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Review

# Back to basic: choosing the appropriate surface disinfectant

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#### Abstract:

From viruses to bacteria, our lives are filled with exposure to germs. In built environment exposure to infectious microorganisms and their byproducts is clearly linked to human health. In the last year, public health emergency surrounding the COVID-19 pandemic, stressed the importance of having good biosafety measures and practices. In fact, to prevent infection from spreading and to maintain the barrier, disinfection and hygiene habits are crucial, especially when the microorganism can persist and survive on surfaces. Contaminated surfaces are called fomites and on them microorganisms can survive even for months. As a consequence fomites serve as second reservoir and transfer pathogens between hosts. The kwowledge of microorganism, type of surface and antimicrobial agent is fundamental to develop the best approach to sanitize fomites and to obtain good disinfection level. Hence, this review has the purpose to briefly describe the organisms, the kind of risk associated with them and the main classes of antimicrobials for surfaces, in order to help choosing the right approach to prevent exposure to pathogens.

**Keywords:** antimicrobial; disinfectant; surface disinfection; fomite; surface contamination; microorganisms

# 1. Introduction

In the build environment, especially considering an indoor lifestyle, to touch objects or surfaces which surround us is integral to everyday life. Such objects or surfaces if contaminated are called fomites and, in the 21th century, their role in disease transfer is higher than ever in human history. Indeed, most microorganisms found in indoor environment are inactive, dormant or dead and either show no impact on human health or are even beneficial. Nevertheless, fomites can become contaminated by pathogenic organisms which have a variety of negative health consequences. In fact, microorganisms can survive even many months and multiply on surfaces or objects [1], leading to development of secondary reservoirs. As a consequence fomites can serve as mechanism for transfer between hosts, just think to doorknobs, elevator buttons, hand rails, phones, keyboards, writing implement etc., that are touched by a person that afterwards will handle other objects (Figure 1).

Furthermore, experimental data show that touching a fomite carries approximately the same risk for the acquisition of a lot of microorganisms (i.e. Methicillin-Resistant *Staphylococcus aureus* - MRSA, Vancomycin-Resistant *Enterococcus* - VRE and *Clostridium difficile*) on hands as touching an infected patient [2–5]. Consequently, preventing transmission of pathogens with disinfection procedures must be carried out not only in the high risk sectors, like laboratories, operating rooms, intensive care units, or food-handling settings, but also for hygienic behaviour in everyday life on floors and on all the surfaces that frequently are touched with hands.

Therefore, environmental disinfection, hygiene habits and the consequent maintenance of barriers are crucial in preventing infection from spreading. To develop effective policies and regulations to minimize the risk of trasmission is strictly necessary to evaluate which organisms are present on the fomites. Furthermore, the choice of the effective

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antimicrobial agent is also based on the risk assessment of the microrganisms and the type of fomites

Public health emergency surrounding the COVID-19 pandemic, stressed the importance of having good biosafety measures and practices, as never before. On these basis, this review has the porpuse to briefly describe the organisms, the kind of risk associated with them and the major characteristic of the main classes of antimicrobials for surfaces in order to help in choosing the right approach to prevent exposure to pathogens.

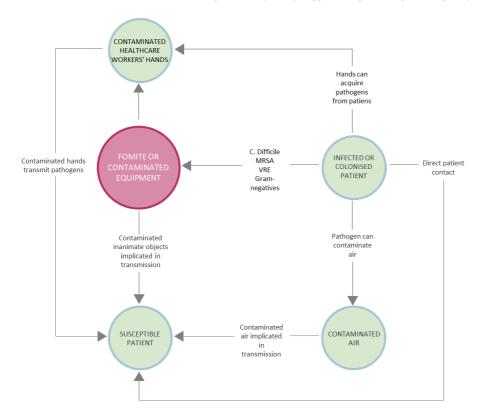


Figure 1. Generic transmission route.

## 2. Most Common Microorganisms on Fomites and Risk Associated with

Primary goal of disinfecting procedures is the inactivation of organisms on fomites. Generally microorganisms belong to diverse group such as bacteria, viral and protozoan species [6]. These biological agents are widely found in the natural environment and, as a result, they can be found either in many work sectors or household contexts. The majority of these microorganisms are harmless; however, some of them or their metabolites may cause diseases. Therefore, the knowledge of these organisms and their survival are fundamental to choose the right antimicrobial agents and to implement effective tactics.

2.1 Bacteria

Bacteria are single-celled organisms (0.3-1.5  $\mu m)$  with independent life and replication cycle. Bacterial cells are generally surrounded by two concentric protective layers: an inner cell membrane and an outer cell wall [7]. The cytoplasmatic membrane shares a similar structure with the eukaryote's one, but there are no sterols. Here, proteins involved in the energy production can be found like some respiratory chain protein as well as photosynthetic protein in photosynthetic bacteria that lack chloroplast. Among the proteins that constitute the cell wall the main one is peptidoglycan (PGN), also known as murein, which provides rigidity to the structure and counteracts the osmotic pressure of the cytoplasm. PGN is characterized by a glucidic backbone of alternating units of two azotated carbohydrates, namely N-acetylglucosamine (GlcNAc) and N-acetylmuramic acid (MurNAc). Each MurNAc is cross-linked to a short amino acid chain, which can vary with different bacterial species [8]. The differences in structural characterization of peptidoglycan define two taxonomic categories: Gram-positive and Gram-negative bacteria (Figure 2).

In Gram-positive bacteria, peptidoglycans make up about 20% of the cell wall dry weight; while in Gram-negative bacteria the thicker peptidoglycan layer contains about 10% of the cell wall dry weight [9]. Furthermore, Gram-positive cell wall has a significant amount (up to 50%) of teichoic and teichuronic acid, which are involved in pathogenesis and play key roles in antibiotic resistance [10].

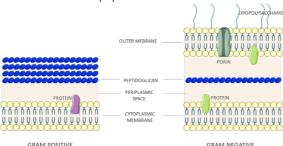


Figure 2. Gram-Negative Versus Gram-Positive Cell Walls.

Certain bacteria may even have a third outermost protective layer called capsule. Whip-like extensions often cover the surfaces of bacteria — long ones called flagella and short ones called pili — in order to become motile and seek out nutrients [11]. An alternative resource exploited by some bacteria is the formation of endospores that are dormant and highly resistant cells able to preserve the genetic material. This ruse helps the bacteria to survive even without nutrient or under extreme stress [12].

Among endospore producing bacteria the most common are the *Bacillus* and *Clostridium* genera [13]. Table 1 reports several endospore forming bacteria and their relative clinical manifestations.

Table 1. Common endospore producing bacteria and their clinical manifestations.

Bacterial species	Clinical manifestation
B. anthracis	anthrax
B. cereus	foodborne illness
B. subtilis	not pathogen
C. botulinum	botulism
C. perfringens	gas gangrene
C. tetani	tetanus

Another bacteria's survival mechanism is the formation of biofilm: clusters of bacteria that

are attached to a surface and/or to each others. During the biofilm development, bacteria secrete extracellular polymeric substances (EPS) which are crucial to the production of an extracellular matrix [14]. This network maintains cohesion between cells and the surface and it protects the accumulation of microorganisms against chemical, biological and mechanical stressors. In this complex arrangement of cells, there are interstitial void spaces in which water flows so nutrients and oxygen diffuse [15]. Since biofilm provides protection from harsh conditions and resistance towards antibiotics, it represents a serious global health concern. Furthermore, biofilm is involved in persistent chronic infections [16,17] and may potentially contribute in their pathogenesis [18].

#### 2 2 Virus

Virus are subcellular organisms with submicrosopic dimension (nm). Their core has either DNA (deoxyribonucleic acid) or RNA (ribonucleic acid) as genetic material. The core is covered by a protein coat [19], called capsid, whose role is to protect it from degradation. Furthermore, the protein coat allows the virus to attack to specific receptor of the host cell. In fact, viruses are obligate intracellular parasites [20], so they need host ribosomes to synthesize viral proteins. Capsid proteins are codified by the viral genome, whose short length entail a limited number of proteins with a specific function. This leads to a capsid constituted by repetitive units of one or few proteins combined in a continuous structure [21], which can have an helicoidal or geometric symmetry. The former is characterized by an helicoidal distribution around the nucleic acid while the latter by a polyhedral or a spherical shape. Besides these styles, a few viruses have a complex architecture like poxviruses, geminiviruses and many bacteriophages [22] (Figure 3).

Furthermore, some viruses show a further shell, called envelope, constituted by viral proteins and lipids. The envelope shields the virus from the immune system's detection and, in addition, facilitates the fusion with the host cell membrane [21].

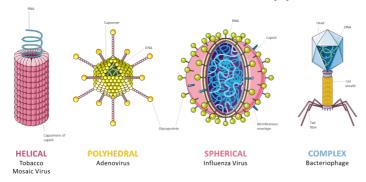
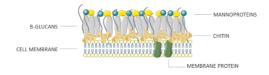


Figure 3. Types of viruses architecture.

## 2.3 Fungi

Fungi are a large group of eukaryotic organisms, mono or pluricellular, that also include yeast and moulds. Since these organisms have a rigid cell wall (rich in chitin and other polysaccharides, especially glucans as depicted in Figure 4) [23], they feed themselves secreting digestive enzymes and by absorbing organic matter from the environment: thus, they are called heterotrophic organisms. Some fungi can live by decomposing dead organic matter (saprobic) while others are parasite of organisms, even fungi, or have developed complex symbionts as in lichens and mycorrhizae [24].



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Figure 4. Fungal wall.

## 2.4 Microbiological risk assessment

According to the Code of Practice to the Safety, Health and Welfare at Work (Biological Agents) Regulation 2020 [25] the biological agents can be classified into four risk groups, reported in Table 2. The classification takes into account:

- Virulence -Ability of the microorganism to penetrate and multiplicate inside the host
- Pathogenicity -Severity of the disease that may result;
- Transmissibility -Capability of the microorganism to be transmitted from one organism to another;
- Treatment -Availability, if any, of effective prophylaxis or therapy.

Table 2. Classification of biological agents.

Risk classification	Description	Examples		
Category 4	Pathogen that may cause severe illness in humans and may be a serious hazard for workers; the biological agent can spread in the community, and usually there are not effective treatments available	Ebola virus, Lassa virus, Smallpox virus	MICF	I-RISK OBES
Category 3	Pathogen that may cause severe ill- ness in humans and be a serious hazard for workers; the biological agent may spread in the commu- nity, but usually effective treat- ments are available	HIV, Bacillus anthracis, HBV, HCV, Mycobacte- rium tuberculosis SARS-CoV-2	CATEGORY 4  CATEGORY 3	
Category 2	Pathogen that may cause pathology in humans and be potential hazard for workers; it's unlikely that can be spread in the community; usually there are effective treatments	Measles virus, Salmo- nella, Legionella		v-risk
Category 1	Pathogen with low probability of developing diseases in human or- ganism	Nonpathogenic strains of Escherichia	MIC	ROBES

Disinfection policies should be also based on risk assessment in order to control cross-contamination while reducing the risk caused by exposure to infectious agents. The  $evaluation \ of \ the \ surface's \ risks \ and \ type \ together \ with \ the \ nature \ of \ the \ pathogen \ agent(s)$ should lead to the use of an appropriate and effective antimicrobial agent. Such approaches must be learned by everyone since their implementation in the routine measure improves both cleaning performance and infection prevention [26].

However, as far as possible, the number of antimicrobials to be used should be limited not

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only for healthy and economic reasons but also to reduce environmental pollution. Not least, the discharge of waste biocides into the environment may promote the development of both biocide and antibiotic resistance [27].

## 3. Factors that Affect the Activity of Antimicrobials

The activity of the antimicrobial agents depends on several factors, some of which are intrinsic qualities of the organism, others derived from the chemicals and external physical environment. More specifically need to be listed:

## Number and type of microorganism

There is no disinfectant that is able to effectively act on all microorganisms classes. So proper choose of chemical germicides is fundamental. Furthermore, there are some microbials that can persist on surfaces showing resistance to these products: for example, the production of endospores or biofilm matrix protects the pathogens from environmental influences[12,28].

### Type and concentration of the antimicrobial

After choosing the proper disinfectant, concentration of the active ingredient is a key factor: the influence of changing in concentration of the active(s) can be measured experimentally, with the determination of the kinetics of inactivation. Moreover, the knowledge of the effect of dilution or concentration on the activity of a sanitazing agent provides some valuable informations that could lead to a reduction of the exposure time.

Furthermore, microbicidal concentration is also a central concept in the microbial resistance field and it is especially important nowadays with increasing knowledge and restrictions on the environmental discharges of potentially harmful chemicals [29].

# pH of the solution

The pH of the solution can affect the efficacy of the disinfection in two ways: a change in the agent itself and a change in the interactions between the microbicide and the microbial cell.

