



Viromes As Genetic Reservoir for the Microbial Communities in Aquatic Environments: A Focus on Antimicrobial-Resistance Genes

Stefano Colombo, Stefania Arioli, Eros Neri, Giulia Della Scala, Giorgio Gargari and Diego Mora*

Department of Food, Environmental and Nutritional Sciences, University of Milan, Milan, Italy

OPEN ACCESS

Edited by:

Télesphore Sime-Ngando,
Centre National de la Recherche
Scientifique (CNRS), France

Reviewed by:

Stefano Amalfitano,
Istituto di Ricerca sulle Acque (CNR),
Italy

Wei Xie,
Tongji University, China

*Correspondence:

Diego Mora
diego.mora@unimi.it

Specialty section:

This article was submitted to
Aquatic Microbiology,
a section of the journal
Frontiers in Microbiology

Received: 01 March 2017

Accepted: 30 May 2017

Published: 15 June 2017

Citation:

Colombo S, Arioli S, Neri E,
Della Scala G, Gargari G and
Mora D (2017) Viromes As Genetic
Reservoir for the Microbial
Communities in Aquatic
Environments: A Focus on
Antimicrobial-Resistance Genes.
Front. Microbiol. 8:1095.
doi: 10.3389/fmicb.2017.01095

Despite studies of viromes isolated from aquatic environments are becoming increasingly frequent, most of them are limited to the characterization of viral taxonomy. Bacterial reads in viromes are abundant but the extent to which this genetic material is playing a role in the ecology of aquatic microbiology remains unclear. To this aim, we developed of a useful approach for the characterization of viral and microbial communities of aquatic environments with a particular focus on the identification of microbial genes harbored in the viromes. Virus-like particles were isolated from water samples collected across the Lambro River, from the spring to the high urbanized Milan area. The derived viromes were analyzed by shotgun metagenomic sequencing looking for the presence, relative abundance of bacterial genes with particular focus on those genes involved in antimicrobial resistance mechanisms. Antibiotic and heavy metal resistance genes have been identified in all virome samples together with a high abundance of reads assigned to cellular processes and signaling. Virome data compared to those identified in the microbiome isolated from the same sample revealed differences in terms of functional categories and their relative abundance. To verify the role of aquatic viral population in bacterial gene transfer, water-based mesocosms were perturbed or not perturbed with a low dose of tetracycline. The results obtained by qPCR assays revealed variation in abundance of *tet* genes in the virome and microbiome highlighting a relevant role of viral populations in microbial gene mobilization.

Keywords: virome, microbiome, antimicrobial resistance genes, mesocosms, Lambro River

INTRODUCTION

Studies of the environments through metagenomics analysis are becoming increasingly frequent and the development of instruments, techniques and databases makes the study and the characterization of viral communities with a shotgun metagenomics approach more and more informative. Water has been described as the major environment for bacteria on earth (Taylor et al., 2011), and bacteriophages outnumber bacteria by a factor ranging from 1 to 10 (Muniesa et al., 2013a). Water is an ideal medium for bacterial life, characterized by accessible dissolved nutrients, as well as protection from desiccation and UV light. In this context, the high frequency of encounters between bacteria and bacteriophages strongly favors, through transduction, the

mobilization of genetic material. Most studies focused on environmental viromes are limited to the characterization of viral taxonomy (Roux et al., 2012; Weynberg et al., 2014; Danovaro et al., 2016). However, viromes harbor huge amounts of bacterial reads which account for up to 90% of the microbial reads (Fancello et al., 2013; Modi et al., 2013; Zablocki et al., 2014; Abeles et al., 2015). Few studies reported that phage metagenomes were enriched with microbial functional genes (Fancello et al., 2013; Modi et al., 2013; Colombo et al., 2016), which clearly reported that phage metagenomes were enriched with microbial functions. The extent to which these virome-associated microbial genes are accessible to members of the environmental microbiota remains unclear. In this context, those factors triggering the mobilization and spreading of microbial genes by the viral communities in water environments are still processes not completely understood. To date the attention was mainly focused on the mobilization of antibiotic-resistance genes (ARGs) (Allen et al., 2010; Muniesa et al., 2013a,b). A part from the water environments, the role of viruses in mobilization of ARGs in the human and mice microbiota after exposure to antibiotics has been recently experimentally documented, showing that the treatment with antibiotics leads to an increase in the ARGs abundance in the virome (Fancello et al., 2013; Modi et al., 2013; Abeles et al., 2015).

Rivers can be polluted by a diverse mixture of antibiotics and other pollutants, metabolites and resistant bacteria through treated and untreated sewage, hospital waste, aquaculture discharges, and agricultural run-off. These aquatic compartments, such as water and sediment, may therefore have a significant role in driving ARGs transfer, ecology and evolution (Taylor et al., 2011; Marti et al., 2014). In this work, we developed a method for the comprehensive characterization of viral and microbial communities by shotgun metagenomics focusing the attention on the presence and relative abundance of microbial gene sequences in viromes isolated from the Lambro River flow, from the spring to the high urbanized/industrialized Milan area. The aim of this work was to describe the relative abundance and the classification in functional categories of the bacterial gene sequences associated with aquatic environment viromes. A special attention was paid to the genes associated with the antimicrobial-resistance mechanisms such as ARGs and heavy metal resistance genes. With the aim to prove the role of viruses in the mobilization of ARGs, tetracycline-resistance (*tet*) genes were monitored in Lambro River water-based mesocosms, perturbed and non-perturbed with a low dose of the antibiotic tetracycline.

MATERIALS AND METHODS

Sampling Procedures

Lambro River catchment is located in the north of Milan with a total drainage area of approximately 1,950 km². Lambro River's spring is situated in the Pre-Alps (1,450 m above sea level) in the Magreglio area, and it flows into a confluence with the Po River (50 m above sea level) with an estimated length of 130 km. The

River catchment average annual rainfall varies between 900 and 1,500 mm (Deksissa et al., 2004).

Three water samples (30 L) were collected in February 2015 along Lambro River. The first sample was collected at the river's spring in Magreglio area (45.935270, 9.264576). This sample due to the characteristics of the area can be considered as belonging to a no urban impact area (A). The second sampling point was near the town of Ponte Lambro (45.49349, 9.13274) approximately 15 km from the Lambro River origin (B). The third sampling point was in Milan in Lambro Park (45.495399, 9.247936) approximately 55 km from the origin (C). Samples were maintained at 6°C for 4 h before processing.

Virus-Like Particle Isolation

Water samples were filtered at 0.2 μm (Pall, Life Sciences, Milan, Italy) with a tangential flow filtration system (TFF) (Quattro systems, Parma, Italy), as previously described in Colombo et al. (2016). The permeate collected after the 0.2 μm TFF, containing all the virus-like particles (VLPs), was subsequently ultrafiltered at 100 kDa (Pall, Life Sciences, Milan, Italy) using the same TFF system; pressure was maintained under 62 kPa in order to avoid VLP damage (Thurber et al., 2009). The retained material containing the VLPs (400 mL) was then precipitated overnight at 4°C using PEG 8000 at a final concentration of 10% (wt/vol) and subsequently centrifuged at 13,000 × *g* for 30 min at 4°C. The pellet was suspended in TE buffer (pH 8.0) and prepared for CsCl gradient centrifugation. The solution was then deposited on top of a 2.5 mL step gradient composed of multiple 0.8 mL CsCl solutions with corresponding densities of 1.70, 1.50, and 1.35 g mL⁻¹. Samples were centrifuged for 2 h at 60,000 × *g* (4°C) in a SW41 swinging bucket rotor (Beckman Instruments Inc., Fullerton, CA, United States) (Thurber et al., 2009). According to the protocol proposed by Thurber et al. (2009), we recovered approximately 2 mL of the 1.5 g mL⁻¹ layer, because material in this density range should be enriched with VLPs.

Extraction and Amplification of VLP DNA for Metagenomic Analysis

Viral DNA was extracted according to a protocol previously described by Thurber et al. (2009). The 1.5 g mL⁻¹ layer collected from the step gradient was treated with DNase (37°C, 12 h), following the manufacturer's instructions (Sigma-Aldrich, Milan, Italy; final concentration, 2.5 U mL⁻¹), to remove residual host and bacterial DNA. To extract the virions, 0.1 volumes of 2 M Tris HCl/0.2 M EDTA, 1 volume of formamide and 100 μL of a 0.5 M EDTA solution was added to the 10 mL sample, and the resulting mixture was incubated at 25°C for 30 min. The sample was subsequently washed with 2 volumes of 70% ethanol and pelleted by centrifugation at 8,000 × *g* for 20 min at 4°C. The pellet was washed twice with 70% ethanol and resuspended in 567 μL of TE buffer, followed by the addition of 30 μL of 10% SDS and 3 μL of a 20 mg mL⁻¹ solution of proteinase K (Fisher Scientific, Waltham, MA, United States). The mixture was incubated for 1 h at 55°C and supplemented with 100 L of 5 M NaCl and 80 μL of a solution of 10% cetyltrimethylammonium bromide/0.7 M NaCl. After a 10 min incubation at 65°C, an equal volume of chloroform

was added, and the mixture was centrifuged (5 min at $8,000 \times g$ at 25°C). The resulting supernatant was transferred to a new tube, and an equal volume of phenol/chloroform/isoamyl alcohol (25:24:1) was added. The mixture was centrifuged (5 min at $8,000 \times g$ at 25°C). The supernatant was recovered, and an equal volume of chloroform was introduced. Following centrifugation, the supernatant was collected, and 0.7 volumes of isopropanol were used to precipitate the DNA. After centrifugation (15 min at $13,000 \times g$ and 4°C), the DNA pellet was washed with $500 \mu\text{L}$ of ice-cold 70% ethanol, air-dried and resuspended in $50 \mu\text{L}$ TE buffer (Thurber et al., 2009).

