

1 Magnetism and NIR dual-response polypyrrole-  
2 coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles for bacteria removal  
3 and inactivation  
4

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23 **Abstract**

24 Microbial contamination of water represents a great threat to the public health that has  
25 attracted worldwide attention. In this work, polypyrrole magnetic nanoparticles  
26 ( $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs) with sterilization properties were fabricated. More specifically, the  
27  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs obtained via aqueous dispersion polymerization and an *in situ*  
28 chemical oxidative polymerization exhibited a cationic surface and high photothermal  
29 conversion efficiency. Using *E. coli* as a model system, it was possible to observe that  
30 more than 50% of bacteria adsorption can be achieved at a dosage of 100  $\mu\text{g}/\text{mL}$  under  
31 magnetic field, and high photothermal sterilization efficacy ( $\sim 100\%$ ) can be obtained  
32 upon NIR exposure at the same dosage for 10 min. Noteworthy, the  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs  
33 can be recycled by magnetism and reused without affecting their photothermal  
34 sterilization capability. This study clearly provides experimental evidence of the great  
35 potential of  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs as stable and reusable nanocomposite material for  
36 photothermal sterilization applications.

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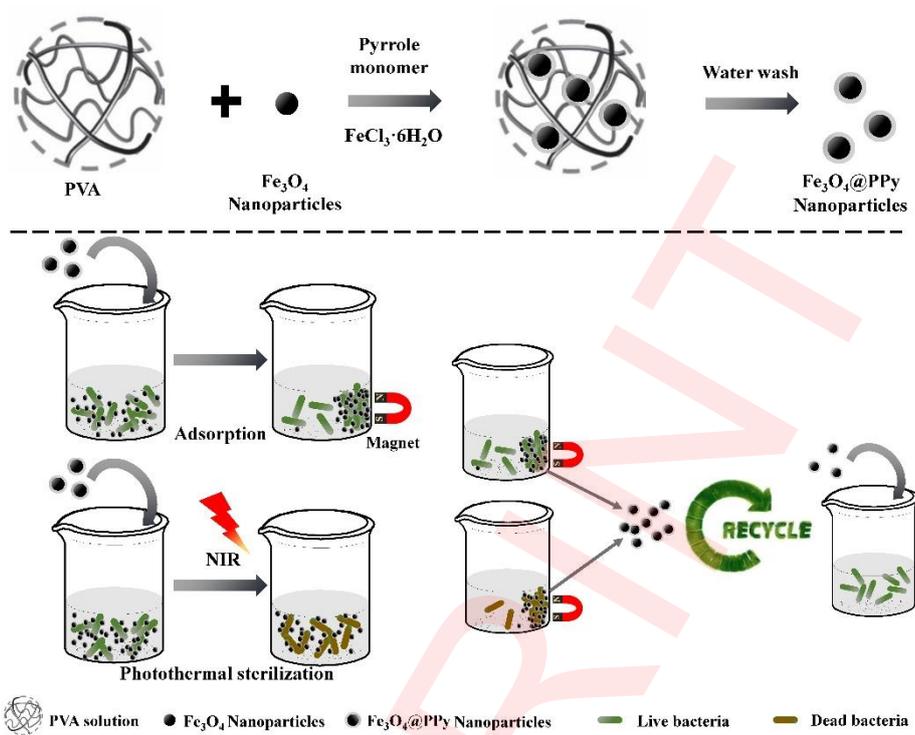
40 *Keywords:* bacteria adsorbent; cationic polymers; *E. coli*; photothermal sterilization;  
41 water safety management.

42

43

### Graphical Abstract

44



## 46 **1. Introduction**

47 It has been estimated that 80 countries in the world suffer from shortage of drinking  
48 water, with approximately 1.2 billion people being exposed to contaminated drinking  
49 water.<sup>1</sup> Proliferation of pathogens bacteria has been regarded as one of the main causes  
50 of water contamination, which can lead to dysentery, colitis, and even uremia, with a  
51 negative impact on the public health and safety.<sup>2</sup> Water safety management is therefore  
52 attracting an increasing attention all over the world. Until now, the approaches used to  
53 disinfect the contaminated water foresee physical and chemical processes. Physical  
54 processes usually include filtration, thermal treatment, and ultraviolet (UV) irradiation.  
55 However, these approaches have their shortages and limitations. For example, they are  
56 time consuming, bio-resources wasting, and their applications are limited.<sup>3</sup> Chemical  
57 processes are based on the use of additives, such as traditional agents (chlorine, chlorine  
58 dioxide, ozone, *etc.*)<sup>4</sup> and novel agents (antibiotics, metal particles, *etc.*).<sup>5</sup> However,  
59 issues associated to their cost and toxicity make these agents an unfavorable option. For  
60 all the above reasons, there is a need to explore more suitable approaches to eliminate  
61 bacteria in drinking water in an efficient way.

62 In the past few years, photothermal transduction agents (PTAs) have obtained widely  
63 recognition in cancer therapy, whereby PTAs now represent a promising alternative to  
64 conventional approaches.<sup>6</sup> PTAs can harvest energy from light and convert the energy  
65 into heat, generating local heat even up to 70 °C.<sup>7</sup> Since most bacteria can be devitalized  
66 above 55 °C,<sup>8</sup> photothermal sterilization mediated by PTAs is a viable option for  
67 microbial control.<sup>9</sup> Sun *et al.* utilized prussian blue as photothermal agent with excellent

68 bacterial inactivation performance.<sup>9a</sup> However, prussian blue can rapidly decompose  
69 under alkaline conditions, which hinder its use for practical purposes.<sup>10</sup> It is notable that  
70 part of the photothermal agents are susceptible to pH, ionic strength, and other adverse  
71 environments, which influence the stability and thermal conversion efficiency, beside  
72 raising serious concerns on the possible formation of toxic by-products.<sup>11</sup> Therefore, it  
73 is of utmost importance to select most suitable photothermal agents in order to achieve  
74 an efficient photothermal sterilization of water.

75 Polypyrrole nanoparticles, a new type of low cost, highly stable, and biocompatible  
76 photothermal agents, have received great attention in photothermal therapy.<sup>12</sup> They  
77 have been demonstrated as wide-coverage materials, being able to interact efficiently  
78 within the entire UV-visible spectrum, up to the near infrared (NIR) region. The low  
79 wavelength coverage (UV-light region) is attributed to the  $\pi - \pi^*$  transition for the  
80 conjugated polypyrrole backbone, whereas the broad wavelength coverage from the  
81 visible region to the NIR region is the characteristic of the bipolaronic polypyrrole.<sup>13</sup>  
82 Moreover, polypyrrole nanoparticles are also stable in alkaline or acidic surroundings  
83 and can bear exposure to severe conditions.<sup>14</sup> Based on these properties, polypyrrole  
84 nanoparticles are well suited to utilize a large portion of the solar spectrum for  
85 photothermal sterilization.

