

52 **Abstract**

53 **Purpose:** The tests currently used for the identification of SARS-CoV-2 include specimens taken
54 from the upper and lower respiratory tract. Although recommendations from the World Health
55 Organization prioritise the usage of a nasopharyngeal swab (NS), nasopharyngeal aspirates (NPA)
56 are thought to be superior in identifying SARS-CoV-2 in children. To our knowledge, however, no
57 paediatric study has been published on the subject. The aim of this study is to evaluate the
58 diagnostic performances of NS referred to NPA for SARS-CoV-2 in children.

59 **Methods:** We calculated the sensitivity and specificity of the NS referred to the NPA of the whole
60 sample and considered both age and collection period as covariates in different analyses.

61 **Results:** We collected 300 paired samples. The NS had a specificity of 97.7% and a sensitivity of
62 58.1%. We found similar results for the group of subjects ≥ 6 years old, while for subjects < 6 years
63 old, the sensitivity was 66.7% and the specificity 97.8%. Considering period as a covariate, the
64 sensitivity and specificity for patients hospitalized in March (31 patients, 52 records) were 70.0%
65 and 97.6%, while for patients involved in the follow up (16 patients, 57 records) they were 57.2%
66 and 89.7%.

67 **Conclusions:** The NS has a low sensitivity in detecting SARS-CoV-2 in children when referred to
68 the NPA, whereas its specificity is high. Our results suggest that in children under 6 years of age
69 NSs should be preferred whenever possible. Though statistically not significant, the sensitivity of
70 the NS rises when performed before the NPA.

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73 **Keywords:** SARS-CoV-2, nasopharyngeal aspirate, nasopharyngeal swab, children.

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103 **INTRODUCTION**

104 In December 2019, the world witnessed the emergence of a novel coronavirus in Wuhan, China.
105 Later named severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the new coronavirus
106 is responsible for a respiratory disease now known as the Coronavirus Disease (COVID-19). After
107 spreading across the globe, the World Health Organization (WHO) declared COVID-19 an
108 international health emergency [1]. In February 2020, the outbreak also involved Italy, which
109 became one of the worst affected countries [2].

110 Cases reported in literature mainly concern adults and the mortality rate is higher in the elderly
111 and in subjects with chronic underlying diseases [3].

112 Data regarding infected children are so far limited. In a study reviewing 44,672 laboratory-
113 confirmed cases of COVID-19, Wu et al. reported that 1% of cases occurred in children from 10 to
114 19 years of age and another 1% in children of 9 years of age or younger, with no deaths reported in
115 the latter group [4]. Overall, children younger than 18 years of age appear to be less vulnerable to
116 the infection, to have milder symptoms, and a less severe disease course compared to adults [4-7]. A
117 Chinese observational study reported that, of the 1391 children younger than 16 years of age tested
118 for SARS-CoV-2, only 171 (8.1%) resulted positive. Among them, 15.8% were asymptomatic
119 while the rest showed only mild symptoms [8]. These observations are similar to what was reported
120 in a recent seroprevalence study which analysed data generated from the first lockdown in
121 Lombardy. More specifically, the study showed a linear increase in the log odds for IgG positivity
122 with age, ranging from 9.1% in 5-year-old children to 12.5% in 20-year-old individuals and ending
123 at around 40% for people over 80 [9].

124 The tests currently used for the direct identification of SARS-CoV-2 include specimens taken
125 from the upper (nasopharyngeal/oropharyngeal swab and nasopharyngeal aspirate) and the lower
126 respiratory tract (bronchoalveolar lavage, tracheal aspirate, sputum) [10-12].

127 Upper respiratory specimens are easily obtainable, require less invasive manoeuvres than lower
128 respiratory specimens, and their collection exposes healthcare workers to a lower risk of infection.
129 Asymptomatic children and patients with mild symptoms are therefore usually tested with this type
130 of sampling. The collection of lower respiratory specimens is instead reserved for symptomatic or
131 severe cases due to the high discomfort caused, the special devices, and skilled operators required to
132 obtain them [10].

