



Prevalence of *Neospora caninum* antibodies in fattening pigs and sows from intensive farms in northern Italy

Luca Villa¹ · Alessia Libera Gazzonis¹ · Carolina Allievi¹ · Sergio Aurelio Zanzani¹ · Michele Mortarino¹ · Maria Teresa Manfredi¹

Received: 15 November 2021 / Accepted: 27 January 2022 / Published online: 4 February 2022
© The Author(s) 2022

Abstract

Neospora caninum (Apicomplexa, Sarcocystidae) is a major cause of reproductive failure in cattle. In pigs, only a few studies investigated the effects of this parasite on reproductive efficiency. Considering the relevance of swine farms in northern Italian regions, an epidemiological survey was designed to investigate the spread of *N. caninum* infection. Three hundred seventy fattening pigs and sows from 23 intensive farms in Lombardy were sampled. Sera were analyzed by a commercial immunofluorescence antibody test. Statistical analysis through univariate and multivariate generalized linear models was conducted to detect farm management practices enhancing the risk of infection. At the farm level, 52.1% (12/23) of the selected farms, 72.7% housing sows and 40% fattening pigs, scored positive. At the individual level, 25 animals (25/370, $P=6.7\%$) were positive to *N. caninum* antibodies: one fattening pig and two sows showed an antibody titer of 1:100, and in two sows, an antibody titer of 1:400 and 1:6400 was evidenced. A higher seroprevalence was detected in sows (17/151, $P=11.2\%$) if compared to fattening pigs (8/219, $P=3.6\%$) (OR = 1.19, P value = 0.000 in sows). Moreover, a higher seroprevalence was recorded in farms with low and moderate sanitary score ($P=100\%$ and $P=64.2\%$, respectively) if compared to farms with high sanitary score ($P=22.2\%$) (OR = 1.24, P value = 0.007 in score = 1 and OR = 1.10, P value = 0.050 in score = 2). This study provides the first data on the circulation of *N. caninum* in intensive swine farms in Italy, demonstrating the spread of the parasite in fattening pigs and sows in Lombardy region.

Keywords Neosporosis · Swine · Intensive farms · IFAT · Biosecurity · Reproductive problems

Introduction

Neospora caninum, an obligate intracellular protozoan, is the causative agent of neosporosis, a severe clinical disease of cattle and dogs worldwide (Dubey 2003). Serological evidence in domestic and wild animals indicates that many species were exposed to this parasite (Almería and López-Gatius 2013). Domestic dogs and wild canids are the definitive hosts; various species were reported as intermediate hosts of the parasite, including ruminants, equids, and swine (Dubey, 2003). *N. caninum* is a major cause of abortion, the main clinical manifestation of bovine neosporosis,

which causes huge economic losses to the dairy and beef industries worldwide (Thilsted and Dubey 1989; Goodswen et al. 2013).

In pigs, few studies investigated the effects of *N. caninum* infection on reproductive efficiency in sows. A major concern in swine farming are reproductive disorders; indeed, the drop in piglets/sow/year causes economic losses. A variety of pathogens are recognized as responsible for reproductive disorders in pigs: however, the pathogenesis of neosporosis and its consequences in the swine species remain unclear (Snak et al. 2019, 2021). A study evidenced an influence of *N. caninum* seropositivity on reproductive parameters in sows, i.e., age at first farrowing, the annual number of deliveries, and stillbirth incidence (Kanga-Waladjo et al. 2009). Recently, Snak et al. (2019) demonstrated that in experimentally infected pigs, the parasite could be trans-placentally transmitted in all phases of gestation, regardless of the time of infection, causing reproductive disorders and abortion with mummified fetuses, especially in the first and second

Section Editor: Sutherland Maciver

✉ Luca Villa
luca.villa@unimi.it

¹ Department of Veterinary Medicine, Università Degli Studi Di Milano, Via dell'Università 6, 26900 Lodi, Italy

gestational thirds. Besides, due to reactivation of the infection, the endogenous vertical transmission was evidenced in the sows inoculated in the final third of gestation. Moreover, *N. caninum* can cause clinical signs in infected female pigs, including hypothermia and leukocytosis in the acute phase of infection; the infection can also acutely reappear in chronically infected swine during pregnancy (Snak et al. 2021).

The occurrence of *N. caninum* natural infection in pigs was reported in some countries throughout the world with varying antibody prevalence depending on the region, the production system, and the diagnostic test employed (Wyss et al. 2000; Damriyasa et al. 2004; Helmick et al. 2002; Azevedo et al. 2010; Bártová and Sedlák 2011; Feitosa et al. 2014; Minetto et al. 2019; Silva et al. 2019; Gui et al. 2020).

