

Oviductal microvesicles and their effect on *in vitro* maturation of canine oocytes



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Abstract

The effect of conditioned medium (CM) or microvesicles (MVs), secreted by multicellular spheroids of oviductal cells, and the involvement of some microRNAs (miRNAs) were investigated in canine oocyte maturation. To generate CM, spheroids were cultured for 3 days. MVs were obtained by ultracentrifugation of CM at 100,000 g and measured for size and concentration by NanoSight instrument. Cumulus-oocyte complexes (COCs) were matured at 38.5°C with 5% CO₂ and 5% of O₂ in synthetic oviductal fluid (SOF) in biphasic systems: for 24 h, with 5.0 µg/mL of LH and for other 48 h with 10% oestrous bitch serum. SOF was used as control (CTR) or supplemented with 10% CM or 25–50–75–100–150 × 10⁶ MVs/mL labeled with PKH-26. Results show that multicellular aggregates secreted shedding vesicles. By fluorescence microscopy, the incorporation of labeled MVs was visible only at 72 h in oocyte cytoplasm. These MVs had a positive effect ($P < 0.05$) on maturation rate (MII) at the concentration of 75 and 100 × 10⁶ MVs/mL compared to CM and CTR (20.34% and 21.82% vs 9.09% and 8.66% respectively). The concentration of 150 × 10⁶ MVs/mL provided only 9.26% of MII. The expression of three specific miRNAs (miR-30b, miR-375 and miR-503) was studied. The lower rate of MII with the higher concentration of MVs is possibly due to the high level of miR-375. In conclusion, the oviductal MVs could be involved in cellular trafficking during oocyte maturation and their possible use *in vitro* could facilitate the exploitation of canine reproductive biotechnologies.

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Q5 Introduction

The limited efficiency of canine oocytes *in vitro* maturation (IVM) is conceivably due to either the peculiar reproductive physiology of the dog or to the deficient maturation media employed up to now. Many ultrastructural changes of oocytes, during acquisition of their competence, are common to all mammalian oocytes (Tesoriero 1982), but it is known that canine oocytes display important differences compared to other mammals. Canine oocytes acquire their competence in the intra- and extra-follicular (oviductal) environments (Holst & Phemister 1971) as they are ovulated spontaneously at an immature diploid stage (germinal vesicle (GV)). Subsequent stages of the meiotic maturation, up to completion of meiosis I, and extrusion of the first polar body (stage of metaphase II-MII) occur in the oviduct. Although the exact time of oocyte maturation has not been determined, it appears not to occur until 2 or 3 days after ovulation (Holst & Phemister 1971). Moreover, canine oocyte shows large amounts of lipid yolk material (Guraya 1965) and the

persistence of the cumulus oophorus for several days, until after fertilization (Holst & Phemister 1971).

In order to reproduce *in vitro* the intratubal conditions, Hewitt and England (1998) were the first researcher who investigated the effect of introducing oviductal elements, such as synthetic oviductal fluid (SOF) to the culture environment or co-culturing with isolated canine oviductal epithelial cells. Afterward, canine IVM has been assessed in co-culture with other cell types (Otoi *et al.* 2000a, Hatoya *et al.* 2006, Saikhun *et al.* 2008) or even after insertion of the oocytes into an explanted oviduct cultured *in vitro* (Luvoni *et al.* 2003). Moreover, to mimic the *in vivo* environment, Songsasen *et al.* (2002) and Silva *et al.* (2009) suggested to culture canine oocytes in low oxygen tension and Nickson *et al.* (1993) the presence of 10% of oestrus bitch serum.

Whatever the maturation system, only 10–20% of oocytes reaches metaphase I (MI) or metaphase II (MII) stages (Luvoni *et al.* 2005). The few MII oocytes are unable to be fertilized and undergo early embryonic development (Viaris de Lesegno *et al.* 2008,

Chebrout *et al.* 2009). Low rates of MII could also be due to low competence of the selected oocytes or inadequate culture conditions (Luvoni *et al.* 2005). Concerning oocytes selection, the age of the donor bitch (Hewitt & England 1998, Songsasen *et al.* 2002), the stage of the reproductive cycle (Yamada *et al.* 1993, Otoi *et al.* 2001), the oocyte diameter (Hewitt & England 1998, Otoi *et al.* 2000b) and the size of the ovarian follicle (Songsasen & Wildt 2005), are factors influencing meiotic competence of canine oocytes cultured *in vitro*. In particular, oocyte diameter may be a useful selection parameter. It has been reported that large oocytes (>120 µm diameter) show a higher frequency of maturation than oocytes <110 µm diameter (Otoi *et al.* 2000a, Ariu *et al.* 2012).

Regarding the IVM *media*, other attempts have been made to identify the factors involved in the resumption and progression of maturation *in vivo*, so that improvements in cultural conditions have been achieved. Indeed, recently, a biphasic system based on a sequential combination of different hormones indicated that MI+MII rates were higher among oocytes cultured in sequential media than those matured in the traditional way (Evecen *et al.* 2011). This system improved cortical granule distribution (an indication of cytoplasmic maturation) and its relationship to the nuclear status (Apparicio *et al.* 2014), although the MII rate remained around 20%. This clearly indicates the need to further study the mechanisms leading to canine oocyte maturation and to identify specific signals that induce *in vivo* meiotic resumption and progression. The co-culture system provided the best results compared to the use of standard TCM-199 medium, but achievements are still far from the maturation rate obtained in the oviductal environment *in vivo* that, in the dog, is beneficial for supporting oocytes maturation. *In vivo*, canine oocytes are exposed to the oviductal secretions and they are in contact with oviductal mucosa for long time, and this may be crucial in the acquisition of oocyte competence. The advantages of the *in vitro* co-culture system suggest that paracrine mechanisms of communication exist between oviductal cells and oocytes. In fact, various growth factors, receptors and binding proteins are secreted by cells in the culture medium, but they could also be produced by the oocytes themselves (Gilchrist *et al.* 2008). Conditioned media (CM), composed of factors secreted by cells during their culture, were mainly developed and used in *in vitro* bovine embryo production. Some of these cell-free media, produced, for example, by culturing granulosa, cumulus or oviductal cells, demonstrated a similar effect in the development of embryos produced by co-culture systems (Rieger *et al.* 1995) or had a positive effect on the quality of *in vitro*-produced bovine embryos (Lopera-Vasquez *et al.* 2016). Conditioned medium is composed of soluble factor, but recent studies have also uncovered the existence of microvesicles (MVs), which are released by cells into the extracellular environment

(Ratajczak *et al.* 2006, Camussi *et al.* 2010). These MVs have been shown to contain proteins, lipids (specifically high levels of sphingomyelins), a variety of RNA species, including microRNAs (miRNAs) and mRNA fragments (Thery *et al.* 2009). MVs could serve as vehicles for the transfer of these molecules between cells (Zhang *et al.* 2009, Raposo & Stoorvogel 2013). The *in vitro* use of feeder cell or CM probably involves MVs that may participate in this required cross-talk, as studied in the bovine *in vitro* embryo production by Lopera-Vasquez *et al.* (2016).

To understand the signals that are carried from epithelial oviductal cells to canine oocytes, we studied the effect of different components of the secretome of this cell line. This effect has never been studied and, since it is difficult to study these communications *in vivo*, we tested these interactions in *in vitro* canine oocyte maturation. Canine oviductal cell secretome, composed of CM *in toto*, MVs and supernatant (SN), collected after removal of MVs, was investigated to know which of these components, if not all, have effects on oocyte maturation, in terms of MII rate. *In vitro* culture was performed taking into account all the best conditions previously described, as the use of oocytes with diameter of 110–120 µm or greater, the biphasic system, the SOF *medium* and the low oxygen tension.

Materials and methods

All chemicals in this study were purchased from Sigma-Aldrich unless otherwise stated, whereas disposable materials were purchased from Euroclone (Milano, Italy).