133 Yang et al. demonstrated that SARS-CoV-2 isolation from the bronchoalveolar lavage fluid
134 (BALF) had a 100% positive rate when collected from severe cases, while isolation from sputum
135 resulted in the highest positive rate when collected from both severe and mild cases, followed by
136 nasal swab [13, 14]. Regarding samples collected from the upper airways, higher viral loads were
137 detected in the nose than in the throat; indeed, recommendations from WHO prioritize
138 nasopharyngeal swab over oropharyngeal swab [10, 15].

139 Zou et al. demonstrated that the viral load in symptomatic children is similar to that of
140 asymptomatic patients, which suggests a potential contagiousness of the latter [15]. Furthermore,
141 they detected higher viral loads in specimens collected soon after the symptom's onset which may
142 indicate a higher risk of transmission in the early stages of infection [10, 15].

143 Although a positive test is highly indicative of infection, a negative test does not rule it out [16].
144 Several factors may contribute to false-negative results including the sampling technique, the
145 transportation process, the potentially limited RNA found in the samples, and the molecular
146 structure making up the coronavirus (e.g. genetic mutations such as D614G acquired by SARS-
147 CoV-2 that naturally selected dominant lineages helping the virus to spread faster and acquire
148 higher levels of virulence) [17, 18]. Testing of specimens from multiple sites may help reduce false-
149 negative results [13].

150 To date, there are no paediatric studies on the identification of SARS-CoV-2 through
151 nasopharyngeal aspirate (NPA). Various studies on major respiratory viruses have shown that in
152 the paediatric population the sensitivity of nasal swabs (NS) is comparable to that of NPAs. There
153 have been some reports, however, that highlight a lower sensitivity of the former compared to the

154 latter for the detection of common viruses such as the Respiratory Syncytial Virus and the
155 Rhinovirus [19-21]. It is generally agreed that the NS is recommended in outpatient settings
156 because of its rapid and less traumatic collection and because it usually does not require any
157 training and additional devices. On the other hand, for hospitalized patients, who could receive
158 unnecessary antibiotic therapy or be subject to additional diagnostic procedures, the NPA is the test
159 of choice due to its higher sensitivity [22, 23]. For this reason, we considered the NPA as our
160 reference test.

161 Our study was aimed at evaluating if the NS could be used instead of the NPA in children. To do
162 this we calculated the concordance and the diagnostic performance of the NS compared the NPA's
163 in accordance to age and order in which the tests were administered.
164

165 **METHODS**

166 **Patients and samples**

167 From March 13th to May 22nd, all children who attended the emergency room and needed to be
168 hospitalized and those who were transferred to our paediatric unit from other wards/hospitals
169 underwent both NS and NPA, acquiring specimens from both nostrils. The tests were performed,
170 sequentially, on admission and after 24 hours by well-trained nurses or doctors. The nasopharyngeal
171 swabs were collected following the procedure published in the New England Journal of Medicine
172 [24], first from one nostril and then from the other, using the Copan-503CS01 nasopharyngeal
173 flocked swab. The nasopharyngeal aspirates were collected from both nostrils using a standard
174 protocol and the Medicoplast mucus extractor 440-ch08. In the laboratory, two assays were
175 performed for the detection of SARS-CoV-2. The Allplex™ 2019-nCoV Assay was used with the
176 Seegene NIMBUS & STARlet instrument, an in vitro diagnostic medical device designed for the
177 qualitative detection of the novel Coronavirus (2019-nCoV) by real-time reverse transcription-
178 polymerase chain reaction (RT-PCR). Starting from 300 µl of both NS and NPA samples, nucleic
179 acid extraction was performed using the STARMag 96 X 4 Universal Cartridge kit and 10 µl of RP-
180 V Internal Control (IC) which was added to each specimen before RNA extraction. The second
181 assay, GeneFinder COVID-19 Plus RealAmp Kit adapted to the ELITE InGenius®(ELITechGroup)
182 instrument is a qualitative one-step RT-PCR that used 200 µl of both samples for automatic and
183 integrated extraction. In this case, the IC was endogenous (RNase p).