Regarding Italy, *N. caninum* infection was reported in various species, i.e., in cattle (Otranto et al. 2003; Rinaldi et al. 2005; Villa et al. 2021) and other domestic species, including dogs, cats, equids, and small ruminants (Ferroglio et al. 2005, 2007a; Villa et al. 2018; Gazzonis et al. 2020), but also in wild mammals and birds (Ferroglio and Rossi 2001; Ferroglio et al. 2007b; Zanet et al. 2013; Gazzonis et al. 2021).

To date, no data on *N. caninum* infection in swine is available for Italy. Considering the relevance of swine farms under the intensive production system in northern Italian regions, this study aimed to investigate the spread of *N. caninum* infection in intensively reared fattening pigs and sows in Lombardy region. Besides, another aim was to evaluate the association between the seroprevalence of *N. caninum* and animal husbandry practices and farm biosecurity procedures to identify potential critical points of the farm system favorable to the parasite circulation.

Materials and methods

Sample collection

The sampling of fattening pigs and sows was performed as previously described for an epidemiological survey on *Toxoplasma gondii* infection in pigs from intensive production system (Gazzonis et al. 2018a).

The survey was carried out in Lombardy, one of the most suitable regions for intensive pig farming in northern Italy. Overall, 219 fattening pigs and 151 sows from 15 and 11 conventional farms, respectively, were sampled at five slaughterhouses; in three farms (Farm.02, Farm.13, and Farm.17), the collection of both sow and fattening pig samples was feasible. For each farm, an average of 16 individuals was sampled (min–max, 3–34). During the slaughtering operations, for each animal, a blood sample was collected

from a jugular vein into tubes without anticoagulants. Blood samples were transported to the laboratory within a few hours; blood was centrifuged (15 min, 2120 × g), and serum was transferred into Eppendorf tubes and stored at –20 °C until serological analysis.

At sampling time, data on farm management were collected, and a “biosecurity score” (ranging from 1 = poor, 2 = moderate, to 3 = optimal) was determined for each farm based on parameters regarding the sanitary procedures applied, as previously described (Gazzonis et al. 2018a).

Serological analysis

Sera samples were analyzed for anti-*N. caninum* antibodies by immunofluorescence antibody test, using slides coated with *Neospora caninum* antigens provided in a commercial kit (MegaScreen® FLUO NEOSPORA caninum, Megacor, Austria), following the manufacturer’s instruction, with slight modifications. An initial screening dilution of 1:50 of serum in PBS was used according to Snak et al. (2019); then, seropositive samples were twofold serially diluted to determine the end-point antibody titer. Briefly, 20 µl of serum dilutions were pipetted on separate antigen wells; 20 µl of the positive and negative controls supplied in the kit were also included in each assay. The slides were incubated for 30 min at 37 °C in a humid chamber. After a washing step with PBS, one drop (20 µl) of FITC anti-pig IgG as conjugate (MegaFLUO® FITC anti-pig IgG Conjugate, Megacor, Austria) was placed onto each well. A subsequent incubation step for 30 min at 37 °C in the dark followed by another washing step was performed. Finally, some drops of mounting medium were added to the coverslips, which were then placed on the slides. The slides were evaluated using a fluorescence microscope (Axioscope 2, Zeiss), comparing each well to the fluorescence of the positive and negative controls, considered as a reference pattern. Only a bright, sharp, and clear, yellow-green fluorescence on the membrane extending to the whole body of *N. caninum* tachyzoites was considered a positive reaction (Fig. 1).

Statistical analysis

Seroprevalence of *N. caninum* infection was calculated at the individual and farm levels according to the considered categories. A farm was considered positive if at least one sampled animal scored positive. Serological data were analyzed to determine which variables could be predictors of *N. caninum* infection; the farm was considered as the statistical unit. Overall, 26 observations were included in the analysis. The only three farms where both sows and