For example, a number of microbicides are effective in their unionized form (Table 3). Thus, pH level would affect their degree of dissociation and would decrease their overall activity. In contrast, other molecules are more effective in their ionized form. Beside these considerations it should also keep in mind that alteration of the pH level could affect the compound's stability.

As a matter of fact, disinfectant products in sanitary field are formulated to guarantee, at certain level of pH, the maximum germicidal efficacy.

Table 3. Effects of pH level on antimicrobial activity.

Activity as environmental pH Classes of disinfectants Mechanisms increases Increase in the degree of dissociation of the Phenols and organic acids molecules Undissociated hypochlorous acid is the most Decreased activity Hypochlorites fast-acting species At low pH, iodine, the most powerful antimi-Iodine crobial species, is the dominating one Quaternary ammonium Increase in the degree of ionization of bacte-Increased activity compounds rial surface groups leading to an increase in (QACs) binding

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### Formulation

The formulation of a disinfectant deeply affects its activity. Several excipients, such as solvents, surfactants, thickeners, chelating agents, colors and fragrances [102-105], can be found in these products; they can interact with the microorganisms or with the active itself and ultimately affect the activity of the formulated product. Most of the information on the effect of different excipients on the activity of disinfectants are not available, since they are often trade secrets.

### · Length of exposure

The microbicidal activity of chemicals usually increases with the rise of the contact time. However, there is not a direct correlation between contact time and microbicidal activity, maybe due to other factors. Contact times for disinfectants are specific for each material and manufacturer. Therefore, all recommendations for use of disinfectants should follow manufacturers' specifications that must be reported on the label.

### Temperature

Temperature can be an important parameter that influences the pathogen' survival. High temperature can impact vital protein and enzymes, as well as the genome. Moreover, high temperature can boost and speed up the germicidal activity of many chemicals resulting in reduced time and improved efficacy. As drawback, high temperature can accelerate the evaporation of the chemicals and also degrade them. Particular care is needed in using and in stocking such chemicals in tropical regions, where their shelf-life may be reduced because of high room temperature;

## Type of surfaces and precleaning process

The location of microorganisms must be considered as well: to sanitize an instrument with multiple pieces or joints and channels is more difficult than a flat surface. Only surfaces that directly contact the germicide will be sanitized. Indeed, the presence of dirt is the principal reason for disinfection failure, since it could interact with the microbicide, reducing its availability or interact with the microorganisms, giving protection. Moreover, material characteristics of the surface may influence the survival of microorganism as well: for example, porous surfaces are more difficult to clean and, consequently, to disinfect. Pretreatment of surfaces, especially when visibly soiled, is fundamental to ensure or improve the microbicidal efficacy of the disinfection procedure.

Beside the activity that is influenced by the factors listed upon, ideally, an antimicrobial agent should: 1) have a wide spectrum against microorganisms; 2) be rapid in its action; 3) be compatible with many materials; 4) be safe for humans and the environment.

# 4. Most Common Antimicrobial Classes

At the present time, there are numerous substances to be used on surfaces that are claimed as antimicrobial agents and they are formulated alone or in combination. The most common disinfectants can be roughly divided as: halogens, alcohols, quaternary ammonium compounds (QACs), peroxigens, ozone and UV. Generally, these antimicrobials damage a specific part of the microorganism as reported in Figure 5.

Figure 5. Mechanisms of biocide actions on microorganisms.

## 4.1. Halogens

## 4.1.1 Chlorine compounds

Historically, the most widely used antimicrobial agents belonging to halogens are chlorine and chlorine releasing compounds.

Since elemental chlorine gas (Cl2) is hazardous it must be banned either from work places or household environment and substituted by chlorine-releasing agents.

The most commonly used chlorine-releasing agent is sodium hypochlorite (NaOCl), universally known as bleach, which is carachterized by high microbicidal efficacy, low toxicity to humans and low cost, but suffers the disadvantages of being irritant and corrosive. Nevertheless, ceramics, methylacrylate, or cement are not sensitive to bleach. More specifically, sodium hypochlorite is potentially bactericidal, virucidal, fungicidal, mycobactericidal, sporicidal. Hence it plays an important role in surface disinfection of healthcare facilities and medical equipments.

The concentration of sodium hypochlorite sold for domestic purposes is around 5-6%, with a pH around 11 and it is irritant; while in higher concentration, 10-15%, with a pH around 13, it burns and it is corrosive. According to the Laboratory biosafety manual [30] published by the World Health Organisation (WHO): "A general all-purpose laboratory disinfectant should have a concentration of 1 g/L available chlorine. A stronger solution, containing 5 g/L available chlorine, is recommended for dealing with biohazardous spillage and in the presence of large amounts of organic matter. Sodium hypochlorite solutions, as domestic bleach, contain 50 g/L available chlorine and should therefore be diluted 1:50 or 1:10 to obtain final concentrations of 1 g/L and 5 g/L, respectively. [...] Surfaces can be decontaminated using a solution of sodium hypochlorite (NaOCl); a solution containing 1 g/L available chlorine may be suitable for general environmental sanitation, but stronger solutions (5 g/L) are recommended when dealing with highrisk situations."

Once sodium hypochlorite dissolves in water (equation 1-3) the two compounds that cause disinfection via oxidation are generated, namely hypochlorite ion (OCl $\cdot$ ), a weak

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base, and its corresponding acid, hypochlorous acid (HOCl), whose percentage is determined by water's pH and which is the most active between the two [31,32]. In fact, hypochlorous acid, due to no electronic charge, better penetrates the microorganism cell wall or any protective layer and effectively kills them by oxidating the side chains of proteins' amino acids [33,34].

$$NaOCl + H_2O \leftrightarrows OCl^- + HOCl$$
 (1)

$$HOCl + H^+ + Cl^- \leftrightarrows Cl_2 + H_2O \tag{2}$$

$$2HOCl + OCl^{-} \rightarrow ClO_{3}^{-} + 2Cl^{-} + H^{+}$$
 (3)

It is also common to express concentration of chlorine compounds in terms of available chlorine or free available chlorine (FAC). The term FAC refers to the mixture of oxidizing chlorine forms that have a chlorine atom in the 0 or -1 oxidation state and are not combined with ammonia or organic nitrogen.

Sodium hypochlorite is characterized by high instability, therefore the FAC value is not so significant: 0.75 grams of activated chlorine evaporate per day. This happens not only when sodium hypochlorite gets heated up, but also when gets in touch with acids, sunlight, specific metals,toxic and corrosive gases, included chlorine itself [35,36].

Sodium hypochlorite solution is an inflammable weak base and these characteristics must be considerate during its use and storage. Because of these reasons, formulation and conditions for the application should minimize the formation of by-products and even chloramines [37].

The overall stoichiometry of degradation is shown in the equation 3:

Thus disinfection's efficacy of chlorine releasing agents depends on the water's pH and FAC. Chlorine disinfection against vegetative bacteria, fungi, and yeast, as well as fungal conidia and viruses is preferable at alkaline NaOCl solutions; although the germicidal efficacy is even greater when pH value is around 5.5 and 8 [37,38]. Furthermore, Kuroiwa et al. [39] proved that, adjusting the pH around 5 by weak acidification with acetic acid, resulted in a shortened killing time of all the B. subtilis JCM1465 spores by one third. On the contrary, this preparation killed all of nonspore-forming bacteria within 30 seconds as quickly as NaClO solution without acidification.

The importance of the pH level is showed in Figure 6. At a 7 pH value the concentration of hypochlorous acid is 80%, while when pH value is around 8 the concentration drop to 20%.

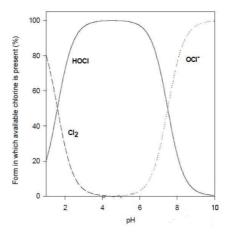


Figure 6. Active chlorine species concentration at different pH values [40].

The pH value of the solution is fundamental either for the bactericidal activity or for the shelf life: at 25–35°C, neutralized-NaOCl solutions (pH 7) expires in a few hours, generated NaOCl (gNaOCl) solutions (produced by electrolysis of a salt (NaCl solution, pH 9) last 6 days, while stabilized NaOCl solutions (pH 9–11) persist more than 30 days [41]. Sodium hypochlorite is widely used, not only as surface antimicrobial but also in water treatment, for water disinfection and for bleaching purposes in textile industry. Furthermore, it can be used to avoid crustaceans and algae formation in cooling towers.

As an alternative, calcium hypoclorite (Ca(OCl)  $_2$ ) also known as HTH (high test hypochlorite) can be used as well. HTH is sold in granular form that, once in solution, achives a pH of 9-11 and it is as stable as NaOCl [41].

Another chlorine releasing agent that has been explored as alternative to sodium, or calcium, hypochlorite is sodium dichloroisocyanurate (NaDCC). This compound is the sodium salt of a chlorinated hydroxytriazine (Figure 7).

Figure 7. Structure of sodium dichloroisocyanurate (NaDCC).

This disinfectant is available as a stable powder which produces solutions that have pH level around 6 and expire within hours [41]. These solutions are more susceptible to inactivation by organic matter than NaOCl [42–44].

NaDCC is often used as a broad spectrum disinfectant since it has been reported to generally achieve similar disinfection activities to chlorine, while results to be less corrosive. On stainless steel Bloomfield *et al.* [45] reported lower ME (microbiocidal effect) values following a 5-minutes exposure to 250 ppm NaDCC compared to NaOCl at the same concentration against *S. aureus* (2.4 vs 4.9 to>6.2 log reduction), *Pseudomonas aeruginosa*(3.7 vs 3.7-4.3 log reduction), and *Enterococcus faecium*(2.2 vs 3.1 log reduction). At 2500 ppm, both NaDCC and NaOCl achieved at least 6 log reduction in each tested organism. Gallandat

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et al. [46] observed similar efficacies of NaOCl, gNaOCl, NaDCC, and HTH (5000 ppm) against both *E. coli* and *Pseudomonas* phage Phi6 after 10-15 minutes on several nonporous surfaces, with minimum 5.9 and 3.1 log reductions, respectively. At higher concentrations, Aarnisalo et al. [47] observed 3.1 and 0.5 log reductions (without/with 2% pork meat) in *Listeria monocytogene* after 30 seconds exposure to 0.04%(w/v) NaDCC and >3.6 and 0.3 log reductions (without/with 2% pork meat) after 30 seconds exposure to 0.2% (w/v) NaOCl. Interestingly, the entry containing hypochlorite as an antibacterial agent and anionactive tensides as cleaning compounds was considered to be much more efficient (3.8 and 2.2 log reductions, without/with 2% pork meat) than the hypochlorite disinfectant, probably due to the inactivation of the NaOCl by the organic matter.

To be effective against bacteria and the spores, an adequate concentration of HOCl is required; in Table 4 are reported the recommended dilutions of each chlorine releasing compound mentioned until now in order to significantly reduce the risk of transmission. The surface conditions, the main advantages and drawbacks have also been considered.

Table 4. Recommended dilutions of commonly used chlorine releasing compounds.

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Chlorine type	Use condition		Advantages	Disadvantages
	Clean condition	Dirty condition		
Sodium hypochlorite solution (5% available chlo-	20 ml/L	100 ml/L	-Can be local (stabilised form)	-Shorter shelf life -Difficult to ship
rine)			-Can be on-side (no stabi- lised form -Does not clog pipes	Low stability (no stabilised form)
High-test hypochlorite (70% available chlorine)	1.4 g/L	7.0 g/L	-Easy to ship -Long shelf life	-Explosive
Sodium dichloroisocyanu- rate powder (60% availa- ble chlorine)	1.7 g/L	8.5 g/L	-Easy to ship -Long shelf life -Does not clog pipes	-Smell
Sodium dichloroisocyanurate tablets (1.5g available		4 tablets per L	-Easy to ship -Long shelf life -Does not clog pipes	-Smell

## 4.1.2 Iodine compounds

Although less reactive than chlorine, iodine solution has a broad spectrum of antimicrobial activity against both gram-negative and gram-positive bacteria, fungi, protozoa, and even bacterial spores [12], while it is not so effective as virucidal [48]. Many investigations identified elemental iodine I2 and hypoiodous acid (HIO) as the two most powerful antimicrobials agents among the several iodine species.

$$I_2 + H_2 0 \leftrightarrows HIO + I^- + H^+$$
 (4)

$$HIO \leftrightarrows IO^- + H^+ \tag{5}$$

$$3HIO + 3OH^- \leftrightarrows IO_3^- + 2I^- + 3H_2O$$
 (6)

The dissociation constant of hypoiodous acid is  $4.5 \times 10^{13}$  and it reveals that the formation of hypoiodite ion (IO-) in aqueous solution is insignificant. The percentages of the species (see equation 4-6) are directly related to pH level of the solution and, to a much lesser extent, to the temperature.

