This is a pre-copyedited, author-produced version of an article accepted for publication in [insert journal title] following peer review. The version of record Cirilli et al. The Multisite PeachRefPop Collection: A True Cultural Heritage and International Scientific Tool for Fruit Trees, Plant Physiology, Volume 184, Issue 2, October 2020, Pages 632–646, is available online at: https://doi.org/10.1104/pp.19.01412

Short title: Multi-site PeachRefPop collection

The multi-site *PeachRefPop* collection: a true cultural heritage and international scientific tool for fruit trees

List of Authors

Marco Cirilli^{1*}, Sabrina Micali^{2*}, Maria José Aranzana³, Pere Arús³, Annarosa Babini⁴, Teresa Barreneche⁵, Marco Bink⁶, Celia M. Cantin³, Angelo Ciacciulli¹, José Enrique Cos-Terrer⁷, Pavlina Drogoudi⁸, Iban Eduardo³, Stefano Foschi⁹, Daniela Giovannini¹⁰, Walter Guerra¹¹, Alessandro Liverani¹⁰, Igor Pacheco¹², Thierry Pascal¹³, Benedicte Quilot-Turion¹³, Ignazio Verde², Laura Rossini^{1a} and Daniele Bassi^{1a}

*equal contribution

^aJoint senior authors

Contact author: Laura Rossini, e-mail: laura.rossini@unimi.it

Affiliations

¹Department of Agricultural and Environmental Sciences (DISAA), University of Milan, Via Celoria 2, Milan, Italy;

²CREA, Research Centre for Olive, Fruit, and Citrus Crops, Via di Fioranello, 52, Rome, Italy;

³IRTA, Centre de Recerca en Agrigenòmica CSIC-IRTA-UAB-UB (CRAG), Campus UAB, Cerdanyola del Vallès (Bellaterra), 08193 Barcelona, Spain;

⁴Phytosanitary Service, Regione Emilia-Romagna, Bologna, Italy;

⁵Université de Bordeaux, INRAE, BFP, F-33140 Villenave d'Ornon, France

⁶Hendrix Genetics Research, Technology & Services B.V., PO Box 114, 5830 AC Boxmeer, Netherlands;

⁷Murcia Institute of Agri-Food Research and Development (IMIDA), La Alberca, Murcia, Spain;

⁸Hellenic Agricultural Organization 'Demeter', Department of Deciduous Fruit Trees, Institute of Plant Breeding and Genetic Resources, Naoussa, Greece;

⁹Centro Ricerche Produzioni Vegetali, Cesena (FC), Italy;

¹⁰CREA, Research Centre for Olive, Fruit, and Citrus Crops, Via Ia Canapona 1 bis, Forlì, Italy;

¹¹Research Centre Laimburg, 39040 Vadena, Italy;

¹²INTA, Universidad de Chile, El Líbano 5524, Macul, Chile;

¹³INRAE, GAFL, F-84143, Montfavet, France.

One sentence summary: Realization of the *PeachRefPop*, the international multi-site reference collection in peach, provides an invaluable tool for scientific studies in perennial species.

Author contributions

Plant material: DB, SF, BQ, TB, TP, PA, MJA, IV, AL, DG; host institutions: DB, MC, SF, LR, PA, IE, JT, PD, IV; phytosanitary controls: AB; site characterization: WG, MC; experimental design: MCAMB; phenotypic data analysis: AL, DG, CC; genotypic data analysis: IP, AC, MT, MJA, SM; core collection design and evaluation: SM, IV, MC; manuscript writing: MC, SM; manuscript revision: DB, LR, IV, SM and all authors for respective parts. Conception and coordination: LR and DB. LR and DB share senior authorship and agree to serve as the authors responsible for contacts.

Abstract

Plants have evolved a range of adaptive mechanisms that adjust their development and physiology to variable external conditions, particularly in perennial species subjected to long-term interplay with the environment. Exploiting the allelic diversity within available germplasm and leveraging the knowledge of the mechanisms regulating genotype interaction with the environment are crucial to address climatic challenges and assist the breeding of novel cultivars with improved resilience. The development of multisite collections is of utmost importance for the conservation and utilization of genetic materials and will greatly facilitate the dissection of genotype-by-environment interaction. Such resources are still lacking for perennial trees, especially with the intrinsic difficulties of successful propagation, material exchange and living collection maintenance. This work describes the concept, design and realization of the first multi-site peach [*Prunus persica* (L.) Batsch] reference collection (*PeachRefPop: PRP*) located across different European countries and sharing the same experimental design. Other than an invaluable tool for scientific studies in perennial species, the *PRP* provides a milestone in an international collaborative project for the conservation and exploitation of European peach germplasm resources and, ultimately, as a true heritage for future generations.

1 Introduction

Since the Roman garden '*hortus*', fruit tree orchards have represented distinctive features of the Mediterranean rural landscape, a synthesis of the interaction among genotype, environment and human customs (**Biasi et al., 2009**). The diversity of pedo-climatic conditions and production systems, along with plasticity of the genotype and human traditions has shaped the selection of a multitude of local cultivars. These materials represent a cultural and genetic heritage of generations of farmers and a 'common good' to preserve for present and future generations.

8 Plants have evolved a range of adaptive mechanisms that adjust their development and 9 physiology to variable external conditions, particularly in perennial species subjected to a long-term 10 environmental exposure and interaction. Climate changes are impacting cultivation environments, raising the need for more resilient cultivars able to maintain performances across variable (and 11 12 often unpredictable) weather conditions (Varshney et al., 2011; Luedeling, 2012; Ramírez and 13 Kallarackal, 2015). Also, increasing the sustainability of fruit production (particularly in terms of 14 resource demands and disease management) requires leveraging knowledge of the interactions between plants, soil, and environmental factors and how they affect productivity and end-product 15 16 quality (Coakley et al., 1999; Singh et al., 2013; Parajuli et al., 2018).

Peach [(*Prunus persica* L. (Batsch)] originated in China (**Li et al., 2019**), later reaching Persia, the Mediterranean Basin, Europe and the Americas, is now the third most cultivated fruit tree species in temperate regions. Beside its importance as a crop, peach is a recognized model for genetic and genomic studies in fruit trees, representing the ideal system for addressing two main challenges in fruit tree breeding:

22 1) understanding and harnessing the allelic diversity within available genepools; noteworthy 23 for peach, the intercompatibility with related species of the Amygdalus subgenus [almond 24 (P. dulcis), P. davidiana, P. kansuensis, etc.] has long been considered a source of natural variability, particularly for the introgression of disease resistances (Gradziel, 25 2002; Foulongne et al., 2003). However, interspecific hybrids have had poor 26 27 applicability in current breeding programs (Cirilli et al., 2017), although new genomics-28 based strategies could change this trend (Serra et al., 2016). Conversely, landraces and local ecotypes could be a source of resilience traits more straightforward to introgress, 29 30 making their preservation and exploitation a suitable strategy for dealing with the 31 changing climatic conditions.

2) systematic dissection of genotype-by-environment (G × E) and/or by-management (G × E
× M) interactions as primary sources of variability for several important quantitative traits
(Bassi et al., 2006; Myles, 2013; Chagné et al., 2014). This is a critical point for genetic
analyses of complex traits, such as genome-wide association studies (GWAS) where
germplasm collections are characterized to identify quantitative trait loci (QTLs) across
different environments, or genome-wide selection (GS), used to predict genomic
estimated breeding values.

The comprehension of genetic, epigenetic and physiological mechanisms as well as the estimation of G × E and/or G × E × M effects requires the development of multisite replicated collections and *ad hoc* experimental designs. The availability of such type of resources is rapidly growing in annual species, while it has not yet been implemented in perennial fruit trees.

During the past century, peach orchard systems have changed dramatically following innovations in orchard design, training systems and agronomic management (**Corelli-Grappadelli**

and Marini, 2008), other than cultivar evolution. Noteworthy, the first reported 'modern' orchard 45 was a peach plantation established in Massa Lombarda (Ravenna, Italy) at the end of the 19th 46 47 century using the white fleshed local cultivar-population 'Buco Incavato' (Bellucci, 1908). In the last decades, considerable breeding efforts have assisted the intensification of cultivation 48 49 techniques and the development of horticultural quality concepts with the introduction of novel, fit-50 for-purpose cultivars (Byrne et al., 2009). In Europe, peach has a long cultivation history, tracing 51 back to the Ancient and Middle ages and characterized by the isolation and propagation of chance seedlings operated by farmers and amateurs, through which each country has set its own pool of 52 53 locally adapted cultivars (Bassi and Layne, 2009). The paradigm shift to the modern controlled-54 crosses approach in early US breeding programs has been the foundation of the dramatic varietal 55 improvement of the last century, beginning with the introduction of seedling materials from China in the mid-19th century (e.g. 'Chinese Cling', progenitor of most modern cultivars) (**Faust and Timon,** 56 **1995**; Byrne et al., 2009). The worldwide spread of improved US materials, favored also by the 57 limited activities in other countries, has resulted in a rapid replacement of landraces and local 58 accessions, particularly in Europe. From the second half of the 20th century, however, novel 59 programs started in several European countries, although they were mostly based on US breeding 60 stocks with a marginal role for local cultivated germplasm. This led to a consequent loss of many 61 local cultivars, in parallel with a progressive narrowing of the genetic bases in modern cultivars 62 (Aranzana et al., 2010; Verde et al., 2013). 63

64 As awareness of genetic erosion in modern plant breeding increased (Fu and Dong, 2015) 65 the conservation and exploitation of genetic resources has become a fundamental aspect in crop breeding (Ford-Lloyd and Jackson, 1986). Considerable efforts have been made in the collection 66 67 and characterization of many plant germplasms (including fruit tree species), along with the 68 development of approaches for their effective management and utilization (Gepts, 2006). The 69 concept of 'core collection', a subset of a germplasm collection of a species that captures most of 70 the genetic diversity while reducing redundancy, has represented an ideal solution for reducing 71 costs and increasing the efficiency of conservation programs (Frankel and Brown, 1984). Several 72 allocation methods have been developed for selecting core collections, attempting to maximize 73 allelic richness or allele coverage (MSTRAT, PowerCore, GenoCore), minimize or maximize 74 genetic distance (GDOpt, SimEli) or simultaneously accommodating for multiple criteria (Core 75 Hunter) (Gouesnard et al., 2001, Kim et al., 2003; Thachuk et al., 2009; Odong et al., 2011; 76 Krishnan et al., 2014). However, the effectiveness of the sampling strategies varied depending on 77 the objective of the core collection, the statistical approach for its definition and the measures for 78 evaluating its guality (Odong et al., 2013). Furthermore, beyond statistical considerations, other 79 aspects are often considered by the institutions hosting the collection, such as historical and socio-80 economic importance, relevance for breeding activities, popularity among growers and consumers, 81 or distinctive phenotypic characteristics.