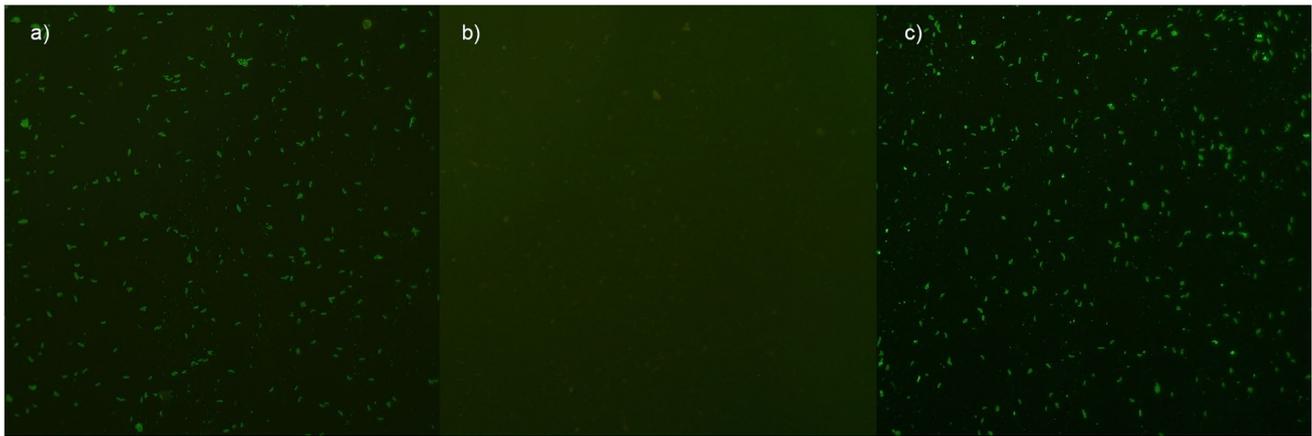


Fig. 1 Immunofluorescence detection images (100×) of **a** positive control, **b** negative control, and **c** one positive sow with an antibody titer of 1:6400 at 1:160 dilution

fattening pigs were sampled, were defined as independent units, since management could considerably vary within the same farm according to the considered productive category.

Separate generalized linear models (GLMs) with negative binomial distribution were performed to verify the influence of farm management on *N. caninum* infection. The intra-herd seroprevalence was considered as the dependent variable; the variables listed in Table 1 were entered as independent variables in the univariate model.

All variables showing a *P* value < 0.1 were entered in multivariate models developed through a backward selection procedure (significance level to remove variables from the model = 0.05), based on Akaike's information criterion (AIC) values. Besides, in the final models, the estimated prevalence values of the biosecurity score levels were compared through pairwise comparisons. Statistical analysis was performed using IBM SPSS Statistics for Windows version 25.0 software (IBM Corp., Armonk, NY, USA).

Table 1 Baseline characteristics related to farm management of pig farms and *Neospora caninum* seroprevalence values for each considered category

Variable	Category	Positive/examined farms	P% (95% CI)
Productive category	Sows	8/11	72.7 (43.44–90.25)
	Fattening pigs	6/15	40.0 (19.82–64.25)
Number of pigs in the farm	≤ 1000	2/8	25.0 (7.15–59.07)
	1000–2500	4/10	40.0 (16.82–68.73)
	≥ 2500	4/8	50.0 (21.52–78.48)
Floor type	Fully slatted	1/2	50.0 (9.45–90.55)
	Partly slatted	1/9	11.1 (1.99–43.50)
	Straw bedding	5/15	33.3 (15.18–58.29)
Animal density (m ²)	1	4/8	50.0 (21.52–78.48)
	1–2	3/11	27.2 (9.75–56.56)
	≥ 2	3/7	42.8 (15.82–74.95)
Possibility to access outdoors	Yes	5/15	33.3 (15.18–58.29)
	No	5/11	45.5 (21.27–71.99)
Presence of rodents	Yes	14/26	53.8 (35.46–71.24)
	No	0/0	0 (–)
Application of pest control	Only internal	3/7	42.8 (15.82–74.95)
	Internal and external	7/19	36.8 (19.15–58.96)
Sanitary score	1	3/3	100 (43.85–100)
	2	9/14	64.2 (38.76–83.66)
	3	2/9	22.2 (6.32–54.74)

Table 2 *Neospora caninum* infection in pigs in northern Italy: serological results including antibody titers reported for each examined farm

Farm	No. of positive/examined animals (antibody titer)		
	Fattening pigs	Sows	Total
1	-	3/7 (1:50, 1:50, 1:100)	3/7
2	1/30 (1:50)	2/4 (1:50, 1:6400)	3/34
3	1/12 (1:50)	-	1/12
4	0/10	-	0/10
5	0/18	-	0/18
6	0/18	-	0/18
7	-	0/30	0/30
8	-	3/5 (1:50, 1:100, 1:400)	3/5
9	0/5	-	0/5
10	0/10	-	0/10
11	2/15 (1:50, 1:100)	-	2/15
12	-	0/4	0/4
13	0/10	2/30 (1:50)	2/40
14	-	1/4 (1:50)	1/4
15	-	4/30 (1:50)	4/30
16	-	0/30	0/30
17	1/25 (1:50)	1/4 (1:50)	2/29
18	0/10	-	0/10
19	2/18 (1:50)	-	2/18
20	0/10	-	0/10
21	1/18 (1:50)	-	1/18
22	0/10	-	0/10
23	-	1/3 (1:50)	1/3
Total	8/219	17/151	25/370
Prevalence (%)	3.65	11.25	6.75