In order to increase the amount of DNA available, whole-genome amplification was performed using reagents and protocols of the Illustra GenomiPhi V2 kit (GE Healthcare, Milan, Italy) to generate sufficient material for metagenomic analysis. Purified VLP DNA (5–15 ng) was mixed with $9 \mu\text{L}$ of sample buffer provided by Illustra GenomiPhi V2 kit and heat-denatured at 95°C for 3 min. Subsequently, $9 \mu\text{L}$ of kit reaction buffer and $1 \mu\text{L}$ of kit enzyme mix were added, and the solution was incubated for 90 min at 30°C . The amplified products were subsequently pooled and purified (UltraClean PCR Clean-UP Kit, MoBio Laboratories, Solana Beach, CA, United States) (Thurber et al., 2009). Finally, $2 \mu\text{L}$ of each DNA solution was quantified using the PowerWave XS Microplate Spectrophotometer at 260 nm (BioTek Instruments, Inc., Winooski, VT, United States) and the Take3 Multi-Volume Plate (BioTek Instruments, Inc., Winooski, VT, United States).

Calculation of Bacterial Contamination Using qPCR

An aliquot of the purified DNA was used as a template in qPCR assays to quantify the amount of contaminating, non-viral DNA. A CFX96 thermocycler (Bio-Rad Laboratories S.r.l., Milan, Italy) was used to quantify the bacterial DNA. The analysis was performed using a Fast Eva Green Supermix SYBR Green PCR Master Mix (Bio-Rad Laboratories) in a reaction volume of $15 \mu\text{L}$ per well. qPCR amplification was carried out with an initial denaturation at 95°C for 5 min, followed by 40 cycles of denaturation at 95°C for 30 s, annealing at 58°C for 30 s and extension at 72°C for 30 s. The primers EubF1 (5'-GTGSTGCAYGGYGTGTCGTCA-3') and EubR1 (5'-GAGGAAGGTGKGGAYGACGT-3') were used to target 16S rRNA to detect any eubacteria. Data were recorded as threshold cycles (C_{tT}) and expressed as mean values; standard deviations were computed using the Bio-Rad CFX Manager. DNA from pure cultures of *Lactococcus garvieae* strain TB25 was subjected to six 10-fold serial dilutions and used as templates for qPCR. Standard curves were generated by plotting the \log_{10} of the bacterial cell numbers against the corresponding C_{tT} values obtained from the amplification of diluted DNA. Calibration curves showed good correlation between C_{tT} values and the number of cells over the considered range (r^2 regression coefficients were between 0.964 and 0.999). The resulting detection limits were between 1.1 and 1.9 \log_{10} bacterial cells per reaction mix (Guglielmetti et al., 2013). On the basis of the

C_{tT} values obtained using viral DNA, the actual concentration of bacterial DNA was determined and expressed as a percentage of the total DNA previously extracted. Bacterial contaminating DNA accounted for less than 0.2% of the total viral DNA extracted.

Extraction of Microbial DNA for Metagenomic Analysis

Water samples were filtered at $0.2 \mu\text{m}$ by TFF, and the obtained permeate was centrifuged for 15 min at $9000 \times g$ and 4°C . The biomass obtained was then suspended in $400 \mu\text{L}$ of TE buffer (pH 8). To this purpose, $15 \mu\text{L}$ of lysozyme (20 mg mL^{-1}) was added before a 60 min incubation at 37°C . Subsequently, $15 \mu\text{L}$ of SDS (20%) and $15 \mu\text{L}$ of proteinase K (20 mg mL^{-1}) (Fisher Scientific) were added, and the mixture was incubated at 55°C for 15 min. The mixture was supplemented with $250 \mu\text{L}$ of phenol and centrifuged at $22,000 \times g$ for 5 min. The phenol phase was removed, and $400 \mu\text{L}$ of chloroform was added to the aqueous supernatant. The centrifugation step was repeated as described above. The supernatant was transferred to a new tube with $40 \mu\text{L}$ of sodium acetate (3 M, pH 5.2) and $800 \mu\text{L}$ of 96% ethanol (-20°C). The mixture was centrifuged for 30 min at $22,000 \times g$ and 4°C . The liquid was discarded, and the pellet was washed with $300 \mu\text{L}$ of 70% ethanol (-20°C). The centrifugation step was repeated as described above, and the liquid was discarded. The DNA was dried and suspended in $40 \mu\text{L}$ of sterile distilled water. The PowerWave XS Microplate Spectrophotometer at 260 nm (BioTek) and the Take3 Multi-Volume Plate (BioTek) were used to quantify $2 \mu\text{L}$ of the DNA solution.

Metagenomic Shotgun Sequencing of Microbial and Viral DNA

A genomic library was generated from $3 \mu\text{g}$ of genomic DNA using the TruSeq DNA PCR-Free Sample Preparation Kit (Illumina) and the MiSeq Reagent, according to the user's guide (Illumina). The quality of the library was estimated prior to loading onto a flow cell, and the library sample was subsequently sequenced using 500 sequencing cycles, according to the MiSeq (Illumina) instructions. The 500 sequencing cycles resulted in an average read length of approximately 250 nucleotides for both paired-end sequences. The MIRA program version 4.0.2 (Chevreux et al., 1999) was used for *de novo* assembly of contigs. Mira options were set as follows: "job = genome, *de novo*, accurate parameters = -nw:cac = no -ge:not = 1 ionor_settings -as:mrpc = 100" ([minimum_reads_per_contig (mrpc) = 100]). Assembled reads were searched for ORFs predicted with PRODIGAL v2_60 linux¹ (Hyatt et al., 2010).

Quality Control of Virome Reads

Reads from metagenomic shotgun sequencing were initially checked for quality using the FASTQC program². Fastq files were trimmed and filtered with FASTX-Toolkit³ and homerTool⁴.

¹<http://prodigal.ornl.gov/>

²<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>

³http://hannonlab.cshl.edu/fastx_toolkit/

⁴<http://homer.salk.edu/homer/ngs/homerTools.html>

The fastq output was filtered for reads with a quality of < 25 , as well as reads of 80 bp. Bases were removed from the ends of the reads until the average quality in a window of 5 bp was > 25 . Contaminant reads (i.e., human reads) were removed with CS-SCORE (Haque et al., 2015). Sequencing metadata, assembly metrics, and annotation statistics are summarized in Supplementary Table S1.

Identification of contaminant reads in viral metagenomes was performed with a BLASTn analysis on the “16SMicrobial” database provided by NCBI⁵. BLASTn was performed with a threshold of 10^{-5} on the *E*-value, and only results with the identity over 90% were kept. Levels of contamination were then expressed in percentage (%) as a number of reads identified with the “16SMicrobial” database over the total number of reads.

Bioinformatics

A BLAST analysis was performed against non-redundant viral database using BLAST+ 2.2.29 <ftp://ftp.ncbi.nlm.nih.gov/blast/executables/blast+/LATEST/>. A database was created using all entries for the taxonomy ID 10239 (Viruses) on NCBI.

Both viral and microbial data underwent a BLAST analysis against both the non-redundant nucleotide and the non-redundant protein database of NCBI; BLASTn, BLASTp, and BLASTx were performed with a threshold of 10^{-5} on the *E*-value. In order to identify reads associated to ARGs, the data were compared with the CARD (Comprehensive Antibiotic Resistance Database); the database is composed of 3228 genes specifically tagged for antibiotic resistance (McArthur et al., 2013). BLASTp and BLASTn were performed in order to identify the antibiotic resistance-encoding reads with a threshold of 10^{-5} on the *E*-value. Only sequences with an amino acid identity $> 30\%$ and a nucleotide identity $> 70\%$ were considered.

Identification of metal resistance reads was performed with BLASTp and BLASTn against BacMet database (Pal et al., 2014). All bioinformatics analyses were performed using the CINECA SCAI (SuperComputing Applications and Innovation) (CINECA, Bologna, Italy), part of the ISCRA VArFaWtr project.

Accession Numbers

The sequence data were submitted to the EMBL-EBI database under the accession number PRJEB14951.

Evaluation of Antibiotic Resistant Colony Frequencies

For the evaluation of antibiotic resistant colony frequencies, samples from the 0.2 μm TFF system retentate (see next paragraph) were used for dilution and plating on two selective media: plate count agar (PCA) and MacConkey (MC) agar (Sigma-Aldrich). Both media were also tested with the addition of antibiotics such as ampicillin, tetracycline, chloramphenicol, and meropenem. The antibiotics were present at a final concentration of 20 $\mu\text{g mL}^{-1}$. Samples were incubated at 20°C for 48 h. The microbial count was carried out in triplicate.

