



Mechanical Properties of Growing Melanocytic Nevi and the Progression to Melanoma

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Abstract

Melanocytic nevi are benign proliferations that sometimes turn into malignant melanoma in a way that is still unclear from the biochemical and genetic point of view. Diagnostic and prognostic tools are then mostly based on dermoscopic examination and morphological analysis of histological tissues. To investigate the role of mechanics and geometry in the morphological dynamics of melanocytic nevi, we study a computation model for cell proliferation in a layered non-linear elastic tissue. Numerical simulations suggest that the morphology of the nevus is correlated to the initial location of the proliferating cell starting the growth process and to the mechanical properties of the tissue. Our results also support that melanocytes are subject to compressive stresses that fluctuate widely in the nevus and depend on the growth stage. Numerical simulations of cells in the epidermis releasing matrix metalloproteinases display an accelerated invasion of the dermis by destroying the basal membrane. Moreover, we suggest experimentally that osmotic stress and collagen inhibit growth in primary melanoma cells while the effect is much weaker in metastatic cells. Knowing that morphological features of nevi might also reflect geometry and mechanics rather than malignancy could be relevant for diagnostic purposes.

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Introduction

Melanocytic nevi are benign proliferations of melanocytes, the skin cells that produce the pigment melanin. They are by definition benign, but 50% of malignant melanomas arise from pre-existing nevi. Current diagnostic methods of melanocytic lesions are based on histopathology and dermoscopy which reveal wide morphological diversity and evolution patterns whose origin is still unknown [1,2]. Leaving aside the controversial cases of Spitz and blue nevi, nevi are categorized from dermoscopic analysis into globular, reticular, structureless brown and mixed patterns (the latter are subdivided into mixed pattern with globular structureless areas in the center or mixed pattern with globules at the periphery) [3]. The classical theory of nevi evolution formulated in 1893 by Unna [4] claims that nevi originate from melanocytes proliferating at the dermo-epidermal junction (lentigo simplex and junctional nevus) that form nests (compound nevus) and eventually migrate completely into the dermis (dermal nevus). More recently, Cramer described an opposite model known as the theory of upward migration [5]. Cramer suggested that precursor cells of melanocytes deriving from pluripotent stem cells of the neural crest wander during embryogenesis along nerves into the dermis, mature here and finally migrate as functional melanocytes into the epidermis. Cramer's last migration step, however, was never broadly accepted. More recently Kittler et al. [6] suggest that nevi can migrate not only vertically but also horizontally, explaining

how nevi can expand in the course of time. The underlying limitation of these theories is that they are based on histopathological observation only, and do not reflect the dynamics of an individual nevus. From a diagnostic point of view it would be extremely useful to correlate the morphological features of nevi to the degree of malignancy or pre-malignancy. This issue is intricate because nevi proliferate in a complex microenvironment that can mediate cell behavior through the composition, structure, and dimensionality of the extracellular matrix (ECM), the polymeric scaffold that surrounds cells within tissues. For example, a malignant phenotype can be reverted into a nonmalignant one by specifically blocking aberrant adhesion of the cancer cell to its extracellular scaffold [7].

Recent research shows that mechanical properties of the tumor microenvironment and of the normal tissue can influence tumor growth and dynamics, in way that is still poorly understood [8–13]. Mechanical stresses such as those experienced by cancer cells during the expansion of the tumor against the stromal tissue have been shown to release and activate growth factors involved in the progression of cancer [14]. Moreover, the stiffness of the matrix surrounding a tumor determines how cancer cells polarize, adhere, contract, and migrate, and thus regulates their invasiveness [15]. Forces exerted by cancer cells as they migrate through the ECM has been quantified accurately using traction force microscopy [16,17]. Another possibility is that mechanical stresses directly

regulate the growth and death rates of cancer cells as shown by Montel et al. who induce osmotic stress by adding dextran, a biocompatible polymer that is not metabolized by cells [11,12]. Several studies in the literature indicate important changes in cellular functioning due to osmotic pressure [18,19], but the stresses involved (in the MPa range) were orders of magnitude larger than those (in the kPa range) studied in Refs. [11,12]. It is interesting to notice that compressive stresses of slightly less than 1 kPa applied through a piston were recently found to induce a metastatic phenotype in cancer cells [13]. While osmotic pressure may have a different origin than compressive mechanical pressure the effect on cells is exactly the same. This was demonstrated experimentally in Refs. [11,12] by applying osmotic pressure directly on the cells, by adding dextran to the growth medium, or indirectly, by placing the cells inside a dialysis bag which was then placed into a dextran solution. In both cases, the effect of pressure on the cells was found to be exactly the same. This is not surprising, since osmotic pressure corresponds indeed to a *real* mechanical pressure on the membrane due to the collisions with solute molecules.

In a recent paper, Simonsen and coauthors showed that interstitial fluid pressure (IFP) was associated with high geometric resistance to blood flow caused by tumor-line specific vascular abnormalities in xenografted tumors from two human melanoma lines with different angiogenic profiles [20]. In another recent paper, Wu and colleagues investigated how nonlinear interactions among the vascular and lymphatic networks and proliferating tumor cells might influence IFP transport of oxygen, and tumor progression [21]. They also investigated the possible consequences of tumor-associated pathologies such as elevated vascular hydraulic conductivities and decreased osmotic pressure differences. All these parameters might affect microenvironmental transport barriers, and the tumor invasive and metastatic potential, opening interesting new therapeutic approaches. In general, understanding the influence of mechanical stress on cancer growth could shed new light on tumor development and progression.

To investigate the role of the environment in the growth of melanocytic nevi, we introduce a computational model of elastic cells proliferating in a layered non-linear elastic medium representing the skin. Several computational models have been introduced in the past to simulate cancer growth in vitro and in vivo. They range from individual cell models [22–26], to lattice models [27–29] and continuum models [29–31]. The aim of our investigation is to correlate the morphological features of the growing nevus with the location of the originating melanocyte and the mechanical properties of the environment. To this end, a natural strategy is provided by individual cell models where each cell can be mechanically deformed and moves according to mechanical forces due to other cells and to the ECM. Models of this kind have been used in the past to simulate the growth of the epidermis [32,33] and the growth of cancer cell colonies inside a confining medium [26].

We first report experimental results on melanoma cell lines showing that a low osmotic pressure reduces proliferation, in agreement with other reports for other tumors [11,12]. We have performed experiments on two cell lines, the first obtained from a primary tumor and the second from a metastatic one, both from the same patient. We find that melanoma cells stemming from a primary tumor are less sensitive to pressure than the metastatic ones. We find a similar results by growing the same cells in collagen coated plates, simulating the effects of the ECM. Melanoma is widely believed to be originating from melanocytes, which are not easily cultivated in vitro. Therefore melanoma cell lines is the most sensible in vitro model for nevi. We expect that

the effect of pressure on proliferation seen in melanoma cells will be even stronger for melanocytes since melanoma cells are much more resistant.

Numerical simulations of the model yield different growth patterns for nevi originating in the epidermis, either close to the basal membrane or to the stratum corneum, or in the dermis. Different initial locations give also rise to different mechanical properties as we show by measuring compressive stresses on the proliferating melanocytes. Compressive stresses are fluctuating strongly inside the nevus, with an average stress that typically increases until the ECM breaks leading to stress drop. A crucial role in the growth pattern is played by the basal membrane, separating the epidermis from the dermis. Depending on its mechanical properties, the membrane can resist the pressure induced by the nevus or fail leading to the invasion of the dermis. In this case, invasion is not correlated with an increased malignancy of the tumor but just to mechanics. This observation of course does not exclude that malignant phenotypes may be induced by mechanical stresses as previously reported in the literature [13]. It is in fact known that malignant cells break the basal membrane by matrix metalloproteinases (MMPs). We simulate this process explicitly showing that it accelerates the invasion of the dermis, but the general morphology remains the same.

Model

We simulate an individual cell based model [24–26] for the growth of melanocytic nevi in the skin. We consider a two dimensional vertical section of the skin (see Figure 1a) consisting of two main layers, the *epidermis* and the *dermis*, separated by the *basal membrane*. The deepest layer of the skin, the hypodermis, is not considered in the present study. In our model, the dermis constitutes the bottom part of the skin, whereas the upper layer is represented by the epidermis and by its most superficial part, the *stratum corneum*. The model describes the skin as a highly organized elastic organ, whose mechanical properties are specific of any of its constituent layers. Periodic boundary conditions are applied to any of the skin layers along the horizontal direction, while open boundary conditions are imposed in the vertical directions.