Results

Serological analysis

Overall, 370 swine from 23 farms, including 219 fattening pigs from 15 farms and 151 sows from 11 farms, were

analyzed for anti-*N. caninum* antibodies. At the farm level, 52.1% (12/23) of the selected farms hosted at least one positive animal; in particular, 72.7% of the farms housing sows and 40% of those housing fattening pigs included in the study scored positive. Serological results for each examined farm are reported in Table 2. At the individual level, 25 animals (25/370, $P = 6.7\%$, 95% CI 4.62–9.78) were positive to *N. caninum* at the initial screening dilution. A higher seroprevalence was detected in sows (17/151, $P = 11.2\%$, 95% CI 7.15–17.29) compared to fattening pigs (8/219, $P = 3.6\%$, 95% CI 1.86–7.04). The intra-herd seroprevalence in *N. caninum* infected farms varied between 13.3 and 60% considering sows and between 3.3 and 13.3% in fattening pigs ($P = 23\%$ and $P = 3\%$, respectively). Moreover, a higher seroprevalence was recorded in farm with a low sanitary score ($P = 100\%$ and $P = 64.2\%$ in farms with score = 1 and score = 2, respectively) if compared to farms with higher sanitary score ($P = 22.2\%$ in farms with score = 3). Among seropositive animals, one fattening pig and two sows showed an antibody titer of 1:100; besides, an antibody titer of 1:400 and 1:6400 was evidenced in two sows, respectively. The serological results related to farm characteristics are reported in Table 1.

Statistical analysis

By univariate analysis, only the variables “productive category” and “biosecurity score” were significantly associated with *N. caninum* infection and were entered in the final multivariate model (Table 3). Indeed, sows were at a higher risk of infection than fattening pigs (OR = 1.19, P value = 0.000). Moreover, the biosecurity score was a predictor of infection, increasing the risk of infection while decreasing the score (OR = 1.24, P value = 0.007 and OR = 1.10, P value = 0.050 in score = 1 and score = 2, respectively). Besides, the pairwise comparison revealed that seroprevalence values of farms with biosecurity score = 1 and score = 2 were statistically different from those of score = 3 (P value < 0.05).

Table 3 Results of the multivariate analysis of the risk factors related to *Neospora caninum* seroprevalence in pigs in Lombardy

Response variable	Category	P%	β^a	Standard error of coefficients	Wald Chi-square	Odds ratio (95% confidence interval)	P value	Akaike information criterion	
Productive category	Sows	72.7	0.18	0.05	13.43	1.19 (1.09–1.31)	0.000	15.63	
	Fattening pigs	40.0	0			1			
Sanitary score	1	100 _a	0.22	0.08	7.17	1.24 (1.06–1.45)	0.007		
	2	64 _a	0.10	0.05		1.10 (1.00–1.22)			0.050
	3	22	0			1			

Seroprevalence values according to each variable followed by the lowercase letter “a” are statistically different from other values without the lowercase letter “a” at P value < 0.05 (generalized linear models’ analysis, pairwise comparison), while they are not statistically different from each other (P value > 0.05, GLM, pairwise comparison)

^aCoefficient

Discussion

This study provides the first data on the circulation of *N. caninum* in intensive swine farms in Italy. A seroprevalence of 6.7% was evidenced at the individual level; at the farm level, 52.1% of the farms were positive. In the present survey, a cut-off of 1:50 for IFAT was applied, according to previous studies (Paré et al. 1995; Snak et al. 2019). Some animals presented higher antibody titer: in particular, one fattening pig and two sows showed an antibody titer of 1:100 and two sows of 1:400 and 1:6400, respectively.

It was evidenced that breeder animals were at higher risk of acquiring the parasite infection: sows were more frequently found seropositive ($P = 11.2\%$) if compared to fattening pigs ($P = 3.6\%$), also considering both the farm level ($P = 72.7\%$ and $P = 40.0\%$, respectively) and the mean intra-herd seroprevalence ($P = 23\%$ and $P = 3\%$, respectively). Indeed, sows are slaughtered at an older age (3–5 years or more) than fattening pigs (approximately 9 months), increasing their risk of acquiring the infection through environmental, horizontal transmission. The same finding was previously revealed in pigs also in other studies (Wyss et al. 2000; Silva et al. 2019; Gui et al. 2020).

In addition to individual characteristics, some features concerning farm management, including sanitary procedures, were associated with *N. caninum* infection. Risk factor analysis highlighted that the farms with a low biosecurity level presented a higher risk of being infected. According to previous studies which demonstrated the association between sanitary procedures and seropositivity for *T. gondii* (Gazzonis et al. 2018a), farms with a poor or moderate sanitary score recorded a seroprevalence of *N. caninum* of 100% and 64.2%, whereas those with a good biosecurity level evidenced a seropositivity of 22.2%. The adoption of adequate hygiene, prevention measures, and biosecurity protocols, including the application of a health management program, vaccination protocols, standard protocols for quarantine for imported animals, protocols for visitors/transporters, sanitary protocols for operators, and application of an all-in/all-out system, proved once again to be fundamental to prevent the risk of infections in the farm, including parasitic infections.