Figure 8 shows  $I_2$  hydrolysis data at different pH values and it is clear that the hightest concentrations of the antimicrobial species are present in the acid range. In fact, when the

solution is alkaline, several iodine species which have no apparent antimicrobial activity can also be generated. Iodate formation could not be a problem if the pH value stays below 8 and the contact time of disinfection is accomplished in the first 30 minutes.

Figure 8. pH-dependant speciation of iodine iodine [49].

Historically iodine solutions or tinctures have been primarily used by health professionals as antiseptics on skin or tissue. Unfortunately aqueous solutions are generally unstable so a combination of iodine and a solubilizing agent or carrier, has been formulated. These combinations, called iodophor, have been used both as antiseptics and disinfectants, retaining the germicidal efficacy of iodine but being more stable and relatively free of toxicity and irritancy [37]. They have been developed to slowly release iodine (L) from the complex, which can be a cationic surfactant, non-ionic, polyoxymer or polyvinylpyrrolidone [50]

The most known and widely used iodophor is povidone-iodine, Figure 9. Regarding this complex Block *et al.* observed 3.14, 3.49, 3.47 and 3.78 log reduction, after 1.5 min for VRE, *E. faecalis* and *S. aureus*, respectively [51].

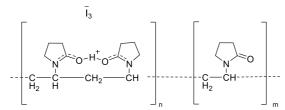


Figure 9. Structure of povidone-iodine complex.

Surfactant iodophor, when used, may add a further detergency activity, even though iodine is chemically less reactive than chlorine. Moreover, surfactant iodophor is less affected by the presence of organic matter than chlorine.

An iodophor, when used at 25 ppm (parts per million of available iodine), is considered to act as a sanitizer, however when the same product is applied at 75 ppm falls into the disinfectant category.

After its releasing, iodine can quickly penetrate the cell wall of a microorganism and oxidize thiol groups leading to disruption of proteins and nucleic acids structures [37].

## 4.2 Alcohols

4.2.1 Alifatic alcohols

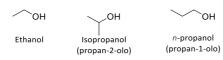


Figure 10. Antimicrobial alcohols.

Among the several aliphatic alcohols that exhibit microbicidal properties ethyl alcohol (ethanol), isopropyl alcohol (isopropanol, propan-2-ol) and *n*-propanol are the most commonly used (Figure 10).

These disinfectants are rapid bactericidal rather than bacteriostatic against vegetative bacteria, included mycobacteria but have no effect on spores. The bactericidal properties of ethanol were examined against several microorganisms for different ranges of time [52]: *P. aeruginosa, Serratia marcescens, E. coli* and *Salmonella typhy* were killed in 10 seconds by all concentrations of ethanol from 40% to 100% (30% for the *E.coli* entry). *S. aureus* and *Streptococcus pyogenes* were slightly more resistant, being killed in 10 seconds with concentrations of 60%–95%. Isopropyl alcohol resulted slightly more bactericidal than ethyl alcohol for *E. coli* and *S. aureus* [53]. Furthermore this category of biocides shows limited fungicidal and virucidal activity specially on lipophilic viruses such as herpes virus, influenza virus, hepatitis B and C viruses [54,55]. Literature data demonstrate that isopropyl alcohol shows its antimicrobial activity against lipid viruses but it is not active against the nonlipid enteroviruses [56]

These alcohols exert their antimicrobial activity by causing proteins denaturation [57,58]. In addition, other modes of action, reported in literature, are the denaturation of dehydrogenases in *E. Coli* and possibly the inhibition of the metabolic processes in *Enterobacter aerogens* [37].

Water plays an important role in the formulation of alcoholic disinfectants because in its absence, proteins are not readily denatured by alcohol. Therefore a 70% solution of alcohol is a much more effective sanitizer than the pure (99%) product [59], but when the concentration drop below 50% there is no practical value [60]. Concentration can be expressed both by weight/weight percentage (%w/w) and, most frequently, by volume/volume percentage (%v/v). This value is important since it is linked to the evaporation rate: higher concentration of alcohol evaporates quickly. The evaporation speed could be an issue if longer contact time is requested, but addition of surfactants [61], or combination with alkali, mineral acids and hydrogen peroxide could overcome this problem [12,24].

Alcohols are fast-acting, easy to use but are not free from limitations that are due to poor detergent properties, toxicity and, of course, their flammability, which is a big concern. The minimum temperature at which vapours above a volatile combustible substance ignite in air when exposed to flame defines the flash point. The higher the concentration, the lower the flash point. For example, the flash points of 70% ethyl and 70% isopropyl alcohol are 20.5°C and 21.0°C, respectively, while the flash point of 30% ethyl alcohol is 29°C [62]. Moreover, even if alcoholic disinfectants are neither corrosive nor staining, they could damage some instruments, by swelling or hardening rubber.

## 4.2.2 Aromatic alcohols

Besides aliphatic alcohols, also aromatic ones exhibit antimicrobial properties being effective in sanitization and disinfection, even in the presence of biological fluids. Phenols are the reference standard for the Rideal–Walker (RW) and Chick–Martin tests for disinfectant evaluation [63].

Phenol ( $C_6H_5OH$ ) is an organic compound that consists of benzene ring bearing a single

f 400 f 401 - 402 r 404

hydroxy substituent. It appears as a white crystalline solid, which is partially water soluble (1 g/15 mL water) [64] and it has a pKa value of 10, that means it is classified as a weak acid.

Phenol exerts its antimicrobial activity against vegetative bacteria, both Gram-positive and negative, fungi and viruses but it is not so effective as sporicidal and against acid-fast bacteria.

The biological activity is related to the undissociated molecule, which induces progressive leakage of essential metabolites, including the release of  $K^*$  [65], leading to membrane damage and consequentially cell lysis, while acting like a protoplasmic poison causing coagulation of the cytoplasm [66].

Phenol is the parent compound but the chemical structure can be modified replacing one of the hydrogen on the aromatic ring with a different functional group (halogen, alkyl, phenyl, benzyl etc.). In Figure 11 are represented several microbicidal phenols.

Figure 11. Several microbiocidal phenols.

4-chloro-2-phenylphenol

The structure activity relationship in the phenol series was investigated by Suter [67]. Regarding the results, it is interesting to notice that the microbiocidal activity increases in derivatives with alkyl chain in *para* position, constituted by a maximum of six carbon atoms, since for longer chain the activity drops probably due to the decrease of water solubility. Nitrophenols were evaluated as well; unfortunately the toxicity increased towards both bacteria and humans and there is also a trend to be inactivated by organic matter. Finally, bisphenolic compounds show activity if they are connected by a methyl linker,

2-benzyl-4-chlorophenol

sulfur or oxygen atom and even if they are directly linked. Augmentation of the efficacy can also be achieved by halogens substitutions.

 $Among all the derivatives, \emph{o-phenylphenol} \ and \ \emph{2-benzyl-4-chlorophenol} \ are \ widely \ used as \ healthcare \ disinfectants.$ 

As disclosed by published reports, commonly used phenolic compounds show, at their use dilution, antimicrobial efficacy against bacteria, fungi, viruses, including HIV [68–71]. However, literature reports also that the phenolic disinfectants 'Stericol' and 'Lysol' show a limited effect on Coxsackie B4, Enterovirus 11 and Poliovirus [72].

Phenols react with certain types of plastic surfaces and they are adsorbed by porous material. If not rinsed thoroughly with water, the alcohol residue can cause skin irritation or depigmentation [73]. Moreover another disadvantage is that phenols are quite expensive, and literature reports demonstrated that they are associated with idiopathic neonatal hyperbilirubinemia in infants [74,75].

## 4.3 Quaternary Ammonium Compounds (QACs)

Quaternary ammonium compounds (QACs) may be considered as amphiphilic substituted compounds, which carry a permanent positive charge nitrogen, counterbalanced by a halide or sulfate moiety. QACs are classified according to the nitrogen substituents, which can include either the type of the carbon chains or the presence of aromatic moieties (Figure 12). The numerous investigations on these chemical structures have increased efficacy while reducing costs.

 $\label{eq:Figure 12.} \textbf{ General structure and common QACs.}$ 

Demand for these disinfectant agents has increased over the decades, furthermore their use is not only limited as germicidal, but they have been widely used also in a variety of industrial, agricultural, clinical applications, and consumer products [76–79].

Their microbicidal activity is due to their adsorption on proteins or acidic phospholipids in the membrane that leads to the formation of hydrophilic voids. The denaturation of essential cell protein causes cytoplasmic membrane permeability and eventually leads to cell disruption [80]. QACs seem also to be involved in the inactivation of energy producing enzyme, furthermore they are able to bind to DNA [81].

Their hydrophobic activity makes them more effective against lipophilic microorganisms. Therefore QACs are solid bactericidal agents, especially against Gram-positive bacteria, and virucidal against enveloped viruses (e.g. herpes simplex, adenovirus, vaccinia) whilst they are not sporicidal and generally not tuberculocidal or virucidal against hydrophilic viruses [82].

QACs are commonly used in ordinary environmental sanitation of noncritical surfaces, such as floors, furnitures, and walls. Scientific literature reports that quaternary ammonium based disinfectants are effective in removing and/or inactivating *S. aureus* and *P. aeruginosa* from computer keyboards, while are not so active against VRE species [83]. Moreover, a recent work by Brown *et al.* [84] demonstrated that the microbial reduction due to QAC's activity on glass continue after contact and wetness time.

However it is important to point up that the efficacy is influenced not only by the compound and surface combinations but even by the product formulation and the water hardness [85]. Indeed, anionic surfactants and high mineral content could lead to insoluble precipitates. Therefore, QAC's formulation is restricted to nonionic or zwitterionic surfactants, which typically are less effective as cleaning ingredients. Furthermore some materials, like cellulose based wipers and gauze pads, absorb these actives, lowering the microbiocidal efficacy [86]. On the other hand, QACs have many advantages like high stability, low colour, odourless and relatively low toxicity (unlike phenols and chlorine bleach). Nevertheless, spraying or fumigation of this chemical disinfectant is not recommended because a few cases disclose occupational asthma as a result of exposure [87–89]. When used, these disinfectant agents are often applied with a cloth or wipe that has been soaked in disinfectant, which may contain mixtures of QACs. Benzalkonium chloride (BAC) is one of the most extensively applied QACs, especially in surface disinfection [90]. BAC's concentration is usually between 0.01 and 1%, but can rise at 15% [91]. Other QACs found in disinfection products have similar concentrations.

# 4.4 Hydrogen Peroxide and Peracids

Figure 13. Structures of biocides peroxigen compounds

Over the years, hydrogen peroxide (H2O2, HP), represented in Figure 13, has extensively been recognized to have antimicrobial properties against a wide variety of microorganisms, such as bacteria, viruses, spores and fungi [92,93]. The mechanism involved in the antibacterial effect of HP ascribes to the release of oxygen free radicals (hydroxyl radical). These radicals are potent oxidising agents that are able to quickly react with bacterial biomolecules, such as thiol groups of proteins, causing irreversible structural modifications and the subsequent cellular death [94]. HP represents one of the most used biocides for different antimicrobial applications, such as disinfection and sterilization, being colourless and odourless and associated with low ecotoxicity. It is a versatile disinfectant, due to the possible employ in several environments including air, water and surfaces. [95]

The most employed formulations of hydrogen peroxide are liquid and gas. Hydrogen peroxide liquid formulations are widely used for sterilization and disinfection processes. Usually, a 6% aqueous solution of hydrogen peroxide is employed for laboratory surfaces cleaning, but its bactericidal and sporicidal efficacy is lower against resistant bacterial spores and protozoan cysts, because of the short exposure time [96]. Hydrogen peroxide solutions are unstable thus suitable stabilizing agents such as benzoic acid are usually added. On the other hand, the production of non-toxic and biodegradable decomposition products (oxygen and water) emerges as an important advantage compared to other disinfectants [97].

Many studies revealed the effectiveness of the vaporized form of HP (HPV) for the surface disinfection [98]. This system inactivates nonenveloped viruses, mycobacteria and some multidrug-resistant microorganisms present in hospital rooms surfaces, reducing the number of contaminated porous and nonporous surfaces to 5-0% [99]. In particular, HPV resulted to be efficient against enteric and respiratory pathogens, including adenovirus type 5, poliovirus Sabin 1, rotavirus SA11, but also *Mycobacterium tuberculosis* and *C. difficile* spores [100]. In addition, HPV is often found in combination with heavy metals like silver ions, which showed an interesting bactericidal activity, resulting to be an useful agent for surface disinfection in hospital settings [93,95]. The hydrogen peroxide solution in nebulization systems was also evaluated for the surface disinfection. It provides a better decrease of the microbial contamination on vertical surfaces compared to horizontal ones. However, the use of aerosol form is limited to the hospital empty spaces, excluding patient rooms, intensive care units and other occupied areas [101].

