Therapeutic Cancer Vaccines Revamping: Technology Advancements and Pitfalls

G. Antonarelli, C. Corti, P. Tarantino, L. Ascione, J. Cortes, P. Romero, E.A. Mittendorf, M.L. Disis, G. Curigliano

PII: S0923-7534(21)04457-4

DOI: https://doi.org/10.1016/j.annonc.2021.08.2153

Reference: ANNONC 745

To appear in: Annals of Oncology

Received Date: 11 May 2021

Revised Date: 21 August 2021

Accepted Date: 29 August 2021

Please cite this article as: Antonarelli G, Corti C, Tarantino P, Ascione L, Cortes J, Romero P, Mittendorf EA, Disis ML, Curigliano G, Therapeutic Cancer Vaccines Revamping: Technology Advancements and Pitfalls, *Annals of Oncology* (2021), doi: https://doi.org/10.1016/j.annonc.2021.08.2153.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2021 European Society for Medical Oncology. Published by Elsevier Ltd. All rights reserved.



1	Therapeutic Cancer Vaccines Revamping: Technology Advancements and Pitfalls
2	
3	Authors: G. Antonarelli 1,2, C. Corti 1,2, P. Tarantino 1,2, L. Ascione 1,2, J. Cortes 3,4, P.
4	Romero 5, E. A. Mittendorf 6, M. L. Disis 7, G. Curigliano 1,2
5	
6	Author Affiliations:
7	1 Division of Early Drug Development for Innovative Therapy, European Institute of Oncology,
8	IRCCS, Milan, Italy
9	2 Department of Oncology and Haematology (DIPO), University of Milan, Milan, Italy
10	3 International Breast Cancer Center (IBCC), Quironsalud Group, Barcelona
11	4 Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain
12	5 Department of Fundamental Oncology, University of Lausanne, Lausanne, Switzerland
13	6 Division of Breast Surgery, Department of Surgery, Brigham and Women's Hospital, Boston,
14	MA, 02115, Breast Oncology Program, Dana-Farber/Brigham and Women's Cancer Center,
15	Boston, MA, 02215
16	7 UW Medicine Cancer Vaccine Institute, University of Washington, Seattle, WA 98109, USA
17	
18	Corresponding author: Prof. Giuseppe Curigliano, MD, PhD, Division of Early Drug
19	Development for Innovative Therapy, European Institute of Oncology, IRCCS, Via Ripamonti
20	435, 20141 Milan, Italy. Phone number: +39 0257489599. E-mail: giuseppe.curigliano@ieo.it.
21	
22	Word count: abstract (171), main text (5019)
23	
24	
25	
26	
27	

29 **Highlights**

- Cancer vaccines have been characterized by positive safety and immunogenicity 30 • 31 profiles but low levels of clinical efficacy
- Novel cancer vaccines strategies entail personalized formulations and effective 32 • combinatorial regimens 33
- 34 Positive momentum from the COVID-19 vaccination campaign can in turn accelerate • 35 cancer vaccine clinical testing
- 36

the second secon

37 Abstract (171)

Cancer vaccines (CVs) represent a long-sought therapeutic and prophylactic immunotherapy
strategy to obtain antigen-specific T-cell responses, and potentially achieve long-term clinical
benefit. However, historically, most CV clinical trials have resulted in disappointing outcomes,
despite promising signs of immunogenicity across most formulations.

42 In the past decade, technologic advances regarding vaccine delivery platforms, tools for 43 immunogenomic profiling and antigen/epitope selection have occurred. Consequently, the 44 ability of CVs to induce tumor-specific and, in some cases, remarkable clinical responses have 45 been observed in early-phase clinical trials. It is notable that the record-breaking speed of 46 vaccine development in response to the coronavirus disease (COVID-19) pandemic mainly 47 relied on manufacturing infrastructures and technological platforms already developed for 48 CVs. In turn, research, clinical data, and infrastructures put in place for the SARS-CoV2 49 pandemic can further speed CV development processes.

50 This review outlines the main technological advancements as well as major issues to tackle 51 in the development of CVs. Possible applications for unmet clinical needs will be described, 52 putting into perspective the future of cancer vaccinology.

53

54 Keywords: Cancer, Vaccines, Immunotherapies, Covid-19, Perspective, Technology,
55 Pandemic

- 56
- 57
- 58
- _ _
- 59
- 60
- 61

62

- 63
- 64

65 Introduction

Cancer immunotherapies (CIs) represent one of the most promising fields in oncology (1).
They aim to enhance immune system recognition of tumor cells, possibly leading to disease
control or survival benefit.

69 CIs include cell therapies, antibodies, cytokines, oncolytic viruses (OVs) and cancer vaccines 70 (CVs) (2) (3). Historically, early CIs attempts had exploited the use of systemic cytokines, 71 which were associated with unfavorable toxicity profiles, limiting clinical applications (4). Over 72 the years, growing knowledge in molecular biology, genomics and cancer immunology has 73 prompted the discovery of novel targets and therapeutic approaches (2) (5) (6). Notable 74 examples are Chimeric Antigen Receptor (CAR) T Cells and Immune Checkpoint Blockade 75 (ICB). Both strategies provided clinical benefits in different malignancies (7) (8) (9) (10), 76 leading to their approval by regulatory agencies as well as to the 2018 Nobel Prize in Medicine 77 (7) (11) (12) (13) (14). Regardless, different CIs are burdened by various escape mechanisms, 78 including antigen-/HLA-loss, metastatic seeding of immunological sanctuaries and 79 unpredictable all-or-none or dissociated responses (15) (16) (17).

CVs have also been tested with the aim of unleashing cancer-specific responses and establishing long-term immunological memory. However, results have been relatively disappointing, with most formulations failing to show clinical benefit and only one CV, Sipuleucel-T (Provenge), being approved by the Food and Drug Administration (FDA) in 2010 (18) (19) (20).

In the past year, the record-breaking speed of vaccine development in response to the coronavirus disease (COVID-19) pandemic relied on manufacturing infrastructure and platforms previously built for CVs (21). New vaccine delivery systems, such as gene-based platforms, have been introduced for the first time into large-scale vaccination campaigns. Their favorable safety, immunogenicity and efficacy profiles have been highlighted by the recordtime approval of BNT162b2 and mRNA-1273, the first two FDA-approved vaccines against COVID-19 (22) (23) (24) (25). This work will focus on the major unresolved issues in cancer vaccinology, providing insights
 concerning technological improvements of different platforms, addressing open areas for
 clinical translation.

95

96 Biological and clinical issues to tackle

97 CVs platforms are classically divided into four types: viral/bacterial-, gene-, peptide-, and cell98 based, as depicted in Figure 1.

