Autoantibody Profile of a Cohort of 54 Italian Patients with Linear IgA Bullous Dermatosis: LAD-1 Denoted as a Major Auto-antigen of the Lamina Lucida Subtype

Emanuele COZZANI^{1#}, Giovanni DI ZENZO^{2#}, Giulia GASPARINI¹, Adele SALEMME², Arianna Fay AGNOLETTI¹, Camilla VASSALLO³, Marzia CAPRONI⁴, Emiliano ANTIGA⁴, Angelo Valerio MARZANO^{5,6}, Riccardo CAVALLI⁷, Corrado OCELLA⁸, Clara DE SIMONE⁹ and Aurora PARODI¹; and the Cutaneous Immunology group of SIDeMaST

¹Section of Dermatology, DISSAL, University of Genoa, and San Martino Policlinic Hospital, Genoa, ²Laboratory of Molecular and Cell Biology, IDI-IRCCS, Rome, ³Department of Clinical-Surgical, Diagnostic, and Pediatric Science, Institute of Dermatology, Fondazione IRCCS Policlinico San Matteo, Pavia, ⁴Section of Dermatology, Department of Health Sciences, University of Florence, Florence, ⁵UCO of Dermatology, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, ⁶Department of Physiopathology and Transplantation, Università degli Studi di Milano, ⁷UOC of Pediatric Dermatology, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, ⁸Dermatology Unit, Department of Surgery and Intensive Care, IRCCS G. Gaslini Children's Hospital, Genoa, and ⁹Institute of Dermatology, Fondazione Policlinico Universitario A. Gemelli IRCCS, Università Cattolica del Sacro Cuore, Rome, Italy [#]Both authors contributed equally to this work.

Linear immunoglobulin A (IgA) bullous dermatosis (LABD) is characterized by the presence of multiple IgA autoantibodies, and a comparatively lesser number of IgG antibodies, directed against different hemidesmosomal antigens. The main autoantigens are LAD-1, LABD-97, BP180 and BP230, type VII collagen and laminin 332. The serology of 54 Italian patients with LABD was studied retrospectively using enzymelinked immunosorbent assay (ELISA), immunoblotting assay, and indirect immunofluorescence on monkey oesophagus and salt-split skin. Among these, indirect immunofluorescence of salt-split skin elicits the greatest sensitivity. Sixty-three percent of the sera were observed to be positive, with a lamina lucida pattern observed in 48%, a sub-lamina densa pattern in 2% and a mixed pattern in 13% of cases. IgA reactivity to LAD-1 on immunoblotting was found in 52% of sera, to BP180-NC16A by ELISA in 32% and to BP230 in 26%. Only 17% of patients had circulating IgG autoantibodies. LAD-1 was determined to be a major autoantigen of the lamina lucida subtype. Combined serological assays demonstrated a high sensitivity (82%), suggesting that this approach could support diagnosis when a biopsy is not feasible or direct immunofluorescence results are negative.

Key words: linear IgA bullous disease; humoral immune response; LAD-1; BP180; BP230; diagnostic sensitivity.

Accepted Jan 9, 2020; Epub ahead of print Feb 3, 2020

Acta Derm Venereol 2020; 100: adv00070.

Corr: Emanuele Cozzani, Section of Dermatology, DISSAL, University of Genoa, Via Pastore 1, 16132, Genoa, Italy. E-mail: emanuele.cozzani@ unige.it

Linear IgA bullous dermatosis (LABD) is an IgAmediated autoimmune bullous disorder occurring in adults and children, characterized by sub-epidermal blisters and linear deposits of IgA at the basement membrane zone (BMZ) (1). Compared with other immunobullous diseases, LABD is immunologically a heterogeneous

SIGNIFICANCE

Linear IgA bullous dermatosis is a rare autoimmune disease, marked by blistering, affecting both children and adults. It is characterized by auto-antibodies targeting various types of skin proteins. We retrospectively studied 54 Italian patients with this disease. The auto-antibody targets were investigated using various blood tests (enzymelinked immunosorbent assay (ELISA), immunoblotting and indirect immunofluorescence on monkey oesophagus and salt-split skin). Indirect immunofluorescence on salt-split skin elicited the greatest diagnostic sensitivity. The most common target was LAD-1, followed by BP180-NC16A and BP230. Eighty-two percent of patients had circulating IgAs, and only 15% possessed circulating IgG auto-antibodies. Combined serological tests supported the diagnosis of linear IgA bullous disease.