⁵<ftp://ftp.ncbi.nlm.nih.gov/blast/db/>

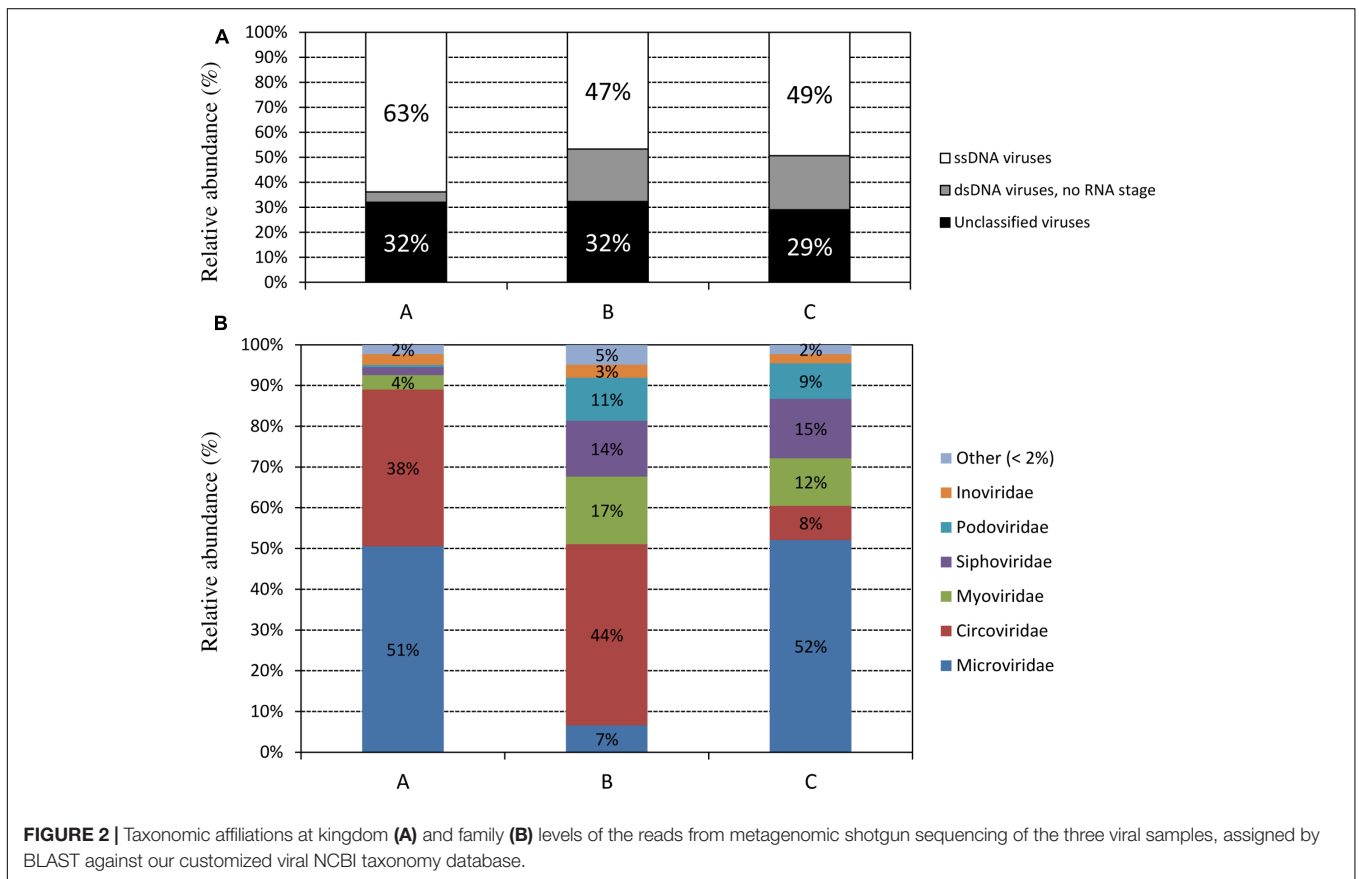
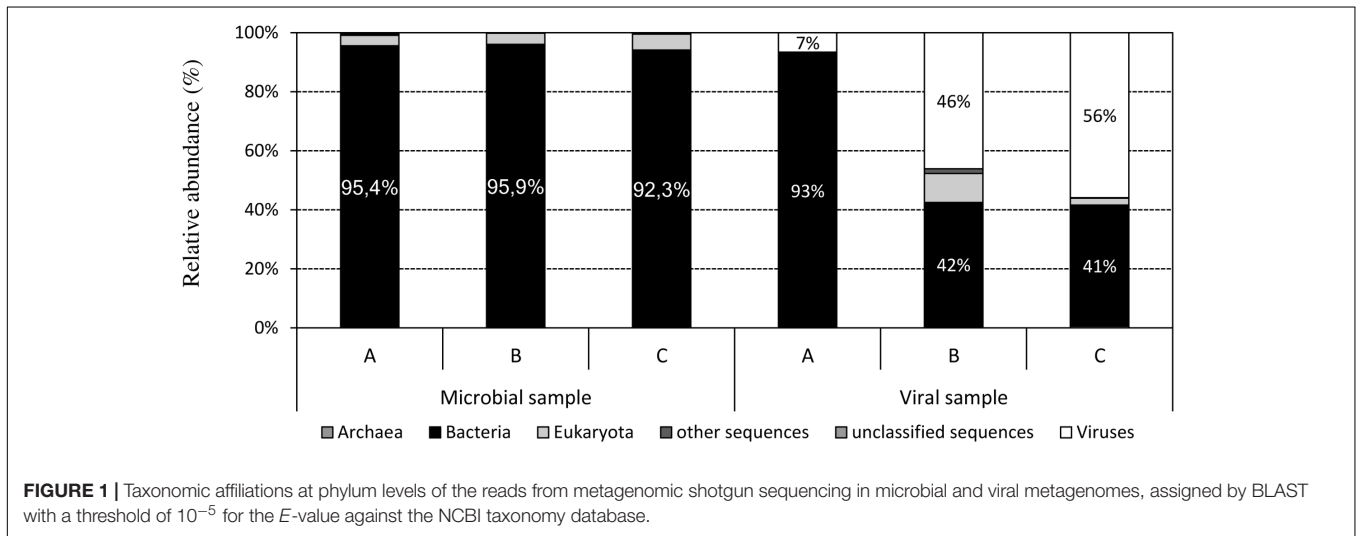
Monitoring of *tet* Genes in Perturbed and Non-perturbed Mesocosms

To understand the role of viral population in the mobilization of *tet*, we prepared mesocosms using water collected from the Lambro River at the Milan sampling site. In detail, the Lambro River water-based mesocosms were prepared with and without the addition of tetracycline used as a selective pressure agent. Because tetracycline resistance-related reads were identified in all three viromes analyzed, and due to the availability of qPCR protocols for detection and quantification of *tet* genes, we choose this antibiotic as a bacterial perturbation agent in the mesocosms. All mesocosms were prepared in 20 L sterilized tanks maintained under static conditions, at environmental temperature (6–10°C), and not exposed directly to the sun light. After sampling the water from the river, eight mesocosms were maintained in the above-described condition for 48 h. After that period, two mesocosms were supplemented with tetracycline (Sigma-Aldrich) at a final concentration of 100 ng L^{-1} and other two mesocosms were used as controls. After 24 h all four mesocosms were treated to collect the viral populations by TFF as previously described. The viral populations collected, were subjected to further purification of VLP and DNA purification as described above. The 0.2 μm TFF retentate, containing the microbiota, was used for the extraction of total DNA as described above. Viral and microbial DNA were used as templates for the qPCR assay to quantify genes associated with tetracycline resistance (*tet* genes) according to the protocols described by Yu et al. (2005) and Fan et al. (2007). After perturbation with 100 ng L^{-1} of tetracycline, the virome and microbiome were collected, and a set of *tet* genes coding for Gram-negative efflux pumps (*tetA-C*, *tetE*, *tetG*, and *tetH*) (Pao et al., 1998; Yu et al., 2005; Fan et al., 2007) were quantified by qPCR. qPCR assays were performed in a CFX96 thermocycler (Bio-Rad Laboratories). The reaction mixture was prepared using a Fast Eva Green Supermix SYBR Green PCR Master Mix (Bio-Rad Laboratories) in a reaction volume of 15 μL per well, using 5 ng of viral or microbial DNA as the template. All qPCR assays were performed in triplicate. The quantification of *tet* genes was shown as the average of the C_t values \pm the standard deviation. Statistically significant differences were determined by an unpaired Student's *t*-test ($*P < 0.05$).

RESULTS

Taxonomic Description of Viral Communities Using the Shotgun Metagenomic Approach

Identification of viral reads in the viromes of the three samples resulted in two different scenarios (Figure 1). The sample A was characterized by the largest abundance of bacterial reads (93%), and only 7% were identified as viral reads. On the contrary, the other two samples showed a lower amount of bacterial reads (41–42%) and a higher level of the viral reads (46–56%). In addition, we would like to underline that ribosomal sequences were detected with a frequency (no. of ribosomal reads on the total of reads in virome) largely below the 0.2% threshold in two



of the three viral metadata. Currently, when $> 0.2\%$ of reads match to 16S ribosomal DNA, a non-viral marker of bacterial DNA, the viromes are considered to have high bacterial DNA content (Roux et al., 2013). We obtained the level of 0.006 and 0.009% of reads match to 16S ribosomal DNA for samples B and C, and 0.35% for sample A. Moreover, in all three samples qPCR quality control assays revealed that the levels of bacterial contaminating DNA in virome samples accounted for less than

0.2% of the total viral DNA extracted, a threshold considered acceptable as suggested by Modi et al. (2013). We therefore exclude that this low level of bacterial DNA contamination in viromes could have affected the subsequent bioinformatic analysis of viromes.

In our samples taxonomy was first associated with the viral reads. Viral reads were mainly identified as ssDNA viruses and as dsDNA viruses (Figure 2A), as expected for bacteriophages.

