



OPEN ACCESS

EDITED BY

Rajesh Katare,
University of Otago, New Zealand

REVIEWED BY

Diana Passaro,
INSERM U1016 Institut Cochin, France
Tomer Itkin,
Tel Aviv University, Israel

*CORRESPONDENCE

Camilla Cerutti
✉ Camilla.Cerutti@ieo.it
Gaia Spinetti
✉ gaia.spinetti@multimedica.it

RECEIVED 19 July 2023

ACCEPTED 12 September 2023

PUBLISHED 17 October 2023


CITATION

Campanile M, Bettinelli L, Cerutti C and Spinetti G (2023) Bone marrow vasculature advanced *in vitro* models for cancer and cardiovascular research.
Front. Cardiovasc. Med. 10:1261849.
doi: 10.3389/fcvm.2023.1261849

COPYRIGHT

© 2023 Campanile, Bettinelli, Cerutti and Spinetti. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Bone marrow vasculature advanced *in vitro* models for cancer and cardiovascular research

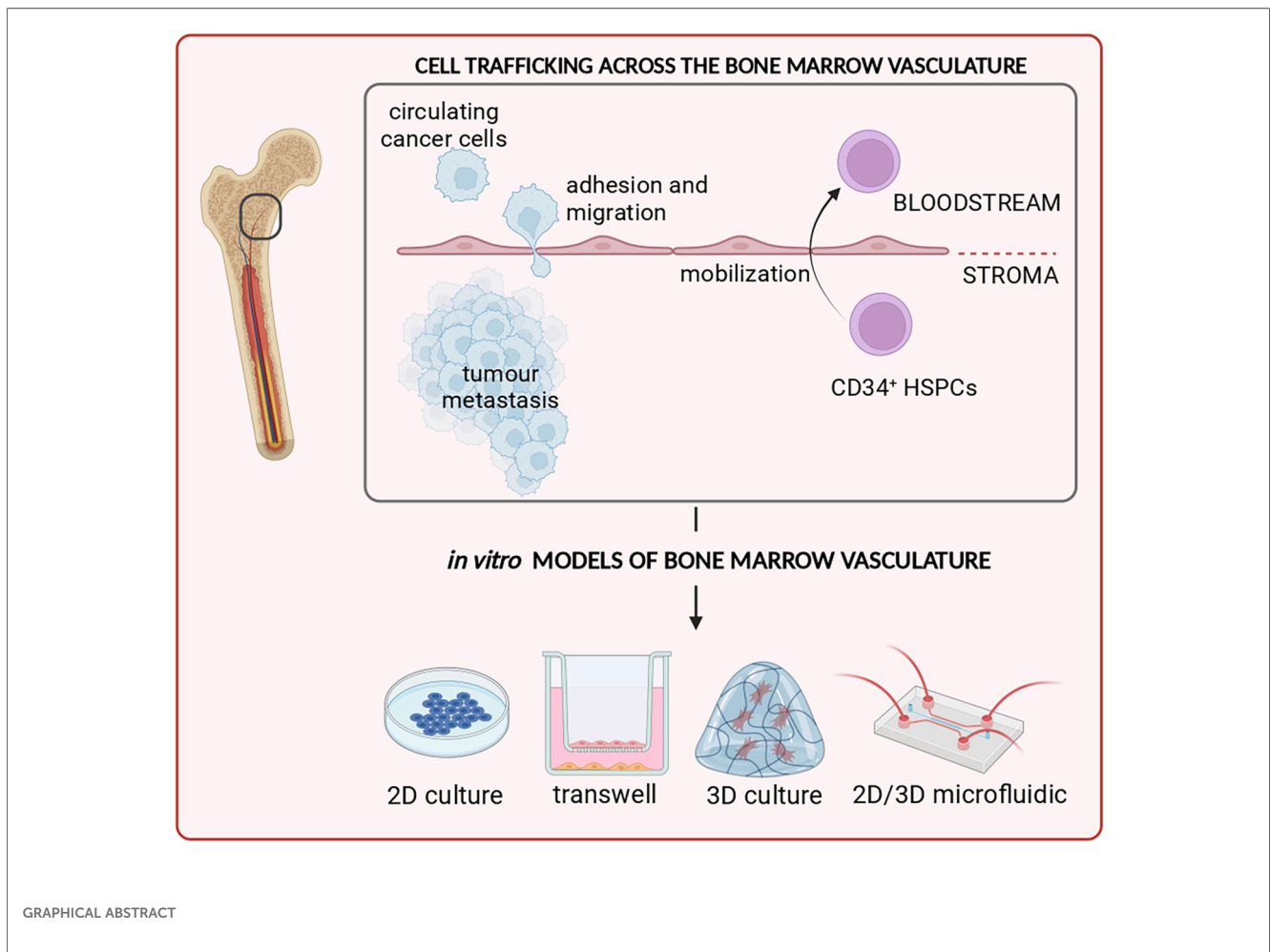
Marzia Campanile¹ , Leonardo Bettinelli^{1,2}, Camilla Cerutti^{2*} and Gaia Spinetti^{1*}

¹Laboratory of Cardiovascular Research, IRCCS MultiMedica, Milan, Italy, ²Department of Experimental Oncology, IRCCS-IEO, European Institute of Oncology, Milan, Italy.

Cardiometabolic diseases and cancer are among the most common diseases worldwide and are a serious concern to the healthcare system. These conditions, apparently distant, share common molecular and cellular determinants, that can represent targets for preventive and therapeutic approaches. The bone marrow plays an important role in this context as it is the main source of cells involved in cardiovascular regeneration, and one of the main sites of liquid and solid tumor metastasis, both characterized by the cellular trafficking across the bone marrow vasculature. The bone marrow vasculature has been widely studied in animal models, however, it is clear the need for human-specific *in vitro* models, that resemble the bone vasculature lined by endothelial cells to study the molecular mechanisms governing cell trafficking. In this review, we summarized the current knowledge on *in vitro* models of bone marrow vasculature developed for cardiovascular and cancer research.

KEYWORDS

bone marrow vasculature, *in vitro* models, HSPC mobilization, cancer, diabetes



1. Introduction

Cardiovascular diseases (CVDs) and cancer are among the leading causes of death in Western countries (1, 2) and together account for over 60% of all deaths in Europe (3). CVDs may arise as a severe complication of diabetes (4) that, being largely diffuse in the global population (5), constitute a serious concern in the cardiovascular field. As for solid cancers such as breast cancer, thanks to treatment improvements and early diagnosis, primary tumors are largely treatable, however with longer life expectancy, cancer recurrence and secondary tumor (metastasis) incidence exponentially increase and are the cause of 90% of cancer-related death (6). In this context, the bone marrow (BM) represents a critical organ in both cardiovascular and cancer research.

The BM resides in the bone cavities and is deputed to new blood cell generation and release in the bloodstream; therefore it is characterized by high cellular trafficking both in health and disease. Moreover, the BM releases stem cells directing tissue regeneration in response to cardiovascular damage, as following myocardial infarction (7). The term “mobilopathy” was coined to indicate the limited CD34⁺ hematopoietic stem progenitor cell (HSPC) inside-out migration (8) observed in diabetic patients and associated with poor CVDs outcome (9–13). To facilitate cell

migration to the bloodstream the BM vasculature presents a fenestrated endothelium at the capillary level, but it may also provide easier access to cancer cells (14–16). The bone is also the primary site where blood cancer begins, such as leukemia, and a site for solid tumors metastasis formation, such as breast cancer (17). Interestingly, numerous studies identified a cross-talk between CVDs and abnormal hematopoiesis, and the central role of BM endothelial cells (ECs) in this process was assessed. CVDs induce phenotypic alteration of ECs, which modify their inflammatory cytokines profile inducing changes in the HSPCs proliferation rate (18). On the other hand, an enhanced myelopoiesis has been linked to CVDs such as atherosclerosis (19).

The understanding of the mechanisms that control hematopoiesis and cell’s trafficking from and to the BM, both in health and diseases is increasing the attention from clinical and basic researchers since they may pave the way to new preventive and therapeutic strategies in the clinic. The structure of human bone marrow vasculature is well known, but the majority of our knowledge of the molecular mechanisms of cellular trafficking derives from animals, which are known to not be able to fully mirroring human physiology (20). Therefore, to move toward more clinically relevant models, new *in vitro* cultures have been developed. The combination of these advanced *in vitro* tools with

human-derived cells will represent a step forward for mechanistic studies and for personalized medicine applications to identify prognostic and therapeutic targets.

2. Bone marrow structure and function

The BM is a vascularized tissue composed of a variety of cell types contributing to its homeostasis and function, including HSPCs, bone marrow mesenchymal stem/stromal cells (BM-MSCs), osteoblasts, osteoclasts, adipocytes, lymphatic and vascular endothelial cells (ECs), and immune cells (21) (Figure 1). The BM's main function is to generate mature blood cells during hematopoiesis, a process that leads to the generation of the different cellular components of the immune system that can traffic in and out of the BM through its vasculature. Common myeloid and lymphoid progenitors generate from the partial differentiation of multipotent progenitors, which are transient-amplifying cells, originating from the asymmetric division of short-term hematopoietic stem cells (ST-HSCs) (22).

The myeloid progenitors differentiate in erythrocytes, granulocytes (basophils, neutrophils, and eosinophils), monocytes, and megakaryocytes (platelets-producing cells), while lymphoid progenitors in lymphocytes (T-cells, B-cells, and natural killer cells) (23). Adult BM-MSCs are a heterogeneous

population of multipotent cells (24) with the ability to differentiate into cells of mesenchymal origin such as adipocytes, chondrocytes, and osteoblasts when exposed to specific growth signals (25, 26) that are forming the BM. Osteoblasts and osteoclasts are the main cell types involved in bone homeostasis and its repair upon injury. Osteoblasts regulate bone formation, and secrete the extracellular matrix (ECM) proteins such as type I collagen, osteopontin, osteocalcin, and alkaline phosphatase, important components of the BM structure, while osteoclast are large multinucleate cells, originating from the hematopoietic lineage, mainly involved in bone resorption (27). BM adipocytes are responsible for filling the medullary canal of long bones (tibia, femur, humerus), and gradually increasing their presence in the marrow during lifespan depending on marrow type (28).

2.1. Bone marrow vascular niche structure and function

HSPCs are organized in specialized BM microenvironments, called "BM niches", that regulate HSPC self-renewal, differentiation, and mobilization (29–32). Two types of BM niches have been identified based on their localization: the endosteal and the perivascular niche which could be further classified as periarterial and perisinusoidal (33) (Figure 1).

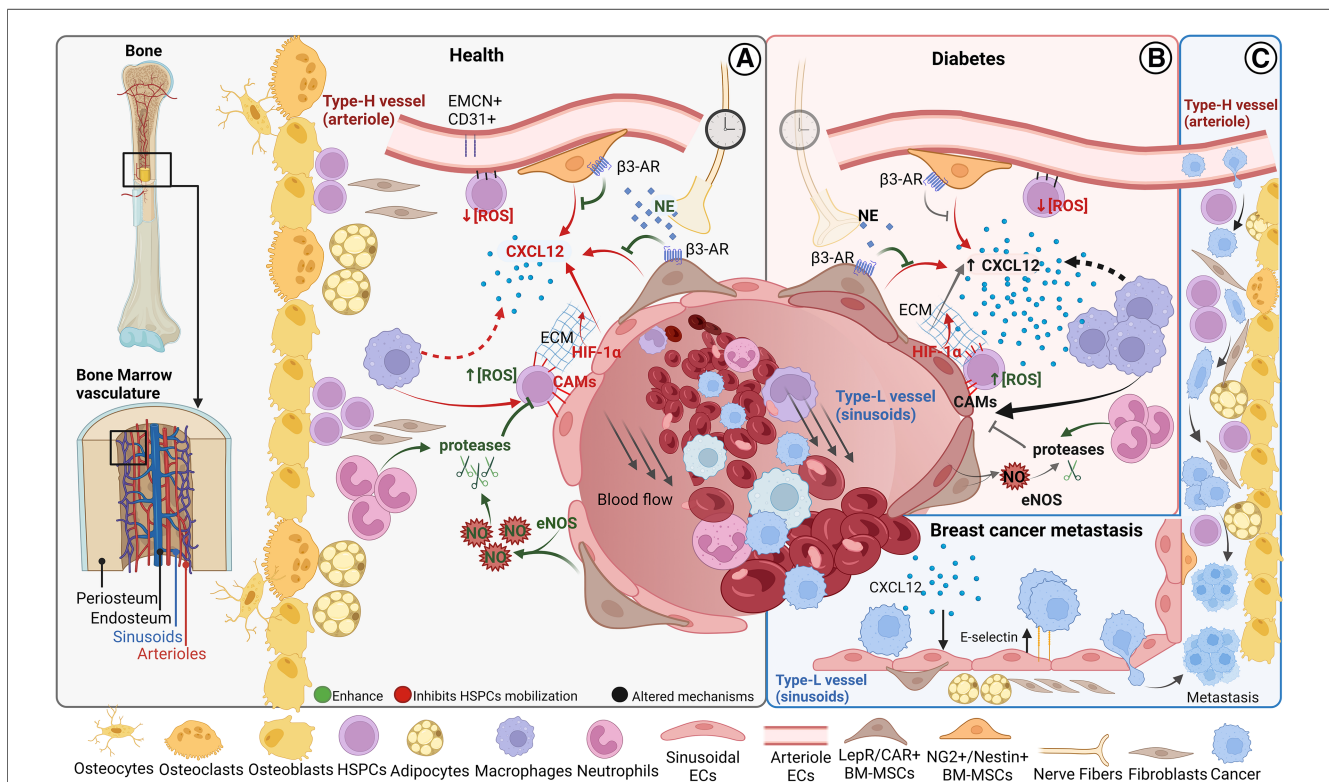


FIGURE 1 Bone marrow cellular composition and main vascular related trafficking mechanisms in homeostasis and disease. Schematic of BM localization and the cellular components (A) BM cell trafficking mechanisms in homeostasis. (B) Altered cell trafficking mechanisms in diabetes (C) Hypothesized steps of cell trafficking in breast cancer bone metastasis formation. BM, bone marrow; HSPCs, hematopoietic stem progenitor cells; CAMs, cell adhesion molecules; CXCL12, CXC chemokine ligand 12; PSCs, perivascular stromal cells; ECs, endothelial cells; NE, norepinephrine; AR, adrenoceptor; NO, nitric oxide. Figure realized with www.biorender.com.

Each niche is characterized by specific oxygen levels, cellular components, and soluble factors that determine a differential modulation of HSPC activity (33, 34). The function of each niche has yet to be fully clarified and the topic remains controversial (33, 35, 36).