disease with pathogenic IgA autoantibodies against different hemidesmosomal antigens (2). Researchers have identified multiple targets over years of research, including collagen VII (3–5), laminin 332 (6), an unidentified 285-kDa protein (7), bullous pemphigoid 230 kDa antigen (BP230) (8), bullous pemphigoid 180 kDa antigen (BP180) (collagen XVII) (8), a 120-kDa protein (linear IgA disease-1 antigen; LAD-1) (9–11) and a 97-kDa protein (LABD-97) (12–14) (**Fig. 1**A). The last 2 antigens were subsequently denoted as parts of the extracellular domain of BP180 ectodomain (10, 15–18).

BP180 is a hemidesmosomal type II transmembrane protein comprising an intracytoplasmic domain (NC16C domain), a transmembrane domain (NC16B domain), and an extracellular region with 15 collagenous domains and 16 non-collagenous domains (NC16A to NC1) (Fig. 1A) (19). The NC16A domain is the main epitope in bullous pemphigoid. BP180 can be cleaved physiologically within the NC16A domain by ADAM9/17 (16, 20) or pathologically by plasmin (21) and neutrophil elastase (22), into a 120-kDa extracellular fragment correspon-



Fig. 1. (A) Schematic representation of human BP180 and its fragments LAD-1 and linear IgA bullous dermatosis (LABD)-97. (B) The shed ectodomain of BP180 (LAD-1) is recognized by IgG and IgA autoantibodies as a 120-kDa polypeptide. Anti-BP180-positive bullous pemphigoid serum bind 120-kDa polypeptide by IgG (lane 1), molecular weight marker (lane 2), an anti-BP180 positive LABD serum bind 120-kDa polypeptide by IgA (lane 3), healthy volunteer serum does not bind 120-kDa polypeptide by IgA (lane 4); (C) IgA reactivity to LAD-1 (120 kDa) detected by immunoblotting: lane 1, healthy volunteer serum; lane 2, LABD control serum; lanes 3–7, LABD patient sera from the present study.

ding to LAD-1 (Fig. 1B). Further processing of LAD-1 within the carboxyl-terminal NC4 domain yields a 97-kDa fragment corresponding to LABD-97 (23, 24).

LABD differs from bullous pemphigoid, in which IgG autoantibodies preferentially target other parts of BP180 (25, 26). Furthermore, it has been shown that IgA autoantibodies from subsets of patients with LABD recognize epitopes within the NC16A region (27, 28).

LABD displays 2 major binding patterns of IgA autoantibodies in indirect immunofluorescence of salt-split skin (IIF-SSS) and in immuno-electron microscopy. The majority of the LABD sera IgA react with the epidermal side of IIF-SSS, corresponding to the lamina lucida type sera, while a fewer number of IgA react with the dermal side of IIF-SSS, corresponding to the sub-lamina densa type sera (29, 30). A limited number of IgA elicit mixed reactivity to both locations. The type VII collagen in anchoring fibrils was determined to be the primary target of IgA antibodies from patients with sub-lamina densa type of LABD (4, 5, 31). In contrast, most patients with lamina lucida type LABD have IgA autoantibodies targeting LABD-97 (12–14) and LAD-1 (9, 10).

The autoimmune heterogeneity considers both targeted antigens as well as the involved Ig class. Apart from autoreactive IgAs, IgGs were also reported to be associated with LABD, which are detected as linear deposits at the BMZ in direct immunofluorescence (DIF) and detected in patients' sera in IB testing or by enzyme-linked immunosorbent assay (ELISA) (6).

The present retrospective study characterized the humoral immune response of 54 Italian patients with LABD using standard approaches, such as IIF-SSS and commercial and adapted ELISAs. In addition, this study suggests that LAD-1 is the major antigen LAD-1 as a major auto-antigen in Caucasian patients affected by the lamina lucida type LABD.

MATERIALS AND METHODS

Patients

Sera were collected from 54 patients with LABD, including children and adults from different Italian clinical centres (Milan,

Pavia, Genoa, Florence and Rome) between 2012 and 2017, for retrospective study. All patients typically demonstrated clinical, histopathological and immunopathological features of LABD (32): (*i*) vesicular or bullous eruption involving skin and mucous membranes; (*ii*) sub-epidermal blisters infiltrated predominantly by neutrophils detected in lesion biopsies; (*iii*) linear IgA-deposit pattern at the BMZ detected by DIF. Patients with both IgA and IgG deposits at the BMZ detected by DIF were excluded. Due to the retrospective design of the study, information on drug-induce LABDs was not available for all patients.