82 In peach, the absence of wild or feral populations makes ex situ collections the main 83 valuable reservoirs of allelic variability for many traits not yet exploited in current breeding 84 programs. Remarkable progress has been achieved in the phenotypic and genotypic characterization of peach genetic resources (Badenes et al., 2015; Cirilli et al., 2018; Yu et al., 85 86 **2018**), taking advantage of genome sequencing and the development of cutting-edge genotyping 87 tools (Verde et al., 2012; Verde et al., 2017, Aranzana et al., 2019). In the framework of the 88 European collaborative project FruitBreedomics (Laurens et al., 2018), a coordinated 89 characterization of peach collections has been accomplished across relevant European 90 repositories (Micheletti et al., 2015; Hernandez-Mora et al., 2017), promoting increased 91 utilization of resources and encouraging the sharing of conservation responsibilities. For example, the Prunus Working Group within the Fruit Network in the European Cooperative Programme on 92 93 Plant Genetic Resources (ECPGR) is dealing with Prunus, including peach (Benediková and 94 Giovannini, 2013). Nevertheless, long-term maintenance of collections remains particularly challenging due to intrinsic vulnerabilities (e.g. direct exposure to environmental variables and 95 pathogens) and costs for in vivo maintenance through vegetative propagation to preserve the 96 97 original genotypes. Moreover, compliance to phytosanitary requirements hampers the sharing of 98 resources among institutions, each having its own stock of materials, resulting in redundancies or 99 risk of loss for unique accessions.

100 This article describes the concept, design and realization of the first multi-site peach 101 reference collection (named as *PeachRefPop: PRP*) across five locations in three European 102 countries (Italy, Spain and Greece). Other than an invaluable tool for scientific studies, the *PRP* 103 provides a milestone of an international collaborative project for the conservation and exploitation 104 of European peach germplasms resources and, ultimately, as a true heritage for future 105 generations.

106

108 **Results**

109 **1.** Criteria for construction of a reference panel of peach accessions and seedlings

The *PRP* collection was built with the aim of selecting a reduced germplasm pool, reflecting the original genetic and phenotypic diversity (**Figure 1**) and the cultural and socio-economic value of peach cultivation, for its exploitation in future breeding programs. A four-step procedure was

- 113 followed (exemplified in **Figure 2**):
- 114 1. Definition of the *PeachRefPop* domain
- 115 2. Establishment of *PeachRefPop* size
- 116 3. Identification of the selection criteria
- 117 4. Choice and allocation of the entries
- 118

119 **1.1 Definition of the** *PeachRefPop* domain

120 To build a resource representing peach diversity and breeding history, the starting point was the 121 genetic material characterized in the framework of the FP7 European project FruitBreedomics 122 (http://fruitbreedomics.com/) in a coordinated effort involving different universities and research 123 institutions across Europe and China. A total of 1,580 Prunus accessions (comprising P. persica 124 and its hybrids with *P. davidiana* and almond) were phenotyped and genotyped with the IPSC9k 125 single nucleotide polymorphism (SNP) array, as previously described (Micheletti et al., 2015). The 126 inclusion of only peach (including P. ferganensis, Verde et al. 2012) among all the available 127 *Prunus* accessions was the leading concept behind the definition of the *PRP* reference collection. 128 Indeed, as a consequence of many factors (genetic diversity, evolution history, mating system, geographical distribution etc.), sampling strategies for the inclusion of wild relatives (e.g. species of 129 130 Amygdalus subgenus) may substantially differ from those for a cultivated species (e.g. peach) 131 (Brown and Marshall, 1995). Moreover, to avoid limitations on the exchange of plant material, the 132 domain was restricted to European repositories. Based on these criteria, the starting panel for 133 building the PRP amounted to a total of 1,262 P. persica accessions (FB_1262). Besides 134 accessions, seedlings from controlled crosses also represent a valuable source of informative 135 materials for both genetic analysis and breeding (or pre-breeding) activities. For this reason, 1,467 136 individuals from 18 progenies and their parents (including an interspecific cross with a P. davidiana 137 accession), also analyzed during the FruitBreedomics project (Hernandez Mora et al., 2017), were 138 considered in the construction process.

139

140 **1.2 Establishment of the** *PeachRefPop* size

The definition of the size is one of the most critical decisions for the establishment of a reference population. For fruit tree crops, the costs of *in vivo* maintenance are particularly onerous, and together with long-term space availability in the field, the main limiting factor of running a germplasm collection. In the perspective of analyzing the interactions between genotype and



Figure 1. **Overview of the range of phenotypic diversity in the** *PeachRefPop*. Columnar and standard tree growth habit (top left and right panel, respectively). Heart shaped, round and flat fruit (top and third row). Range of fruit flesh, skin color and overcolor (second and bottom rows). Variation in flower morphology and color (third and fourth rows). Fruit size variation (fourth row, first and last panels).

environment and/or management practices, or performing genetic studies such as GWAS and GS, an adequate panel size and experimental design are key factors for the power and reliability of statistical analyses. On the other hand, for agrobiodiversity conservation purposes, the least number of accessions to include in a core set depends on the level of genetic repetitiveness present in the original germplasm pool. The first step towards the establishment of the *PRP* size was the assessment of the allelic richness and redundancy observed at marker loci. Two series of core collections of incremental size were generated, one based on the genetic diversity



Figure 2. Graphical summary of the overall scheme followed for selecting the *PeachRefPop* collection. From the starting panel of 1,262 accessions, 169 accessions were selected combining two sets: 69 accessions extracted from genetic and phenotypic diversity analyses and taking into account availability of whole genome resequencing (WGRS) data; 100 accessions selected by an empirical strategy from an experts panel considering breeding and traditional value along with genetic structure. These were supplemented with 214 seedlings from crossing populations of scientific importance and their respective 20 parents. The total number of entries in the *PeachRefPop* amounts to 403.

(Maximization method, OPT), the other through random sampling (RAN). The maximization 152 procedure (M strategy by Schoen and Brown, 1993), is based on the sampling of the total allelic 153 154 diversity observed at marker loci in the least number of entries. By plotting the genetic diversity measured over the core size, a convex curve was obtained, indicating the presence of redundancy 155 156 across the European peach germplasm collection. The inflection point, corresponding to a plateau 157 in the increase of diversity, was observed at the level of core 26. At this core size, 99.9% of the total genetic diversity was captured in the core obtained with the M method in comparison to 93.5% 158 with random sampling (Figure 3). The outperformance of the optimized versus the random 159 160 selection was observed across all the core sizes, indicating that the OPT maximization strategy 161 was more efficient and was preferred for conservation purposes in our germplasm.



Figure 3. Assessment of allelic redundancy observed at marker loci in the starting panel (FB_1262). Core collections of incremental size were generated, based on the Maximization (OPT) and random sampling (RAN) methods in MSTRAT software, using a set of 445 SNPs. Datapoints represent averaged values over 5 independent repetitions for each size.

- According to some recent works in peach (reviewed in **Aranzana et al., 2019**), a number of about 100 – 150 unrelated accessions usually provides an adequate resolution for identifying major loci or developing prediction models. In light of all the above premises, an ideal target number of 400 entries was deemed adequate for allocating a minimum of 150 accessions and a maximum of 250
- seedlings from progenies (including the parents) based on the outputs of selection criteria.
- 167

168 **1.3 Identification of the selection criteria**

169 In spite of the genetic redundancy observed and excluding the rare cases of synonymy, the vast 170 majority of the accessions are not overlapped across the various collections, being conserved for a 171 multitude of reasons and purposes, including scientific research, agrobiodiversity preservation or 172 support to breeding activities. To reconcile these reasons with the aim of creating a feasible, 173 usable and multi-purpose reference collection to be shared among European institutions, a mixed 174 approach was considered for selecting the accessions. A subset of entries was sampled using an 175 analytical strategy, based on the criteria of maximizing genetic (and phenotypic) diversity, also 176 taking into account the availability of whole genome re-sequencing data (WGRS); the remaining 177 entries were selected using an empirical strategy, leveraging the knowledge of an experts panel 178 (e.g. breeders, experienced scientists and curators of each repository) and considering the 179 traditional and historical value at national and/or regional levels, the relevance for breeders, 180 growers and consumers, taking into account agronomical or pomological characteristics. Moreover, 181 to maintain a balanced representation of the genetic structure of the whole collection, the empirical 182 selection of accessions was partially supported by information on population structure [Structure 183 and principal component analysis (PCA) analysis available from Micheletti et al., 2015]. 184 Complementing the choice of accessions, seedlings were selected based on the availability of detailed genotypic and/or phenotypic information, genetic background, scientific relevance and, 185 186 above all, priority traits for breeding.