It is now well established that the majority of HSPCs localize near the vasculature (37–42), lined by BM endothelial cells (BMECs), but it is still debated whether HSPCs are mostly associated with sinusoids or arterioles (37, 41, 43). Blood vessels are distributed throughout the bone tissue except in cartilaginous areas such as the growth plate. The bone marrow vasculature, mostly consisting of dense and highly branched sinusoidal vessels, provides the barrier between the hematopoietic compartment and the peripheral circulation (15, 44). Sinusoids are also called type L vessels and are mainly located in the diaphysis of long bones, whereas arteriolar type H vessels are characteristic of metaphysis (37, 45, 46). Type H vessels, which are fed directly by arterioles and, therefore, exhibit higher partial pressure of oxygen (pO₂) and blood velocity than type L vessels, are interconnected at their distal ends, near the growth plate, by structures termed loops or arches (44). Sinusoids are larger and fenestrated, resulting in much more permeable vessels compared to arterioles (15). The higher permeability is associated with a higher reactive oxygen species (ROS) state of HSPCs (34) (Figure 1), which mediates HSPC mobilization (47, 48). The work of Itkin et al. demonstrated that HSPC migration toward the bloodstream occurs exclusively at sinusoids (34). Importantly, sinusoidal and columnar vessels are interconnected and form a single vascular network (44).

Type H ECs express higher levels of endothelial markers CD31 (PECAM1) and endomucin compared to type L ECs (45, 46). Type H vessels couple angiogenesis and osteogenesis and regulate bone development in growing bones. Type H vessels are surrounded by a large number of osteoprogenitors and secrete osteogenic factors, such as platelet-derived growth factors (PDGFs), fibroblast growth factor1 (FGF1), and transforming growth factors (TGFs), to support osteoprogenitor cell survival and proliferation, providing a resource of osteoblasts for bone formation. Type H vessels also support vessel-associated osteoclasts through a RANKL–RANK signaling pathway, promoting cartilage resorption and directional bone formation (49).

Beyond the type of vessel and ECs, different perivascular BM-MSCs discriminate the periarterial from perisinusoidal niche. Around arterioles, BM-MSCs are positive to Neuron Glial 2 (NG2) and Nestin whereas perisinusoidal BM-MSCs are leptin receptor (LepR)-expressing CXC chemokine ligand 12 (CXCL12)-abundant reticular (CAR) cells (41, 42). Pericytes are defined as perivascular cells distributed along the microvessel walls and behave as mesenchymal stem cells (MSCs) *in vitro* displaying adipogenic, osteogenic, and chondrogenic potential (50). Given this definition, both LepR/CAR cells and NG2⁺/Nestin⁺ cells could be considered pericyte subtypes (50, 51), even if in some studies pericytes have been confined to smooth muscle α -actin (α -SMA) positive cells (52). Other mature hematopoietic cells, such as megakaryocytes (MKs), macrophages, and neutrophils are also

present near the vasculature and could be considered part of the perivascular niche (Figure 1) (53).

In this review, we will describe the *in vitro* models of bone marrow vasculature developed so far for cardiovascular and cancer research with a particular focus on cell trafficking of angiogenic BM stem cells and cancer cells across the BM vasculature describing the mechanisms involved. Providing details of the *in vitro* approaches used we aim at highlighting the current limits and the comparative benefits.

3. Bone marrow cellular trafficking in cardiovascular and cancer research

3.1. Bone marrow HSPCs mobilization mechanisms

BM CD34⁺ stem cells can support neoangiogenesis restoring blood flow and facilitating inflammatory response. They are physiologically mobilized in response to infections and tissue damage (54, 55) and their release can be pharmacologically stimulated too (56). Granulocyte colony-stimulating factor (G-CSF) is the most used substance for BM stem cell mobilization and has been widely used to study the HSPC mobilization process in animal models (56).

G-CSF mobilizes HSPCs through two principal mechanisms (i) reducing BM levels of the chemoattractant chemokine CXCL12 and (ii) disrupting the HSPCs cell adhesion molecules (CAMs)-dependent attachment to the BM microenvironment.

In the BM, CXCL12 is principally released by perivascular BM-MSCs, in a circadian fashion through the norepinephrine/ β 3-adrenoreceptor signalling pathway (57–59), and ECs (35) (Figure 1). The ECs CXCL12 secretions are required for HSPCs maintenance in the BM niche (35), and hypoxia-inducible factor-1 α (HIF-1 α) may play a role in this mechanism (60, 61) (Figure 1). Also, macrophages have been linked to HSPC retention in the BM through the CXCL12 signalling (62–64) (Figure 1).

HSPCs are anchored to the BM niche through the expression of CAMs (65) (Figure 1). Macrophage depletion induces HSPC mobilization also reducing the expression of CAMs and CAMs ligands (62). Neutrophil-derived matrix metalloproteinase-9 (MMP-9), neutrophil elastase, and cathepsin-G directly cleave some CAMs *in vitro* (65, 66) highlighting the mechanism by which neutrophils induce HSPCs mobilization in response to G-CSF in treated mice (66–68) MMP-9 activity is modulated by levels of nitric oxide, which is produced by endothelial nitric oxide synthase (eNOS) (69). Suppression of eNOS in BM significantly decreased endothelial progenitor cells' release into the bloodstream (70, 71). In BM eNOS is predominantly expressed in panendothelial cell antigen antibody (MECA32) positive cells, suggesting that MECA32⁺ cells' expression of eNOS influences HSPCs mobilization through activation of MMP-9 (71). It is still debated the role of the osteogenic niche in cellular mobilization (35, 57, 59, 61, 62, 64, 65).

3.2. Altered bone marrow HSPCs mobilization in diabetes

Type 2 diabetes mellitus (T2DM) not only results in altered BM structure and stem cell count (72–74) but also, in most investigations reduced CD34⁺ circulating cells were observed in T2DM (11, 72, 75, 76) and type 1 diabetes (77). A meta-regression analysis shows that G-CSF-induced HSPCs mobilization is inhibited in diabetic patients (78). Indeed, diabetes impairs the stem cell mobilization mechanism at different levels. In diabetic mice, G-CSF administration fails to reduce CXCL12 levels (Figure 1). This abnormality in G-CSF regulation of CXCL12 levels is rescued by direct pharmacologic inhibition of CXCL12-CXCR4 signalling, suggesting that diabetic mobilopathy is caused by impairment in CXCL12 levels regulation and not in HSPCs intrinsically (79). Two studies demonstrated that diabetes deregulates stem cell motility through autonomous nervous system impairment (79, 80) (Figure 1). The laboratory of Paolo Madeddu, MD, demonstrated that substance P/neurokinin 1 receptor (NK1R) nociceptive signalling is implied in HSPCs mobilization and that nociceptive neuropathy reflects the decrease in NK1R-HSPCs circulating cells and reduced recruitment of NK1R-HSPCs to the ischemic site in diabetic model (81, 82).

Vessel degeneration observed in diabetes is thought to be largely caused by ECs dysfunction triggered by hyperglycaemic conditions (83). In the context of the BM niche, ECs express less CXCL12 in diabetic animal models (84) (Figure 1). On the other hand, there is an increase in the number of pro-inflammatory CD169⁺ macrophages, that correlated with HSPCs mobilopathy, in the diabetic mice model compared to wild-type (85).

Moreover, diabetes could potentially increase CAMs levels suppressing eNOS activity (86) (Figure 1), indeed a higher number of VCAM1 positive HSPCs have been observed in diabetic mice (87).

In vitro models mimicking BM vasculature-CD34⁺ cell interaction could allow a deeper understanding of the aforementioned mechanisms of cellular mobilization depicting the target mechanisms to enhance it in conditions in which it is impaired.

3.3. Bone marrow cancer metastasis

The BM vascular niche plays a crucial role in the development and progression of hematologic (88, 89), and solid cancer (90). Despite the medical progress in primary tumor treatments, cancer metastasis is the leading cause of cancer-related death. Metastasis is a complex process that involves several stages, including the invasion of surrounding tissue by cancer cells, entry into the bloodstream or lymphatic system, and eventual attachment to the target organ endothelium, where circulating tumor cells (CTCs) may arrest. CTCs that firmly adhere to ECs can form metastases by squeezing through small gaps between ECs and entering the secondary tissue via cell extravasation.

Once in the tissue, disseminated tumor cells (DTCs) may establish a new colony by multiplying and growing, combined with new blood vessel formation to support their growth. Breast cancer is the most common cancer in women worldwide and has a high propensity to form metastasis in the bone. After years of latency, 70%–90% of the patients presented bone lesions (91). Breast CTCs are released from the primary tumor and subtypes of resistant cells reach the BM long before the metastasis formation. Oestrogen receptor (ER), progesterone receptor, and human epidermal growth factor receptor 2 (HER2) triple-positive breast cancer subtypes have a high risk of bone metastasis (92). Recent evidence suggests that the interaction between epithelial to mesenchymal transition (EMT) transcription factors like Zinc Finger E-Box Binding Homeobox 1 (ZEB1) and ER α promote breast cancer bone metastasis (93). The precise mechanisms by which CTCs migrate within the bone marrow are not yet fully understood. It has been hypothesized that breast CTCs migrate across the BM sinusoidal walls, invade and survive in the BM stroma, and then reach the endosteal niche, rich in arterioles type H-vessels, the site for breast cancer bone metastasis (90, 94) (Figure 1). Type H-vessel ECs express chemokines like CXCL12 and adhesion molecule E-selectin that induce DTCs migration, retention, and activation across the type H vessels (95–97) (Figure 1). Breast DTCs remodel the BM niche and type H vessels to support metastasis by releasing factors like G-CSF (98). These events lead to breast bone metastasis, that are mainly osteolytic (bone resorption-lytic), but also osteoblastic (increased bone formation) affecting normal BM function and patient poor outcome. Current therapies are still palliative, and few approved therapies are available to patients. Preclinical studies show that E-selectin inhibitor suppresses tumor cell homing to the bone and outgrowth of micrometastasis (91). Despite these recent findings to unravel breast cancer metastasis formation in the BM, further understanding of all metastatic steps is essential for developing effective therapies to prevent or treat breast cancer bone metastasis. As pointed out earlier for BM stem cell mobilization in diabetes, recent advances in *ex-vivo* microfluidic vascular models in combination with emerging BM vascular models might provide insight into the underlying mechanisms of bone metastasis formation and therapeutic targets for effective clinical treatments.

4. Bone marrow vascular *in vitro* models

4.1. 2D bone marrow *in vitro* models

Traditional 2D cultures and cocultures have been the main systems used in biology, including vascular research. *In vitro* vascular models of BM included both murine and human-derived vascular and BM niche cells cultured under static or dynamic conditions. Many 2D static models of BM vasculature employed non-BM ECs, both in murine and human culture. For example, a rat 2D model of BM was obtained seeding primary

BM-MSCs and primary aortic ECs, on the opposite side of a semipermeable membrane (99).

Murine and human-derived cells were used in combination in hybrid models by coculturing the widely used and characterized primary human vein ECs derived from the umbilical cord (HUVECs), representing the vascular endothelium, with primary mouse BM-derived HSPCs (100), or mouse leukemic initiating cells (LICs) (120). Kobayashi et al. also employed human

primary ECs isolated from the skin and aorta as vascular components of the bone marrow model (100).

In another hybrid 2D model, murine primary BM cells were grown on collagen, then a porous membrane was laid on top, and sealed with a microfluidic channel layer. Here, HUVECs were seeded on a basement membrane extracts coating and grown to form a vessel wall-mimicking monolayer (121) (Figure 2C).

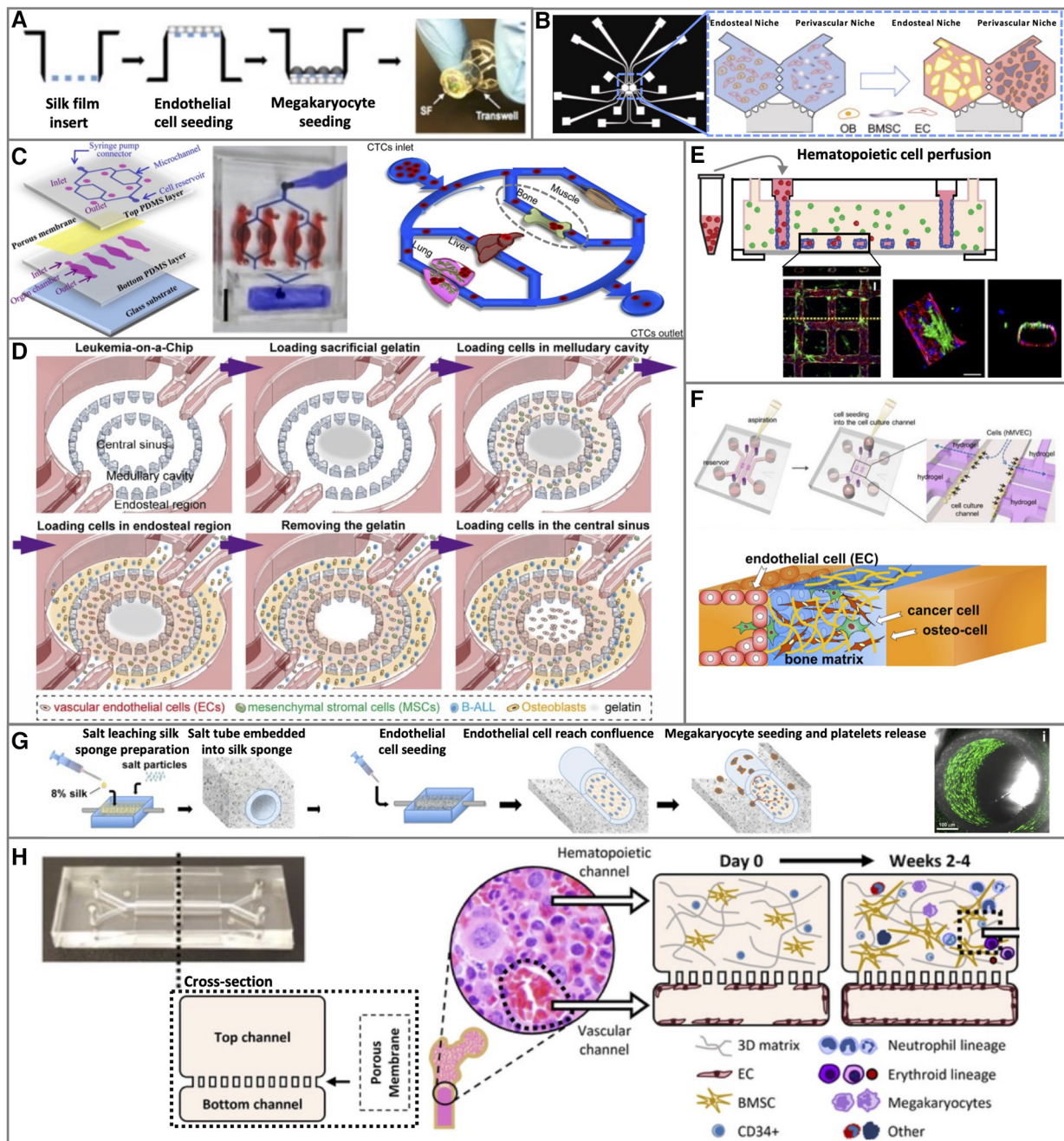


FIGURE 2 Representative 2D and 3D bone marrow vascular *in vitro* models. (A) 2D static, (C) 2D microfluidic, and (B,D–G) 3D BM vascular *in vitro* models. Figure adapted from: (A) Di Buduo et al.; *Blood* (105). (B) Glaser et al.; *Biomaterials* (130). (C) Kong et al.; *Oncotarget* (121). (D) Ma et al.; *Science Advances* (123). (E) Kotha et al.; *Stem Cell Research & Therapy* (134). (F) Bersini et al.; *Biomaterials* (131). (G) Di Buduo et al.; *Blood* (105). (H) Chou et al.; *Nature Biomedical Engineering* (119).