Dermis

The dermis is arbitrarily divided into two anatomical regions: the papillary and reticular dermis. We consider only the papillary layer, whose thickness we set to $150\mu\text{m}$. We model the dermis as a network of cells with an average radius $\sim 15\mu\text{m}$, arranged in a disordered lattice and connected by non-linear springs. The disordered lattice is obtained starting from a triangular lattice, displacing the nodes randomly by a small amount. The resulting configuration is triangulated by a Voronoi algorithm. Nearest neighbors cells are connected by non-linear springs with a rest length equal to the resulting distance between the displaced cells. Cells here represents fibroblasts, macrophages, and adynocytes present inside the papillary layer, while springs model the effect of the ECM, composed by collagen fibers and elastin. We set the average length of these springs at $\sim 60\mu\text{m}$. The dermis also contains several irregularities such as blood vessels, lymph vessels, nerve endings and skin appendages such as air follicles, small hair muscles and sebaceous glands, but we ignore these details in the present model. The mechanical behaviour of the dermis has been studied experimentally using in-vivo suction and tension tests as reported in [34–36]. In these studies, the mechanical response of the dermis layer to an external stress was found to obey to the following non-linear *James-Green-Simpson* pressure-displacement

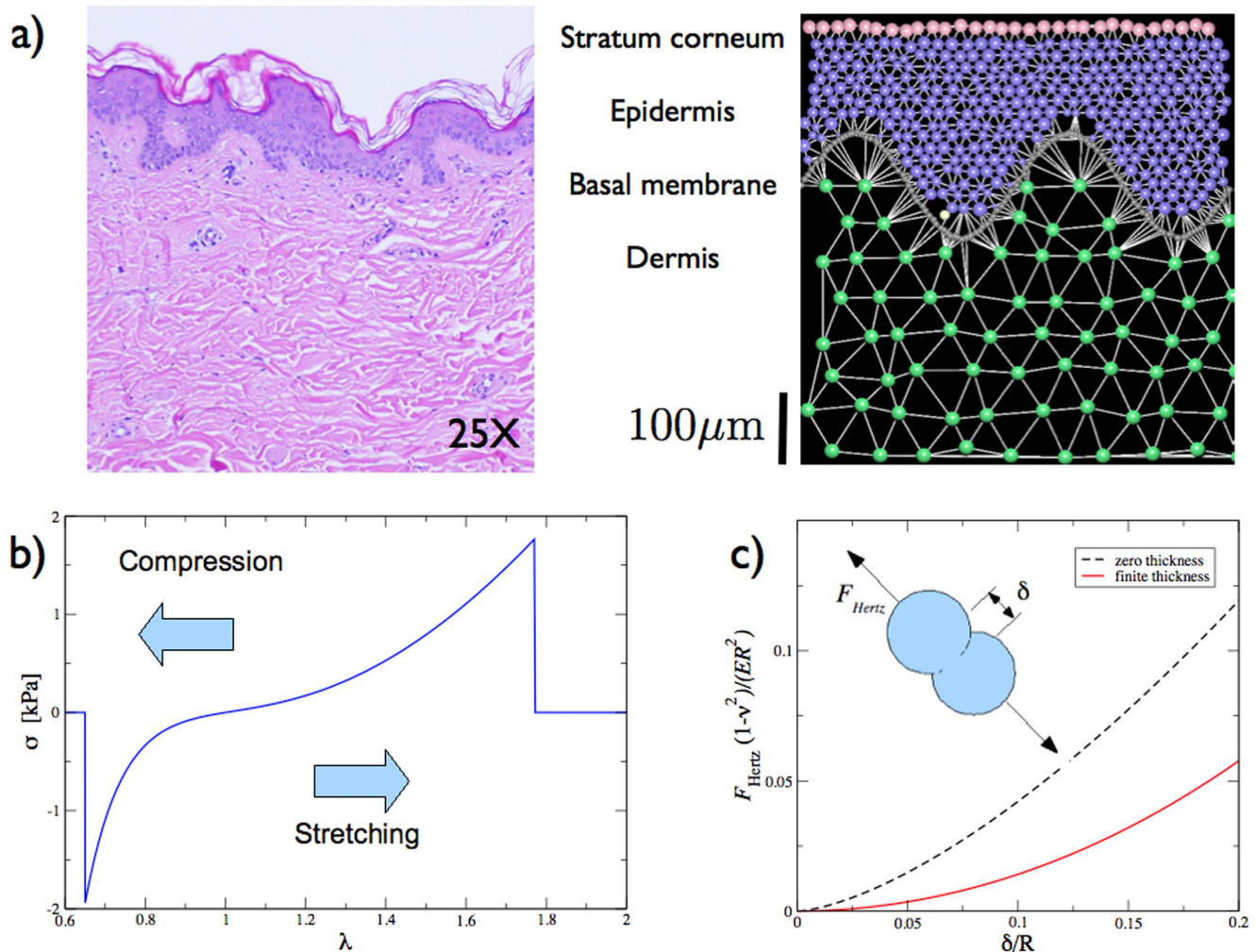


Figure 1. Computational model. a) A two dimensional section of the skin (left image courtesy of Dr. Claudio Clemente) is simulated as a non-linear elastic layered material (right). The top layer is the stratum corneum (pink cells), resting on the epidermis (blue cell). The epidermis is separated from the dermis (green cells) by a basal membrane (grey). In a typical simulation we consider a melanocyte (yellow) duplicating inside the skin, either in the epidermis, or in the dermis. b) The connecting fiber have a non-linear elastic behavior with a fracture strength set at $\sigma_Y = 1.8$ kPa. c) When in contact, the cells interact by a finite-thickness Hertz law. The corresponding zero-thickness law is reported for comparison.
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relationship [37]

$$\sigma = 2C_{10} \left(\lambda - \frac{1}{\lambda^2} \right) + 6C_{11} \left(\lambda^2 - \lambda - 1 + \frac{1}{\lambda^2} + \frac{1}{\lambda^3} - \frac{1}{\lambda^4} \right) \quad (1)$$

where σ and λ represent respectively the stress and the stretch of the tissue, the strain being just $\epsilon = \lambda - 1$. The expression (1) is derived from the strain energy density function usually adopted to mimic the non-linear behaviour of the skin [38]. Here we use the same law to model the deformation of the ECM springs, setting their elastic constant to $C_{10} = 0.1125$ kPa and $C_{11} = 0.315$ kPa, and introduce a failure stress at $\sigma_Y = 1.8$ kPa, at which the spring breaks. The non-linear elastic behavior of the springs is illustrated in Fig 1b, showing that when the stress overcomes σ_Y the spring breaks and the stress goes to zero.

Epidermis

The epidermis is a multilayered tissue composed of the stratum granulosum, stratum spinosum and stratum corneum. Since the elastic properties of the stratum corneum are considerably different

from the other two, we will consider it separately. The thickness of the epidermis (stratum granulosum and stratum spinosum) is $150 \mu\text{m}$ and it is composed mainly by keratinocytes, whose average radius is $\sim 20 \mu\text{m}$. In the model, keratinocytes are placed at the nodes of a disordered lattice, obtained as discussed above for the dermis. The turnover rate of melanocytes is assumed to be much faster than the natural turnover rate of keratinocytes, modelled explicitly in Ref. [32] but ignored here. This is justified since the typical turnover for melanoma cells is 1–2 days [42] which is much smaller than the turnover of keratinocytes (50–70 days). To simulate the packing of keratinocytes in the epidermis, we chose the radius of each cell after the Voronoi triangulation so that no neighboring cell overlaps. Cells are then coupled by non-linear springs accounting for the neo-Hookean behavior observed in *in vivo* experiments [34–36]. Experiments suggest that the non-linear pressure-displacement relation in Eq. (1) captures the mechanical properties of the epidermal layer with a good accuracy using the same parameters values obtained for the dermis [34,35]. In the model, we link the cells by non-linear springs with $C_{10} = 0.1125$ kPa, $C_{11} = 0.315$ kPa and $\sigma_Y = 1.8$ kPa. The average

length of the links between cells coincides with the average cell radius, hence two keratinocytes may often be in contact and will thus interact as we discuss below.

Stratum Corneum

The stratum corneum is composed of dead corneocytes of an average radius of $\sim 20\mu\text{m}$. Although it may consist of up to 15–20 layers of corneocytes, in our simplified numerical model only one single layer is taken into consideration. Experiments [34–36] have provided evidence that the stratum corneum mechanical properties are consistent with a linear Hookean law. Here, we consider elastic cells linked by linear springs with Young's modulus $E = 1.5\text{kPa}$ and failure stress set to $\sigma_Y = 2\text{kPa}$. The stratum corneum cells are linked to the epidermis by linear springs whose Young's modulus is $E = 1.6125\text{kPa}$.

Basal membrane

The basal membrane constitutes an elastic sheet separating the dermis from the epidermis. It has a complex structure composed of the lamina reticularis and lamina basale connected by collagen fibrils; usually the thickness is around $0.5 - 1\mu\text{m}$. In our model the basal membrane is represented by an assembly of fictitious cells of radius $0.5\mu\text{m}$, connected by linear springs with Young's modulus which we set either 0.15kPa or 0.3kPa . The failure stress of the springs is set to $\sigma_Y = 2\text{kPa}$. The fictitious cells composing the basal membrane have Hertzian repulsive interactions with the other cells thus providing an effective barrier to their motion. To represent the highly corrugated structure of the membrane, we use a simple periodic function with amplitude equal to $7.5\mu\text{m}$ and period $30\mu\text{m}$. Dermis and epidermis cells are connected to the basal membrane from below and above respectively, these springs obey to a linear Hookean force with Young's modulus equal to $E = 0.18725\text{kPa}$. In the model, the basal membrane can be broken mechanically, but we also consider the production of MMPs. To model this effect, we stipulate that at each time step each cell composing the nevus can dissolve one link with a probability that decreases with the distance from the cell:

$$P_{MMP} = p_{break} \left(\frac{r_{break}}{r} \right)^2, \quad (2)$$

with $p_{break} = 10^{-2}$ and $r_{break} = 30\mu\text{m}$.