Experimental design

This study explores the MII production rate of canine oocytes co-cultured with oviductal epithelial cell in monolayer or in multicellular spheroids. Moreover, this study investigates the effects of supplementing the IVM medium with 10% of CM *in toto* or SN or with different amounts of MVs. The ability of oocytes to uptake labeled MVs and the identification of some miRNAs, responsible for oocytes maturation in other species, both in oviductal cells and in the MVs, are also investigated.

Animals

To isolate epithelial cell and oocytes, oviducts and ovaries were recovered from 64 healthy adult domestic bitches (over 12 months old) referred to the University Veterinary Hospital for ovariectomy at owners convenience. In this study, after approval by the Ethical Committee of the University of Milan and written owner's consent, all procedures were carried following standard veterinary practice and in accordance to 2010/63 EU directive on animal protection and Italian Law (D.L. No. 116/1992).

To collect epithelial cells, oviducts were obtained from bitches at oestrous stage, while to collect oocytes, ovaries of bitches at all different stages of oestrous cycle were used.

The stage of oestrous cycle of these animals was determined through the evaluation of ovarian morphology. Three stages were differentiated:

- Anaestrus, when ovaries did not show apparent follicles or corpora lutea.
- Estrus (follicular stage), when on the ovarian surface there were one or more 2–8 mm in diameter follicles.
- Diestrus, when one or more corpora lutea were present on the ovarian surface.

In addition, the oestrous stage was confirmed by vaginal cytology and blood serum progesterone concentration.

Cell isolation and culture

To obtain oviductal epithelial cells, ovaries with ovarian bursa were maintained in physiological saline solution (0.9% NaCl), supplemented with 1% penicillin (100 IU/mL)–streptomycin (100 mg/mL) at 38°C and transported to the laboratory within 1 h. Oviducts were dissected free from ovarian bursa fat and washed in fresh physiological saline solution. The oviductal lumen was rinsed with phosphate-buffered saline (PBS), supplemented with 10% fetal calf serum (FCS), by means of a fine blunt needle to remove possible oocytes within the lumen. Then, two methods were assessed to isolate epithelial cells for co-culture in canine oocyte IVM. In the first method, oviducts were slit lengthways and the luminal surface was lightly scraped with a microblade. In the second method, oviducts were squeezed out by sterile tweezers to release epithelial cells into the culture medium composed of high-glucose Dulbecco modified Eagle's medium (HG-DMEM) supplemented with 10% of FCS, 2 mM L-glutamine and penicillin (100 IU/mL)–streptomycin (100 mg/mL). This method has been reported for the sampling of oviductal epithelial cells in the cow (Eystone & First 1989, Xu *et al.* 1992). By the first method, only isolated cells were collected. These cells were stained with trypan blue to assess their viability, and concentration was measured using a Burker chamber. Approximately, 60×10^4 cells/mL/well were seeded and cultured in monolayer (Fig. 1A).

By the second method, in addition to isolated cells, oviductal exfoliated mucosal portions were observed at microscopy. They were collected through a 10 μ L micropipette tip and seeded in different wells of a 4-well dish. The culture of these structures was performed in suspension (Fig. 1B) for three days until they organized themselves into multicellular spheroids. The presence of cilia maintained them in constant rotary motion (Fig. 1C).

For both kinds of epithelial cells, the culture was performed at 38.5°C in an atmosphere with 5% of CO₂ and 90% of humidity with culture medium. At the time of their formation, multicellular spheroids were collected and used, in addition to canine IVM, for electron microscopy analysis, immunocytochemistry and CM production.

Transmission electron microscopy

For histological and ultrastructural analyses, multicellular spheroids were fixed in a mixture of 2% paraformaldehyde and 2.5% glutaraldehyde in 0.1 M sodium cacodylate buffered solution at pH 7.4 for 1 h at room temperature. After washing

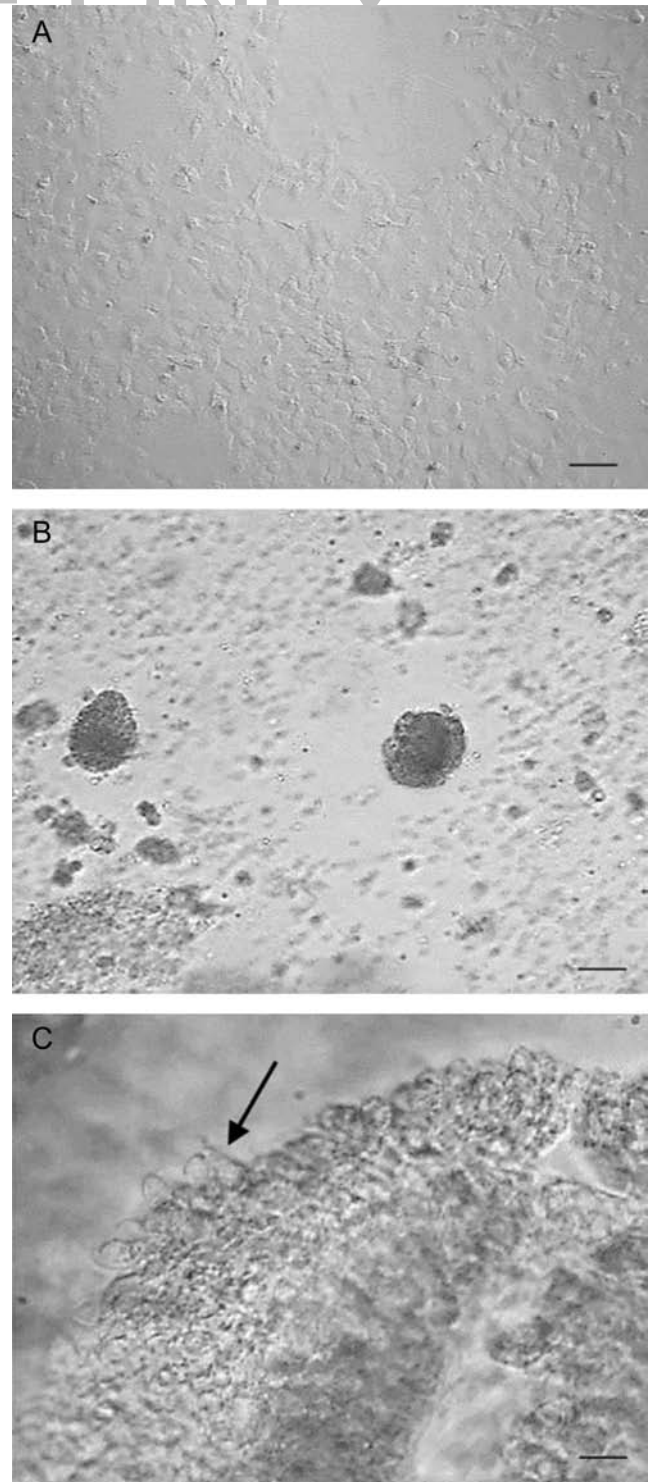


Figure 1 Morphology of oviductal cells. (A) Monolayer of epithelial cells. (B) Multicellular spheroids cultured in suspension. (C) Arrow indicates cilia. Light microscope images. Magnification 20 \times . Scale bar of figure (A) = 20 μ m; scale bar of figure (B) = 30 μ m; scale bar of figure (C) = 2 μ m.

in the same buffer, samples were post-fixed in 1% OsO₄, 1.5% potassium ferrocyanide in 0.1 M cacodylate for 1 h in dark condition on ice. After several washing in distilled water,

samples were stained with 0.5% uranyl acetate in water overnight at 4°C and, finally, were dehydrated in a graded ethanol series (30, 50, 70, 80, 90, 96% for 5 min each and washed three times with absolute ethanol for 10 min each). The samples were infiltrated with ethanol and resin (Araldite-Epon) at volumetric proportions of 1:1 for 2 h, and then in 100% Epon twice for 1 h each and, finally, polymerized at 60°C for 48 h. Sectioning was performed using an Ultracut E microtome (Reichert, Austria). Semithin sections of about 0.5 µm were collected onto microscope slides and stained with methylene blue-azure II and basic fuchsine (D'Amico *et al.* 2005). For the ultrastructural analysis, sections of 70 nm were collected on 300-mesh uncoated copper grid. All thin sections were not counterstained and observed with a Zeiss LEO 912ab Energy Filtering TEM operating at 120 kV. Digital images were acquired using a CCD-BM/1K system operating with the iTEM (Olympus Soft Imaging Solutions).

