

1 **Life history stage-specific diet shifts by dung beetles revealed using DNA metabarcoding**

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16 **Author's contributions:** GIHK, ML, PT and EC conceived and designed the study, collected and
17 analysed the data and wrote the manuscript; GFF, FB, AB, DR extracted and amplified DNA and
18 contributed to the writing of the paper.

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30 **Abstract**

31 Life history changes typically lead to changes in resource use. Such shifts are not well understood
32 in the dung beetles, despite recognized differences in larval and adult feeding ability. We use the
33 flightless dung beetle *Circellium bacchus* to explore such shifts, identifying dung sources of adults
34 using DNA metabarcoding and compare this with published accounts of larval dung sources. *C.*
35 *bacchus* is traditionally considered to specialise on the dung of large herbivores for both larval and
36 adult feeding. We successfully extracted mammal DNA from 151 adult *C. bacchus* samples (out of
37 172 collected), this representing 16 mammal species (ranging from elephants to small rodents),
38 many of which are hitherto undescribed in the diet. Adult *C. bacchus* showed clear dung source
39 preferences, especially for large herbivores inhabiting dense-cover vegetation. Our approach also
40 confirmed the presence of cryptic taxa in the study area, and we propose this may be used for
41 biodiversity survey and monitoring purposes. Murid rodent feces were the most commonly fed-
42 upon dung source (77.5%) for adult *C. bacchus*, differing markedly from the large and
43 megaherbivore dung sources used for larval rearing. These findings support the hypothesis of life
44 history specific shifts in resource use in dung beetles, and reveal a hitherto unsuspected, but
45 ecologically important, role of these dung beetles in consuming rodent feces. The differences in
46 feeding abilities of the larval and adult life history stages have profound consequences for their
47 resource use and foraging strategies, and hence the ecological role of dung beetles. This principle
48 and its ecological consequences should be explored in other scarabaeid dung beetle species.

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50 **Keywords:** biodiversity survey, *Circellium bacchus*; coprophagy, environmental DNA, next
51 generation sequencing, megaherbivores, mitochondrial DNA, Scarabaeinae, rodent feces.

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58 **Introduction**

59 Ontogenetic shifts give rise to ecological shifts, particularly in species with complex life histories
60 (Werner 1988), and understanding such shifts is important to understanding the ecological role of
61 these species. This is particularly relevant given that more than 80% of animal species show such
62 ontogenetic shifts (Werner 1988). The massively diverse (nearly 6 000 species) and ecologically
63 important scarabaeid dung beetle family exhibits complex life histories, with changes in the feeding
64 ability of the different life history stages. Thus, while the larvae are typically able to ingest and
65 digest relatively coarse plant fragments from the dung of herbivores, the adults' mouth parts
66 constrain them to ingesting liquid and fine (< 130 µm) particulate matter (Holter 2016). However,
67 the use of dung by dung beetles, and by extension their contribution to the detrital food chain and
68 nutrient cycling, has focused almost exclusively on those sources used for larval rearing, with
69 limited attention to the dung sources of adults. Here, we use Africa's largest (up to 50 mm in
70 length) telecoprid species, the flightless *Circellium bacchus*, and DNA metabarcoding, to identify
71 the diet sources of adults and compare this with published accounts of larval dung resources
72 (Kryger et al. 2006).

73 The dung sources of *C. bacchus*, for adult consumption and brood ball construction, has
74 been described anecdotally or estimated using opportunistic observations of beetles feeding on or
75 preparing dung balls from various sources. The anecdotal information is that flightless dung
76 beetles rely on elephant *Loxodonta africana*, Cape buffalo *Syncerus caffer* and black rhinoceros
77 *Diceros bicornis* dung, and based on this, Chown *et al.* (1995) concluded that this species depends
78 on black rhinoceros dung for its persistence. Kryger *et al.* (2006) observed flightless dung beetles
79 consuming dung of elephant, buffalo, rhino, "various antelope", monkey *Chlorocebus pygerythrus*,
80 human, hare *Lepus* sp. and ostrich *Struthio camelus*. This sampling was not systematic in terms of
81 availability of different dung sources. Using a limited cafeteria-style experiment, Kryger et al.
82 (2006) also estimated dung source preferences by *C. bacchus* for adult feeding and brood ball
83 construction, showing that dung preferences varied among elephant, black rhinoceros, buffalo and
84 cattle *Bos taurus*. Kryger *et al.* (2006; p. 201) concluded there is "distinct preference for feeding on
85 elephant dung early in the morning" and that "cattle/buffalo dung was preferred later in the day".

86 For brood ball material, bovids (buffalo and cattle) were apparently preferred over megaherbivores
87 (elephant and rhino; Kryger et al. 2006). Based on the above, there are two contrasting views with
88 regard to the diet resource use of this species: it is either a large mammalian herbivore specialist
89 or is a mammalian generalist.