184 For samples analysed with the GeneFinder COVID-19 Plus RealAmp Kit, we considered positive
185 Ct values ≤ 45 and negative Ct > 45 , while for the Allplex™ 2019-nCoV Assay positive findings
186 were considered when Ct values were ≤ 40 and negative when Ct values were > 40 . A weak
187 positivity was defined for Ct values of 40-45 or 37-40, depending on the method.

188 There was not a defined order for obtaining the specimens. More specifically, we performed first
189 the NS and then the NPA in March 2020, whereas we collected first the NPA and then the NS
190 during the follow-up. A total of 134 patients were included in the study. Thirteen among the latter
191 and two outpatient children were followed by collecting paired specimens until both came out
192 negative 24 hours apart. Thus 300 paired specimens (NS/NPA) were collected from 136 patients
193 (134 hospitalized and 2 outpatients) and were tested for SARS-CoV-2.
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195 **Statistical analyses**

196 All the statistical analyses were performed with R (v. 3.6.2) [25]. In order to estimate the
197 incidence of the positive cases on hospitalized patients, we calculated the proportion of the positive
198 cases (patients who had a positive result of NS or a positive result of NPA) in hospitalized patients
199 and its 95% confidence interval using the binomial distribution [26].

200 Analyses on diagnostic tests results (NS and NPA) concerned the concordance between the
201 results of the two tests. Furthermore, we calculated the mismatch for positive and negative values
202 and the sensitivity and specificity of NS (considering NPA as reference). In order to evaluate
203 whether sensitivity, specificity and mismatch of the NS were influenced by the patients' age, we
204 considered age as a covariate (coded as 0 from 0 to 5 years old and as 1 for more than 5 years old).

205 Since the order of collecting specimens was different between patients hospitalized in March, and
206 patient of the follow up, for the purpose of detecting any change in sensitivity, specificity and
207 mismatches according to the order of execution of the tests, we considered the period as covariate
208 (codified as 0 for the March period and as 1 for the Follow up).

209 Concerning the performance of the NS (considering NPA as reference test), descriptive summary
210 measures were also calculated: likelihood ratio positive (LR+) and negative (LR-). Likelihood ratios
211 compare the probability that a patient with positive NPA at the date of the test has a particular NS
212 test result as compared to someone with NPA negative. LR+ is the ratio between the probability of
213 a true positive result on the probability of false positive result. LR- is the ratio between the
214 probability of a false negative result on the probability of a true negative result [27].

215 Tests with very high LR+ and very low LR- have greater discriminating ability: in particular tests
216 with LRs >10 or <0.1 are very useful in establishing or excluding a diagnosis [28].

217 The analyses were performed by generalized estimating equation models (GEE) with family
218 binomial to take account of the correlation among diagnostic tests on the same patients [29]. To
219 estimate the percentage of concordance, the model response was coded as 1 if the results of the two
220 diagnostic tests agreed and as 0 otherwise. To estimate the sensibility and the mismatch for positive
221 values only the records with a positive result of the NPA were used, while to estimate specificity
222 and negative mismatch only the records with a negative result of the NPA were used. In both cases,
223 the model response was the result of the NS (coded as 0 if it was negative and as 1 if it was
224 positive) [30].

225 The influence of age and test period were estimated using the Wald test on the respective model
226 coefficients with a 5% significance level (two tailed test).

227 Due to the absence of a reliable prevalence value for COVID-19 in children, we could not
228 calculate the positive and negative predictive values (VPP, VPN).

229

230 **RESULTS**

231 For this study's purposes, we considered positive to SARS-CoV-2 every patient whose NPA or
232 NS or NPA/NS resulted positive or weak positive.