Immunocytochemistry of multicellular spheroids

Indirect immunofluorescence was performed to assess the expression of cytokeratin, a protein normally found in cells of epithelial origin and vimentin, a cytoskeletal protein expressed in cells of mesenchymal origin. Multicellular spheroids were washed in PBS supplemented with bovine serum albumin (BSA 0.03 mg/mL), fixed in 4% paraformaldehyde (20–30 min at 4°C) and washed in PBS for 10 min. Samples were permeabilized with 0.3% Triton X100, in PBS for 30 min at room temperature (RT) and washed in PBS supplemented with 1% BSA (PBS/1% BSA). Non-specific binding was blocked by incubating the samples in 20% normal donkey serum, 1% BSA and 0.1% Triton X100 in PBS for 30 min at RT. The samples were incubated 1:1000, overnight at 4°C with a mouse monoclonal anti-Pan-cytokeratin, (AE1/AE3 Zymed San Francisco, CA, USA) or a mouse monoclonal anti vimentin (VIM, clone 3B4 Agilent Technologies Dako, Denmark, 1:1000) in PBS/1% BSA. Samples were extensively washed in PBS/1% BSA and incubated for 1 h at RT with a donkey anti-mouse secondary antibody conjugated with Alexa Fluor 488 (Invitrogen), diluted 1:500 in PBS containing 1% BSA. Samples were rinsed twice in PBS/1% BSA and mounted in an antifade medium (Vecta Shield; Vector Laboratories, Inc., Burlingame, CA, USA) supplemented with DAPI dilactate (1 µg/mL). Samples were analyzed on an epifluorescence microscope (Eclipse E600; Nikon) equipped with a 40× objective, a digital camera and software (NIS, Elements Imaging Software; Nikon). Negative controls were performed by omitting the primary antibody. The excitation wavelength for DAPI was 330–350 nm and for Alexa Fluor488 (FITC) was 465–495 nm.

Conditioned medium, microvesicles and supernatant production

CM, MVs and SN were obtained by multicellular spheroids only. Multicellular spheroids were cultured for 3 days with 1 mL per well of a serum-free culture medium (Lonza, Milano, Italy) added with 1% penicillin (100 IU/mL)–streptomycin (100 mg/mL) and 2 mM L-glutamine. Conditioned media of three different samples were collected, pooled and centrifuged

at 2500 g for 20 min to remove the cellular debris. A portion of this CM *in toto* was stored at –20°C. To obtain MVs and SN, another aliquot of CM was ultracentrifuged (Beckman Coulter Optima L – 100K) at 100,000 g for 1 h at 4°C and SN was collected and stored at –20°C for later use. The pellet was washed in serum-free medium 199 containing N-2-hydroxyethylpiperazine-N-2-ethanesulfonic acid (HEPES) 25 mM and submitted to a second ultracentrifugation under the same conditions. The resulting pellet, composed of MVs, was fractionated for MVs analysis or used for the *in vitro* test.

Measurements of MVs

Size and concentration of MVs secreted by multicellular spheroids were evaluated by NanoSight LM10 instrument (Nanoparticle tracking analysis, NTA, NanoSight Ltd., Amesbury, UK), which permits discrimination of microparticles less than 1 µm in diameter. The software (NTA 2.0 analytic software) allows the analysis of video images of particle movement under Brownian motion and the calculation of diffusion coefficient, sphere equivalent and hydrodynamic radius of particles by using the Stokes–Einstein equation. This instrument is equipped with a 405 nm laser and a high-sensitivity sCMOS camera (OrcaFlash2.8, Hamamatsu C11440, NanoSight Ltd). Videos were collected and analyzed using the NTA software with the minimal expected particle size, minimum track length and blur setting, all set to automatic. Ambient temperature was recorded manually and did not exceed 25°C. Five microliters of each sample were diluted in sterile physiological solution to a final volume of 1 mL. Samples were analyzed within 15 min of the initial dilution with a delay of 10 s between sample introduction and the start of the measurement. For each sample, multiple videos of 30-s duration were recorded generating replicate histograms that were averaged.

MVs labeling with PKH-26

To trace MVs *in vitro* by fluorescence microscopy, MVs from spheroidal aggregates were labeled with the red fluorescence aliphatic chromophore intercalating into lipid bilayers, PKH-26 dye. Briefly, after ultracentrifugation, the MV pellet was diluted to 1 mL with PKH-26 kit solution and 2 µL of fluorochrome were added to this suspension and incubated for 15 min at 38.5°C. Thereafter, 7 mL of serum-free HG-DMEM were added to the suspension that was ultracentrifuged again at 100,000 g for 1 h at 4°C. The final pellet was immediately resuspended in HG-DMEM and stored at –20°C.

miRNA analyses by RNA extraction and PCR amplification

MV pellet was subjected to RNase digestion to remove extraneous ribonucleic acids (Skog *et al.* 2008). Total RNA was isolated from pool of different MVs and multicellular spheroid preparations using the NucleoSpin mRNA kit (Macherey-Nagel, Germany), in combination with TRIzol lysis and purification of small and large RNA in one fraction (total RNA). RNAs were quantified using a NanoDrop ND-1000

spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA). RNA quality was checked using the Agilent Bioanalyser 2100 (Agilent), where the presence of small RNAs was verified in both MV and cell samples.

RNAs from all samples were reverse transcribed with the miScript Reverse Transcription Kit. The cDNA was then pre-amplified by using miScript PreAMP PCR Kit (all from Qiagen), following manufacturer instruction with some modification: miScript PreAMP Primer Mix was replaced with miR-specific primers: hsa-miR-30b, has-miR-375 and cfa-miR-503, SNORD95 as forward primer, and miScript Universal Primer as reverse primer in separate reactions. *Homo sapiens* hsa miRNAs were perfectly homologous with *Canis lupus familiaris* cfa miRNA sequence for miR-30b and miR-375. PCR was performed on pre-amplified products by using PCR Master Mix (2x) (Thermo Fisher Scientific), with same primer couple: hsa-miR-30b, hsa-miR-375, cfa-miR-503 and SNORD95 in combination with miScript Universal Primer. The small nucleolar snoRNA, C/D Box 95 SNORD95 was used as positive control. Negative controls using water in place of the pre-Amp product were performed alongside each reaction. The cycling conditions were 3 min at 95°C, followed by 35 cycles of 30s at 95°C, 30s at 58°C and 1 min at 72°C, and finally 7 min at 72°C. The amplified PCR products were separated electrophoretically on 2.5% agarose gels, and visualized under UV, by using GeneRuler 50bp as DNA ladder (Thermo Fisher Scientific).

Bitch serum production

~~In order to reproduce the most faithfully as possible the in vivo conditions,~~ we used serum obtained by three bitches at peri-ovulation oestrous stage, as determined by progesterone concentration and vaginal cytology. The serum was prepared from blood that after clotting was centrifuged at 3000g for 10 min. The supernatant was collected and serum was heat-inactivated for 30 min at 56°C and stored at -20°C until use. Serum progesterone concentration was evaluated by ELFA method validated for use in the dog by Enzyme Linked Fluorescent Assay (MiniVidas, Biomerieux) (Brugger *et al.* 2011).