99 Most CV clinical trials so far have utilized cellular-, viral- or peptide-based platforms, thanks to 100 pre-existing knowledge regarding safety, immunogenicity and manufacture (18) (28). Until late 101 2014, 451 CV clinical trials had been conducted, with gene-based formulations representing 102 less than 5% (29). Remarkably, the ratio of phase III to II CV clinical trials was as low as 1-to-103 21, highlighting a tight bottleneck in drug development processes (30). In addition, most phase 104 III trials ultimately failed to demonstrate efficacy data (29). For example, the MAGRIT trial 105 failed to show increased disease-free survival (DFS) in patients with surgically-resected non-106 small cell lung cancer (NSCLC) receiving a recombinant anti-MAGE-A3 protein vaccine 107 compared to placebo (31). Similarly, the TeloVac trial did not show improved overall survival 108 (OS) in locally-advanced/metastatic pancreatic patients by adding GV1001, a peptide-based 109 vaccine against telomerase, compared to standard-of-care chemotherapy (32). Altogether, 110 these results possibly originated from non-optimized vaccination strategies, non-ideal antigen 111 selection or greatly unexplored combinatorial strategies.

Nonetheless, despite numerous other CVs failing to prove efficacy in phase III trials, the entire field never came to a halt (33). Indeed, strong evidence originating from virtually all trials across different platforms supported favorable toxicity profiles and immunogenicity (30). These were the two foundational pillars onto which cancer vaccinology resumed, envisioning novel platforms and trial designs to eventually attain positive efficacy results (29) (34). After widespread disenchantments, a rejuvenation of the entire field has been witnessed, driven by key aspects warranting optimization to meet expectations (35).

119

120 **Antigen choice**. There are two classes of antigens (Ag) to target: tumor-associated (TAAs) 121 and tumor-specific. The former are non-mutated proteins, being either overexpressed (i.e. 122 human epidermal growth factor receptor 2, HER2, or telomerase), part of tissue differentiation 123 (i.e. tyrosinase, Melan-A) or cancer-germline (i.e. MAGEs, NY-ESO), and not necessarily 124 tumor specific. In contrast, the latter, comprised of oncoviral antigens and neoantigens 125 (NeoAg), are tumor-specific, as shown in Figure 2. NeoAg arise from nonsynonymous 126 mutations, which can be functionally relevant. Indeed, "driver" mutations confer cell-growth 127 advantages and are clonally selected, as opposed to "passenger" mutations, which do not 128 affect the replication rate, hence are less susceptible to clonal selection (36) (37). Past CV 129 trials preferentially targeted TAAs over NeoAg since TAAs were harder to identify (33). 130 However, since TAAs are non-mutated proteins, immune responses tap into T cell repertoires 131 that are subject to central-/peripheral-tolerance (22).

In contrast, phase I/II clinical trials have shown that immune responses against NeoAg are not subjected to central or peripheral tolerance, alongside favorable toxicity profiles (38) (39) (40) (41). In the past years, faster and cheaper availability of Next Generation Sequencing (NGS) technology and more refined bioinformatic prediction tools enabled clinical testing of personalized CVs targeting tumor-specific NeoAg, yielding promising early clinical data (42).

137

138 **Tumor heterogeneity**. Tumors are not single biological entities, as they evolve under diverse 139 intrinsic and extrinsic pressures mainly imposed by treatment, immunity, metabolism and the 140 tumor microenvironment (TME) (43) (44). Importantly, tumors with a high degree of intratumor 141 heterogeneity (ITH) have been linked to worse prognosis (45) (46) (47) (48) (49) (50). Genetic 142 and/or non-genetic mechanisms feed different types of ITH: spatial-, temporal-, 143 immunological- and behavioral- ITH (43). As a major determinant of therapy resistance, ITH 144 should be a fundamental aspect to consider to better guide therapeutic strategies (44) (51). 145 Theoretically, one approach to reduce ITH would be to conduct CV trials in patients with low-146 tumor burden and in front line settings (52). Similarly to Immune Checkpoint Blockers (ICBs), 147 CVs may elicit more robust immune responses in treatment-naïve patients (53) (54).

Moreover, a bolder and promising proposal is to use CVs for immunoprevention of cancer, particularly in individuals with preneoplastic lesions (55) (56) (57).

150 Importantly, gene-based vaccines have the potential to better target ITH compared to the other 151 platforms by eliciting Ag-specific immune responses against different epitopes (58). This could 152 be achieved by inserting sequences encoding for different NeoAg within each vaccination shot 153 (59). In addition, gene-based platforms potentially allow for indefinite rounds of vaccinations, 154 following tumor evolution over time (60). In this setting, the emergence of immunodominance, 155 whereby immune responses preferentially focus on a fraction of the possible epitopes of an 156 unknown protein, could probably blunt the effect of chasing ITH by inserting multiple 157 sequences for diverse NeoAg, and the eventual emergence of such phenomenon shall be 158 precisely assessed (56) (57).

159

Immunomodulation: from "cold" to "hot" TME. A major hurdle undermining therapeutic CV 160 161 efficacy are immune-evasive mechanisms, particularly in solid tumors (63). Recognized 162 mechanisms are T cell exhaustion in inflamed tumors, lack of T cell infiltration in immune 163 excluded tumors and defective antigen presentation processes in immune desert tumors (64). 164 Recently, novel technologies to unravel TME complexity and heterogeneity have been 165 developed, such as mass cytometry and single-cell -omics (i.e. epigenomics, transcriptomics, 166 metabolomics, proteomics) (59). However, in this context, spatial localization of a given cell within the TME is often lost during sample preparation and sequencing procedure (67). Thus, 167 168 development of novel systems-based approaches of simultaneous single-cell analyses 169 combined with spatial microarchitectural information remains a primary technological 170 challenge (68) (69) (70). Potential strategies to heat up the TME and improve antitumor 171 immunity are summarized in Figure 3.

Of note, interactions between tumor cells and their TME can destroy normal tissue homeostasis and shift the TME towards the metastasis-promoting state. In addition, metastatic tumors growing in different organs may consist of significantly different immune infiltrates as compared with the primary tumor site, triggering differential responses to immunotherapies

(75). For example, comparing the immune infiltrates of patient-matched primary versus
metastatic breast cancers (BCs) evidenced a higher content of Tumor Associated
Macrophages (TAMs) across in metastatic sites, suggesting TAMs as a potential therapeutic
target for metastatic BCs (76).

180 In addition, many studies have focused on how tumor metabolism imposes an 181 immunosuppressive TME status (77). Recognized TME metabolic derangements are, for 182 example, low pH, hypoxia, aerobic glycolysis, fatty acid biosynthesis, accumulation of lactate 183 and kynurenines as well as deprivation of glucose, glutamine and tryptophan (78). Targeting 184 such metabolic alterations has been explored in preclinical models, while human translation 185 studies have mainly investigated the role indoleamine-2,3-dioxygenase (IDO) inhibition (79). 186 IDO is a rate-limiting enzyme upregulated in response to IFN-gamma that catabolizes 187 tryptophan, blunting T cell responses (80). IDO inhibitors have shown limited efficacy when 188 utilized as single agents, thereby numerous clinical trials are ongoing investigating 189 combinations with ICBs or CVs (81) (82).