Indirect immunofluorescence

IIF was performed on monkey oesophagus (MO) samples using commercial slides (BioSystems S.A. Barcelona, Spain). IIF-SSS was performed using a commercial kit (Immco Diagnostics, NY, USA). All sera were diluted in the ration 1:20 with phosphatebuffered saline (PBS). The slides were incubated with the diluted sera for 30 min. After washing in PBS, the slides were coated with fluorescein isothiocyanate-conjugated goat anti-human IgA or IgG (Kallestad Diagnostic, Chaska, MN, USA) for 30 min. Following a further PBS wash, the slides were mounted in buffered glycerine and examined under a fluorescence microscope (Leitz, Wetzlar, Germany). An IIF titre greater than 1:40 was considered positive.

Enzyme-linked immunosorbent assay for IgG and IgA detection

LABD IgG autoantibodies against BP180-NC16A and BP230 were characterized using commercial ELISA kit (MBL Co, Nagoya, Japan), according to the manufacturer's protocol. To compare the results obtained from different plates, optical densities (ODs) of the test samples were adjusted according to positive and negative control samples supplied in each kit. The protocol had been modified to an extent for the detection of IgA autoantibodies against BP180 and BP230. Briefly, BP180 and BP230 microwell strips were incubated with LABD patient sera diluted 50 times and appropriate control sera. After washing, the bound antibodies were detected using 10.000-fold diluted HRP-conjugated anti-human IgA antibody (Sigma-Aldrich, St Louis, MO, USA). Colour development was achieved using 3,3',5,5'-tetramethylbenzidine (Sigma) and after the reaction ceased, OD was measured at 450 nm with the correlation wavelength set at 620 nm using a microplate reader (Bio-Rad Laboratories Inc., Hercules, CA, USA). The cut-off value was determined to be the mean OD measured in a cohort of serum samples collected from 30 healthy volunteers and 3 standard deviations.

LAD-1 preparation and immunoblotting analysis

LAD-1, the 120-KDa soluble ectodomain of BP180, was isolated from the culture medium of normal keratinocytes derived from

human epidermis). Keratinocytes were grown at confluence and shed ectodomain of BP180 released in the medium was precipitated by the addition of ammonium sulphate (30% saturated solution) and incubated overnight at 4°C. Precipitates were collected, re-suspended in 25 mmol/l Tris-HCL, pH 7.8, 65 mmol/l NaCl and 5 mmol/LEDTA and dialysed against the same buffer overnight at 4°C. LAD-1 was electrophoresed on a 6% polyacrylamide gel in reducing conditions.

Immunoreactivity was detected after transfer to polyvinylidene difluoride membrane (Immobilon-P, Millipore Corporation, Billerica, MA, USA), by incubation with a 1:20 dilution of patient or control sera, followed by incubation with alkaline phosphatase-labelled secondary antibodies against IgA.

RESULTS

Study population

Serum samples were collected from 54 patients with LABD (12 children and 42 adults), 30 females and 24 males, with overall mean age 41 ± 31 years (age range 1–93 years).

Indirect immunofluorescence results

Presence of IgA at the BMZ was detected in 8 out of 54 (15%) serum samples from LABD patients, as tested by IIF on MO, while 34/54 (63%) serum samples from LABD patients were IgA-positive, as tested by IIF-SSS on human skin (**Table I**). In particular, 26/54 (48%), 1/54 (2%) and 7/54 (13%) serum samples were determined to have lamina lucida, sub-lamina densa and mixed patterns, respectively (Table I). Considering adults and children individually: serum samples collected from 6/12 (50%) children and 28/42 (67%) adults reacted with IIF-SSS.

With regards to IgG, none of the serum samples reacted with IIF on MO, while 1/54 (2%) serum samples (which was extracted from an adult) reacted to the epidermal side of IIF-SSS on human skin.