86 Magnetic nanoparticles have attracted great interest due to their intrinsic properties.<sup>15</sup>  
87 In the field of water management, they can be easily dispersed and recovered from the  
88 aqueous medium by on/off switching of an external magnetic field, with great  
89 advantage in terms of no losses, no energy consumption, and no by-products

90 formation.<sup>16</sup> Among the different types of magnetic nanoparticles, Fe<sub>3</sub>O<sub>4</sub>-based  
91 nanoparticles have been widely used for a variety of applications, especially due to their  
92 low cost and biocompatibility.<sup>17</sup> Because of the negative charge of the cell membrane  
93 of bacteria, using positively-charged magnetic nanoparticles is expected to promote the  
94 nanoparticle/bacteria interaction, thus improving the removal efficiency.<sup>18</sup>

95 In this study, polypyrrole magnetic nanoparticles (Fe<sub>3</sub>O<sub>4</sub>@PPy NPs) were prepared  
96 via aqueous dispersion polymerization and an *in situ* chemical oxidative polymerization  
97 method. The proposed nanomaterials have been conceived as an alternative approach  
98 for water sterilization in the broad context of water safety management. To this scope,  
99 the obtained nanoparticles were characterized in terms of morphology, dispersibility in  
100 water, conversion efficiency of the absorbed NIR photons into heat, bacteria removal  
101 efficiency, and photothermal sterilization ability. The reusability of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs  
102 was also evaluated.

103

## 104 **2. Experimental**

### 105 **2.1. Materials**

106 Ethylene glycol, polyvinyl alcohol (PVA) and pyrrole were purchased from Aladdin  
107 chemicals company (Shanghai, China). Iron chloride hexahydrate (FeCl<sub>3</sub>·6H<sub>2</sub>O),  
108 trisodium citrate and sodium acetate were purchased from Beijing chemical reagents  
109 company (Beijing, China). All the chemicals and reagents were of analytical grade.  
110 Deionized (DI) water (18.25 MΩ·cm) was acquired from a Milli-Q water-purification  
111 system (Bedford, MA, USA).

112

## 113 2.2. Preparation of the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs

### 114 2.2.1. Synthesis of Fe<sub>3</sub>O<sub>4</sub> nanoparticles

115 The Fe<sub>3</sub>O<sub>4</sub> nanoparticles (Fe<sub>3</sub>O<sub>4</sub> NPs) were prepared via a modified solvothermal  
116 reaction as previously reported.<sup>17a</sup> Briefly, 1.3 g FeCl<sub>3</sub>·6H<sub>2</sub>O and 0.40 g trisodium  
117 citrate were dissolved in 40 mL ethylene glycol by means of an ultrasonicator (KQ-  
118 400KDV, Kunshan, China). Then, 2.4 g sodium acetate was added to the above solution  
119 and stirred at 1000 rpm for 30 minutes. The obtained homogeneous solution was sealed  
120 in a Teflon-lined stainless-steel autoclave (100 mL volume) and heated at 200 °C for  
121 12 h. After cooling to room temperature, the products were washed three times with  
122 ethanol and water, and the final nanoparticles were dried under vacuum for 12 h at 45 °C.

### 123 2.2.2. Synthesis of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs

124 The Fe<sub>3</sub>O<sub>4</sub>@PPy NPs were synthesized via an *in situ* chemical oxidative  
125 polymerization method in the presence of Fe<sub>3</sub>O<sub>4</sub> @NPs. Briefly, 3.0 g PVA was  
126 dissolved in 30 mL hot DI water under mechanical agitation until a clear sticky liquid  
127 was obtained. After cooling to room temperature, 100 mg of Fe<sub>3</sub>O<sub>4</sub> NPs was redispersed  
128 in the PVA solution under ultrasonication at room temperature for 10 min. Then, the  
129 solution was equilibrated for 1 h before 200 μL of pyrrole monomer was added. After  
130 further stirring for 30 min, 1.6 g FeCl<sub>3</sub>·6H<sub>2</sub>O was used as an oxidant to initiate the  
131 polymerization. The polymerization proceeded at room temperature for 6 h to obtain  
132 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs. The so obtained nanoparticles were collected by magnetic separation  
133 and then washed with hot water for three times to remove pyrrole in excess and other  
134 reagents. The final nanoparticles were dried under vacuum for 12 h at 45 °C.

## 136 2.3. Characterization

137 The size and morphology of Fe<sub>3</sub>O<sub>4</sub> NPs and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs were determined by  
138 transmission electron microscopy (TEM) (H-7650, Hitachi Instruments, Japan)  
139 operating at an acceleration voltage of 100 kV. Thermogravimetric analysis (TGA) was  
140 carried out with a thermogravimetric analyzer (449C, TA Instruments, USA) at a  
141 heating speed rate of 10 °C/min under N<sub>2</sub> atmosphere from room temperature to 800 °C.  
142 Fourier transform infrared (FTIR) spectroscopy was performed using an infrared  
143 spectrometer (Nicolet IS 5, Thermo Scientific, USA) with KBr as background over the  
144 range of 4000–400 cm<sup>-1</sup>. The magnetic properties of nanoparticles were assessed using  
145 a quantum design system (PPMS-9, Quantum Design Company, USA) at room  
146 temperature with a maximum magnetic field of 10 kOe. Hydrodynamic diameter and  
147 zeta potential measurements were run by dynamic light scattering (DLS) equipment  
148 (ZetaPALS, Brookhaven Instruments, USA). The UV-vis spectra ranging from 400 nm  
149 to 900 nm were obtained with a UV-vis spectrophotometer (UV-2600, Shimadzu  
150 Instruments, China).

151

#### 152 **2.4. Photothermal performance of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs**

153 The photothermal performance of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was firstly evaluated by  
154 acquiring the real-time photothermal heating curves exposed to NIR laser irradiation  
155 (808 nm, 2 W/cm<sup>2</sup>). Fe<sub>3</sub>O<sub>4</sub>@PPy NPs aqueous dispersions at different concentrations  
156 (10, 20, 50, 100 µg/mL) were irradiated for 10 min. The temperature change was  
157 monitored and recorded every 30 s using a submerged digital thermometer with a  
158 thermocouple microprobe. The heating curve was made by plotting the measured  
159 temperature versus time. DI water irradiated by NIR laser was used as control. In  
160 addition, the effect of radiant power at different levels (0.5, 1, 1.5, 2 and 2.5 W/cm<sup>2</sup> for  
161 10 min exposure) on photothermal performance was also determined. The temperature

162 change was monitored and recorded every 30 s, and the heating curve was plotted. The  
163 photothermal conversion efficiency of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was measured by recording the  
164 temperature change of dispersion under continuous irradiation of an 808 nm laser (2  
165 W/cm<sup>2</sup>) until the temperature of the solution reached a steady-state evolution, followed  
166 by spontaneous cooling to room temperature. The photothermal conversion efficiency  
167 was calculated according to the calculation method reported by Chang et al.<sup>19</sup> The  
168 photostability of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was analyzed by the LASER ON and LASER OFF  
169 procedure with NIR laser described by Li et al. with a little modification.<sup>20</sup> The LASER  
170 ON procedure was operated for 10 min, and then the samples were let to cool to room  
171 temperature for 30 min with no exposure to NIR laser irradiation (LASER OFF). This  
172 cycle was continuously repeated for 5 times. The real-time temperature increase of  
173 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs upon the NIR laser irradiation (2 W/cm<sup>2</sup>) was visualized by IR  
174 thermal camera (Ti25, Fluke, USA).