This is the first study investigating *N. caninum* infection in swine species in Italy. However, considering the Italian epidemiological scenario, previous studies reported the circulation of the parasite in other species. Indeed, in cattle, serological studies evidenced a herd prevalence of 44.1% and 77.8% and individual seropositivity values of 11% and 30.8%, respectively (Otranto et al. 2003; Rinaldi et al. 2005). A recent survey reported a molecular prevalence of *N. caninum* of 27.8% in aborted bovine fetuses in cattle farms in Lombardy region, confirming that the parasite circulates in

the study area (Villa et al. 2021). Concerning small ruminants, Gazzonis et al. (2020) reported herd and individual prevalence values of 89.4% and 19.3% in sheep and 32.1% and 5.7% in goats in Lombardy, respectively. Moreover, other domestic species in northern Italian regions were also exposed to the parasite with variable seroprevalence values, i.e., 30.2% in dogs (Ferroglio et al. 2007a), 31.9% in cats (Ferroglio et al. 2005), and 0.4% in horses (Villa et al. 2018). Among wild mammals, Ferroglio and Rossi (2001) confirmed serological exposure to *N. caninum* in wild ruminants (5.9% in chamois, 2.3% in roe deer, and 1.9% in red deer), whereas parasitic DNA was detected in 10.3% of rodents (Ferroglio et al. 2007b) and 2.8% of eastern cottontail rabbits (Zanet et al. 2013); besides, Gazzonis et al. (2021) reported a molecular prevalence of 3.6% in wild birds in northern Italy. All these data confirm the circulation of *N. caninum* in both domestic and wild animals in Italy.

To compare seroprevalence data, differences in study populations, diagnostic techniques, and protocols should always be considered. The values recorded in other European studies resulted similar but lower than those of our study: 1–3% in Switzerland (Wyss et al. 2000), 3% in the Czech Republic (Bártová and Sedlák 2011), 0.04% in Germany (Damriyasa et al. 2004), and 0% in England (Helmick et al. 2002). Outside Europe, some studies were conducted in Brazil, where the seroprevalence varied between 3.1–3.2% (Azevedo et al. 2010; Feitosa et al. 2014), 13.5% (Minetto et al. 2019), and 18.9% (Silva et al. 2019). Moreover, a study from China recorded a prevalence of 1.9%, and parasitic DNA was detected in the brain tissues of three pigs (Gui et al. 2020).

To date, only a few studies investigated the effects of *N. caninum* infection in sows. A survey on wandering sows in Senegal evidenced an influence of *N. caninum* seropositivity on reproductive parameters, i.e., age at first farrowing, the annual number of deliveries, and stillbirth incidence (Kamga-Waladjo et al. 2009).

Furthermore, serological studies on the spread of *N. caninum* were also conducted in European wild boars' populations (Almería et al. 2007; Bártová et al. 2006; Reiterová et al. 2016). Considering that wild boar populations are nowadays in expansion in terms both of the number of animals and habitat range, the increased frequency of contacts among wild boars, livestock, and humans could influence the transmission of zoonotic and animal-specific pathogens, as previously demonstrated in this area for other similar protozoa infections (Gazzonis et al. 2018b, 2019a). In the same study area, a recent molecular survey reporting the presence of *N. caninum* in brain tissue of wild birds of prey not only suggests the involvement of avian species in the parasite life cycle but also the environmental circulation of *Neospora*, indicating a possible role of these wild

populations in the epidemiology of the parasite infection at the interface of the domestic and sylvatic life cycle (Gazzonis et al. 2021).

Indeed, in analogy to other intermediate hosts, domestic pigs probably become infected with *N. caninum* by ingesting food or drinking water contaminated by sporulated oocysts shed from dogs or tissues containing cysts of other intermediate hosts (e.g., micromammals). Besides, the parasite may be transmitted trans-placentally (congenital vertical transmission) from an infected dam to the fetuses during pregnancy, as experimentally demonstrated (Jensen et al., 1998; Snak et al. 2019).

Experimental studies evidenced that all pigs seroconverted after inoculation with Nc1 strain (Dubey et al. 1996; Jensen et al. 1998; Snak et al. 2019, 2021). Moreover, Jensen et al. (1998) also detected the parasite in two fetuses providing the first indication of transplacental transmission of *N. caninum* in pigs. A recent study (Snak et al. 2019) confirmed that in pigs, *N. caninum* could be transmitted via the placenta, causing reproductive disorders, in particular mummified fetuses, especially in the first and second gestational thirds. Furthermore, in the same study, parasitic DNA was detected in milk and amniotic fluid, suggesting a role of these matrices in the protozoan transmission, as also evidenced for both *T. gondii* and *N. caninum* in other species, including ruminants and equids (Björkman et al. 1997; González-Warleta et al. 2011; Mancianti et al. 2014; Gazzonis et al. 2018c, 2019b). It was also demonstrated that *N. caninum* can cause clinical signs in adults; indeed, experimentally infected sows showed hyperthermia followed by hypothermia and leukocytosis in the acute phase (Snak et al. 2021).