233 Out of the 134 patients hospitalized, 18 children tested positive (prevalence 13.4%, 95% CI:
234 8.2%-20.4%). Among the latter, 13 of them and 2 outpatient children were followed by collecting
235 paired specimens until both resulted negative 24 hours apart.

236 We collected 600 samples in total (equal to 300 paired): 43 positive NPA, 31 positive NS, 257
237 negative NPA, and 269 negative NS.

238 Of the 300 paired specimens evaluated: 276 were concordant, 24 were discordant, so the naïve
239 concordance was 92.0% (95% CI: 88.3%-94.6%).

240 The mismatch negative NS and positive NPA was greater than the mismatch positive NS and
241 negative NPA (about 42% and about 2% respectively), see Table 1.

242 The NS's specificity was greater than its sensitivity, suggesting the NS test was more suitable to
243 rule-in positive NPA patients than in ruling-out negative NPA patients (sensitivity was about 58%
244 and specificity was about 98%). The LR+ was 25.3 and LR- was 0.43, which means that a patient
245 with a positive result from an NPA is 25.3 times more likely to have a positive result from an NS
246 than someone with a negative result from an NPA and that a patient with a positive NPA is 0.43
247 times as likely to have a negative NS than someone with a negative NPA (or that a patient with a
248 negative NPA is about 2 times more likely to have a negative NS than someone with a positive
249 NPA). Considering the high value of the LR+ (greater than 10), the NS is expected to be useful in
250 establishing the positivity of SARS-CoV-2; however, the NS is probably not very useful in
251 excluding the infection, as shown by the higher than 0.1 LR-.

252 Considering age as a covariate, its effect on the above-mentioned measures was not statistically
253 significant for all models at the 5% significance level. More specifically, considering the mismatch
254 between positive NPAs and negative NSs we obtained a Z value (Wald statistic) of 0.493 ($p =$
255 0.483), for the mismatch between negative NPAs and positive NSs we obtained a Z value of 0.03 (p

256 = 0.87) while considering sensitivity and specificity we found Z values of 0.49 ($p = 0.48$) and 0.03
257 ($p = 0.87$), respectively.

258 Concerning sensitivity, specificity, LR+ (21.9), and LR- (0.46), results regarding subjects ≥ 6
259 years old were similar to those for “all records” (Table 1).

260 The mismatch between negative NSs and positive NPAs was smaller for subjects < 6 years old
261 (about 33%) than for all subjects and subjects ≥ 6 years old, while the mismatch between positive
262 NSs and negative NPAs was similar (about 2%) among the three groups (Table 1).

263 The specificity of the NS for this age group was greater than its sensitivity, suggesting the test
264 was more suitable to detect positive NPA patients than negative NPA patients (the test had a
265 sensitivity of about 67% and a specificity of about 98%). In addition, the sensitivity was greater
266 than the sensitivity calculated for all subjects and for subjects ≥ 6 years old (Table 1). The LR+ was
267 30.3 and LR- was 0.34.

268 As described above, we considered patients hospitalized in March (31 patients and 52 records)
269 and patients involved in follow up (16 patients and 57 records).

270 Regarding hospitalized children in March, we retrieved 52 paired specimens: 10 positive NPAs, 8
271 positive NSs, 42 negative NPAs, 44 negative NSs. Of the 52 paired specimens evaluated, 48 were
272 concordant and 4 were discordant; thus, the naïve concordance was 92.3% (95% CI: 81.7% -
273 97.0%). The results for these records showed that the smallest mismatch was between negative NSs
274 and positive NPAs (30%). The mismatch between positive NSs and negative NPAs was similar to
275 the previously presented results (about 2%). While we found a similar specificity to that of our
276 other results, the sensitivity was higher (70%), see Table 1. The LR+ was 29.2 and the LR- was
277 0.31.