175

## 176 **2.5. Bacteria culture and preparation**

177 The *E. coli* strains were used as a model microorganism in this study. They were  
178 provided by the Institute of Applied Microbiology, Heilongjiang Academy of Science  
179 (China). Bacteria cultures were maintained on an agar plates at 4 °C and subcultured  
180 monthly. Before each experiment, *E. coli* were activated in nutrient broth at 37 °C up  
181 to the mid-log phase. The bacteria suspensions were centrifuged at 3502 g for 10 min  
182 and washed with saline buffer (0.9 wt% NaCl in water). The *E. coli* cells were then  
183 poured into a buffer solution to achieve the required concentration.

184

## 185 **2.6. Bacteria removal efficiency**

186 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs at different concentrations were added in 10<sup>6</sup> colony forming unit

187 (CFU) bacteria suspension (3 mL) and thoroughly dispersed for 10 minutes to promote  
188 interaction between NPs and bacteria. An external magnet was then placed on the wall  
189 of the cuvette, the NPs set aside by the magnetic field, and the clear suspension  
190 separated and collected in a beaker. The removed suspension was diluted appropriately  
191 and seeded on agar plates. Thereafter, the CFU of the bacteria suspension was counted  
192 by the plate counting method. Briefly, 100  $\mu$ L of the diluted suspension was placed onto  
193 a sterile solid medium and cultured at 37 °C for 24 h. Bare bacteria suspensions (i.e.,  
194 which did not enter in contact with Fe<sub>3</sub>O<sub>4</sub>@PPy NPs) were used as a control.

195

## 196 **2.7. Photothermal sterilization**

197 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs at different concentrations were added in 10<sup>6</sup> CFU bacteria  
198 suspension (3 mL) and then irradiated with 808 nm wavelength at a power density of 2  
199 W/cm<sup>2</sup> for 10 min. The bacteria suspension without irradiation was used as a control.  
200 After irradiation, the suspensions were used for estimating the photothermal  
201 sterilization activity of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs by plate counting method, following the same  
202 procedure described above.

203

## 204 **2.8. Recyclability**

205 After a first photothermal sterilization test, Fe<sub>3</sub>O<sub>4</sub>@PPy NPs were collected by a  
206 magnetic force for 10 min, and then subjected to a 3 min water bath sonication for  
207 removal of impurities. A second photothermal sterilization test was thus performed, up  
208 to a total of four cycles. At the end of each cycle, the photothermal sterilization activity  
209 of the recycled Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was determined by plate counting method. The UV-  
210 vis spectra of the recycled Fe<sub>3</sub>O<sub>4</sub>@PPy NPs exposed to NIR irradiation and magnetic

211 separation were also recorded.

212

## 213 **2.9. Bacteria morphology change observation**

214 The interaction between *E. coli* and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs and the morphological change  
215 of *E. coli* after NIR irradiation were observed by TEM. More specifically, the *E.*  
216 *coli*/Fe<sub>3</sub>O<sub>4</sub>@PPy NP complexes were first collected and the excess (unbound) *E. coli*  
217 cells and impurities washed out with DI water. At this point, the interaction between *E.*  
218 *coli* and NPs was evaluated by TEM. The *E. coli* and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs suspensions  
219 after NIR irradiation were centrifuged and washed with DI water for morphological  
220 change observations by TEM. The untreated *E. coli* were taken as control.

221

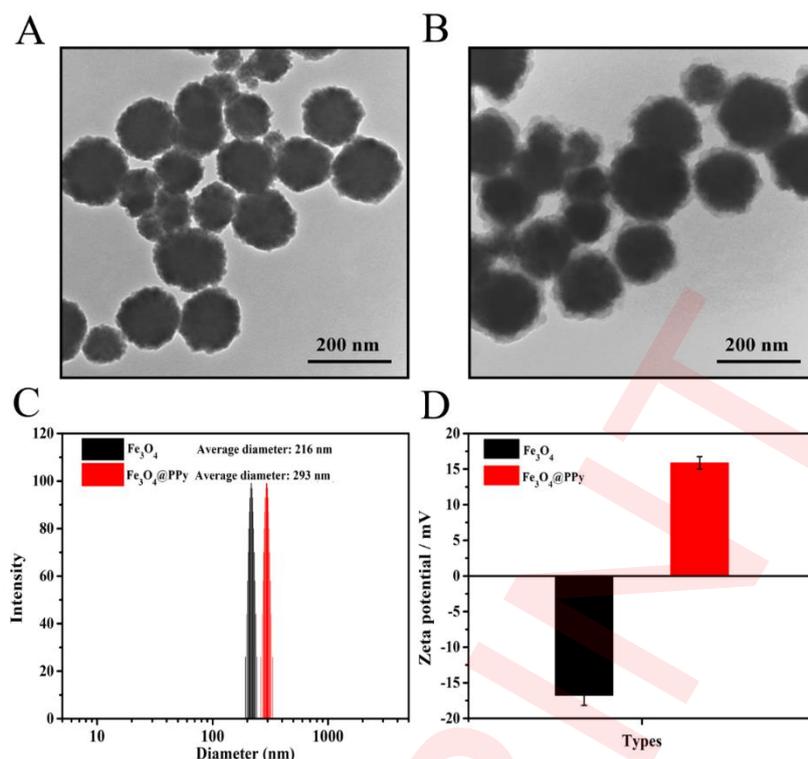
## 222 **3. Results and discussion**

### 223 **3.1. Preparation and characterization of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs**

224 Amongst various magnetic nanoparticles, experimental evidence suggests that those  
225 obtained by the modification of carboxylate groups exhibit favorable stability,  
226 biocompatibility, and dispersibility.<sup>21</sup> In this study, carboxylate groups-modified  
227 magnetite nanoparticles were synthesized via a modified solvothermal reaction. Then,  
228 an *in situ* chemical oxidative polymerization method was used to fabricate Fe<sub>3</sub>O<sub>4</sub>@PPy  
229 NPs.