Conclusions

N. caninum circulates widely in the pig farms of northern Italy, where natural parasite infection occurs both in fattening pigs and in sows. Considering the importance of pig farming in the study area, linked to valuable production, the impact of *N. caninum* infection in intensive pig farming should be further investigated, particularly in breeding sows, due to its possible involvement in reproductive problems. Indeed, the correct diagnosis of the cause of abortion is important for adopting appropriate control and prevention measures. Among the further perspectives, the molecular detection and multilocus microsatellite genotyping of *N. caninum* would indicate the spatial distribution and mutual connections between the parasite's isolates from different species.

Author contribution LV, ALG, MM, and MTM conceived and designed the study. LV and ALG collected the samples. LV, ALG, CA, and SAZ performed the laboratory analyses. LV wrote the first

draft of the manuscript. LV, ALG, MM, and MTM revised and edited the manuscript. All authors read and approved the final manuscript.

Data availability The data that support the findings of this study are available from the corresponding author upon reasonable request.

Code availability Not applicable.

Declarations

Ethics approval All procedures were approved by the Institutional Animal Care and Use Committee of Università degli Studi di Milano (“Organismo Preposto al Benessere degli Animali,” Prot. no. OPBA_34_2017).

Consent to participate Not applicable.

Consent for publication Not applicable.

Informed consent Not applicable.

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Almería S, López-Gatius F (2013) Bovine neosporosis: clinical and practical aspects. *Res Vet Sci* 95:303–309. <https://doi.org/10.1016/j.rvsc.2013.04.008>
- Almería S, Vidal D, Ferrer D, Pabón M, Fernández-de-Mera MIG, Ruiz-Fons F, Alzaga V, Marco I, Calvete C, Lavin S, Gortazar C, López-Gatius F, Dubey JP (2007) Seroprevalence of *Neospora caninum* in non-carnivorous wildlife from Spain. *Vet Parasitol* 143:21–28. <https://doi.org/10.1016/j.vetpar.2006.07.027>
- Azevedo SS, Pena HFJ, Alves CJ, Guimaraes Filho AAM, Oliveira RM, Maksimov P, Schares G, Gennari SM (2010) Prevalence of anti-*Toxoplasma gondii* and anti-*Neospora caninum* antibodies in swine from Northeastern Brazil. *Rev Bras Parasitol Vet* 19:80–84
- Bártová E, Sedlák K, Literák I (2006) Prevalence of *Toxoplasma gondii* and *Neospora caninum* antibodies in wild boars in the Czech Republic. *Vet Parasitol* 142:150–153. <https://doi.org/10.1016/j.vetpar.2006.06.022>
- Bártová E, Sedlák K (2011) Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in slaughtered pigs in the Czech Republic. *Parasitology* 138:1369–1371. <https://doi.org/10.1017/S0031182011001041>
- Björkman C, Holmdahl OJ, Uggla A (1997) An indirect enzyme-linked immunoassay (ELISA) for demonstration of antibodies