90 DNA metabarcoding is increasingly used to identify taxa in sampled material, such as plant
91 species in the dung of herbivores, prey species in the gut contents of carnivores or in soil samples
92 (review in Pompanon et al. 2012; Shehzad et al. 2012; Taberlet et al. 2012a; Yoccoz et al. 2012;).
93 Animal dung includes not only the DNA of ingested food items but also DNA from the animal
94 providing the dung (Shehzad et al. 2012; De Barba et al. 2014). Thus, we expected adult *C.*
95 *bacchus* to ingest DNA from the animal species on whose dung they had fed, and that this would
96 be present in the feces and could be used to identify the source of the dung. We used this
97 approach to test the hypotheses that the flightless dung beetle is a large herbivore dung specialist
98 or alternatively is a generalist, and that adult beetle diets differ from that reported for the larvae (i.e.
99 the sources used for brood ball construction; from Kryger et al. 2006). We demonstrate substantial
100 shifts in resource use and preference by the adults compared to the larvae, this despite the fact
101 that adults are responsible for the acquisition of the material for both adults and larvae. Our
102 findings confirm the detection of dung source DNA in dung beetle feces, represent the first
103 systematic survey of this dung beetle's adult diet and provide novel insights into dung beetle diet
104 preferences. Importantly, we provide evidence for life history level shifts in diet and reveal a cryptic
105 functional role for this species.

106

107 **Study design**

108 Sampling

109 Flightless dung beetles were sampled in the Main Camp and Colchester sections
110 (collectively 26 500 ha in area) of the Addo Elephant National Park, South Africa, between January
111 and February 2014, a period of high dung beetle activity (Kryger et al. 2006). *C. bacchus* is within a
112 monotypic genus of uncertain taxonomic position and is unusual by virtue of its flightlessness and
113 strict ectothermy (Chown et al. 1995; Davis et al. 2008a). The fragmented status of the population,

114 apparent contraction in distribution range and slow reproduction (Chown et al. 1995) has led to
115 suggestions that this species should be considered threatened (Kryger et al. 2006). Although
116 protected in some conservation areas, most notably the AENP, tourist activities represent an
117 additional threat through roadkills (Hayward et al. 2010). These attributes have led to this beetle
118 attracting scientific interest and conservation concern, as well as achieving charismatic fauna
119 status for wildlife viewing among tourists (Kerley et al. 2003) and legal protection; these latter two
120 achievements being uncommon among terrestrial invertebrates.

121 The AENP is about 60 km north east of Port Elizabeth on the south east coast, annual
122 rainfall is 450 mm pa, with temperatures varying between summer maxima of ca. 32°C and winter
123 minima of ca. 5°C (Weather SA). The AENP is recognised as supporting the largest population of
124 flightless dung beetles (Kryger et al. 2006). In addition, the Main Camp and Colchester sections
125 (which form a discrete, fenced unit) support a wide diversity of mammals (52 species, excluding
126 volant and fossorial species; Swanepoel 1975; Boshoff et al. 2002; Hayward et al. 2007), most
127 prominent among the herbivores being elephant, black rhino and buffalo, while the apex predators
128 are represented by lions *Panthera leo*, leopards *P. pardus*, and spotted hyaena *Crocuta crocuta*.
129 There is also a diverse avian and reptile fauna.

130 Dung beetle sampling comprised locating individuals active on roads or trails across the
131 study area (irrespective of habitat type), i.e. sampled beetles were not associated with dung balls
132 or dung. When picked up, dung beetles either defecate within about 5 seconds or take much
133 longer (pers obs). Fecal samples from beetles that defecated on being picked up were wrapped in
134 Kimwipes paper (Kimberly-Clark) and immediately placed into labelled plastic vials containing silica
135 gel. Dung beetles that did not defecate were released within 30 seconds of being picked up and
136 not sampled. The fecal samples were preserved dry in silica gel until DNA extraction. It was difficult
137 to prevent possible human contamination during the sampling, i.e. we did not wear gloves and
138 facial masks. Instead we elected to remove such potential contamination at the data analysis
139 stage.

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143 DNA extraction, amplification and sequencing

144 DNA extractions were carried out using a phosphate buffer protocol, modified from Taberlet *et al.*
145 (2012b). Each fecal sample was put in an Eppendorf tube containing 500 μL of saturated
146 phosphate buffer (Na_2HPO_4 ; 0.12 M; pH 8), and shaken gently for 15 min (45 rpm). The resulting
147 mixture was centrifuged at 11 000 g for 10 min. The next steps were performed using the
148 NucleoSpin[®] Soil kit (Macherey-Nagel, Düren, Germany) following the manufacturer's instructions,
149 and skipping the lysis steps. Four hundred μL of the supernatant was added to 250 μL of SB
150 buffer, loaded onto the extraction column, and washed once with SB and SW1 buffers, and twice
151 with SW2 buffers. The elution was done with 100 μL of SE buffer. A negative extraction control
152 was included into each batch of 23 dung beetle fecal samples, using the phosphate buffer as
153 starting material.

154 For DNA amplifications, we used a primer pair targeting a short but informative fragment of
155 the 16S mitochondrial gene of mammals (Giguet-Covex *et al.* 2014). The forward and reverse
156 primer sequences are 5'-CGAGAAGACCCTATGGAGCT-3' and 5'-CCGAGGTCRCCCAACC-3',
157 respectively. To discriminate samples and PCR replicates after sequencing, both forward and
158 reverse primers were tagged with 8-nucleotide labels (hereafter designated as "tag") with at least
159 three nucleotide differences among each of them. Furthermore, three additional random
160 nucleotides were added on the 5'-end of each primer, in order to allow efficient detection of the
161 different clusters during the sequencing step. For each sample and each replicate, the same tag
162 was used on both primers, i.e. on both sides of the PCR product (Schnell *et al.* 2015, Taberlet *et*
163 *al.* 2018).