230 As shown in **Figure 1A** and **Figure 1B**, both Fe<sub>3</sub>O<sub>4</sub> NPs and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs  
231 exhibited uniform spherical shape. Compared with Fe<sub>3</sub>O<sub>4</sub> NPs, a thin layer coated on  
232 the surface of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was clearly visible, indicative of the successful  
233 modification of the nanoparticles with pyrrole polymers. Particle size measurement by  
234 TEM revealed that the mean size of Fe<sub>3</sub>O<sub>4</sub> NPs and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was 185 ± 15 nm

235 and  $210 \pm 23$  nm, respectively. The slight size increase for  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs can be  
236 attributed to the shell structure of pyrrole polymer. The hydrate particle sizes of  $\text{Fe}_3\text{O}_4$   
237 NPs and  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs were  $216 \pm 18$  nm and  $293 \pm 27$  nm, respectively (**Figure**  
238 **1C**). The larger particle size measured by DLS compared to TEM images was due to  
239 the fact that the mean diameter of NPs by DLS was calculated from the diffusional  
240 properties of dynamic nanoparticles in hydrated state, whereas in the case of TEM  
241 images, they were captured from nanoparticles in dried state.<sup>22</sup> The zeta potential of  
242  $\text{Fe}_3\text{O}_4$  NPs was  $-16.77 \pm 1.40$  mV, but it increased up to  $15.89 \pm 0.87$  mV after  
243 modification with polypyrrole (**Figure 1D**), which behaved as a positively-charged  
244 polymer.  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs exhibited a core-shell architecture, which was attributed to  
245 the commonion effect of the oxidant  $\text{Fe}^{3+}$  aggregated around the surface of  $\text{Fe}_3\text{O}_4$  NPs  
246 and the complexation processes of  $\text{Fe}_3\text{O}_4$  core with polypyrrole. The common-ion effect  
247 and complexation processes ensured the success of the particles preparation, and  
248 enhanced the stability of the  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs.<sup>23</sup>



249

250 **Figure 1.** TEM images of Fe<sub>3</sub>O<sub>4</sub> NPs (A) and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs (B), hydrodynamic  
 251 diameter of Fe<sub>3</sub>O<sub>4</sub> NPs and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs (C), and zeta potential of Fe<sub>3</sub>O<sub>4</sub> NPs and  
 252 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs (D).

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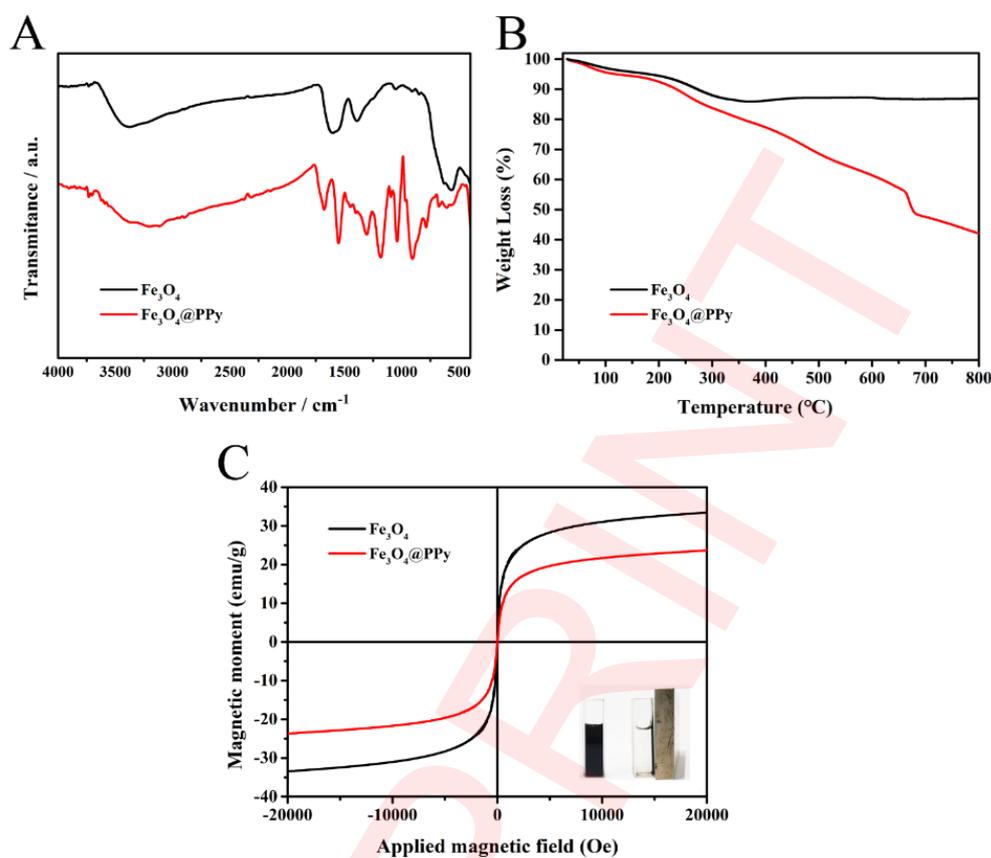
254 The FT-IR spectroscopy results of the Fe<sub>3</sub>O<sub>4</sub> NPs and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs are shown in  
 255 **Figure 2A.** For Fe<sub>3</sub>O<sub>4</sub> NPs, the bands at 563 and 420 cm<sup>-1</sup> were both associated with  
 256 the Fe-O stretching, while the bands centered at 1601 and 1390 cm<sup>-1</sup> were assigned to  
 257 the carboxylate group. The bands at 1052 and 3376 cm<sup>-1</sup> were ascribed to the vibrating  
 258 mode of both C-H and O-H functional groups.<sup>24</sup> The characteristic absorption bands of  
 259 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs at 1556 and 1403 cm<sup>-1</sup> were attributed to the fundamental stretching  
 260 vibration of pyrrole rings, whereas the band at 1297 cm<sup>-1</sup> was attributed to C-N  
 261 stretching vibration. The other peaks at 1195, 1043, 910 and 786 cm<sup>-1</sup> were due to the  
 262 in-plane, out-of-plane, C-H and N-H bending vibrations. The shifting of the peak

263 associated with the Fe-O stretching from  $563\text{ cm}^{-1}$  to  $605\text{ cm}^{-1}$  indicates the presence  
264 of  $\text{Fe}_3\text{O}_4$  as well as the interaction of  $\text{Fe}_3\text{O}_4$  with polypyrrole.<sup>25</sup> All these peaks  
265 demonstrated the successful preparation of  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs.