Collection of ovaries and compact COCs retrieval

Ovaries were collected from bitches during routine ovariectomy at University Veterinary clinics. No attempt was made to divide the oocytes according to the stage of the oestrous cycle of the animals. After excision, ovaries were put in a sterile container in physiological solution supplemented with 1% penicillin–streptomycin. Within 1 h of collection, ovaries were washed several times in fresh PBS and ovarian bursa was removed. Ovaries were put in TCM-199 medium with 25 mM HEPES supplemented with 0.1% BSA and 1% penicillin–streptomycin at 38.5°C and were sliced repeatedly by scalpel to release oocytes.

After collection, immature oocytes were screened under stereomicroscope and selected based on their diameter and morphology. Damaged oocytes were eliminated (Fig. 2A). The selected oocytes were 110–120 µm or more in diameter,

darkly pigmented and completely surrounded by two or more layers of cumulus cells (Fig. 2B).

In vitro maturation of oocytes

Compact COCs were cultured in 4-wells dishes containing 500 µL of culture medium under 500 µL of equilibrated paraffin oil for a maximum of 30 oocytes per well. The incubation took place in incubator with 5% CO₂, 5% O₂ and 90% N₂ for 72 h.

The culture medium used was SOF manufactured in laboratory and composed of NaCl (1.1 M), KCl (72 mM), KH₂PO₄ (12 mM), MgSO₄ (7.4 mM), DL-lactic acid (60%) (50 mM), NaHCO₃ (250 mM), phenol red (260 mM), sodium pyruvate (100 mM), CaCl₂ di-hydrate (178 mM), sodium HEPES (125 mM), acid HEPES (125 mM), L-glutamine (30.8 mM), glycine (500 mM), alanine (84.2 mM), non-essential MEM (100x), BME (100x), myo-inositol (2.8 mM), sodium tri-citrate di-hydrate (340 mM), FBS (2%), BSA (0.005 g/mL), pyruvate (11 mg/mL) and gentamicin (5 µL/mL).

The medium was filtered and allowed to equilibrate for 1 h in the same incubator before being used. SOF was used in

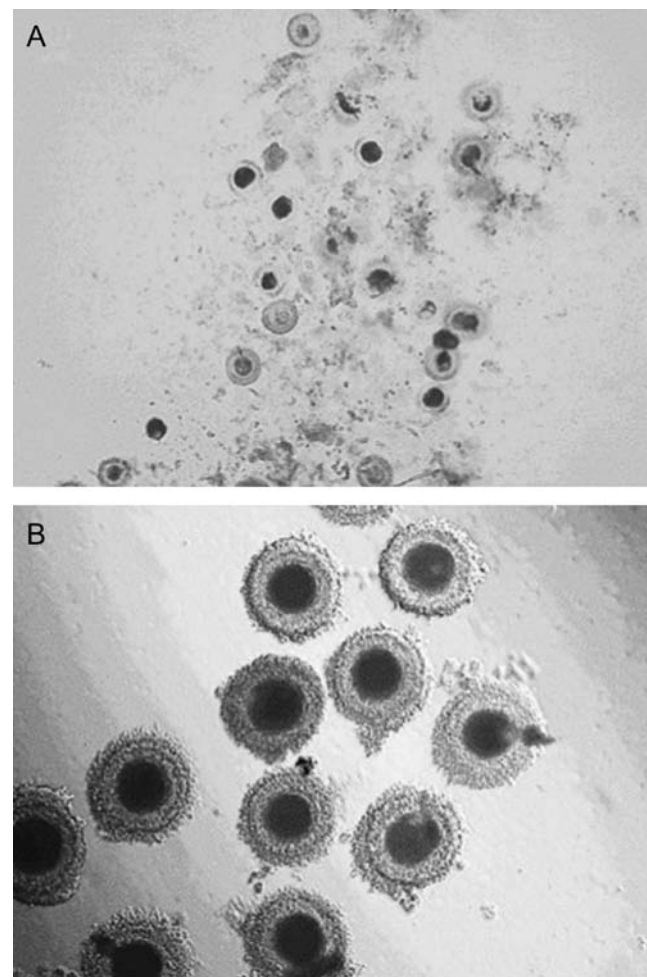


Figure 2 Morphology of oocytes. (A) Canine oocytes discarded after collection. (B) Canine oocytes selected for IVM.

a biphasic system: for 24 h, COCs were cultured with 5.0 µg/mL of LH (Lutropin, Vetoquinol) and for other 48 h with 10% oestrous bitch serum with progesterone level higher than 10 ng/mL.

In this second step, we tested four experimental conditions:

Group 1) co-culture with epithelial cells in monolayer

Group 2) co-culture with multicellular spheroids

Group 3) culture with 10% of CM secreted by multicellular spheroids

Group 4) culture with 10% of SN secreted by multicellular spheroids

Group 5) culture with different amounts (25–50–75–100–150 × 10⁶/mL) of MVs secreted by multicellular spheroids. This amount was calculated based on microliters of pellet obtained after centrifugation and on the total amount of MVs found after analysis by NanoSight.

As control group, SOF in biphasic system without CM, SN or MVs was used.

Assessment of meiotic maturation by Hoechst 33342

At the end of the culture period, irrespective of their morphology, oocytes were removed from the culture drop and washed in PBS for three times. Then, oocytes were denuded by incubation for 10 min at 38.5°C in TCM 199 with 20% FCS containing 80 IU/mL hyaluronidase and by aspiration in and out of finely drawn glass pipettes. All oocytes were stained for 15 min at RT in a small drop of PBS supplemented with 10 µg/mL bis-benzimide (Hoechst 33342) on a glass slide. Particular attention was paid to avoid sample exposure to light during staining in order to reduce photo bleaching. The nuclear configuration and chromatin morphology were evaluated under fluorescence microscopy (Olympus BX51) with UV light (353–365 nm excitation and 460 nm emission) to determine the stage of meiotic maturation. Chromatin configurations were classified as previously described (Kim *et al.* 2004). GV stage is the so-called meiotic block. Stages indicative of meiosis resumption were germinal vesicle break down (GVBD) (organization and condensation of chromatin into chromosomes), metaphase I (alignment of chromosomes on meiotic spindle) and metaphase II, which was indicated by the presence of the 2nd metaphase plate (MII) and the first polar body (PB) was extruded. Oocytes with irregular chromatin distribution or absence of chromatin were considered as indicative of degenerated or non-identifiable chromatin material (DEG).

Evaluation of MV uptake by oocytes

To confirm MVs incorporation into oocyte, a group of oocyte was cultured in SOF supplemented with MVs previously labeled with PKH-26, at a concentration of 100 × 10⁶ MVs/mL for 48 h of IVM. Oocytes were stained with Hoechst 33343 (10 µg/mL) and MVs incorporation was evaluated under fluorescence by an Olympus BX51 microscope. Labeled MVs were excited at 550 nm while emission wavelength was set at 567 nm. Hoechst 33342 dye was excited at 353–365 nm while the emission wavelength was set at 460 nm.

Statistical analysis

Percentages of oocytes reaching various stages of nuclear maturation under the different experimental conditions were statistically analyzed by chi-squared test in a large contingency table and by the Yates correction for continuity. In addition, Fisher's exact test was performed when appropriate (observations <5). A probability level of $P < 0.05$ was considered as significantly different. Data were analyzed with GraphPad InStat 3.00 for Windows (GraphPad Software).

Results

TEM

The semithin section indicated that multicellular spheroids are formed by clusters of cells tightly in contact. Several apical protrusions, as well as elongated cilia, were located on the outer cell side of the cluster. Non-ciliated cells showed a more intense staining than ciliated cells (Fig. 3).

TEM observation showed the presence of epithelial cells, ciliated and not ciliated. Ciliated cells were characterized by some endocytic vesicle in the apical region (Fig. 4A and B). Non-ciliated cells often displayed microvilli on the apical protrusion and were characterized by an electron-dense cytoplasm containing a lobate nucleus rich in dispersed heterochromatin, dark granules of secretion in their supranuclear cytoplasm and elongated mitochondria (Fig. 4B and C). The presence of cell junctions in the apical portion of these cells indicates the polarization of these cells (Fig. 4B) that are also closely connected to each other through membrane interdigitations (Fig. 4A, B and C). Cells with a large nucleus with disperse heterochromatin and voluminous vesicles containing electron-lucent secrete were detected under the epithelial cells (Fig. 4B and D).