Among them approximately 30% of the reads were classified only at the kingdom level. The remaining reads showed a taxonomy distribution as follows: *Microviridae* was the most abundant family identified both in samples B (51%) and C (52%) while only 7% in sample A. Similarly, *Circoviridae* were present in a large amount in samples A and B (38 and 44%, respectively), while only the 8% of the identified viral reads were recognized in sample C. To a lesser extent *Myoviridae* and *Siphoviridae* were present in a similar abundance in samples A and C (17 and 12%; 14 and 15%, respectively) while only at a low amount in the sample B (4 and 2%) (Figure 2B).

A separate taxonomic identification of the microbial reads in the three viromes has been performed (Figure 3). As observed by other authors (Rosario et al., 2009; López-Bueno et al., 2009; Roux et al., 2012; Fancello et al., 2013; Modi et al., 2013) bacterial reads in viromes are abundant and accounted for up to 90% of the microbial reads. We therefore focused the attention on their taxonomic identification. *Flavobacteriaceae* was the only family that stood out of the three samples (37% in the A sample in comparison to 6 and 12% in samples B and C, respectively). Other notable families identified were *Chitinophagaceae* (18% in sample B), *Neisseriaceae* (17% in sample A), and *Lactobacillaceae* (12% in sample A). Virome of sample C was the richest regarding bacteria with 294 different bacterial families represented, followed by viromes of samples A and B showing 290 and 236 bacterial families, respectively.

Functional Characterization of Virome Data

Metabolism-related reads in viromes showed a range of abundance between 15 and 37%, whereas information storage and processing-related reads ranged between 20 and 36% (Figure 4). The abundance of reads assigned to cellular processes and signaling was similar in all three samples (25–25–22%). Focusing the attention at the subcategories (Figure 4B), we observed a similar distribution of WHAT? in all samples for the most of them with the exception of nucleotide transport and metabolism-related reads, which accounted for up to 56% in sample C (11 and 14% in the other two samples).

Mobilome-associated reads showed an abundance ranged between 5 and 12%. This category, as defined by NCBI-COG, mainly groups reads that encode for phage-related proteins as terminase, capsid, tail and other as transposase and protein involved in initiation of plasmid replication. In detail, phage-related ORFs accounted for 20–59% of the mobilome, whereas transposase and protein involved in plasmid replication showed a relative abundance between 33 and 37% of the mobilome.

Functional Characterization of Microbiome Data

As for analysis of virome, functional classification of microbiome reads was performed using the COG database. No changes were observed for each functional category among the different samples (Figure 4A). Metabolism-related reads were the most represented (37–42%), reads associated with “cellular processes and signaling” were the second most observed (25–31%), and

reads related to “information storage and processes” were the third most observed (15–19%) (Figure 4A). Reads included in the “mobilome” category accounted for 2–3% of the total: among these, up to 65% were identified as transposase-related reads, while from 12 to 25% were classified as phage-related reads. Among the metabolism-related category, none was predominant with respect to the others: amino acid transport and metabolism (19–21%), carbohydrate transport and metabolism (14–18%), inorganic ion transport and metabolism (14–18%), lipid transport and metabolism (12–13%), and energy production (12–17%) (Figure 4B).

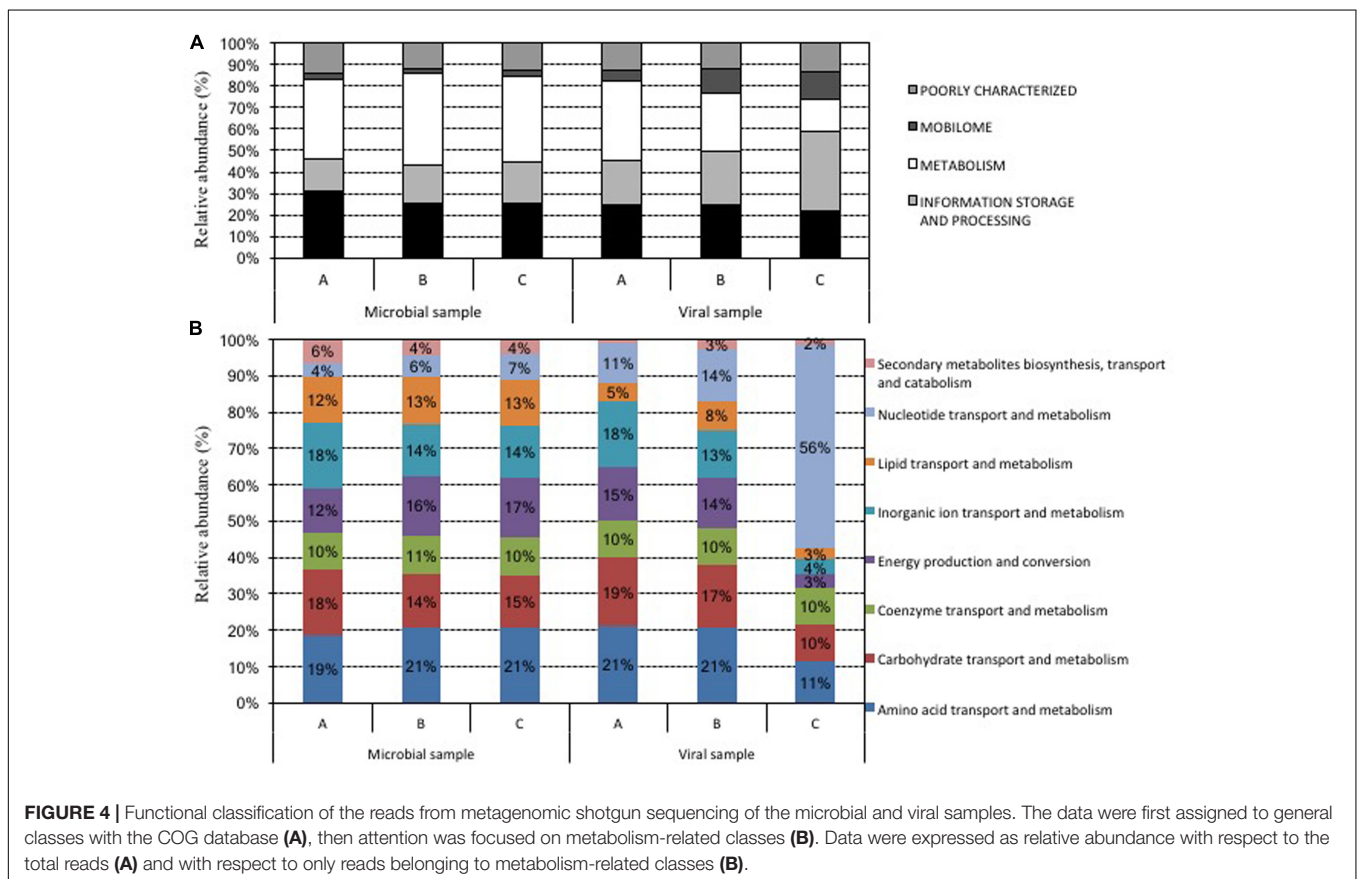
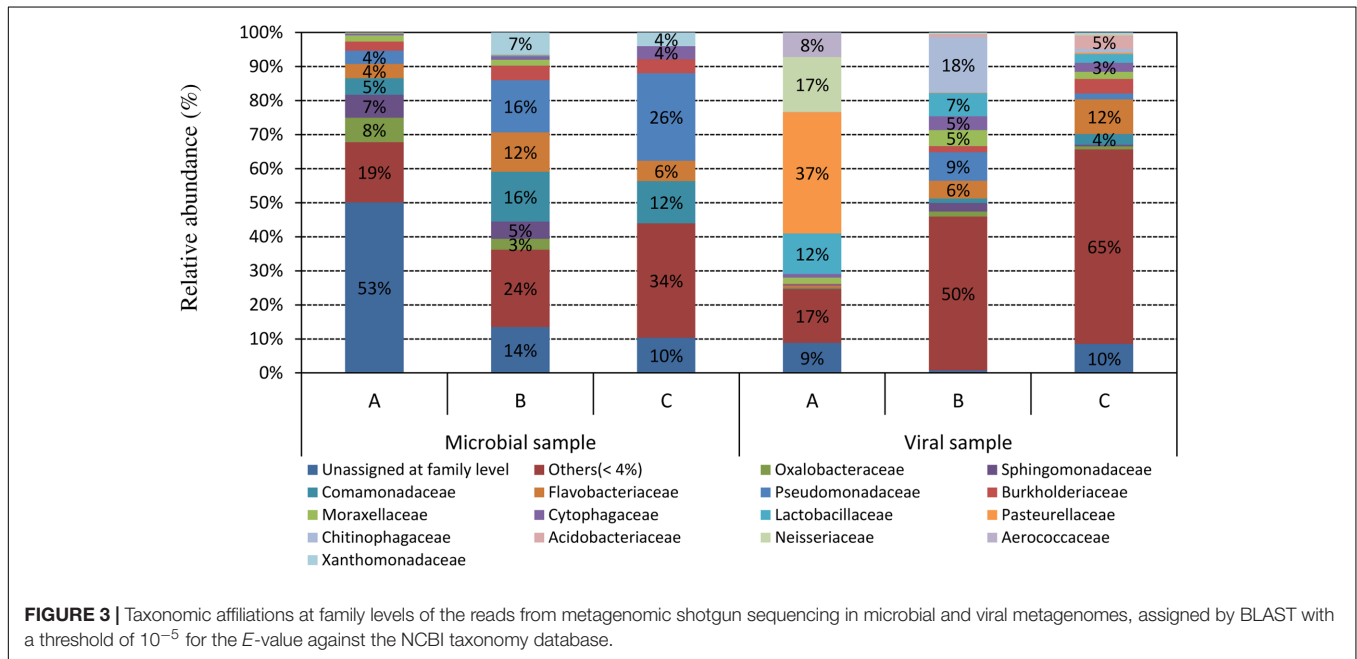
Identification of Antibiotic Resistance Genes in the Microbiome and Virome

In viromes we observed a relative abundance of reads associated with ARGs ranging from 0.48% (sample C) to 1.92% (sample A) (calculated as the relative abundance of reads associated with ARGs over the total reads identified within NCBI) (Figure 5A). The genes coding for DNA synthesis inhibitors and protein synthesis inhibitor classes showed an abundance ranging from 15 to 38%, whereas ARGs belonging to efflux pumps and genes modulating antibiotic efflux ranged between 24 and 57%. Aminocoumarin and glycopeptide resistance genes were two of the most represented in the three samples. Tetracycline-resistance genes ranged between 2 and 12% (Figure 5B).