266 **Figure 2B** showed the TGA plots of the  $\text{Fe}_3\text{O}_4$  NPs and  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs. The  
267 different stages of weight loss were correspondingly due to different physicochemical  
268 degradation phenomena of the nanoparticles. The first weight loss of both  $\text{Fe}_3\text{O}_4$  NPs  
269 and  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs ranging from  $64\text{ }^\circ\text{C}$  to  $142\text{ }^\circ\text{C}$  might be attributed to the loss of  
270 adsorbed water. The second weight loss of  $\text{Fe}_3\text{O}_4$  NPs, ranging from  $280\text{ }^\circ\text{C}$  to  $400\text{ }^\circ\text{C}$ ,  
271 might be possibly generated by the degradation of magnetic nanoparticles. Conversely,  
272 the second weight loss (from  $260\text{ }^\circ\text{C}$  to  $670\text{ }^\circ\text{C}$ ) observed for the  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs might  
273 be attributed to the degradation of polypyrrole. Finally, the third weight loss appeared  
274 only in the  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs was plausibly due to the degradation of the core part of  
275 magnetic nanoparticles.

276 The magnetic properties of  $\text{Fe}_3\text{O}_4$  NPs and  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs were determined by the  
277 measurement of the magnetic hysteresis curve at 300K (room temperature). As shown  
278 in **Figure 2C**, both  $\text{Fe}_3\text{O}_4$  NPs and  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs had the typical superparamagnetic  
279 behavior. At the same time, it was not surprising that the modification of  $\text{Fe}_3\text{O}_4$  NPs  
280 surface with polypyrrole weakened the magnetic properties, due to the nonmagnetic  
281 features of polypyrrole. The saturation magnetization value of the  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs was  
282  $23.87\text{ emu/g}$ , which was strong enough for magnetic separation.<sup>26</sup> The visual photos of  
283 the  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs suspensions before and after the application of an external  
284 magnetic field were displayed in the inset of **Figure 2C**. The  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs can be  
285 well-dispersed in the aqueous solution without any external magnetic field; when a  
286 magnet was placed on the wall of the cuvette, the  $\text{Fe}_3\text{O}_4@\text{PPy}$  NPs rapidly moved  
287 toward the cuvette wall and the suspension turned from black to totally transparent, thus

288 confirming that the polypyrrole was tightly bound to the  $\text{Fe}_3\text{O}_4$  NPs, with virtually no  
289 residual (unbound) pyrrole polymer in the solution.



290

291 **Figure 2.** FTIR spectra (A) and TGA curves (B) of  $\text{Fe}_3\text{O}_4$  NPs and  $\text{Fe}_3\text{O}_4@PPy$  NPs,  
292 and magnetic hysteresis curve of the  $\text{Fe}_3\text{O}_4$  NPs and  $\text{Fe}_3\text{O}_4@PPy$  NPs at 300 K (C).

293 The insert in the figure is the photograph of  $\text{Fe}_3\text{O}_4@PPy$  NPs suspensions before and  
294 after an external magnetic field treatment.

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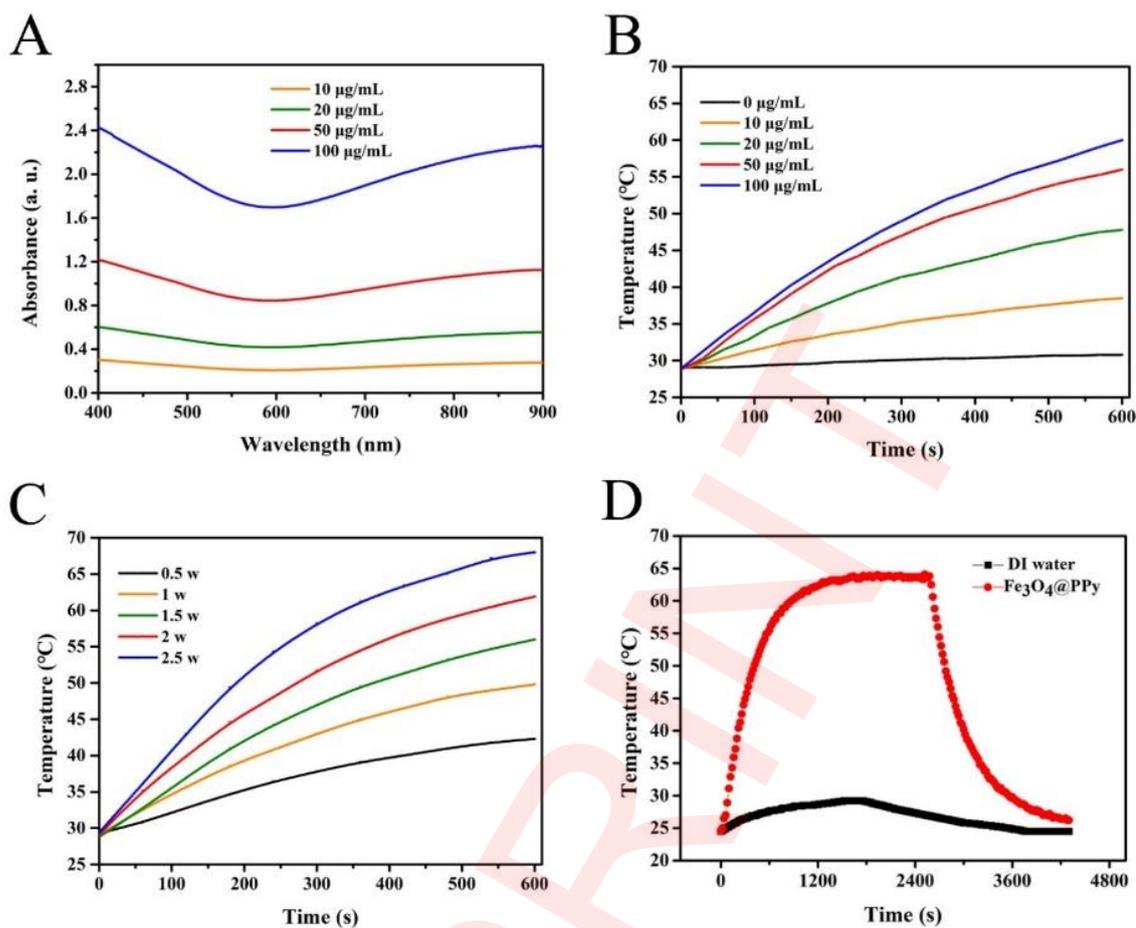
296 An ideal photothermal nanomaterial should have strong spectral absorption and high  
297 photothermal conversion ability.<sup>12a</sup> The spectral absorption of  $\text{Fe}_3\text{O}_4@PPy$  NPs was  
298 firstly measured by UV-vis spectra. As shown in **Figure 3A**, the  $\text{Fe}_3\text{O}_4@PPy$  NPs  
299 possessed a strong and broad absorption in the range from 400 nm to 900 nm, which  
300 endowed the  $\text{Fe}_3\text{O}_4@PPy$  NPs with both NIR-driven and full-spectrum applications.