- to *Neospora caninum* in serum and milk of cattle. *Vet Parasitol* 68:251–260. [https://doi.org/10.1016/S0304-4017\(96\)01076-X](https://doi.org/10.1016/S0304-4017(96)01076-X)
- Damriyasa IM, Bauer C, Edelhofer R, Failing K, Lind P, Petersen E, Schares G, Tenter AM, Volmer R, Zahner H (2004) Cross-sectional survey in pig breeding farms in Hesse, Germany: seroprevalence and risk factors of infections with *Toxoplasma gondii*, *Sarcocystis* spp. and *Neospora caninum* in sows. *Vet Parasitol* 126:271–286. <https://doi.org/10.1016/j.vetpar.2004.07.016>
- Dubey JP, Lindsay DS, Adams DS, Gay JM, Baszler TV, Blagburn BL, Thulliez P (1996) Serologic responses of cattle and other animals infected with *Neospora caninum*. *Am J Vet Res* 57:329–336
- Dubey JP (2003) Review of *Neospora caninum* and neosporosis in animals. *Korean J Parasitol* 41:1–16. <https://doi.org/10.3347/kjp.2003.41.1.1>
- Feitosa TF, Vilela VLR, Melo LRB, Almeida Neto JL, Souto DVO, Morais DF, Athayde ACR, Azevedo SS, Pena HFJ (2014) *Toxoplasma gondii* and *Neospora caninum* in slaughtered pigs from Northeast, Brazil. *Vet Parasitol* 202:305–309. <https://doi.org/10.1016/j.vetpar.2014.03.015>
- Ferroglio E, Rossi L (2001) Prevalence of *Neospora caninum* antibodies in wild ruminants from the Italian Alps. *Vet Rec* 148:754–755. <https://doi.org/10.1136/vr.148.24.754>
- Ferroglio E, Guiso P, Pasino M, Accossato A, Trisciuglio A (2005) Antibodies to *Neospora caninum* in stray cats from north Italy. *Vet Parasitol* 131:31–34. <https://doi.org/10.1016/j.vetpar.2005.04.012>
- Ferroglio E, Pasino M, Ronco F, Benà A, Trisciuglio A (2007a) Seroprevalence of antibodies to *Neospora caninum* in urban and rural dogs in north-west Italy. *Zoonoses Public Health* 54:135–139. <https://doi.org/10.1111/j.1863-2378.2007.01033.x>
- Ferroglio E, Pasino M, Romano A, Grande D, Pregel P, Trisciuglio A (2007b) Evidence of *Neospora caninum* DNA in wild rodents. *Vet Parasitol* 148:346–349. <https://doi.org/10.1016/j.vetpar.2007.06.031>
- Gazzonis AL, Marangi M, Villa L, Ragona ME, Olivieri E, Zanzani SA, Giangaspero A, Manfredi MT (2018a) *Toxoplasma gondii* infection and biosecurity levels in fattening pigs and sows: serological and molecular epidemiology in the intensive pig industry (Lombardy, northern Italy). *Parasitol Res* 117:539–546. <https://doi.org/10.1007/s00436-017-5736-z>
- Gazzonis AL, Villa L, Riehn K, Hamedy A, Minazzi S, Olivieri E, Zanzani SA, Manfredi MT (2018b) Occurrence of selected zoonotic food-borne parasites and first molecular identification of *Alaria alata* in wild boars (*Sus scrofa*) in Italy. *Parasitol Res* 117:2207–2215. <https://doi.org/10.1007/s00436-018-5908-5>
- Gazzonis AL, Zanzani SA, Stradiotto K, Olivieri E, Villa L, Manfredi MT (2018c) *Toxoplasma gondii* antibodies in bulk tank milk samples of caprine dairy herds. *J Parasitol* 104:560–565. <https://doi.org/10.1645/17-44>
- Gazzonis AL, Gjerde B, Villa L, Minazzi S, Zanzani SA, Riccaboni P, Sironi G, Manfredi MT (2019a) Prevalence and molecular characterisation of *Sarcocystis miescheriana* and *Sarcocystis suihominis* in wild boars (*Sus scrofa*) in Italy. *Parasitol Res* 118:1271–1287. <https://doi.org/10.1007/s00436-019-06249-2>
- Gazzonis AL, Zanzani SA, Villa L, Manfredi MT (2019b) *Toxoplasma gondii* in naturally infected goats: monitoring of specific IgG levels in serum and milk during lactation and parasitic DNA detection in milk. *Prev Vet Med* 170:104738. <https://doi.org/10.1016/j.prevetmed.2019.104738>
- Gazzonis AL, Villa L, Manfredi MT, Zanzani SA (2020) Spatial analysis of infections by *Toxoplasma gondii* and *Neospora caninum* (Protozoa: Apicomplexa) in small ruminants in northern Italy. *Animals* 9:916. <https://doi.org/10.3390/ani9110916>
- Gazzonis AL, Villa L, Lubian E, Ressegotti S, Grilli G, Raimondi S, Zanzani SA, Manfredi MT (2021) Molecular survey on *Toxoplasma gondii* and *Neospora caninum* infection in wild birds of prey. *Microorganisms* 9:736. <https://doi.org/10.3390/microorganisms9040736>
- González-Warleta M, Castro-Hermida JA, Carro-Corral C, Mezo M (2011) Anti-*Neospora caninum* antibodies in milk in relation to production losses in dairy cattle. *Prev Vet Med* 101:58–64. <https://doi.org/10.1016/j.prevetmed.2011.04.019>
- Gui BZ, Lv QY, Ge M, Li RC, Zhu XQ, Liu GH (2020) First report of *Neospora caninum* infection in pigs in China. *Transbound Emerg Dis* 67:29–32. <https://doi.org/10.1111/tbed.13358>
- Goodswen SJ, Kennedy PJ, Ellis JT (2013) A review of the infection, genetics, and evolution of *Neospora caninum*: from the past to the present. *Infect Genet Evol* 13:133–150. <https://doi.org/10.1016/j.meegid.2012.08.012>
- Helmick B, Otter A, McGarry J, Buxton D (2002) Serological investigation of aborted sheep and pigs for infection by *Neospora caninum*. *Res Vet Sci* 73:187–189. [https://doi.org/10.1016/S0034-5288\(02\)00093-0](https://doi.org/10.1016/S0034-5288(02)00093-0)
- Jensen L, Jensen TK, Lind P, Henriksen SA, Uggla A, Bille-Hansen V (1998) Experimental porcine neosporosis. *APMIS* 106:475–482. <https://doi.org/10.1111/j.1699-0463.1998.tb01374.x>
- Kamga-Waladjo AR, Chatagnon G, Bakou SN, Boly H, Diop PEH, Tainturier D (2009) *Neospora caninum* antibodies and its consequences for reproductive characteristics in wandering sows from Senegal, West Africa. *Asian J Anim Vet Adv* 4:263–266. <https://doi.org/10.3923/ajava.2009.263.266>
- Mancianti F, Nardoni S, Papini R, Mugnaini L, Martini M, Altomonte I, Salari F, D’Ascenzi C, Dubey JP (2014) Detection and genotyping of *Toxoplasma gondii* DNA in the blood and milk of naturally infected donkeys (*Equus asinus*). *Parasit Vectors* 7:165. <https://doi.org/10.1186/1756-3305-7-165>
- Minetto MK, Witter R, Oliveira ACS, Minetto JA, Barros ML, Aguiar DM, Pacheco RC (2019) Antibodies anti-*Toxoplasma gondii* and anti-*Neospora caninum* in backyard pigs from the state of Mato Grosso, Brazil. *Rev Bras Parasitol Vet* 28:403–409. <https://doi.org/10.1590/S1984-29612019050>
- Otranto D, Llazarri A, Testini G, Traversa D, di Regalbono AF, Badan M, Capelli G (2003) Seroprevalence and associated risk factors of neosporosis in beef and dairy cattle in Italy. *Vet Parasitol* 118:7–18. <https://doi.org/10.1016/j.vetpar.2003.10.008>
- Paré J, Hietala SK, Thurmond MC (1995) Interpretation of an indirect fluorescent antibody test for diagnosis of *Neospora* sp. infection in cattle. *J Vet Diagn Invest* 7:273–275. <https://doi.org/10.1177/104063879500700222>
- Reiterová K, Špilovská S, Blaňarová L, Derdákóvá M, Čobádiová A, Hisira V (2016) Wild boar (*Sus scrofa*) - reservoir host of *Toxoplasma gondii*, *Neospora caninum* and *Anaplasma phagocytophilum* in Slovakia. *Acta Parasitol* 61:255–260. <https://doi.org/10.1515/ap-2016-0035>
- Rinaldi L, Fusco G, Musella V, Veneziano V, Guarino A, Taddei R, Cringoli G (2005) *Neospora caninum* in pastured cattle: determination of climatic, environmental, farm management and individual animal risk factors using remote sensing and geographical information systems. *Vet Parasitol* 128:219–230. <https://doi.org/10.1016/j.vetpar.2004.12.011>
- Silva M, Snak A, Reiter JC, Junior GS, Cristani J, Moura AB (2019) Occurrence of antibodies against *Neospora caninum* in sows and factors associated with infection in commercial herds in two regions of the state of Santa Catarina, Brazil. *Semin Ciênc Agrár* 41:697–702. <https://doi.org/10.5433/1679-0359.2020v41n2p697>
- Snak A, Junior GS, Pilati GVT, Kroetz CC, Consoni W, Cristani J, de Moura AB (2019) Does *Neospora caninum* cause reproductive problems in pigs? *Vet Parasitol* 275:108934. <https://doi.org/10.1016/j.vetpar.2019.108934>
- Snak A, Henrique SM, Sebolt APR, Cristani J, Sato ME, Miletti MC, de Moura AB (2021) Experimental infection of tachyzoites of