164 Two PCRs per sample and per control were carried out, including the fecal sample extracts,
165 the extraction negative controls, the PCR negative controls, and the PCR positive controls (DNA
166 extract from *Didelphis marsupialis*). We used the AmpliTaq Gold[®] 360 Master Mix (Applied
167 Biosystems[™], Foster City, CA, USA), in a final volume of 20 μl containing 2 μl of DNA extract
168 (including the extraction negative controls), 0.2 μM of each primer, and 0.16 μL of bovine serum
169 albumin (BSA, Roche Diagnostic, Basel, Switzerland). To reduce the amplification of human DNA,

170 we added a human blocking oligonucleotide (5'-CCACCGAAATTTTAAATGCAGGTTTGGTAGTT-
171 C3-3') in each PCR, at a final concentration of 2 μ M. The design of this blocking oligonucleotide
172 was done according to Vestheim & Jarman (2008). The PCR cycling parameters were: 10 minutes
173 at 96°C for activating the polymerase, and then 45 cycles with denaturation for 30 s at 96°C,
174 annealing for 30 s at 50°C, elongation for 60 s at 72°C, with a final extension for 420 s. All PCR
175 products, including samples and controls, were mixed together and purified (MinElute™ PCR
176 purification kit, Qiagen, Hilden, Germany). The library preparation and the sequencing was
177 outsourced (Fasteris SA, Geneva, Switzerland). The library was prepared using the MetaFast
178 protocol (www.fasteris.com/metafast) and the sequencing carried out on the HiSeq 2500
179 sequencing platform (Illumina, San Diego, CA, USA) with a paired-end approach (2 x 125 bp).

180

181 Sequence data analysis

182 The sequence reads were analyzed using OBITools (Boyer et al. 2016). First, the direct and
183 reverse reads corresponding to a single molecule were aligned and merged using the
184 *illuminapairedend* program, taking into account data quality during the alignment and the
185 consensus computation. Primers and tags were then identified using the *ngsfilter* program. Only
186 the amplified region of the sequences with a perfect match on tags and a maximum of two errors
187 on primers were recorded for the subsequent analysis, keeping the information about sample
188 names. Strictly identical sequences were clustered together using the *obiuniq* program, keeping
189 the information about their distribution among samples. Sequences shorter than 60 bp or longer
190 than 90 bp, or with occurrence lower than 1000 in the whole dataset were excluded using the
191 *obigrep* program. Potential PCR/sequencing errors were identified and removed using the *obiclean*
192 program. We kept only sequences identified at least once as "head" or "singleton" in the different
193 PCRs (i.e. "head" are sequences that are at least twice as abundant as other sequences differing
194 by a single change, "singleton" are sequences that have no other sequences differing by a single
195 change; see Boyer et al. 2016 for further explanations). Taxon assignment was achieved using the
196 *ecotag* program. The reference database for the taxonomic assignment was built by extracting the
197 relevant part of the mitochondrial 16S gene from EMBL nucleotide library (release 126) using the

198 *ecoPCR* program (Ficetola et al. 2010). All sequences with a best identity lower than 0.86 when
199 compared to any sequence in the reference database were removed, as they potentially
200 correspond to chimeras or to non-specific amplifications (Taberlet et al. 2018). Mammalian DNA
201 was considered as present in a scat sample if at least one of two PCR replicates showed more
202 than 100 sequence reads.

203 Finally, we inspected the automatic taxonomic assignments of sequences manually and
204 considered species-level identities reliable only if these matched near-perfectly ($\geq 98\%$ identity) to
205 a single species in the reference database. Close, but non-identical, matches (88-98% identity)
206 were consistently made at the genus level, and checked against the occurrence of these taxa in
207 the study area.

208

209 Herbivore dung production

210 Estimates of large herbivore dung production were derived from the SANParks mammal census
211 data for 2013 for the Main Camp and Colchester sections of the AENP, derived from systematic
212 aerial counts (SANParks Unpublished data). These dung production estimates were calculated for
213 each censused megaherbivore and ungulate species, as well as the undetected blue duiker
214 *Philantomba monticola* (for a total of 11 large herbivore species). Dung production estimates were
215 based on mass specific models of herbivore food intake (Owen-Smith 1992), based on $\frac{3}{4}$ adult
216 female body mass for each species (Hayward and Kerley 2005), adjusted for ruminant/hindgut
217 fermenter digestive efficiency (Owen-Smith 1992).

218

219 Statistical analysis

220 Dung beetle diet was described as the frequency-of-occurrence of diet sources across fecal
221 samples at genus or species level. To assess the adequacy of our sample sizes, we generated an
222 accumulation curve (with 50 random resamplings) of diet sources recorded per sample (Online
223 Resource 1). However, because this curve did not reach a clear asymptote, we estimated the total
224 number of diet sources with a non-parametric species richness estimator (Foggo et al. 2003).

225 Differences between observed and expected counts provided an estimate of the variation in diet
226 information at the upper-limit of sampling effort.