 Table I. Immunological test results in 54 patients with linear IgA

 bullous dermatosis (LABD)

Immunological test	n (%)
Indirect immunofluorescence (basement membrane zone), IgA	
Monkey oesophagus	8 (15)
Indirect immunofluorescence-salt-split skin, IgA	
Overall indirect immunofluorescence-salt-split skin IgA positive	34 (63)
Split skin exclusively epidermal side (lamina lucida)	26 (48)
Split skin exclusively dermal side (sub-lamina densa)	1 (2)
Split skin both sides	7 (13)
Indirect immunofluorescence (basement membrane zone), IgG	
Monkey oesophagus	0
Split skin exclusively epidermal side	1 (2)
Split skin exclusively dermal side	0
Split skin both sides	0
Immunoblotting, IgA	
LAD-1 (120kDa)	28 (52)
Enzyme-linked immunosorbent assay, IgA	
BP180-NC16A	17 (32)
BP230	14 (26)
Enzyme-linked immunosorbent assay, IgG	
BP180-NC16A	3 (6)
BP230	5 (9)

IgA reactivity profile

Overall, IB results indicated IgA reactivity to LAD-1 in 28/54 serum samples (52%) (Table I, Fig. 1C). Anti-LAD-1 IgA were detected by IB in 8/12 (67%) serum samples from children and from 20/42 (48%) serum samples adults. BP180-NC16A ELISA results were positive in 17/54 patients (32%) and BP230 ELISA results were positive in 14/54 patients (26%) (Table I).

Considering the IIF-SSS pattern groups, serum samples from patients with lamina lucida binding pattern had variable reactivity to different epidermal antigens, as shown in **Table II**. Some of the sera samples reacted with multiple epidermal antigens (Table II). Altogether, IgA from 24/26 (92%) samples binding to lamina lucida reacted with at least 1 of the epidermal antigens studied.

IgG reactivity profile

ELISA with BP180-NC16A and BP230 were positive for 3/54 (6%) and 5/54 patients (9%), respectively (Table I). Overall, circulating IgG antibodies anti-BP180-NC16A and/or anti-BP230 detected by ELISA were detected in 9/54 (17%) serum samples, while the total number of serum samples with IgG autoantibodies detected either by DIF, IIF-SSS or ELISA were 7/54 (13%).

DISCUSSION

These results confirm the heterogeneity of IgA autoantibody (autoAbs) targets in LABD. The predominant Ig class auto-antibody in the present cohort of patients with LABD was IgA, whereas IgG was present in only 22% of patients. Auto-reactive IgAs in serum samples primarily targeted LAD-1, while targeting BP180-NC16A and BP230 to a lesser extent.

The processed extracellular part of BP180 represents the primary antigenic stimulus in LABD, indicating that the processing induces neo-epitope formation (33). Extracellular regions of BP180 produced by removal of intra-cytoplasmic and transmembrane domains were reported to be recognized by 83% of LABD serum samples (26). Schumann et al. demonstrated that IgA targeted the

Table II. IgA reactivity profile in 34 indirect immunofluorescence
salt-split skin IgA-positive patients

Autoantigens	Epidermal side (lamina lucida) (n = 26) n (%)	Dermal side (sublamina densa) (n = 1) n (%)	Both sides (<i>n</i> = 7) <i>n</i> (%)		
Immunoblotting, IgA					
LAD-1(120kDa)	19 (73)	0	3 (43		
ELISA, IgA					
BP180-NC16A	10 (38)	0	1 (14)		
BP230	10 (38)	0	0		
Combined - reactivity profile					
LAD-1+BP180+BP230	5 (1)	0	0		
LAD-1 +BP180	1 (4)	0	1 (13)		
LAD-1 +BP230	1 (3.7)	0	0		

4/5 E. Cozzani et al.