Peracetic acid (CH<sub>3</sub>COOOH), Figure 13, is an organic pexoxide with activity against mycobacteria, viruses, spores, molds at low concentrations. It results to be a more potent antimicrobial agent than hydrogen peroxide [102,103]. Peracetic acid is a strong oxidizing agent that provides innocuous decompositions by-products: acetic acid and hydrogen peroxide. Generally, it is employed as surface disinfectant and for the medical devices sterilization [104]. A 15% aqueous solution of a mixture of peracetic acid, acetic acid, hydrogen peroxide, and water is commonly commercially available for the application as disinfectant [97].

Figure 13 reports also performic acid (CH<sub>2</sub>O<sub>3</sub>), which is another well-known disinfectant carachterized by virucidal, bactericidal, sporicidal and fungicidal activity, useful in hospital environments and food industry [105]. In a similar way to peracetic acid, performic acid liquid formulation includes formic acid, hydrogen peroxide and water, with production of non-toxic by-products. The main limit of performic acid solution application is due to its instability, which requires the instant preparation before use [106].

## 4.5 Ozone

Ozone (Os) is an inorganic gas, an allotropic form of oxygen, that represents one of the most potent oxidising agent, mainly used for the disinfection of water systems but also for the decontamination of surfaces in healthcare settings and medical industries [107–109]. Ozone effectively inactivates bacteria, viruses, molds and protozoa by: producing hydroxyl free radicals that can react with glycoproteins; disrupting the integrity of cell membrane; oxidizing enzyme's thiol groups thus interfering with their activity; damaging DNA [110]. *P. fluorescens, S. aureus*, enteropathogenic *E. coli, S. typhimurium*, stomatitis virus, encephalomyocarditis virus, *Vibrio cholerae* and *Shigella flexner*i are among the most sensitive microorganisms to the ozone treatment. Moreover, a quicker inactivation is observed when they are suspended in phosphate-buffered saline solutions [111].

Ozone (O<sub>3</sub>) spontaneously decomposes into oxygen (O<sub>2</sub>) and single reactive oxygen atom, associated to the antimicrobial activity. On the other hand, the use of gaseous form for disinfection is not convenient for operator safety, due to the exposure time to high concentrations of the gas [112]. Ozone solutions in water (ozonated water) allow to obtain a liquid form useful for a safe and effective surfaces disinfection, even if its low stability limited the applications [110]. In fact, the acqeous form shows a short half-life at 20°C,

approximately 20–30 minutes, after which it converts into oxygen molecule; while the gaseous form results to have more stability and a longer half-life (12 hours) [113]. The main aspects that affect the ozone stability are temperature, pH and ozone-oxidizable materials. To reduce the decomposition rate of the gas, several ozone generators were designed to produce stabilized form of aqueous  $O_3$  and to extend its half-life up to a few hours [114].

The effectiveness of aqueous and gaseous  $O_3$  against manure-based pathogens (MBP) were assessed for several contaminated surfaces. Aqueous ozone achieved a good reduction of MBP contamination on plastic and metal surfaces after 4 minutes of exposure, but not in more complex surfaces [110]. In a recent study, aqueous ozone demonstrated its efficacy also against several isolates of SARS-CoV-2 after 5 minutes of incubation, resulting a new potential alternative for the disinfection of outdoor surfaces contaminated by this virus [114,115]. Synergistic effects have been shown between ozone and ultraviolet, hydrogen peroxide or negative air ions, in order to increase the production of hydroxyl radicals and to improve the antimicrobial activity [116]. Zoutman et al. evaluated the efficacy of ozone in combination with hydrogen peroxide in vapour form for steel surface disinfection, demonstrating a high level of decontamination in short exposure time against the most common hospital-associated microorganisms [117]. The combination of O3 at low concentration and ultraviolet also demonstrated synergistic effects on E. coli and Escherichia virus MS2 inactivation, highlighting the potential antimicrobial properties of this mixture couple for the development of new disinfectants [118]. The use of ozone generators may be associated to the production of negative air ions (NAI) and nitrogen oxides that displayed bacteriostatic properties and a reduction of microbial populations, alone and in combination with the O<sub>3</sub> [119].

## 4.6 UV

Ultraviolet (UV) is an electromagnetic radiation characterized by a wavelength from 10 to 400 nm, longer than X-rays but shorter than visible light. Three bands of UV light have been identified: UVA (400–315 nm), UVB (315–280 nm) and UVC (280–100 nm). UVC is also called ultraviolet germicidal irradiation (UVGI) for its antimicrobial properties [120]. In fact, since many years UV radiation has been employed for the disinfection and sterilization, mainly the wavelength of 250 nm that has revealed better performance [121]. Nevertheless, different inactivation responses have been observed for several pathogens types including bacteria, viruses, fungi, and spores, even multidrug-resistant (MDR) strains of Acinetobacter baumannii, and C. difficile spores [122]. The efficacy of the decontamination is also related with the UVC amount and exposure time. For example, best inactivation response for bacteria is at 254 nm, while higher wavelengths are required for viruses and protozoa (260–270 nm) [123].

The mechanisms involved in the antimicrobial effects of UV light are based on photochemistry. Microorganism biomolecules, mainly nucleotides, absorb the photon energy emitted by UV light which cause to them chemical modifications and cellular damage through three potential routes: photohydration of DNA, photosplitting (breaking the DNA) or photodimerization [120]. Usually, when thymine bases adjacent to other ones are excited by a UV light, several covalently linked dimers are generated, blocking the DNA replication process. Anyway, UV is not able to kill microorganisms but make they unable to duplicate and induce infections [124].

During the years the use of several UVC light-based devices for the cleaning and disinfection especially in hospital settings is increased because of its associated advantages, among which the absence of residues after treatment, the broad spectrum activity and rapid exposure times [125]. Today, mercury vapor arc lamps and xenon lamps represent the most frequently used UVC devices (100–280 nm). The first one emits a continuous UVC light at low pressure (approximately 254 nm), while xenon lamps generate a pulsed light at high intensity [126]. However, the UV irradiation at 254 nm can cause eyes and skin damages, so the treatment must be performed in unoccupied rooms. Alternatively, 222 nm UVC light could be used, since it is poorly absorbed by the eyes and skin.

Hiroki Kitagawa *et al.* validated the effectiveness of UVC radiation at 222 nm against SARS-CoV-2 contaminations, highlighting the possibility to carried out the disinfection process also in occupied rooms and spaces [127].

New technologies have been reported with the aim to improve the effectiveness of surface decontamination using UVGI. A novel portable UVC devise has been assessed on several surfaces including plastic, bedrail, stainless steel, chrome-plated and porcelain objects. High level of bacterial inactivation has been observed against MRSA on bedrail and against VRE on chrome and stainless steel [128]. Another study has described the efficacy of a new portable pulsed ultraviolet (UV) radiation generator for the surface cleaning, towards the most common nosocomial bacteria, including *P. aeruginosa, A. baumannii, S. aureus and B. cereus.* A potent antibacterial activity has been detected after a short exposure time, revealing as an advantageous new method of sanitation [129,130]. Moreover, the UV technology leads to the development of the UVC reflective wall, aimed to reduce the time of irradiation. The exposure time decreases from 25 to 5 minutes for MRSA and from 43 to 9 minutes for *C. difficile* spores if UVC generator (254 nm) is located in a room coated by a specific reflective agent for UVC light [131].

The different mechanisms of action, the antimicrobial and cellular effects of the described antimicrobial agents are summarized in Table 5 together with the main advantages and disadvantages.

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Table 5. Summary of advantages and disadvantages of common surface disinfectant.

Disinfectant	Mechanism of action	Cellular effect	Antimicrobial ef- fect	Advantages	Disadvantages
Chlorine compounds	Oxidation of side chains aminoacids in proteins	Unfolding tertiary structure and protein aggregation	Bactericidal, fungi- cidal, virucidal sporicidal	-Not flammable -Fast-acting -Low-cost -Resistant to water hardness -Relatively stable	-Salt residues -Corrosive to metals -Affected by organic ma -Fabric discoloration -Potential production of trih thane -Irritating odor at high cond
Iodine com- pounds	Oxidation of thiol groups to disulfides in proteins	Modification of struc- tural protein and/or al- terations in enzyme ac- tivities	Bactericidal, viruci- dal	-Not flammable	-Limited spectrum of acti -Degradation of silicone cat -Staining for surfaces
Alcohols	Denaturation and precipitations of cytoplasmic and membrane proteins	Alteration in metabolic processes, membrane damage	Bactericidal, fungi- cidal, virucidal	-Fast-acting -Noncorrosive -Nonstaining -Suitable for small surfaces disinfection	-Not sporicidal -Affected by organic ma -No cleaning propertie -Deterioration of some instru -Flammable -Rapid evaporation
Phenols	Denaturation of cytoplasmic and membrane proteins	Leakage of essential metabolites, release of K*, membrane dam- age, cytoplasmic coag- ulation	Bactericidal, fungi- cidal, virucidal	-Low costs -Not flammable -Nonstaining	-Rapid absorption by porous als and irritate tissues -Potential depigmentation of -Hyperbilirubinemia in in
Quaternary am- monium com- pounds	Binding to phosphates and fatty acid chains in phos- pholipids of cell membrane and DNA	Depolarization, membrane damage, cytoplasmic coagulation	Bactericidal, fungi- cidal, virucidal (en- veloped viruses)	-Good cleaning agents -Surface compatible -Long antimicrobial activity -Low costs	-Not sporicidal -Affected by water hardr -Asthma after benzalkonium exposure -Affected by organic ma

Disinfectant	Mechanism of action	Cellular effect	Antimicrobial ef- fect	Advantages	Disadvantages
Hydrogen peroxide and peracids	Oxidation of thiol groups to disulfides in proteins	Modification of struc- tural protein and/or al- terations in enzyme ac- tivities	Bactericidal, fungi- cidal, virucidal	-Fast-acting -Safe for workers -Non-toxic by products -Surface compatible -Nonstaining -Odourless -Not flammable	-More expensive compared to other disinfectants -Not sporicidal al low concentrations
Ozone	Oxidation of thiol groups in proteins and interaction with purine and pyrimidine bases	Modification of struc- tural protein, altera- tions in enzyme activi- ties and/or DNA dam- ages	Bactericidal, moldi- cidal, virucidal, protozocidal	-Fast-acting	-Gaseous form <sub>not safe</sub> -Low stability solutions form -Reacted with organic matter
UV light	chemical modifications of nucleotides caused by pho- ton energy emitted	DNA damages (photo- hydration, photosplit- ting, photodimeriza- tion)	bacteria, fungi, vi- ruses, spores	-Absence of residues or by products -Fast-acting	-No microbiocidal effect -Eyes and skyn damages for UV irra- diation at 254-nm

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#### 5. Antimicrobial Surfaces

To date, several strategies have been proposed to prevent microorganisms from adhering to surface or to kill the ones that manage to attach them. Furthermore, minimize the biofilm formation should be a further goal [132]. Nonetheless, it is necessary to take into account that bacterial colonization of surfaces is a key process of corrosion, infection, fermentation and fouling [133].

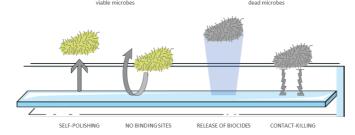


Figure 14. General classes of antimicrobial surfaces.

New strategies to control and hopefully avoid the adhesion of microorganisms on surfaces (Figure 14) are inspired by nature, a source that appears to be almost unlimited, and it has attracted a large amount of interest in the past decades. Indeed a current trend is based on natural materials such as plant leaves and insect cuticles. For example, the leaves of Nelumbo nucifera, commonly known as lotus, exhibit superhydrophobicity and self-cleaning abilities. The characteristics that afford this self-cleaning capability are the lipid's layer that covered the surface. This results in a high water contact angle  $(\theta > 150^\circ)$  and a low tilting angle  $(\theta < 10^\circ)$ , which are parameters needed to lead the water droplet to roll off [134]. In this way the water droplets collect dirt as they move over the leaf . Many other plants exhibit very similar properties to that of the lotus leaf, Indian canna, taro and cabbage leaves.

Similarly, insect surfaces are covered by a layer of lipophilic cuticle. Some insects, e.g. dragonflies or cicada, self-assemble this barrier into three-dimensional nanoarray structures, which enable air to be trapped in and hence exhibit a high water contact angle [135,136]. Furthermore, the turbulent conditions during their flight enhance these self-cleaning properties. Artificial surfaces can be produce to possess similar properties, causing water to behave in a similar way, therefore bacterial cells could be removed before they could adhere to the surface [133,137].

Other interesting approaches use bio-functionalization or surface coatings to give or enhance antibacterial properties: solid heavy metals, such as silver [138,139], copper [140–142] or zinc [143,144], and its alloys have been widely used as antimicrobial agents for millennia due to their intrinsically strong antibacterial activity.