187

188 **1.4 Choice, evaluation and description of the** *PeachRefPop* accessions

189 Capturing the maximum amount of genetic diversity present in the entire collection while reducing 190 redundancy was the primary driver for sampling the first PRP subset (the core set). For this 191 purpose, the advanced M method, implemented in the software PowerCore (Kim et al., 2007) 192 through a modified heuristic algorithm, was used to select a core from the initial panel of 193 accessions, based on a set of 3,894 filtered SNPs previously described in Micheletti et al., (2015). 194 After superimposing 17 accessions with available whole-genome re-sequencing data, an ideal core 195 of 69 accessions (PwC 69) was extracted, representing a sampling size of 5.5% (Supplemental 196 Table 1). Considering the many variables that could affect the actual availability of materials for 197 grafting, a flexible approach was further developed to rank each accession of the whole panel 198 based on genotypic and phenotypic diversity. Four different sets made up of 100 cores of 70 199 entries each were constructed with MSTRAT by setting different combinations of genotypic (9) 200 subsets of SNPs extracted approximately every 1.8 Mb to avoid linkage between them) and 201 phenotypic data (7 qualitative and 10 quantitative traits, following transformation of the latter into 202 categories) (Supplemental Table 2). Accessions were ranked in groups according to the average 203 frequency of inclusion across the four sets (Supplemental Table 3). Combining the core 204 population extracted by PowerCore with the MSTRAT ranking list resulted in a shortlist of 69 205 accessions (41 and 28, respectively, indicated as Core_69), ensuring the inclusion of the maximum 206 possible level of genetic diversity. For the completion of the final *PRP_X* panel, the remaining 100 207 accessions (Priority 100) were empirically selected by experts, following the above specified 208 criteria.

Estimates of genetic diversity were used to compare the starting panel FB_1262, the core collection obtained by PowerCore (PwC_69) and the final set of *PRP* accessions (*PRP_X*),

composed by joining Priority 100 and Core 69 subsets. In addition, Core Hunter software was 211 used to create additional core sets, either of 69 and 169 entries, based on the optimization of 212 213 various criteria, including allelic coverage (CV 169) and three distance-based algorithms A-NE (AN 69 and AN 169), E-NE (EN 69 and EN 169) and E-E (EE 69 and EE 169). Concerning 214 215 parameters accounting for allelic diversity, all sets showed high and similar values for the allelic 216 coverage (CV), while the number of effective alleles (N_e) and expected heterozygosity (H_e) were 217 slightly lower for the Priority 100 subset (**Table 1**). The Shannon-Weaver diversity index (SH) was 218 comparable among the different subsets, ranging between 0.595 in EE 169 and 0.534 in 219 Priority 100. SH generally displays higher values in the presence of a reduced redundancy (Peet, 220 **1975**). In contrast, values of observed heterozygosity (H_0) tended to be more variable, ranging 221 from a minimum of 0.202 in PwC_69 to a maximum of 0.318 in AN_69. According to Odong et al. 222 (2013), distance-based criteria were used for further evaluations, such as the minimization of A-NE 223 distance, particularly indicated for generalist collections (as the *PRP*), and maximization of either 224 E-E or E-NE, both suitable for core collection representing the extremes of the entire collection. A-225 NE distance generally tends to decrease along with the increase of core size, being minimized in 226 the AN 169 and AN 69 core sets (0.137 and 0.172, respectively), a priori optimized using this 227 selection criterion. Despite the relative low performance of both Priority 100 and Core 69 (0.188 and 0.195, respectively), the PRP_X set showed low values for this index (0.165), most probably 228 229 as a consequence of the increased size. Regarding E-E and E-NE, PRP_X (as well as 230 Priority 100) showed lower values, particularly for E-NE distance, indicating the presence of a 231 certain redundancy within the panel.

232 The population structure of peach germplasm was well represented in the PRP_X , in 233 agreement with the presence of clusters of breeding-derived accessions (further separated in 234 peach- and nectarine-type groups), Occidental traditional and admixed entries with prevalent 235 Oriental origins (Figure 4A). Structure was also preserved in the other core sets, except for that 236 selected through the E-E distance algorithm, tending to oversample the admixed group 237 (Supplemental Figure 1). PCA was also run to check the distribution of the PRP_X with respect to 238 the other sets, and the first two components explained 15.9 and 8.4%, respectively, of the total 239 variance detected. In the scatter plot, 95% confidence ellipses show almost overlapping areas 240 (except for EE 169), confirming that the PRP X panel was well distributed to represent the 241 structure of the starting germplasm (Figure 4B). Finally, a Neighbor-joining (NJ) tree, based on the 242 dissimilarity matrix between the whole FB 1262 panel, was also built to assess the distribution of 243 PRP accessions (Figure 4C).

A number of accessions of historical and regional importance, mostly belonging to the Occidental traditional cluster, were included. For example, French cultivars dating from late Middle Age ('Grosse Mignonne', 'Millecoton de Septembre', 'Reine des Verges', 'Brugnon Violet') (**Okie et al., 2008**), traditional non-melting Spanish cultivars ('Amarillo de Agosto 1', 'Calante', 'Campiel',



Figure 4. Genetic structure and phylogenetic analysis of PeachRetPop accessions. A) Population structure estimated in the whole panel (FB_1262) and PeachRetPop accessions (PRP_X), as estimated for K (number of a priori cluster) equal to 4; B) PCA analysis of the subsets with a core size of 169 entries. Scores for each accession were obtained from the work of Micheletti *et al.*, 2015. The 95% confidence ellipses in the scatter plot were estimated using PAST software. C) NJ phylogenetic tree. Blue squares indicate accessions with traditional and historical value, violet circles indicate the other PeachRetPop accessions, and colors reflect the population structure.

²⁴⁸ 'Jesca', 'Groc Abel', 'Groc Alto') (Badenes et al., 1998; Wünsch et al., 2006) and the Italian

249 'Crasiommolo Rosso' (a white fleshed nectarine belonging to the 'Sbergie' type) and 'Poppa di 250 Venere', first reported at the end of eighteenth century (Majoli, 1790 - 1810). The richness of the 251 Italian peach germplasm was also widely represented by materials from several regions, including Sicily ('Imera', 'Tardiva di Ficarazzi', 'Settembrina di Bivona', 'Gialla di Moavero') (Marchese et al 252 253 2005), Campania ('Zingara Nera'), Apulia ('Percoco di Turi'), Liguria ('Michelini'), Emilia-Romagna 254 ('Buco Incavato', 'San Varano 2' and 'San Varano 3', 'Rosa del West', this last used for the 255 preparation of the famous cocktail 'Bellini') and Tuscany ('Regina di Londa') (Gallesio, 2003; Monte et al., 2006; Liverani and Giovannini, 2016). Early breeding materials, mainly from US 256 257 programs and founders of most of the currently cultivated materials are also included, along with 258 worldwide commercial cultivars (Supplemental Table 4).

Finally, *PRP* accessions encompassed a wide range of phenotypic variability for traits related to fruit quality, resistance or tolerance against major diseases (brown rot, powdery mildew, leaf curl, aphids and Sharka disease), tree growth habit and phenology (**Figure 5 and Supplemental Table 4**).

263

1.5 Choice and description of the *PeachRefPop* progenies

265 Seedlings from 15 cross populations from the research and breeding activities of some European 266 universities and institutions were also added. Most of these materials were already described in 267 depth (Hernandez-Mora et al., 2017). The leading criterion for the choice of seedlings was the 268 effective segregation of priority traits in peach, mainly related to phenology (fruit developmental 269 period, maturity date), fruit quality (fresh weight, soluble solid content, titratable acidity, texture and 270 aroma) and disease resistance (brown rot, powdery mildew, green peach aphids and Plum Pox Virus (PPV)) (Table 2). A range of breeding materials was considered, such as F1, F2, BC1 271 272 populations as well as hybrids with P. davidiana, particularly interesting as a source of PPV 273 resistance (Decroocg et al., 2005).

274

275 2. Experimental design and orchard sites description

The *PRP* was established in 5 institutions from 3 countries (Greece, Italy and Spain) (Figure 6A):

- Institute of Agrifood Research and Technology (IRTA) in Gimenells, Catalonia region, Spain
 (ES)
- II. Murcia Institute of Agri-Food Research and Development (IMIDA) in Mula, Murcia region,
 Spain (ES)
- III. Centro di Ricerca per le Produzioni Vegetali (CRPV) in Imola, Emilia-Romagna region, Italy
 (IT)
- IV. Institute of Plant Breeding and Genetic Resources (IPB&GR) in Naoussa, Imathia region,
 Greece (GR)
- 285 V. Research Centre for Olive, Fruit, and Citrus Crops (CREA) in Rome, Italy (IT).



Figure 5. Distribution of main phenotypic traits in the *PeachRefPop* accessions. In the maturity date plot, UE, E, M, L and VL indicates ultra-early, early, medium, late and very late ripening accessions, respectively; SSC, soluble solid content; in the fruit texture plot, four major texture groups are shown: non-melting (NM), melting (M), slow-softening (SwS) and stony hard (SH).

For each accession and seedling, a single mother plant was propagated through grafting on a common 'GF677' rootstock by the same nursery. Plants were grafted in the same year (2015) to obtain trees of the same age. To ensure an adequate compromise between the number of replicate trees and sustainable costs of maintenance, an augmented design with replicated control checks was adopted in all sites except Rome, hosting a partial copy of the *PRP* (accessions only, without randomization).