In Rafii's lab, a method to maintain long-term survival and facilitates organ-specific purification of primary ECs was generated, based on the expression of E4ORF1 in serum/cytokine-free conditions (122). E4ORF1-HUVECs were employed to develop the vascular niche of an engineered platform hosting other cells types: human leukemic and embryonal carcinomas cells (122), BM-derived LT-HSCs (101) or hCD34⁺ cord blood (CB) stem and progenitor cells (102).

A coculture of HUVECs with human primary HSPCs was performed in a 50 μm wide microwell allowing one EC and one HSPC to be dispensed per well (103).

HUVECs or primary endothelial colony-forming cells have been used for tri-cultures with primary human BM-MSCs and primary hematopoietic CB-derived CD133⁺ CD34⁺ cells (104).

An improved transwell system has been developed mounting common transwell with silk films, functionalized by surface coating or entrapment of ECM components (collagen, fibrinogen, fibronectin, or laminin). In this model, CB-derived primary ECs and MKs were seeded on opposite sides of the functionalized membrane (105) (Figure 2A).

A higher level of the biological complexity of the 2D BM vascular *in vitro* model was achieved by introducing the primary BMECs isolated from human BM aspirates. These cells were cultured as monolayers with primary HSPCs, both as direct coculture and in transwell (106).

4.2. 3D bone marrow *in vitro* models

The 2D systems are reproducible and easy to use, however, they have limitations in mimicking fundamental *in vivo* BM vascular features and complexities such as cell-cell or cell-matrix interactions, hampering the biological accuracy to model the BM vasculature and its surrounding niche. Recently, the development of 3D models and organ-on-a-chip fabrication allowed the recapitulation of the *in vivo* features of the vasculature more closely.

4.2.1. Murine 3D bone marrow *in vitro* models

BM vascular 3D murine *in vitro* models include both hybrid (murine and human cells) and murine-derived cells (Tables 1, 2). Rat primary BM-MSCs in collagen gel were cocultured with primary rat aortic ECs. The ECs were either bound to dextran-coated Cytodex 3 microcarrier beads embedded in a 3D fibrin gel underneath to allow vascular sprouting (99), or cultured on the surface of collagen gel-BM-MSCs modules grown under shear stress (107). In this model, the empty spaces created by the collagen modules randomly packed were considered perfusable EC-lined channels (107).

Murine tri-culture has been realized by coculturing mouse ECs from the yolk sack (C166 cells) with BM-MSCs and leukemic B-ALL cells in a 3D fibrin hydrogel, in the two inner regions of a 3 concentric rings microfluidic chip. The niche cells, BM-MSCs, and ECs were placed in the middle ring area, while the B-ALL cells were in the central region (123).

A murine tetra-culture was generated in a microfluidic chip with two channels divided by a semi-porous membrane to physically separate the vascular and the BM niche components. The ECs C166 were grown as a monolayer on the lower side of the membrane under fluidic conditions (30 $\mu\text{l/h}$). On the apical side, the BM niche composed of primary BM HSPCs or whole BM cells and primary BM-MSCs in a fibrin-collagen gel was topped with a layer of BM-MSCs-derived osteoblasts (108).

An engineered murine BM vascular *in vitro* model was generated directly *in vivo*, implanting subcutaneously a polydimethylsiloxane (PDMS) device with a central cylindrical cavity functioning as a scaffold, resulting in a bone-like cylindrical disk tissue with a central region of blood-filled marrow. This engineered BM was surgically removed 8 weeks after implantation, punctured in multiple places to allow access to the culture medium, and then cultured in a 5 layers PDMS microfluidic device. Here, a compatible central cylindrical chamber to accommodate the engineered BM was encapsulated in a sandwich-like structure with two porous membranes and microfluidic channels (109).

One of the most important vascular features consists in the hemodynamic forces applied to the vascular walls formed by ECs, therefore advanced vascular models require shear stress to mimic *in vivo* blood flow. Two hybrid human-murine 3D microfluidic BM *in vitro* models that include shear stress on the endothelium were generated. The first is a bioreactor-on-a-chip composed of an upper and a lower microfluidic channel separated by a series of pillars (2 μm apart). The upper channel was injected with a 3D Matrigel or alginate gel embedding murine primary MKs, and a monolayer of primary HUVECs was cultured on fibronectin in the lower channel to mimic the BM vessel wall. In this model, the endothelial monolayer was directly exposed to a controlled flow (12.5 $\mu\text{l/h}$) by syringe pumps (110). In the second one, a 3-channel microfluidic model developed by Jeon and colleagues a flow of 2 $\mu\text{l/min}$ was introduced via the lateral channel to condition the central channel vasculature network flow resulting in wall shear stress of 0.25 dynes/cm² (124).

4.2.2. Human 3D bone marrow *in vitro* models

To increase the complexity of the 3D BM vascular models, a great effort was put into isolating and/or employing human-derived cells both in static and microfluidic conditions (Tables 1, 2). These models mainly combined non-BM-derived primary ECs with BM stromal cells by culturing HUVECs or a few different primary ECs such as ECs from CB or derma as an endothelial component. Therefore, the human BM niche-stromal components were the real BM tissue-specific cells in these models.

A tri-culture of HUVECs mixed with primary CD138⁻ BM-MSCs and patient-derived cancer cells was performed in a fibrinogen scaffold generated from patient BM-derived fibrinogen, naturally found in the plasma of BM supernatant (125).

HUVECs-MSCs coculture has been also used to generate a 3D static and dynamic tetra-culture with osteoblasts differentiated from primary umbilical cord-derived MSCs, and primary CB-derived CD34⁺ HSPCs. Oliveira et al. generated a BM in

liquefied capsules, to facilitate cell movement and self-organization, following three main sequential steps: (i) engineering the endosteal niche by seeding human MSCs with surface-functionalized microparticles, (ii) bioencapsulation of the endosteal niche with HUVECs and HSPCs in alginate, and (iii) production of a multilayered membrane layer-by-layer followed by core liquefaction (111). Another 3D BM static model employed spheres was, generated by the magnetic levitation method, to avoid the use of scaffolds or exogenous matrices and to allow the cells to form the extracellular matrix (108).

Primary ECs from derma, primary BM-MSCs, and primary CD34⁺ HSPCs from CB were combined in a tri-culture, assembled in the 3D Bio Assembler™ System that mixed all cells (112).

A static penta-culture 3D BM model was realized by embedding in Matrigel or alginate hydrogel primary cells: CB-derived HSPCs and EPCs, BM-MSCs, osteoblasts, and BM-MSCs-derived adipocytes (113).

The outstanding research of Palikuqi et al. identified a new method to obtain a durable hemodynamic, self-organizing, large-volume 3D vascular system in a Matrigel-free matrix, composed of a mixture of laminin, entactin, and type-IV collagen (LEC matrix). It is based on the “reset” of vascular endothelial cells (R-VECs) through the transient reactivation of the embryonic-restricted ETS variant transcription factor 2 (ETV2). These findings could lead to a revolution in the organoid field thanks to the ability to vascularize organoids and tumoroids (136). In the context of *in vitro* BM modeling, it would open the possibility of recreating the vascular niche in a complex 3D environment for pharmaceutical studies but also offer the opportunity to develop functional and perfused implantable tissues *ex vivo*.

A vascularized BM organoid based on human induced pluripotent stem cells (iPSCs) was developed in a hydrogel composed of collagen I and IV, and Matrigel. The used medium was supplemented with vascular endothelial growth factor C and the characterization reveals the presence of iPSC-derived ECs, MSC, HSPCs, myelomonocytic, megakaryocytes and immature erythrocytes with human BM cells characteristics. Moreover, this human BM organoid were engrafted with different types of cells to mimic human diseases; specifically: iPSC-derived HSPCs derived from patients with myelofibrosis or healthy individuals, primary cancer cells from patients affected by multiple myeloma, acute lymphoblastic leukemia (ALL), and myeloid leukemias (126).

The aforementioned ring shape microfluidic chip (123), used to culture murine cells, was also employed in an all-human fibrin hydrogel-based 3D model. In this case, HUVECs were seeded in the central sinus region, a mixture of HUVECs, primary BM-MSCs, and leukemia cancer cells in the inner ring, and a mixture of osteoblasts (hFOB cells) and leukemic cancer cells leukemia in the outer ring channel (123) (Figure 2D).

A 3D hydroxyapatite porous scaffold placed in, and, connected to a perfusion bioreactor was used to coculture adipose tissue-derived stromal vascular fraction, which includes primary mature ECs, endothelial progenitor cells, pericytes, and MSCs, with osteo-differentiated primary BM-MSCs. This engineered

vascularized BM niche was perfused at the rate of 300 µl/min with a suspension of CD34⁺ CB-HSPCs (114).

4.2.3. Human 3D bone marrow self-assembled vascular networks *in vitro* models

To mimic the *in vivo* vascular structure, 3D human BM *in vitro* models were developed based on self-assembled vascular networks (Tables 1, 2). HUVECs have been cocultured in tri-cultures with primary BM-MSCs and cancer cells in a hydrogel drop of approximately 200–300 Pa stiffness, optimal for the development of a robust endothelial network (127).

Microfluidic human BM 3D *in vitro* models are characterized by controlled and/or induced flow for nutrient exchange in all-human cell cultures. 3D vascular models made of capillary-like structure networks were formed by HUVECs cocultured with primary BM-MSCs. The mesenchymal cells adopted pericyte-like localizations among the network system, both in a 3D decellularized bone matrix (128) and in fibrin matrixes within 3D microfluidic chips with different designs and geometries (115–117).

Maturano-Kruik et al. developed a capillary-like structures network model seeding the ECs and MSCs within a 3D decellularized bone tissue's trabecular scaffold that after a maintenance phase, was placed in the microfluidic chip and exposed to a controlled flow (128).

The microfluidic chips of Jeon et al. (115) and Mykuliak et al. (117) were designed with a central gel channel, for HUVEC-MSC coculture, between two lateral channels for culture medium, separated by pillars. In this last HUVEC-MSC coculture model, the shear stress on ECs was exerted by interstitial flow and it was estimated to be 0.0065 Pa (117).

Carrion et al. used a 3D penta-channel microfluidic chip with three central parallel gel channels and two lateral channels for medium supply. The central gel chamber was filled with fibrin gel only, and the two adjacent channels with fibrin-embedded ECs and MSCs respectively (116).

Capillary-like structures were also obtained by culturing HUVECs in a 3D gel in the absence of perivascular MSCs (103, 129). In one case, cells were cultured in a microfluidic chip with three parallel microchannels, partitioned by trapezoid-shaped pillars. The central channel was filled with collagen gel, and one lateral channel with ECs monolayer, grown on the pillar-collagen interface of the central channel to provide the space for angiogenic sprouting and neo-vessel formation. The other lateral channel was seeded with leukemia cancer cells followed by injection in the channel reservoirs of BM-MSCs (HS5 cells) in collagen gel (129).

In a second work, the microfluidic chip was composed of two lateral media channels and three main parallel gel channels (103). The two lateral channels mimic the BM vascular niche and the endosteal niche, hosting the vascular network generated by HUVECs and hFOB osteoblasts in collagen fibrin gel, respectively. The central channel was loaded with primary HSPCs in collagen fibrin gel. Another lateral channel, separated by the medium channel from the vascular compartment, holds mitomycin C-treated fibroblasts (NHLF cells) and was used as a cytokine-secreting compartment (103).

The BM perivascular and endosteal niches were also simultaneously reproduced in a microfluidic device with two hexagonal chambers connected by three symmetric two-way ports that allow nutrient diffusion and cell migration. Primary CB-derived ECs were cultured in both chambers in fibrin hydrogel to form a vascular network, mixed either with hFOB 1.19 osteoblasts to form the endosteal niche or primary BM-MSCs to form the perivascular niche on-a-chip (130) (Figure 2B).

The biological complexity of a 3D BM *in vitro* model can be enhanced by adding other cell types in culture, to better mimic the tissue microenvironment. Other models characterized by the formation of a BM vascular network and its niche were realized in 3D microfluidic devices with parallel 3- or 5-channels. In the central channel, HUVECs were cultured with primary BM-MSCs, osteo-differentiated primary BM-MSCs, and primary HSPCs (118) or with primary BM-MSCs and macrophages (115) in ECM gel. Hence, this type of model allows the mixing of the vascular and the endosteal niches' cellular components. In the central channel of the 3-channel microfluidics device, a microvascular network with highly branched structures was grown in 3D fibrin, supported by medium from the two lateral channels (115). In the central channel of the 5-channel microfluidic system BM-M-SCs were cultured on a mix of collagen and dopamine-HCl to differentiate in osteoblasts as a monolayer, then a fibrin-collagen hydrogel with a mixture of endothelial, mesenchymal and hematopoietic cells was injected on top of the osteoblasts to obtain a 3D human BM-on-a-chip. The two side channels serve for medium supply, and the two external channels were filled with BM-MSCs in fibrin-collagen hydrogel to provide further support to the BM niche (118).

4.2.4. Human 3D bone marrow endothelial cell-lined *in vitro* models

To mimic the vascular geometry and endothelial distribution *in vivo*, 3D vascular microfluidic models were designed to grow ECs on the channel walls to generate an endothelial monolayer creating a lumen.

A microfluidic device with three parallel central channels and two lateral channels for medium supply was designed to create a lumen (137). In this device, a HUVEC monolayer was cultured on Matrigel in the central channel to form a vascular wall-like geometry lumen with a squared section where medium and circulating cells were flown through (131). In the side channel, in contact with the endothelial central channel, osteo-differentiated primary BM-MSCs were cultured in 3D collagen to generate an osteo-conditioned microenvironment (131) (Figure 2F).

A 3D 1-channel BM microfluidic model, part of a body-on-a-chip, consists of primary HSPCs embedded in a 3D fibrin gel and a HUVECs monolayer placed on opposite site of a porous membrane. Here the endothelial monolayer was directly exposed to controlled shear stress (0.2 dynes/cm²) (132).