Usually these approaches focus on a nano-size particulate form of the metal: larger surface allows a better contact with the target microbe cells, while enabling more efficient release of the particles. Among these materials, copper is one of the most frequently used due to its efficiency in "contact killing": microorganism survives only a few minutes on these kind of surfaces [145,146]. Obviously, the higher the copper concentration, the faster and more efficient is the antimicrobial activity. Nevertheless, to promote the activity other factors have to be taken into account: both extrinsic, such as protocols and operators, and intrinsic [147].

The major issue with the use of metallic ions is that their interactions are non-specific, which is a major concern from a biocompatibility and cytotoxicity point of view. Furthermore, the leaching components may contaminate and accumulate in the environment,

promoting bacteria's resistance.

Further studies are still required to find the best enhancing parameters like high temperature or high humidity, the metal's physical form, or coating techniques [148].

More recently, another innovative approach based on photosensitizer compounds, hasbeen developed for preventing bacterial colonization. These biocides exert their action after activation by a light source [149]. UVA-induced antimicrobial activity can also be achieved with metals [150,151]; the main mechanisms driving the activity are the formation of highly reactive species like superoxide and hydroxyl radicals and the slow release of metal ions.

The most common techniques that can be applied to incorporate biocides in the surface involve the impregnation of the antimicrobial into the coating. The simultaneous encapsulation of different antimicrobials in one matrix has proven to be more efficient than entrap only one [152]. Layer by layer (LbL) technique is another powerful strategy for surface engineering, which allows to control the leaching characteristics of a biocide [153]. In addition, slow-releasing systems, release-on-command systems and non-leaching systems have also been developed. Commonly employed polymers are polyoxazolines with methyl (PMOZ), ethyl (PEOZ), and propyl (PPOZ) [154], polyacrylamide [155] or poly ethylene glycol-PEG [156]). It has been experimentally proven that antimicrobial properties are also shown by surfactant type polymers and some naturally derived polymers, like chitosan [157]. Different molecules used to chemically modify surface are describe in Figure 15. The building blocks of these polymers can differ from the nature, the molecular weight and the chain length. These are critical parameters that need to be optimized with other factors which may influence the effectiveness of the antimicrobial, like the surface charge density and the hydrophilic/hydrophobic balance.

Figure 15. Chemical structure of some common monomers and polymers used for surface treatment

The physical principle is that polymer brushes act as a steric barrier against bacterial attachment. Indeed some polymers provide an unfavorable surface for bacterial interaction, specially cationic polymers. They have shown effectiveness against bacterial infection but their long term use disclose toxicity as a concern. Their mechanism totally relies on their charge that attract and "capture" negatively charged bacterial cells, and this interaction damages the bacterial membrane, giving a bacteriostatic, and eventually a bactericidal effect.

To improve the antimicrobial efficacy several agents, such as small compounds, peptides and enzymes, can be introduced into polymer molecules [158]. Probably, polymers of QACs represent the class that has received more attention over the years [159,160].

Ideally, a coating of antimicrobial polymer must exhibit a broad antimicrobial spectrum in brief contact's time and it must remain effective over the lifespan's article while avoiding leaching into the environment or decomposition in toxic products. Furthermore, it shouldn't be toxic nor irritating to those who are handling it and not water soluble (for water disinfection application) [161].

Figure 16 summarize all the approaches that involve changes of the chemical and/or physical properties of the surface in order to have a biocide effect.

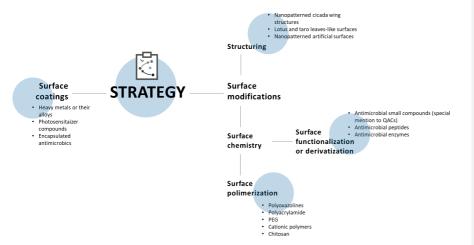


Figure 16. Different approaches in the design of antimicrobial surfaces.

## 6. Current And Future Issues

Antimicrobials are a precious resource that effectively keep harmful microorganisms at bay. Unfortunately, nowadays, biocidal products are perceived as either direct and indirect threats. The direct one is due to the dissemination of resistant strains: the concept of bacterial resistance to biocides is not novel and the first evidence has been reported in the early 1950s [162]. This phenomenon has been associated with the increasing exposure to biocides; furthermore several investigations describe a possible linkage between antimicrobial agents and the occurrence of antibiotic cross- and co- resistance [163,164]. The indirect threat regards the transfer of genes which confers resistance to a suscepbtible strain, enhancing its resistance level. For example, the extensive use of quaternary ammonium compounds has been blamed for the spread of QAC-resistance bacteria, both Gram-positive and negative. Resistance's mechanisms to this class of compounds is underexplored, however efflux pump and alteration of membrane composition are among the predominant ones [165,166].

Another example of antimicrobial resistance can be found in the tolerance to oxidizing biocides, like chlorine, hydrogen peroxide and paracetic acid, which has also been described [167]. Resistance to these agents can result from the overproduction of enzymes which increases the defense towards radical-mediated damage or protects from biofilm's alterations.

The selective pressure towards disinfectants may occur also when biocides are discharged into the environment, themselves or their residues [168,169]. McBain *et al.* [170] investigate the effects of triclosan use on the domestic-drain biofilm ecosystems. They found out that the biocide did not significantly lower the total counts but altered the bacterial composition, due to innate resistance or insusceptibility of some species able to degrade triclosan. Hospital wastewaters have been investigated as well [171,172], since they are characterized by high concentration of antibiotics and disinfectants.

However, the lack of data on the majority of antimicrobial compounds prevents to clearly identify the risk arising from the increase and indiscriminate use of these biocides.

In conclusions, the consciousness that the perfect antimicrobial agent may not yet exist the right choice and the appropriate use of the current chemicals are necessary to avoid both resistance and environmental issue. For this purpose a deep knowledge of the antimicrobial agent together with the type of surface would result in an effective and suitable disinfection level.

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#### References

- Kramer, A.; Schwebke, I.; Kampf, G. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. *BMC Infect. Dis* 2006, 6, 130
- Hayden, M.K.; Blom, D.W.; Lyle, E.A.; Moore, C.G.; Weinstein, R.A. Risk of Hand or Glove Contamination After Contact With Patients Colonized With Vancomycin-Resistant Enterococcus or the Colonized Patients' Environment. Infect. Control Hosp. Epidemiol. 2008, 29, 149–154, doi:10.1086/524331.
- 3. Duckro, A.N.; Blom, D.W.; Lyle, E.A.; Weinstein, R.A.; Hayden, M.K. Transfer of vancomycin-resistant enterococci via health care worker hands. *Arch. Intern. Med.* 2005, 165, 302–307, doi:10.1001/archinte.165.3.302.
- Guerrero, D.M.; Nerandzic, M.M.; Jury, L.A.; Jinno, S.; Chang, S.; Donskey, C.J. Acquisition of spores on gloved hands after contact with the skin of patients with Clostridium difficile infection and with environmental surfaces in their rooms. Am. J. Infect. Control 2012, 40, 556–558, doi:10.1016/j.ajic.2011.08.002.
- Stiefel, U.; Cadnum, J.L.; Eckstein, B.C.; Guerrero, D.M.; Tima, M.A.; Donskey, C.J. Contamination of Hands with Methicillin-Resistant Staphylococcus aureus after Contact with Environmental Surfaces and after Contact with the Skin of Colonized Patients Infect. Control Hosp. Evidemiol. 2011. 32. 185–187. doi:10.1086/657944.
- Janeway, C.A. Jr; Travers, P.; Walport, M. Immunobiology: The Immune System in Health and Disease. 5th edition. New York: Garland Science; 2001. Infectious agents and how they cause disease, 27114.
- 7. Silhavy, T.J.; Kahne, D.; Walker, S. The bacterial cell envelope. Cold Spring Harb. Perspect. Biol. 2010, 2.
- 8. Auer, G.K.; Weibel, D.B. Bacterial Cell Mechanics. Biochemistry 2017, 56, 3710–3724, doi:10.1021/acs.biochem.7b00346.
- 9. Esko, J.D.; Doering, T.L.; Raetz, C.R. Eubacteria and Archaea. In Essentials of Glycobiology, 2nd ed.; Gerald, W. H., Marilynn, E. E., Cold Spring Harbor Laboratory Press, 2009, Chapter 20.
- Brown, S.; Santa Maria, J.P.; Walker, S. Wall teichoic acids of gram-positive bacteria. Annu. Rev. Microbiol. 2013, 67, 313–336, doi:10.1146/annurev-micro-092412-155620.
- 11. Dufrêne, Y.F.; Persat, A. Mechanomicrobiology: how bacteria sense and respond to forces. Nat. Rev. Microbiol. 2020, 18, 227–240.
- 12. Nicholson, W.L.; Fajardo-Cavazos, P.; Rebeil, R.; Slieman, T.A.; Riesenman, P.J.; Law, J.F.; Xue, Y. Bacterial endospores and their significance in stress resistance. *Antonie Van Leeuwenhoek* 2002; 81, 27-32.
- Mallozzi, M.; Viswanathan, V.K.; Vedantam, G. Spore-forming Bacilli and Clostridia in human disease. Future Microbiol. 2010, 5, 1109–1123.
- Chen, Y.; Norde, W.; van der Mei, H.C.; Busscher, H.J. Bacterial cell surface deformation under external loading. MBio 2012, 3, doi:10.1128/mBio.00378-12.
- 15. Donlan, R.M. Biofilms: Microbial life on surfaces. Emerg. Infect. Dis. 2002, 8, 881–890.
- Sharma, D.; Misba, L.; Khan, A.U. Antibiotics versus biofilm: An emerging battleground in microbial communities. Antimicrob. Resist. Infect. Control 2019, 8, 1–10.
- 17. Bjarnsholt, T. The role of bacterial biofilms in chronic infections. APMIS. Suppl. 2013, 1–51, doi:10.1111/apm.12099.
- Høiby, N. A short history of microbial biofilms and biofilm infections. APMIS 2017, 125, 272–275.
- Gelderblom, H.R. Structure and Classification of Viruses in Medical Microbiology. 4th ed. Galveston (TX): University of Texas Medical Branch at Galveston; 1996. Chapter 41.
- 20. Summers, W.C. Virus Infection. In Encyclopedia of Microbiology; Elsevier Inc., 2009; 546–552.
- Lucas, W. Viral Capsids and Envelopes: Structure and Function. In Encyclopedia of Life Sciences; John Wiley & Sons, Ltd: Chichester, UK, 2010.
- 22. Louten, J. Virus Structure and Classification. In Essential Human Virology; Elsevier, 2016; 19–29.
- 23. Ruiz-Herrera, J.; Ortiz-Castellanos, L. Cell wall glucans of fungi. A review. Cell Surf. 2019, 5.
- Taylor, T.N.; Osborn, J.M. The importance of fungi in shaping the paleoecosystem. Rev. Palaeobot. Palynol. 1996, 90, 249–262, doi:10.1016/0034-6667/9500086-0.
- Classification of Biological Agents Health and Safety Authority Available online: https://www.hsa.ie/eng/publications\_and\_forms/publications/biological\_agents/biological\_agents\_code\_of\_practice\_2020.html (accessed on Apr 20, 2021).
- Hayden, M.K.; Bonten, M.J.M.; Blom, D.W.; Lyle, E.A.; van de Vijver, D.A.M.C.; Weinstein, R.A. Reduction in Acquisition of Vancomycin-Resistant Enterococcus after Enforcement of Routine Environmental Cleaning Measures. Clin. Infect. Dis. 2006, 42, 1552–1560, doi:10.1086/503845.
- Dettenkofer, M.; Block, C. Hospital disinfection: Efficacy and safety issues. Curr. Opin. Infect. Dis. 2005, 18, 320–325.
- 28. Stewart, P.S.; Costerton, J.W. Antibiotic resistance of bacteria in biofilms. Lancet 2001, 358, 135–138.
- Johnson, A.C.; Jin, X.; Nakada, N.; Sumpter, J.P. Learning from the past and considering the future of chemicals in the environment. Sci. 2020, 367, 384-387.
- 30. World Health Organization Laboratory biosafety manual Third edition; 2004.

 Albrich, J.M.; McCarthy, C.A.; Hurst, J.K. Biological reactivity of hypochlorous acid: Implications for microbicidal mechanisms of leukocyte myeloperoxidase. Proc. Natl. Acad. Sci. U. S. A. 1981, 78, 210–214, doi:10.1073/pnas.78.1.210.

