Klucinski W. The occurrence of dynamic structural disorders in the pharynx and larynx, at rest and during exercise, in horses diagnosed with mild and moderate equine asthma (inflammatory airway disease). *Pol J Vet Sci*. 2018;21:203-211.
76. Koblinger K, McDonald NK, Wasko A, et al. Endoscopic assessment of airway inflammation 637 in horses. *J Vet Intern Med*. 2011;25:1118-1126.
77. Newton JR, Wood JLN. Evidence of an association between inflammatory airway disease and EIPH in young thoroughbreds 566 during training. *Equine Vet J*. 2002;34:417-424.
78. Da Silva KM, Otaka JNP, Goncalves CAP, et al. Association between exercise-induced pulmonary hemorrhage and inflammatory airway disease in polo ponies. *J Equine Sci*. 2017;28:55-59.
79. Boshuizen B, De Bruijn M, Dewulf J, et al. Sport horses with inflammatory airway disease (IAD) with predominantly eosinophils or mast cells are more predisposed to poor performance, exercise induced pulmonary haemorrhage (EIPH) and have shorter careers when compared to horses with neutrophilic iad. *Equine Vet J*. 2014;46:7.
80. McClure SR, Glickman LT, Glickman NW. Prevalence of gastric ulcers in show horses. *J Am Vet Med Assoc*. 1999;215:1130-1133.
81. McKenzie E. Hematology and serum biochemistry of the equine athlete. In: Hinchcliff KW, Kaneps AJ, Geor RJ, eds. *Equine Sports Medicine and Surgery*. 2nd ed. Philadelphia, PA: Saunders Elsevier; 2014:921-929.
82. Vincent TL, Newton JR, Deaton CM, et al. Retrospective study of predictive variables for maximal heart rate (HR_{max}) in horses undergoing strenuous treadmill exercise. *Equine Vet J Suppl*. 2006;36:146-152.
83. Perrson SGB. Heart rate and blood lactate responses to submaximal treadmill exercise in the normally performing Standardbred trotter—age and sex variations and predictability from the total red blood cell volume. *Zentralb Veterinarmed A*. 1997;44:125-132.
84. Nieto JE, Snyder JR, Vatistas NJ, Jones JH. Effect of gastric ulceration on physiologic responses to exercise in horses. *Am J Vet Res*. 2009;70:787-795.

How to cite this article: Lo Feudo CM, Stucchi L, Conturba B, Stancari G, Zucca E, Ferrucci F. Medical causes of poor performance and their associations with fitness in Standardbred racehorses. *J Vet Intern Med*. 2023;1-14. doi:[10.1111/jvim.16734](https://doi.org/10.1111/jvim.16734)