227 Large herbivore dung source preference by flightless dung beetles was estimated using
228 Jacob's index (Jacobs 1974) based on the proportion of estimated dung production and proportion
229 of records of dung beetle consumption of the dung for each of the censused herbivore species.
230 Jacob's index varies between + 1, for maximum preference (i.e. where dung consumption is
231 greater than dung production), and – 1, for maximum avoidance (i.e. where consumption is less
232 than production). Preferences were calculated for each taxon, and these data were used to
233 calculate Jacob's index for the digestive morphology guilds (ruminants, hindgut fermenters),
234 feeding guilds (grazers, browsers, mixed feeders) and habitat use guilds (open habitat, closed
235 habitat, mixed use of open and closed – see Online Resource 2 for guild data).

236

237 **Results**

238 DNA metabarcoding results

239 We analyzed a total of 172 dung beetle fecal samples, together with eight extraction negative
240 controls using phosphate buffer as starting material, four extraction negative controls using
241 Kimwipes paper as starting material, three PCR negative controls, and four PCR positive controls.
242 After the *illumina-paired-end* and *ngsfilter* programs (assembling forward and reverse reads, and
243 identifying primers and tags), we obtained a total of 3,967,490 sequences. The dereplication
244 yielded 163,798 different sequences. Removing the sequences occurring only once in the whole
245 dataset decreased the number of different sequences to 48,189. The automatic filtering and
246 taxonomic assignment described in Study design yielded 68 molecular operational taxonomic
247 units. The obvious contaminants were then removed. Among these, human and human-related
248 sequences represented about 35% of the dataset at this stage. This level of human contamination
249 was expected according to our sampling protocol (see Study design). Additionally, we observed a
250 few cow *Bos taurus* and pig *Sus scrofa* sequences, these being known contaminants in PCR kits
251 (Leonard et al. 2007) and not occurring in the study site. We also obtained red deer *Cervus*
252 *elaphus* contamination in a single replicate of five dung beetle fecal samples. This contamination

253 most probably comes from the hundreds of red deer scats that were extracted the day before in the
254 same laboratory, as there are no known cervid populations in or around AENP. After removing
255 human, pig, cow and red deer contaminants, the number of MOTUs decreased to 37. A final
256 manual inspection of the remaining sequences yielded 16 putative mammalian species distributed
257 among the 151 dung beetle fecal samples that produced usable sequences (see Online Resource
258 3). With the exception of two genera of the Murid family (*Micaelamys* and *Otomys* - the latter
259 following the monogeneric treatment of the Otomyini (Taylor et al. 2004), and one member of the
260 Bovidae (*Cephalophus*) that could only be identified to genus level, the majority of diet sources
261 were identified to species-level.

262 These 16 taxa accounted for 86.5% of the variation in dung beetle dietary information at the
263 upper limit of sampling effort. This suggests that our sample size was appropriate to describe the
264 sources of the diet.

265

266 Dung source use

267 Despite the marker also potentially amplifying bird, reptile and amphibian DNA, we detected only
268 mammal DNA in *C. bacchus* feces. The identified mammal diet sources ranged in body size from
269 the elephant (2 000 - 6 000 kg) to the 43 g striped field mouse *Rhabdomys pumilio* (Fig. 1), and
270 spanned 6 taxonomic orders. Murid rodents provided 77.5% of the diet sources. In terms of broad
271 feeding guilds, these dung source taxa were dominated by herbivores, with only a single record of
272 one carnivore species' (*Canis mesomelas*) scat being consumed. The DNA sequences from dung
273 beetle feces indicate the presence of some previously unrecorded taxa in the study area. These
274 include a third species of *Otomys* (only two are considered confirmed for the study area;
275 Swanepoel 1975), and a record for the duiker genus *Cephalophus*. It is noteworthy that one of the
276 dung sources species, the common warthog *Phacochoerus africanus*, is an introduced species that
277 is now considered invasive in the AENP and surrounds (Mgqatsa 2010). These records represent
278 only 27% of the 52 non-volant, non-fossorial mammal species currently recorded as occurring in
279 the study area, indicating selectivity of dung source use by *C. bacchus*. Contrary to expectations,
280 the frequency of dung source use does not increase with body size for the 16 taxa included in the

281 analyses ($R^2 = 0.02$, $F_{1,14} = 0.27$, $P = 0.613$; % Frequency of occurrence = $9.11 - (1.20 * \log(\text{body}$
282 mass (kg))).

283

284 Dung source preferences

285 The flightless dung beetle showed clear preferences or avoidance for dung sources among the
286 large herbivore species for which we have both dung production and consumption estimates (Fig.
287 2). The traditionally considered important or preferred sources of dung (elephant, black rhino,
288 buffalo) were all avoided, while the dung of three smaller bovids and the two suids (the latter
289 including the invasive warthog) were preferred, but that of three other large bovids was avoided
290 (Fig. 2; Online Resource 2).