Accessions and seedlings were arranged in two blocks (M1 and M2) according to the following design (**Figure 6B**):

- the M1 block, composed of two sub-blocks (M1.1 and M1.2) each including the entire 294 295 PRP collection of 169 accessions plus 20 cross parents (A group) and 214 seedlings (S group) for 296 a total of 403 genotypes in each sub-block. Taking into account the physical layout of the design for 297 each location/field combination (i.e., the number of rows and the number of positions per row) each 298 accession and seedling was randomly assigned to a position within a sub-block (as illustrated for 299 Gimenells location in **Supplemental Table 5**). To assess and correct for spatial variation within and 300 between experimental sites, the three accessions 'Big Top', 'Springcrest', and 'Nectaross' from the 301 A group were included with a higher replication and randomly distributed over M1.1 and M1.2 sub-302 blocks (at least 5 additional trees of each genotype for each sub-block).



Figure 6. Experimental design and *PeachRefPop* orchards layout. A) Google maps satellite images of the established *PeachRefPop* orchards across the different European sites; B) Experimental design of multisite *PeachRefPop*. A schematic example is provided for Gimenells location. Accessions (A) and seedlings (S) in each block and sub-block were completely randomized. The M1.1 and M1.2 sub-blocks each include a full copy of the collection (189 accessions, 214 seedlings) plus replicate checks (C) of the accessions 'Big Top', 'Nectaross' and 'Springcrest' (5 additional trees for each sub-block). The M2.1 and M2.2 sub-blocks include half of the *PeachRefPop* collection and each site has a different half, chosen according to a pairwise design scheme. To this end, accessions (excluding control checks), and seedlings were randomly assigned to eight disjoint subgroups (A1 to A4 and S1 to S4) of approximately equal size and 4 of them assigned so that each site shares at least one A or one S group with the other sites. In the example, each M2 sub-block at Gimenells is composed of A1 and A2 (46 and 46 accessions, respectively, plus the 3 checks for a total of 95), S1 and S2 (54 and 54 seedlings, respectively, for a total of 108) other than 3 additional replicates for each of the 3 checks (9 trees). Gimenells shares the A1 and A2 with Naoussa.

- the **M2 block**, composed of two sub-blocks (M2.1 and M2.2) each including half of the PRP collection (85 accessions plus 10 cross-parents and 112 seedlings). In each site, the M2.1 and M2.2 sub-blocks include the same set of entries (i.e. the same half of the collection), randomly assigned in each sub-block, plus the replicated control checks previous described (at least 3 additional trees of each entry for each sub-block). The composition of the M2 block is not the same 308 across sites (i.e. each site has a different half of the collection), chosen according to a pairwise 309 design scheme (Figure 6B): firstly, excluding control checks, accessions and seedlings of the PRP 310 collection were randomly divided into eight disjoint subgroups - A1 to A4 (of 48, 47, 47 and 48 accessions, respectively) and S1 to S4 (of 54, 53, 54 and 53 seedlings, respectively) - then 4 311 312 subgroups were assigned so that each site shares at least one A or one S with the other sites. For 313 example, Imola location shares A4 and S3 with Naoussa, A3 with Mula and S2 with Gimenells. 314 This partial replication design is such that, within the full design, all subgroups were well 315 connected.

316 The geographic location of each site as well as the basic climate and soil parameters are 317 shown in **Table 3**. The sites covered a range of latitude from about 38° N in Mula (south-eastern 318 Spain) to 44° N in Imola (northern Italy), while altitude spanned from near sea level in Imola and 319 Rome (53 and 73 m, respectively) to 278 m at the Mula site. Although all sites are included in the 320 Mediterranean zone, climates widely range from semi-arid in Mula (warm winter and hot summer) 321 to sub-continental in Imola and Naoussa (with moderately cold winter). Average monthly 322 temperatures (1999 - 2018 time series) varied from the colder regimes of Naoussa (1.7 ± 3.9°C 323 and $23.5 \pm 2.9^{\circ}$ C in the coldest and hottest month, January and August, respectively) to the 324 warmer conditions of Mula (7.9 \pm 3.7 °C and 27.1 \pm 2.6 °C, respectively) (Figure 7A). The fulfillment 325 of chilling requirement (i.e. the period of cold temperatures needed for overcoming endo-326 dormancy) is a parameter of utmost relevance for peach reproductive phenology. According to the 327 Chilling Hours (CH) model (Weinberger, 1950), assigning one hour for each hourly temperature 328 between 0 and 7.2° C threshold, accumulation patterns widely ranged from 1,762 ± 124 CH at 329 Naoussa to 693 ± 159 at Mula. Also, precipitation differently affected the selected sites, with Imola 330 having the wettest conditions (964 ± 218 mm per year) and both Spanish locations having the 331 driest (Gimenells 361 ± 95 and Mula 336 ± 75) (Figure 7B).

332



Figure 7. **Climatic profiles of** *PeachRefPop* **sites.** A) Trend of minimum and maximum daily air temperatures at the five locations (averaged from 1999 – 2018 time series). Thick lines show smoothed mean temperatures. B) Average monthly precipitations (in mm).

334 Discussion

335 The concept of the PRP arises from the growing awareness about current and common 336 issues on ex situ peach conservation across European institutions. Fluctuations in funding 337 availability and intrinsic constraints of living orchard collections threaten the long-term preservation 338 of diversity resources, causing a progressive loss of valuable materials. Reference or core 339 collections have been designed for several fruit tree species, for example olive (Khadari et al., 340 2003; El Bakkali et al., 2012; Belaj et al., 2012), grape (Laucou, et al., 2011), cherry (Campoy 341 et al., 2016), apple (Gross et al., 2013; Lassois et al., 2016) and apricot (Krichen et al., 2012). 342 Nevertheless, they have mainly been created for improving resource allocation in the context of a 343 single institution or repository. The development of a trans-national and shared strategy provides

344 the most promising opportunity in the conservation approach. Actual establishment of the PRP has 345 required huge coordination efforts and faced the effective availability of materials, the difficulties of 346 their exchange and the success of clonal propagations (particularly for old, often unique, accessions). The sampling strategy for the PRP has been defined to accommodate multiple 347 348 purposes while maintaining the maximum possible diversity compared to the starting panel. The 349 final panel of accessions was assembled by the combination of two different subsets: the first 350 (Core 69), ensuring the preservation of the total allele number with the minimum number of 351 accessions, was extracted by widely adopted maximization strategies, either using a class 352 coverage criterion (in PowerCore) or Shannon-Weaver index (SH index (in MSTRAT), with the 353 latter penalizing redundancy. The second subset, accommodating for other scopes (Priority 100), 354 was chosen by experts with a robust knowledge on the genetic structure in peach, providing a 355 reliable criterion for assisting selection. As a whole, genetic analysis supports that PRP 356 composition is highly representative of the diversity of peach germplasms present in European 357 collections, as it retains all the allelic variability present within the starting panel, specifically targets 358 defined genetic clusters according to the genetic structure and includes most relevant phenotypic 359 traits. Indeed, differences among the various sampling strategies were negligible for allelic coverage (CV), expected heterozygosity and SH index, revealing a buffer effect towards 360 361 optimization criteria. Such effect could be expected, since peach has experienced a severe 362 domestication bottleneck with a reduction of genetic diversity, followed by a strong artificial 363 selection during domestication and modern improvement (Verde et al., 2013; Yu et al., 2018; Li et 364 al., 2019). This is also reflected in the narrow genetic bases of peach germplasm available across 365 main European repositories. Thus, the high level of allelic redundancy allows selecting many 366 different subpopulations able to retain the same amount of genetic variation. In spite of this, a 367 preliminary validation using distance-based criterion not used in the selection stage showed a 368 minimized A-NE index, the most indicative for evaluating the quality of multipurpose collections 369 (Odong et al., 2013). Conversely, E-E and, particularly, E-NE indices resulted less optimized, due 370 to a certain redundancy on the Priority 100 subset (i.e. a higher number of genotypes providing 371 unique alleles). This was mainly due to the inclusion of accessions of traditional and breeding 372 values, respectively belonging to the Occidental Traditional and Occidental breeding clusters, 373 characterized by a very narrow genetic background. Clearly, the inclusion of these materials is 374 crucial in the overall perspective of balancing diversity and usefulness, as they integrated various 375 fundamental qualities, such as popularity, prestige, tradition and breeding. A similar mixed strategy 376 was also recently optimized for creating a core collection for Swiss pear germplasm (Urrestarazu 377 et al., 2019).

Climate challenges in peach growing areas increase the need for resilient cultivars able to maintain productivity while showing an enhanced capacity for adaptation to sub-optimal conditions. Nevertheless, resilience and adaptive traits often have a complex inheritance and a strong 381 interaction with the environment or cultivation practice (Kissoudis et al., 2016). The partitioning of 382 phenotypic variation into genotypic, environmental and their interaction components involves ad 383 hoc experimental designs and integration of field data on a common set of genetic materials under 384 a range of different environmental/management conditions. Multi-environment trials (METs) have 385 been extensively used to study GxExM interactions, carry out GWAS and develop GS models for 386 complex traits in annual crops (Malosetti et al., 2013; Gutierrez et al., 2015; Zhu et al., 2018; 387 Bustos-Korts et al., 2019) or study GxE interactions in forest trees (Li et al., 2017). In contrast, 388 such experimental designs are lagging in fruit trees, largely because of the need for large and 389 diverse germplasm sets for quantitative genetics analyses and the above-mentioned difficulties in 390 material propagation and exchange. The PRP aims to fill this gap, as the replicated design and the 391 different pedo-climatic conditions across sites are particularly indicated for the dissection of 392 interactions between genotype and environment and/or management practice. The PRP locations 393 cover major climatic zones of the Mediterranean area, from semi-arid conditions of southern Spain 394 to sub-continental of northern Italy and Greece determining a broad range of temperatures and 395 precipitation patterns. In particular, sites are characterized by a different rate and amount of chilling 396 and heat accumulation, which will be particularly useful for the dissection of traits associated to 397 reproductive phenology, such as blooming and fruit ripening time. The proximity of experimental 398 sites to major production areas provide an added value for the translation of scientific outcomes. 399 The inclusion of both accessions and seedlings from various crosses allows development and 400 testing of statistical approaches for genomics-assisted breeding, such as joint linkage-association 401 analysis (Yu et al., 2008; Lu et al., 2010) and genome-wide selection (Resende et al., 2012; van 402 Nocker and Gardiner, 2014), or systematic QTL validation (Peace et al., 2014). Also, the 403 integration of omics (including epi-genomics) data may improve our understanding of physiological 404 changes in response to environmental stimuli and constraints.