soluble ectodomain more efficiently than the full-length protein in patients with LABD; 50% of the adult LABD serum samples and 100% of child LABD serum samples recognized this target (11). Likewise, in our experiments, 48% of adult LABD serum samples and 67% of child LABD serum samples reacted with LAD-1. In line with previous study, serum samples from children with LABD seem to particularly react with LAD-1 in comparison with adults; this finding could suggest a major role of neo-epitopes present in the LAD-1 antigen in the disease pathogenesis in children. However, due to the low number of patients in our cohort (8 patients) and in the cohort from Schumann's study (7 patients), no conclusion can be drawn. This issue should be investigated further on a larger cohort. In the present study, approximately 50% of the patients were positive for anti-LAD-1 IgA, and this proportion increased to 73% when only lamina lucida type patients were considered. Moreover, there was a strong correlation (90%) between lamina lucida positive sera and reactivity to BP180 (both LAD-1 and NC16A) and BP230. IgA positive against BP180-NC16A was detected consistently in a significant number (38%) of patients, which is in line with previous reports (6, 27, 28, 34). Notably, only 50% of these patients had targeted both LAD-1 and BP180-NC16A, while 50% exclusively targeted BP180-NC16A. Also, as anticipated, most patients positive for LAD-1 IgA (detected by IB) were negative for BP180-NC16A IgA (detected by ELISA). Notably, autoAbs directed against BP230 were more frequently detected in patients with LABD when BP180 autoAbs were also present. This association could depend on an intermolecular epitope spreading phenomenon (35, 36). Specifically, the pathological progression could induce damage in epithelial cells and result in abnormal exposure of the intracellular BP230, inducing the production of autoAbs (35). Undoubtedly, IB for 285 kDa antigen, LABD-97, full-length BP180 and type VII collagen might have vielded significant results; however, due to economic and technical limitations these tests were not conducted.

The concentration of circulating IgGs reactive to BMZ antigens was significantly low (17%). This is in sharp contrast with the results reported recently by Ohata et al., in which positive circulating IgG autoantibodies were detected either by IIF, IB or ELISA in 53% of their serum samples (6). The current study probably observed lower rates of auto-reactive circulating IgGs, as unlike the protocol adopted by Ohata et al., we did not test for IgGs directed against a wider variety of autoantigens. Considering LABD and BP to be opposite ends of the same spectrum, as suggested by Ohata et al. (6), might be inaccurate; however, these 2 diseases are overlap significantly in terms of clinical and serological aspects (37).

The higher positivity rates of IIF-SSS compared with IIF on MO confirmed the higher sensitivity of IIF-SSS in detecting auto-reactive IgAs. Our results confirm that the lamina lucida type is the most frequent variant of LABD. This is in line with a previous report suggesting that IgA reactivity is more prevalent on the epidermal end than on the dermal end of IIF-SSS (6).

The present study denotes that serological diagnosis of LABD patients can be performed using different assays for detection of IgG and IgA circulating autoantibodies (IIF, IB on LAD-1, BP180 and BP230 ELISAs). Notably, the combined sensitivity allows detection of autoantibodies in 83% of patients (45/54). Specifically, the most efficient process hierarchy involves DIF, followed by IIF-SSS as the most sensitive approach, followed by IB on LAD-1 and ELISA on IgA BP180, and IgA BP230 ELISA performed in case of negative results (which in one case was the only positive serological test). Notably, none of the LABD serum samples that resulted negative for circulating IgA antibodies in IIF, IB for LAD-1 or ELISA for BP180, were positive for IgG ELISAs.

This study has certain limitations; it focused specifically on the epidermal antigens targeted in LADB, and only a limited number of paediatric patients were studied.

Conclusion

LABD is characterized by a heterogeneous humoral response, largely comprising IgA auto-antibodies and a lesser number of IgG autoantibodies. It is yet to be established if these auto-antibodies elicit pathogenicity. Several antigens of the BMZ are targeted by auto-antibodies in LABD. LAD-1 is a major auto-antigen of the lamina lucida subtype, particularly in children. DIF remains the most preferred diagnostic method for detection of LABD. However, the combination of several serological assays could an important approach to support diagnosis, especially in cases in which a biopsy is not feasible or the DIF results are negative.

REFERENCES

- Zone JJ. Clinical spectrum, pathogenesis and treatment of linear IgA bullous dermatosis. J Dermatol 2001; 28: 651–653.
- Horváth B, Niedermeier A, Podstawa E, Müller R, Hunzelmann N, Kárpáti, et al. IgA autoantibodies in the pemphigoids and linear IgA bullous dermatosis. Exp Dermatol 2010; 19: 648–653.
- Zambruno G, Manca V, Kanitakis J, Cozzani E, Nicolas JF, Giannetti A. Linear IgA bullous dermatosis with autoantibodies to a 290 kd antigen of anchoring fibrils. J Am Acad Dermatol 1994; 31: 884–888.
- Tsuchisaka A, Ohara K, Ishii N, Nguyen NT, Marinkovich MP, Hashimoto T. Type VII collagen is the major autoantigen for sublamina densa-type linear IgA bullous dermatosis. J Invest Dermatol 2015; 135: 626–629.
- Utsunomiya N, Chino T, Oyama N, Utsunomiya A, Yamaguchi Y, Takashima W, et al. Sublamina densa-type linear IgA bullous dermatosis with IgA autoantibodies specific for type VII collagen: a case report and clinicopathological review of 32 cases. Dermatol Online J 2017; 15: 23.
- 6. Ohata C, Ishii N, Koga H, Nakama T. A clinical and serological study of linear IgA bullous dermatosis without linear immunoglobulin deposition other than IgA at the basement membrane zone using direct immunofluorescence. Br J Der-

matol 2017; 177: 152-157.