In microbiome, the relative abundance of reads associated with ARGs ranged between 8 and 12% (calculated as the relative abundance of reads associated with ARGs over the total reads identified within NCBI). Nevertheless, the three microbiomes showed a similar distribution of the five drug classes as shown in Figure 5A (here the relative abundance is calculated for the total amount of reads associated with ARGs): ARGs that encode efflux pumps were the most abundant (~47%), followed by cell wall synthesis and disruption ARGs (23%), and finally DNA synthesis inhibitors (17%). Looking specifically at the antibiotic resistance genes identified from the microbial metagenome, an identical situation was observed: up to 45% of the ARGs identified were genes that encode for efflux pumps or genes modulating the efflux systems (Figure 5A). As previously described for viromes, aminocoumarin, and glycopeptide resistance genes were the most represented in all three microbiome samples with amounts ranging from 12 to 16%. Sequences belonged to beta lactam coding genes, and chloramphenicol and tetracycline-resistance genes were also found (Figure 5B).

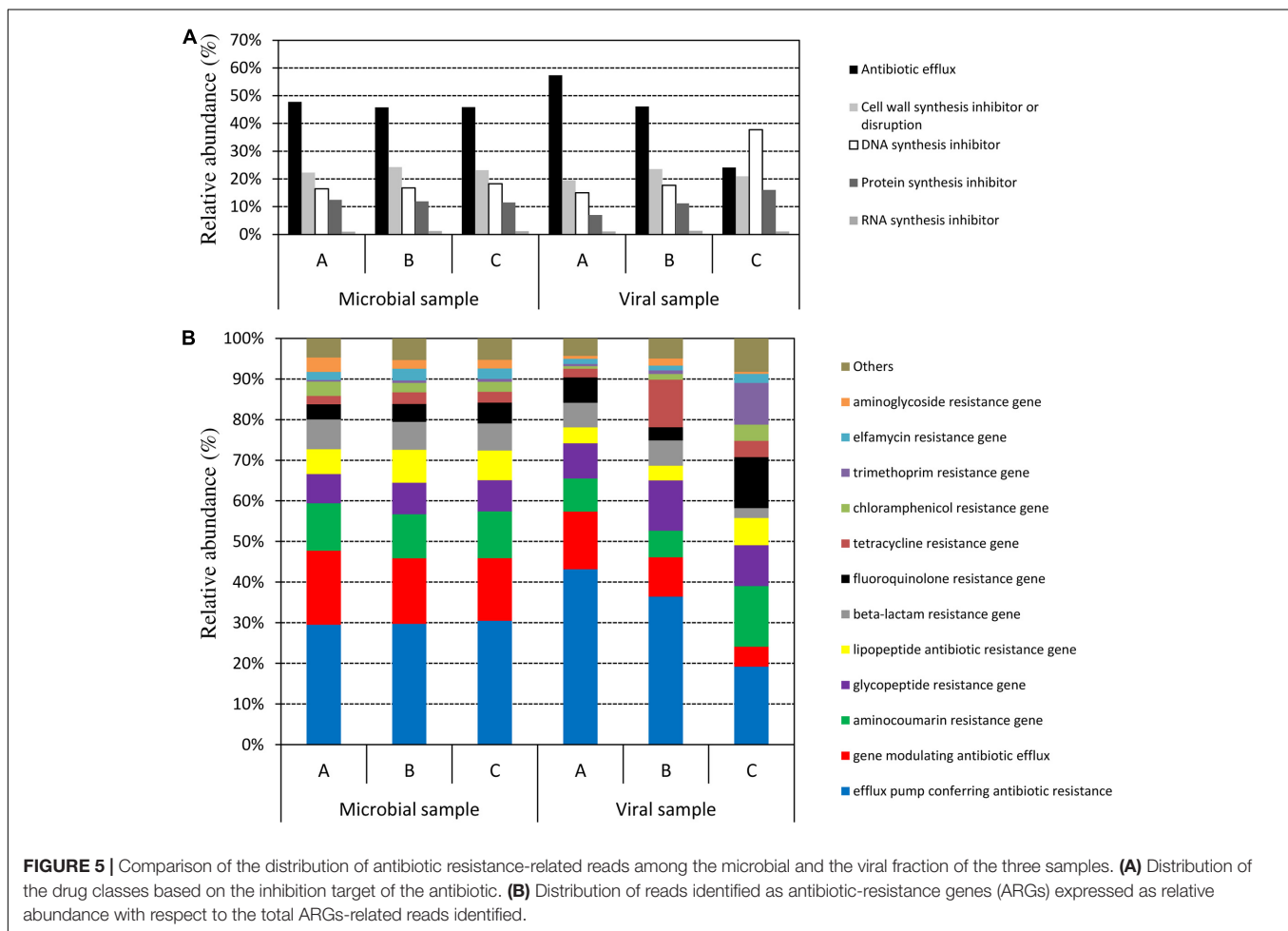
Identification of Heavy Metal and Biocide Resistance Genes

From the spring to Milan area, the Lambro River crosses increasingly industrialized areas that negatively affect the point-source pollution levels in its waters, specifically pesticides, metals, organochlorines, and organic compounds. For these reasons, microbiomes and viromes were analyzed for the identification of genes related to resistance mechanisms against heavy metals (MRGs). In the microbiomes, the relative abundance of MRGs ranged from 9% in sample A to 14% in sample C. In viromes, MRGs ranged between 3% in sample A to 1% in



sample C. Fifty-six different resistance entries were identified with a similar pattern of relative abundance among the six different metagenomes (Supplementary Table S2). Among the genes associated with metal resistance the highest abundances

identified were against copper (~13% in the three microbiomes while up to 20% in virome sample B), zinc (an average of 13% in the microbiomes and from 8 to 14% in the viromes), iron (8% in the microbiomes; up to 16% in virome sample A), and



nickel (7% in the microbiomes; from 5 to 13% in the viromes). Biocide resistance genes were identified in a much lower amount with respect to the MRGs: among them the most abundant identified were resistance genes against hydrogen peroxide (2% in the microbiomes and up to 4% in the viromes).

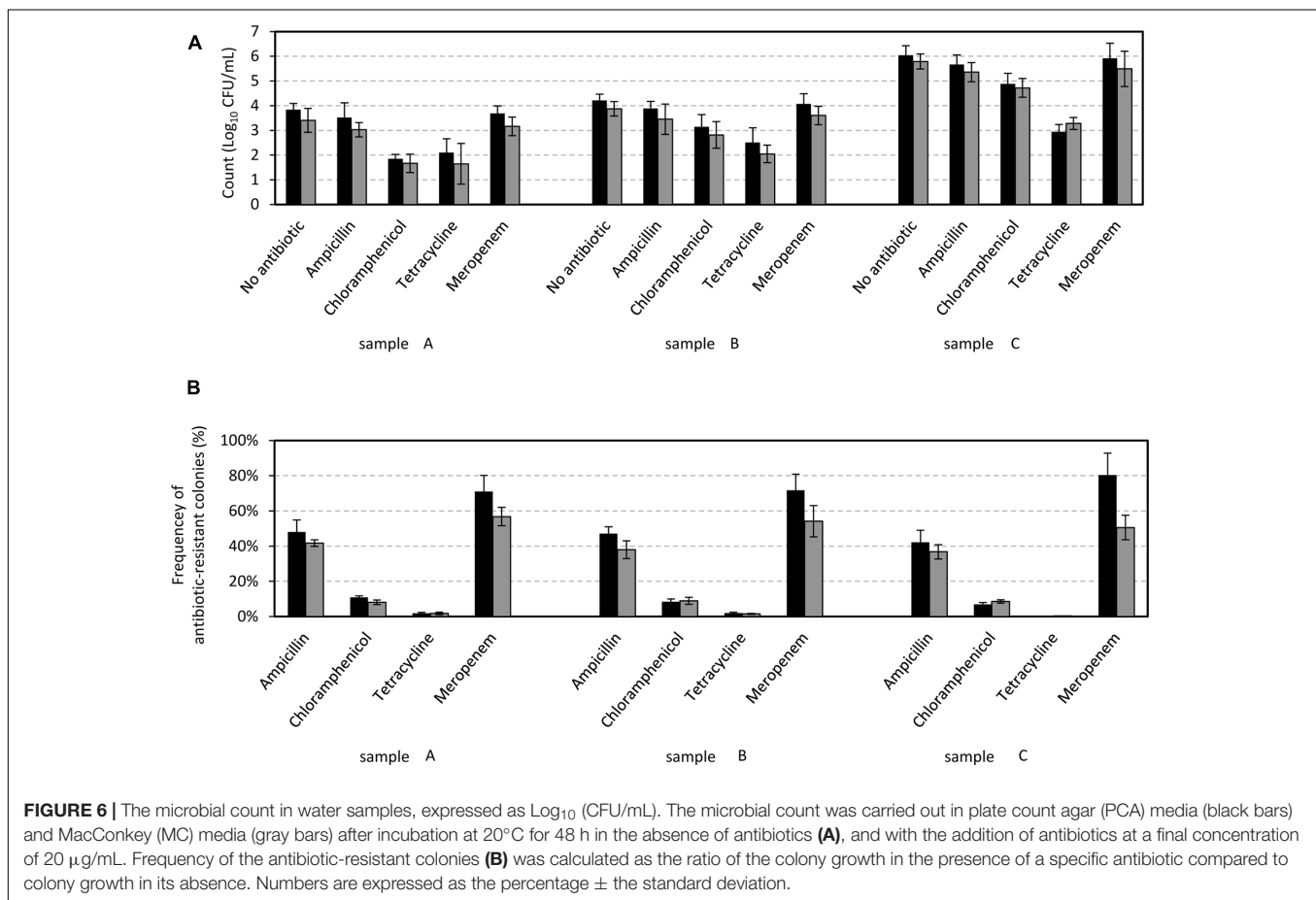
Frequency of Antibiotic Resistant Colonies in Water Samples