26 of 30

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- Brazis, A.R.; Leslie, J.E.; Kabler, P.W.; Woodward, R.L. The inactivation of spores of Bacillus globigii and Bacillus anthracis by free available chlorine. Appl. Microbiol. 1958, 6, 338–342, doi:10.1128/aem.6.5.338-342.1958.
- 33. Hawkins, C.L.; Pattison, D.I.; Davies, M.J. Hypochlorite-induced oxidation of amino acids, peptides and proteins. *Amino Acids* 2003, 25, 259–274.
- 34. Winter, J.; Ilbert, M.; Graf, P.C.F.; Özcelik, D.; Jakob, U. Bleach Activates a Redox-Regulated Chaperone by Oxidative Protein Unfolding. Cell 2008, 135, 691–701, doi:10.1016/j.cell.2008.09.024.
- Adam, L.C.; Suzuki, K.; Gordon, G.; Fábián, I. Hypochlorous Acid Decomposition in the pH 5-8 Region. *Inorg. Chem.* 1992, 31, 3534–3541, doi:10.1021/ic00043a011.
- 36. White, G. C., Handbook of Chlorination and Alternative Disinfectants , John Wiley & Sons Ltd 1999; Vol. 77;
- Heseltine, P. Disinfection, Sterilization, and Preservation, 5th ed. SS Block, ed.; Philadelphia: Lippincott Williams & Wilkins, 2001; 1,504 pages. Infect. Control Hosp. Epidemiol. 2002, 23, 109–109, doi:10.1017/s0195941700084289.
- 38. Fukuzaki, S. Mechanisms of Actions of Sodium Hypochlorite in Cleaning and Disinfection Processes. *Biocontrol Sci.* **2006**, *11*, 147–157, doi:10.4265/bio.11.147.
- Kuroiwa, K.; Nakayama, H.; Kuwahara, T.; Tamagawa, K.; Hattori, K.; Murakami, K.; Korai, H.; Ohnishi, Y. Augmenting effect
  of acetic acid for acidification on bactericidal activity of hypochlorite solution. Lett. Appl. Microbiol. 2003, 36, 46–49,
  doi:10.1046/j.1472-765X.2003.01261.x.
- ECHA Regulation (EU) No 528/2012 concerning the making available on the market and use of biocidal products Evaluation of active substances Assessment Report Cholecalciferol PT 14 (Rodenticides); 2018;
- Iqbal, Q.; Lubeck-Schricker, M.; Wells, E.; Wolfe, M.K.; Lantagne, D. Shelf-Life of Chlorine Solutions Recommended in Ebola Virus Disease Response. PLoS One 2016, 11, e0156136, doi:10.1371/journal.pone.0156136.
- 42. Coates, D. A comparison of sodium hypochlorite and sodium dichloroisocyanurate products. J. Hosp. Infect. 1985, 6, 31–40, doi:10.1016/S0195-6701(85)80015-3.
- 43. Bloomfield, S.F.; Uso, E.E. The antibacterial properties of sodium hypochlorite and sodium dichloroisocyanurate as hospital disinfectants. *J. Hosp. Infect.* **1985**, *6*, 20–30, doi:10.1016/S0195-6701(85)80014-1.
- Coates, D. Comparison of sodium hypochlorite and sodium dichloroisocyanurate disinfectants: neutralization by serum. J. Hosp. Infect. 1988, 11, 60–67, doi:10.1016/0195-6701(88)90040-0.
- Bloomfield, S.F.; Arthur, M.; Begun, K.; Patel, H. Comparative testing of disinfectants using proposed European surface test methods. Lett. Appl. Microbiol. 1993, 17, 119–125, doi:10.1111/j.1472-765X.1993.tb01439.x.
- Gallandat, K.; Wolfe, M.K.; Lantagne, D. Surface Cleaning and Disinfection: Efficacy Assessment of Four Chlorine Types Using Escherichia coli and the Ebola Surrogate Phi6. Environ. Sci. Technol. 2017, 51, 4624–4631, doi:10.1021/acs.est.6b06014.
- Aarnisalo, K.; Salo, S.; Miettinen, H.; Suihko, M.L.; Wirtanen, G.; Autio, T.; Lundén, J.; Korkeala, H.; Sjöberg, A.M. Bactericidal efficiencies of commercial disinfectants against listeria monocytogenes on surfaces. J. Food Saf. 2000, 20, 237–250, doi:10.1111/j.1745-4565.2000.tb00302.x.
- Mbithi, J.N.; Springthorpe, V.S.; Sattar, S.A. Chemical Disinfection of Hepatitis A Virus on Environmental Surfaces. Appl. Environ. Microbiol. 1990, 56, 3601-3604;
- 49. Bichsel, Y. Behavior of iodine species in oxidative processing during drinking water treatment, Dipl. Chem. ETH, Swiss Federal Institute Of Technology Zurich, 2000.
- Al-Adham, I.; Haddadin, R.; Collier, P. Types of Microbicidal and Microbistatic Agents. In Russell, Hugo & Ayliffe's: Principles and Practice of Disinfection, Preservation and Sterilization; John Wiley & Sons, 2012; pp. 5–70.
- Block, C.; Robenshtok, E.; Simhon, A.; Shapiro, M. Evaluation of chlorhexidine and povidone iodine activity against methicillinresistant Staphylococcus aureus and vancomycin-resistant Enterococcus faecalis using a surface test. J. Hosp. Infect. 2000, 46, 147–152, doi:10.1053/jhin.2000.0805.
- 52. Morton, H.E. The relationship of concentration and germicidal efficiency of ethyl alcohol. *Ann. N. Y. Acad. Sci.* 1950, 53, 191–196, doi:10.1111/j.1749-6632.1950.tb31944.x.
- 53. Coulthard CE, S.G. The germicidal effect of alcohol with special reference to its action on bacterial spores. Pharm. J. 1936, 79–81.
- 54. Rutala, W.A.; Weber, D.J. Guideline for Disinfection and Sterilization in Healthcare Facilities, 2008.
- Doerrbecker, J.; Friesland, M.; Ciesek, S.; Erichsen, T.J.; Mateu-Gelabert, P.; Steinmann, J.; Pietschmann, T.; Steinmann, E. Inactivation and survival of hepatitis C virus on inanimate surfaces. J. Infect. Dis. 2011, 204, 1830–1838, doi:10.1093/infdis/jir535.
- Chemical Disinfectants | Disinfection & Sterilization Guidelines | Guidelines Library | Infection Control | CDC Available
  online: https://www.cdc.gov/infectioncontrol/guidelines/disinfection/disinfection-methods/chemical.html# (accessed on Jan 8,
  2021).
- Gilbert, P.; McBain, A.J. Potential impact of increased use of biocides in consumer products on prevalence of antibiotic resistance. Clin. Microbiol. Rev. 2003, 16, 189–208.
- 58. Fraise, A.P. Choosing disinfectants. J. Hosp. Infect. 1999, 43, 255–264.
- 59. Harrington, C.; Walker, H. The Germicidal Action of Alcohol. *Bost. Med. Surg. J.* **1903**, *148*, 548–552, doi:10.1056/nejm190305211482102.
- Salvage, R.; Hull, C.M.; Kelly, D.E.; Kelly, S.L. Use of 70% alcohol for the routine removal of microbial hard surface bioburden in life science cleanrooms. Future Microbiol. 2014, 9, 1123–1130, doi:10.2217/FMB.14.73.

27 of 30

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61. Rutala, W.A.; Weber, D.J. Monitoring and improving the effectiveness of surface cleaning and disinfection. *Am. J. Infect. Control* 2016, 44, e69–e76, doi:10.1016/j.ajic.2015.10.039.

- 62. Rotter, M.L. Arguments for alcoholic hand disinfection. J. Hosp. Infect. 2001, 48, S4-S8, doi:10.1016/S0195-6701(01)90004-0.
- 63. Withell, E.R. The evaluation of bactericides. J. Hyg. (Lond). 1942, 42, 339–353, doi:10.1017/S0022172400035555.
- 64. O'Neil, M. The Merck index: an encyclopedia of chemicals, drugs, and biologicals; 13th ed.; Merck: Whitehouse Station N.J., 2001;.
- Lambert, P.A.; Hammond, S.M. Potassium fluxes, first indications of membrane damage in micro-organisms. *Biochem. Biophys. Res. Commun.* 1973, 54, 796–799, doi:10.1016/0006-291X(73)91494-0.
- Judis, J. Mechanism of action of phenolic disinfectants IV. Effects on induction of and accessibility of substrate to β-galactosidase in Escherichia coli. J. Pharm. Sci. 1965, 54, 417–420, doi:10.1002/jps.2600540315.
- Suter, C.M. Relationships between the structure and the bactericidal properties of phenols. Chem. Rev. 1941, 28, 269–299, doi:10.1021/cr60090a004.
- Terleckyj, B.; Axler, D.A. Quantitative neutralization assay of fungicidal activity of disinfectants. Antimicrob. Agents Chemother. 1987. 31, 794–798. doi:10.1128/AAC.31.5.794.
- Martin, L.S.; McDougal, J.S.; Loskoski, S.L. Disinfection and Inactivation of the Human T Lymphotropic Virus Type III/Lymphadenopathy-Associated Virus. J. Infect. Dis. 1985, 152, 400–403, doi:10.1093/infdis/152.2.400.
- 70. Sattar, S.A.; Springthorpe, V.S. Survival and disinfectant inactivation of the human immunodeficiency virus: A critical review.
- Rev. Infect. Dis. 1991, 13, 430–447.
  Sagripanti, J.L.; Eklund, C.A.; Trost, P.A.; Jinneman, K.C.; Abeyta, C.; Kaysner, C.A.; Hill, W.E. Comparative sensitivity of 13 species of pathogenic bacteria to seven chemical germicides. Am. J. Infect. Control 1997, 25, 335–339, doi:10.1016/S0196-
- 6553(97)90026-2.
  72. Narang, H.K.; Codd, A.A. Action of commonly used disinfectants against enteroviruses. *J. Hosp. Infect.* 1983, 4, 209–212, doi:10.1016/0195-6701(83)90052-X.
- Rutala, W.A.; Weber, D.J. Disinfection and Sterilization in Health Care Facilities: An Overview and Current Issues. *Infect. Dis. Clin.*, 30, 609-637 doi:10.1016/j.idc.2016.04.002.
- 74. Doan, H.M.; Keith, L.; Shennan, A.T. Phenol and Neonatal Jaundice. Pediatrics 1979, 64, 324-325
- 75. Calafat, A.M.; Weuve, J.; Ye, X.; Jia, L.T.; Hu, H.; Ringer, S.; Huttner, K.; Hauser, R. Exposure to bisphenol A and other phenols in neonatal intensive care unit premature infants. *Environ. Health Perspect.* 2009, 117, 639–644, doi:10.1289/ehp.0800265.
- Zhang, C.; Cui, F.; Zeng, G. ming; Jiang, M.; Yang, Z. zhu; Yu, Z. gang; Zhu, M. ying; Shen, L. qing Quaternary ammonium compounds (OACs): A review on occurrence, fate and toxicity in the environment. Sci. Total Environ. 2015, 518, 352–362.
- 77. Ying, G.G. Fate, behavior and effects of surfactants and their degradation products in the environment. *Environ. Int.* **2006**, 32, 417–431.
- Tezel, U.; Pavlostathis, S.G. Role of Quaternary Ammonium Compounds on Antimicrobial Resistance in the Environment. In Antimicrobial Resistance in the Environment; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2011; 349–387.
- Fumagalli, L.; Regazzoni, L.G.; Straniero, V.; Valoti, E.; Aldini, G.; Vistoli, G.; Carini, M.; Picozzi, C. Stressed degradation studies of domiphen bromide by LC-ESI-MS/MS identify a novel promising antimicrobial agent. J. Pharm. Biomed. Anal. 2018, 159, 224– 228, doi:10.1016/j.jpba.2018.06.055.
- Wessels, S.; Ingmer, H. Modes of action of three disinfectant active substances: A review. Regul. Toxicol. Pharmacol. 2013, 67, 456–467, doi:10.1016/j.yrtph.2013.09.006.
- Zinchenko, A.A.; Sergeyev, V.G.; Yamabe, K.; Murata, S.; Yoshikawa, K. DNA compaction by divalent cations: Structural specificity revealed by the potentiality of designed quaternary diammonium salts. *ChemBioChem* 2004, 5, 360–368, doi:10.1002/cbic.200300797.
- 82. Gerba, C.P. Quaternary ammonium biocides: Efficacy in application. Appl. Environ. Microbiol. 2015, 81, 464–469.
- Rutala, W.A.; White, M.S.; Gergen, M.F.; Weber, D.J. Bacterial Contamination of Keyboards: Efficacy and Functional Impact of Disinfectants. Infect. Control Hosp. Epidemiol. 2006, 27, 372–377, doi:10.1086/503340.
- 84. Brown, E.; Dhanireddy, K.; Teska, P.; Eifert, J.; Williams, R.C.; Boyer, R. Influence of drying time on prewetted disinfectant towelettes to disinfect glass surfaces. *Am. J. Infect. Control* **2020**, *48*, 846–848, doi:10.1016/j.ajic.2019.11.006.
- 85. Cousins, C.M.; Clegg, L.F.L. The effect of water hardness and temperature on water sterilization by mixtures of detergents and quaternary ammonium compounds. *J. Appl. Bacteriol.* **1956**, *19*, 250–255, doi:10.1111/j.1365-2672.1956.tb00075.x.
- Song, X.; Vossebein, L.; Zille, A. Efficacy of disinfectant-impregnated wipes used for surface disinfection in hospitals: A review. Antimicrob. Resist. Infect. Control 2019, 8, 1-14.
- 87. Purohit, A.; Kopferschmitt-Kubler, M.C.; Moreau, C.; Popin, E.; Blaumeiser, M.; Pauli, G. Quaternary ammonium compounds and occupational asthma. *Int. Arch. Occup. Environ. Health* **2000**, 73, 423–427, doi:10.1007/s004200000162.
- Schyllert, C.; Rönmark, E.; Andersson, M.; Hedlund, U.; Lundbäck, B.; Hedman, L.; Lindberg, A. Occupational exposure to chemicals drives the increased risk of asthma and rhinitis observed for exposure to vapours, gas, dust and fumes: A crosssectional population-based study. Occup. Environ. Med. 2016, 73, 663–669, doi:10.1136/oemed-2016-103595.
- 89. Zock, J.P.; Plana, E.; Jarvis, D.; Antó, J.M.; Kromhout, H.; Kennedy, S.M.; Künzli, N.; Villani, S.; Olivieri, M.; Torén, K.; et al. The use of household cleaning sprays and adult asthma: An international longitudinal study. *Am. J. Respir. Crit. Care Med.* 2007, 176, 735–741, doi:10.1164/rccm.200612-1793OC.
- 90. Kampf, G.; Degenhardt, S.; Lackner, S.; Jesse, K.; von Baum, H.; Ostermeyer, C. Poorly processed reusable surface disinfection tissue dispensers may be a source of infection. *BMC Infect. Dis.* **2014**, *14*, 1-8 doi:10.1186/1471-2334-14-37.