Immunocytochemistry

Multicellular spheroids exhibited positive immunostaining for cytokeratin, normally found in cells of epithelial origin. Cells also exhibited immunostaining for vimentin, a cytoskeletal protein expressed in cells of mesenchymal origin (Fig. 5). This result is indicative of a mixed cell population of both epithelial and mesenchymal cells in these tri-dimensional structures.

Isolation and measurements of MVs

NanoSight analysis determined the presence of MVs with size ranging from 50 nm to 670 nm for the three samples, with an average of 234 ± 23 nm. MVs production corresponds to 540 × 10⁶ particles/mL of *medium*.

According to size, these MVs can be defined as shedding vesicles, i.e. vesicles budded directly from the plasma membrane of oviductal cells.

DOOF ONLY

Oocytes retrieval and culture

From 64 ovariectomized bitches, 17 were in anoestrus, 12 in oestrus and 35 in diestrus. A total of 1375 oocytes were obtained from all animals, with an average of 10.74 oocytes per ovary. Oocytes were selected according to their diameter and morphology and, based on these criteria, almost 50% of oocytes were eliminated and the selected oocytes were spread randomly among different culture *media*, not taking into account the oestrous cycle stage of the bitches.

In all experimental conditions, at the end of the IVM period, some oocytes showed expansion of cumulus oophorus (Fig. 8).

Based on nuclear morphology showed by Hoechst staining, oocytes were classified as GV (Fig. 9A), germinal vesicle breakdown (GVBD; Fig. 9B), metaphase I (MI; Fig. 9C) and metaphase II (MII; Fig. 9D). Furthermore, degenerated oocytes (DEG) also were counted (Fig. 9E).

The results of the experiments are given in Table 1, with the rates of different nuclear stages.

The maturation rate in the control *medium* was 8.66% and all data obtained in other conditions were compared to this result.

The co-culture system with epithelial cells in monolayer provided 10.71% of MII that is not statistically different ($P < 0.05$) compared to CTR, while co-culture with multicellular spheroids gave 20% of MII that was significantly higher ($P < 0.05$) than MII rate of CTR, monolayer of epithelial cells, CM (9.09%) and SN (7.14%).

The best conditions for obtaining MII oocytes were the co-culture with MVs at concentrations varying between 75 and 100×10^6 /mL. Maturation rates were respectively 20.34% and 21.82%, and they were statistically greater ($P < 0.05$) than control, monolayer of epithelial cells, CM and SN but similar to the rate of co-culture with multicellular spheroids. Higher concentrations of MVs (150×10^6) showed statistically ($P < 0.05$) lower maturation rates (9.26%) compared to the others MV concentrations.

The rate of oocytes that remained at the GV stage was the same in all the experimental conditions. The rate of oocytes with GVBD was statistically higher ($P < 0.05$) in CTR, in oocytes matured on monolayer and in the presence of CM and SN (45.24, 47.56, 38.64 and 44.05% respectively), but their MII rate was statistically lower ($P < 0.05$) than co-culture system with multicellular spheroids and with MVs concentration that provided the best maturation rates.

Evaluation of MV uptake by oocytes

MVs were abundantly incorporated by cumulus cells, but not in the oocyte cytoplasm, after 48 h of culture

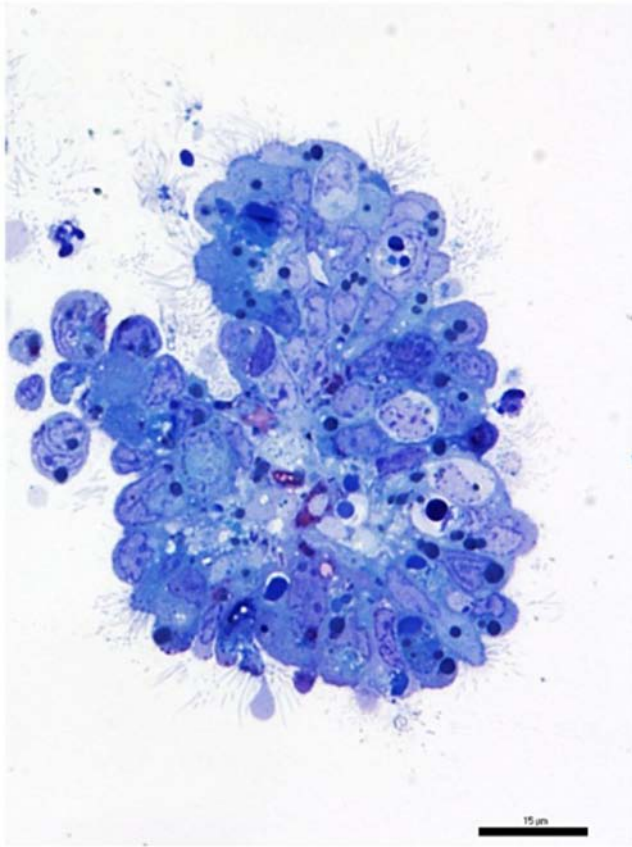


Figure 3 Semithin section. (A) Multicellular spheroids at magnification 63x. Scale bar 15 µm. (B, C, D, E and F) Multicellular spheroids at magnification 100x. Scale bar 10 µm. A cluster of cells tightly in contact formed these aggregates. Several apical protrusions, as well as elongated cilia, were located on the outer cell side of the cluster. Magnification 200x.

Q8



Figure 6 shows an example of NanoSight analysis on MVs released by oviductal multicellular spheroids.

miRNA analyses by RNA extraction and PCR amplification

Expression of specific miRNA was determined by PCR assay that showed the presence of miR-30b, miR-375 and miR-503 in both MVs and multicellular spheroids (Fig. 7).

Progesterone level of bitch serum

According to hormonal evaluation by MiniVidas system, progesterone levels of the three bitches on peri-ovulation oestrus stage were 8, 12 and 15 ng/mL, with an average of 11.67 ng/mL. The serum samples were used pooled.

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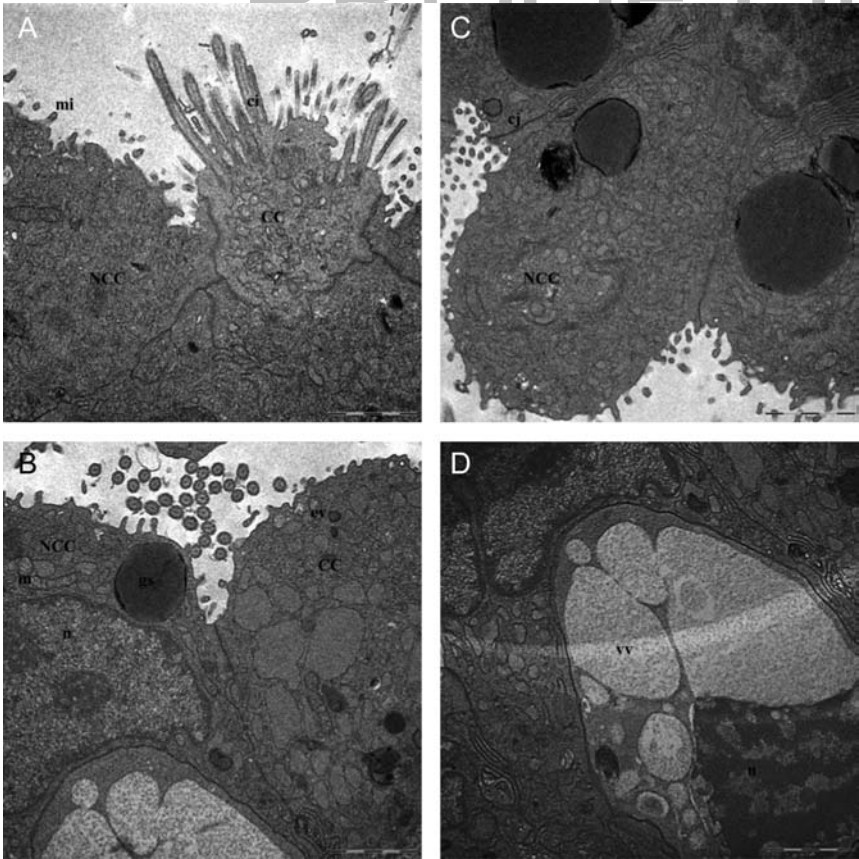