190 Another key determinant of cancer-immune interactions is the concomitant use of drugs with 191 intrinsic TME-modulator (TMEm) functions (83). One example are cyclin-dependent kinase 192 inhibitors, which have been linked with improved antigen presenting processes and pro-193 inflammatory cytokine secretion, ultimately favoring immune escape via PD-L1 upregulation 194 (84). In order to fully uncover immunological aftermaths of diverse drugs, their TMEm functions 195 need to be tested and validated in disease-specific pre-clinical models (85). For example, the 196 CD73-adenosine axis has recently been recognized as a key druggable immunomodulatory 197 pathway activated in glioblastoma and pancreatic ductal adenocarcinoma murine models, 198 paving the way for clinical testing (i.e. ARC-8, NCT04104672) (77) (78).

Ultimately, recent studies suggest that the gut microbiome plays a critical role in modulating immune responses, also affecting response to ICB therapies. For example, commensal *Prevotella heparinolytica* has been shown to promote IL17-producing cells, accelerating myeloma progression in a preclinical model (88). Moreover, gut colonization with *Firmicutes* and *Faecalibacterium genus* was associated with improved clinical response rates and colitis

in ICB-treated melanoma patients (89). Similarly, the microbiota composition was shown to influence anti-tumor efficacy of neoantigen CVs (90). In the field of pharmacobiotics, *Bifidobacterium longum* supplementation on anti-PD1 therapy was recently suggested to curb tumor growth in a BC murine model (91). Importantly, the impact of the microbiome needs to be assessed across various tumor subtypes, as its effects are likely to differ.

209

210 **Technology advancements and pitfalls**

211 Platform-related Improvements. Substantial technological improvements have affected 212 mainly gene- and viral-based platforms. Gene-based vaccines have historically lagged behind, 213 mainly due to scarce safety data, large-scale manufacturing experience, reduced 214 immunogenicity of early formulations, their instability and poor uptake/specificity (92). 215 However, their use has blossomed in recent years, mainly due to three major areas of 216 technological advancements: structure optimization, novel delivery systems and refined 217 epitope prediction tools (93) (94) (95) (96). mRNA production has been standardized, with 218 simpler manufacture processes obviating the need for cell culture or viral vector production 219 (97). Additionally, mRNA-based platforms in the pipeline allow for quick sequence adaptations 220 in response to emerging resistance mutations (92). Lastly, another advantage of gene-based 221 platforms is that they do not need exogenous, immunogenic cargos, potentially allowing for 222 indefinite dosing/booster shots (92). Concerning RNA-structural optimization, recent 223 technological advances aim at avoiding detrimental immune activation as well as increasing 224 safety, biodistribution and immunogenicity profiles (98). Major breakthroughs have been 225 sequence optimization, allowing for enhanced transcription and in vivo stability, Good 226 Manufacturing Practice grade purification systems to avoid toxic leftovers, and the insertion of 227 modified nucleotides with higher translation capacity and lower immunogenicity (via TLR7 228 avoidance), such as N1-methyl-pseudouridine (1m Ψ) (98). Such modifications may expand 229 the clinical indications of RNA-based vaccines: for example, a non-inflammatory, m1 Ψ -based, 230 vaccine structure design recently showed disease protection in a pre-clinical model of 231 experimental autoimmune encephalitis (99).

232 In addition, mRNA sequences can be utilized not only to encode for tumor-specific antigens, 233 but also immunomodulators (i.e. cytokines, ligands), monoclonal/bispecific antibodies, small 234 interfering RNA, CART constructs, or combinations (93). Indeed, several phase I/II trials are 235 currently testing such approaches (54) (87). For example, mRNA-2752, a lipid nanoparticle 236 (LNP)-loaded with OX40 ligand (OX40L, also known as tumor necrosis factor superfamily, 237 member 4 ligand), IL23 and IL36y, is being tested against several malignancies in combination 238 with durvalumab, demonstrating an acceptable safety profile, pro-inflammatory cytokine 239 release, together with some cases of tumor shrinkage (NCT03739931) (100).

240 In parallel, improvements have recently been made also in the field of viral-vector vaccines 241 (101), which typically utilize either live or non-replicating vectors (102). Major innovations 242 include the introduction of different viral vectors, such as adenoviruses (Ad) (i.e. non-human 243 primate, NHP), parvoviruses (i.e. adeno-associated viruses, AAV) and poxviruses (i.e. 244 Modified vaccinia Ankara) (103). Such platforms allow for remarkable versatility, carrying the 245 genetic information for antigen expression and induce potent T-cell responses (104) (105). A 246 major limitation is the high prevalence of pre-existing immunity against the vector itself, 247 possibly reducing overall efficacy by limiting multiple vaccinations (102). To overcome this, 248 prime/boost approaches based on two different viruses immunologically non-cross-reacting 249 ("heterologous prime/boost") have shown promising results in humans (106) (107) (108). 250 Alternatively, use of serotypes with low-prevalence is also advised (109). Moreover, complex 251 manufacturing pipelines based on cell-culture systems and the possibility of residual viral 252 replication also remain open areas of research and technological development (110).

Moreover, viral-based approaches also pertain OVs (72). Historically, they have been used as *in situ* vaccination agents to elicit immune responses against multiple, unpredicted epitopes, given their natural ability to replicate within cancer cells (72). Notably, the only OV being granted regulatory approval has been Talimogene laherparepvec (T-VEC) (111). Subsequently, efforts have been made to arm OVs with immunomodulating agents, to couple them with immune-stimulating agents and/or to elicit Ag-specific responses (112). Remarkable responses and tumor immune infiltration have been recently documented with

HSV-1 G207 in pediatric high-grade gliomas (113). Moreover, the use of a genetically-modified Maraba Virus (MR1) has been validated in pre-clinical models to boost immune responses when administered after Ad-based vaccination, posing the rationale to test Ad:MR1 primeboost combinations in humans (NCT02285816) (NCT02879760) (114). Ultimately, strategies aiming at eliciting Ag-specific responses via OVs exploit virion-coating with peptides of interest, exploiting either electrostatic forces (i.e. negatively charged virions and positively charged poly-lysine peptides) or membrane-anchoring (98) (99).

267

Novel delivery vehicles. Innovative compounds have been introduced in clinical trials, 268 269 especially for RNA-based platforms, such as protamine combined, lipoplexes (LPX), or LNPs 270 (58) (117). Among them, LNPs stand out as a major nanomedicine advancement, as 271 witnessed by their implementation in the development of COVID-19 vaccines (65) (73). In 272 particular, BNT162b2 and mRNA-1273 exploit LNPs as vectors for spike protein-encoding 273 mRNAs and are currently being administered in worldwide vaccination campaigns (22) (23). 274 For the first time, such gene-based vaccines have been linked with remarkable safety profiles 275 in the general population, as well as in special sub-groups such as cancer patients, pregnant 276 women, and the elderly (26) (119) (120). Remarkably, antibody persistence was also detected 277 up to six months after the completion of the second vaccination boost (25) (86) (87). Briefly, 278 LNPs protect RNA-sequence from degradation and allow for stringent spatial-temporal control. 279 In addition, their lipid/moiety composition could be further modified to promote cell/organ-280 specific targeting and adjuvant properties, further expanding the potential use of gene-based 281 vaccines (97) (123).