291 Dung source preference was negatively related to body size for the 11 large herbivore
292 species for which preference data were available ($R^2 = 0.52$, $F_{1,9} = 9.75$, $P=0.013$; $D = 1.63-(0.74 *$
293 $\log(\text{body mass (kg)})$). In terms of the digestive morphology guilds, ruminants were marginally
294 preferred ($D = 0.19$) over hindgut fermenters ($D = - 0.19$), although there was a roughly equal
295 distribution of both guilds in either the preference or avoidance category (c.f. Fig. 2 and Online
296 Resource 2). The dung of the browsing guild was consistently preferred ($D = 0.52$), while that of
297 the mixed feeders ($D = - 0.31$) and the grazers ($D = - 0.08$) were avoided. The clearest patterns
298 were among guilds of species characteristic of different habitats (Fig. 2): dung of species that use
299 dense, closed vegetation was consistently preferred ($D = 0.63$), while that of open habitat species
300 was consistently avoided ($D = - 0.86$). The dung of species that show mixed use of open and
301 dense habitats was avoided ($D = - 0.29$), although this was not always the case (Fig. 2).

302

303 **Discussion**

304 The findings we present emerge from a novel approach for exploring the functional role of dung
305 beetles, and support the hypothesis of DNA material from the dung source occurring in dung beetle
306 feces. DNA metabarcoding has rarely been applied to describing the diet of invertebrates based on
307 feces (but see Ibanez et al. 2013; Gomez and Kolokotronis 2016; Kaunisto et al. 2017; Rodgers et
308 al. 2017) and can clearly be applied to identifying diet sources for coprophagous taxa, as well as

309 detritivores. DNA metabarcoding can also be applied to the identification of the source of fecal
310 material, and hence to identifying the source of dung balls, even those not associated with an
311 identifiable dung deposit. Based on this, it is clear that DNA metabarcoding-based studies have the
312 potential to considerably expand our understanding of the functional role of species that are
313 otherwise difficult to study. Furthermore, DNA metabarcoding also allowed the possible
314 identification of two taxa apparently unrecorded at the study site, the third *Otomys* species and the
315 *Cephalophus*, although we could not resolve which species these represent. The former is not
316 surprising, as de Graaff (1974) recorded *O. angoniensis*, in addition to *O. irroratus* and *O.*
317 *unisulcatus*, but this was later discounted (Swanepoel and Branch 1982) due a lack of further
318 records. In contrast, the natural range limit of the closest *Cephalophus* is at least 1000 km north-
319 east of the study site. A possible explanation for this record is that representatives of this genus
320 may have been introduced by neighbouring landowners, with escapees subsequently invading the
321 AENP. Both these possibilities require further testing.

322 The DNA metabarcoding of dung beetle feces also served to detect some taxa in this
323 national park, whose presence is missed by conventional census strategies. Thus, *Philantomba*
324 *monticola*, the blue duiker is a small, dense vegetation-dwelling antelope known to occur in the
325 AENP, and was not recorded in the aerial census (SANParks Unpublished data), but was detected
326 in the dung beetle diet. This detection of cryptic species may also extend to identifying species that
327 may be absent. The Cape grysbok *Raphicerus melanotis*, although recorded in the AENP
328 historically (Penzhorn 1971), was not detected in the aerial census or the dung beetle diet. Given
329 that this species is characteristic of dense cover and uses latrines (Kerley et al. 2010), its dung
330 may be expected to serve as an attractive food source (see later). Its absence from both survey
331 approaches calls into question its persistence in the area. Based on this, we suggest that DNA
332 metabarcoding of dung beetle feces can be used as an efficient and cost effective biodiversity
333 survey and monitoring tool, as proposed for DNA extracted from leeches (family Haemadipsidae;
334 Schnell et al. 2012) and carrion flies (families Calliphoridae and Sarcophagidae; Calvignac-
335 Spencer et al. 2013).

336 The dung sources consumed by adult *C. bacchus* presented here differ substantially from
337 published findings, which have emphasized the importance of megaherbivore dung. In particular,
338 the absence of any records of *C. bacchus* feeding on the dung of the black rhinoceros conflicts
339 with speculation by Chown *et al.* (1995) that *C. bacchus* was dependent on the black rhinoceros for
340 a reliable dung source. These differences may reflect the lack of systematic approaches in
341 previous studies, the emphasis on studying dung beetles at large dung sources, or
342 misidentification of the dung sources. In addition to these sampling artifacts, the variation in
343 observed dung use may reflect life history level variation in the use of dung by *C. bacchus*. These
344 issues are expanded on below.

345 The high levels of elephant dung production in the AENP (over 50% of estimated large
346 herbivore dung production – Online Resource 4) means that sampling *C. bacchus* dung use based
347 on non-stratified sampling of dung sources (as apparently done by Kryger *et al.* (2006), and the
348 various anecdotal descriptions of dung use by *C. bacchus*) would result in an overestimate of the
349 importance of particularly this source for dung beetles. Clearly, well-designed, systematic sampling
350 is needed to reliably assess resource use in cases where resources show such overbearing
351 variances in availability. Furthermore, most published studies of resource use by *C. bacchus* have
352 been focussed on observing beetles forming either brood or feeding balls from dung piles (e.g.
353 Kryger *et al.* 2006). A consequence of this approach (in addition to the resource availability bias
354 raised above) is that the use of smaller fecal deposits (e.g. rodent droppings) would be completely
355 overlooked, as such deposits would rarely be substantial enough to form feeding or brood balls. A
356 corollary of this is that the use of rodent droppings by *C. bacchus* may have been entirely
357 overlooked in the past as observers were not sensitive to the need to monitor *C. bacchus* feeding
358 behaviour when not engaged in dung ball formation. Finally, a further sampling artefact may arise
359 due to the misidentification of dung being used by *C. bacchus*, although the extent of this is
360 typically not known. The approach used in the present study, however, avoided this risk.