278 Concerning the follow-up samples, there were 57 paired specimens: 28 positive NPAs, 19
279 positive NSs, 29 negative NPAs, and 38 negative NSs. Of the 57 paired specimens evaluated, 42
280 were concordant and 15 were discordant; thus, the naïve concordance was 73.7 % (95% CI: 61.8% -
281 82.9%). In this case, the mismatch between negative NSs and positive NPAs was similar to the
282 analyses on all subjects and on subjects ≥ 6 years old (about 43%); the mismatch, however, between
283 positive NSs and negative NPAs was greater than all previous results (about 10%). Here, the
284 specificity was greater than the sensitivity (about 90% for the specificity and about 57% for the
285 sensitivity), although the former was also the lowest among all previous analyses (Table 1). The
286 LR+ was 5.6 and LR- was 0.48.

287 The impact of period on the above-mentioned measures was not statistically significant at a 5%
288 significance level for all models. Especially for the mismatch between positive NPAs and negative
289 NSs we obtained a Z value (Wald statistic) of 0.520 ($p = 0.471$) and for the mismatch between
290 negative NPAs and positive NSs we had a Z value of 1.8 ($p = 0.180$). The Z values for sensitivity
291 and specificity were respectively of 0.52 ($p = 0.47$) and 1.8 ($p = 0.180$).

292

293 **DISCUSSION**

294 According to our results, the NS has a low sensitivity in the detection of SARS-CoV-2 in
295 children. At the same time, the NS has both a high specificity and a high LR+, which means that a
296 positive NS has a good reliability in detecting who has a positive NPA.

297 Despite the influence of age and collection period resulted not statistically significant, the
298 difference between the results of the two age groups and of the two collection periods could suggest
299 a potential impact of the two factors, which should be evaluated in larger case series.

300 Regarding the order with which the tests were obtained, the sensitivity and specificity in
301 hospitalized children in March were higher than those of follow up patients. This may imply that
302 executing an NS before an NPA may result in a greater probability of identifying SARS-CoV-2.
303 This may be explained by the fact that the NPA, through the thin catheter of the mucus extractor, is
304 able to collect a larger amount of secretions from the lower parts of the upper respiratory tract and
305 number of deeper cells compared to the NS [31, 32], which may lead to reduced or inadequate
306 samples to identify the virus when the NS is used after the NPA. Our results were not statistically

307 significant probably due to the small and non-uniform sample examined, so further studies,
308 involving a larger sample, are necessary to strengthen this evidence.

309 Concerning age, the sensitivity of the NS was highest in the group of children younger than 6
310 years. This means that a negative NS has a good reliability in detecting the patient who has a
311 negative NPA. Moreover, patients younger than 6 years have the highest LR+, while the specificity
312 is similar to that found in the other analyzed groups. The NS, therefore, is suitable to identify
313 children younger than 6 years with a positive NPA. According to these results, performing NSs in
314 this age group is better for identifying a SARS-CoV-2 infection. Although our results point towards
315 a better identification of the SARS-CoV-2 infection using the NS in children under 6 years, in our
316 experience, [21, 32], performing an NPA in young children is simpler than performing an NS: the
317 aspiration of mucus from the nasopharynx using the small catheter resulted less unpleasant
318 compared to brushing against the nasopharyngeal wall using the NS. At the same time, the NPA
319 requires a catheter, an aspiration trap, a vacuum source, and specialized training, which are only
320 available in a hospital setting. On the other hand, for the NS no additional training or devices are
321 needed.

322 Our study has some limitations. Firstly, the different order with which the specimens were
323 obtained, which implies that the data are not uniform. Secondly, the small sample. Another
324 limitation is the lack of data regarding signs and symptoms of patients who underwent NPA and NS
325 and the resulting inability to describe a correlation between the isolation of SARS-CoV-2 and
326 clinical features. Finally, to our knowledge, an analysis on the relationship between the viral load
327 and the infectivity has not yet been reported. It has been, however, demonstrated that the
328 identification of the virus on a specimen does not necessarily correlate with infectivity and there are
329 indeed multiple reports which attest a prolonged viral shedding after symptoms resolution in
330 COVID-19 [33]. In our department, among children whose tests were positive, three continued to
331 be positive to either NS or NPA or both for the following 9 weeks. These findings are in line with
332 several other studies that demonstrated the prolonged viral shedding of children [34, 35].