282

Bioinformatics and novel antigen prediction tools. NeoAg may be exploited not only to indirectly estimate the likelihood of response to ICBs in certain tumors, but also to design personalized therapeutic CVs (124) (125). To do so, standardized bioinformatic tools able to identify and prioritize possible tumor-specific mutations have been developed (125). However, not all mutations result in neoepitopes that are recognized by the immune system, owing to

288 human leukocyte antigen (HLA) restriction/immunodominance (68) (69). Therefore, HLA 289 typing is also required to foresee potentially immunogenic epitopes (42). HLA class I-binding 290 epitopes are predicted through algorithms and computational approaches trained on peptide-291 binding affinity data (42) (128). Such algorithms have also been tested on mass spectrometry 292 (MS) data of peptides presented on specific mono-allelic HLA-expressing cell lines to increase 293 accuracy (125) (129) (130). Besides MS, methods for high-throughput detection of mutation-294 associated epitopes, such as mass cytometry and T-cell receptor clonotyping, are also being 295 successfully implemented (42). Additionally, recent advances in big data analysis and artificial 296 intelligence are contributing to improve neoepitope prediction (131) (132). In particular, deep 297 learning approaches have been applied to large HLA peptide and genomic datasets from 298 various human tumors (e.g. NetMHCpan, NetMHCIIpan) to create a computational model of 299 antigen presentation (71) (72) (95) (131). Moreover, large-scale cancer proteomic data 300 sharing efforts such as the Clinical Proteomic Tumor Analysis Consortium (CPTAC), the 301 Tumor Neoantigen Selection Alliance (TESLA) and the HLA Ligand Atlas of healthy human 302 tissues will facilitate the enumeration of targetable tumor NeoAg (135) (136).

303 Several obstacles currently make the design of therapies targeting NeoAg difficult. For 304 example, among the vast number of putative NeoAg, only a small fraction is ultimately 305 validated, efficiently presented or shown to be immunogenic (137). In fact, prediction tools are 306 more specific for MHC-I compared to MHC-II molecules, possibly due to a longer sequence 307 and open ends of the latter (42). Also, additional evidence suggests that many tumor-specific 308 epitopes may arise from non-translated sequences, for which most in silico tools have not yet 309 been optimized (138). Lastly, further studies to improve understanding of the factors that can 310 affect NeoAg expression, presentation and immunogenicity are necessary (42).

311

312 Getting Cancer Vaccines to The Clinic

313 **Gene-based CVs**. Clinical results regarding nucleoside-based CVs have been 314 heterogeneous, due to the large number of phase I/II trials enrolling a limited number of 315 patients, diverse primary endpoints and fast-emerging technological advancements (92).

316 In general, gene-based vaccines comprise about 22% of vaccines in preclinical development 317 (37/166) and about 37% of those in clinical development (10/27). Importantly, two of them 318 received FDA-licensing for COVID-19 (139) (140). In this context, two landmark clinical trials 319 targeting TAAs and NeoAg, provided first evidence of efficacy as therapeutic approaches for 320 cancer (40) (137). Indeed, in the phase I Lipo-MERIT trial (NCT02410733), 89 advanced, ICB-321 treated, melanoma patients received mRNA-based CV against up to four TAAs (141). 322 Remarkably, Th1-skewed, polyclonal T cell responses following vaccination were observed, 323 along with synergy with anti-PD-1 in ICB-experienced patients, ultimately resulting in durable 324 responses rates (35% in the combination group). Notably, the RNA was optimized to achieve 325 highest expression in immature dendritic cells (96) and the liposomal delivery system elicited 326 TLR-7-mediated type-I interferon responses, easing T-cell expansion (142). The phase I 327 Individualized Cancer Immunotherapies (IVAC) MUTANOME trial (NCT02035956), testing an 328 RNA-based platform targeting two TAAs and up to ten NeoAg in thirteen advanced melanoma 329 patients, showed the emergence of T-cell responses in vaccinated patients, with a reduction 330 in the cumulative rate of metastatic events (40). Of note, polyclonal T-cell responses were 331 detected in all patients in both CD4 and CD8 compartments, and evidence of synergy with 332 ICB (40).

Altogether, these trials provided evidence about heavily pre-treated, high-tumor burden, patient populations, highlighting the potential of gene-based platforms and their synergism with traditional immunotherapies.

336

Viral-vector CVs. Several viral vectors have been evaluated in CV clinical studies (143). For example, a Gorilla Adenovirus (GAd)-derived, NeoAg-based CV was recently shown to synergize with ICB in preclinical tumor models, leading up to disease eradication (144) (145). Importantly, viral vectors can be armed with multiple antigens of interest, such as PSA/MUC-1/brachyury in metastatic castration-resistant prostate cancer patients (NCT03481816), or with regulated immunomodulator expression, such as gene-switches for IL-12 delivery in a preclinical model of glioma (146) (147). In addition, two NHP Ad vectors are in clinical

development for the delivery of NeoAg CVs: Chimpanzee (ChAd68) and GAd20. Preliminary
results in patients with advanced tumors have demonstrated robust and consistent induction
of CD8 T cells against multiple NeoAg upon vaccination with ChAd68 (Granite, NCT03639714,
NCT03953235).

348 The above-mentioned induction of anti-vector immunity has been overcome by heterologous 349 prime/boost. Such trials elicited higher immune responses than repeated vaccination with an 350 individual viral vector (148). Both self-amplifying RNA and Modified Vaccinia Ankara (MVA) 351 technologies are currently being used to boost NHP Ad vectors in clinical trials 352 (NCT03639714). In this regard, the NHP/MVA prime/boost regimen with two vectors (GAd20 353 and MVA) is currently evaluated with a NeoAg-based vaccine for high microsatellite instable 354 (MSI-H) tumors (NCT04041310). Instead, the Nous-209 vaccine is based on concomitant 355 administration of four viral vectors encoding for 209 shared NeoAg peptides among patients 356 with MSI-H tumors (145).

Adenovirus-vectored vaccines are also being tested to elicit responses in the central nervous system. In this context, a phase I, dose-escalation study was conducted with DNX-2401 (Delta-24-RGD) in 37 patients with recurrent high-grade glioma, resulting in 20% survival at more than three years (149). Overall, the entire field of viral-based CV is advancing thanks to the exploitation of novel viral species and innovative strategies with other vaccination approaches, prompting their application in the oncologic setting.