With the aim to evaluate the frequency of antibiotic resistant colonies of the aquatic microbial communities, we first quantified the total cultivable microbial abundance by plating on two different media: PCA and MC. The total bacterial count obtained revealed an increase of cultivable bacteria from the spring to the water samples collected close to Milan city. On PCA, the bacterial count ranged between 7×10^3 CFU L⁻¹ (3.8 log₁₀) $\pm 4 \times 10^2$ in sample A and 1×10^6 Log₁₀CFU L⁻¹ (6.1 log₁₀) $\pm 7 \times 10^4$ in sample C (Figure 6A). Similarly, on MC agar the Gram-negative bacterial count ranged between 2×10^3 CFU L⁻¹ (3.4 log₁₀) $\pm 4 \times 10^2$ in sample A and 6×10^5 CFU L⁻¹ (5.8 log₁₀) $\pm 3 \times 10^4$ in sample C. Focusing on the frequencies of the antibiotic resistant colonies, surprisingly no significant differences were observed among the different samples. In PCA media the highest frequencies were observed

in presence of beta lactamic antibiotics: meropenem-resistant colonies reached up to 70% of the total, and ampicillin-resistant colonies were approximately 47%. However, chloramphenicol-resistant colonies accounted for 7–11%, and tetracycline-resistant colonies were under 2% in all three samples (Figure 6B). We therefore conclude that the frequency of antibiotic resistant colonies was not affected by the increasingly industrialized and urbanized areas crossed by the Lambro River from samples A to C. However, the absolute number of antibiotic-resistant colonies increased significantly from samples A to C area as a consequence of the increase in the total bacterial count in the water samples.

Changes in *tet* Genes in Virome- and Microbiome-Exposed and Non-exposed to Low Dose of Tetracycline in Lambro River Water-Based Mesocosms

With the aim of following changes in tetracycline-resistant gene (*tet*) in the viromes and microbiomes, we prepared mesocosms using water collected from the Lambro River in the Milan sampling site. The results obtained revealed a significant reduction of *tetA-C*, *tetG*, and *tetH* genes (higher C_t values) in viromes collected from mesocosms that were exposed

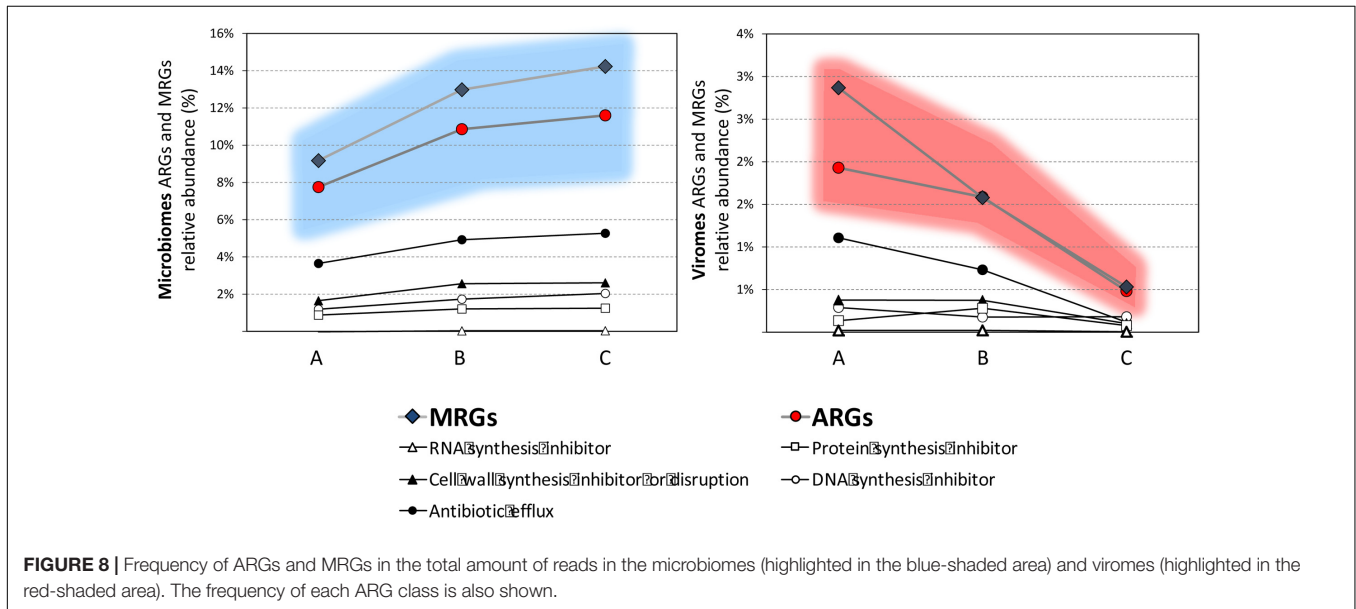
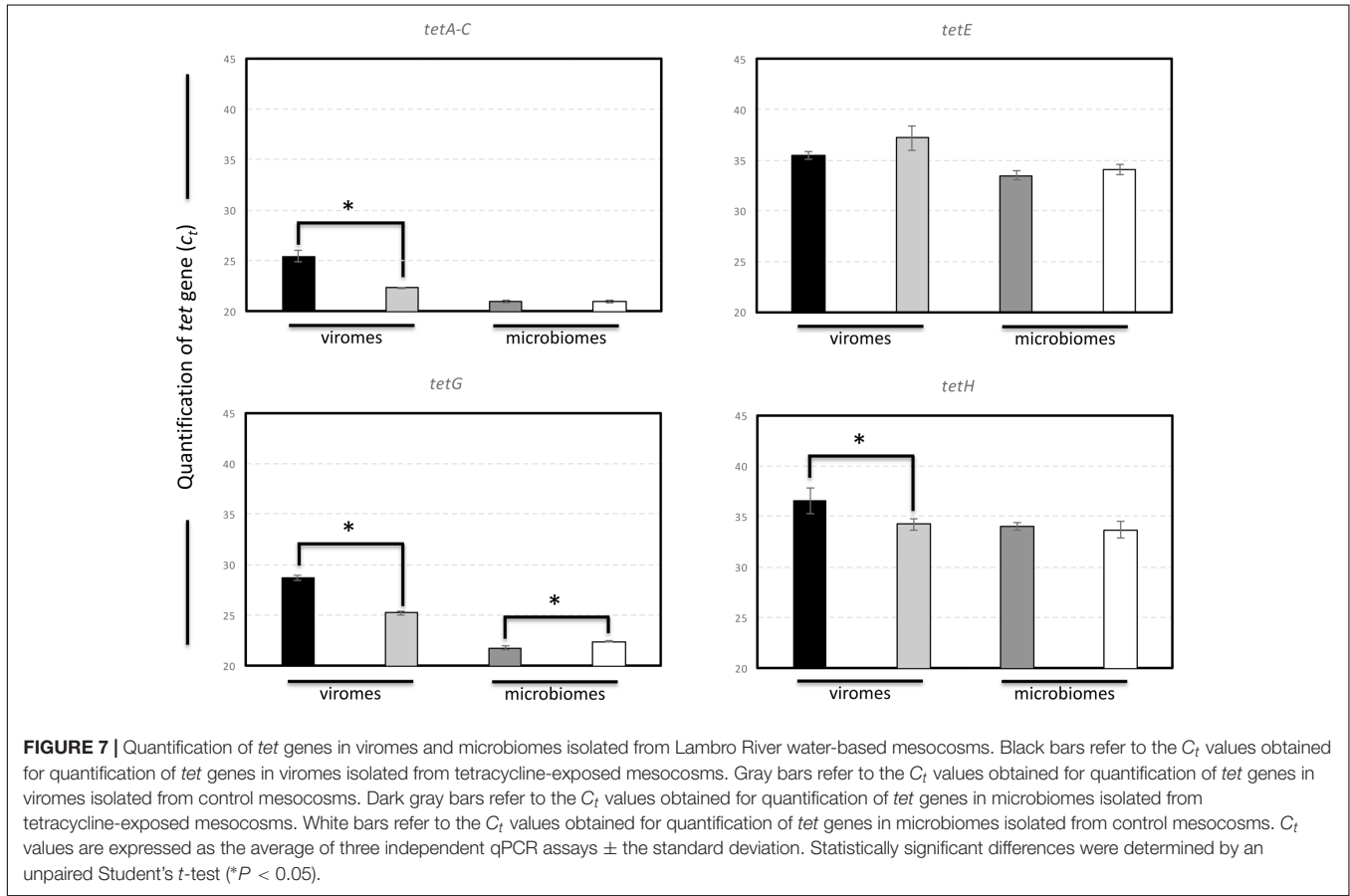


to tetracycline compared to *tet* genes quantified in viromes collected from non-perturbed mesocosms (lower C_t values), used as control. Interestingly, no significant changes in the abundance of *tet* genes were detected in microbiomes with the exception of *tetG*, which was higher in the microbiomes collected from tetracycline-exposed mesocosm compared to the controls. No significant changes in the frequency of tetracycline-resistant colonies were detected between the two mesocosms after tetracycline addition. The overall data obtained suggested that the environmental perturbation with tetracycline determined a mobilization of *tet* genes, the majority of which decreased in viral populations. Conversely, an increase of *tet* genes in the microbiomes was detected only for *tetG* (Figure 7).