405 The *PRP* multi-site experimental design was established with a complete randomization of 406 genotypes (accessions and seedlings) within each sub-block and replicate checks to account for 407 spatial variability. The rationale behind the choice of such design mainly derived from the 408 possibility of a direct comparison with standard reference varieties. A drawback of this approach is 409 the relatively few degrees of freedom for experimental errors, lowering the power to detect 410 differences among genotypes. The use of alpha designs (Patterson and Williams, 1976) and 411 derived row-column designs (John and Eccleston, 1986) might be statistically more powerful, 412 especially to estimate contrasts between genotypes and improve estimation of spatial variation, 413 e.g. due to different soil composition within the orchard. The identification of optimal designs for a 414 large number of genotypes is still challenging (Cullis et al., 1998). The PRP will allow validation of 415 the performance of this experimental design on fruit trees and a foundation for future planning of 416 multisite collections.

The PRP has been grafted on a single 'GF677' rootstock, a P. amygdalus × P. persica 417 418 hybrid. 'GF677' is the most widespread, mainly for its growth vigor, excellent affinity, adaptation to 419 limestone soils and tolerance to drought and replanting (Reighard and Loreti, 2008). While the 420 choice of a single rootstock is justified by the need of simplifying the experimental design, this 421 precludes assessment of scion-by-rootstock interaction. A number of Prunus rootstocks are 422 currently available for peach, some of them harboring interesting traits for resistance to soil 423 pathogens or abiotic stress conditions. Their integration into feasible experimental designs will be 424 the next challenge.

425 In perspective, the PRP should fulfill several purposes, from research to education and 426 traineeship of young breeders. A better understanding of diversity is expected to encourage the 427 use of broad-ranging germplasm (maybe also in other existing ex situ collections) in breeding 428 programs. In the last decades, the mission of many agriculture-oriented institutions has shifted 429 from the traditional focus of establishing horticultural collections to a wider target of preserving germplasm resources and agricultural heritage (Hammer et al., 2003; Havens et al., 2006). This 430 431 objective is of utmost importance for fruit tree species of ancient cultivation history, such as peach. 432 For these reasons, a number of traditional and local cultivars (either old or relatively modern) has 433 been included in the PRP, as a safeguard of an integral part of the rural landscape and collective 434 memory. Since information and descriptions about local germplasms are scarce and often restricted to cultivation areas, their choice has been directly handled by curators of each repository, 435 436 with the aid of experienced breeders.

437

438 Materials and Methods

439 Datasets

440 A set of 1,262 accessions was selected as representative of the peach [Prunus persica (L.) Batsch] 441 germplasm maintained in collections of four different European countries (Supplemental Table S3). 442 The complete list of institutions providing plant materials, SNP genotyping and phenotypic data for 443 seven monogenic traits have been previously described (Micheletti et al., 2015). SNP genotyping 444 data were obtained from the Genome Database for Rosaceae 445 (https://www.rosaceae.org/publication datasets accession number tfGDR1013). Phenotypic data reported in Supplemental Table 4 were obtained from Micheletti et al. (2015) and the 446 447 FruitBreedomics database (http://bioinformatics.tecnoparco.org/fruitbreedomics/).

448 Construction of core subsets

The advanced M (maximization) strategy implemented in PowerCore v. 1.0 (**Kim et al., 2007**) using 3,894 SNP markers was carried out to extract a core subset able to capture all the alleles observed in the entire collection. The size of the final core collection depends on the level of variability and redundancy present in the whole panel and cannot be set *a priori*. Seventeen kernel accessions with available whole-genome re-sequencing data were superimposed through the 454 'preferential selection' tool, which retains the accessions defined by the user without validation. 455 The standard M strategy implemented in MSTRAT (Gouesnard et al., 2001) was also applied. 456 MSTRAT algorithm selects a subset of *n* accessions from the *N* accessions of the entire collection 457 by maximizing the number of alleles (and/or trait classes) at each locus. The sampling size 458 estimated with PowerCore was set as default parameter and four sets of 100 core collections were 459 constructed by using different combinations of genotypic and phenotypic data. Due to the restraints 460 in the number of variables MSTRAT is able to manage, different subsets of approximately 100 461 SNPs each were obtained through an *ad hoc* developed Perl script program, by extracting 1 SNP 462 every 1,800 Kbp, corresponding to the max boundary for linkage disequilibrium (LD) found in some 463 subpopulations of the original plant material (Micheletti et al 2015). Seven qualitative and 10 464 quantitative traits (these last transformed into qualitative categories) were used as phenotypic data. 465 For each run, the core size was set to 70 and 100 independent replicates with 100 iterations were 466 generated. The Shannon-Weaver diversity index was used as a second criterion to classify core 467 subsets. Redundancy was assayed through the 'Redundancy' tool implemented in MSTRAT, which 468 samples two different sets of core collections of increasing size, as defined by the user, through 469 the application of the maximization strategy or random sampling. For this analysis a subset of 445 470 SNP markers was pruned from the whole set of 4271 using Plink v1.07 with a window size of 50, a 471 shift of 7 and a variance inflation factor (VIF) of 2. Redundancy was assayed in the whole panel of 472 accessions with a step of 5 in the first 100, 5 repetitions and 50 iterations. The Mixed Replica 473 search algorithm implemented in the Core Hunter II software (De Beukelaer et al., 2012) was 474 used to generate a core collection of fixed size (either of 69 and 169 entries) based on the 475 optimization of the Modified Rogers' (MR) distance measure (Wright, 1978), with a weight of 1.0. 476 For the evaluation of the quality of the different core subsets, genetic distance-based criteria were 477 considered: the average genetic distance between all the entries of each core collection (E-E); the 478 average distance between each entry and the nearest neighboring entry for each core collection 479 (N-E); the average distance between each genotype of the entire collection and the nearest entry 480 in each core collection (A-NE). The quality of each collection increased for lower value of A-NE 481 (the maximum representation is obtained for AN = 0, when each accession is represented by itself 482 or by an identical duplicate), and higher value both for E-NE (maximizes the average distance 483 between each selected individual and the closest other selected item in the core) and E-E 484 (maximizes the average distance between each pair of selected individuals in the core.).

485 Genetic diversity and population analyses

Genetic diversity measures were performed using GenAlex 6.41 (**Peakall et al., 2006**) and include: number of effective alleles (N_e , the number of equally frequent alleles required to give the observed level of heterozygosity), levels of observed (H_o) and expected (H_E) heterozygosity, and the Shannon-Weaver index (SH). Allelic coverage was calculated by the function CV implemented in Core Hunter II software. Population structure was inferred using a model-based clustering 491 algorithm ADMIXTURE v1.22 (Alexander et al., 2009). From SNP data, the software identifies K a 492 priori genetic clusters provided by the user, and for each individual it estimates the probability of 493 membership to each cluster. A preliminary analysis was performed by inputting successive values 494 of K from 2 to 6. The value of K that maximized the predictive accuracy was chosen based on a 495 10-fold cross-validation procedure with 10 different fixed initial seeds (Supplemental Figure 2). 496 Data of Principal Component Analysis (PCA) were retrieved from a previous work (Micheletti et 497 al., 2015). The 95% confidence ellipses in the scatter plot were estimated using PAST software 498 (Hammer et al., 2001). Phylogenetic tree was built from a pairwise genetic distance matrix 499 between individuals clustered with NJ method in TASSEL (Bradbury et al., 2007). Bootstrap 500 replicate and tree reconstruction were performed in MEGA7 software (Kumar et al., 2016).

501 Experimental design and pedo-climatic analyses

502 In the experimental design, randomization was performed with the Genstat software 503 (https://genstat.kb.vsni.co.uk/knowledge-base/hcitegen/). Meteorological time-series from 1999 to 504 2018 were obtained from ECMWF (European Centre for Medium-Range Weather Forecasts), 505 except for Mula (Murcia, Spain), for which data were available from a nearby weather station. 506 Hourly temperature series were obtained by linear interpolation of available tri-hourly data and 507 expressed in degrees Celsius. Cumulative precipitations were averaged and expressed in mm per 508 month or year. Chill accumulation was calculated according to the Chilling Hours Model 509 (Weinberger, 1950) as the sum of hourly temperatures between 0 and 7.2 °C during the dormant 510 season (15 November - 31 March). Soil texture was expressed according to USDA (United States 511 Department of Agriculture) classification. Mineral composition, pH and organic matter content were 512 determined according to standard procedures for soil analysis.

513

514 Supplemental Data

515

516 **Supplemental Figure S1**. Population structure estimated in the core sets AN_169, EE_169, 517 EN 169 and CV 169.