361 A more interesting functional explanation of these differences may lie in the understanding of
362 the differences in feeding strategies of the dung beetle adult and larval life history stages. Larvae
363 are able to ingest and digest cellulose-rich material from the coarse particulate matter in dung

364 balls, which are provided by the adults (Davis et al. 2008b). In contrast, adults are constrained in
365 their ability to ingest coarse material, and rely instead on ingesting fluid and extremely fine
366 particulate matter for their nutrition (Holter 2000). Thus, the brood ball preparation and adult
367 nutritional requirements place differing constraints on the use of dung resources. The former has a
368 limit in terms of the minimum amount of dung required to form a brood ball - these range from 22-
369 85 g of dry mass (e.g. Kryger et al. 2006). In contrast, adult feeding does not have such a volume
370 constraint, and fluid content, particle size and the C:N ratio is more important (Holter 2016). Based
371 on this, we hypothesize that *C. bacchus* exhibits different foraging strategies, depending on
372 whether the focus is on brood ball formation or adult feeding. Thus, for brood ball formation the
373 dung source must be large, and foraging *C. bacchus* need to locate dung from species that either
374 have large dung boli (e.g. elephant, buffalo) or that use latrines (e.g. black rhino). In this context, *C.*
375 *bacchus* can be hypothesized to be following a quantity maximization strategy in their dung source
376 preferences for brood ball preparation. This would explain the published focus on the use of the
377 dung of these taxa by *C. bacchus*. In contrast, feeding adult *C. bacchus*, not having such a volume
378 constraint, may either adopt a quality maximization strategy or a time/effort minimization strategy
379 for their location and use of dung resources. Furthermore, differences in dung sources for brood
380 ball formation and adult feeding have the potential to reduce competition between feeding adults
381 and those forming brood balls, thus effectively increasing resource availability.

382 There is limited information to test which of these strategies is employed by feeding adult *C.*
383 *bacchus*. Ruminant and rodent feces are characterized by smaller particle size than that of hindgut
384 fermenters (Owen-Smith 1992), but the filtering out of coarse material during ingestion by adults
385 (Holter 2016) makes this less important. Limited data suggest better C:N ratios in ruminant than
386 megaherbivore (hindgut fermenters) feces (Holter and Scholtz 2007), although such information is
387 not available for the smaller non-ruminants and rodents. Available preference data is limited to the
388 larger herbivore species, and does indicate a preference by *C. bacchus* for feeding on ruminant
389 feces. This provides partial support for the quality maximization strategy. Relevant data for the
390 time/effort minimization strategy as an explanation for dung source use by adult *C. bacchus* is
391 currently not available and would require quantified field observations of foraging effort.

392 An additional aspect of dung preferences displayed by *C. bacchus* relates to the
393 preference displayed for dung of herbivores that typically use densely vegetated habitat (Fig. 2).
394 This may reflect habitat-specific competitive abilities of this dung beetle. The fact that *C. bacchus* is
395 flightless, means that it is ectothermic and limited to walking when foraging, and is at a competitive
396 disadvantage with heterothermic, flying dung beetles for both the location and use of dung deposits
397 (Chown et al. 1995). The flightlessness of *C. bacchus* has been interpreted as an adaptation to
398 densely wooded habitats (Chown et al. 1995), and such dense vegetation hinders access for the
399 flying dung beetles. Based on these hypotheses, it may be predicted that *C. bacchus* has a
400 competitive advantage when foraging in dense vegetation, and this should be reflected in its
401 feeding preferences for dung from herbivores characteristic of such dense, closed vegetation. This
402 is supported by diet source and preference data provided here.

403 The important role of dung beetles in breaking down nutrients otherwise trapped in fecal
404 deposits is widely recognised, especially for large vertebrate dung (Nichols et al. 2009). The
405 energy equivalence rule (Damuth 1981) suggests that large and small mammal dung deposition
406 may be of similar orders of magnitude. Thus, mechanisms for the breakdown of rodent dung
407 should also be important in ecosystem functioning. Based on this, and observations of the
408 extensive use of rodent dung by adult *C. bacchus*, we suggest that *C. bacchus* adults and larvae
409 may collectively be serving an important role in the breakdown of mega-, meso- and
410 microherbivore dung. Following Nichols *et al.* (2009), *C. bacchus* may be important for maintaining
411 ecological processes and possible ecological cascades, a hitherto unrecognized functional role
412 with regard to rodent feces.

413 The findings presented here not only expand our understanding of the functional role of *C.*
414 *bacchus*, but also highlight the value of DNA-metabarcoding in exploring such patterns. In addition,
415 the hypothesis of ontogenetic shifts leading to ecological shifts is supported. Considering that the
416 larval/adult differences in feeding abilities are plesiomorphic, we expect such ecological shifts
417 across the diversity of scarabaeid dung beetles. Clearly, the differences in feeding abilities of the
418 larval and adult life history stages has profound consequences for their resource use and foraging

419 strategies, and hence the ecological role of dung beetles. This principle and its ecological
420 consequences needs to be explored in other scarabaeid dung beetle species.