A similar microfluidic model included also the primary BM-MSCs with primary HSPCs embedded in fibrin in the top compartment, and a HUVEC monolayer grown under shear

stress (1.2–1.6 µl/min) as a vascular compartment in the bottom (119) (Figure 2H).

The BM endosteal niche was partially recapitulated in a 3D microfluidic chip, as part of a vascularized multi-organ tissue system (133). Here, the bone tissue was obtained by coculturing both BM-MSCs differentiated osteoblasts and primary CD14⁺ monocytes-derived osteoclasts within a decellularized bone scaffold on top. To mimic blood vessel walls, the porous membrane was seeded with pericytes-mimicking BM-MSCs on the upper surface and with HUVECs mixed to a small fraction of BM-MSCs on the lower side grown under shear stress (1.88 dynes/cm² max) (133).

A 3D microfluidic vascular network that mimics the vasculature geometry was created by growing HUVECs on a collagen gel grid characterized by a series of squared blocks design, to allow the formation of a rounded-shaped monolayer under gravity-mediated flow. In the collagen gel, the BM components were embedded as primary BM-MSC or BM fibroblasts cell lines (134) (Figure 2E).

A tubular-shaped vascular microfluidic device made with a silk microtube surrounded by a porous silk sponge was seeded with primary CB- or derma-derived ECs in the silk microtube lumen and exposed to shear stress. Then, primary HSPCs-derived human MKs were seeded into the silk sponge (105) (Figure 2G).







Finally, the 3D BM vascular *in vitro* model, which better mimics the *in vivo* BM vasculature was generated using organ-specific primary BMECs, isolated via magnetic separation from frozen human BM-MNCs. Arterial and sinusoidal ECs were isolated via double selection for CD146⁺/NG2⁺ or CD146⁺/NG2^{lo/-}, respectively (135). These cells were grown to replicate the major 3D BM niches in a deconstructed way in a microfluidic chip designed with 4 non-communicating parallel channels that can be exposed to control flow, where the four different niche cell populations were encapsulated in ECM-derived hydrogel: (I) arteriolar BMECs, (II) sinusoidal BMECs, (III) primary BM-MSCs and (IV) osteoblasts derived from osteo-differentiated primary BM-MSCs (135).

In the next sections, we describe how the reported models have been employed for either cardiovascular or cancer research.

4.3. Bone marrow vascular models used for cardiovascular research

Modeling the BM vasculature is essential to study inside-out cell mobilization and for the comprehension of mechanisms regulating this process important in both cancer and cardiovascular research. Many laboratories have focused on developing *in vitro* models of the BM vasculature that can be used for cardiovascular studies. These, range from cultures that allow HSPCs maintenance *in vitro* to study hematopoiesis (108, 109, 111–113, 118, 119), to devices to investigate cell-cell interaction (103, 107, 114, 115) and vasculogenesis (99, 115–117). Advanced systems recapitulate key features of BM damage (109, 118, 119) or diseases (119). Other interesting research has developed devices for platelet production *in vitro* that can be employed for clinical purposes (105, 110).

TABLE 1 Bone marrow vascular *in vitro* models for cardiovascular-related research.

Species	<i>In vitro</i> model features					Cell type				Topic	Ref	
	Human 	ECM	Static	Microfluidic	Shear stress	Endothelial 	Mesenchymal stem/stromal 	Hematopoietic/progenitor 	Osteoblasts 			Monocytes/Macrophages Adipocytes Fibroblasts 
CARDIOVASCULAR-RELATED												
2D BONE MARROW <i>in vitro</i> MODELS												
Rat	2D	-	✓		NO	Primary aortic ECs	Primary BM-MSCs				Angiogenesis	(99)
Mouse & Human (hybrid)	2D	Coating	✓		NO	HUVECs or HMECs or HAECs Gelatin		Primary HSPCs (BM)			Mechanisms of ECs-HSPCs interaction	(100)
Mouse & Human (hybrid)	2D	-	✓		NO	E4ORF1+ HUVECs		Murine primary LT-HSPCs (BM)			LT-HSCs expansion	(101)
Human	2D	-	✓		NO	E4ORF1+ HUVECs		Primary HSPCs (cord blood)			HSPCs expansion	(102)
Human	2D	Coating	✓		NO	HUVECs Fibronectin		Primary HSPCs (cord blood)			HSPC polarization	(103)
Human	2D	-	✓		NO	HUVECs or ECFCs (cord blood)	Primary BM-MSCs	Primary HSPCs (cord blood)			HSPC expansion <i>in vitro</i> ; cell-cell interaction	(104)
Human	2D	ECM-coated silk films	✓		NO	EPCs (cord blood)				Primary MKs	Platelet production	(105)
Human	2D	Coating	✓		NO	HUVECs or BMECs Gelatin	Primary BM-MSCs	Primary HSPCs (cord blood or peripheral blood)			HSPCs proliferation/differentiation	(106)
3D BONE MARROW <i>in vitro</i> MODELS												
murine 3D <i>in vitro</i> models												
Rat	3D	FIBRIN COLLAGEN	✓		NO	Primary aortic ECs	Primary BM-MSCs				Angiogenesis	(99)
Rat	3D	COLLAGEN		✓	YES	Primary aortic ECs	Primary BM-MSCs				Cell-cell interaction	(107)
Mouse	3D	FIBRIN-COLLAGEN		✓	YES	C166 cell line Yolk sack	Primary BM-MSCs	Primary HSPCs (BM)			Maintenance of HSPCs	(108)
Mouse	3D	Tissue explant		✓	NO		Primary bone marrow cells (BMCs)				Model of BM damage	(109)
Mouse & human (hybrid)	3D	ALGINATE or MATRIGEL		✓	YES	HUVECs Fibronectin				Primary MK (mouse) or hiPSC derived MKs	Platelet production	(110)

(Continued)

TABLE 1 Continued















Species	In vitro model features				Shear stress	Cell type				Topic	Ref
	Human 	ECM	Static	Microfluidic		Endothelial 	Mesenchymal stem/stromal 	Hematopoietic/progenitor 	Osteoblasts 		
human 3D in vitro models											
Human	3D	ALGINATE	✓		DYNAMIC	HUVECs		Primary HSPCs (cord blood)	Osteo-diff Primary BM-MSCs	Maintenance of HSPCs, cell-cell interaction	(111)
Human	3D	ECM cell- produced	✓		NO	HDMECs	Primary HSPCs (cord blood)	Primary HSPCs (cord blood)		Maintenance of HSCs	(112)
Human	2D	ALGINATE or MATRIGEL	✓		NO	EPCs (cord blood)		Primary HSPCs (cord blood)	Osteoblast-diff primary BM-MSCs	Maintenance of HSPCs	(113)
Human	2D	ECM coated silk films	✓		NO	EPCs (cord blood)			CD61 ⁺ MKs	Maintenance of HSPCs	(114)
human 3D self-assembled vascular networks in vitro model											
Human	3D	FIBRIN	✓		NO	HUVECs	Primary BM-MSCs			Angiogenesis	(115)
Human	3D	FIBRIN	✓		NO	HUVECs	Primary BM-MSCs			Angiogenesis	(116)
Human	3D	FIBRIN	✓		YES	HUVECs	Primary BM-MSCs			Angiogenesis	(117)
Human	3D	FIBRIN-COLLAGEN	✓		NO	HUVECs		Primary HSPCs (CORD BLOOD)	hFOB cell line	HSPCs polarity	(103)
Human	3D	FIBRIN-COLLAGEN	✓		NO	HUVECs	Primary BM-MSCs	Primary HSPCs (BM)	Osteoblast-diff primary BM-MSCs	Maintenance of HSPCs; model of BM damage	(118)
human 3D endothelial cell-lined in vitro models											
Human	3D	FIBRIN	✓		YES	HUVECs Fibronectin Collagen	Primary BM-MSCs	Primary HSPCs (peripheral blood)		Model of BM damage and diseases	(119)
Human	3D	FIBRIN-COLLAGEN-LAMININ	✓		YES	EPCs (cord blood) or HMECV-d			Primary MKs	Platelet generation	(105)

TABLE 2 Bone marrow vascular *in vitro* models for cancer research.

Species	<i>In vitro</i> model features					Shear stress	Cell type	Disease	Ref
	Human 	ECM	Static	Microfluidic	Endothelial				
Murine 									
Cancer									
2D Bone MARROW in vitro MODELS									
Mouse & Human (hybrid)	2D	-		✓	HUVECs	NO		Leukemia (AML)	(120)
Rat & Human (hybrid)	2D	Coating		✓	HUVECs Cultrex	YES	Primary BM cells Collagen	Breast triple neg	(121)
Human	2D	-		✓	EAORF1+ HUVECs	NO		Leukemia (APL) and embryonic carcinoma	(122)
3D BONE MARROW in vitro MODELS									
murine 3D in vitro models									
Mouse	3D	FIBRIN		✓	OP9 cell line (BM)	NO		Leukemia (ALL)	(123)
Mouse & human (hybrid)	3D	FIBRIN		✓	HUVECs	YES	Primary BM- MSCs	Brest triple negative (macrophages)	(124)
human 3D in vitro models									
Human	3D	FIBRIN		✓	HUVECs	NO	Primary BM- MSCs	Multiple myeloma	(125)
Human	3D	COLLAGEN- MATRIGEL		✓	iPSC-derived ECs	NO	iPSC-derived MSCs	Myelofibrosis, multiple myeloma, leukemia (ALL)	(126)
Human	3D	FIBRIN		✓	HUVECs	NO	Primary BM- MSCs	Leukemia (ALL)	(123)
human 3D self-assembled vascular networks in vitro models									
Human	3D	StarPEG- HEPARIN HYDROGEL		✓	HUVECs	NO	Primary BM- MSCs	Leukemia (AML)	(127)
Human	3D	BONE MATRIX		✓	HUVECs	YES	Primary BM- MSCs	Breast triple neg	(128)
Human	3D	COLLAGEN		✓	HUVECs	NO	HS5 cell line (BM)	Leukemia (AML-CML)	(129)
Human	3D	FIBRIN		✓	ECFCs (cord blood)	NO	Primary BM- MSCs	Breast triple neg	(130)

(Continued)

TABLE 2 Continued

Species	In vitro model features						Cell type				Disease	Ref	
	Murine 	Human 	ECM	Static	Microfluidic	Shear stress	Endothelial 	Mesenchymal stem/stromal 	Hematopoietic/progenitor 	Osteoclasts/Osteoblasts 			Monocytes/macrophages 
human 3D endothelial cell-lined in vitro models													
Human		3D	COLLAGEN		✓	NO	HUVECs Matrigel			Osteoblast-differentiated BM-MSCs		Breast triple neg	(131)
Human		3D	FIBRIN and COLLAGEN		✓	YES	HUVECs Collagen and Fibronectin		Primary HSPCs CD34 ⁺ (cord blood)			Cancer drugs	(132)
Human		3D	BONE MATRIX		✓	YES	HUVECs Fibronectin	Primary BM-MSCs		Osteoblast-differentiated BM-MSCs		Cancer drugs	(133)
Human		3D	COLLAGEN		✓	YES	HUVECs	H95 or HS27a cell lines (BM)			Primary monocytes (peripheral blood)	Leukemia (AML)	(134)
Human		3D	HYALURONIC ACID-GELATIN		✓	NO	Primary BMAECs and BMSECs	Primary BM-MSCs		Osteoblast-differentiated primary BM-MSCs		Leukemia (AML)	(135)

Two different approaches, involving mouse primary cell growth in a BM-rich microenvironment, were able to maintain the proportions and characteristics of HSPCs for up to 7 and 14 days, respectively (108, 109). However, to finely study human hematopoiesis human cells are required (138, 139). Two unconventional methods were adopted to generate spherical-shaped human BM cocultures (111, 112). Organotypic multicellular spheres developed through magnetic levitation in the absence of scaffold and exogenous matrices stimulate cells to produce their extracellular matrix. This work opens the door to the realization of more complex organoids applicable in the field of regenerative medicine (112). The cellular release of ECM components and HSPCs supportive factors was observed also in the other spherical-shaped culture, the “human bone marrow-in-a-liquefied-capsule” when placed under dynamic fluid (111).

Braham et al., developed a fully primary human cell 3D coculture useful to understand the interaction of HSPCs with other BM cells and the ECM in normoxic, hypoxic, and hyperoxic conditions. They show that Matrigel is a better substrate for coculturing HSPCs with osteo-vascular components compared to alginate (113). The interaction of HSPCs with the BM microenvironment is crucial for *in vitro* long-term culture maintenance and in deciding their fate (140). A human 3D microfluidic device was able to recapitulate the *in vivo* observed HSPCs polarization (141) proving to be a great model to study HSPC polarization mechanisms, which are still unknown (103).

Butler et al., demonstrate the fundamental role of ECs in cells expansion, in the E4ORF1-ECs based vascular niche engineered platform ECs resulted to enhance proliferation and maintenance of human leukemic and embryonal carcinomas cells (122), BM-derived LT-HSCs (101) and cord blood-derived hCD34⁺ stem and progenitor cells (102) in serum/cytokine-free conditions.

Born and colleagues demonstrate that a self-assembled vascular structure not only maintains the HSPCs in their undifferentiated state but also preserved the osteogenic potential of BM-MSCs (114). The influence of ECs on BM-MSCs was evaluated also in a rat-based 3D dynamic system demonstrating that the combination of shear stress with the vascular component induces BM-MSC differentiation in α -SMA⁺ pericytes and stimulates their migration through ECs (107). This model is of great interest as the presence of BM-perivascular cells is fundamental to mimicking BM vasculature. A similar study conducted with human primary ECs, and BM-MSCs, cultured in 3D microfluidic condition (115) confirms the transition of MSCs toward α -SMA⁺ mural cells when cocultured with ECs and that this transition is further enhanced by the presence of angiogenic factors such as angiopoietin-1 and TGF- β . Interestingly, the α -SMA⁺ mural cells colocalize with ECs suggesting a contact-dependent mechanism to induce BM-MSCs differentiation. These last studies prove the important support of ECs on BM-MSCs fate.

Other research has instead focused on the influence of BM-MSCs on ECs and the vasculogenesis process. Jeon et al., demonstrate that the vascular component developed a higher number of network branches and display a reduced vessel diameter in the presence of BM-MSCs (115). Rat BM-MSCs embedded in a 3D microenvironment enhanced sprouting and

proliferation of ECs through paracrine signals (99). A human-based microfluidic chip with cells embedded in fibrin gel confirmed the capability of BM-MSCs to sustain ECs vascular network formation. Moreover, they have shown that BM-MSCs promote basement membrane deposition by ECs and that the perivascular localization of BM-MSCs occurs through the interaction of their integrin adhesion receptor α 6 β 1 and basement membrane laminin (116). Analyzing the fibroblast effects on EC microvasculature they observed that fibroblasts enhance vascular network formation even faster than BM-MSCs (116). The interaction of ECs with MSCs derived from BM or adipose tissue was compared in a human cell-based 3D microfluidic model. Both types of MSCs support network formation, but the BM-MSCs resulted in a more organized, interconnected, and denser ECs network and in higher expression of genes characteristic of pericyte and ECs compared to the coculture of ECs with adipose-derived MSCs (117).