Figure 4 TEM observation. (A) Ciliated cells in longitudinal section. (B) Ciliated cells were characterized by a diffuse presence of rough endoplasmic reticulum throughout the cytoplasm and some endocytic vesicle in the apical region. (C) Non-ciliated cells often displayed microvilli on the apical protrusion and were characterized by an electron-dense cytoplasm containing a lobate nucleus rich in dispersed heterochromatin, dark granules of secretion in their supranuclear cytoplasm, elongated mitochondria and a rich endoplasmic reticulum. The presence of cell junctions in the apical portion of these cells indicates the polarization of these cells that are also closely connected to each other through interdigitations of membrane. (D) Cells with a large nucleus with disperse heterochromatin and voluminous vesicles containing electron-lucent secrete were detected under the epithelial cells (Fig. 3B and D). CC, ciliated cells; ci, cilia; cj, cell junction; ev, endocytic vesicle; gs, granule of secretion; m, mitochondria; mi, microvilli; n, nucleus; NCC, non-ciliated cells; vv, voluminous vesicle.

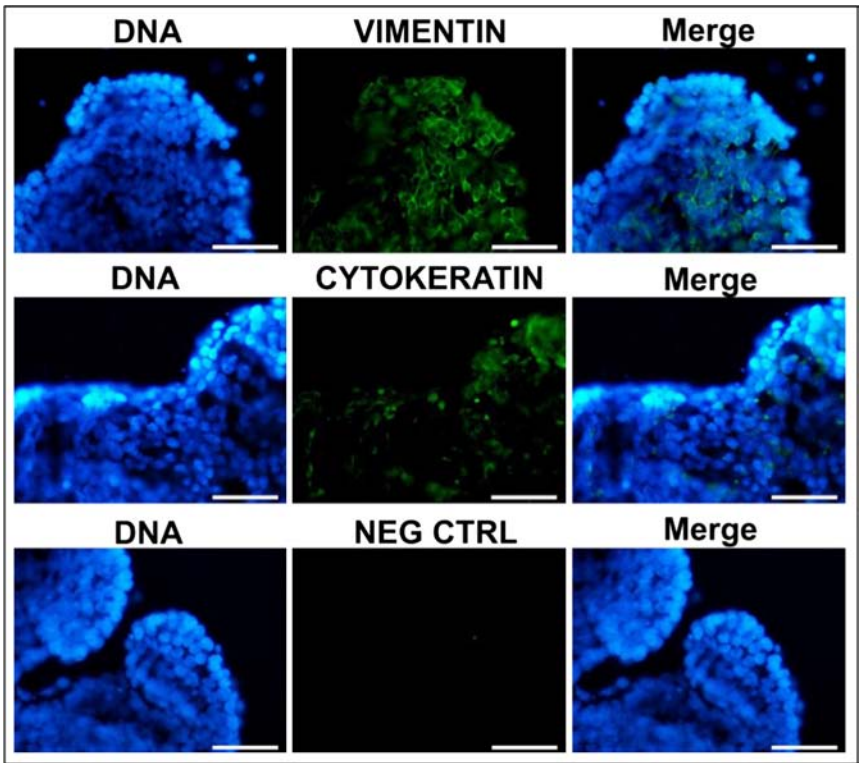


Figure 5 Immunocytochemistry. Multicellular spheroids exhibited positive immunostaining for cytokeratin and vimentin. Figures in the left column were obtained by staining with DAPI. Figures in center column show specific staining and control (CTRL). Figures in right column were merge images. Magnification 40x. Scale bar 50µm.

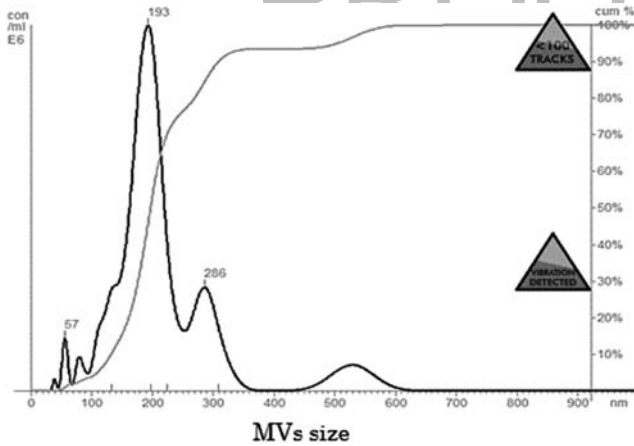


Figure 6 Measurement of MVs. NanoSight analysis relates MV size (in abscissa) to their concentration (in ordinate).

(Fig. 10A). The incorporation of MVs within the cytoplasm was visible 72 h after the start of IVM, as shown in the Fig. 10C and D.

Discussion

The study of the interactions of oviductal cells and their secretions with gametes and embryos at cellular and molecular level is fundamental in the comprehension of early reproductive mechanisms and in the development and refinement of reproductive biotechnologies (Gualtieri *et al.* 2012).

The objective of the present study was to understand the effect of secretome of epithelial oviductal cells on canine oocyte IVM. We focused our attention on epithelial cells of the isthmus–ampullar oviductal tract because *in vivo* this oviductal environment in the dog has some unique qualities that are beneficial for extending oocyte viability and supporting their maturation but at the moment is not given to know which factors are so important and where they are produced. In addition, for this purpose, IVM was performed taking into account all the most effective conditions reported in literature and previously described. In the biphasic system, COCs were cultured for 24 h with LH to simulate the LH peak, which occurs in the early estrus, and that leads to ovulation. *In vivo*, in conjunction with the LH surge, a concomitant gradual increase in serum concentrations of

progesterone becomes apparent, and for this reason, for other 48 h, the oestrous bitch serum with progesterone level higher than 10 ng/mL was added. In addition to these experimental conditions, in this second step, COCs were cocultured with oviductal epithelial cells that were cultivated in two different ways: in monolayer or in tri-dimensional multicellular spheroids in suspension. The results showed that co-culture with epithelial cells in monolayer provided statistically ($P < 0.05$) less maturation rate (MII) compared to co-culture in the presence of multicellular spheroids (10.71% vs 20.00%). Both IVM conditions were statistically ($P < 0.05$) better than control in SOF medium alone, without helper cells (8.66%). The difference between the two kinds of oviductal epithelial cells could be explained taking into account that, during culture, oviductal monolayers can de-differentiate losing cell polarity, cell height, ciliation, secretory activity and responsiveness to hormones, showing signs of alteration after 24–48 h post confluence (Rottmayer *et al.* 2006, Sostaric *et al.* 2008, Gualtieri *et al.* 2009, Ulbrich *et al.* 2010). For this reason, we tried to use the multicellular spheroids that spontaneously aggregate after the squeezing of canine oviduct. These multicellular spheroids are heterogeneous and mainly composed of epithelial, ciliated and non-ciliated cells. As demonstrated by TEM analysis, these cells maintain their polarity and their secretory activities evidenced by the occurrence of several secretory granules. The presence of several protrusions and short scattered microvilli suggests that they could mostly be derived from region of the ampulla (de Santis *et al.* 2011). In addition to these cells, multicellular spheroids are also composed of stromal cells, positive to vimentin.

Considering the better maturation rate, these multicellular spheroids seem to better mimic what normally occur *in vivo*, probably thanks to the cell-to-cell interactions between epithelial cells and between epithelial and mesenchymal cells inside these aggregates.