333

334 **CONCLUSIONS**

335 The NS has a low sensitivity in detecting SARS-CoV-2 in children when referred to NPA, both in
336 the overall analysis and in that according to age. Its specificity on the other hand is high. This
337 means that a positive NS can be reliable, but that a negative NS cannot rule out the presence of
338 SARS-CoV-2 since the proportion of false negatives is substantial.

339 Though statistically not significant, we found that when the NS is performed before the NPA its
340 sensitivity rises, which may be due to the fact that NSs performed before NPAs are richer in
341 secretion, cells, and therefore viruses.

342 Although statistical significance was not reached, our results suggest that the use of the NS for
343 the detection of SARS-CoV-2 should be preferred whenever possible in children younger than 6
344 years, thanks to its high LR+.

345 As far as we know, this is the first study dealing with diagnostic performance of NS referred to
346 NPA for detecting SARS-CoV-2 in children. Further analyses are mandatory to transfer these
347 findings to our clinical practice.

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364 **Authors' contributions:** Prof Marchisio, Castaldi and Biganzoli conceptualized and designed the
365 study, reviewed and revised the manuscript. Drs Di Pietro and Capecchi collected data, wrote and
366 reviewed the manuscript. Drs Biganzoli, Luconi, Marano and Boracchi analyzed the statistical data.
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409 **REFERENCES**