Commentato [S1]: Spazio in piu

 Pereira, B.M.P.; Tagkopoulos, I. Benzalkonium chlorides: Uses, regulatory status, and microbial resistance. Appl. Environ. Microbiol. 2019, 85.

- Baldry, M.G.C. The bactericidal, fungicidal and sporicidal properties of hydrogen peroxide and peracetic acid. J. Appl. Bacteriol. 1983, 54, 417–423, doi:10.1111/j.1365-2672.1983.tb02637.x.
- Absalan, A.; Ehrampoush, M.; Davoudi, M.; Vakili, T.; Ebrahimi, A. Antibacterial effects of hydrogen peroxide and silver composition on selected pathogenic enterobacteriaceae. *Int. J. Environ. Health Eng.* 2012, 1, 23, doi:10.4103/2277-9183.96148.
- Steinberg, D.; Heling, I.; Daniel, I.; Ginsburg, I. Antibacterial synergistic effect of chlorhexidine and hydrogen peroxide against Streptococcus sobrinus, Streptococcus faecalis and Staphylococcus aureus. J. Oral Rehabil. 1999, 26, 151–156, doi:10.1046/j.1365-2842 1999 00343 x
- 95. Martin, N.L.; Bass, P.; Liss, S.N. Antibacterial properties and mechanism of activity of a novel silver-stabilized hydrogen peroxide. *PLoS One* **2015**, *10*, 1–20, doi:10.1371/journal.pone.0131345.
- 96. McDonnell, G. The Use of Hydrogen Peroxide for Disinfection and Sterilization Applications; Mcdonnell, G.. "The Use of Hydrogen Peroxide for Disinfection and Sterilization Applications." in Patai's Chemistry of Functional Groups, 2014, 1-34.
- 97. Fraise, A.P.; Lambert, P.A.; Maillard, J.Y. Russell, Hugo and Ayliffe's Principles and Practice of Disinfection, Preservation and Sterilization: Fourth Edition; John Wiley & Sons. 2008.
- 98. Totaro, M.; Casini, B.; Profeti, S.; Tuvo, B.; Privitera, G.; Baggiani, A. Role of hydrogen peroxide vapor (HPV) for the disinfection
- of hospital surfaces contaminated by multiresistant bacteria. *Pathogens* **2020**, *9*, 408 doi:10.3390/pathogens9050408.

  99. Weber, D.J.; Kanamori, H.; Rutala, W.A. "No touch" technologies for environmental decontamination: Focus on ultraviolet
- devices and hydrogen peroxide systems. *Curr. Opin. Infect. Dis.* **2016**, *29*, 424–431, doi:10.1097/QCO.00000000000000284.

  100. Tuladhar, E.; Terpstra, P.; Koopmans, M.; Duizer, E. Virucidal efficacy of hydrogen peroxide vapour disinfection. *J. Hosp. Infect.* **2012**, *80*, 110–115, doi:10.1016/j.jhin.2011.10.012.
- 101. Orlando, P.; Cristina, M.L.; Dallera, M.; Ottria, G.; Vitale, A.; Badolati, G. Surface disinfection: Evaluation of the efficacy of a nebulization system spraying hydrogen peroxide. J. Prev. Med. Hyg. 2008, 49, 116–119, doi:10.15167/2421-4248/jpmh2008.49.3.127.
- Domínguez Henao, L.; Turolla, A.; Antonelli, M. Disinfection by-products formation and ecotoxicological effects of effluents treated with peracetic acid: A review. Chemosphere 2018, 213, 25–40, doi:10.1016/j.chemosphere.2018.09.005.
- 103. Cutts, T.; Kasloff, S.; Safronetz, D.; Krishnan, J. Decontamination of common healthcare facility surfaces contaminated with SARS-CoV-2 using peracetic acid dry fogging. *J. Hosp. Infect.* **2021**, *109*, 82–87, doi:10.1016/j.jhin.2020.12.016.
- Mcdonnell, G.; Russell, A.D. Antiseptics and disinfectants: Activity, action, and resistance. Clin. Microbiol. Rev. 1999, 12, 147–179, doi:10.1128/cmr.12.1.147.
- Rutala, W.A.; Weber, D.J. New disinfection and sterilization methods. *Emerg. Infect. Dis.* 2001, 7, 348–353, doi:10.3201/eid0702.010241.
- 106. Gehr, R.; Chen, D.; Moreau, M. Performic acid (PFA): Tests on an advanced primary effluent show promising disinfection performance. *Water Sci. Technol.* **2009**, *59*, 89–96, doi:10.2166/wst.2009.761.
- 107. Rickloff, J.R. An evaluation of the sporicidal activity of ozone. Appl. Environ. Microbiol. 1987, 53, 683–686, doi:10.1128/aem.53.4.683-686.1987.
- 108. Martinelli, M.; Giovannangeli, F.; Rotunno, S.; Trombetta, C.M.; Montomoli, E. Water and air ozone treatment as an alternative sanitizing technology. J. Prev. Med. Hyg. 2017, 58, E48–E52, doi:10.15167/2421-4248/jpmh2017.58.1.757.
- Breidablik, H.J.; Lysebo, D.E.; Johannessen, L.; Skare; Andersen, J.R.; Kleiven, O.T. Ozonized water as an alternative to alcohol-based hand disinfection. J. Hosp. Infect. 2019, 102, 419–424, doi:10.1016/j.jhin.2019.01.026.
- Megahed, A.; Aldridge, B.; Lowe, J. The microbial killing capacity of aqueous and gaseous ozone on different surfaces contaminated with dairy cattle manure. PLoS One 2018, 13, 1–22, doi:10.1371/journal.pone.0196555.
- Burleson, G.R.; Murray, T.M.; Pollard, M. Inactivation of Viruses and Bacteria by Ozone, With and Without Sonication. Appl. Microbiol. 1975, 29, 340–344, doi:10.1128/am.29.3.340-344.1975.
- 112. Albert, S.; Amarilla, A.A.; Trollope, B.; Sng, J.D.J.; Setoh, Y.X.; Deering, N.; Modhiran, N.; Weng, S.H.; Melo, M.C.; Hutley, N.; et al. Assessing the potential of unmanned aerial vehicle spraying of aqueous ozone as an outdoor disinfectant for SARS-CoV-2. *Environ. Res.* 2021, 196, 110944, doi:10.1016/j.envres.2021.110944.
- 113. Megahed, A.; Aldridge, B.; Lowe, J. Comparative study on the efficacy of sodium hypochlorite, aqueous ozone, and peracetic acid in the elimination of Salmonella from cattle manure contaminated various surfaces supported by Bayesian analysis. *PLoS One* 2019, 14, 1–15, doi:10.1371/journal.pone.0217428.
- 114. Albert, S.; Amarilla, A.A.; Trollope, B.; Sng, J.D.J.; Setoh, Y.X.; Deering, N.; Modhiran, N.; Weng, S.H.; Melo, M.C.; Hutley, N.; et al. Assessing the potential of unmanned aerial vehicle spraying of aqueous ozone as an outdoor disinfectant for SARS-CoV-2. *Environ. Res.* 2021, 196, 110944, doi:10.1016/j.envres.2021.110944.
- 115. Grignani, E.; Mansi, A.; Cabella, R.; Castellano, P.; Tirabasso, A.; Sisto, R.; Spagnoli, M.; Fabrizi, G.; Frigerio, F.; Tranfo, G. Safe and Effective Use of Ozone as Air and Surface Disinfectant in the Conjuncture of Covid-19. *Gases* 2020, 1, 19–32, doi:10.3390/gases1010002.
- Selma, M. V.; Allende, A.; López-Gálvez, F.; Conesa, M.A.; Gil, M.I. Disinfection potential of ozone, ultraviolet-C and their combination in wash water for the fresh-cut vegetable industry. Food Microbiol. 2008, 25, 809–814, doi:10.1016/j.fm.2008.04.005.
- 117. Zoutman, D.; Shannon, M.; Mandel, A. Effectiveness of a novel ozone-based system for the rapid high-level disinfection of health care spaces and surfaces. *Am. J. Infect. Control* **2011**, *39*, 873–879, doi:10.1016/j.ajic.2011.01.012.

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118. Fang, J.; Liu, H.; Shang, C.; Zeng, M.; Ni, M.; Liu, W. E. coli and bacteriophage MS2 disinfection by UV, ozone and the combined UV and ozone processes. Front. Environ. Sci. Eng. 2014, 8, 547–552, doi:10.1007/s11783-013-0620-2.