- 518 **Supplemental Figure S2**. Predictive accuracy (cross-validation error) of population stratification in
- 519 both *PRP_X* and FB_1262 as determined by Admixture software.
- 520 **Supplemental Table S1**. PowerCore output.
- 521 **Supplemental Table S2**. MSTRAT outputs for the four settings.
- 522 **Supplemental Table S3**. Accession ranking by MSTRAT frequencies.
- 523 **Supplemental Table S4**. *PeachRefPop* accessions description.
- 524 **Supplemental Table S5**. Layout of sub-block M1.1 as illustrated for Gimenells location
- 525
- 526 Acknowledgments

527 We wish to thank Claudio Buscaroli and Martina Lama for field assistance, Remo Chiozzotto for 528 lab assistance and Michela Troggio (Fondazione Edmund Mach) for genotypic analyses. We are 529 grateful to Fosco Vesely for extracting climate series. This work has been partially supported by 530 the PRIMA-FREECLIMB international project (DB, IE, PD, BQT), the European Union-funded 531 project "FruitBreedomics: Integrated approach for increasing breeding efficiency in fruit tree crops" 532 (Grant #FP7-265582) (DB, LR, PA, IV, TB, BQT, WG), and in the framework of MAS.PES, an 533 Italian project aimed at apricot and peach breeding (DB, SF). We thank the INRAE's 'Prunus 534 Biological Resources Center' for preserving and managing the peach collections and the Fruit Tree 535 Experimental Unit (UEA) of INRAE-Nouvelle Aquitaine-Bordeaux for growing the trees and grants 536 from the Spanish Ministry of Economy and Competitiveness (MINECO/FEDER projects AGL2012-537 40228-C02-01 and RTA2015-00050-00-00) and the CERCA Programme-Generalitat of Catalonia 538 (PA). We also thank the Italian National Centre of Fruit Germplasm (Centro Nazionale per il 539 Germoplasma Frutticolo, CNGF) at CREA - Rome for preserving and maintaining peach 540 collections, and the Italian Ministry of Agriculture (MIPAAF) for its financial support through the 541 RGV-FAO program (IV).

542

543 **Dedication**

544 This work is dedicated to the memory of our colleague Chiara Ferrandi, who recently passed away.

545

546

547 Tables

548

Table 1. Genetic analysis and parameters for the different core subsets. N_e : number of effective alleles); SH: Shannon-Weaver diversity index; H_o : observed heterozygosity; H_e : expected heterozygosity; CV: percentage allelic coverage; MR distance: average Modified Rogers genetic distance; E-E: average entry-to-entry distance; A-NE: average distance between each genotype of the collection and the nearest entry, E-NE: average distance between each entry and the nearest entry.

Satisama	N	eп	ы	ы	cv	MR distance			
Set name	IN _e	эп	Πο	Π _e		EE	A-NE	E-NE	
FB_1262	1.621	0.547	0.292	0.367	0.995	0.285	-	0.131	
PwC_69	1.675	0.574	0.202	0.39	0.987	0.318	0.203	0.237	
EE_69	1.705	0.587	0.234	0.401	0.992	0.347	0.209	0.229	
AN_69	1.645	0.560	0.318	0.378	0.977	0.286	0.172	0.210	
EN_69	1.704	0.587	0.269	0.402	0.991	0.334	0.207	0.275	
CV_169	1.638	0.556	0.285	0.375	0.995	0.302	0.163	0.212	
EE_169	1.721	0.595	0.224	0.408	0.994	0.330	0.183	0.191	

PRP_X	1.647	0.563	0.270	0.379	0.988	0.290	0.165	0.180
Priority_100	1.597	0.534	0.283	0.356	0.979	0.277	0.188	0.179
Core_69	1.713	0.593	0.247	0.406	0.988	0.303	0.195	0.212
EN_169	1.683	0.578	0.277	0.394	0.993	0.315	0.175	0.256
AN_169	1.643	0.559	0.300	0.377	0.987	0.290	0.137	0.203

Table 2. Description of the progenies used for establishing the *PeachRefPop* collection. Trait
abbreviations: FD, flowering date; MD, maturity date; SSC, soluble solid content; FW, fruit weight;
BR, brown rot; TA, titratable acidity; SwS, slow-softening texture; PM, powdery mildew; PPV, Plum
Pox Virus; GPA, green peach aphid; SH, stony hard texture.

Cross (parents)	Acronym	Institution	Type of Progeny	Seedlings #	Trait(s)	
'Bolero' × 'Oro A'	В×О	UMIL – Milan	F1	9	MD, SSC, FW, skin overcolor, aroma	
'Contender' × 'Elegant Lady'	C × EL	UMIL – Milan	F1	14	BR, MD	
'Max 10' × 'Rebus 028'	M×R	UMIL – Milan	F1	9	MD, TA, SSC, FW, SwS	
'Sweetfire' x 'Garcica'	Sf × G	UMIL – Milan	F1	15	MD, TA, SSC, FW, SwS	
'Belbinette' × 'Nectalady'	Bb × NI	IRTA – Lleida	F1	20	FD, MD, TA, SSC, FW	
'Big Top' × 'Nectaross'	Bt × Nr	IRTA – Lleida	F1	19	FD, MD, TA, SSC, FW	
'Big Top' × 'Armking'	Bt × Ak	IRTA – Lleida	F1	18	FD, MD, TA, SSC, FW	
'Subirana' x 'Feraude'	PN643	IRTA – Lleida	F1	7	Fruit shape	
'Summergrand' x ' <i>P. davidiana</i> P1908'	SD	INRA – Avignon	F1	6	PM, PPV	
'Zephyr' × [(('Summergrand' (S) × ' <i>P.</i> <i>davidiana</i> P1908')) x S]	BC2	INRA – Avignon	BC2	13	FD, PM, PPV, TA, SSC, FW	
'Pamirskij 5' × 'Rubira'	P × R	INRA – Avignon	F2	13	PM, GPA, foliage colour	
FRF 1495 x FRF 1148 (Ma 16-03-059)	POP1376	CREA – Forli	F1	17	PM; fruit pubescence	
IFF 983 x Ma 25-01-042	POP1115	CREA – Forli	F1	17	TA, SwS and aroma	
FRF 1695 x FRF 1681	POP1095	CREA – Forli	F1	19	SH	
FRF 813 × FRF 691	POP1039	CREA – Forli	F1	18	skin overcolor	

Table 3. Basic pedo-climatic features of the five *PeachRefPop* locations. Features include geographic coordinates, altitude, average annual minimum and maximum temperatures and cumulative annual precipitations (data series 1999 – 2018). Chilling accumulation was calculated according to the Chilling Hours (CH) model as the sum of hourly temperatures between 0 and 7.2 °C during the dormant season; S.O.M, soil organic matter content.

	• ••••	Altitude (m)		Soil				
Site	Geographical coordinates		Avg. Annual Temperature (min - max, °C)	Cumulative Precipitation (mm)	Chilling accumulation (CH)	Texture	рН	S.O.M. (%)
CREA - Rome (IT)	41°47' N - 12°33' E	79	11.2 - 18.9	731 ± 155	1171 ± 224	sandy - loam	7.7	1.9
CRPV - Imola (IT)	44°20' N - 11°45' E	53	9.5 - 17.9	964 ± 218	1753 ± 195	silty - loam	7.2	1.5
IMIDA - Mula (ES)	38°3' N - 1°25' O	278	12.0 - 24.5	336 ± 75	693 ± 159	clay	7.8	2.6
IPB&GR - Naoussa (GR)	40°37' N - 22°06' E	119	8.1 - 17.2	818 ± 160	1762 ± 124	sandy - loam	6.8	2.5
IRTA - Gimenells (ES)	41°39' N - 0°23' E	259	9.1 - 18.9	361 ± 95	1637 ± 133	sandy - loam	7.7	2.6

566

567 Figure Legends

568

Figure 1. Overview of the range of phenotypic diversity in the *PeachRefPop*. Columnar and standard tree growth habit (top left and right panel, respectively). Heart shaped, round and flat fruit (top and third row). Range of fruit flesh, skin color and overcolor (second and bottom rows). Variation in flower morphology and color (third and fourth rows). Fruit size variation (fourth row, first and last panels).

574 Figure 2. Graphical summary of the overall scheme followed for selecting the PeachRefPop collection. From the starting panel of 1,262 accessions, 169 accessions were selected combining 575 576 two sets: 69 accessions extracted from genetic and phenotypic diversity analyses and taking into 577 account availability of whole genome resequencing (WGRS) data; 100 accessions selected by an 578 empirical strategy from an experts panel considering breeding and traditional value along with 579 genetic structure. These were supplemented with 214 seedlings from crossing populations of 580 scientific importance and their respective 20 parents. The total number of entries in the 581 PeachRefPop amounts to 403.

- **Figure 3**. Assessment of allelic redundancy observed at marker loci in the starting panel (FB_1262). Core collections of incremental size were generated, based on the Maximization (OPT) and random sampling (RAN) methods in MSTRAT software, using a set of 445 SNPs. Datapoints represent averaged values over 5 independent repetitions for each size.
- **Figure 4.** Genetic structure and phylogenetic analysis of *PeachRefPop* accessions. A) Population structure estimated in the whole panel (FB_1262) and *PeachRefPop* accessions (*PRP_X*), as estimated for K (number of a priori cluster) equal to 4; B) PCA analysis of the subsets with a core size of 169 entries. Scores for each accession were obtained from the work of **Micheletti et al.**, **2015**. The 95% confidence ellipses in the scatter plot were estimated using PAST software. C) NJ phylogenetic tree. Blue squares indicate accessions with traditional and historical value, violet circles indicate the other *PeachRefPop* accessions, and colors reflect the population structure.

Figure 5. Distribution of main phenotypic traits in the *PeachRefPop* accessions. In the maturity date plot, UE, E, M, L and VL indicates ultra-early, early, medium, late and very late ripening accessions, respectively. SSC, Soluble solids content. In the fruit texture plot, four major texture groups are shown: non-melting (NM), melting (M), slow-softening (SwS) and stony hard (SH).