DISCUSSION

The role of viruses in controlling and shaping microbial communities is getting more and more evident (Modi et al., 2013; Muniesa et al., 2013a,b; Danovaro et al., 2016). In most of these studies viromes were characterized in terms of abundance of VLPs, and in terms of relative abundance of the taxonomical classes of viruses, without a deep description of the microbial genes harbored by viral genomes in terms both of taxonomy and function. Some authors hypothesize

that phageome may represent a community-based mechanism for protecting microbial community, preserving its functional robustness during environmental stress (Modi et al., 2013). Modi et al. (2013) explored the murine gut phageome as a potential genetic reservoir for bacterial adaptation following antibiotic treatment. The authors observed that antibiotic treatment leads to the enrichment of phage-encoded genes that confer resistance via disparate mechanisms to the administered drug. Based on all these considerations, we decided to explore the quality and abundance of microbial gene sequences in aquatic viromes isolated from river samples in order to set up a protocol suitable to describe the phage-bacterial ecological network. Lambro River was chosen as a model because it is known as a high environmental risk area for more than 20 years due to contamination with heavy metals and persistent organic pollutants. In all water samples collected we were able to isolate and concentrate the microbiota and the viral populations adopting a TFF-based procedure. A CsCl gradient centrifugation followed by a DNase treatment of concentrated VLPs limited under the 0.2% threshold the contamination (qPCR evaluated) of bacterial DNA in viral DNA extraction. This extremely low level of contamination was reflected, after viral genome amplification and shotgun sequencing, in values lower than 0.2‰ of abundance of reads associated to ribosomal genes on the total of reads in two out of three viromes analyzed.



These values of contamination are comparable and sometimes lower than those measured in viromes already available in public database (Supplementary Table S3) (López-Bueno et al., 2009; Rosario et al., 2009; Fancello et al., 2013; Zablocki et al., 2014).

As reported by other authors, viromes harbored a huge amount of phage-encoded bacterial sequences (López-Bueno et al., 2009; Rosario et al., 2009; Roux et al., 2012; Fancello et al., 2013; Modi et al., 2013). The functional classification analysis carried out on viromes and microbiomes highlights

interesting differences in the relative abundance of the different gene classes. Whereas the three microbiomes showed a very similar distribution for each gene functional class, the three viromes showed potential differences. The reads associated with genes coding for functions related to the microbial metabolism apparently decreased from sample A (37%) to sample C (15%), which was enriched with reads associated with genes coding for nucleotide transport and metabolism category (56% compared to 11 and 14% in samples A and B, respectively). The ecological reasons for these differences are not known, but the bacterial genes present in the viromes surely reflect the gene content harbored in the genomes of the bacterial taxa subjected to viral lytic cycles. Concerning the abundance of ARGs- and MRGs-associated reads (abundance of ARGs and MRGs reads on the total amounts of reads) (Figure 8), we apparently observed their increase in the three microbiomes from samples A to C and an opposite trend in the three viromes (Figure 8).

The increase of ARGs and MRGs-associated reads from samples A to C could be coinciding with the increased industrialization/urbanization level of the geographical area where samples were collected. This association, if confirmed by further studies, is in agreement with a previously formulated hypothesis that considers the exposure of environmental landscapes to pollutants (antibiotics, metals, and disinfectants) a risk for the selection and appearance of multidrug resistant strains (Berendonk et al., 2015). Moreover, a link between heavy metals and the maintenance/acquisition of antibiotic resistances has been documented (Knapp et al., 2011; Chen et al., 2015; Teixeira et al., 2016). This correlation is caused by cross- and co-resistance phenomena. Cross-resistance occurs when the same mechanism reduces the susceptibility to antibiotics and metals simultaneously, whereas co-resistance happens when different resistance mechanisms (ARGs and MRGs) are co-localized in the same genetic locus, and therefore they are co-transferred in case of horizontal gene transfer mechanisms (Baker-Austin et al., 2006). The contamination of the Lambro River with heavy metals is well documented (Camusso et al., 1995; Galli et al., 1998; Barghigiani et al., 2001; Farkas et al., 2007; Viganò et al., 2008), and the toxic effects of these pollutants are also well known (Tchounwou et al., 2012). In biological systems, heavy metals have been reported to affect cellular structures, several physiological activities (Wang and Shi, 2001), and metal ions have been found to cause DNA damage. Therefore, the presence of heavy metals in water environment represents a selective pressure that could force the co-selection of MRGs and ARGs in the microbiome in the absence of antibiotic molecules or in the presence of low concentrations of antibiotic molecules. In this context, it is worth mentioning the increased relative abundance of mobilome-associated reads in the viromes of sample A, thus increasing the probability of transduction events in that environment.

In a previous study carried out on an aquaculture water sample, we observed that microbial ORFs identified in the virome did not reflect the microbiome taxonomy, as determined by 16S rRNA gene profiling or shotgun metagenomic analysis, suggesting that microbial genes mobilized in the genome of

viruses could be considered to be remnants of past recombination events rather than a picture of the current microbial diversity (Colombo et al., 2016). Likewise, in this study we observed the same phenomena (Figure 3) in all three samples analyzed, thereby confirming that viromes represent a picture of past microbial populations.

To understand whether the microbial genes associated with the viromes were accessible to members of the aquatic microbiota, we carried out a set of mesocosms focusing our attention on the mobilization of *tet* genes. We observed significant changes in the abundance of *tet* genes in viromes collected from mesocosms exposed to low dose of antibiotic compared to the abundance of these genes in the control mesocosms not exposed to tetracycline. These data are not sufficient to demonstrate gene mobilization from virome to microbiome as a consequence of the selective pressure introduced using a low dose of tetracycline, but they represent the first evidence of the modulation of specific genes within viral populations as a consequence of a specific environmental perturbation. Further studies are necessary in order to understand the mechanisms that drive the gene flow between microorganisms and the complex viral populations.

CONCLUSION

We have developed an integrated approach suitable for the exploration of microbiomes and viromes for microbial genes with a focus on ARGs and MRGs starting from fresh water river samples. This approach could represent a useful tool for the characterization of phage-bacterial ecological network in water environments, and for the understanding of environmental constraints which are relevant in the mobilization of ARGs and MRGs.

AUTHOR CONTRIBUTIONS

SC and GG have carried out the bioinformatic analysis, SC performed the virome isolation. SC, GD, and EN carried out the mesocosms-based and the plate-count experiments. SA, GD, and EN have analyzed the data. DM and SC have planned the experimental design and they wrote the manuscript.

ACKNOWLEDGMENTS

All bioinformatics analyses were performed using the facilities of CINECA SCAI (SuperComputing Applications and Innovation) (CINECA, Bologna, Italy) as part of the ISCRA VArFaWtr project.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fmicb.2017.01095/full#supplementary-material>