To develop effective drugs to recover BM damage or treat BM diseases it is important to mimic the BM alteration characteristics of the pathology. The mouse cell-based model of Torisawa et al. faithfully recapitulates BM damage observed in live irradiated mice and the BM recovery after administration of G-CSF, a largely used drug to treat victims of radiation accidents (109, 142). In a 3D human osteovascular niche on-a-chip the endosteal component demonstrates a protective activity on HSPCs reducing their apoptosis following ionizing radiation exposure (118).

Chou and colleagues developed an advanced human microfluidic BM-on-a-chip suitable to study drug and irradiation toxicity as well as BM recovery. In this chip, able to maintain myeloerythroid proliferation and differentiation, under dynamic condition was recapitulated the reduction in neutrophils and total cell count induced by the chemotherapeutic agent 5-fluorouracil; and also neutrophils and erythroid cytotoxicity induced by AZD2811, a potential cancer drug currently in phase II of clinical development (119). The hypothesis that AZD2811 selectively targets dividing neutrophil and erythroid precursors was assessed inside this microfluidic chip. Moreover, the observed differences in cytotoxicity between low and high levels of γ -radiation resemble human radiation sensitivity. Interestingly, the damaged cells can recover inside the model, an important characteristic for pharmaceutical research (119).

Finally, HSPCs derived from Shwachman–Diamond syndrome (SDS) patients were cocultured with normal BM-MSCs and ECs. SDS is a rare genetic disorder resulting in bone marrow failure, characterized by neutropenia and other BM-cell defects. The SDS-on-a-chip faithfully reproduces the disease, opening new frontiers for translational, drug development, and cytotoxicity studies (119).

One of the problems of the clinical practice that is trying to be solved by these new *in vitro* technologies is represented by platelet transfusion. Nowadays, all transfusion platelets are obtained by human donors and can cause immunogenic reactions in the receiving patient. Two laboratories realized two different bioreactors able to reproduce thrombopoiesis, in both cases the presence of vascular shear stress enhanced platelet collection

(105, 110). Thon et al., utilized human induced pluripotent stem cells (iPSCs)-derived MKs that could potentially produce an unlimited number of functional human platelets and overcome the immunogenicity problem (110). The model of Buduo et al., is of great interest because they designed a functionalized silk-based BM system that could replace the endothelial layer, required to promote thrombopoiesis *in vitro*, facilitating the scaling-up for future massive platelet production (105).

4.4. Bone marrow vascular models used for cancer research

The BM is the semi-solid tissue where hematologic cancers such as leukemia and myelomas originate (143, 144). The BM is also one of the most common sites for solid tumor metastasis as breast and prostate cancer (145) (Figure 1). 2D and 3D BM *in vitro* vascular models have been employed to study both liquid or hematologic cancers and solid cancer metastasis.

4.4.1. Hematologic cancers

Leukaemia and myeloma begin during hematopoiesis with cell genetic alterations. Most leukaemia involves white blood cells, and some affect other types of blood cells. When these cells over-proliferate, they crowd out normal BM cells with consequent hematopoiesis alterations leading to the development of the disease. Leukemic cells leave the bone marrow niche through the BM vasculature to enter the bloodstream, where they can spread to other organs. Leukaemia is classified based on the speed of progression, that which could be acute or chronic, and on the type of blood cell involved, myeloid or lymphoid (146).

Acute leukaemias (AL) are characterized by immature blood cells with high intrinsic proliferative potential that leads to a fast progression of the disease. Altered immature blood cells with a myeloid commitment, cells able to differentiate into MKs, red or non-lymphocyte white blood cells during hematopoiesis in health, cause acute myeloid leukemia (AML), which is the most common type of acute leukemia in adults (147).

In leukemic patients it has been observed that an increased BM vascularization is a key feature, however, this event has not been largely studied for solid tumor progression and metastasis (148). Angiogenesis was studied with a 3D BM microfluidic chip showing that ECs (HUVECs) sprouting and neo-vessel formation were increased by the presence of AML cells (HL60 cells) compared to control (EC medium). In the presence of HS5BM-SCs, to mimic the BM niche, angiogenesis was more pronounced, and, in coculture with AML cells, it was further enhanced, but characterized by a larger number of isolated endothelial tips without multicellular stalks, reflecting what was found in AML patient biopsies. ECs migrated for longer distances in the collagen matrix, with a higher number of endothelial tips but lower vascularization were found using chronic myeloid leukemia (CML) cells (K562 cells) in the absence of stromal cells (129). CML originates from partially mature myeloid white blood cells and is characterized by slower progression compared to AML. In turn, ECs play an important

role in CML. Coculture of HUVECs with a murine AML cell line in a 2D model showed that ECs support the expansion of rare leukemic cells, known as LICs representing a subset of self-renewing cells able to generate an aggressive AML (120). This was also shown in the 3D BM vascular model developed by Bray et al., where both AML cell lines (KG1a, MOLM13, MV4-11, OCI-AML3) and primary patient-derived AML cells exerted preference to adhere and proliferate along the endothelial network (HUVECs-BM-SCs), further highlighting the importance of the interaction between AML cells and the vascular niche. In accordance with these findings, this model enabled further research on drug resistance, one of the main challenges in clinical treatment. It has been observed that the 3D tri-culture model was more resistant to two antineoplastic drugs commonly used to treat AML compared to simpler 2D and 3D monocultures (AML cells only) (127). In a perfusable 3D microfluidic vascular model with tubular HUVECs vessels cocultured with stromal BM fibroblast cell lines (HS27a or HS5), patient-derived primary AML cells adhesion and extravasation were tested under shear stress in combination with primary monocytes. Interestingly, AML cells did not exhibit preferential adhesion or extravasation in the presence of either stromal cell type, in contrast, leukemic cells without monocytes adhered and extravasated more to and across vessels cocultured with HS27a when compared to HS5 cells. These findings underline the importance of complex 3D models that mimic *in vivo* multicellular components (134) (Figure 2E).

To explore the AML cells-BM niche components specific interaction, a deconstructed microfluidic BM niches on-a-chip (NOC) with arterial (BMAECs), mesenchymal (BM-SCs), sinusoidal (BMSECs) and osteoblastic niches was perfused with AML cells (MOLM13). AML cells were preferentially lodged within the osteoblastic and the arterial niches rather than into the mesenchymal or sinusoidal niches (135). Acute lymphoblastic leukaemia (ALL) is another form of acute leukaemia and the most common cancer in childhood, characterized by the overproduction of immature and dysfunctional lymphoblast in the BM (149). A ring-shaped leukaemia-on-a-chip 3D BM model (Figure 2D) that resembles the *in vivo* spatial architecture and cellular composition of the leukaemia BM tissue was developed and adopted to study the heterogeneity in B-ALL human BM microenvironments based on their chemo resistance. It has been found that Ph⁺ SUP B-ALL cells cocultured with BM niche cells in the biomimetic device were more resistant to conventional chemotherapeutic than REH B-ALL, consistent with the insensitivity of Ph⁺ B-ALL to these types of drugs in the clinic. Furthermore, the leukemia-on-a-chip 3D BM model was designed to rescue the niche cells for downstream assay like single-cell RNAseq, a very powerful tool to characterize the BM microenvironment with limited cell input number. Finally, using a simpler ring-shaped murine 3D BM model with ECs and BM-MSCs it was observed that B-ALL cells attracted ECs. Niche cells promoted B-ALL cell cluster formation and reduced their motility over time (123).

The second most common hematologic malignancy is the MM, where clonal terminally differentiated B lymphocytes undergo abnormal proliferation followed by accumulation in the BM

crowding out healthy blood cells. In MM abnormal monoclonal antibodies, paraproteins, are produced causing overly thick blood and kidney issues (150). To investigate the pathophysiology of MM, a 3D tissue-engineered BM (3DTEBM) derived from MM patients-BM supernatant that incorporated the BM components MM (MM1s, H929, RPMI8226 or primary), stromal, and ECs was established. It was observed that the presence of both stromal and ECs increased MM cell proliferation compared to single culture or coculture using other commercially available 3D systems, thanks to the 3DTEBM scaffold with fibrinogen cross-linking (125).

The classical feature of BM myelofibrosis consist in the deposition of reticulin and collagen fibers by marrow stroma (151) and the iPSC-based organoid model was able to recapitulate them when the donor cells derived from patients with myelofibrosis but not when derived from healthy donor: increased soluble TGF β levels, collagen 1 and α -SMA and reduced vascularization was observed (126).

4.4.2. Solid cancer

Solid tumors such as breast cancer can spread from the primary site intravasating in the bloodstream to form secondary tumors at distant sites called metastasis. Metastasis is a multistep process where circulating tumor cells (CTCs) interact with ECs, forming the microvascular vessel wall, adhering and extravasate extravasating into the target organ to form metastasis (Figure 1). These key events of metastasis formation take place under hemodynamic shear stress exerted on ECs by blood flow (152). Triple-negative breast cancer (negative for oestrogen receptor, progesterone receptor, and HER2) is a highly invasive type of breast cancer, resistant to both hormonal therapy and treatments against HER2 (Trastuzumab), that develops metastasis in the bone with high frequency (153). Numerous BM vascular models *in vitro* were developed to study metastasis formation by human triple negative breast cancer cells (MDA-MB-231) (121, 124, 128, 130, 131). It has been shown that CTCs firm adhesion to ECs (HUVECs) under shear stress was higher when HUVECs were cocultured with BM niche specialized cells compared to HUVECs cocultured with muscle cells, resulting in cancer cells organotropism for bone vasculature (121) (Figure 2C). In a model of self-assembled capillary-like network (HUVECs), cancer cell extravasation in the matrix increased in the presence of BM cells (BM-MSCs and osteoblasts). In contrast, it was observed a decreased extravasation in a shear stress-preconditioned vascular network or in the presence of inflammatory macrophages (inflammation model) within the BM niche (124). The extravasation step was further investigated in a microfluidic model with HUVECs forming a vascular lumen. MDA-MB-231 cells transmigrated across the HUVECs monolayer into the osteo-differentiated BM-MSC gel and travelled within the matrix more compared to the control. Extravasated cancer cells were able to proliferate and form micrometastasis of various sizes, ranging from 4 cells to 60 cells or more (131). A 3D BM vascular model was employed to investigate cancer cell migration to BM-specific niches. An increased MDA-MB-231 cell migration to the BM niches compared to the control was observed, without differences between perivascular and endosteal niches. The migrated

MDA-MB-231 cells expressed more Ki-67, a proliferation marker, than non-migrated cells, consistent with high Ki-67 expression in bone lesions (130) (Figure 2B). All these 3D BM vascular models showed how the BM niche is important for the breast cancer cell metastatic process and enabled to highlight the cancer cell organotropism. Finally, a 3D model with a bone matrix scaffold, to maintain the ECs viability and branching without the need of for specialized growth-promoting conditions culture medium, was developed to study metastatic colonization of MDA-MB-231 cells under static and interstitial flow conditions. It was reported that the interstitial flow promoted vascular branching and prevented cancer cell growth rate within the niche (128). The BM vasculature-on-a-chip models were also employed for drug toxicity studies. Two models were realized as part of a body on a chip (132, 133) to test the uptake and side effects of common chemotherapeutic drugs given to treat several cancers such as cisplatin and doxorubicin. The perfusable vasculatures were challenged in the luminal side when in contact with the apical bone marrow niche. Cisplatin was flowed through the vascular channel causing myeloid toxicity in the HSPCs BM niche, reducing the total number of cells in the HSPCs BM niche, in particular neutrophils and erythrocytes, recapitulating the known side effects of cisplatin in patients (neutropenia and anaemia) *in vitro* (132). In a vascularized bone model which includes a lot of components in common with the BM endosteal niche, osteoblasts were found to be more sensitive than osteoclasts when exposed to doxorubicin, as observed in pre-clinical studies, and, as expected, the endothelium showed decreased resistance (133).

5. Conclusions and future directions

Despite animal models, such as the broadly used murine models, are a powerful tool for genetic and physiological studies, they do not always mirror human biology and human medical disease conditions. About 8% is the rate of successful translation of drugs from animal testing to human treatments (20). Thus, there is an unmet urgent need to develop physiologically relevant humanized models for hematopoietic and cardiovascular disorders. *In vitro* modeling has become a major focus of research in the field of vascular and cancer research in recent years, as scientists strive to understand the cellular and molecular mechanisms in human cells essential for the development of effective preventive and treatment strategies without the need for animal or human testing. Moreover, the advent of new strategies that enable a punctual characterization of the BM can not be ignored. Single-cell RNA sequencing has been used to identify and describe the characteristics of BM cells deriving from human and mouse (154–156) providing an exceptional tool for the validation of *in vitro* models especially in the context of disease modeling.

In vitro models can be used to simulate hemodynamic shear stress, an important component in cellular trafficking, providing insight into the disease mechanisms and how these might be prevented or treated. Furthermore, *in vitro* models can be used to investigate the effects of drugs or other treatments on the pathophysiology of specific organs. In this review, we

summarized both murine and human 2D and 3D bone marrow vascular *in vitro* models with their cellular composition and experimental function that may be suitable to study *in vitro* cellular mobilization and homing in the BM via its vasculature, a central process in cardiovascular diseases and cancer. From this compilation emerged that these *in vitro* systems aimed to mimic at best the complexity of the BM environment. However, behind including ECs as vascular components, other BM cell types are required to establish a more *in vivo*-like and reliable model as different cell types release important factors orchestrating cellular fate, as explained in the first paragraphs of this review. Moreover, to study human disease and its mechanisms like cell trafficking, it is essential to implement the *in vitro* models technologies with organ-specific human-derived cells. Thus, to recapitulate the BM environment in an *in vitro* model it is essential to include: (i) a monolayer of ECs with a fluid flow applied on it to mimic the blood flow; (ii) the presence of perivascular cells surrounding the ECs such as pericytes, and the immune components (macrophages and/or neutrophils); (iii) all components need to be human and derived from the BM or that display BM-specific characteristics. Among the thirty-three literature works considered in this review, none include all these characteristics together. In the work of Jeon et al. (124) the shear stress was applied to the ECs layer and the model included almost all the cellular components necessary to replicate the vascular niche, however, the ECs were not BM-derived and the immune component originates from mice. As Jeon et al., all the 2D and 3D human BM vasculature *in vitro* models described, except for the works of Rafii et al. (106) and Aleman et al. (135), hold ECs derived from other vascular districts. Therefore, there is a need for a more standardized protocol for human bone marrow endothelial and perivascular cell isolation, or iPS-derived cell differentiated in BM cells to improve these advanced *in vitro* models of BM vasculature. Success in the combination of these advanced *in vitro* BM vasculature models with human/patients-derived cells will allow the study of physiological and pathological mechanisms that can be relevant to pre-clinical studies. Furthermore, these new generation of *in vitro* models can be used to investigate the effects of drugs or other treatments on cellular trafficking at the BM vasculature, paving the way for more effective treatments for cancer and cardiovascular diseases.