- 7. Wojnarowska F, Whitehead P, Leigh IM, Bhogal BS, Black MM. Identification of the target antigen in chronic bullous disease of childhood and linear IgA disease of adults. Br J Dermatol 1991; 124: 157–162.
- Ghohestani RF, Nicolas JF, Kanitakis J, Claudy A. Linear IgA bullous dermatosis with IgA antibodies exclusively directed against the 180- or 230-kDa epidermal antigens. J Invest Dermatol 1997; 108: 854–858.
- 9. Collier P, Wojnarowska F, Allen J, Kirtschig G. Molecular overlap of the IgA target antigens in the subepidermal blistering diseases. Dermatology 1994; 189: 105–107.
- Marinkovich MP, Taylor TB, Keene DR, Burgeson RE, Zone JJ. LAD-1, the linear IgA bullous dermatosis autoantigen, is a novel 120-kDa anchoring filament protein synthesized by epidermal cells. J Invest Dermatol 1996; 106: 734–738.
- Schumann H, Baetge J, Tasanen K, Wojnarowska F, Schäcke H, Zillikens D, et al. The shed ectodomain of collagen XVII/ BP180 is targeted by autoantibodies in different blistering skin diseases. Am J Pathol 2000; 156: 685–695.
- Zone JJ, Taylor TB, Kadunce DP, Meyer LJ. Identification of the cutaneous basement membrane zone antigen and isolation of antibody in linear immunoglobulin A bullous dermatosis. J Clin Invest 1990; 85: 812–820.
- Zone JJ, Taylor TB, Kadunce DP, Chorzelski TP, Schachner LA, Huff JC, et al. IgA antibodies in chronic bullous disease of childhood react with 97 kDa basement membrane zone protein. J Invest Dermatol 1996; 106: 1277–1280.
- Dmochowski M, Hashimoto T, Bhogal BS, Black MM, Zone JJ, Nishikawa T. Immunoblotting studies of linear IgA disease. J Dermatol 1993; 6: 194–200.
- Pas HH, Kloosterhuis GJ, Heeres K, van der Meer JB, Jonkman MF. Bullous pemphigoid and linear IgA dermatosis sera recognize a similar 120-kDa keratinocytes collagenous glycoproteins with antigenic cross-reactivity to BP180. J Invest Dermatol 1997; 108: 423–429.
- Hirako Y, Usukura J, Uematsu J, Hashimoto T, Kitajima Y, Owaribe K. Cleavage of BP180, a 180 kDa bullous pemphigoid, yields a 120 kDa collagenous extracellular polypeptide. J Biol Chem 1998; 273: 9711–9717.
- Schäcke H, Schumann H, Hammami-Hauasli N, Raghunat M, Bruckner-Tuderman L. Two forms of collagen XVII in keratinocytes. A full-length transmembrane protein and a soluble ectodomain. J Biol Chem 1998; 273: 25937–25943.
- Zone JJ, Taylor TB, Meyer LJ, Petersen MJ. The 97-kDa linear IgA bullous disease antigen is identical to a portion of the extracellular domain of the 180 kDa bullous pemphigoid antigen, BPAG2. J Invest Dermatol 1998; 110: 207–210.
- Hashimoto T, Ishii N, Tsuruta D. Production of neoepitopes by dynamic structural changes on BP180/Type XVII collagen. J Invest Dermatol 2017; 137: 2462–2464.
- Franzke C-W, Tasanen K, Schäcke H, Zhou Z, Tryggvason K, Mauch C, et al. Transmembrane collagen XVII, an epithelial adhesion protein, is shed from the cell surface by ADAMs. EMBO J 2002; 21: 5026.
- 21. Schmidt E, Wehr B, Tabengwa EM, Reimer S, Brocker E-B, Zillikens D. Elevated expression and release of tissue-type, but not urokinase-type, plasminogen activator after binding of autoantibodies to bullous pemphigoid antigen 180 in cultured human keratinocytes. Clin Exp Immunol 2004; 135: 497.
- 22. Verraes S, Hornebeck W, Bernard P, Polette M, Borradori L.