Cell mechanics

Cells are modelled as non-linear elastic spheres and, for the sake of simplicity, we employ the same mechanical constants for all the cells composing the epidermis, the dermis and the nevus. When two cells are in contact they repel by a finite-thickness Hertzian law [39]

$$F_{Hertz} = \frac{4}{3} \frac{E}{(1-\nu^2)} \sqrt{R} \delta^{3/2} \frac{(1 + 1.15\omega^{1/3} + 9.5\omega + 9.288\omega^2)}{1 + 2.3\omega}, \quad (3)$$

where E is the cell's Young's modulus, set to $E \simeq 1\text{kPa}$ [40], ν is the Poisson's ratio, $R \equiv \frac{R_1 R_2}{R_1 + R_2}$ with R_1 and R_2 radii of the contact cells, δ is the indentation depth and $\omega = 0.1 \frac{\delta}{R}$ (see Fig. 1c).

We notice that cells are often modelled as incompressible ($\nu = 0.5$) [40], but the value of Poisson ratio is difficult to estimate experimentally. Individual cell models use compressible cells as we do and employ $\nu < 0.5$ [26,41]. Here we use $\nu = 0.33$ but its precise value is not really important in the framework of our simulations:

changing ν would just imply a modification of the pre-factor of Eq. 3, leaving qualitatively unaffected the numerical results hereby displayed. We notice that, although Eq.(3) refers to the experimental situation of a microsphere indenting a finite-sized soft gel, it captures the non-linear corrections to the usual Hertzian law, needed when large deformations occur. In most simulations, nevi cells are interacting by Hertzian repulsion only, while the other cells are typically linked also by non-linear spring. We have also tested for the effect of adhesion between nevi cells introducing a radial force $f_{adhesion} = k_{ad}\delta$, with $k_{ad} = 1.5 \times 10^{-5}\text{N/m}$. Adhesive forces only act when the indentation depth δ is larger than $-20\mu\text{m}$.

Interactions with the ECM

In the simpler implementation of the model, nevi cells have no direct interaction with the ECM. An indirect interaction exists since nevi cells interact with other cells which are held together by the ECM. This indirect interaction is, however, weak inside the dermis. To overcome this limitation, we also consider a direct interaction between nevi cells and the ECM. In practice, we follow the same strategy used to model the basal membrane and place three fictitious cells with radius $5\mu\text{m}$ on each ECM link. In this way, nevi cells interact directly with the ECM.

Growth dynamics

The quasi-static growth of the nevus is simulated by randomly selecting a single cell among those belonging to the dermis or epidermis. This cell represents the first proliferating cell and therefore does not present any linkage with the surrounding skin network. The growth process is obtained by duplication of the initial cell, so that the new cell is still in contact with the first one, but its growth direction is picked at random, i.e. lattice-free: such a choice is motivated by the fact a proliferative population will always eventually fill the lattice, suppressing cell-to-cell crowding effects [29]. The insertion of a new cell encompasses an overall rearrangement of the entire network, which is achieved by minimization of the stress within the system. Since growing cells are completely disconnected from the surrounding skin tissue, they only interact through Hertzian-like forces (Eq. 3) and possibly through adhesion. Two different choices of protocol are applied to the duplication mechanism: a cell can either be picked randomly among those composing the nevus, or the cell can be selected with a rate that depends on the compressive stress. In particular, the probability $P_i(N) \propto e^{-\frac{\sigma_i(N)}{\langle\sigma(N)\rangle}}$ is assigned to each cell, where $\sigma_i(N)$ represents compressive stress of the i -th cell and $\langle\sigma(N)\rangle$ is the average stress. Finally, the cell that duplicates is selected with probability $P_i(N)$.

Results

Experiments

Osmotic pressure affects the growth of primary melanoma cells. We test the effect of low osmotic pressure (from 0.1 kPa to 1 kPa) on cellular proliferation of primary and metastatic human melanoma cell lines, IgR39 and IgR37, respectively at short (3 days) and long time (6 days). As shown in Fig. 2, the treatment with 1 kPa for 6 days decreases the cellular proliferation of IgR39 while IgR37 are only slightly affected. According to Table 1, the osmotic pressure increases cell death to necrosis while the percentage of cells in early apoptosis slightly decreases at low pressures and then returns to the original value at higher pressure (Table 1). We are tempted to attribute this last result to a statistical fluctuation.

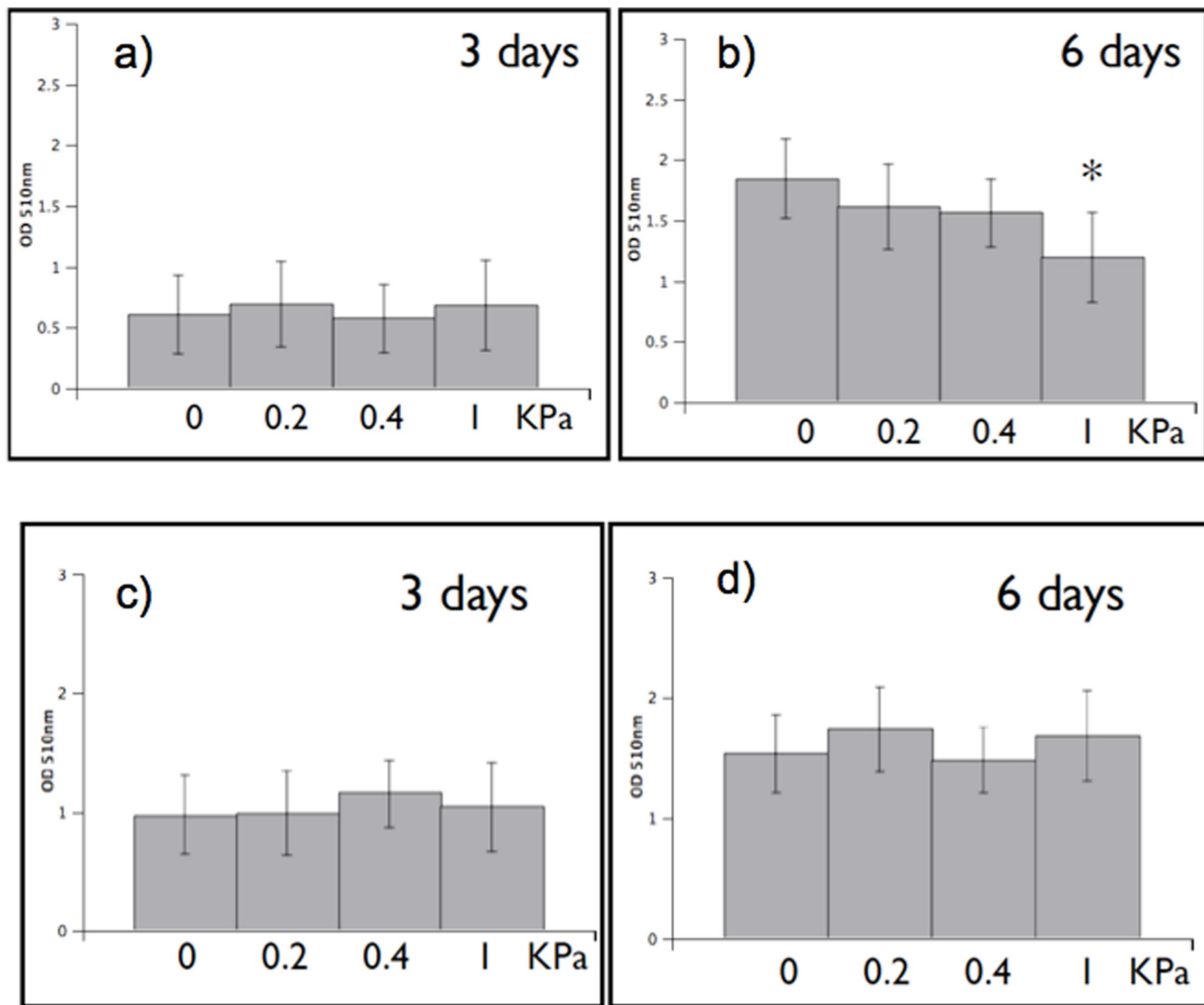


Figure 2. Effect of osmotic pressure on cell proliferation in melanoma. 500 cells/well were plated on 96 multiwells (IgR39 and IgR37). The day after plating the cells were submitted to different osmotic pressure (from 0.2 to 1 kPa) for 3 or 6 days. At the end of the incubation the cells were fixed with 50%TCA for 2 hours at 4°C and air-dry at room temperature. Thus, the cells were incubated with 0.05% SBR solution for 30 minutes at room temperature and then quickly rinsed four times with 1% acidic acid to remove unbound dye. Finally, the protein-bound dye was solubilized with 10 mM TRIS and OD was measured at 510 nm with microplate reader 550 (Bio-Rad). a) The growth of IgR39 (non-metastatic) cells is not affected by pressure after 3 days, b) but cells grow significantly less after 6 days. c) IgR37 (metastatic) cells are unaffected by pressure both after c) 3 or d) 6 days. Statistically significant results according to the KS test ($p < 10^{-2}$) are denoted with *. doi:10.1371/journal.pone.0094229.g002

Table 1. Effect of osmotic pressure on cell death.