References

- Ahmad FB, Anderson RN. The leading causes of death in the US for 2020. *JAMA*. (2021) 325:1829–30. doi: 10.1001/jama.2021.5469
- Santos JV, Souza J, Valente J, Alonso V, Ramalho A, Viana J, et al. The state of health in the European Union (EU-28) in 2017: an analysis of the burden of diseases and injuries. *Eur J Public Health*. (2020) 30:573–8. doi: 10.1093/eurpub/ckz203
- OECD/European Union. *Health at a Glance: Europe 2018: State of Health in the EU Cycle*. Paris/European Union, Brussels: OECD Publishing (2018). Available from: https://doi.org/10.1787/health_glance_eur-2018-en
- Dal Canto E, Ceriello A, Rydén L, Ferrini M, Hansen TB, Schnell O, et al. Diabetes as a cardiovascular risk factor: an overview of global trends of macro and micro vascular complications. *Eur J Prev Cardiol*. (2019) 26:25–32. doi: 10.1177/2047487319878371
- Sun H, Saeedi P, Karuranga S, Pinkepank M, Ogurtsova K, Duncan BB, et al. IDF Diabetes atlas: global, regional and country-level diabetes prevalence estimates for 2021 and projections for 2045. *Diabetes Res Clin Pract*. (2022) 183:109119. doi: 10.1016/j.diabres.2021.109119
- Guan X. Cancer metastases: challenges and opportunities. *Acta Pharm Sin B*. (2015) 5:402–18. doi: 10.1016/j.apsb.2015.07.005
- Wojakowski W, Tendera M, Michalowska A, Majka M, Kucia M, Maślankiewicz K, et al. Mobilization of CD34/CXCR4 +, CD34/CD117 +, c-met+stem cells, and mononuclear cells expressing early cardiac, muscle, and endothelial markers into peripheral blood in patients with acute myocardial infarction. *Circulation*. (2004) 110:3213–20. doi: 10.1161/01.CIR.0000147609.39780.02
- DiPersio JF. Diabetic stem-cell “mobilopathy”. *N Engl J Med*. (2011) 365:2536–8. doi: 10.1056/NEJMcibr112347
- Ling L, Shen Y, Wang K, Jiang C, Fang C, Ferro A, et al. Worse clinical outcomes in acute myocardial infarction patients with type 2 diabetes mellitus: relevance to impaired endothelial progenitor cells mobilization. *PLoS One*. (2012) 7:1–10. doi: 10.1371/journal.pone.0050739

Author contributions

MC: Writing – original draft, Writing – review & editing. LB: Writing – original draft. CC: Conceptualization, Writing – original draft, Writing – review & editing. GS: Conceptualization, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article.

MC and GS are supported by the Italian Ministry of Health (Ricerca Corrente and 5xmille to the IRCCS MultiMedica). CC is the recipient of the International Cancer Research Fellowship ICARE-2 funded from by the European Union’s Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 800924 (“Single-cell epigenetic and molecular signatures in human breast cancer metastasis formation”). The funders had no role in the design of the review.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of *Frontiers*, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher’s note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

10. Rigato M, Bittante C, Albiero M, Avogaro A, Fadini GP. Circulating progenitor cell count predicts microvascular outcomes in type 2 diabetic patients. *J Clin Endocrinol Metab.* (2015) 100:2666–72. doi: 10.1210/jc.2015-1687
11. Fadini GP, Rigato M, Cappellari R, Bonora BM, Avogaro A. Long-term prediction of cardiovascular outcomes by circulating CD34⁺ and CD34⁺CD133⁺ stem cells in patients with type 2 diabetes. *Diabetes Care.* (2017) 40:125–31. doi: 10.2337/dc16-1755
12. Fadini GP, de Kreutzenberg S, Agostini C, Boscaro E, Tiengo A, Dimmeler S, et al. Low CD34⁺ cell count and metabolic syndrome synergistically increase the risk of adverse outcomes. *Atherosclerosis.* (2009) 207:213–9. doi: 10.1016/j.atherosclerosis.2009.03.040
13. Schmidt-Lucke C, Rössig L, Fichtlscherer S, Vasa M, Britten M, Kämper U, et al. Reduced number of circulating endothelial progenitor cells predicts future cardiovascular events: proof of concept for the clinical importance of endogenous vascular repair. *Circulation.* (2005) 111:2981–7. doi: 10.1161/CIRCULATIONAHA.104.504340
14. Nguyen DX, Bos PD, Massagué J. Metastasis: from dissemination to organ-specific colonization. *Nat Rev Cancer.* (2009) 9(4):274–84. doi: 10.1038/nrc2622
15. Kopp HG, Aveilla ST, Hooper AT, Rafii S. The bone marrow vascular niche: home of HSC differentiation and mobilization. *Physiology* (2005) 20:349–56. doi: 10.1152/physiol.00025.2005
16. Peng Y, Wu S, Li Y, Crane JL. Type H blood vessels in bone modeling and remodeling. *Theranostics.* (2020) 10:426–36. doi: 10.7150/thno.34126
17. Frank S, Carton M, Dubot C, Campone M, Pistilli B, Dalenc F, et al. Impact of age at diagnosis of metastatic breast cancer on overall survival in the real-life ESME metastatic breast cancer cohort. *Breast.* (2020) 52:50–7. doi: 10.1016/j.breast.2020.04.009
18. Rohde D, Vandoorne K, Lee I-H, Grune J, Zhang S, McAlpine CS, et al. Bone marrow endothelial dysfunction promotes myeloid cell expansion in cardiovascular disease. *Nat Cardiovasc Res.* (2022) 1(1):28–44. doi: 10.1038/s44161-021-00002-8
19. Swirski FK, Nahrendorf M. Bone marrow takes center stage in cardiovascular disease. *Circ Res.* (2016) 119:701–3. doi: 10.1161/CIRCRESAHA.116.309584
20. Marshall LJ, Bailey J, Cassotta M, Herrmann K, Pistollato F. Poor translatability of biomedical research using animals — a narrative review. *Altern Lab Anim.* (2023) 51:102–35. doi: 10.1177/02611929231157756
21. Biswas L, Chen J, De Angelis J, Singh A, Owen-Woods C, Ding Z, et al. Lymphatic vessels in bone support regeneration after injury. *Cell.* (2023) 186:382–397.e24. doi: 10.1016/j.cell.2022.12.031
22. Wang LD, Wagers AJ. Dynamic niches in the origination and differentiation of haematopoietic stem cells. *Nature Reviews Molecular Cell Biology.* (2011) 12(10):643–55. doi: 10.1038/nrm3184
23. Riether C, Schurch C, Ochsenschein AF. Regulation of hematopoietic and leukemic stem cells by the immune system. *Cell Death Differ.* (2015) 22:187–98. doi: 10.1038/cdd.2014.89
24. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini FC, Krause DS, et al. Minimal criteria for defining multipotent mesenchymal stromal cells: The international society for cellular therapy position statement. *Cytotherapy.* (2006) 8:315–7. doi: 10.1080/14653240600855905
25. Mohamed-Ahmed S, Fristad I, Lie SA, Suliman S, Mustafa K, Vindenes H, et al. Adipose-derived and bone marrow mesenchymal stem cells: a donor-matched comparison. *Stem Cell Res Ther.* (2018) 9:1–15. doi: 10.1186/s13287-018-0914-1
26. Uccelli A, Moretta L, Pistoia V. Mesenchymal stem cells in health and disease. *Nat Rev Immunol.* (2008) 8(9):726–36. doi: 10.1038/nri2395
27. Levi B, Longaker MT. Mechanisms of bone development and repair. *Nat Rev Mol Cell Biol.* (2020) 21(11):696–711. doi: 10.1038/s41580-020-00279-w
28. Piotrowska K, Tarnowski M. Bone marrow adipocytes—role in physiology and various nutritional conditions in human and animal models. *Nutrients.* (2021) 13:1412. doi: 10.3390/nu13051412
29. Nombela-Arrieta C, Manz MG. Quantification and three-dimensional microanatomical organization of the bone marrow. *Blood Adv.* (2017) 1:407–16. doi: 10.1182/bloodadvances.2016003194
30. Oh M, Nör JE. The perivascular niche and self-renewal of stem cells. *Front Physiol.* (2015) 6:367. doi: 10.3389/fphys.2015.00367
31. Kiel MJ, Morrison SJ. Uncertainty in the niches that maintain haematopoietic stem cells. *Nat Rev Immunol.* (2008) 8:290–301. doi: 10.1038/nri2279
32. Morrison SJ, Scadden DT. The bone marrow niche for haematopoietic stem cells. *Nature.* (2014) 505:327–34. doi: 10.1038/nature12984
33. Zhao M, Li L. Dissecting the bone marrow HSC niches. *Cell Res.* (2016) 26(9):975–6. doi: 10.1038/cr.2016.71
34. Itkin T, Gur-Cohen S, Spencer JA, Schajnovitz A, Ramasamy SK, Kusumbe AP, et al. Distinct bone marrow blood vessels differentially regulate haematopoiesis. *Nature.* (2016) 532:323–8. doi: 10.1038/nature17624
35. Ding L, Morrison SJ. Haematopoietic stem cells and early lymphoid progenitors occupy distinct bone marrow niches. *Nature.* (2013) 495:213–35. doi: 10.1038/nature11885
36. Zhang J, Niu C, Ye L, Huang H, He X, Tong W-G, et al. Identification of the haematopoietic stem cell niche and control of the niche size. *Nature.* (2003) 425:836–41. doi: 10.1038/nature02041
37. Acar M, Kocherlakota KS, Murphy MM, Peyer JG, Oguro H, Inra CN, et al. Deep imaging of bone marrow shows non-dividing stem cells are mainly perisinusoidal. *Nature.* (2015) 526:126–30. doi: 10.1038/nature15250
38. Kokkaliaris KD, Kunz L, Cabezas-Wallscheid N, Christodoulou C, Renders S, Camargo F, et al. Adult blood stem cell localization reflects the abundance of reported bone marrow niche cell types and their combinations. *Blood.* (2020) 136:2296–307. doi: 10.1182/blood.2020066574
39. Kiel MJ, Yilmaz ÖH, Iwashita T, Yilmaz OH, Terhorst C, Morrison SJ. SLAM Family receptors distinguish hematopoietic stem and progenitor cells and reveal endothelial niches for stem cells. *Cell.* (2005) 121:1109–21. doi: 10.1016/j.cell.2005.05.026
40. Kiel MJ, Radice GL, Morrison SJ. Lack of evidence that hematopoietic stem cells depend on N-cadherin-mediated adhesion to osteoblasts for their maintenance. *Cell Stem Cell.* (2007) 1:204–17. doi: 10.1016/j.stem.2007.06.001
41. Kunisaki Y, Bruns I, Scheiermann C, Ahmed J, Pinho S, Zhang D, et al. Arteriolar niches maintain haematopoietic stem cell quiescence. *Nature.* (2013) 502:637–43. doi: 10.1038/nature12612
42. Sugiyama T, Kohara H, Noda M, Nagasawa T. Maintenance of the hematopoietic stem cell pool by CXCL12-CXCR4 chemokine signaling in bone marrow stromal cell niches. *Immunity.* (2006) 25:977–88. doi: 10.1016/j.immuni.2006.10.016
43. Méndez-Ferrer S, Michurina TV, Ferraro F, Mazloom AR, MacArthur BD, Lira SA, et al. Mesenchymal and hematopoietic stem cells form a unique bone marrow niche. *Nature.* (2010) 466:829. doi: 10.1038/nature09262
44. Chen J, Hendriks M, Chatzis A, Ramasamy SK, Kusumbe AP. Bone vasculature and bone marrow vascular niches in health and disease. *J Bone Miner Res.* (2020) 35:2103–20. doi: 10.1002/jbmr.4171
45. Mosteo L, Storer J, Batta K, Searle EJ, Duarte D, Wiseman DH. The dynamic interface between the bone marrow vascular niche and hematopoietic stem cells in myeloid malignancy. *Front Cell Dev Biol.* (2021) 9:1–24. doi: 10.3389/fcell.2021.635189
46. Kusumbe AP, Ramasamy SK, Adams RH. Coupling of angiogenesis and osteogenesis by a specific vessel subtype in bone. *Nature.* (2014) 507:323–8. doi: 10.1038/nature13145
47. Tesio M, Golan K, Corso S, Giordano S, Schajnovitz A, Vagima Y, et al. Enhanced c-met activity promotes G-CSF-induced mobilization of hematopoietic progenitor cells via ROS signaling. *Blood.* (2011) 117:419–28. doi: 10.1182/blood-2009-06-230359
48. Golan K, Vagima Y, Ludin A, Itkin T, Cohen-Gur S, Kalinkovich A, et al. S1p promotes murine progenitor cell egress and mobilization via S1P1-mediated ROS signaling and SDF-1 release. *Blood.* (2012) 119:2478–88. doi: 10.1182/blood-2011-06-358614
49. Owen-woods C, Kusumbe A. Fundamentals of bone vasculature: specialization, interactions and functions. *Semin Cell Dev Biol.* (2022) 123:36–47. doi: 10.1016/j.semcdb.2021.06.025
50. Attwell D, Mishra A, Hall CN, O'Farrell FM, Dalkara T. What is a pericyte? *J Cereb Blood Flow Metab.* (2016) 36:451–5. doi: 10.1177/0271678X15610340
51. Kunisaki Y. Pericyte biology in different organs. *Adv Exp Med Biol.* (2019) 1122:101–14. doi: 10.1007/978-3-030-11093-2_6
52. Baryawno N, Przybylski D, Kowalczyk MS, Kfoury Y, Severe N, Gustafsson K, et al. A cellular taxonomy of the bone marrow stroma in homeostasis and leukemia. *Cell.* (2019) 177:1915–1932.e16. doi: 10.1016/j.cell.2019.04.040
53. Bruns I, Lucas D, Pinho S, Ahmed J, Lambert MP, Kunisaki Y, et al. Megakaryocytes regulate hematopoietic stem cell quiescence through CXCL4 secretion. *Nat Med.* (2014) 20:1315–20. doi: 10.1038/nm.3707
54. Skirecki T, Mikaszewska-Sokolewicz M, Godlewska M, Dołęgowska B, Czubak J, Hoser G, et al. Mobilization of stem and progenitor cells in septic shock patients. *Sci Rep.* (2019) 9:3289. doi: 10.1038/s41598-019-39772-4
55. Liao R, Pfister O, Jain M, Mouquet F. The bone marrow—cardiac axis of myocardial regeneration. *Prog Cardiovasc Dis.* (2007) 50:18–30. doi: 10.1016/j.pcad.2007.03.001
56. D'Amario D, Leone AM, Borovac JA, Cannata F, Siracusanò A, Niccoli G, et al. Granulocyte colony-stimulating factor for the treatment of cardiovascular diseases: an update with a critical appraisal. *Pharmacol Res.* (2018) 127:67–76. doi: 10.1016/j.phrs.2017.06.001
57. Katayama Y, Battista M, Kao WM, Hidalgo A, Peired AJ, Thomas SA, et al. Signals from the sympathetic nervous system regulate hematopoietic stem cell egress from bone marrow. *Cell.* (2006) 124:407–21. doi: 10.1016/j.cell.2005.10.041
58. García-García A, Korn C, García-Fernández M, Domingues O, Villadiego J, Martín-Pérez D, et al. Dual cholinergic signals regulate daily migration of

hematopoietic stem cells and leukocytes. *Blood*. (2019) 133:224–36. doi: 10.1182/blood-2018-08-867648