- 410
- 411 1. World Health Organization. Coronavirus disease 2019 (COVID-19). Situation Report 51.
412 Published March 11, 2020. Accessed July 27, 2020. [https://www.who.int/docs/default-](https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200311-sitrep-51-covid-19.pdf?sfvrsn=1ba62e57_10)
413 [source/coronaviruse/situation-reports/20200311-sitrep-51-covid-](https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200311-sitrep-51-covid-19.pdf?sfvrsn=1ba62e57_10)
414 [19.pdf?sfvrsn=1ba62e57_10](https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200311-sitrep-51-covid-19.pdf?sfvrsn=1ba62e57_10).
- 415 2. Epicentro. Istituto Superiore della sanità. Dati della Sorveglianza integrata COVID-19 in
416 Italia. Published February 27, 2020. Accessed July 27, 2020.
417 <https://www.epicentro.iss.it/coronavirus/sars-cov-2-dashboard>
- 418 3. Wang L, Wang Y, Ye D, Liu Q (2020) Review of the 2019 novel coronavirus (SARS-CoV-
419 2) based on current evidence. *Int J Antimicrob Agents* 55:105948.
- 420 4. Wu Z, McGoogan JM (2020) Characteristics of and important lessons from the coronavirus
421 disease 2019 (COVID-19) outbreak in China: summary of a report of 72 314 cases from the
422 Chinese Center for Disease Control and Prevention. *JAMA* 323:1239-1242.
- 423 5. Ludvigsson JF (2020) Systematic Review of COVID-19 in Children Shows Milder Cases
424 and a Better Prognosis Than Adults. *Acta Paediatr* 109:1088-1095.
- 425 6. Liu W, Zhang Q, Chen J, Xiang R, Song H, Shu S (2020) Detection of Covid-19
426 in Children in Early January 2020 in Wuhan, China. *N Engl J Med* 382:1370-1371.
- 427 7. Brodin P (2020) Why is COVID-19 so mild in children? *Acta Paediatr* 109:1082-1083.
- 428 8. Lu X, Zhang L, Du H, Zhang J, Li YY, Qu J, Zhang W, Wang Y, Bao S, Li Y, Wu C, Liu
429 H, Liu D, Shao J, Peng X, Yang Y, Liu Z, Xiang Y, Zhang F, Silva RM, Pinkerton KE,
430 Shen K, Xiao H, Xu S, Wong GWK; Chinese Pediatric Novel Coronavirus Study Team
431 (2020) SARS-CoV-2 infection in children. *N Engl J Med* 382:1663-1665.
- 432 9. Pagani G, Conti F, Giacomelli A, Bernacchia D, Rondanin R, Prina A, Scolari V, Gandolfi
433 CE, Castaldi S, Marano G, Ottomano C, Boracchi P, Biganzoli EM, Galli M (2020)
434 Seroprevalence of SARS-CoV-2 IgG significantly varies with age: results from a mass
435 population screening (SARS-2-SCREEN-CdA). *MedRxiv*.
436 <https://doi.org/10.1101/2020.06.24.20138875>
- 437 10. Mawaddah A, Gendeh HS, Lum SG, Marina MB (2020) Upper Respiratory Tract Sampling
438 in COVID-19. *Malays J Pathol* 42:23-35.
- 439 11. Loeffelholz MJ, Tang YW (2020) Laboratory Diagnosis of Emerging Human Coronavirus
440 Infections - The State of the Art. *Emerg Microbes Infect* 9:747-756.
- 441 12. Hanson KE, Caliendo AM, Arias CA, Englund JA, Lee MJ, Loeb M, Patel R, El Alayli A,
442 Kalot MA, Falck-Ytter Y, Lavergne V, Morgan RL, Murad MH, Sultan S, Bhimraj A,
443 Mustafa RA (2020) Infectious Diseases Society of America Guidelines on the Diagnosis of
444 COVID-19. *Clin Infect Dis* ciaa760.
- 445 13. Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W (2020) Detection of SARS-CoV-2 in
446 Different Types of Clinical Specimens. *JAMA* 323:1843-4.
- 447 14. Yang Y, Yang M, Shen C (2020) Evaluating the accuracy of different respiratory specimens
448 in the laboratory diagnosis and monitoring the viral shedding of 2019-nCoV infections.
449 *MedRxiv*. <https://doi.org/10.1101/2020.02.11.20021493>.
- 450 15. Zou L, Ruan F, Huang M, Liang L, Huang H, Hong Z, Yu J, Kang M, Song Y, Xia J, Guo
451 Q, Song T, He J, Yen HL, Peiris M, Wu J (2020) SARS-CoV-2 Viral Load in Upper
452 Respiratory Specimens of Infected Patients. *N Engl J Med* 382:1177-1179.
- 453 16. Zitek T (2020) The Appropriate Use of Testing for COVID-19. *West J Emerg Med* 21:470-
454 472.
- 455 17. Winichakoon P, Chaiwarith R, Liwsrisakun C, Salee P, Goonna A, Limsukon A,
456 Kaewpoowat Q (2020) Negative Nasopharyngeal and Oropharyngeal Swabs Do Not Rule
457 Out COVID-19. *J Clin Microbiol* 58:e00297-20.
- 458 18. Callaway E (2020) The coronavirus is mutating - does it matter? *Nature* 585(7824):174-177.