	3 days		6 days	
	Necrosis	Early Apoptosis	Necrosis	Early Apoptosis
0 kPa	0.47	1.17	0.47	9.48
0.2 kPa	0.36	0.32	0.38	6.51
1 kPa	1.1	0.94	1.91	9.56

Percentage of IgR39 cells in necrosis or early apoptosis after 3 or 6 days for various osmotic pressures in a typical experiment.

doi:10.1371/journal.pone.0094229.t001

We also perform a colony formation experiment using the crystal violet assay on primary and metastatic melanoma cells, following the prescriptions of Ref.[42]. In Fig 3a we report the cumulative colony size distribution $P(s)$, where s is the number of cells in each colony, for IgR39 cells with or without osmotic pressure. Each distribution is obtained by combining the result of colonies obtained in six different wells. Typical images of the colonies are reported in Fig 3c. We fit the distributions with the solution of a continuum time branching process in which each cell divides with rate γ which yields a cumulative colony size distribution after t days given by [42]

$$P(s) = (1 - e^{-\gamma t})^s \text{ for } s \geq 1. \quad (4)$$

From the fit, we estimate the average division rate as $\gamma = 0.60$ divisions/day in normal conditions and $\gamma = 0.51$ divisions/day under an osmotic pressure of 1 kPa. In Fig. 3b we report the average colony size with its standard error for IgR39 and IgR37 cells with and without osmotic pressure. While osmotic pressure hinders colony growth in both cases, the effect is much stronger for primary tumor cells than in metastatic ones.

Effect of collagen on the growth of melanoma cells. To confirm that the general effect of stress we observe on growing melanoma cells is not restricted to osmotic pressure, we perform proliferation experiments using collagen coated plates simulating the effect of the extracellular matrix on the cells. As shown in Fig. 4, the presence of collagen significantly reduces proliferation

in primary melanoma cells (IgR39) while the effect is much weaker in metastatic cells (IgR37). We next induce an osmotic pressure of 1 kPa in the collagen coated plates and find no additional significant result with respect to the case with collagen alone. We can interpret this result by assuming that collagen already induces a strong stress on the cells so that additional osmotic stress has no significant effect of the growth. We can thus suggest that osmotic pressure and collagen both act in similar ways on melanoma cells: they reduce proliferation but the effect is much stronger in primary cells than in metastatic ones.

Numerical simulations

Morphology of melanocytic nevi depends on the location of the initiating cell. We perform numerical simulations of the growth of melanocytic nevi for different conditions. In particular, we study the morphology resulting from the growth using different locations of the initial seed. As illustrated in Fig 5, we consider four type of initial locations: deep inside the epidermis (Fig. 5a and movie S1), in the epidermis but close to the basal membrane, either in the minima (Fig. 5b, Fig. 5e and and movies S2–S3) or on the maxima of the membrane (Fig. 5c, Fig. 5f and and movie S4–S5), or inside the dermis (Fig. 5d and movie S6). For initiating cells close to the basal membrane, we consider two different scenarios with a strong (Fig. 5b and 5c) or weak membrane (Fig. 5e and 5f). Fig. 5 indicates that the morphology of the nevus depends substantially on the location of the initiating cell. When the nevus starts up in the epidermis the growth is mostly horizontal and far from the dermis (Fig. 5a). For initiating cells residing close to a

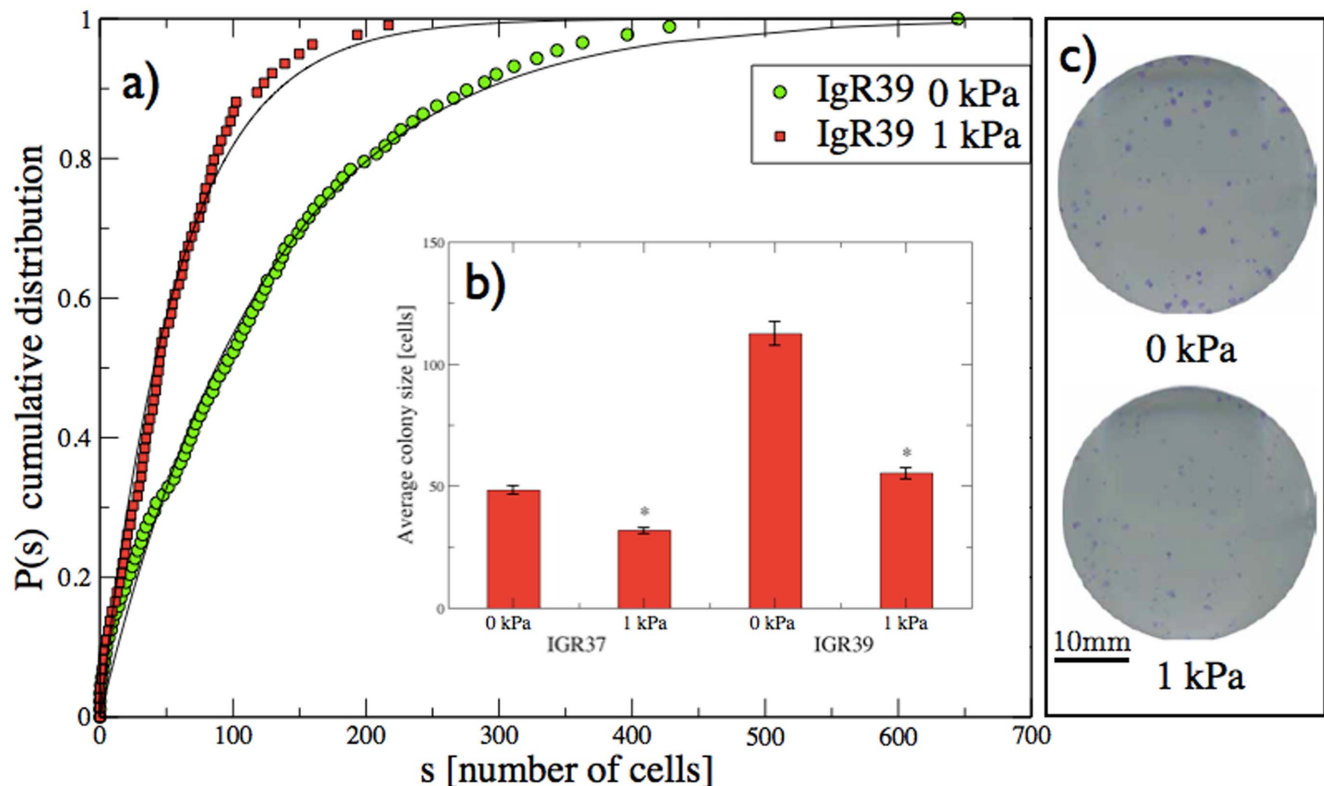


Figure 3. Effect of osmotic pressure on colony formation in melanoma. a) The cumulative distributions of colony size obtained from IgR39 cells under 1 kPa osmotic pressure with respect to the control (0 kPa). The curves are the fit with a continuous time branching process model (see Ref [42]) yielding a division rate of $\gamma = 0.60$ that is reduced to $\gamma = 0.51$ under osmotic pressure. b) The average value of the colony size distribution with the associated standard error for IgR39 and IgR37 cells. Statistically significant results according to the KS test ($p < 10^{-2}$) are denoted with *. c) The images show two representative examples of the colonies for 0 kPa and 1 kPa conditions. doi:10.1371/journal.pone.0094229.g003