59. Méndez-Ferrer S, Lucas D, Battista M, Frenette PS. Haematopoietic stem cell release is regulated by circadian oscillations. *Nature*. (2008) 452:442–7. doi: 10.1038/nature06685
60. Ceradini DJ, Kulkarni AR, Callaghan MJ, Tepper OM, Bastidas N, Kleinman ME, et al. Progenitor cell trafficking is regulated by hypoxic gradients through HIF-1 induction of SDF-1. *Nat Med*. (2004) 10:858–64. doi: 10.1038/nm1075
61. Hoggatt J, Pelus LM. Mobilization of hematopoietic stem cells from the bone marrow niche to the blood compartment. *Stem Cell Research and Therapy*. (2011) 2(2):13. doi: 10.1186/scrt54
62. Winkler IG, Sims NA, Pettit AR, Barbier V, Nowlan B, Helwani F, et al. Bone marrow macrophages maintain hematopoietic stem cell (HSC) niches and their depletion mobilizes HSCs. *Blood*. (2010) 116:4815–28. doi: 10.1182/blood-2009-11-253534
63. Chow A, Lucas D, Hidalgo A, Méndez-Ferrer S, Hashimoto D, Scheiermann C, et al. Bone marrow CD169⁺ macrophages promote the retention of hematopoietic stem and progenitor cells in the mesenchymal stem cell niche. *J Exp Med*. (2011) 208:761–71. doi: 10.1084/jem.20101688
64. Christopher MJ, Rao M, Liu F, Woloszynek JR, Link DC. Expression of the G-CSF receptor in monocytic cells is sufficient to mediate hematopoietic progenitor mobilization by G-CSF in mice. *J Exp Med*. (2011) 208:251–60. doi: 10.1084/jem.20101700
65. Kulkarni R, Kale V. Physiological cues involved in the regulation of adhesion mechanisms in hematopoietic stem cell fate decision. *Front Cell Dev Biol*. (2020) 8:611. doi: 10.3389/fcell.2020.00611
66. Lévesque JP, Hendy J, Takamatsu Y, Williams B, Winkler IG, Simmons PJ. Mobilization by either cyclophosphamide or granulocyte colony-stimulating factor transforms the bone marrow into a highly proteolytic environment. *Exp Hematol*. (2002) 30:440–9. doi: 10.1016/S0301-472X(02)00788-9
67. Pelus LM, Bian H, King AG, Fukuda S. Neutrophil-derived MMP-9 mediates synergistic mobilization of hematopoietic stem and progenitor cells by the combination of G-CSF and the chemokines GRO β /CXCL2 and GRO β T/CXCL2 Δ 4. *Blood*. (2004) 103:110–9. doi: 10.1182/blood-2003-04-1115
68. Lévesque J, Hendy J, Takamatsu Y, Simmons PJ, Bendall LJ. Disruption of the CXCR4/CXCL12 chemotactic interaction during hematopoietic stem cell mobilization induced by G-CSF or cyclophosphamide. *J Clin Invest*. (2003) 111:187–96. doi: 10.1172/JCI15994
69. Ridnour LA, Windhausen AN, Isenberg JS, Yeung N, Thomas DD, Vitek MP, et al. Nitric oxide regulates matrix metalloproteinase-9 activity by guanylyl-cyclase-dependent and -independent pathways. *Proc Natl Acad Sci U S A*. (2007) 104:16898–903. doi: 10.1073/pnas.0702761104
70. Kazakov A, Müller P, Jagoda P, Semenov A, Böhm M, Laufs U. Endothelial nitric oxide synthase of the bone marrow regulates myocardial hypertrophy, fibrosis, and angiogenesis. *Cardiovasc Res*. (2012) 93:397–405. doi: 10.1093/cvr/cvr305
71. Aicher A, Heeschen C, Mildner-Rihm C, Urbich C, Ihling C, Technau-Ihling K, et al. Essential role of endothelial nitric oxide synthase for mobilization of stem and progenitor cells. *Nat Med*. (2003) 9:1370–6. doi: 10.1038/nm948
72. Fadini GP, Boscaro E, De Kreutzenberg S, Agostini C, Seeger F, Dimmeler S, et al. Time course and mechanisms of circulating progenitor cell reduction in the natural history of type 2 diabetes. *Diabetes Care*. (2010) 33:1097–102. doi: 10.2337/dc09-1999
73. Mangialardi G, Ferland-McCollough D, Maselli D, Santopaolo M, Cordaro A, Spinetti G, et al. Bone marrow pericyte dysfunction in individuals with type 2 diabetes. *Diabetologia*. (2019) 62(7):1275–90. doi: 10.1007/s00125-019-4865-6
74. Spinetti G, Cordella D, Fortunato O, Sangalli E, Losa S, Gotti A, et al. Global remodeling of the vascular stem cell niche in bone marrow of diabetic patients: implication of the microRNA-155/FOXO3a signaling pathway. *Circ Res*. (2013) 112:510–22. doi: 10.1161/CIRCRESAHA.112.300598
75. Terenzi DC, Al-Omran M, Quan A, Teoh H, Verma S, Hess DA. Circulating pro-vascular progenitor cell depletion during type 2 diabetes. *JACC Basic Transl Sci*. (2019) 4:98–112. doi: 10.1016/j.jacbs.2018.10.005
76. Egan CG, Lavery R, Caporali F, Fondelli C, Laghi-Pasini F, Dotta F, et al. Generalised reduction of putative endothelial progenitors and CXCR4-positive peripheral blood cells in type 2 diabetes. *Diabetologia*. (2008) 51:1296–305. doi: 10.1007/s00125-008-0939-6
77. Boscaro F, D'Anna M, Bonora BM, Tresso S, Cappellari R, Avogaro A, et al. Effects of glucose variability on hematopoietic stem/progenitor cells in patients with type 1 diabetes. *J Endocrinol Invest*. (2021) 44:119–26. doi: 10.1007/s40618-020-01278-6
78. Fadini GP, Avogaro A. Diabetes impairs mobilization of stem cells for the treatment of cardiovascular disease: a meta-regression analysis. *Int J Cardiol*. (2013) 168:892–7. doi: 10.1016/j.ijcard.2012.10.089
79. Ferraro F, Lymperi S, Méndez-Ferrer S, Saez B, Spencer JA, Yeap BY, et al. Diabetes impairs hematopoietic stem cell mobilization by altering niche function. *Sci Transl Med*. (2011) 3:104ra101. doi: 10.1126/scitranslmed.3002191
80. Busik JV, Tikhonenko M, Bhatwadekar A, Opreanu M, Yakubova N, Caballero S, et al. Diabetic retinopathy is associated with bone marrow neuropathy and a depressed peripheral clock. *J Exp Med*. (2009) 206:2897–906. doi: 10.1084/jem.20090889
81. Amadesi S, Reni C, Katare R, Meloni M, Oikawa A, Beltrami AP, et al. Role for substance P-based nociceptive signaling in progenitor cell activation and angiogenesis during ischemia in mice and in human subjects. *Circulation*. (2012) 125:1774–86. doi: 10.1161/CIRCULATIONAHA.111.089763
82. Dang Z, Maselli D, Spinetti G, Sangalli E, Carnelli F, Rosa F, et al. Sensory neuropathy hampers nociception-mediated bone marrow stem cell release in mice and patients with diabetes. *Diabetologia*. (2015) 58:2653–62. doi: 10.1007/s00125-015-3735-0
83. Paneni F, Beckman JA, Creager MA, Cosentino F. Diabetes and vascular disease: pathophysiology, clinical consequences, and medical therapy: part i. *Eur Heart J*. (2013) 34:2436–46. doi: 10.1093/eurheartj/ehf149
84. Hoyer FF, Zhang X, Coppin E, Vasamsetti SB, Modugu G, Schloss MJ, et al. Bone marrow endothelial cells regulate myelopoiesis in diabetes Mellitus. *Circulation*. (2020) 142:244–58. doi: 10.1161/CIRCULATIONAHA.120.046038
85. Albiero M, Poncina N, Ciciliot S, Cappellari R, Menegazzo L, Ferraro F, et al. Bone marrow macrophages contribute to diabetic stem cell mobilopathy by producing oncostatin M. *Diabetes*. (2015) 64:2957–68. doi: 10.2337/db14-1473
86. Gallagher KA, Liu ZJ, Xiao M, Chen H, Goldstein LJ, Buerk DG, et al. Diabetic impairments in NO-mediated endothelial progenitor cell mobilization and homing are reversed by hyperoxia and SDF-1 α . *J Clin Invest*. (2007) 117:1249–59. doi: 10.1172/JCI29710
87. Katagi M, Terashima T, Ohashi N, Nakae Y, Yamada A, Nakagawa T, et al. Malfunctioning CD106-positive, short-term hematopoietic stem cells trigger diabetic neuropathy in mice by cell fusion. *Commun Biol*. (2021) 4:1–14. doi: 10.1038/s42003-021-02082-5
88. Batsivari A, Luydmila M, Haltalli R, Passaro D, Pospori C, Lo Celso C, et al. Cell niche to diverse stresses. *Nat Cell Biol*. (2020) 22:7–17. doi: 10.1038/s41556-019-0444-9
89. Méndez-Ferrer S, Bonnet D, Steensma DP, Hasserjian RP, Ghoobrial IM, Gribben JG, et al. Bone marrow niches in haematological malignancies. *Nat Rev Cancer*. (2020) 20:285–98. doi: 10.1038/s41568-020-0245-2
90. Satcher RL, Zhang XHF. Evolving cancer–niche interactions and therapeutic targets during bone metastasis. *Nat Rev Cancer*. (2022) 22(2):85–101. doi: 10.1038/s41568-021-00406-5
91. Hofbauer LC, Bozec A, Rauner M, Jakob F, Perner S, Pantel K. Novel approaches to target the microenvironment of bone metastasis. *Nat Rev Clin Oncol*. (2021) 18:488–505. doi: 10.1038/s41571-021-00499-9
92. Xiao W, Zheng S, Yang A, Zhang X, Zou Y, Tang H, et al. Breast cancer subtypes and the risk of distant metastasis at initial diagnosis: a population-based study. *Cancer Manag Res*. (2018) 10:2147/CMAR.S176763
93. Mohammadi Ghahhari N, Sznurkowska MK, Hulo N, Bernasconi L, Aceto N, Picard D. Cooperative interaction between ER α and the EMT-inducer ZEB1 reprograms breast cancer cells for bone metastasis. *Nat Commun*. (2022) 13:2104. doi: 10.1038/s41467-022-29723-5
94. Suva LJ, Washam C, Nicholas RW, Griffin RJ. Bone metastasis: mechanisms and therapeutic opportunities. *Nature Publishing Group*. (2011) 7(4):208–18. doi: 10.1038/nrendo.2010.227
95. Esposito M, Mondal N, Greco TM, Wei Y, Spadazzi C, Lin S, et al. Mesenchymal–epithelial transition and wnt metastasis. *Nat Cell Biol*. (2019) 21:627–39. doi: 10.1038/s41556-019-0309-2
96. Carrascal A, Silva M, Sackstein R, Pascoal C, Serrano I, Jos M, Videira PA. Inhibition of fucosylation in human invasive ductal carcinoma reduces E-selectin ligand expression, cell proliferation, and ERK1/2 and p38 MAPK activation. *Mol Oncol*. (2018) 12:579–93. doi: 10.1002/1878-0261.12163
97. Price TT, Burness ML, Sivan A, Warner MJ, Cheng R, Lee CH, et al. Dormant breast cancer micrometastases reside in specific bone marrow niches that regulate their transit to and from bone. *Sci Transl Med*. (2016) 8:1–23. doi: 10.1126/scitranslmed.aad4059
98. Yip RKH, Rimes JS, Capaldo BD, Vaillant F, Mouchemore KA, Pal B, et al. Mammary tumour cells remodel the bone marrow vascular microenvironment to support metastasis. *Nat Commun*. (2021) 12(1):6920. doi: 10.1038/s41467-021-26556-6
99. Chamberlain MD, Gupta R, Sefton MV. Bone marrow-derived mesenchymal stromal cells enhance chimeric vessel development driven by endothelial cell-coated microtissues. *Tissue Eng Part A*. (2012) 18:285–94. doi: 10.1089/ten.tea.2011.0393
100. Kobayashi H, Butler JM, O'Donnell R, Kobayashi M, Sen Ding B, Bonner B, et al. Angiocrine factors from akt-activated endothelial cells balance self-renewal and differentiation of haematopoietic stem cells. *Nat Cell Biol*. (2010) 12:1046–56. doi: 10.1038/ncb2108
101. Butler JM, Nolan DJ, Vertes EL, Varnum-Finney B, Kobayashi H, Hooper AT, et al. Endothelial cells are essential for the self-renewal and repopulation of notch-dependent hematopoietic stem cells. *Cell Stem Cell*. (2010) 6:251–64. doi: 10.1016/j.stem.2010.02.001