- 459 19. Sung RY, Chan PK, Choi KC, Yeung AC, Li AM, Tang JW, Ip M, Tsen T, Nelson EA
460 (2008) Comparative Study of Nasopharyngeal Aspirate and Nasal Swab Specimens for
461 Diagnosis of Acute Viral Respiratory Infection. *J Clin Microbiol* 46(9):3073-6.
- 462 20. Meerhoff TJ, Houben ML, Coenjaerts FE, Kimpen JL, Hofland RW, Schellevis F, Bont LJ
463 (2010) Detection of Multiple Respiratory Pathogens During Primary Respiratory Infection:
464 Nasal Swab Versus Nasopharyngeal Aspirate Using Real-Time Polymerase Chain Reaction.
465 *Eur J Clin Microbiol Infect Dis* 29:365-71.
- 466 21. Faden H (2010) Comparison of Midturbinate Flocked-Swab Specimens With
467 Nasopharyngeal Aspirates for Detection of Respiratory Viruses in Children by the Direct
468 Fluorescent Antibody Technique. *J Clin Microbiol* 48:3742-3.
- 469 22. Macfarlane P, Denham J, Assous J, Hughes C (2005) RSV Testing in Bronchiolitis: Which
470 Nasal Sampling Method Is Best? *Arch Dis Child* 90:634-5.
- 471 23. Lambert SB, Whiley DM, O'Neill NT, Andrews EC, Canavan FM, Bletchly C, Siebert DJ,
472 Sloots TP, Nissen MD (2008) Comparing Nose-Throat Swabs and Nasopharyngeal
473 Aspirates Collected From Children With Symptoms for Respiratory Virus Identification
474 Using Real-Time Polymerase Chain Reaction. *Pediatrics* 122:e615-20.
- 475 24. Marty FM, Chen K, Verrill KA (2020) How to Obtain a Nasopharyngeal Swab Specimen. *N*
476 *Engl J Med* 382:e76.
- 477 25. R Development Core Team, R (2011) R: A language and environment for statistical
478 computing. R Foundation for Statistical Computing. Vienna, Austria. [https://www.r-](https://www.r-project.org/)
479 [project.org/](https://www.r-project.org/)
- 480 26. Clopper CJ, Pearson ES (1934) The use of confidence or fiducial limits illustrated in the
481 case of the binomial. *Biometrika* 26:404–413.
- 482 27. Ranganathan P, Aggarwal R (2018) Understanding the properties of diagnostic tests-Part 2:
483 Likelihood ratios. *Perspect Clin Res* 9: 99–102.
- 484 28. Deeks JJ, Altman DG (2004) Diagnostic tests 4: Likelihood ratios. *BMJ* 329:168–9.
- 485 29. Liang KY, Zeger SL (1984) Longitudinal data analysis using generalized linear models.
486 *Biometrika* 73:13-22.
- 487 30. Leisenring W, Sullivan Pepe M, Longton G (1997) A marginal regression modelling
488 framework for evaluating medical diagnostic tests. *Statistics in medicine* 1263-1281.
- 489 31. Chan KH, Peiris JSM, Lim W, Nicholls JM, Chiu SS (2008) Comparison of Nasopharyngeal
490 Flocked Swabs and Aspirates for Rapid Diagnosis of Respiratory Viruses in Children. *J Clin*
491 *Virol* 42:65-9.
- 492 32. Abu-Diab A, Azzeh M, Ghneim R, Ghneim R, Zoughbi M, Turkuman S, Rishmawi N, Issa
493 AE, Siriani I, Dauodi R, Kattan R, Hindiyyeh M (2008) Comparison Between Pernal
494 Flocked Swabs and Nasopharyngeal Aspirates for Detection of Common Respiratory
495 Viruses in Samples From Children. *J Clin Microbiol* 46:2414-7.
- 496 33. Widders A, Broom A, Broom J (2020) SARS-CoV-2: The viral shedding vs infectivity
497 dilemma. *Infect Dis Health* S2468-0451(20)30028-6.
- 498 34. Liu P, Cai J, Jia R, Xia S, Wang X, Cao L, Zeng M, Xu J (2020) Dynamic surveillance of
499 SARS-CoV-2 shedding and neutralizing antibody in children with COVID-19. *Emerg*
500 *Microbes Infect* 9:1254-1258.
- 501 35. Lu Y, Li Y, Deng W, Liu M, He Y, Huang L, Lv M, Li J, Du H (2020) Symptomatic
502 Infection Is Associated With Prolonged Duration of Viral Shedding in Mild Coronavirus
503 Disease 2019: A Retrospective Study of 110 Children in Wuhan. *Pediatr Infect Dis J*
504 39:e95-e99.