363

364 Peptide-based CVs. Historically, most peptide-based CVs tested so far in the clinic showed 365 variable signs of immunogenicity and clinical activity (150). Two major improvements in this 366 field have been the introduction of novel adjuvants as well as the use of synthetic long peptides 367 (SLP) (151) (152) (153). As opposed to short peptides, SLPs do not directly bind to MHC 368 class I molecules; indeed they require antigen presenting processing for presentation to 369 cytotoxic T lymphocytes with proper immune-stimulatory co-receptors (154). Moreover, SLPs 370 also allow for multi-epitope targeting, as shown for TAS0314, a peptide containing four TAAs 371 from SART2 and SART3 proteins, in a pre-clinical model of SART2₉₃₋₁₀₁-expressing 372 melanoma (155). Other formulations of this approach are currently undergoing evaluation373 (156).

374 In addition, peptide-based CV formulations can also target patient-specific NeoAg. In this 375 scenario, one of the most advanced CV products is represented by NeoVax, comprised of up 376 to 20 different SLPs with the immunostimulatory adjuvant poly-ICLC (a synthetic dsRNA viral 377 mimic that acts as a TLR3 agonist) (38). Clinical trials in advanced melanoma and 378 glioblastoma have both demonstrated the emergence of polyfunctional, specific, Th1-skewed 379 responses post-vaccination (NCT01970358) (NCT02287428) (37) (134). In the melanoma 380 trial, four patients out of six had no recurrence up to 25 months after vaccination; while the 381 two relapsing patients showed complete tumor regression after ICB therapy (38). These 382 studies highlighted the potential of such peptide-based CV formulations, which are currently 383 tested also in combination with other immunotherapies (NCT02950766) beina 384 (NCT03929029) (42). Moreover, another promising trial is represented by the Phase Ib NT-385 002, assessing a personalized NeoAg CV, NEO-PV-01, targeting up to 20 NeoAg predicted 386 by bioinformatic analysis, as a first line therapy for advanced non-squamous NSCLC with 387 carboplatin, pemetrexed and pembrolizumab (NCT03380871) (158) (159) (160). Authors 388 reported Ag-specific and durable (up to 1 year) immune responses, with approximately 55% 389 of vaccine peptides eliciting measurable immune responses. Remarkably, overall response 390 rates in the intention-to-treat and the vaccination populations were 37% and 57%, respectively 391 (159).

Overall, concerning peptide-based CV formulations, research in the field of adjuvants as well
as in the discovery of ideal antigenic targets is still needed to further improve immunogenicity
and, ultimately, clinical efficacy.

395

Biotech and industrial perspectives. A key aspect of CV development efforts is the capacity
of making early and objective treatment choices in order to select ideal candidates, a specific
platform, eventual combinatorial agents and vaccination schedules (161).

399 For RNA-based CVs, different aspects still need to be thoroughly assessed to boost their 400 efficacy. One is their design, as LNP:mRNA mass ratio can be adjusted (from 10:1 to 30:1), 401 implying; for example, a significant amount of LNP for multi-Ag candidates in a given dose 402 (42) (98). Moreover, differences in safety and immunogenicity profiles between non-replicating 403 mRNA and self-amplifying mRNA vaccine sequences are largely unknown, and may have 404 implications to improve sequence optimizations upon iterative development schemes (97). In 405 addition early-phase clinical trials need to precisely capture the inflammatory components of 406 the different mRNA vaccine formulations, given that several intracellular immune sensors are 407 activated by RNA, in order to optimize the benefit (immunogenicity, efficacy) while reducing 408 the risk (safety) profiles (162). In this regard, safety and tolerability may limit multi-antigen 409 approaches, and here pre-clinical studies will be crucial for development. Lastly, limited data 410 exist on repeated administrations of mRNA vaccines in humans (161). As the entire field 411 accrues more data from human studies and current COVID-19 vaccination programs, potential 412 long-term safety and immunogenicity issues will need to be accurately collected and critically 413 discussed (86) (118) (140).

414 Considering biotech and industrial implications, viral-based CVs face different issues. 415 Importantly, manufacturing pipelines are more complex and require laborious cell-culture 416 methods imposing complex purification and microbiological constraints (164). Moreover, 417 replication-defective viruses need thorough (pre-)clinical validation regarding their replication 418 capacity and the absence of systemic disease manifestations in frail sub-populations, such as 419 immunocompromised individuals (165). In addition, vaccine-related disease enhancement has 420 been described in some pre-clinical models for SARS-CoV or respiratory syncytial virus 421 vaccines, and must be always considered (166) (167). Moreover, punctual reports concerning 422 toxicities as novel viruses are introduced in clinical practice must be thoroughly monitored by 423 regulatory agencies (168).

Key aspects to consider in the development of a vaccination strategy, given a certain kind of mutation obtained from sequencing studies are summarized in **Figure 4** (41) (118). For example, tumor mutational burden (TMB) high tumors should benefit more from multiple,

427 personalized rounds of vaccinations against different predicted neo-epitopes, possibly in 428 combination with ICBs or TME-altering drugs (83). Instead, off-the-shelf vaccination strategies 429 should be envisioned in TMB-low tumors both prophylactically, against viral-associated 430 epitopes or mutations conferring resistance to chronic concomitant therapies, or 431 therapeutically, against known driver mutations. In general, these considerations must be 432 continuously updated and re-evaluated considering the fast-changing technologic advances 433 and data procurement.

434

435 Future Perspectives

436 Since most CV clinical trials are small scale, data originating from them will need to be 437 standardized in order to allow comparability and build large-scale reference datasets 438 regarding immunogenicity, biomarkers and efficacy readouts (18). In this way, patient 439 stratification could identify subgroups potentially gaining benefit from CV programs (170). In 440 parallel, attempts to translate predictive biomarkers identified from ongoing research in ICB-441 treated patients may also help identifying patients most likely to respond to CVs. For example, 442 a recent meta-analysis of transcriptomic and clinical data from >1000 ICB-treated patients 443 across various malignancies, identified clonal TMB, CXCL9/CXCL13 expression, CCND1 444 amplification and TRAF2 loss being predictive of ICB-response (171).

445 Another concern is to match vaccination strategies with tumor biology/genetics. In fact, CVs 446 can either be utilized as ready-to-use, off-the-shelf drugs, or as personalized products based 447 on sequencing data (18). Cancer biology and data originating from large longitudinal 448 sequencing studies in multiple malignancies must instruct different vaccination strategies in 449 different clinical settings (169). For example, vaccination strategies relying on products 450 tailored on sequencing data should be most suited for patients carrying biomarkers predicting 451 positive response to ICBs (171). Conversely, tumors showing high oncogene addiction are 452 characterized by driver mutations, which fall in specific loci and harbour few recurrent genetic 453 alterations (18). Theoretically, these tumors could benefit from vaccination strategies aimed 454 at targeting such recurrent driver mutations, possibly in the (neo)adjuvant setting, by means

455 of ready-to-use, off-the-shelf CV products released on histologic information. In this regard, in 456 2019 Moderna & Merck opened a phase I clinical trial with the aim of targeting the four most 457 common KRAS alterations in NSCLC, pancreatic and CRC patients by means of a mRNA-458 based vaccine with or without pembrolizumab (NCT03948763). Importantly, the use of off-the-459 shelf CV products can also be applied in the prophylactic setting in patients under chronic 460 treatment with targeted or endocrine therapies, to avoid recurrent mutations causing loss of 461 response, as well as in patients with genetic cancer syndromes (i.e. FAP, Lynch Syndrome) 462 (172) (173) (174). Moreover, driver mutations, and their therapy-resistance mutation, may not 463 be the solely targets in oncogene-driven tumors. In fact, oncogenic pathways often co-operate 464 with other mutant proteins to promote disease progression (175) (176). For example, KRAS 465 exploits mutant TP53 in fostering disease growth in a pre-clinical model of pancreatic ductal 466 adenocarcinoma (87).

