

# AUTHOR QUERIES

***Authors please note: Authors are responsible for any page charges as outlined in the acceptance letter or as indicated on the Instructions for Authors (<https://www.ahajournals.org/atvb/reviced-accepted-manuscripts>). Unless you have selected open access for your article, or it is otherwise noted on the acceptance letter, page charges are as follows: \$70 per black and white page (print articles only) or \$35 per page (online-only articles only). For each color page (print only), please add \$653/page. If there are any concerns regarding these charges, these should be addressed within 48 hours of receiving the s-proof. Author(s) will be invoiced for all color and page charges post publication. If you have selected open access for your article, please refer to details in the queries below.***

## AUTHOR PLEASE ANSWER ALL QUERIES

- AQ1–Please note only those terms that are used 5 times or more can be abbreviated, except trial names and proteins, which should be expanded at first use but then can be abbreviated throughout regardless of how many times they appear.
- AQ2–Please turn to page 2 of your proof and review the running head, which will appear in the upper right-hand margins of odd-numbered pages. Running heads must be 50 or fewer characters in length, including spaces and punctuation. If your original short title was longer than 50 characters, we may have shortened it. Please modify if necessary (but observe our length guidelines).
- AQ3–Please confirm that all authors are included in the correct order in the byline and that all names are spelled correctly, including special characters, accents, middle initials, and degrees, if applicable. For indexing purposes, confirm author names have been correctly identified as given names (blue), surnames (red), and suffixes (black). Color in the byline will not appear on the final published version. Note that journal style discourages listing honorary degrees (FAHA, FRCP, etc.) in the byline; please delete such degrees from the author byline.
- AQ4–Please note that as per journal style, in the first occurrence of a protein name in the text, protein abbreviation should appear first, followed by its expansion in parentheses. Therefore, please check and add expansion for the protein names within parentheses next to the first use of the protein name in the abstract, text, table footnotes, and in figure legends.
- AQ5–Please define “APTT, FXII, and PT” in text.
- AQ6–Please confirm that all authors’ institutional affiliations (including city/state/country locations) are correct as shown in the affiliations footnote.
- AQ7–Please carefully review any Acknowledgments, Sources of Funding, and/or Disclosures listed at the end of the manuscript (before the References), and confirm that they are accurate and complete for all authors.
- AQ8–Please provide volume and page range for reference 6.

AQ9–Please provide volume for reference 10.

# Arteriosclerosis, Thrombosis, and Vascular Biology

## LETTER TO THE EDITOR

### Letter by Brambilla et al Regarding Article, "Platelets Promote Thromboinflammation in SARS-CoV-2 Pneumonia"

Marta Brambilla, Paola Canzano, Elena Tremoli, Marina Camera

#### To the Editor:

We read with great interest the work recently published by Taus *et al*<sup>1</sup> highlighting the central role played by platelets in the thromboinflammation that characterizes coronavirus disease 2019 (COVID-19) pneumonia, being primed to spread proinflammatory and procoagulant activity.

The authors reported an increased potential of platelets to release cytokines, chemokines, and growth factors, highlighting their possible involvement in the local and systemic inflammation as well as in immune modulation. They also provided evidence that in patients with COVID-19 platelets are characterized by high expression levels of P-selectin, and this leads to an increased number of platelet-leukocyte aggregates. This feature has been consistently reported also in other studies published in the recent past months.<sup>2-6</sup> Despite this platelet activated phenotype, no other major alteration in the platelet-dependent hemostatic process (except for a reduced expression of collagen-induced fibrinogen receptor  $\alpha\text{IIb}\beta\text{3}$ ) was reported. Finally, the authors addressed the platelet involvement in the COVID-19 associated procoagulant phenotype by separately analyzing the intrinsic and extrinsic pathway of blood coagulation. They stated that the platelet procoagulant contribution is mediated by an increased FXII activity that correlated with APTT. Conversely, the authors suggested that the extrinsic pathway of coagulation plays a minor role in the prothrombotic condition associated with the disease since in patients with COVID-19 they found a prolonged PT associated with normal FVII activity.

It should be considered on this regard, however, that PT cannot evaluate the contribution of the endogenous TF (tissue factor), the main trigger of the extrinsic coagulation also present in human platelets. Furthermore, a direct evaluation of the platelet-associated expression of TF nor of phosphatidylserine, the phospholipid required for the assembly of the coagulation factors, was not performed due to limitations of safety protocols.

Some recently published data come in help filling this gap. In particular, it has been shown that the procoagulant role of platelets in patients with COVID-19 is sustained more by TF than by phosphatidylserine exposure. We have indeed provided evidences, by means of a whole blood flow cytometry analysis approach, that in these patients the number of TF expressing platelets is more than twice greater than that measured in healthy subjects ( $P < 0.001$ ).<sup>6</sup> The protein is functionally active being able to sustain thrombin generation through the activation of the extrinsic coagulation pathway. Unlike the TF expressing platelets, the number of phosphatidylserine positive platelets in patients with COVID-19 was comparable to that measured in healthy donors.<sup>6,7</sup> Interestingly, incubating healthy volunteers' platelets with patients plasma led to an increase in TF expression in the platelets.<sup>6</sup>

It is worth mentioning in this context that the assessment of the contribution of platelet-associated TF to the prothrombotic status found in COVID-19, as well as in any other clinical settings, needs some technical advice. Indeed, TF-positive platelets are those with the highest mean platelet volume,<sup>8</sup> and this is consistent both in healthy subjects as well as in pathological conditions.<sup>9</sup> This platelet fraction is easily lost during platelet-rich plasma preparation, if this is performed with high-speed centrifugation ( $>100$  g) lasting longer than 10 minutes, as described in the article by Taus *et al*. Therefore, evaluation of TF-positive platelets in whole blood allows a more accurate analysis, avoiding the risk of losing platelets. If platelets need to be analyzed in platelet-rich plasma, it must be representative as much as possible of whole blood to obtain unbiased results; this can be evaluated by assessing the percentage of recovered platelets as well as the mean platelet volume.<sup>8</sup>

Finally, the contribution of the extrinsic pathway to thrombosis in patients with COVID-19 is also supported by recently published data showing a significant increase

For Disclosures, see page XXX.