REFERENCES

- Abeles, S. R., Ly, M., Santiago-Rodriguez, T. M., and Pride, D. T. (2015). Effects of long term antibiotic therapy on human oral and fecal viromes. *PLoS ONE* 10:e0134941. doi: 10.1371/journal.pone.0134941
- Allen, H. K., Donato, J., Wang, H. H., Cloud-Hansen, K. A., Davies, J., and Handelsman, J. (2010). Call of the wild: antibiotic resistance genes in natural environments. *Nat. Rev. Microbiol.* 8, 251–259. doi: 10.1038/nrmicro.2312
- Baker-Austin, C., Wright, M. S., Stepanauskas, R., and McArthur, J. V. (2006). Co-selection of antibiotic and metal resistance. *Trends Microbiol.* 14, 176–182. doi: 10.1016/j.tim.2006.02.006
- Barghigiani, C., Ristori, T., and Scerbo, R. (2001). Assessment of water pollution and suitability to fish life in six Italian rivers. *Environ. Monit. Assess.* 66, 187–205. doi: 10.1023/A:1026447914322
- Berendonk, T. U., Manaia, C. M., Merlin, C., Fatta-Kassinos, D., Cytryn, E., Walsh, F., et al. (2015). Tackling antibiotic resistance: the environmental framework. *Nat. Rev. Microbiol.* 13, 310–317. doi: 10.1038/nrmicro.3439
- Camusso, M., Viganò, L., and Balestrini, R. (1995). Bioconcentration of trace metals in rainbow trout: a field study. *Ecotoxicol. Environ. Saf.* 31, 133–141. doi: 10.1006/eesa.1995.1053
- Chen, S., Li, X., Sun, G., Zhang, Y., Su, J., and Ye, J. (2015). Heavy metal induced antibiotic resistance in bacterium LSJC7. *Int. J. Mol. Sci.* 16, 23390–23404. doi: 10.3390/ijms161023390
- Chevreur, B., Wetter, T., and Suhai, S. (1999). Genome sequence assembly using trace signals and additional sequence information. *German Conf. Bioinform.* 99, 45–56.
- Colombo, S., Arioli, S., Guglielmetti, S., Lunelli, F., and Mora, D. (2016). Virome-associated antibiotic-resistance genes in an experimental aquaculture facility. *FEMS Microbiol. Ecol.* 92:fiw003. doi: 10.1093/femsec/fiw003
- Danovaro, R., Dell'Anno, A., Corinaldesi, C., Rastelli, E., Cavicchioli, R., and Krupovic, M. (2016). Virus-mediated archaeal hecatomb in the deep seafloor. *Sci. Adv.* 2:e1600492. doi: 10.1126/sciadv.1600492
- Deksissa, T., De Pauw, D., and Vanrolleghem, P. A. (2004). Dynamic in-stream fate modeling of xenobiotic organic compounds: a case study of linear alkylbenzene sulfonates in the Lambro River. Italy. *Environ. Toxicol. Chem.* 23, 2267–2278. doi: 10.1897/03-466
- Fan, W., Hamilton, T., Webster-Sesay, S., Nikolich, M. P., and Lindler, L. E. (2007). Multiplex real-time SYBR Green I PCR assay for detection of tetracycline efflux genes of Gram-negative bacteria. *Mol. Cell. Probes* 21, 245–256. doi: 10.1016/j.mcp.2006.12.005
- Fancello, L., Trape, S., Robert, C., Boyer, M., Popgeorgiev, N., Raoult, D., et al. (2013). Viruses in the desert: a metagenomic survey of viral communities in four perennial ponds of the Mauritanian Sahara. *ISME J.* 7, 359–369. doi: 10.1038/ismej.2012.101
- Farkas, A., Erratico, C., and Viganò, L. (2007). Assessment of the environmental significance of heavy metal pollution in surficial sediments of the River Po. *Chemosphere* 68, 761–768. doi: 10.1016/j.chemosphere.2006.12.099
- Galli, P., Crosa, G., and Occhipinti Ambrogio, A. (1998). Heavy metals concentrations in acanthocephalans parasites compared to their fish host. *Chemosphere* 37, 2983–2988. doi: 10.1016/S0045-6535(98)00340-3
- Guglielmetti, S., Fracassetti, D., Taverniti, V., Del Bo', C., Vendrame, S., Klimis-Zacas, D., et al. (2013). Differential modulation of human intestinal bifidobacterium populations after consumption of a wild blueberry (*Vaccinium angustifolium*) drink. *J. Agric. Food Chem.* 61, 8134–8140. doi: 10.1021/jf402495k
- Haque, M. M., Bose, T., Dutta, A., Reddy, C. V., and Mande, S. S. (2015). CS-SCORE: rapid identification and removal of human genome contaminants from metagenomic datasets. *Genomics* 106, 116–121. doi: 10.1016/j.ygeno.2015.04.005
- Hyatt, D., Chen, G. L., Locascio, P. F., Land, M. L., Larimer, F. W., and Hauser, L. J. (2010). Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. doi: 10.1186/1471-2105-11-119
- Knapp, C. W., McCluskey, S. M., Singh, B. K., Campbell, C. D., Hudson, G., and Graham, D. W. (2011). Antibiotic resistance gene abundances correlate with metal and geochemical conditions in archived Scottish soils. *PLoS ONE* 6:e27300. doi: 10.1371/journal.pone.0027300
- López-Bueno, A., Tamames, J., Velázquez, D., Moya, A., Quesada, A., and Alcamí, A. (2009). High diversity of the viral community from an Antarctic lake. *Science* 326, 858–861. doi: 10.1126/science.1179287
- Marti, E., Huerta, B., Rodríguez-Mozaz, S., Barceló, D., Jofre, J., and Balcázar, J. L. (2014). Characterization of ciprofloxacin-resistant isolates from a wastewater treatment plant and its receiving river. *Water Res.* 15, 67–76. doi: 10.1016/j.watres.2014.05.006
- McArthur, A. G., Waglechner, N., Nizam, F., Yan, A., Azad, M. A., Baylay, A. J., et al. (2013). The comprehensive antibiotic resistance database. *Antimicrob. Agents Chemother.* 57, 3348–3357. doi: 10.1128/AAC.00419-13
- Modi, S. L., Lee, H. H., Spina, C. S., and Collins, J. J. (2013). Antibiotic treatment expands the resistance reservoir and ecological network of the phage metagenome. *Nature* 499, 219–223. doi: 10.1038/nature12212
- Muniesa, M., Colomer-Lluch, M., and Jofre, J. (2013a). Could bacteriophages transfer antibiotic resistance genes from environmental bacteria to human-body associated bacterial populations? *Mob. Genet. Elements* 3:e25847. doi: 10.4161/mge.25847
- Muniesa, M., Colomer-Lluch, M., and Jofre, J. (2013b). Potential impact of environmental bacteriophages in spreading antibiotic resistance genes. *Future Microbiol.* 8, 739–751. doi: 10.2217/fmb.13.32
- Pal, C., Bengtsson-Palme, J., Rensing, C., Kristiansson, E., and Larsson, D. G. (2014). BacMet: antibacterial biocide and metal resistance genes database. *Nucleic Acids Res.* 42, D737–D743. doi: 10.1093/nar/gkt1252
- Pao, S. S., Paulsen, I. T., and Saier, M. H. (1998). Major facilitator superfamily. *Microbiol. Mol. Biol. Rev.* 62, 1–34.
- Rosario, K., Nilsson, C., Lim, Y. W., Ruan, Y., and Breitbart, M. (2009). Metagenomic analysis of viruses in reclaimed water. *Environ. Microbiol.* 11, 2806–2820. doi: 10.1111/j.1462-2920.2009.01964.x
- Roux, S., Enault, F., Robin, A., Ravet, V., Personnic, S., Theil, S., et al. (2012). Assessing the diversity and specificity of two freshwater viral communities through metagenomics. *PLoS ONE* 7:e33641. doi: 10.1371/journal.pone.0033641
- Roux, S., Krupovic, M., Debroas, D., Forterre, P., and Enault, F. (2013). Assessment of viral community functional potential from viral metagenomes may be hampered by contamination with cellular sequences. *Open Biol.* 3:130160. doi: 10.1098/rsob.130160
- Taylor, N. G., Verner-Jeffreys, D. W., and Baker-Austin, C. (2011). Aquatic systems: maintaining, mixing and mobilising antimicrobial resistance? *Trends Ecol. Evol.* 26, 278–284. doi: 10.1016/j.tree.2011.03.004
- Tchounwou, P. B., Yedjou, C. G., Patlolla, A. K., and Sutton, D. J. (2012). Heavy metals toxicity and the environment. *EXS* 101, 133–164. doi: 10.1007/978-3-7643-8340-4_6
- Teixeira, P., Tacão, M., Alves, A., and Henriques, I. (2016). Antibiotic and metal resistance in a ST395 *Pseudomonas aeruginosa* environmental isolate: a genomics approach. *Mar. Pollut. Bull.* 110, 75–81. doi: 10.1016/j.marpolbul.2016.06.086
- Thurber, R. V., Haynes, M., Breitbart, M., Wegley, L., and Rohwer, F. (2009). Laboratory procedures to generate viral metagenomes. *Nat. Protoc.* 4, 470–483. doi: 10.1038/nprot.2009.10
- Viganò, L., Patrolocco, L., Polesello, S., and Pagnotta, R. (2008). Interactions between trophic and toxic factors in a polluted urban river. *Ecotoxicol. Environ. Saf.* 69, 49–57. doi: 10.1016/j.ecoenv.2007.01.001
- Wang, S., and Shi, X. (2001). Molecular mechanisms of metal toxicity and carcinogenesis. *Mol. Cell. Biochem.* 222, 3–9. doi: 10.1023/A:1017918013293
- Weynberg, K. D., Wood-Charlson, E. M., Suttle, C. A., and van Oppen, M. J. (2014). Generating viral metagenomes from the coral holobiont. *Front. Microbiol.* 5:206. doi: 10.3389/fmicb.2014.00206
- Williamson, S. J., Allen, L. Z., Lorenzi, H. A., Fadrosch, D. W., Bami, D., Thiagarajan, M., et al. (2012). Metagenomic exploration of viruses throughout the Indian Ocean. *PLoS ONE* 7:e42047. doi: 10.1371/journal.pone.0042047
- Yu, Z., Michel, F. C. Jr., Hansen, G., Wittum, T., and Morrison, M. (2005). Development and application of real-time PCR assays for quantification of genes encoding tetracycline resistance. *Appl. Environ. Microbiol.* 71, 6926–6933. doi: 10.1128/AEM.71.11.6926-6933.2005

Zablocki, O., van Zyl, L., Adriaenssens, E. M., Rubagotti, E., Tuffin, M., Cary, S. C., et al. (2014). High-level diversity of tailed phages, eukaryote-associated viruses, and virophage-like elements in the metaviromes of antarctic soils. *Appl. Environ. Microbiol.* 80, 6888–6897. doi: 10.1128/AEM.01525-14

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Colombo, Arioli, Neri, Della Scala, Gargari and Mora. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.