- the NC1 strain of *Neospora caninum* in female swine. *Parasitol Res* 120:1049–1057. <https://doi.org/10.1007/s00436-021-07054-6>
- Thilsted JP, Dubey JP (1989) Neosporosis-like abortions in a herd of dairy cattle. *J Vet Diagn Invest* 1:205–209. <https://doi.org/10.1177/104063878900100301>
- Villa L, Gazzonis AL, Alvarez-Garcia G, Diezma-Diaz C, Zanzani SA, Manfredi MT (2018) First detection of anti-*Besnoitia* spp specific antibodies in horses and donkeys in Italy. *Parasitol Int* 67:640–643. <https://doi.org/10.1016/j.parint.2018.06.008>
- Villa L, Maksimov P, Luttermann C, Tuschy M, Gazzonis AL, Zanzani SA, Mortarino M, Conraths FJ, Manfredi MT, Schares G (2021) Spatial distance between sites of sampling associated with genetic variation among *Neospora caninum* in aborted bovine fetuses from northern Italy. *Parasit Vectors* 14:47. <https://doi.org/10.1186/s13071-020-04557-6>
- Wyss R, Sager H, Muller N, Inderbitzin F, Konig M, Audige L, Gottstein B (2000) Untersuchungen zum Vorkommen von *Toxoplasma gondii* und *Neospora caninum* unter fleischhygienischen Aspekten. *Schweiz Arch Tierheilk* 142:95–108. <https://doi.org/10.5169/seals-590222>
- Zanet S, Palese V, Trisciuglio A, Cantón Alonso C, Ferroglio E (2013) *Encephalitozoon cuniculi*, *Toxoplasma gondii* and *Neospora caninum* infection in invasive eastern cottontail rabbits *Sylvilagus floridanus* in northwestern Italy. *Vet Parasitol* 197:682–684. <https://doi.org/10.1016/j.vetpar.2013.05.014>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.