421

422

423 **Competing interests.** We declare that we have no competing interests

424

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429

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433

434 **Data accessibility.** Data are available from the Dryad Digital Repository: [http://dx.doi.org/](http://dx.doi.org/§§§§§§§§§§§§§§)
435 [§§§§§§§§§§§§§§](http://dx.doi.org/§§§§§§§§§§§§§§) (Kerley et al. [§§§§](http://dx.doi.org/§§§§)).

436

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535

536 **Figure headings**

537 **Fig. 1** Frequency of occurrence of diet sources recorded in flightless dung beetle *Circellium*
538 *bacchus* fecal samples in the Main Camp and Colchester Sections of the Addo Elephant National
539 Park.

540

541 **Fig. 2** Relative preferences for dung of each of the censused large herbivores (in the Main Camp
542 and Colchester Sections of the Addo Elephant National Park) by flightless dung beetles *Circellium*
543 *bacchus*, estimated using Jacob's index, where $D > 0$ indicates preference and $D < 0$ avoidance.
544 See Online Resource 2 for species details.

545

546 **Supplementary material**

547 **Online Resource 1** Mean accumulation curve (50 random resamplings) of diet sources (putative
548 species) recorded per dung beetle fecal sample.

549

550 **Online Resource 2** Broad guild status (feeding, digestive morphology and habitat use guilds) for
551 the herbivorous mammal species recorded in the 2013 SANParks census of the Main Camp and
552 Colchester Sections of the Addo Elephant National Park, and potentially providing dung sources
553 for the flightless dung beetle *Circellium bacchus* (see Fig. 2). The blue duiker, although not
554 included in the census, did serve as a dung source (see Fig. 1) and is included here. Feeding guild
555 comprises browser, grazer or mixed feeder (Online Resource 5a), the digestive morphology guild
556 comprises ruminant or hind gut fermenter (Online Resource 5a), and habitat use comprises open
557 habitat use, closed habitat use or mixed open and closed habitat use (Online Resource 5a-c).

558

559 **Online Resource 3** Mammalian sequences (fragment of mitochondrial 16S gene) obtained from
560 dung beetle *Circellium bacchus* fecal samples collected in Addo Elephant National Park. The
561 scientific name attributed to each sequence is deduced from the best identity found in the release
562 126 of the EMBL database, and taking into account occurrence of the mammalian species
563 occurring in the study area. For *Otomys* sp. 3 and *Rhabdomys pumilio*, we had four and three
564 different sequence variants, respectively. We assumed that these closely related variants belong to
565 the same species, but in the absence of extra data we cannot exclude the presence of additional
566 species within *Otomys* or *Rhabdomys*.