301 The absorption at 900 nm was the characteristic of the bipolaronic band transitions of  
302 pyrrole polymers,<sup>27</sup> and the absorbance of the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs at 900 nm was in a high  
303 level. This demonstrated that the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs had the properties of polypyrrole,  
304 which has potential in photothermal applications.

305

### 306 **3.2. Photothermal properties of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs**

307 To evaluate the photothermal properties of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs, the photothermal  
308 conversion ability was measured under NIR laser exposure (808 nm, 2.0 W/cm<sup>2</sup>) by  
309 monitoring the real-time temperature changes. As shown in **Figure 3B**, a temperature  
310 increase was observed with increasing the concentration of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs from 10  
311 to 100 µg/mL. Moreover, the temperature increase was also dependent on the power  
312 density of the incident NIR light, as shown in **Figure 3C**. In particular, 10 min of NIR  
313 laser irradiation of 100 µg/mL Fe<sub>3</sub>O<sub>4</sub>@PPy NPs solution made possible a temperature  
314 increase from 28.9 °C to 60.1 °C. In comparison, the temperature of only water (i.e., in  
315 the absence of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs) upon 808 nm laser for 10 min only had a 2.1 °C  
316 increment. Here, the temperature variation of water can be assumed to be negligible  
317 compared to the effect observed for the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs exposed to the same  
318 irradiation power density. For the determination of the photothermal conversion  
319 efficiency of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs, we further measured the temperature change under  
320 continuous irradiation, until a steady-state temperature level was achieved. At this point,  
321 the laser was turned off to record the equilibrium temperature (**Figure 3D**).



322

323 **Figure 3.** UV-vis spectra of the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs (A), temperature increase of the

324 different concentrations of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs (B), temperature changes of the

325 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs with different laser power densities (C), and photothermal heating

326 curves of the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs and DI water (D).

327

328 The relationship between cooling time and negative natural logarithm of the

329 temperature driving force is shown in **Figure 4A**. The time constant for the heat transfer

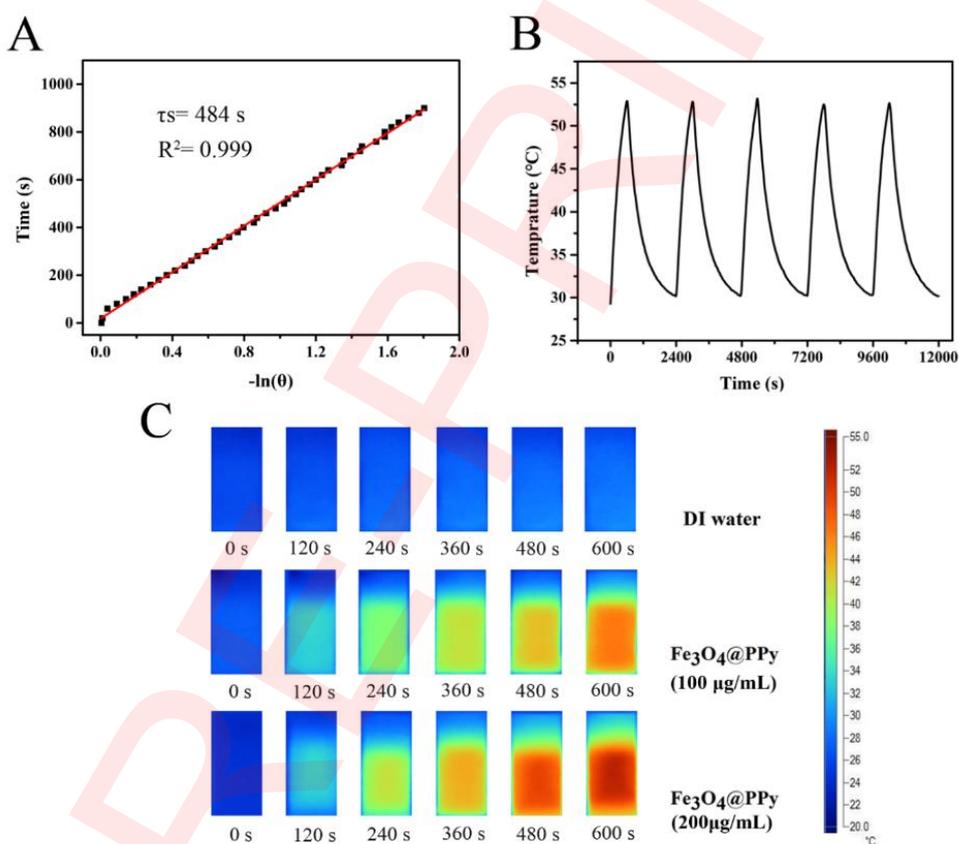
330 of the system was determined to be  $\tau_s = 484$  s. According to the generated results, the

331 calculated photothermal conversion efficiency of the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was 17%. The

332 “on-off” temperature curves of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs during five cycle examination were

333 presented in **Figure 4B**. The negligible fluctuation in temperature at each irradiation

334 cycle demonstrated the stability of the photothermal performance of the Fe<sub>3</sub>O<sub>4</sub>@PPy  
335 NPs. The real-time thermal images of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs exposed to irradiation are  
336 shown in **Figure 4C**. The color of the solutions gradually changed from blue  
337 (corresponding to low temperature) to dark red (corresponding to high temperature),  
338 clearly indicating that the temperature of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs solution increased with  
339 increasing both the irradiation time and sample concentration. These results indicated  
340 that the prepared Fe<sub>3</sub>O<sub>4</sub>@PPy NPs featured high photothermal conversion and good  
341 photothermal stability properties, which can be effectively used in photothermal  
342 applications.



343

344 **Figure 4.** Photothermal conversion efficiency (A) and photostability (B) of  
345 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs and thermal images of DI water and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs (C).