This suggests that the oviduct epithelium actively participates in the production of a unique biochemical milieu able to promote the *in vivo* oocyte maturation. Many growth factors, receptors and binding proteins are secreted into the culture medium by these so-called ‘helpers’ cells’ (Van Langendonck *et al.* 1996, Izquierdo *et al.* 1999) and the potential of these medium, called CM, is already known for a long time, mainly for *in vitro* embryo production. Ijaz *et al.* (1994) and

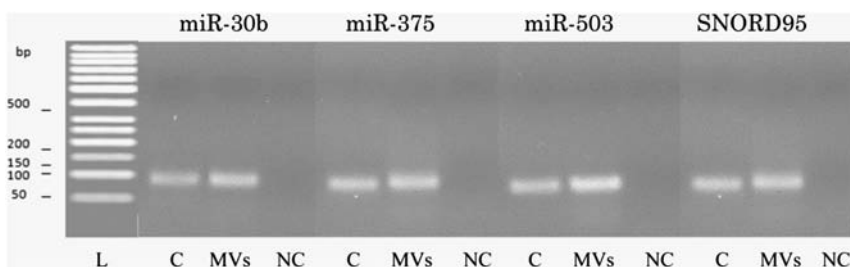


Figure 7 Identification of mi-RNAs. PCR analysis, identified three specific miRNAs (miR-30b, miR-375, miR-503), both in MVs and oviductal cells. C is cells, MVs is microvesicles and NC is negative control. SNORD95 was used as positive control.

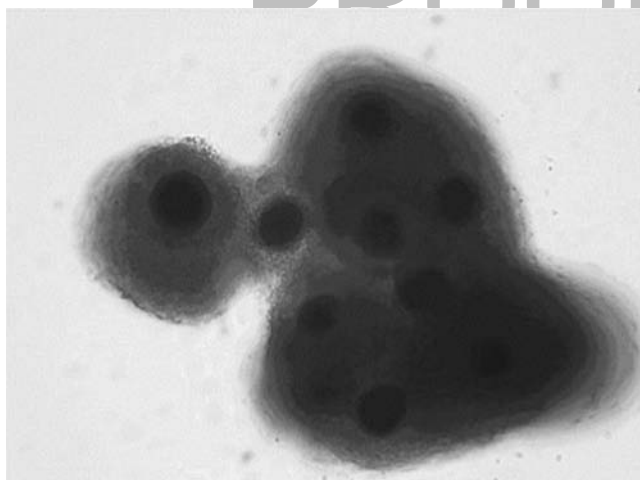


Figure 8 Morphology of matured oocytes. Cumulus expansion of oocytes cultured with 100×10^6 MVs/mL.

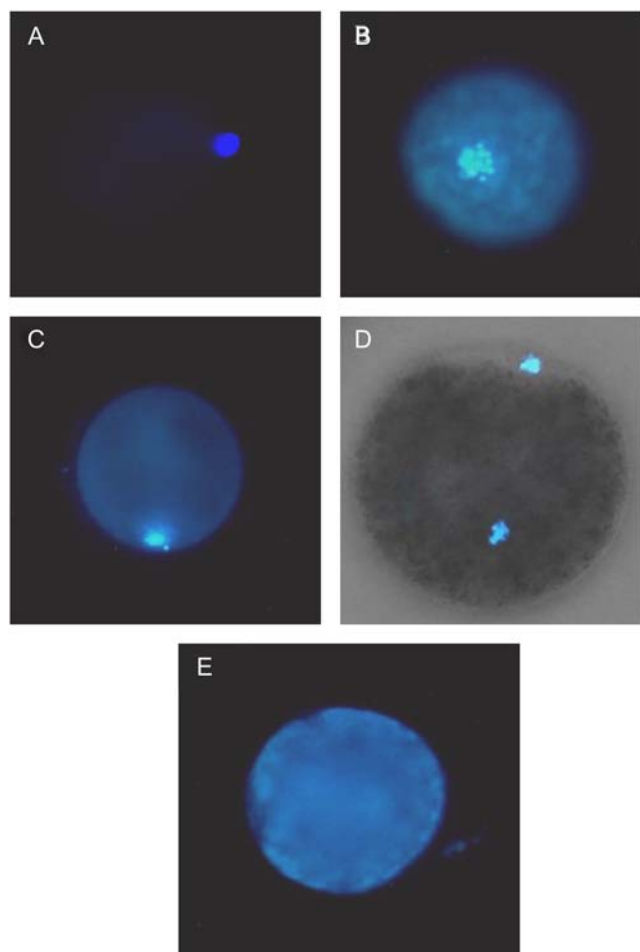


Figure 9 Nuclear stages of oocytes after 72 h of culture. Oocytes stained with Hoechst 33342 and evaluated under fluorescence microscopy, demonstrating (A) germinal vesicle (GV), (B) germinal vesicle break down (GVBD), (C) metaphase I (MI), (D) metaphase II (MII), (E) degenerated (DEG) oocytes. Magnification 20x.

Zhu *et al.* (1994) defined the CM as a complex matrix of secreted products from cells, in particular, growth factors, cytokines and glycoproteins that may influence the development of early preimplantation embryos. Some experiments have shown that the use of CM provides embryo developmental rates similar to those obtained in the co-culture systems (Kobayashi *et al.* 1992, Mermillod *et al.* 1993, Lee *et al.* 2003), with the benefit of not having animal cells in the embryo culture. CM was also studied on porcine gamete to reduce polyspermy rates and improve embryo development (Bureau *et al.* 2000).

To date, the effect of different components of oviductal CM on canine IVM has not been investigated. Since *in vivo* canine oocytes interacts with the oviduct for a prolonged time and, *in vitro*, multicellular spheroids provided the better maturation rate, this study investigated the effect of different components of multicellular spheroid secretome on the canine oocyte MII rate.

CM is composed of soluble factors plus MVs. Consequently, CM *in toto*, MVs and SN, collected after removal of MVs, were investigated to recognize which of these components, if not all, have had effects on oocyte maturation, in terms of MII rate.

At first, the presence of MVs in CM produced by multicellular spheroids was identified. By their size, it was possible to classify these MVs as shedding vesicles, also known as ectosomes or microparticles, that originate from direct budding and 'blebbing' of the plasma membrane. These MVs are different from exosomes that arise from the cell membrane endosomal compartment and are released into the extracellular space after fusion of multivesicular bodies with the plasma membrane. Exosomes tend to be homogeneous in size (30–120 nm) while shedding vesicles are more heterogeneous (ranging from 100 nm to 1 μ m). The multicellular spheroid-derived MVs had an average size of 234 ± 23 nm and in each ml of CM, 54×10^6 MVs were present. Obviously, these data refer to *in vitro* study and no information about their production during *in vivo* conditions is available.

The different components of secretome were added in the second step of the biphasic IVM system matching the co-culture with epithelial cells in monolayer or by multicellular spheroids. The results of our study suggested that the exposure of canine COCs to the various fractions of secretome provided dissimilar rates of MII. Indeed, CM and SN gave respectively 9.09% and 7.14% of MII that were not statistically different ($P < 0.05$) from the rate provided by epithelial cells in monolayer (10.71%). The first insight about these results indicated that, probably, CM and SN were used too much diluted (10% of SOF composition) to provide an adequate amount of soluble and non-soluble factors, but the maturation in whole CM or SN, or in lower rate, did not provide MII attainment (data not shown).

Table 1 Effects of different experimental conditions on IVM of canine oocytes.