Respective contribution of neutrophil elastase and matrix metalloproteinase 9 in the degradation of BP180 (type XVII collagen) in human bullous pemphigoid. J Invest Dermatol 2001; 117: 1091.

- Hirako Y, Nishizawa Y, Sitaru C, Opitz A, Marcus K, Meyer HE, et al. The 97-kDa (LABD97) and 120-kDa (LAD-1) fragments of bullous pemphigoid antigen 180/type XVII collagen have different N-termini. J Invest Dermatol 2003; 121: 1554.
- Hofmann SC, Voith U, Schönau V, Sorokin L, Bruckner-Tuderman L, Franzke C-W. Plasmin plays a role in the in vitro generation of the linear IgA dermatosis antigen LADB97. J Invest Dermatol 2009; 129: 1730.
- 25. Izumi K, Nishie W, Mai Y, Wada M, Natsuga K, Ujiie H, et al. Autoantibody profile differentiates between inflammatory and noninflammatory bullous pemphigoid. J Invest Dermatol 2016; 136: 2201.
- 26. Csorba K, Schmidt S, Florea F, Ishii N, Hashimoto T, Hertl M, et al. Development of an ELISA for sensitive and specific detection of IgA autoantibodies against BP180 in pemphigoid diseases. Orphanet J Rare Dis 2011; 6: 31.
- Zillikens D, Herzele K, Georgi M, Schmidt E, Chimanovitch I, Schumann H, et al. Autoantibodies in a subgroup of patients with linear IgA disease react with the NC16A domain of BP180. J Invest Dermatol 1999; 113: 947–953.
- Schmidt E, Herzele K, Schumann H, Wesselmann U, Chimanovitch I, Bruckner-Tuderman L, et al. Linear IgA disease with circulating IgA antibodies against the NC16A domain of BP180. Br J Dermatol 1999; 140: 964–966.
- Bhogal BS, Wojnarowska F, Marsden RA, Das A, Black MM, McKee PH. Linear IgA bullosus dermatosis of adults and children: an immunoelectron microscopic study. Br J Dermatol 1987; 117: 289–296.
- Prost C, De Leca AC, Combemale P, Labeille B, Martin N, Cosnes A, et al. Diagnosis of adult linear IgA dermatosis by immunoelectron microscopy in 16 patients with linear IgA deposits. J Invest Dermatol 1989; 92: 39–45.
- Wojnarowska F, Collier PM, Allen J, Millard PR. The localization of the target antigens and antibodies in linear IgA disease is heterogeneous, and dependent on the methods used. Br J Dermatol 1995; 132: 750–757.
- Egan CA, Zone JJ. Linear IgA bullous dermatosis. Int J Dermatol 1999; 38: 818–827.
- Toyonaga E, Nishie W, Izumi K, Natsuga K, Ujiie H, Iwata H, et al. C-terminal processing of collagen XVII induces neoepitopes for linear IgA dermatosis autoantibodies. J Invest Dermatol 2017; 137: 2552–2559.
- 34. Lin MS, Fu CL, Olague-Marchan M, Hacker MK, Zillikens D, Giudice GJ, et al. Autoimmune responses in patients with linear IgA bullous dermatosis: both autoantibodies and T lymphocytes recognize the NC16A domain of the BP180 molecule. Clin Immunol 2002; 102: 310–319.
- 35. Di Zenzo G, Thoma-Uszynski S, Calabresi V, Fontao L, Hofmann SC, Lacour JP, et al. Demonstration of epitopespreading phenomena in bullous pemphigoid: results of a prospective multicenter study. Invest Dermatol 2011; 131: 2271–2280.
- Didona D, Di Zenzo G. Humoral epitope spreading in autoimmune bullous diseases. Front Immunol 2018; 9: 779.
- Antiga E, Caproni M, Fabbri P. Linear immunoglobulin a bullous dermatosis: need for an agreement on diagnostic criteria. Dermatology 2013; 226: 329–332.

<u>cta **D**erma</u>to-Venereologica