© 2021 American Heart Association, Inc.

Arterioscler Thromb Vasc Biol is available at [www.ahajournals.org/journal/atvb](http://www.ahajournals.org/journal/atvb)

in circulating extracellular vesicle-TF activity that is associated with severity of disease and mortality.<sup>10</sup> Notably, these findings complement well with the evidence that the TF-positive microvesicles derive mainly from platelets and erythrocytes, further highlighting the role of platelet activation in the thrombotic events described in patients with COVID-19.<sup>6</sup>

In conclusion, we believe that the very interesting data reported by Taus *et al* could be reinterpreted with a different perspective taking also into account the recently proposed interdependency of the intrinsic and extrinsic pathway on the fibrin formation process in flowing blood.<sup>11</sup> Indeed, while not excluding a potential role of polyphosphates in the activation of FXII, as pointed out by the authors, it could be also hypothesized that the increased FXII activity observed is the result of the activation of the extrinsic pathway supported by increased TF-positive platelet levels measured in patients with COVID-19.

## ARTICLE INFORMATION

### Affiliations

Centro Cardiologico Monzino IRCCS, Milan, Italy (M.B., P.C., E.T., M.C.). Department of Pharmaceutical Sciences, Università degli Studi di Milano, Italy (M.C.).

### Sources of Funding

None.

### Disclosures

None.

## REFERENCES

1. Taus F, Salvagno G, Canè S, Fava C, Mazzaferri F, Carrara E, Petrova V, Barouni RM, Dima F, Dalbeni A, et al. Platelets promote thromboinflammation

in SARS-CoV-2 pneumonia. *Arterioscler Thromb Vasc Biol.* 2020;40:2975–2989. doi: 10.1161/ATVBAHA.120.315175

2. Venter C, Bezuidenhout JA, Laubscher GJ, Lourens RJ, Steenkamp J, Kell DB, Pretorius E. Erythrocyte, platelet, serum ferritin, and p-selectin pathophysiology implicated in severe hypercoagulation and vascular complications in COVID-19. *Int J Mol Sci.* 2020;21:8234.
3. Manne BK, Denorme F, Middleton EA, Portier I, Rowley JW, Stubben C, Petrey AC, Tolley ND, Guo L, Cody M, et al. Platelet gene expression and function in patients with COVID-19. *Blood.* 2020;136:1317–1329. doi: 10.1182/blood.2020007214
4. Barrett TJ, Lee AH, Xia Y, Lin LH, Black M, Cotzia P, Hochman J, Berger JS. Platelet and vascular biomarkers associate with thrombosis and death in coronavirus disease. *Circ Res.* 2020;127:945–947. doi: 10.1161/CIRCRESAHA.120.317803
5. Zhang S, Liu Y, Wang X, Yang L, Li H, Wang Y, Liu M, Zhao X, Xie Y, Yang Y, et al. SARS-CoV-2 binds platelet ACE2 to enhance thrombosis in COVID-19. *J Hematol Oncol.* 2020;13:120. doi: 10.1186/s13045-020-00954-7
6. Canzano P, Brambilla M, Porro B, Cosentino N, Tortorici E, Vicini S, Poggio P, Cascella V, Pengo MF, Veglia F, et al. Platelet and endothelial activation as potential mechanisms behind the thrombotic complications of COVID-19 patients. *JACC Basic Transl Sci.* 2020;in press
7. Denorme F, Manne BK, Portier I, Petrey AC, Middleton EA, Kile BT, Rondina MT, Campbell RA. COVID-19 patients exhibit reduced procoagulant platelet responses. *J Thromb Haemost.* 2020;18:3067–3073.
8. Brambilla M, Rossetti L, Zara C, Canzano P, Giesen PLA, Tremoli E, Camera M. Do methodological differences account for the current controversy on tissue factor expression in platelets? *Platelets.* 2018;29:406–414. doi: 10.1080/09537104.2017.1327653
9. Brambilla M, Camera M, Colnago D, Marezi G, De Metrio M, Giesen PL, Balduini A, Veglia F, Gertow K, Biglioli P, et al. Tissue factor in patients with acute coronary syndromes: expression in platelets, leukocytes, and platelet-leukocyte aggregates. *Arterioscler Thromb Vasc Biol.* 2008;28:947–953. doi: 10.1161/ATVBAHA.107.161471
10. Rosell A, Havervall S, von Meijenfeldt F, Hisada Y, Aguilera K, Grover SP, Lisman T, Mackman N, Thalin C. Patients with COVID-19 have elevated levels of circulating extracellular vesicle tissue factor activity that is associated with severity and mortality. *Arterioscler Thromb Vasc Biol.* 2020;ATVBAHA120315547. doi: 10.1161/ATVBAHA.120.315547
11. Brouns SLN, van Geffen JP, Campello E, Swieringa F, Spiezia L, van Oerle R, Provenzale I, Verdoold R, Farndale RW, Clemetson KJ, et al. Platelet-primed interactions of coagulation and anticoagulation pathways in flow-dependent thrombus formation. *Sci Rep.* 2020;10:11910. doi: 10.1038/s41598-020-68438-9