102. Butler JM, Gars EJ, James DJ, Nolan DJ, Scandura JM, Rafi S. Development of a vascular niche platform for expansion of repopulating human cord blood stem and progenitor cells. *Blood*. (2012) 120:1344–7. doi: 10.1182/blood-2011-12-398115
103. Bessy T, Candelas A, Souquet B, Saadallah K, Schaeffer A, Vianay B, et al. Hematopoietic progenitors polarize in contact with bone marrow stromal cells in response to sdf1. *J Cell Biol*. (2021) 220(11):e202005085. doi: 10.1083/jcb.202005085
104. Radtke S, Görgens A, da Conceição Castro SV, Kordelas L, Königer A, Dürig J, et al. Human multipotent hematopoietic progenitor cell expansion is neither supported in endothelial and endothelial/mesenchymal co-cultures nor in NSG mice. *Sci Rep*. (2019) 9:1–12. doi: 10.1038/s41598-019-49221-x
105. Di Buduo CA, Wray LS, Tozzi L, Malara A, Chen Y, Ghezzi CE, et al. Programmable 3D silk bone marrow niche for platelet generation ex vivo and modeling of megakaryopoiesis pathologies. *Blood*. (2015) 125:2254–64. doi: 10.1182/blood-2014-08-595561
106. Rafi S, Shapiro F, Pettengell R, Ferris B, Nachman R, Moore M, et al. Human bone marrow microvascular endothelial cells support long-term proliferation and differentiation of myeloid and megakaryocytic progenitors. *Blood*. (1995) 86:3353–63. doi: 10.1182/blood.V86.9.3353.bloodjournal8693353
107. Khan OF, Chamberlain MD, Sefton M V. Toward an in vitro vasculature: differentiation of mesenchymal stromal cells within an endothelial cell-seeded modular construct in a microfluidic flow chamber. *Tissue Eng Part A*. (2012) 18:744–56. doi: 10.1089/ten.tea.2011.0058
108. Sharipol A, Lesch ML, Soto CA, Frisch BJ. Bone marrow microenvironment-on-chip for culture of functional hematopoietic stem cells. *Front Bioeng Biotechnol*. (2022) 10:1–15. doi: 10.3389/fbioe.2022.855777
109. Torisawa YS, Spina CS, Mammoto T, Mammoto A, Weaver JC, Tat T, et al. Bone marrow-on-a-chip replicates hematopoietic niche physiology in vitro. *Nat Methods*. (2014) 11:663–9. doi: 10.1038/nmeth.2938
110. Thon JN, Mazutis L, Wu S, Sylman JL, Ehrlicher A, Machlus KR, et al. Platelet bioreactor-on-a-chip. *Blood*. (2014) 124:1857–67. doi: 10.1182/blood-2014-05-574913
111. Oliveira CS, Nadine S, Gomes MC, Correia CR, Mano JF. Bioengineering the human bone marrow microenvironment in liquefied compartments: a promising approach for the recapitulation of osteovascular niches. *Acta Biomater*. (2022) 149:167–78. doi: 10.1016/j.actbio.2022.07.001
112. Mejía-Cruz CC, Barreto-Durán E, Pardo-Pérez MA, Jimenez MC, Rincón J, Vanegas K, et al. Generation of organotypic multicellular spheres by magnetic levitation: model for the study of human hematopoietic stem cells microenvironment. *Int J Stem Cells*. (2019) 12:51–62. doi: 10.15283/ijsc18061
113. Brahm MVJ, Li Yim ASP, Garcia Mateos J, Minnema MC, Dhert WJA, Öner FC, et al. A human hematopoietic niche model supporting hematopoietic stem and progenitor cells in vitro. *Adv Healthc Mater*. (2019) 8:1–14. doi: 10.11648/j.am.20190801.11
114. Born G, Nikolova M, Scherberich A, Treutlein B, García-García A, Martin I. Engineering of fully humanized and vascularized 3D bone marrow niches sustaining undifferentiated human cord blood hematopoietic stem and progenitor cells. *J Tissue Eng*. (2021) 12:204173142110448. doi: 10.1177/20417314211044855
115. Jeon JS, Bersini S, Whisler JA, Chen MB, Dubini G, Charest JL, et al. Generation of 3D functional microvascular networks with human mesenchymal stem cells in microfluidic systems. *Integr Biol*. (2014) 6:555–63. doi: 10.1039/C3IB40267C
116. Carrion B, Huang CP, Ghajar CM, Kachgal S, Kniazeva E, Jeon NL, et al. Recreating the perivascular niche ex vivo using a microfluidic approach. *Biotechnol Bioeng*. (2010) 107:1020–8. doi: 10.1002/bit.22891
117. Mykuliak A, Yrjänäinen A, Mäki AJ, Gebraad A, Lampela E, Kääriäinen M, et al. Vasculogenic potency of bone marrow- and adipose tissue-derived mesenchymal stem/stromal cells results in differing vascular network phenotypes in a microfluidic chip. *Front Bioeng Biotechnol*. (2022) 10:1–16. doi: 10.3389/fbioe.2022.764237
118. Nelson MR, Ghoshal D, Mejías JC, Rubio DF, Keith E, Roy K. A multi-niche microvascularized human bone marrow (hBM) on-a-chip elucidates key roles of the endosteal niche in hBM physiology. *Biomaterials*. (2021) 270:120683. doi: 10.1016/j.biomaterials.2021.120683
119. Chou DB, Frisimant V, Milton Y, David R, Pop-Damkov P, Ferguson D, et al. On-chip recapitulation of clinical bone marrow toxicities and patient-specific pathophysiology. *Nat Biomed Eng*. (2020) 4:394–406. doi: 10.1038/s41551-019-0495-z
120. Poulos MG, Gars EJ, Gutkin MC, Kloss CC, Ginsberg M, Scandura JM, et al. Activation of the vascular niche supports leukemic progression and resistance to chemotherapy. *Exp Hematol*. (2014) 42:976–986.e3. doi: 10.1016/j.exphem.2014.08.003
121. Kong J, Luo Y, Jin D, An F, Zhang W, Liu L, et al. A novel microfluidic model can mimic organ-specific metastasis of circulating tumor cells. *Oncotarget*. (2016) 7:78421–32. doi: 10.18632/oncotarget.9382
122. Seandel M, Butler JM, Kobayashi H, Hooper AT, White IA, Zhang F, et al. Generation of a functional and durable vascular niche by the adenoviral E4ORF1 gene. *Proc Natl Acad Sci U S A*. (2008) 105:19288–93. doi: 10.1073/pnas.0805980105
123. Ma C, Witkowski MT, Harris J, Dolgalev I, Sreeram S, Qian W, et al. Leukemia-on-a-chip: dissecting the chemoresistance mechanisms in B cell acute lymphoblastic leukemia bone marrow niche. *Sci Adv*. (2020) 6:1–14. doi: 10.1126/sciadv.aba5536
124. Jeon JS, Bersini S, Gilardi M, Dubini G, Charest JL, Moretti M, et al. Human 3D vascularized organotypic microfluidic assays to study breast cancer cell extravasation. *Proc Natl Acad Sci U S A*. (2015) 112:214–9. doi: 10.1073/pnas.1417115112
125. de la Puente P, Muz B, Gilson RC, Azab F, Luderer M, King J, et al. 3D tissue-engineered bone marrow as a novel model to study pathophysiology and drug resistance in multiple myeloma. *Biomaterials*. (2015) 73:70–84. doi: 10.1016/j.biomaterials.2015.09.017
126. Khan AO, Rodriguez-Romera A, Reyat JS, Olijnik AA, Colombo M, Wang G, et al. Human bone marrow organoids for disease modeling, discovery, and validation of therapeutic targets in hematologic malignancies. *Cancer Discov*. (2023) 13:364–85. doi: 10.1158/2159-8290.CD-22-0199
127. Bray LJ, Binner M, Körner Y, Von Bonin M, Bornhäuser M, Werner C. A three-dimensional ex vivo tri-culture model mimics cell-cell interactions between acute myeloid leukemia and the vascular niche. *Haematologica*. (2017) 102:1215–26. doi: 10.3324/haematol.2016.157883
128. Marturano-Kruik A, Nava MM, Yeager K, Chramiec A, Hao L, Robinson S, et al. Human bone perivascular niche-on-a-chip for studying metastatic colonization. *Proc Natl Acad Sci U S A*. (2018) 115:1256–61. doi: 10.1073/pnas.1714282115
129. Zheng Y, Sun Y, Yu X, Shao Y, Zhang P, Dai G, et al. Angiogenesis in liquid tumors: an in vitro assay for leukemic-cell-induced bone marrow angiogenesis. *Adv Healthc Mater*. (2016) 5:1014–24. doi: 10.1002/adhm.201501007
130. Glaser DE, Curtis MB, Sariano PA, Rollins ZA, Shergill BS, Anand A, et al. Organ-on-a-chip model of vascularized human bone marrow niches. *Biomaterials*. (2022) 280:121245. doi: 10.1016/j.biomaterials.2021.121245
131. Bersini S, Jeon JS, Dubini G, Arrigoni C, Chung S, Charest JL, et al. A microfluidic 3D in vitro model for specificity of breast cancer metastasis to bone. *Biomaterials*. (2014) 35:2454–61. doi: 10.1016/j.biomaterials.2013.11.050
132. Herland A, Ben MM, Das D, Somayaji RM, Prantil-Baun R, Novak R, et al. Quantitative prediction of human drug pharmacokinetic responses using multiple vascularized organ chips coupled by fluid transfer. *Nat Biomed Eng*. (2020) 4(4):421–36. doi: 10.1038/s41551-019-0498-9
133. Ronaldson-Bouchard K, Teles D, Yeager K, Tavakol DN, Zhao Y, Chramiec A, et al. A multi-organ chip with matured tissue niches linked by vascular flow. *Nat Biomed Eng*. (2022) 6:351–71. doi: 10.1038/s41551-022-00882-6
134. Kotha SS, Hayes BJ, Phong KT, Redd MA, Bomsztyk K, Ramakrishnan A, et al. Engineering a multicellular vascular niche to model hematopoietic cell trafficking. *Stem Cell Res Ther*. (2018) 9(1):77. doi: 10.1186/s13287-018-0808-2
135. Aleman J, George SK, Herberg S, Devarasetty M, Porada CD, Skardal A, et al. Deconstructed microfluidic bone marrow on-a-chip to study normal and malignant hemopoietic cell-niche interactions. *Small*. (2019) 15(43):e1902971. doi: 10.1002/smll.201902971
136. Palikuqi B, Nguyen DHT, Li G, Schreiner R, Pellegata AF, Liu Y, et al. Adaptable haemodynamic endothelial cells for organogenesis and tumorigenesis. *Nature*. (2020) 585:426–32. doi: 10.1038/s41586-020-2712-z
137. Shin Y, Han S, Jeon JS, Yamamoto K, Zervantonakis IK, Sudo R, et al. Microfluidic assay for simultaneous culture of multiple cell types on surfaces or within hydrogels. *Nat Protoc*. (2012) 7:1247–59. doi: 10.1038/nprot.2012.051
138. Reichert D, Friedrichs J, Ritter S, Kaübler T, Werner C, Bornhäuser M, et al. Phenotypic, morphological and adhesive differences of human hematopoietic progenitor cells cultured on murine versus human mesenchymal stromal cells. *Sci Rep*. (2015) 5:1–14. doi: 10.1038/srep15680
139. van Pel M, Fibbe WE, Schepers K. The human and murine hematopoietic stem cell niches: are they comparable? *Ann N Y Acad Sci*. (2016) 1370:55–64. doi: 10.1111/nyas.12994
140. Bennaceur-Griscelli A, Pondarré C, Schiavon V, Vainchenker W, Coulombel L. Stromal cells retard the differentiation of CD34⁺CD38^{low}/neg human primitive progenitors exposed to cytokines independent of their mitotic history. *Blood*. (2001) 97:435–41. doi: 10.1182/blood.V97.2.435
141. Coutu DL, Kokkaliaris KD, Kunz L, Schroeder T. Three-dimensional map of nonhematopoietic bone and bone-marrow cells and molecules. *Nat Biotechnol*. (2017) 35:1202–10. doi: 10.1038/nbt.4006
142. Hofer M, Pospíšil M, Komurková D, Hoferová Z. Granulocyte colony-stimulating factor in the treatment of acute radiation syndrome: a concise review. *Molecules*. (2014) 19:4770–8. doi: 10.3390/molecules19044770
143. Yamaguchi T, Kawamoto E, Gaowa A, Park EJ, Shimaoka M. Remodeling of bone marrow niches and roles of exosomes in leukemia. *Int J Mol Sci*. (2021) 22(4):1881. doi: 10.3390/ijms22041881
144. Mukkamalla SKR, Malipeddi D. Myeloma bone disease: a comprehensive review. *Int J Mol Sci*. (2021) 22:1–21. doi: 10.3390/ijms22126208
145. Yin JJ, Pollock CB, Kelly K. Mechanisms of cancer metastasis to the bone. *Cell Res*. (2005) 15:57–62. doi: 10.1038/sj.cr.7290266

146. Whiteley AE, Price TT, Cantelli G, Sipkins DA. Leukaemia: a model metastatic disease. *Nat Rev Cancer*. (2021) 21:461–75. doi: 10.1038/s41568-021-00355-z
147. De KI, Abdul-Hay M. Acute myeloid leukemia: a comprehensive review and 2016 update. *Blood Cancer J*. (2016) 6:e441–e441. doi: 10.1038/bcj.2016.50
148. Schmidt T, Carmeliet P. Angiogenesis: a target in solid tumors, also in leukemia? *Hematology Am Soc Hematol Educ Program*. (2011) 2011:1–8. doi: 10.1182/asheducation-2011.1.1
149. Iacobucci I, Mullighan CG. Genetic basis of acute lymphoblastic leukemia. *J Clin Oncol*. (2017) 35:975–83. doi: 10.1200/JCO.2016.70.7836
150. Brigle K, Rogers B. Pathobiology and diagnosis of multiple myeloma. *Semin Oncol Nurs*. (2017) 33:225–36. doi: 10.1016/j.soncn.2017.05.012
151. Zahr AA, Salama ME, Carreau N, Tremblay D, Verstovsek S, Mesa R, et al. Bone marrow fibrosis in myelofibrosis: pathogenesis, prognosis and targeted strategies. *Haematologica*. (2016) 101:660–71. doi: 10.3324/haematol.2015.141283
152. Reymond N, D'Água BB, Ridley AJ. Crossing the endothelial barrier during metastasis. *Nat Rev Cancer*. (2013) 13:858–70. doi: 10.1038/nrc3628
153. Bianchini G, De Angelis C, Licata L, Gianni L. Treatment landscape of triple-negative breast cancer — expanded options, evolving needs. *Nat Rev Clin Oncol*. (2022) 19:91–113 doi: 10.1038/s41571-021-00565-2
154. Oetjen KA, Lindblad KE, Goswami M, Gui G, Dagur PK, Lai C, et al. Human bone marrow assessment by single-cell RNA sequencing, mass cytometry, and flow cytometry. *JCI Insight*. (2018) 3:1–9. doi: 10.1172/jci.insight.124928
155. Baccin C, Al-Sabah J, Velten L, Helbling PM, Grünschlager F, Hernández-Malmierca P, et al. Combined single-cell and spatial transcriptomics reveal the molecular, cellular and spatial bone marrow niche organization. *Nat Cell Biol*. (2020) 22:38–48. doi: 10.1038/s41556-019-0439-6
156. Tikhonova AN, Dolgalev I, Hu H, Sivaraj KK, Hoxha E, Cuesta-Domínguez Á, et al. The bone marrow microenvironment at single-cell resolution. *Nature*. (2019) 569:222–8. doi: 10.1038/s41586-019-1104-8