- 119. Fan, L.; Song, J.; Hildebrand, P.D.; Forney, C.F. Interaction of ozone and negative air ions to control micro-organisms. J. Appl. Microbiol. 2002, 93, 144–148, doi:10.1046/j.1365-2672.2002.01683.x.
- Gray, N.F. Ultraviolet Disinfection in Microbiology of Waterborne Diseases. Second Edi.; Academic Press, 2014. 617-630.; Elsevier, 2013
- 121. Bolton, J.R.; Dussert, B.; Bukhari, Z.; Hargy, T.; Clancy, J.L. Inactivation of Cryptosporidium parvum by medium-pressure ultraviolet light in finished drinking water. *Proc. Am. Water Work. Assoc. Annu. Conf.* 1998, A, 389–403.
- 122. Rutala, W.A.; Gergen, M.F.; Weber, D.J. Room Decontamination with UV Radiation. *Infect. Control Hosp. Epidemiol.* 2010, 31, 1025–1029, doi:10.1086/656244.
- 123. Lindblad, M.; Tano, E.; Lindahl, C.; Huss, F. Ultraviolet-C decontamination of a hospital room: Amount of UV light needed. Burns 2020, 46, 842–849, doi:10.1016/j.burns.2019.10.004.
- 124. Oguma, K.; Katayama, H.; Ohgaki, S. Photoreactivation of Escherichia coli after low- or medium-pressure UV disinfection determined by an endonuclease sensitive site assay. *Appl. Environ. Microbiol.* 2002, 68, 6029–6035, doi:10.1128/AEM.68.12.6029-6035 2002
- Casini, B., Tuvo, B., Cristina, M. L., Spagnolo, A. M., Totaro, M., Baggiani, A., & Privitera, G. P. Evaluation of an ultraviolet C (UVC) light-emitting device for disinfection of high touch surfaces in hospital critical areas. Int. J. Environ. Res. Public Health, 2010, 16710, 2572
- Kumar, A.; Sagdeo, A.; Sagdeo, P.R. Possibility of using ultraviolet radiation for disinfecting the novel COVID-19. Photodiagnosis Photodyn. Ther. 2021, 34, 102234, doi:10.1016/j.pdpdt.2021.102234.
- 127. Kitagawa, H.; Nomura, T.; Nazmul, T.; Omori, K.; Shigemoto, N.; Sakaguchi, T.; Ohge, H. Effectiveness of 222-nm ultraviolet light on disinfecting SARS-CoV-2 surface contamination. *Am. J. Infect. Control* **2021**, 49, 299–301, doi:10.1016/j.ajic.2020.08.022.
- Jelden, K.C.; Gibbs, S.G.; Smith, P.W.; Hewlett, A.L.; Iwen, P.C.; Schmid, K.K.; Lowe, J.J. Comparison of hospital room surface disinfection using a novel ultraviolet germicidal irradiation (UVGI) generator. J. Occup. Environ. Hyg. 2016, 13, 690–698, doi:10.1080/15459624.2016.1166369.
- 129. Umezawa, K.; Asai, S.; Inokuchi, S.; Miyachi, H. A comparative study of the bactericidal activity and daily disinfection house-keeping surfaces by a new portable pulsed UV radiation device. Curr. Microbiol. 2012, 64, 581–587, doi:10.1007/s00284-012-0110-v.
- 130. Kovach, C.R.; Taneli, Y.; Neiman, T.; Dyer, E.M.; Arzaga, A.J.A.; Kelber, S.T. Evaluation of an ultraviolet room disinfection protocol to decrease nursing home microbial burden, infection and hospitalization rates. *BMC Infect. Dis.* **2017**, 17, 1–8, doi:10.1186/s12879-017-2275-2.
- 131. Rutala, W.A.; Gergen, M.F.; Tande, B.M.; Weber, D.J. Rapid Hospital Room Decontamination Using Ultraviolet (UV) Light with a Nanostructured UV-Reflective Wall Coating. *Infect. Control Hosp. Epidemiol.* 2013, 34, 527–529, doi:10.1086/670211.
- 132. Tiller, J.C. Antimicrobial surfaces. *Adv. Polym. Sci.* **2010**, 240, 193–217.
- Linklater, D.P.; Baulin, V.A.; Juodkazis, S.; Crawford, R.J.; Stoodley, P.; Ivanova, E.P. Mechano-bactericidal actions of nanostructured surfaces. Nat. Rev. Microbiol. 2021, 19, 8–22.
- 134. K. Webb, H.; Hasan, J.; K. Truong, V.; J. Crawford, R.; P. Ivanova, E. Nature Inspired Structured Surfaces for Biomedical Applications. Curr. Med. Chem. 2011, 18, 3367–3375, doi:10.2174/092986711796504673.
- Nguyen, S.H.T.; Webb, H.K.; Hasan, J.; Tobin, M.J., Crawford, R.J.; Ivanova, E.P. Dual role of outer epicuticular lipids in determining the wettability of dragonfly wings. Colloids Surfaces B Biointerfaces 2013, 106, 126–134, doi:10.1016/j.colsurfb.2013.01.042.
- 136. Ivanova, E.P.; Nguyen, S.H.; Webb, H.K.; Hasan, J.; Truong, V.K.; Lamb, R.N.; Duan, X.; Tobin, M.J.; Mahon, P.J.; Crawford, R.J. Molecular Organization of the Nanoscale Surface Structures of the Dragonfly Hemianax papuensis Wing Epicuticle. *PLoS One* **2013**, *8*, 67893, doi:10.1371/journal.pone.0067893.
- 137. Jenkins, J.; Mantell, J.; Neal, C.; Gholinia, A.; Verkade, P.; Nobbs, A.H.; Su, B. Antibacterial effects of nanopillar surfaces are mediated by cell impedance, penetration and induction of oxidative stress. *Nat. Commun.* **2020**, *11*, 1–14, doi:10.1038/s41467-020-15471-x.
- Knetsch, M.L.W.; Koole, L.H. New strategies in the development of antimicrobial coatings: The example of increasing usage of silver and silver nanoparticles. *Polymers.* 2011, 3, 340–366.
- 139. Maillard, J.Y.; Hartemann, P. Silver as an antimicrobial: facts and gaps in knowledge. Crit. Rev. Microbiol. 2013, 39, 373–383.
- Weaver, L.; Noyce, J.O.; Michels, H.T.; Keevil, C.W. Potential action of copper surfaces on meticillin-resistant Staphylococcus aureus. I. Appl. Microbiol. 2010. 109. 2200–2205. doi:10.1111/j.1365-2672.2010.04852.x.
- 141. Noyce, J.O.; Michels, H.; Keevil, C.W. Potential use of copper surfaces to reduce survival of epidemic meticillin-resistant Staphylococcus aureus in the healthcare environment. J. Hosp. Infect. 2006, 63, 289–297, doi:10.1016/j.jhin.2005.12.008.
- Ruparelia, J.P.; Chatterjee, A.K.; Duttagupta, S.P.; Mukherji, S. Strain specificity in antimicrobial activity of silver and copper nanoparticles. Acta Biomater. 2008, 4, 707–716, doi:10.1016/j.actbio.2007.11.006.
- 143. Jin, S.E.; Jin, H.E. Antimicrobial activity of zinc oxide nano/microparticles and their combinations against pathogenic microorganisms for biomedical applications: From physicochemical characteristics to pharmacological aspects. *Nanomaterials* **2021**, *11*, 1–35.
- 144. Silva, B.L. da; Abuçafy, M.P.; Manaia, E.B.; Junior, J.A.O.; Chiari-Andréo, B.G.; Pietro, R.C.L.R.; Chiavacci, L.A. Relationship between structure and antimicrobial activity of zinc oxide nanoparticles: An overview. Int. J. Nanomedicine 2019, 14, 9395–9410.

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145. Grass, G.; Rensing, C.; Solioz, M. Metallic copper as an antimicrobial surface. Appl. Environ. Microbiol. 2011, 77, 1541–1547.

- Valeria Prado, J.; Roberto Vidal, A.; Claudia Durán, T. Aplicación de la capacidad bactericida del cobre en la práctica médica. Rev. Med. Chil. 2012, 140, 1325-1332.
- Vincent, M.; Hartemann, P.; Engels-Deutsch, M. Antimicrobial applications of copper. Int. J. Hyg. Environ. Health 2016, 219, 585-591, doi:10.1016/j.ijheh.2016.06.003.
- 148. Villapún, V.M.; Dover, L.G.; Cross, A.; González, S. Antibacterial metallic touch surfaces. Materials. 2016, 9, 736
- Santos, M.R.E.; Mendonça, P. V.; Branco, R.; Sousa, R.; Dias, C.; Serra, A.C.; Fernandes, J.R.; Magalhães, F.D.; Morais, P. V.; Coelho, J.F.J. Light-Activated Antimicrobial Surfaces Using Industrial Varnish Formulations to Mitigate the Incidence of Nosocomial Infections. ACS Appl. Mater. Interfaces 2021, 13, 7567–7579, doi:10.1021/acsami.0c18930.
- Wu, X.; Huang, Y.Y.; Kushida, Y.; Bhayana, B.; Hamblin, M.R. Broad-spectrum antimicrobial photocatalysis mediated by titanium dioxide and UVA is potentiated by addition of bromide ion via formation of hypobromite. Free Radic. Biol. Med. 2016, 95, 74-81, doi:10.1016/j.freeradbiomed.2016.03.012.
- 151. Visnapuu, M.; Rosenberg, M.; Truska, E.; Nõmmiste, E.; Šutka, A.; Kahru, A.; Rähn, M.; Vija, H.; Orupõld, K.; Kisand, V.; et al. UVA-induced antimicrobial activity of ZnO/Ag nanocomposite covered surfaces. Colloids Śurfaces B Biointerfaces 2018, 169, 222-232, doi:10.1016/j.colsurfb.2018.05.009.
- 152. Eby, D.M.; Luckarift, H.R.; Johnson, G.R. Hybrid antimicrobial enzyme and silver nanoparticle coatings for medical Instruments. ACS Appl. Mater. Interfaces 2009, 1, 1553-1560, doi:10.1021/am9002155.
- Rudra, J.S.; Dave, K.; Haynie, D.T. Antimicrobial polypeptide multilayer nanocoatings. J. Biomater. Sci. Polym. Ed. 2006, 17, 1301-
- 1315, doi:10.1163/156856206778667433
- 154. Waschinski, C.J.; Herdes, V.; Schueler, F.; Tiller, J.C. Influence of satellite groups on telechelic antimicrobial functions of polyoxazolines. Macromol. Biosci. 2005, 5, 149-156, doi:10.1002/mabi.200400169
- Fundeanu, I.; van der Mei, H.C.; Schouten, A.J.; Busscher, H.J. Polyacrylamide brush coatings preventing microbial adhesion to silicone rubber. Colloids Surf. B. 2008, 64, 297-301, doi:10.1016/j.colsurfb.2008.02.005.
- Chapman, R.G.; Ostuni, E.; Liang, M.N.; Meluleni, G.; Kim, E.; Yan, L.; Pier, G.; Warren, H.S.; Whitesides, G.M. Polymeric thin films that resist the adsorption of proteins and the adhesion of bacteria. Langmuir 2001, 17, 1225-1233, doi:10.1021/la001222d.
- D'Almeida, M.; Attik, N.; Amalric, J.; Brunon, C.; Renaud, F.; Abouelleil, H.; Toury, B.; Grosgogeat, B. Chitosan coating as an antibacterial surface for biomedical applications. PLoS One 2017, 12, doi:10.1371/journal.pone.0189537.
- 158. Alves, D.; Olívia Pereira, M. Mini-review: Antimicrobial peptides and enzymes as promising candidates to functionalize biomaterial surfaces. Biofouling 2014, 30, 483-499.
- Zubris, D.; Minbiole, K.; Wuest, W. Polymeric Quaternary Ammonium Compounds: Versatile Antimicrobial Materials. Curr. Top. Med. Chem. 2016, 17, 305-318, doi:10.2174/1568026616666160829155805.
- Grigoras, A.G. Natural and synthetic polymeric antimicrobials with quaternary ammonium moieties: a review. Environ. Chem. Lett. 2021, 1, 3
- Kenawy, E.R.; Worley, S.D.; Broughton, R. The chemistry and applications of antimicrobial polymers: A state-of-the-art review. Biomacromolecules 2007, 8, 1359-1384.
- 162. Chapman, I.S. Biocide resistance mechanisms. Int. Biodeterior. Biodegrad. 2003, 51, 133-138, doi:10.1016/S0964-8305(02)00097-5.
- Amsalu, A.; Sapula, S.A.; Lopes, M.D.B.; Hart, B.J.; Nguyen, A.H.; Drigo, B.; Turnidge, J.; Leong, L.E.; Venter, H. Efflux pumpdriven antibiotic and biocide cross-resistance in pseudomonas Aeruginosa isolated from different ecological Niches: A case study in the development of multidrug resistance in environmental hotspots. Microorganisms 2020, 8, 1-18, doi:10.3390/microorganisms8111647
- 164. Paul, D.; Chakraborty, R.; Mandal, S.M. Biocides and health-care agents are more than just antibiotics: Inducing cross to coresistance in microbes. Ecotoxicol. Environ. Saf. 2019, 174, 601-610, doi:10.1016/j.ecoenv.2019.02.083.
- Jennings, M.C.; Forman, M.E.; Duggan, S.M.; Minbiole, K.P.C.; Wuest, W.M. Efflux Pumps Might Not Be the Major Drivers of OAC Resistance in Methicillin-Resistant Staphylococcus aureus. Chem Bio Chem 2017, 18, 1573–1577, doi:10.1002/cbic.201700233.
- Morrison, K.R.; Allen, R.A.; Minbiole, K.P.C.; Wuest, W.M. More QACs, more questions: Recent advances in structure activity relationships and hurdles in understanding resistance mechanisms. Tetrahedron Lett. 2019, 60, 150935.
- Dukan, S.; Touati, D. Hypochlorous acid stress in Escherichia coli: Resistance, DNA damage, and comparison with hydrogen eroxide stress. J. Bacteriol. 1996, 178, 6145-6150, doi:10.1128/jb.178.21.6145-6150.1996.
- Ribeiro, M.; Simões, L.C.; Simões, M. Biocides. In Encyclopedia of Microbiology; Elsevier, 2019; pp. 478–490.
- Siedlecka, A. Antibiotic and Disinfectant Resistance in Tap Water Strains Insight into the Resistance of Environmental Bacteria. Polish J. Microbiol. 2021, 70, 57-67, doi:10.33073/pjm-2021-004.
- 170. McBain, A.J.; Bartolo, R.G.; Catrenich, C.E.; Charbonneau, D.; Ledder, R.G.; Price, B.B.; Gilbert, P. Exposure of sink drain microcosms to triclosan: Population dynamics and antimicrobial susceptibility. Appl. Environ. Microbiol. 2003, 69, 5433-5442, doi:10.1128/AEM.69.9.5433-5442.2003.
- 171. Kümmerer, K. Resistance in the environment. J. Antimicrob. Chemother. 2004, 54, 311-320.
- 172. Baquero, F.; Martínez, J.-L.; Cantón, R. Antibiotics and antibiotic resistance in water environments. Curr. Opin. Biotechnol. 2008, 19, 260-265

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