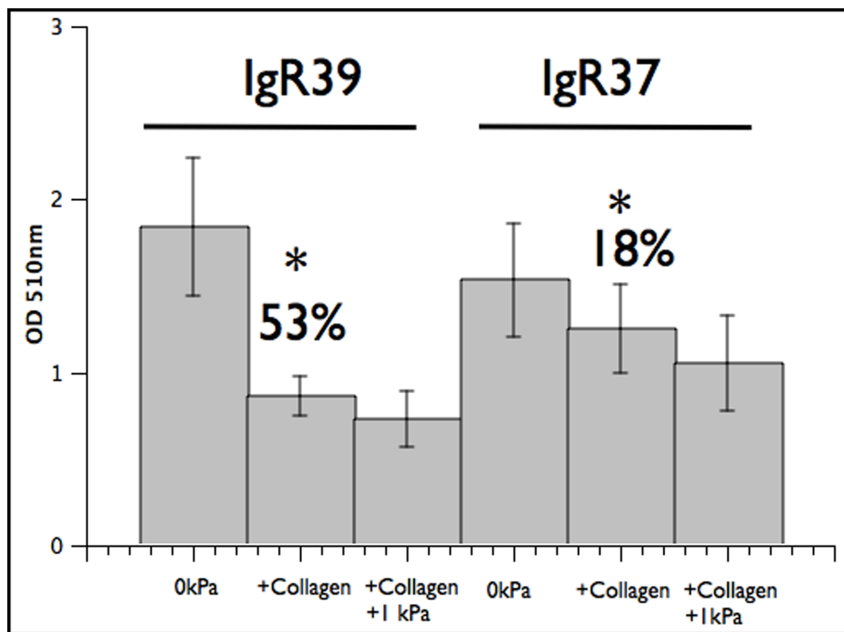


Figure 4. Effect of collagen coating on cell proliferation in melanoma. 500 cells/well were plated on 96 collagen coated multiwells (IgR39 and IgR37) and submitted to 1 kPa osmotic pressure for 6 days. Cell growth was evaluated as in Fig.2. The graph shows that the presence of collagen slows cell growth both in primary (IgR39) and metastatic (IgR37) cells but the effect is much stronger for primary cells. Additional osmotic pressure does not significantly alter the results. Statistically significant results, according to the KS test ($p < 10^{-2}$), are denoted with *. doi:10.1371/journal.pone.0094229.g004

strong basal membrane the growth follows the profile of the membrane (Fig. 5b) or expands vertically in the epidermis (Fig. 5c) depending on whether the initiating cells lies in the minima or maxima defined by the membrane profile. When the membrane is weak, however, it is easily broken by the growing cells that then

invade the dermis (Fig. 5e and Fig. 5f). The growth is more radial when the initiating cells is in the minimum of the basal membrane (Fig. 5e) and more vertical when it lies close to the maximum (Fig. 5e).

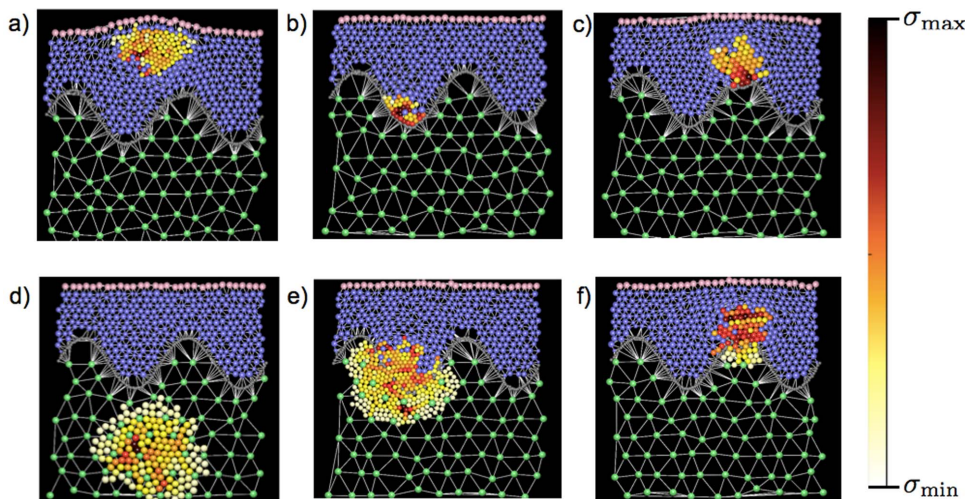


Figure 5. Morphology of nevi for different locations of the initiating cell for random growth. Illustration of the results of numerical simulations for nevi grown from melanocytes located in different positions in the skin and for different mechanical properties of the basal membrane. Here growth occurs randomly and independently on compressive stress. Growing melanocytes are shown with a varying color that reflects their compressive stresses σ according to the color bar. The maximum $\sigma_{\max}$ and minimum $\sigma_{\min}$ for each configuration are equal to (in kPa): a) 0.61–0.77, b) 0.0043–0.21, c) 0.085–0.57, d) 0.0013–0.149, e) 0.0047–0.37, f) 0.0067–0.72. a) Nevi grown from melanocytes residing inside the epidermis tend to grow horizontally and do not spread towards the basal membrane. b) Nevi grown on the minima of a strong basal membrane tend to grow roughly parallel to the membrane itself. c) Nevi growing from the maxima of a strong basal membrane tend to grow vertically in the epidermis. d) Dermal nevi tend to have radial shape. When the basal membrane is weak, e) nevi growing from the minima of the basal membrane invade the dermis in a radial fashion, while f) when they start from maxima of the basal membrane the invasion of the dermis occurs more vertically. doi:10.1371/journal.pone.0094229.g005

Mechanical stresses in growing melanocytic nevi. We record the compressive stresses sustained by the melanocytes during the growth of the simulated nevi. A pictorial view of the stress in each cell is shown in Fig. 5, where the color scale depicts compressed cells with a scale going from yellow (low stress) to red (high stress). The images show that stresses are in general heterogeneously distributed and, as illustrated by the supplementary movies, fluctuate considerably after each division step. We notice that typically the most stressed cells are located in the inner part of the nevus, reproducing a general feature observed in tumor spheroids [11,12]. Furthermore, we see the formation of stress chains, typical of granular assemblies [43]. To better quantify the evolution of the compressive stress we measure the average compressive stress σ experienced by the cells as a function of the number of cells in the nevus. The results reported in Fig. 6 correspond to the different cases explored in Fig. 5. In all cases, pressure builds up as cells are duplicating until at some point the connective tissue breaks drastically reducing the average compressive stress experienced by the cells. This effect is particularly clear in the case of a nevus growing in the epidermis (see Fig. 6a). The large error bars reported in Fig. 6 indicate that stresses vary widely between individual cells but also among different realizations of the process.

To better quantify the stress fluctuations experienced by individual cells, we report in Fig. 7 an estimate of the distribution (probability density function) $\rho(\sigma)$ of the compressive stresses σ for different initial conditions. The distribution is sampled over different realizations and over different cells. Since the distribution is expected to change with the number of cells present in the nevus (as shown from the error bars in Fig. 6), we consider only the case for which the average pressure is highest (the peak stress in each

curve in Fig. 6). In all cases the distribution displays an approximately exponential tail (i.e. $\rho(\sigma) \simeq \exp(-\sigma/\sigma_0)$).

Compressive stresses are much higher for nevi growing in the epidermis than in the dermis. This is due to the general assumption made in the model that cells are more packed in the epidermis than in the dermis. When we include a direct interaction between the nevi cells and the ECM, as discussed in the model section, compressive stresses in the dermis increases but the general features of the results are unchanged (see Fig. 8 and Movie S7). We have also tested for the effect of adhesion between nevi cells and found very little differences with respect to the case in which adhesion is absent (see Fig. 8).

Rupture of the basal membrane. The results reported in Fig. 5e and 5f refer to the case in which the basal membrane breaks due to the action of mechanical forces. Simulations of the rupture of the basal membrane induced by MMPs are reported in Fig. 9 and in Movie S8. In particular, the Movie S8 illustrates how the effect of MMPs leads to a much rapid invasion of the dermis. This is because MMPs typically break the basal membrane in different locations, while when the membrane ruptures mechanically in one location the stress in the rest of the membrane is released preventing further rupture. Finally, we notice that the average compressive stress in the nevus is reduced when MMPs are present (see Fig. 9b).

Stress dependent growth. Inspired by the experimental results on primary melanoma that indicating that compressive stress inhibits growth, we perform the simulations with a stress dependent division rate, so that compressed cells divide less. The morphology of the resulting nevi changes only slightly with respect of the previous case (compare Fig. S1 with Fig. 5), but we observe the development of incipient finger-like structures (see in particular Fig. S1c and S1f). We also measure the average compressive stress

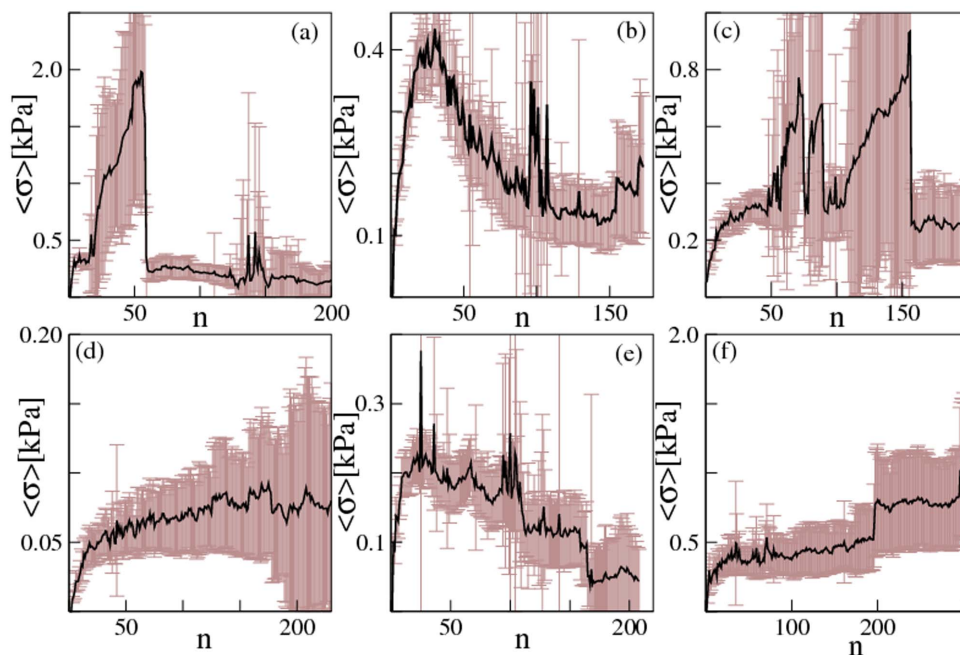


Figure 6. Mechanical stresses in randomly growing nevi. We report the evolution of the average compressive stress experienced by the melanocytes composing the nevus as a function of the size of the nevus, quantified by the total number of cells n , for the same conditions as in Fig. 5. The error bars represent the standard error of the mean. The stress is averaged over all the cells in a nevus and over at least ten statistically independent realizations of the growth process. The different panels represent different initial locations: a) in the middle of the epidermis, b) in the minima of a strong basal membrane, c) in the maxima of a strong basal membrane, d) in the dermis, e) in the minima of a weak basal membrane, f) in the maxima of a weak basal membrane.