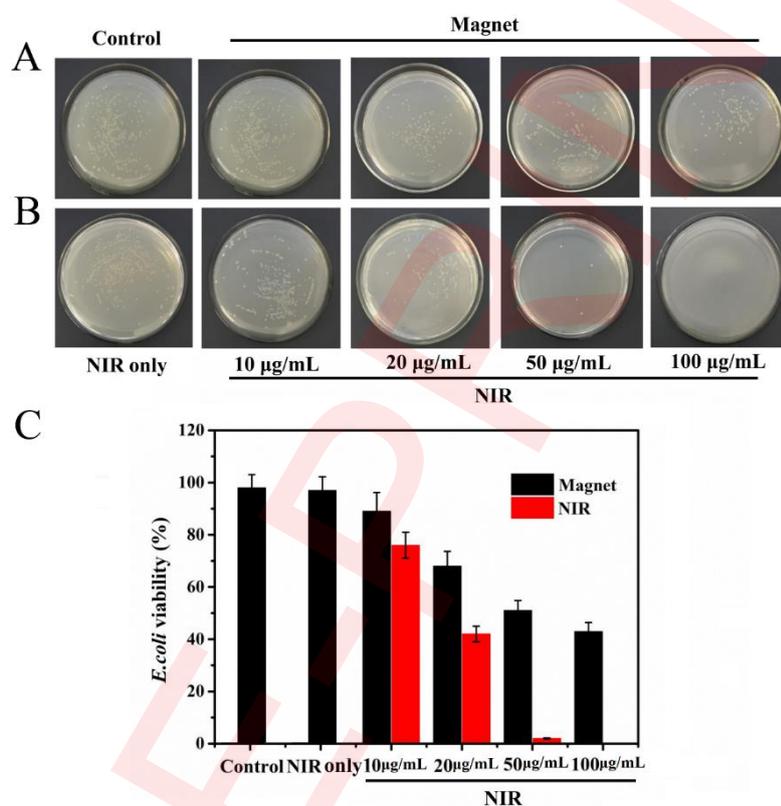
346

### 347 3.3. Bacteria removal efficiency of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs

348 Many approaches have been undertaken in water safety.<sup>3c, 5c, 28</sup> Adsorbent materials,  
349 in particular, outperformed the conventional chemical flocculant precipitation and  
350 disinfection processes used in water treatment.<sup>29</sup> Magnetic nanoparticles have been  
351 regarded as a kind of novel adsorbent materials due to their ease of removal of target  
352 species from water with an external magnetic field, high surface area and efficient  
353 adsorption capacity. With special reference to bacterial adsorbent materials, the  
354 negatively charged molecules on the bacteria cell wall provide anionic sites for  
355 electrostatic binding of cationic molecules.<sup>30</sup> According to this principle, cationic  
356 magnetic adsorbent materials were fabricated with the purpose to effectively capture  
357 and remove bacteria in water systems.<sup>1, 31</sup>

358 In the present study, *E. coli* was taken as a model microorganism to determine the  
359 bacteria removal efficiency of the prepared cationic magnetic nanoparticles  
360 (Fe<sub>3</sub>O<sub>4</sub>@PPy NPs). **Figure 5A** shows the remaining *E. coli* after separation of the  
361 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs/*E. coli* complex with an external magnetic field, whereas the  
362 statistical analysis of the bacteria viability was displayed in **Figure 5C**. The decreased  
363 bacteria viability demonstrated the increased bacteria removal efficiency with the  
364 increase of the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs concentration. After 10 min of interaction and 10 min  
365 of magnetic adsorption, more than 50% *E. coli* was removed at a 100 µg/mL  
366 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs concentration. The *E. coli* removal efficiency was previously tested  
367 using magnetic graphene composites. Compared with the pure Fe<sub>3</sub>O<sub>4</sub> NPs, the magnetic  
368 graphene composites showed better removal performance against *E. coli*, which was  
369 mostly attributed to the much larger surface area of the graphene nanoparticles.  
370 However, it must be noted that the bacteria removal efficiency was still not effective,  
371 because more than 15 min were necessary to achieve an effective removal of the *E.*

372 *coli*.<sup>31b</sup> In our study, the simultaneous establishment of electrostatic interactions  
 373 between the cationic Fe<sub>3</sub>O<sub>4</sub>@PPy NPs and the peptidoglycans and proteins of the  
 374 bacteria cell membrane,<sup>30</sup> boosted the aggregation and complex formation between the  
 375 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs and *E. coli* cells. In turn, the complex can be easily separated using  
 376 an external magnetic field, thus enhancing the bacteria removal efficiency.<sup>32</sup> Therefore,  
 377 the cationic Fe<sub>3</sub>O<sub>4</sub>@PPy NPs, with the advantages of ease of handling and favorable  
 378 adsorption performance, can be taken as a promising bacteria adsorbent material.



379  
 380 **Figure 5.** Photographs of residual *E. coli* after magnetic adsorption treatment (A),  
 381 survival *E. coli* after photothermal treatment (B), and *E. coli* viability (C).

382

### 383 3.4. Photothermal sterilization property of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs

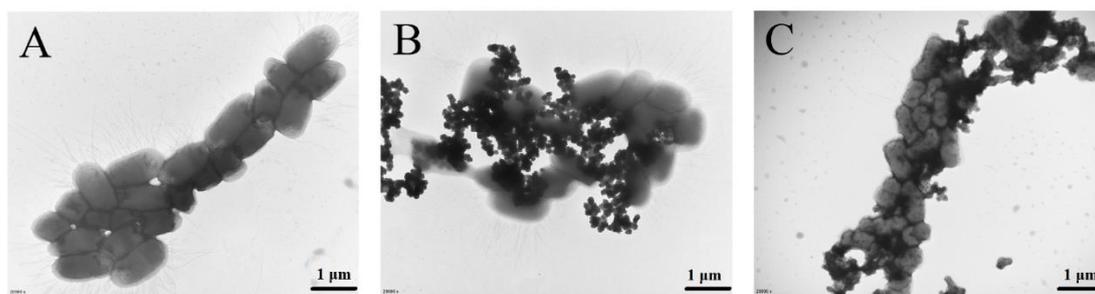
384 Based on our previous works, we took on enormous effort to expand the application  
 385 of the photothermal nanoparticles.<sup>12a, 33</sup> Due to the excellent photothermal performance

386 of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs, we further investigated their photothermal sterilization induced by  
387 NIR radiation of the electromagnetic spectrum. The survived *E. coli* strains after NIR  
388 irradiation is shown in **Figure 5B**, whereas the results in **Figure 5C** demonstrated that  
389 the photothermal sterilization performance of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs increased with  
390 increasing NPs concentration. After continuous irradiation for 10 min, 100% of *E. coli*  
391 strains were inactivated with a concentration of 100 µg/mL Fe<sub>3</sub>O<sub>4</sub>@PPy NPs. The  
392 sterilization mechanism of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was mostly due to the hyperthermia on  
393 bacteria. It has been reported that 55 °C was the threshold temperature, above which  
394 denaturation of proteins and synthesis of heat-shock proteins would occur. Long-time  
395 heat treatments would break the intracellular circulation of bacteria and lead to their  
396 death.<sup>34</sup> In the present work, after 10 min NIR irradiation, the temperature of 100 µg/mL  
397 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs rose to 60.1 °C, which was enough to inactivate the bacteria. Arising  
398 from these results, as expected Fe<sub>3</sub>O<sub>4</sub>@PPy NPs exhibited effective photothermal  
399 sterilization activity, which can be profitably exploited for the generation of a new class  
400 of photothermal sterilization agents against bacteria in aqueous medium.