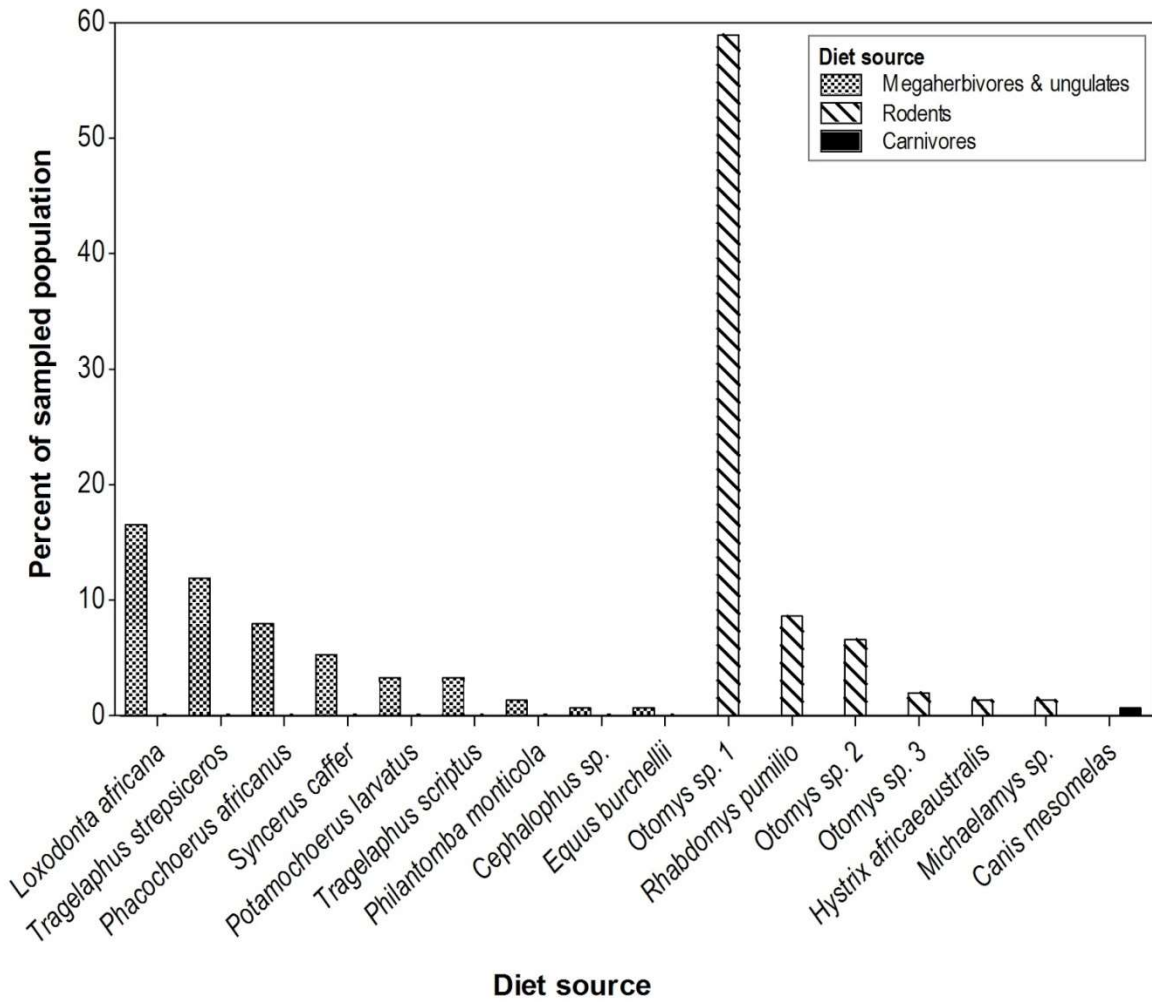
567

568 **Online Resource 4** Estimated relative dung production by large herbivores (elephants and
569 ungulates) in the Main Camp and Colchester Sections of the Addo Elephant National Park, based
570 on census data (SANParks Unpublished data), $\frac{3}{4}$ female body mass and digestive efficiency.

571

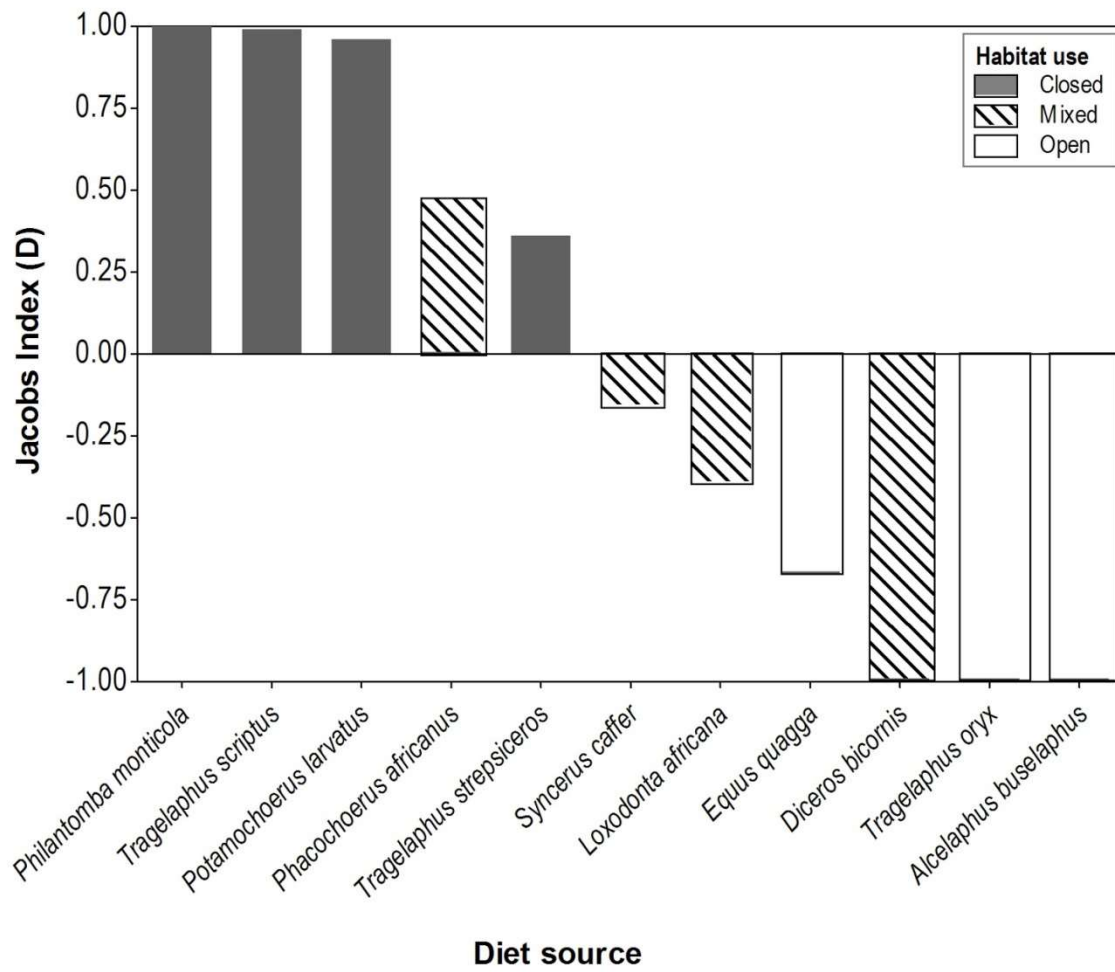
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575

576 **Figure 1**



577

578 **Figure 2**