467 The choice of combinatorial agents must consider tumor biology and TME-specific 468 derangements, as previously discussed, together with the clinical setting in which these are 469 introduced. This is because any combinatorial drug comes at the cost of possible added side 470 effects, and the toxicity/benefit ratio varies from patient to patient and from early to advanced settings. For example, in early stages, combinatorial regimens should aim at increasing CV 471 472 immunogenicity and foster the formation and persistence memory T cell subsets. In the 473 metastatic setting, instead, combinatorial drugs should achieve disease control in the short to 474 medium term, allowing CVs to stimulate T-cell specific immunity and/or re-invigorate ICB-475 driven responses.

476 Of note, growing evidence suggests a higher benefit of ICB-therapies in circulating tumor DNA 477 (ctDNA)-positive patients in several malignancies, with a favorable prognostic role of ctDNA 478 seroconversion rates (177)(178). Consolidated data concerning eventual 479 prognostic/predictive roles of this biomarker could possibly instruct for the use of ctDNA 480 seroconversion rate as a primary endpoint of (neo)adjuvant CV clinical trials, alongside long-481 term survival data (i.e. PFS, OS) supporting eventual regulatory approvals.

Finally, in order to build positive momentum in the cancer vaccination field, four aspects should be strengthened: research & technology, clinical scenario, trial comparability and global preparedness (**Figure 5**).

485

486 **Conclusions**

Suboptimal clinical trial designs, the use of CVs as single agents, sometimes with weak antigens, as well as the enrolment of advanced, heavily pre-treated patients, have been just some of the reasons that led to poor clinical trial results so far (179). Nonetheless, enormous progress has been made in both oncology and vaccinology (53) (65) (71).

491 First, unprecedented in-depth, running, characterization of cancer genetics, including genetic 492 determinants of therapy resistance, and the introduction of novel immunotherapies or TME-493 altering drug to combine CVs with in future clinical trials, have broadened the spectrum of both 494 TAA or NeoAg targets (180). Moreover, bioinformatic prediction tools are becoming more 495 refined with the growing availability of tumor mutations alongside HLA sequencing population 496 libraries (i.e. IPD-IMGT/HLA Database) (181) (182) (183). Second, technological advances in 497 the vaccinology field are occurring, especially regarding formulations (gene-, viral-, peptide-498 based) and delivery systems, contributing to the time-record introduction of effective vaccines 499 in the COVID-19 pandemic (21). In this scenario, research and innovation efforts to address 500 COVID-19 provided large-scale evidence about the favorable safety and immunogenicity 501 profiles of these vaccine platform technologies and point to the need to accompany CVs with 502 interventions at the level of the suppressive TME. This momentum could, in turn, speed up 503 the development of CVs employing novel technologies, which are showing promising, 504 although immature, signs of efficacy in early Phase I trials (37) (39) (131).

505 Importantly, the choice of ideal endpoints to allow for a hypothetical regulatory approval of 506 such agents remains a matter of debate, as whether safety profiles should be considered 507 according to a platform-based approach or to the single vaccine product. For these reasons, 508 testing as per classical phase I-III schedule is still to be addressed.

509 Overall, there is rising optimism that technological advancements, data accumulating from 510 worldwide vaccination campaigns, strengthened production processes and, importantly, 511 clinical results from ongoing phase II/III trials will clarify the ultimate role of CVs in cancer 512 treatment in the ensuing years.

Journal Prevention

Funding: This work did not receive any specific grant from funding agencies in the public,commercial, or not-for-profit sectors.

515

Acknowledgements: GA contributed to the literature search, conception, design of the article and drafted the first version of the manuscript. CC, PT and LA contributed to the literature search, conception, design of the article and provided critical revisions of the manuscript. GC contributed to conception and design of the article, provided critical revisions of the manuscript and supervision. Figure 1, Figure 2 and Figure 3 were created with biorender.com. All the authors provided critical revisions of the manuscript and final approval to the submitted work.

523 **Competing Interests:** GA, CC, LA and PT have no potential conflicts of interest to disclose. 524 JC reports consulting/advisor fees or honoraria from: Roche, Celgene, Cellestia, AstraZeneca, 525 Biothera Pharmaceutical, Merus, Seattle Genetics, Daiichi Sankyo, Erytech, Athenex, 526 Polyphor, Lilly, Servier, Merck Sharp&Dohme, GSK, Novartis, Eisai, Pfizer, Samsung Bioepis, 527 Ariad pharmaceuticals, Baxalta GMBH/Servier Affaires, Bayer healthcare, Guardanth health, 528 Piqur Therapeutics, Puma C, Queen Mary University of London. PR serves as consultant for 529 MaxiVax, Enterome and Inotrem. EAM is on the SAB for Astra-Zeneca/Medimmune, Celgene, 530 Genentech, Genomic Health, Merck, Peregrine Pharmaceuticals, SELLAS Lifescience, and 531 Tapimmune and has clinical trial support to her former institution (MDACC) from Astra-Zeneca/Medimmune, EMD-Serono, Galena Biopharma and Genentech as well as Genentech 532 533 support to a SU2C grant, as well as sponsored Research Support to the laboratory from GSK 534 and Eli Lilly. MLD has stock ownership of VentiRX and Epithany. MLD has received grant 535 funding from EMD Serono, VentiRx, Seattle Genetics, and Celgene. GC served as consultant 536 or advisor for Roche, Lilly, and Bristol-Myers Squibb, served on the speaker's bureau for 537 Roche, Pfizer, and Lilly, received travel funding from Pfizer and Roche, and received honoraria 538 from Roche, Pfizer, Lilly, Novartis, and SEAGEN, all outside the submitted work. EAM has 539 been compensated for participation on SABs for Exact Sciences, Merch and Roche; 540 uncompensated participation on steering committees for BMS, Lilly and Roche.