Figure 6. Experimental design and *PeachRefPop* orchards layout. A) Google maps satellite images of the established *PeachRefPop* orchards across the different European sites; B) Experimental design of multisite *PeachRefPop*. A schematic example is provided for Gimenells location. Accessions (A) and seedlings (S) in each block and sub-block were completely randomized. The M1.1 and M1.2 sub-blocks each include a full copy of the collection (189 accessions, 214 seedlings) plus replicate checks (C) of the accessions 'Big Top', 'Nectaross' and 603 'Springcrest' (5 additional trees for each sub-block). The M2.1 and M2.2 sub-blocks include half of 604 the PeachRefPop collection and each site has a different half, chosen according to a pairwise 605 design scheme. To this end, accessions (excluding control checks), and seedlings were randomly 606 assigned to eight disjoint subgroups (A1 to A4 and S1 to S4) of approximately equal size and 4 of 607 them assigned so that each site shares at least one A or one S group with the other sites. In the 608 example, each M2 sub-block at Gimenells is composed of A1 and A2 (46 and 46 accessions, 609 respectively, plus the 3 checks for a total of 95), S1 and S2 (54 and 54 seedlings, respectively, for 610 a total of 108) other than 3 additional replicates for each of the 3 checks (9 trees). Gimenells 611 shares the A1 and S1 groups with Mula, S2 with Imola and A2 with Naoussa. 612 Figure 7. Climatic profiles of *PeachRefPop* sites. A) Trend of minimum and maximum daily air

- 613 temperatures at the five *PeachRefPop* locations (averaged from 1999 2018 time series). Thick
- 614 lines show smoothed mean temperatures. B) Average monthly precipitations (in mm).

616

Parsed Citations

Alexander DH, Novembre J, Lange K (2009) Fast model-based estimation of ancestry in unrelated individuals. Genome Res 19: 1655-1664

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Aranzana MJ, Decroocq V, Dirlewanger E, Eduardo I, Gao ZS, Gasic K, Iezzoni A, Jung S, Prieto H, Tao R, Verde I, Abbott AG, Arús P (2019) Prunus genetics and applications after de novo genome sequencing: achievements and prospects. Hortic Res 6: 58.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Badenes ML, Martínez-Calvo J, Llácer G (1998) Analysis of peach germplasm from Spain. Acta Hort 465: 243-250.

Pubmed: <u>Author and Title</u> Google Scholar: Author Only Title Only Author and Title

Belaj A, del Carmen Dominguez-García M, Atienza SG, Urdíroz NM, De la Rosa R, Satovic Z, Martín A, Kilian A, Trujillo I, Valpuesta V, Del Río C (2012) Developing a core collection of olive (Olea europaea L.) based on molecular markers (DArTs, SSRs, SNPs) and agronomic traits. Tree Genet Genomes: 8(2): 365-378.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Benediková D, Giovannini D (2013) Review on genetic resources in the ECPGR Prunus Working Group. Acta Hortic 981: 43-51.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Biasi R, Botti F, Cullotta S, Barbera G (2012) The role of Mediterranean fruit tree orchards and vineyards in maintaining the traditional agricultural landscapes. Acta Hortic 940: 79-88

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Bradbury PJ, Zhang Z, Kroon DE, Casstevens TM, Ramdoss Y, Buckler, ES (2007) TASSEL: software for association mapping of complex traits in diverse samples. Bioinformatics 23(19): 2633-2635.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Campoy JA, Lerigoleur-Balsemin E, Christmann H, Beauvieux R, Girollet N, Quero-García J, Dirlewanger E, Barreneche, T (2016) Genetic diversity, linkage disequilibrium, population structure and construction of a core collection of Prunus avium L. landraces and bred cultivars. BMC Plant Biol 16(1): 49.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Chagné D, Dayatilake D, Diack R, Oliver M, Ireland H, Watson A, Gardiner SE, Johnston JW, Schaffer RJ and Tustin S (2014) Genetic and environmental control of fruit maturation, dry matter and firmness in apple (Malus × domestica Borkh.). Hortic Res 1: 14046.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Cirilli M, Geuna F, Babini AR, Bozhkova V, Catalano L, Cavagna B, Dallot S, Decroocq V, Dondini L, Foschi S, Ilardi V, Liverani A, Mezzetti B, Minafra A, Pancaldi M, Pandolfini T, Pascal T, Savino VN, Scorza R, Verde I, Bassi D (2016) Fighting Sharka in peach: current limitations and future perspectives. Front Plant Sci 7: 1290.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Cirilli M, Rossini L, Geuna F, Palmisano F, Minafra A, Castrignanò T, Gattolin S, Ciacciulli A, Babini AR, Liverani A, Bassi D (2017) Genetic dissection of Sharka disease tolerance in peach (P. persica L. Batsch). BMC Plant Biol 17(1): 192.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Cirilli M, Flati T, Gioiosa S, Tagliaferri I, Ciacciulli A, Gao Z, Gattolin S, Geuna F, Maggi F, Bottoni P, Rossini L, Bassi D, Castrignanò T, Chillemi G (2018) PeachVar-DB: a curated collection of genetic variations for the interactive analysis of peach genome data. Plant Cell Physiol 59(1): e2-e2.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Coakley SM, Scherm H, Chakraborty S (1999) Climate change and plant disease management. Ann Rev Phytopathol 37(1): 399-426.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Corelli-Grappadelli L, Marini RP (2008) Orchard Planting Systems. In: Layne DR, Bassi D, editors. The Peach: Botany, Production and Uses. Wallingford: CABI. pp. 264–288.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Cullis B, Gogel B, Verbyla A, Thompson R (1998) Spatial analysis of multi-environment early generation variety trials. Biometrics 54: 1-

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Decroocq V, Foulongne M, Lambert P, Gall O, Mantin C, Pascal T, Schurdi-Levraud V, Kervella J (2005) Analogues of virus resistance genes map to QTLs for resistance to sharka disease in Prunus davidiana. Mol Genet Genom 272: 680-689.

Pubmed: Author and Title

Google Scholar: Author Only Title Only Author and Title

El Bakkali A, Haouane H, Moukhli A, Costes E, Van Damme P, Khadari B (2013) Construction of core collections suitable for association mapping to optimize use of Mediterranean olive (Olea europaea L.) genetic resources. PLoS One 8(5): e61265.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Faust M, Timon B (1995) Origin and dissemination of peach. Hortic Rev 17: 331-379.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Ford-Lloyd B, Jackson M (1986) Plant Genetic Resources: An Introduction to Their Conservation and Use, Edward Arnold, London.

Franco J (2006) Sampling strategies for conserving maize diversity when forming core subsets using genetic markers. Crop Sci 46(2):854-864.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Frankel OH, Brown AHD (1984) Current plant genetic resources – a critical appraisal. In: Genetics: New Frontiers (vol IV). New Delhi, India: Oxford and IBH Publishing

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Fu YB, Dong YB (2015) Genetic erosion under modern plant breeding: case studies in Canadian crop gene pools. In Genetic Diversity and Erosion in Plants (89-104). Springer, Cham.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Gallesio G (2003) Il trattato del pesco di Giorgio Gallesio. In: Baldini, E. (ed.) Gli inediti trattati del pesco e del ciliegio. Complementi scientifici della 'Pomona Italiana' di Giorgio Gallesio. Accademia dei Georgofili, Florence, Italy, 9-146.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Gepts P (2006) Plant genetic resources conservation and utilization. Crop Sci 46(5): 2278-2292.

Pubmed: <u>Author and Title</u> Google Scholar: Author Only Title Only Author and Title

Grondona MO, Cressie N (1991) Using spatial considerations in the analysis of experiments. Technometrics 33: 381-392.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Gross BL, Volk GM, Richards CM, Reeves PA, Henk AD, Forsline PL, Szewc-McFadden A, Fazio G, Chao CT (2013) Diversity captured in the USDA-ARS national plant germplasm system apple core collection. J Am Soc Hortic Sci 138(5): 375-381.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Khadari B, Breton C, Moutier N, Roger J, Besnard G, Bervillé A, Dosba F (2003) The use of molecular markers for germplasm management in a French olive collection. Theor Appl Genet 106(3): 521-529.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Kim KW, Chung HK, Cho GT, Ma KH, Chandrabalan D, Gwag JG, Kim TS, Cho EG, Park YJ (2007) PowerCore: a program applying the advanced M strategy with a heuristic search for establishing core sets. Bioinformatics 23(16): 2155-2162.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Kissoudis C, van de Wiel C, Visser R, van der Linden G (2016) Future-proof crops: challenges and strategies for climate resilience improvement. Curr Opin Plant Biol 30: 47-56.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Krishnan RR, Sumathy R, Ramesh S, Bindroo B, Naik GV (2014) SimEli: Similarity elimination method for sampling distant entries in development of core collections. Crop Sci 54(3): 1070-1078.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Krichen L, Audergon JM, Trifi-Farah N (2012) Relative efficiency of morphological characters and molecular markers in the establishment of an apricot core collection. Hereditas 149(5): 163-172.

8.