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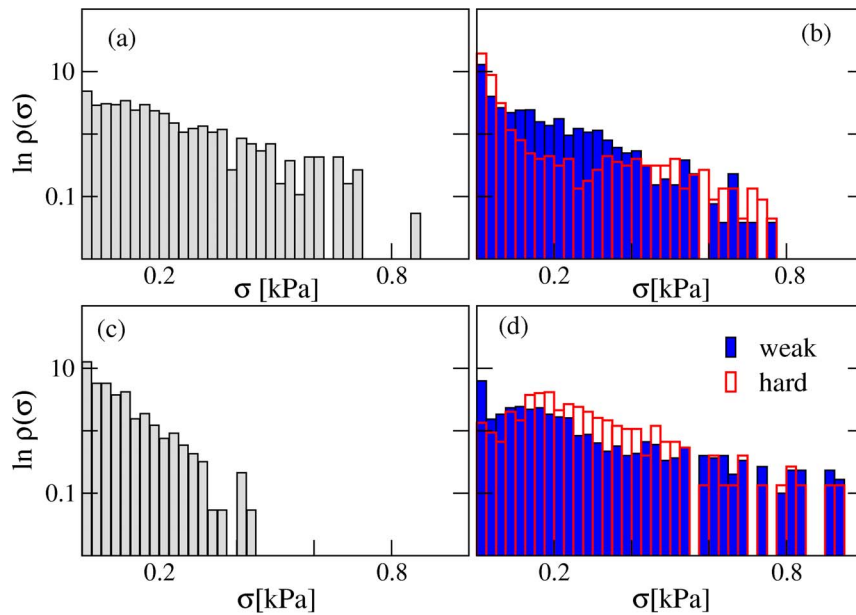


Figure 7. Compressive stress distribution in nevi. We report the distribution $\rho(\sigma)$ of the compressive stresses σ (in kPa) experienced by melanocytes in a nevus. The results are sampled over different realizations of the process. Since the distribution also changes with time, we only consider the time at which the average compressive stress $\langle \sigma \rangle$ is highest. The different panels represent different initial conditions corresponding to Fig. 5: a) in the middle of the epidermis, b) in the minima of the basal membrane (for strong and weak membranes) c) in the dermis, d) in the maxima of the basal membrane (for strong and weak membranes).
doi:10.1371/journal.pone.0094229.g007

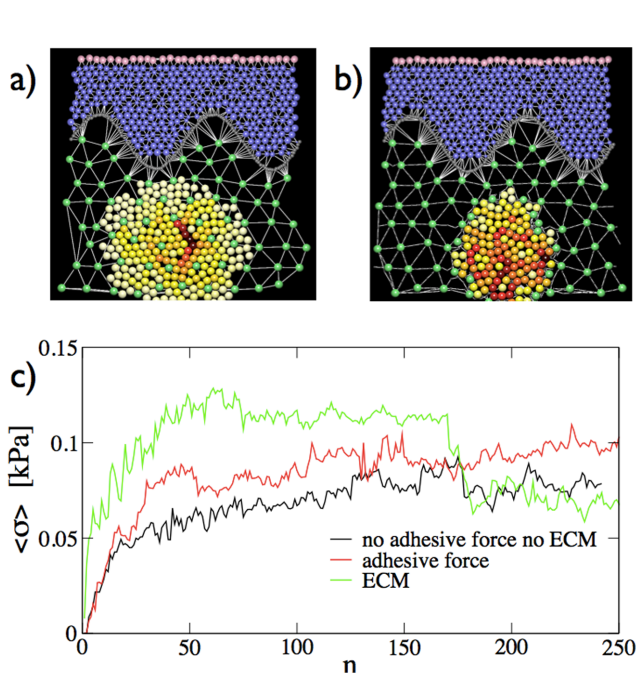


Figure 8. Role of adhesion forces and direct interactions with the ECM. Morphology of a nevus grown in the dermis a) in presence of adhesive forces between nevi cells and b) considering direct interactions with the ECM modelled as discussed in the methods section. c) The evolution of the average compressive stress experienced by the melanocytes composing the nevus for cases a) and b) is compared with a simulation in which no adhesive forces and interactions with the ECM are present.
doi:10.1371/journal.pone.0094229.g008

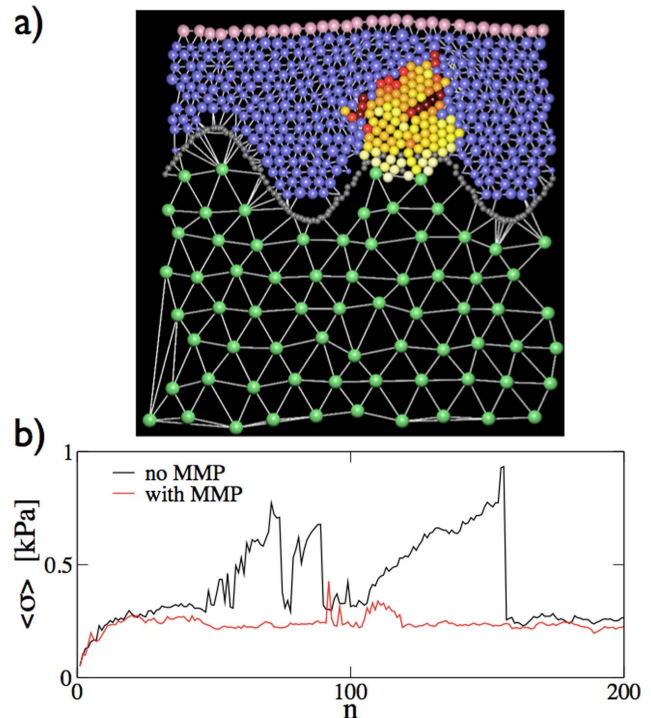


Figure 9. Model of MMP induced breaking of the basal membrane. a) We report the typical morphology of a nevus starting from the epidermis with a strong basal membrane that can however be broken by MMPs. b) The evolution of the average compressive stress in presence of MMPs is compared with the case in which MMPs are not present.
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and find that it is typically smaller than in the previous case (see Fig. S2). This is due to the fact that in this case growth occurs in regions that are less compressed so that there is more space to accommodate new cells.

Comparison with histological images. In Fig. 10, we report two examples of common melanocytic nevi as revealed by histological sections. In Fig. 10a, we suggest that an intraepithelial junctional nevus that is all confined in the epidermis (compare with Fig. 5b). Views at different magnifications show that nevi cells are closely packed and press against the basal membrane as indicated by rounded pocket shown in the left panel. In Fig. 10b, we report an example of dermal nevus (compare with Fig. 5d). In this case, cells are less packed and are scattered all through the dermis without pressing on the basal membrane. These images are in good qualitative with the results of our model that show a tendency for nevi cells to press from the epidermis into the dermis rather than the reverse. This effect is likely due to the different mechanical properties of the two skin layers, as assumed in the model.

Discussion

Melanoma is a very aggressive tumor, being chemio- and radio-resistant once it becomes metastatic [44]. Its incidence is increasing both in Europe and in US, making it a current challenging problem for research. It is commonly believed that nevi may progress into melanoma, or alternatively regress by differentiating [45]. Since diagnosis is mostly based on dermoscopic and histological analysis, it is extremely important to correlate the morphological features of nevi to the degree of malignancy or pre-malignancy of the lesions. Since nevi proliferate inside a complex microenvironment, we investigated the role of mechanics and geometry on the morphology and internal stresses of melanocytic nevi. Using human melanoma cell lines, we confirm also for melanoma a negative effect of the mechanical stress on cellular proliferation as already reported for other tumors [8,9,11,12]. We show that osmotic pressure is more effective in primary melanoma cells than in metastatic ones, suggesting that metastatic cells come from a subpopulation more aggressive and

insensitive to the mechanical properties of the environment. We corroborate this result by experiments of cell growth in collagen coated plates where again primary cells are more affected than metastatic ones. This suggests that cells composing melanocytic nevi may be sensitive to stress as well, an issue that we investigated computationally.