401

### 402 **3.5. Morphology observation of *E. coli* and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs**

403 TEM images were used to acquire a deep understanding of the interaction between  
404 *E. coli* and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs upon magnetism, as well as the morphology changes of *E.*  
405 *coli* after NIR irradiation (**Figure 6**). *E. coli* cells without any treatment exhibited an  
406 intact and smooth morphology (**Figure 6A**). After adding the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs into the  
407 *E. coli* suspension, the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs adsorbed onto the *E. coli* surface (**Figure 6B**).  
408 The morphology of the *E. coli* was damaged severely after NIR irradiation, as  
409 confirmed by the shrinkage of the bacteria cytoplasm followed by rupture of the cell  
410 wall and membrane (**Figure 6C**).



411

412 **Figure 6.** TEM images of *E. coli* cells (A), *E. coli* cells and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs treated  
 413 with an external magnetic field (B) and *E. coli* cells and Fe<sub>3</sub>O<sub>4</sub>@PPy NPs treated with  
 414 NIR irradiation (C).

415

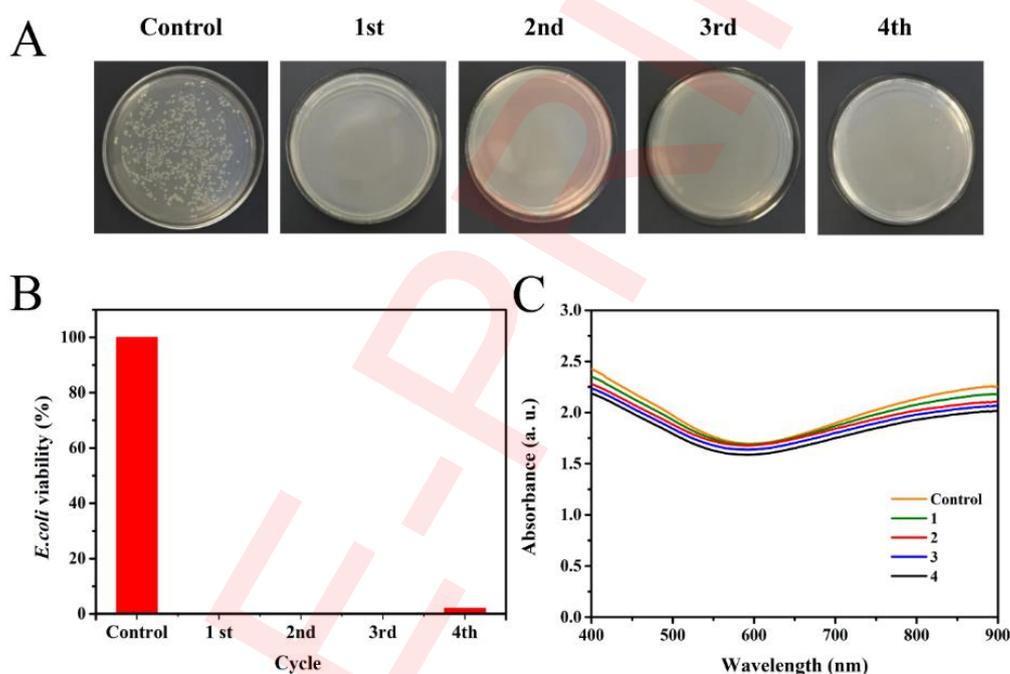
416 As explained before, the adsorption of the positively-charged NPs on the negatively-  
 417 charged bacteria were boosted by the cationic pyrrole polymer, which easily adsorbed  
 418 onto the surface of the *E. coli* membrane through electrostatic interactions. Based on  
 419 this interaction ability, the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs can be suggested as an adsorbent material  
 420 for bacteria removal upon application of an external magnetic field. In addition, the  
 421 prepared adsorbent materials can simultaneously absorb light energy and convert it into  
 422 thermal energy, hence playing a local heating effect. The photothermal effect of  
 423 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs led to an increase of the temperature of bacteria suspensions above  
 424 the threshold temperature of the bacteria survival. As a consequence, the bacteria  
 425 membrane was damaged, the intracellular components leaked out, the vital movements  
 426 were impaired, and the bacteria were eventually inactivated.<sup>35</sup> Under the synergistic  
 427 action of the adsorption and heat effect, the photothermal sterilization property of  
 428 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was made more effective.

429

### 430 3.6. Recyclability of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs for photothermal sterilization

431 The magnetic properties of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs allowed for their easy separation and

432 reutilization using an external magnetic field. The recyclability of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs for  
433 photothermal sterilization was determined by inactivation tests mediated by recycled  
434 NPs. As shown in **Figure 7A-7B**, over 99% of bacteria were inactivated using recycled  
435 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs for each cycle with the same photothermal procedure. The spectral  
436 absorption of the reused Fe<sub>3</sub>O<sub>4</sub>@PPy NPs also showed no significant variations after  
437 four cycles (**Figure 7C**). These results demonstrate the high stability and recyclability  
438 of the prepared Fe<sub>3</sub>O<sub>4</sub>@PPy NPs. Therefore, it can be further confirmed that the  
439 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs can be considered a reliable alternative in the photothermal  
440 sterilization of water.



441

442 **Figure 7.** Photographs of survival *E. coli* after photothermal sterilization with  
443 recycled (four cycles) Fe<sub>3</sub>O<sub>4</sub>@PPy NPs (A), *E. coli* viability for each cycle (B) and  
444 UV-vis spectra of recycled Fe<sub>3</sub>O<sub>4</sub>@PPy NPs after NIR irradiation and magnetic  
445 separation (C)

446

447

#### 448 **4. Conclusions**

449 In this work, we have successfully developed Fe<sub>3</sub>O<sub>4</sub>@PPy NPs for inactivation of  
450 bacteria suspended in aqueous media. The sterilization activity of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs was  
451 tested successfully against *E. coli* in a water system, and was explained in terms of: i)  
452 adsorption capability of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs mediated by electrostatic interaction between  
453 NPs and bacteria surface; and ii) high photothermal conversion efficiency of  
454 Fe<sub>3</sub>O<sub>4</sub>@PPy NPs and a local heating effect, due to their strong absorption from UV to  
455 NIR regions, thus making possible the conversion of large portion of solar energy to  
456 thermal energy, which can lead the bacteria suspension temperature above the threshold  
457 of the bacteria survival. In addition, the excellent photothermal stability and  
458 recyclability of Fe<sub>3</sub>O<sub>4</sub>@PPy NPs make them an appealing system for multiple uses  
459 without affecting the ultimate photothermal sterilization ability. We believe that the  
460 excellent performance of the Fe<sub>3</sub>O<sub>4</sub>@PPy NPs, based on the dual adsorption behavior  
461 and photothermal sterilization property, will provide a new pathway for the water safety  
462 management.

463

#### 464 **Conflicts of interest**

465 There are no conflicts to declare.

466

#### 467 **Acknowledgements**

468 The authors gratefully acknowledge the financial supports by The National Natural  
469 Science Foundation (81901868), the Fundamental Research Funds for the Central  
470 Universities (2572018BU03) and the financial support by National Key R&D Program  
471 of China (2017YFD0600706).

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