Hammer Ø, Harper DA, Ryan PD (2001) PAST: paleontological statistics software package for education and data analysis. Palaeontol Electron 4(1): 9

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Laucou V, Lacombe T, Dechesne F, Siret R, Bruno JP, Dessup M, Ortigosa P, Parra P, Roux C, Santoni S, Varès D, Péros JP, Boursiquot JM, This P (2011) High throughput analysis of grape genetic diversity as a tool for germplasm collection management. Theor Appl Genet 122(6): 1233-1245.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Li Y, Suontama M, Burdon RD, Dungey HS (2017) Genotype by environment interactions in forest tree breeding: review of methodology and perspectives on research and application. Tree Genet Genomes 13: 60-78

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Malosetti M, Ribaut JM, van Eeuwijk FA (2013) The statistical analysis of multi-environment data: modeling genotype-by-environment interaction and its genetic basis. Front Physiol 4: 44.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Marchese A, Tobutt KR, Caruso T (2005) Molecular characterisation of Sicilian Prunus persica cultivars using microsatellites. J Hortic Sci Biotech 80: 121-129

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Mora JRH, Micheletti D, Bink M, Van de Weg E, Cantín C, Nazzicari N, Caprera A, Dettori MT, Micali S, Banchi E, Campoy JA, Dirlewanger E, Lambert P, Pascal T, Troggio M, Bassi D, Rossini L, Verde I, Quilot-Turion B, Laurens F, Arús P, Aranzana MJ (2017) Integrated QTL detection for key breeding traits in multiple peach progenies. BMC Genomics 18(1): 404.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Li Y, Cao K, Zhu G, Fang W, Chen C, Wang X, Zhao PP, Guo J, Ding T, Guan L, Zhang Q, Guo W, Fei Z, Wang L (2019). Genomic analyses of an extensive collection of wild and cultivated accessions provide new insights into peach breeding history. Genome Biol 20: 36 Pubmed: Author and Title

Google Scholar: <u>Author Only Title Only Author and Title</u>

Liverani A, Giovannini D (2016) Pesco. In: Fideghelli, C. Atlante dei fruttiferi autoctoni italiani, volume III. ISBN 978-88-99595-35-7 (Italian)

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Foulongne M, Pascal T, Arus P, Kervella J (2003) The potential of Prunus davidiana for introgression into peach [Prunus persica (L.) Batsch] assessed by comparative mapping. Theoret Appl Genet 107: 227-238.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Gradziel TM (2002) Almond species as sources of new germplasm for peach improvement. Acta Hort 592: 81-88

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Gouesnard B, Bataillon TM, Decoux G, Rozale C, Schoen DJ, David JL (2001) MSTRAT: An algorithm for building germ plasm core collections by maximizing allelic or phenotypic richness. J Hered 92(1): 93-94.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Gutiérrez L, Germán S, Pereyra S, Hayes PM, Pérez CA, Capettini F, Locatelli A, Berberian NM, Falconi EE, Estrada R, Fros D, Gonza V, Atamirano H, Huerta-Espino J, Neyra E, Orjeda G, Sandoval-Islas S, Singh R, Turkington K, Castro AJ (2015) Multi-environment multi-QTL association mapping identifies disease resistance QTL in barley germplasm from Latin America. Theor Appl Genet 128(3): 501-516.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

John JA, JA Eccleston (1986). Row-column a-designs. Biometrika 73: 301-306.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Laurens F, Aranzana MJ, Arus P, Bassi D, Bink M, Bonany J, Caprera A, Corelli-Grappadelli L, Costes E, Durel CE, Mauroux JB, Muranty H, Nazzicari N, Pascal T, Patocchi A, Peil A, Quilot-Turion B, Rossini L, Stella A, Troggio M, Velasco R, van de Weg E (2018) An integrated approach for increasing breeding efficiency in apple and peach in Europe. Hortic Res 5(1): 11.

Pubmed: Author and Title

Google Scholar: Author Only Title Only Author and Title

Li Y, Cao K, Zhu G, Fang W, Chen C, Wang X, Zhao P, Guo J, Ding T, Guan L, Zhang Q, Guo W, Fei Z, Wang L (2019) Genomic analyses of an extensive collection of wild and cultivated accessions provide new insights into peach breeding history. Genome Biol 20(1): 36.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Luedeling E (2012) Climate change impacts on winter chill for temperate fruit and nut production: a review. Sci Hortic 144: 218-229. Pubmed: Author and Title

Google Scholar: <u>Author Only Title Only Author and Title</u>

Kumar S, Stecher G, Tamura K (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol, 33(7): 1870-1874.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Myles S (2013) Improving fruit and wine: what does genomics have to offer? Trends Genet 29: 190-196.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Majoli C (1790-1810). Plantarum collectio juxta Linnaeanum systema a Lectore Caesare Majolio Hyeronimino digesta et depicta. Forlì.

Micheletti D, Dettori MT, Micali S, Aramini V, Pacheco I, Da Silva Linge C, Foschi S, Banchi E, Barreneche T, Quilot-Turion B, Lambert P, Pascal T, Iglesias I, Carbó J, Wang L, Ma R, Li XW, Gao ZS, Nazzicari N, Troggio M, Bassi D, Rossini L, Verde I, Laurens F, Arús P, Aranzana MJ (2015) Whole-genome analysis of diversity and SNP-major gene association in peach germplasm. PLoS ONE 10: e0136803. Pubmed: Author and Title

Google Scholar: <u>Author Only Title Only Author and Title</u>

Moing A, Poessel JL, Svanella-Dumas L, Loonis M, Kervella J (2003) Biochemical basis of low fruit quality of Prunusdavidiana, a pest and disease resistance donor for peach breeding. J Am Soc Hortic Sci 128: 55-62.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Monte M, Sottile F, Barone E, Caruso T, Bazzoni A (2006) The Sicilian Peach (Prunus persica L. Batsch) Germplasm: Horticultural Characteristics and Sanitary Status. Acta Hort 713: 57-60.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Odong T, van Heerwaarden J, Jansen J, van Hintum TJ, van Eeuwijk F (2011) Statistical techniques for defining reference sets of accessions and microsatellite markers. Crop Sci 51(6):2401-2411.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Okie WR, Bacon T, Bassi D (2008) Fresh Market Cultivar Development. In: Layne DR, Bassi D, editors. The Peach: Botany, Production and Uses. Wallingford: CABI: 264–288.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Parajuli R, Thoma G, Matlock MD (2018) Environmental sustainability of fruit and vegetable production supply chains in the face of climate change: A review. Sci Total Environ 650(2): 2863-2879

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Patterson HD, Williams ER (1976) A new class of resolvable incomplete block designs. Biometrika 63: 83-92.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only</u> <u>Author and Title</u>

Peace CP, Luby JJ, van de Weg WE, Bink MCAM, lezzoni AF (2014) A strategy for developing representative germplasm sets for systematic QTL validation, demonstrated for apple, peach, and sweet cherry. Tree Genet Genomes 10(6):1679-1694.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Peakall ROD, Smouse PE (2006) GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. Mol Ecol Notes 6(1): 288-295.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Peet RK (1975) Relative diversity indices. Ecology 56(2): 496-498.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Purcell S, Neale B, Todd-Brown K., Thomas L, Ferreira MAR, Bender D, Maller J, Sklar P, de Bakker PIW, Daly MJ, Sham PC (2007) PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet 81(3): 559-575. Pubmed: Author and Title

Google Scholar: Author Only Title Only Author and Title

Ramírez F, Kallarackal J (2015) Responses of fruit trees to global climate change. SpringerBriefs in Plant Science.

Reighard G, Loreti F (2008) Rootstock development. In Layne, D., and D. Bassi (eds.) The peach: Botany, production and uses. CABI Publishing, Wallingford, Oxon, UK.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Schoen DJ, Brown AHD (1995) Maximising genetic diversity in core collections of wild relatives of crop species. Core collections genetic resources (Hodgkin T, Brown AHD, van Hintum TJL, Morales EAV, eds.). John Wiley & Sons, Chichester, UK, 55-77. Pubmed: <u>Author and Title</u>

Google Scholar: <u>Author Only</u> <u>Title Only</u> <u>Author and Title</u>

Serra O, Donoso JM, Picañol R, Batlle I, Howad W, Eduardo I, Arús P (2016) Marker-assisted introgression (MA) of almond genes into the peach background: a fast method to mine and integrate novel variation from exotic sources in long intergeneration species. Tree Genet Genomes 12: 96

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Singh HCP, Rao NKS, Shivashankar KS (Eds.) (2013). Climate-Resilient Horticulture: Adaptation and Mitigation Strategies (pp. 81-88). Springer India.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Thachuk C, Crossa J, Franco J, Dreisigacker S, Warburton M, Davenport GF (2009) Core Hunter: an algorithm for sampling genetic resources based on multiple genetic measures. BMC Bioinformatics 10(1): 243.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Urrestarazu, J, Kägi C, Bühlmann A, Gassmann J, Santesteban LG, Frey JE, Kellerhals M, Miranda C (2019) Integration of expert knowledge in the definition of Swiss pear core collection. Sci Rep 9(1): 8934.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Varshney RK, Bansal KC, Aggarwal PK, Datta SK, Craufurd PQ (2011) Agricultural biotechnology for crop improvement in a variable climate: hope or hype? Trends Plant Sci 16(7): 363-371.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Verde I et al (2012) Development and evaluation of a 9K SNP array for peach by internationally coordinated SNP detection and validation in breeding germplasm. PLoS ONE 7: e35668.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Verde I et al (2013) The high-quality draft genome of peach (Prunus persica) identifies unique patterns of genetic diversity, domestication and genome evolution. Nat Genet 45: 487–U47.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Verde I et al (2017) The Peach v2.0 release: high-resolution linkage mapping and deep resequencing improve chromosome-scale assembly and contiguity. BMC Genomics 18: 225.

Pubmed: Author and Title Google Scholar: Author Only Title Only Author and Title

Weinberger JH (1950) Chilling requirements of peach varieties. Proc Amer Soc Hort Sci 56: 122-128

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Yu Y et al. (2018) Genome re-sequencing reveals the evolutionary history of peach fruit edibility. Nat Commun 9: 5404.

Pubmed: <u>Author and Title</u> Google Scholar: <u>Author Only Title Only Author and Title</u>

Zhu XM, Shao XY, Pei YH, Guo XM, Li J, Song XY, Zhao MA (2018) Genetic diversity and genome-wide association study of major ear quantitative traits using high-density SNPs in maize. Frontiers in plant science. Front Plant Sci 9: 966.

Pubmed: Author and Title

Google Scholar: <u>Author Only Title Only Author and Title</u>