We have devised a computational model for the growth of melanocytic nevi in a layered tissue representing the skin. Numerical simulations of the model show that the morphology of the resulting nevus depends considerably on the environment in which the growth takes place, in the epidermis close or far from the basal membrane or in the dermis. This is interesting since environmental properties, in this case the mechanical behavior of the tissue, is rarely considered in the progression of nevi. Simulations indicate that nevi are subject to fluctuating compressive stresses due to the tissue elasticity. If we introduce a stress dependence proliferation rate, we observe a decrease in the overall stress. Our computational model is appropriate to describe a benign nevus, since we do not consider active motion of the cells but only their quasi-equilibrium displacement in response to elastic forces. To model the growth of melanoma cells, one should implement their mobility and their ability to degrade the surrounding extracellular matrix, by expressing MMPs. We have incorporated this last aspect in the simulations showing that the invasion of the dermis occurs more rapidly. It is remarkable that many intriguing features observed in melanoma, such as the formation of rough tumor boundaries and the invasion of the dermis from the epidermis, are observed from simple rules combining mechanics and geometry, although we simplified many of the irregularities present in the dermis. As a matter of fact we expect that, while the overall shape of the nevus can be influenced by the degree of accuracy in the dermis description, the qualitative picture remains unchanged. A limitation of our model lies in its two-dimensional nature that, while it greatly simplifies the numerical calculations, could affect the values of the quantities we measure. We expect, however, that the general phenomenology we observe should be unchanged in a more realistic three-dimensional situation.

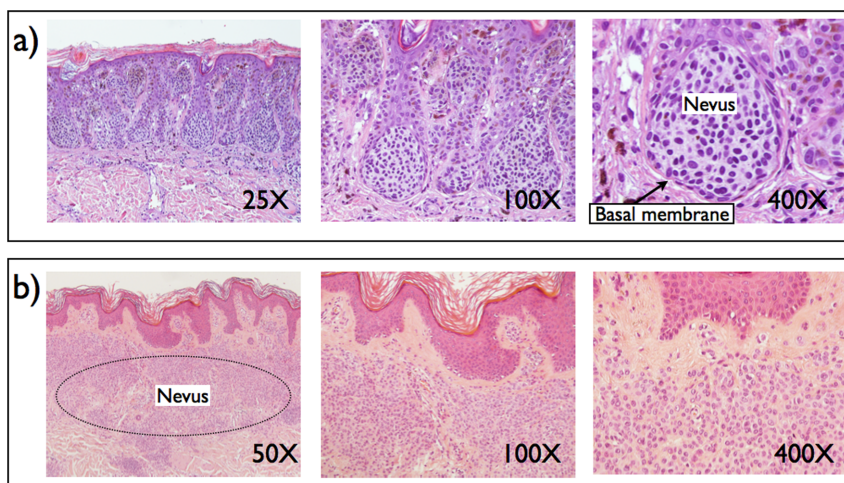


Figure 10. Histological images of melanocytic nevi. We illustrate the morphology observed by optical microscopy at different magnifications on histological sections of two types of nevi. a) Intraepithelial junctional nevi are confined in the epidermis and press against the basal membrane forming characteristic rounded pockets of densely packed cells (compare with Fig. 5b) b) Dermal nevi are confined in the dermis (compare with Fig. 5d). Cells are loosely packed and do not touch the basal membrane. The basal membrane is located at the separation between the upper (epidermis) and lower (dermis) skin layers and indicated by an arrow in panel a). Images courtesy of Dr. Claudio Clemente.

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Materials and Methods

Dextran solution

A master solution of dextran at 10% (w/v) was formed (Dextran from Leuconostoc spp, Fluka) and the diluted to the desired concentration with complete medium. Transformation from dextran concentration to osmotic pressure was performed according to the calibration curve measured in Ref. [46].

Cell lines

Human IGR39 and IGR37 cells were obtained from Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH and cultured as previously described [47]. IGR39 was derived from a primary amelanotic cutaneous tumor and IGR37 from an inguinal lymph node metastasis in the same patient.

Cell growth with collagen

The cells were plated on pre-coated collagen type I dishes (Sigma) according to the manufacturers instructions.

Colony growth

Cells are plated on 6 multiwells, fixed with 3.7% paraformaldehyde (PFA) for 5 minutes and then stained for 30 min with 0.05% crystal violet solution. After two washing with tap water, the plates are drained by inversion for a couple of minutes. In order to control the merging of different colonies, the experiments are performed with different initial cell concentrations as described recently by our group [42]. Data analysis of the resulting colonies has been performed according to Ref. [42].

Sulforhodamine B colorimetric assay for cytotoxicity screening

The sulforhodamine B (SRB) assay is used for cell density determination based on the cellular protein content according to Ref. [48]

Apoptosis detection

The Annexin V-FITC Apoptosis Detection kit by Sigma was used to detect apoptotic and necrotic cells by detecting annexin V-FITC and propidium iodide by flow cytometry, respectively. Briefly the cells were incubated with Annexin V-FITC and propidium iodide at room temperature for 10 minutes and protect by light. Then the cells were immediately analysed by flow cytometry (FACS Aria flow cytometer (Becton, Dickinson and Company, BD, Mountain View, CA). Data were analyzed using FlowJo software (Tree Star, Inc., San Carlos, CA).

Histological analysis of bioptic tissues

Tissue specimens are immediately fixed in neutral buffered formalin, embedded in paraffin, sectioned and subjected to histopathological characterization after hematoxylin-eosin staining.

Statistical analysis

Statistical significance is evaluated according to the Kolmogorov-Smirnov non-parametric test with $p < 10^{-2}$.

Model simulations

The model is simulated through a custom made python code. To find mechanical equilibrium for the system, we use the fire relaxation method [49]. To speed up the code the relaxation routine has been written in C. Graphics rendering is done using

Povray. Simulation and visualization codes are available at <https://github.com/alexalemi/cancersim>.

Supporting Information

Figure S1 Morphology of nevi for different locations of the initiating cell for pressure dependent growth.

Illustration of the results of numerical simulations for nevi grown from melanocytes located in different positions in the skin and for different mechanical properties of the basal membrane. The conditions are the same as in Fig. 5, but here the growth depends on the compressive stress acting on each cell. Growing melanocytes are shown with a varying color that reflects their compressive stresses according to the color bar. The maximum $\sigma_{\max}$ and minimum $\sigma_{\min}$ for each configuration are equal to (in kPa): a) $7.46 \cdot 10^{-6} - 0.13$, b) $8.2410^{-6} - 0.52$, c) $0.0016 - 0.63$ d) $0.00015 - 0.136$ e) $0.00039 - 0.043$ f) $0.0022 - 0.428$. (TIFF)

Figure S2 Mechanical stresses in pressure dependent growth of nevi.

We report the evolution of the average compressive stress experienced by the melanocytes composing the nevus as a function of the size of the nevus, quantified by the number of cells n , for the same conditions as in Fig. S1. The error bars represent the standard error of the mean. The different panels represent different initial locations: a) in the middle of the epidermis, b) in the minima of a strong basal membrane, c) in the maxima of a strong basal membrane, d) in the dermis, e) in the minima of a weak basal membrane, f) in the maxima of a weak basal membrane. (TIFF)

Movie S1 Growth of a nevus starting from a melanocyte placed deep inside the epidermis.

(AVI)

Movie S2 Growth of a nevus starting from a melanocyte placed in the epidermis at the minimum of a weak basal membrane.

(AVI)

Movie S3 Growth of a nevus starting from a melanocyte placed in the epidermis at the minimum of a strong basal membrane.

(AVI)

Movie S4 Growth of a nevus starting from a melanocyte placed in the epidermis at the maximum of a weak basal membrane.

(AVI)

Movie S5 Growth of a nevus starting from a melanocyte placed in the epidermis at the maximum of a strong basal membrane.

(AVI)

Movie S6 Growth of a nevus starting from a melanocyte placed deep inside the dermis.

(AVI)

Movie S7 Growth of a nevus starting from a melanocyte placed deep inside the dermis considering a direct interaction with the ECM.

(AVI)

Movie S8 Growth of a nevus starting from a melanocyte placed in the epidermis at the maximum of a strong basal membrane that can be broken by MMPs.