541 **References**

- 542 1. Couzin-Frankel J. Breakthrough of the year 2013. Cancer immunotherapy. Science. 2013
 543 Dec 20;342(6165):1432–3.
- 544 2. Chen DS, Mellman I. Oncology Meets Immunology: The Cancer-Immunity Cycle.
 545 Immunity. 2013 Jul;39(1):1–10.
- Zhang Y, Zhang Z. The history and advances in cancer immunotherapy: understanding
 the characteristics of tumor-infiltrating immune cells and their therapeutic implications.
 Cellular & Molecular Immunology. 2020 Aug;17(8):807–21.
- Berraondo P, Sanmamed MF, Ochoa MC, Etxeberria I, Aznar MA, Pérez-Gracia JL, et al.
 Cytokines in clinical cancer immunotherapy. British Journal of Cancer. 2019
 Jan;120(1):6–15.
- 552 5. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from 553 immunosurveillance to tumor escape. Nat Immunol. 2002 Nov;3(11):991–8.
- Marshall HT, Djamgoz MBA. Immuno-Oncology: Emerging Targets and Combination
 Therapies. Front Oncol [Internet]. 2018 [cited 2021 Mar 7];8. Available from: https://www.frontiersin.org/articles/10.3389/fonc.2018.00315/full
- Neelapu SS, Locke FL, Bartlett NL, Lekakis LJ, Miklos DB, Jacobson CA, et al.
 Axicabtagene Ciloleucel CAR T-Cell Therapy in Refractory Large B-Cell Lymphoma.
 New England Journal of Medicine. 2017 Dec 28;377(26):2531–44.
- 8. Maude SL, Laetsch TW, Buechner J, Rives S, Boyer M, Bittencourt H, et al.
 Tisagenlecleucel in Children and Young Adults with B-Cell Lymphoblastic Leukemia. N
 Engl J Med. 2018 Feb 1;378(5):439–48.

- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved
 Survival with Ipilimumab in Patients with Metastatic Melanoma. New England Journal
 of Medicine. 2010 Aug 19;363(8):711–23.
- 10. Wolchok JD, Chiarion-Sileni V, Gonzalez R, Rutkowski P, Grob J-J, Cowey CL, et al.
 Overall Survival with Combined Nivolumab and Ipilimumab in Advanced Melanoma.
 New England Journal of Medicine. 2017 Oct 5;377(14):1345–56.
- 569 11. Waldman AD, Fritz JM, Lenardo MJ. A guide to cancer immunotherapy: from T cell basic
 570 science to clinical practice. Nat Rev Immunol. 2020 Nov;20(11):651–68.
- 571 12. Dougan M, Dranoff G, Dougan SK. Cancer Immunotherapy: Beyond Checkpoint
 572 Blockade. Annual Review of Cancer Biology. 2019;3(1):55–75.
- 573 13. First-Ever CAR T-cell Therapy Approved in U.S. | Cancer Discovery [Internet]. [cited 2021
 574 Apr 27]. Available from: https://cancerdiscovery.aacrjournals.org/content/7/10/OF1
- 575 14. The Nobel Prize in Physiology or Medicine 2018 [Internet]. NobelPrize.org. [cited 2021
 576 Apr 26]. Available from: https://www.nobelprize.org/prizes/medicine/2018/press577 release/
- 578 15. Majzner RG, Mackall CL. Tumor Antigen Escape from CAR T-cell Therapy. Cancer
 579 Discov. 2018 Oct 1;8(10):1219–26.
- 580 16. Paulson KG, Voillet V, McAfee MS, Hunter DS, Wagener FD, Perdicchio M, et al. Acquired
 581 cancer resistance to combination immunotherapy from transcriptional loss of class I
 582 HLA. Nature Communications. 2018 Sep 24;9(1):3868.
- 583 17. Vago L, Perna SK, Zanussi M, Mazzi B, Barlassina C, Stanghellini MTL, et al. Loss of
 584 Mismatched HLA in Leukemia after Stem-Cell Transplantation. New England Journal of
 585 Medicine. 2009 Jul 30;361(5):478–88.

- 18. Jou J, Harrington KJ, Zocca M-B, Ehrnrooth E, Cohen EEW. The Changing Landscape of
- 587 Therapeutic Cancer Vaccines—Novel Platforms and Neoantigen Identification. Clin 588 Cancer Res. 2021 Feb 1;27(3):689–703.
- 19. Rosenberg SA, Yang JC, Restifo NP. Cancer immunotherapy: moving beyond current
 vaccines. Nat Med. 2004 Sep;10(9):909–15.
- 20. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, et al. Sipuleucel-T
 immunotherapy for castration-resistant prostate cancer. N Engl J Med. 2010 Jul
 29;363(5):411–22.
- 594 21. Wherry EJ, Jaffee EM, Warren N, D'Souza G, Ribas A. How Did We Get a COVID-19
 595 Vaccine in Less Than 1 Year? Clin Cancer Res. 2021 Apr 15;27(8):2136–8.
- 596 22. Hollingsworth RE, Jansen K. Turning the corner on therapeutic cancer vaccines. npj
 597 Vaccines. 2019 Feb 8;4(1):1–10.
- 23. Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, et al. Safety and
 Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. N Engl J Med. 2020 Dec
 31;383(27):2603–15.
- 601 24. Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, et al. Efficacy and Safety
 602 of the mRNA-1273 SARS-CoV-2 Vaccine. N Engl J Med. 2021 Feb 4;384(5):403–16.
- 603 25. Dagan N, Barda N, Kepten E, Miron O, Perchik S, Katz MA, et al. BNT162b2 mRNA Covid-604 19 Vaccine in a Nationwide Mass Vaccination Setting. New England Journal of Medicine 605 [Internet]. 2021 Feb 24 [cited 2021 Apr 27]; Available from: https://www.nejm.org/doi/10.1056/NEJMoa2101765 606
- 607 26. Shimabukuro TT, Kim SY, Myers TR, Moro PL, Oduyebo T, Panagiotakopoulos L, et al.
 608 Preliminary Findings of mRNA Covid-19 Vaccine Safety in Pregnant Persons. N Engl J
 609 Med. 2021 Apr 21;

- 27. Hu Z, Ott PA, Wu CJ. Towards personalized, tumour-specific, therapeutic vaccines for
 cancer. Nat Rev Immunol. 2018 Mar;18(3):168–82.
- 612 28. Pollard AJ, Bijker EM. A guide to vaccinology: from basic principles to new developments.
 613 Nat Rev Immunol. 2021 Feb;21(2):83–100.
- Shemesh CS, Hsu JC, Hosseini I, Shen B-Q, Rotte A, Twomey P, et al. Personalized
 Cancer Vaccines: Clinical Landscape, Challenges, and Opportunities. Mol Ther. 2021
 Feb 3;29(2):555–70.
- 30. Tan AC, Goubier A, Kohrt HE. A quantitative analysis of therapeutic cancer vaccines in
 phase 2 or phase 3 trial. J Immunother Cancer. 2015;3:48.
- 31. Vansteenkiste JF, Cho BC, Vanakesa T, De Pas T, Zielinski M, Kim MS, et al. Efficacy of
 the MAGE-A3 cancer immunotherapeutic as adjuvant therapy in patients with resected
 MAGE-A3-positive non-small-cell lung cancer (MAGRIT): a randomised, double-blind,
 placebo-controlled, phase 3 trial. Lancet Oncol. 2016 Jun;17(6):822–35.
- 32. Middleton G, Silcocks P, Cox T, Valle J, Wadsley J, Propper D, et al. Gemcitabine and
 capecitabine with or without telomerase peptide vaccine GV1001 in patients with locally
 advanced or metastatic pancreatic cancer (TeloVac): an open-label, randomised, phase
 3 trial. Lancet Oncol. 2014 Jul;15(8):829–40.
- 33. Mocellin S, Mandruzzato S, Bronte V, Lise M, Nitti D. Part I: Vaccines for solid tumours.
 The Lancet Oncology. 2004 Nov 1;5(11):681–9.
- 34. Morse MA, Chui S, Hobeika A, Lyerly HK, Clay T. Recent developments in therapeutic
 cancer vaccines. Nature Clinical Practice Oncology. 2005 Feb;2(2):108–13.
- 35. Romero P, Banchereau J, Bhardwaj N, Cockett M, Disis ML, Dranoff G, et al. The Human
 Vaccines Project: A roadmap for cancer vaccine development. Sci Transl Med. 2016
 Apr 13;8(334):334ps9.