Media	Assessed oocytes (n ^o)	Meiotic stages				
		GV (%)	GVBD (%)	MI (%)	MII (%)	DEG (%)
IVM control SOF+LH/SOF+serum	82	17 (20.73) ^a	39 (47.56) ^a	5 (6.1)	3 (8.66) ^a	18 (21.95)
SOF+LH/SOF+serum+monolayer of epithelial cells	84	18 (21.43) ^a	38 (45.24) ^a	6 (7.14)	9 (10.71) ^a	13 (15.48)
SOF+LH/SOF+serum+multicellular spheroids	79	10 (12.66) ^a	23 (29.11) ^b	10 (12.66)	16 (20.00) ^b	20 (25.32)
SOF+LH/SOF+serum+10% CM	88	15 (17.05) ^a	34 (38.64) ^a	9 (10.23)	8 (9.09) ^a	22 (25.00)
SOF+LH/SOF+serum+10% SN	84	22 (26.19) ^a	37 (44.05) ^a	6 (7.14)	6 (7.14) ^a	13 (15.48)
SOF+LH/SOF+serum+25×10 ⁶ MVs	54	15 (27.78) ^a	14 (25.93) ^b	10 (18.52)	3 (5.56) ^a	12 (22.22)
SOF+LH/SOF+serum+50×10 ⁶ MVs	58	9 (15.52) ^a	10 (17.24) ^b	11 (18.97)	5 (8.62) ^a	23 (39.65)
SOF+LH/SOF+serum+75×10 ⁶ MVs	59	7 (11.86) ^a	17 (28.81) ^b	7 (11.86)	12 (20.34) ^b	16 (27.12)
SOF+LH/SOF+serum+100×10 ⁶ MVs	55	6 (10.91) ^a	16 (29.10) ^b	9 (16.36)	12 (21.82) ^b	12 (21.82)
SOF+LH/SOF+serum+150×10 ⁶ MVs	54	6 (11.11) ^a	12 (22.22) ^b	10 (18.52)	5 (9.26) ^a	21 (38.89)

Different superscripts in columns indicate $P < 0.05$.

The CM is rich of soluble and non-soluble signal but can be considered a spent medium because, during cell metabolism, some molecules (for example the level of glucose) decrease (Pereira *et al.* 2014), and this could influence the oocyte metabolism negatively.

MVs were tested at different amounts, and taking into account that one milliliter of CM contained 54×10^6 particles/mL, the experiment was performed using between 25 and 150×10^6 MVs per mL of SOF. The better results were obtained at the concentration of 75 and 100×10^6 MVs/mL (20.34% and 21.82% of MII respectively) that were not statistically different ($P < 0.05$) from the rate provided by co-culture with multicellular spheroids (20%). The higher concentration of MVs (150×10^6 MVs/mL) gave statistically ($P < 0.05$) lower rate of MII (9.26) compared to the other two amounts.

To understand the good efficacy of MVs, we performed other two studies in order to investigate if oocytes were able to uptake these MVs and to check the presence, either in epithelial oviductal cells or in MVs, of some miRNAs, which emerge from the literature to be important for oocyte maturation. Certainly, cumulus cells are involved in paracrine mechanism as a bridge between oviduct and oocytes considering that they incorporated a significant amount of labeled MVs before the oocyte uptake. Moreover, recently, it was

identified that the miRNA-378 present in the cumulus cells is involved in the maturation of porcine oocyte. Its overexpression alters the cumulus expansion and decreases the expression of genes associated either to the cumulus expansion or to the oocyte maturation (Pan *et al.* 2015). About the involvement of miRNA in oocyte maturation, Tesfaye *et al.* (2009) showed that between the immature and mature bovine oocyte, a difference of 59 miRNAs exists, of which, 31 are preferably expressed in the immature oocyte and 28 in the mature ones. Our study found that oviductal cells and their MVs contain miR-30b, miR-375 and miR-503 that can be considered particularly interesting for the canine IVM oocytes because it is involved in follicular growth and oocyte maturation. Pathway analysis revealed that these miRNAs target factors such as WNT, MAPK, ERβ, TGFβ, neurotrophin and ubiquitin (Silveira *et al.* 2012, Sohel *et al.* 2013, Santonocito *et al.* 2014). Review of Machtinger *et al.* (2016) reports that WNT molecules are glycoproteins involved in follicular growth, luteogenesis and steroidogenesis. The members of the TGFβ superfamily, including inhibin, activin, bone morphogenetic protein 15 (BMP15) and growth differentiation factor 9 (GDF9), are expressed in the oocyte from very early stages and are responsible for follicular growth and oocyte

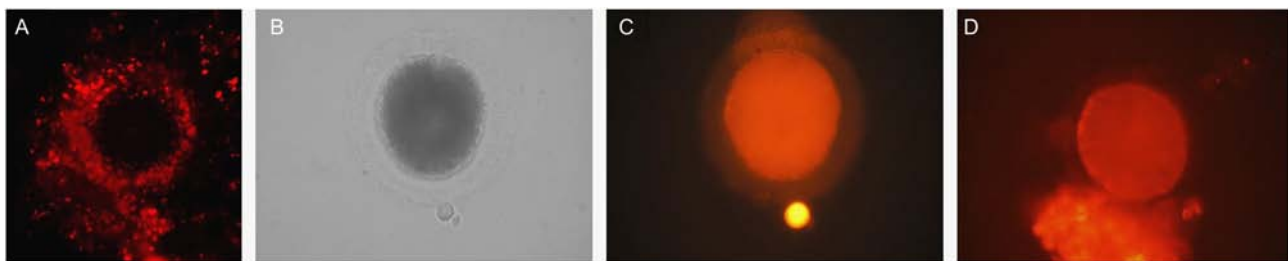


Figure 10 Incorporation of MVs by oocytes. (A) MVs, labeled with PKH-26, are present within the cumulus cells, since 48 h of culture but not in oocyte cytoplasm. (B) Oocyte observed under transmitted visible light evidencing one cell of cumulus oophorus. (C) The same oocyte observed under fluorescent microscopy. The considerable incorporation of MVs within the cumulus cell and into the cytoplasm of oocyte at MI stage after 72 of IVM become clearly visible. (D) Oocyte with MVs incorporated in its cytoplasm and its partial cumulus oophorus.

maturation. The MAPK pathway stimulates granulosa cell proliferation and cumulus expansion. Furthermore, together with the ER β pathway, the MAPK pathway promotes the resumption of meiosis in the oocyte. The ubiquitin-mediated pathway modulates oocyte meiotic maturation and early mitotic division in embryos, while the neurotrophin-signaling pathway regulates oogenesis and follicle formation. Considering that canine oocyte maturation occurs in the oviduct, it is conceivable that these miRNAs are present in the oviductal cells to be transferred to the oocytes for the purpose of their maturation. This is different compared to other species, for example in women, where these miRNAs were found in follicular fluid (place designed for oocyte maturation) and accumulate in granulosa cells and/or the cumulus cells (Sohel *et al.* 2013).

Through identification of these three miRNAs, perhaps it would be possible to explain why higher MVs doses lowered the MII rate to 9.26%. Two studies reported high levels of miR-375 in exosomes from older vs younger mares. This miRNA targets TGF β , suppressing the TGF β pathway leading to impaired oocyte maturation in older females (Knight & Glister 2006, da Silveira *et al.* 2012). An excessive dose of MVs, and in our case of the miR-375 would, therefore, have a deleterious, rather than beneficial, effect on oocyte maturation.

Conclusions

The results of this study show that even in the absence of cell-cell contact, the secretome produced by oviductal cells can exert beneficial actions on target cells, in this case represented by oocytes. Apparently, not the soluble components of the CM but the content of the microvesicles, with specific miRNAs, can contribute to the canine oocyte maturation.

Obviously, more studies will be needed to better understand the total cargo of miRNA contained in oviductal cells and their MVs and to understand if other miRNAs, involved in oocyte maturation, are preferably compartmentalized in these MVs. In this case, the addition of MVs or, preferably, the addition of specific miRNAs, or other biomolecules that may be involved in maintaining oocyte vitality, could potentially improve the rate of *in vitro* maturation, giving an improvement to the reproductive *in vitro* technologies in the canine species.

Moreover, this study provides a useful starting point for further studies related to the paracrine mechanisms of communication between oviduct and oocytes.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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