(AVI)

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References

- Baxter LL, Pavan WJ (2013) The etiology and molecular genetics of human pigmentation disorders. *Wiley Interdiscip Rev Dev Biol* 2: 379–92.
- Kim JK, Nelson KC (2012) Dermoscopic features of common nevi: a review. *G Ital Dermatol Venerol* 147: 141–8.
- Zalaudek I, Manzo M, Savarese I, Docimo G, Ferrara G, et al. (2009) The morphologic universe of melanocytic nevi. *Semin Cutan Med Surg* 28: 149–56.
- Unna PG (1893) Naevi and naevocarcinome. *Berl Klin Wochenschr* 30.
- Cramer SF (1988) The melanocytic differentiation pathway in congenital melanocytic nevi: theoretical considerations. *Pediatr Pathol* 8: 253–65.
- Kittler H, Selteneim M, Dawid M, Pehamberger H, Wolff K, et al. (2000) Frequency and characteristics of enlarging common melanocytic nevi. *Arch Dermatol* 136: 316–20.
- Weaver VM, Petersen OW, Wang F, Larabell CA, Briand P, et al. (1997) Reversion of the malignant phenotype of human breast cells in three-dimensional culture and in vivo by integrin blocking antibodies. *J Cell Biol* 137: 231–45.
- Helminger G, Netti PA, Lichtenbeld HC, Melder RJ, Jain RK (1997) Solid stress inhibits the growth of multicellular tumor spheroids. *Nat Biotech* 15: 778–783.
- Cheng G, Tse J, Jain RK, Munn LL (2009) Micro-environmental mechanical stress controls tumor spheroid size and morphology by suppressing proliferation and inducing apoptosis in cancer cells. *PLoS ONE* 4: e4632.
- Samuel MS, Lopez JI, McGhee EJ, Croft DR, Strachan D, et al. (2011) Actomyosin-mediated cellular tension drives increased tissue stiffness and β -catenin activation to induce epidermal hyperplasia and tumor growth. *Cancer Cell* 19: 776–91.
- Montel F, Delarue M, Elgeti J, Malaquin L, Basan M, et al. (2011) Stress clamp experiments on multicellular tumor spheroids. *Phys Rev Lett* 107: 188102.
- Montel F, Delarue M, Elgeti J, Vignjevic D, Cappello G, et al. (2012) Isotropic stress reduces cell proliferation in tumor spheroids. *New Journal of Physics* 14: 055008.
- Tse JM, Cheng G, Tyrrell JA, Wilcox-Adelman SA, Boucher Y, et al. (2012) Mechanical compression drives cancer cells toward invasive phenotype. *Proc Natl Acad Sci U S A* 109: 911–6.
- Paszek MJ, Zahir N, Johnson KR, Lakins JN, Rozenberg GI, et al. (2005) Tensional homeostasis and the malignant phenotype. *Cancer Cell* 8: 241–54.
- Hoffman BD, Grashoff C, Schwartz MA (2011) Dynamic molecular processes mediate cellular mechanotransduction. *Nature* 475: 316–323.
- Zaman MH, Trapani LM, Sieminski AL, Siemeski A, Mackellar D, et al. (2006) Migration of tumor cells in 3d matrices is governed by matrix stiffness along with cell-matrix adhesion and proteolysis. *Proc Natl Acad Sci U S A* 103: 10889–94.
- Koch TM, Münster S, Bonakdar N, Butler JP, Fabry B (2012) 3d traction forces in cancer cell invasion. *PLoS One* 7: e33476.
- Racz B, Reglodi D, Fodor B, Gasz B, Lubics A, et al. (2007) Hyperosmotic stress-induced apoptotic signaling pathways in chondrocytes. *Bone* 40: 1536–43.
- Nielsen MB, Christensen ST, Hoffmann EK (2008) Effects of osmotic stress on the activity of MAPKs and PDGFR-beta-mediated signal transduction in NIH-3T3 fibroblasts. *Am J Physiol Cell Physiol* 294: C1046–55.
- Simonsen TG, Gaustad JV, Leinaas MN, Rofstad EK (2012) High interstitial fluid pressure is associated with tumor-line specific vascular abnormalities in human melanoma xenografts. *PLoS One* 7: e40006.
- Wu M, Frieboes HB, McDougall SR, Chaplain MAJ, Cristini V, et al. (2013) The effect of interstitial pressure on tumor growth: coupling with the blood and lymphatic vascular systems. *J Theor Biol* 320: 131–51.
- Walker DC, Southgate J, Hill G, Holcombe M, Hose DR, et al. (2004) The epitheliome: agent-based modelling of the social behaviour of cells. *Biosystems* 76: 89–100.
- Holcombe M, Adra S, Bicak M, Chin S, Coakley S, et al. (2012) Modelling complex biological systems using an agent-based approach. *Integr Biol (Camb)* 4: 53–64.
- Galle J, Loeffler M, Drasdo D (2005) Modeling the effect of deregulated proliferation and apoptosis on the growth dynamics of epithelial cell populations in vitro. *Biophysical Journal* 88: 62–75.
- Drasdo D, Höhme S (2005) A single-cell-based model of tumor growth in vitro: monolayers and spheroids. *Physical Biology* 2: 133.
- Drasdo D, Höhme S (2012) Modeling the impact of granular embedding media, and pulling versus pushing cells on growing cell clones. *New Journal of Physics* 14: 055025.
- Simpson MJ, Binder BJ, Haridas P, Wood BK, Treloar KK, et al. (2013) Experimental and modelling investigation of monolayer development with clustering. *Bull Math Biol* 75: 871–89.
- Treloar KK, Simpson MJ, Haridas P, Manton KJ, Leavesley DI, et al. (2013) Multiple types of data are required to identify the mechanisms influencing the spatial expansion of melanoma cell colonies. *BMC Syst Biol* 7: 137.
- Plank MJ, Simpson MJ (2012) Models of collective cell behaviour with crowding effects: comparing lattice-based and lattice-free approaches. *J R Soc Interface* 9: 2983–96.
- Chatelain C, Balois T, Ciarletta P, Amar MB (2011) Emergence of microstructural patterns in skin cancer: a phase separation analysis in a binary mixture. *New Journal of Physics* 13: 115013.
- Eikenberry S, Thalhauser C, Kuang Y (2009) Tumor-immune interaction, surgical treatment, and cancer recurrence in a mathematical model of melanoma. *PLoS Comput Biol* 5: e1000362.
- Adra S, Sun T, MacNeil S, Holcombe M, Smallwood R (2010) Development of a three dimensional multiscale computational model of the human epidermis. *PLoS One* 5: e8511.
- Thingnes J, Lavelle TJ, Hovig E, Omholt SW (2012) Understanding the melanocyte distribution in human epidermis: an agent-based computational model approach. *PLoS One* 7: e40377.
- Hendriks FM, Brokken D, van Emmeren J, Oomens C, Baajens F, et al. (2003) A numerical-experimental method to characterize the non-linear mechanical behaviour of the human skin. *Skin Research and Technology* 9: 274–283.
- Hendriks FM (2005) Mechanical behaviour of human epidermal and dermal layers in vivo. *Technische Universiteit Eindhoven: Ph. D. Thesis*.
- Hendriks FM, Brokken D, Oomens C, Bader D, Baajens F (2006) The relative contributions of different skin layers to the mechanical behaviour of human skin in vivo using suction experiments. *Medical Engineering & Physics* 28: 259–266.
- James AG, Green A, Simpson G (1975) Strain energy functions of rubber. i. characterization of gum vulcanizates. *Journal of Applied Polymer Science* 19: 2033–2058.
- MSC Software Corporation (2001) Volume A: Theory and user information, Version 2001. MSC.MARC, 1 edition.
- Long R, Hall MS, Wu MM, Hui CH (2011) Effects of gel thickness on microscopic indentation measurements of gel modulus. *Biophysical Journal* 101: 643–650.
- Lekka M, Laidler P, Gil D, Lekki J, Stachura Z, et al. (1999) Elasticity of normal and cancerous human bladder cells studied by scanning force microscopy. *Eur Biophys J* 28: 312–6.
- Drasdo D, Kree R, McCaskill J (1995) Monte Carlo approach to tissue-cell populations. *Physical Review E* 52: 6635–6657.
- Baraldi MM, Alemi AA, Sethna JP, Caracciolo S, La Porta CAM, et al. (2013) Growth and form of melanoma cell colonies. *Journal of Statistical Mechanics: Theory and Experiment* 2013: P02032.
- Howell D, Behringer RP, Veje C (1999) Stress fluctuations in a 2d granular couette experiment: A continuous transition. *Phys Rev Lett* 82: 5241–5244.
- La Porta CAM (2009) Mechanism of drug sensitivity and resistance in melanoma. *Curr Cancer Drug Targets* 9: 391–397.
- Clark WH, Elder DE, Guerry D, Epstein MN, Greene MH, et al. (1984) A study of tumor progression: the precursor lesions of superficial spreading and nodular melanoma. *Hum Pathol* 15: 1147–65.
- Bonnet-Gonnet C, Belloni L, Cabane B (1994) Osmotic pressure of latex dispersions. *Langmuir* 10: 4012–4021.
- Taghizadeh R, Noh M, Huh YH, Ciusani E, Sigalotti L, et al. (2010) CXCR6, a newly defined biomarker of tissue-specific stem cell asymmetric self-renewal, identifies more aggressive human melanoma cancer stem cells. *PLoS One* 5: e15183.
- Vichai V, Kirtikara K (2006) Sulforhodamine B colorimetric assay for cytotoxicity screening. *Nat Protoc* 1: 1112–6.
- Bitzek E, Koskinen P, Gähler F, Moseler M, Gumbsch P (2006) Structural relaxation made simple. *Phys Rev Lett* 97: 170201.

Author Contributions

Conceived and designed the experiments: CAMLP. Performed the experiments: EC CAMLP. Analyzed the data: CAMLP AT SZ. Wrote the paper: AT SZ CAMLP. Designed the computational model: AT JPS SZ. Wrote the simulation code: AAA. Performed numerical simulations: AT.