- 36. Bozic I, Antal T, Ohtsuki H, Carter H, Kim D, Chen S, et al. Accumulation of driver and
 passenger mutations during tumor progression. PNAS. 2010 Oct 26;107(43):18545–
 50.
- 637 37. Campbell PJ, Getz G, Korbel JO, Stuart JM, Jennings JL, Stein LD, et al. Pan-cancer
 638 analysis of whole genomes. Nature. 2020 Feb;578(7793):82–93.
- 639 38. Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, et al. An immunogenic personal
 640 neoantigen vaccine for patients with melanoma. Nature. 2017 Jul 13;547(7662):217–
 641 21.
- 39. Carreno BM, Magrini V, Becker-Hapak M, Kaabinejadian S, Hundal J, Petti AA, et al.
 Cancer immunotherapy. A dendritic cell vaccine increases the breadth and diversity of
 melanoma neoantigen-specific T cells. Science. 2015 May 15;348(6236):803–8.
- 40. Sahin U, Derhovanessian E, Miller M, Kloke B-P, Simon P, Löwer M, et al. Personalized
 RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer.
 Nature. 2017 Jul 13;547(7662):222–6.
- 41. Hilf N, Kuttruff-Coqui S, Frenzel K, Bukur V, Stevanović S, Gouttefangeas C, et al. Actively
 personalized vaccination trial for newly diagnosed glioblastoma. Nature. 2019
 Jan;565(7738):240–5.
- 42. Blass E, Ott PA. Advances in the development of personalized neoantigen-based
 therapeutic cancer vaccines. Nat Rev Clin Oncol. 2021 Jan 20;
- 43. Vitale I, Shema E, Loi S, Galluzzi L. Intratumoral heterogeneity in cancer progression and
 response to immunotherapy. Nat Med. 2021 Feb;27(2):212–24.
- 44. Dagogo-Jack I, Shaw AT. Tumour heterogeneity and resistance to cancer therapies.
 Nature Reviews Clinical Oncology. 2018 Feb;15(2):81–94.

657	45. Vitale I, Sistigu A, Manic G, Rudqvist N-P, Trajanoski Z, Galluzzi L. Mutational and
658	Antigenic Landscape in Tumor Progression and Cancer Immunotherapy. Trends Cell
659	Biol. 2019 May;29(5):396–416.

- 46. Hua X, Zhao W, Pesatori AC, Consonni D, Caporaso NE, Zhang T, et al. Genetic and
 epigenetic intratumor heterogeneity impacts prognosis of lung adenocarcinoma. Nat
 Commun. 2020 May 18;11(1):2459.
- 47. Patel AP, Tirosh I, Trombetta JJ, Shalek AK, Gillespie SM, Wakimoto H, et al. Single-cell
 RNA-seq highlights intratumoral heterogeneity in primary glioblastoma. Science. 2014
 Jun 20;344(6190):1396–401.
- 48. Lin D-C, Mayakonda A, Dinh HQ, Huang P, Lin L, Liu X, et al. Genomic and Epigenomic
 Heterogeneity of Hepatocellular Carcinoma. Cancer Res. 2017 May 1;77(9):2255–65.
- 49. Li S, Garrett-Bakelman FE, Chung SS, Sanders MA, Hricik T, Rapaport F, et al. Distinct
 evolution and dynamics of epigenetic and genetic heterogeneity in acute myeloid
 leukemia. Nat Med. 2016 Jul;22(7):792–9.
- 50. Dentro SC, Leshchiner I, Haase K, Tarabichi M, Wintersinger J, Deshwar AG, et al.
 Characterizing genetic intra-tumor heterogeneity across 2,658 human cancer genomes.
 Cell. 2021 Apr 15;184(8):2239-2254.e39.
- 51. Marusyk A, Janiszewska M, Polyak K. Intratumor Heterogeneity: The Rosetta Stone of
 Therapy Resistance. Cancer Cell. 2020 Apr 13;37(4):471–84.
- 52. Kim SI, Cassella CR, Byrne KT. Tumor Burden and Immunotherapy: Impact on Immune
 Infiltration and Therapeutic Outcomes. Front Immunol [Internet]. 2021 Feb 1 [cited 2021
- 678 Apr 27];11. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7882695/
- 53. Marra A, Viale G, Curigliano G. Recent advances in triple negative breast cancer: the
 immunotherapy era. BMC Med. 2019 May 9;17(1):90.

- 54. Chiang AC, Herbst RS. Frontline immunotherapy for NSCLC the tale of the tail. Nature
 Reviews Clinical Oncology. 2020 Feb;17(2):73–4.
- 55. Enokida T, Moreira A, Bhardwaj N. Vaccines for immunoprevention of cancer. J Clin
 Invest. 2021 May 3;131(9).
- 56. Lollini P-L, Nicoletti G, Landuzzi L, De Giovanni C, Nanni P. Immunoprevention and
 immunotherapy of mammary carcinoma. Breast J. 2010 Oct;16 Suppl 1:S39-41.
- 57. Lohmueller JJ, Ham JD, Kvorjak M, Finn OJ. mSA2 affinity-enhanced biotin-binding CAR
 T cells for universal tumor targeting. Oncoimmunology. 2017;7(1):e1368604.
- 58. Beck JD, Reidenbach D, Salomon N, Sahin U, Türeci Ö, Vormehr M, et al. mRNA
 therapeutics in cancer immunotherapy. Molecular Cancer. 2021 Apr 15;20(1):69.
- 59. Akram A, Inman RD. Immunodominance: a pivotal principle in host response to viral
 infections. Clin Immunol. 2012 May;143(2):99–115.
- 60. Draper SJ, Heeney JL. Viruses as vaccine vectors for infectious diseases and cancer. Nat
 Rev Microbiol. 2010 Jan;8(1):62–73.
- 695 61. Tokuyasu TA, Huang J-D. A primer on recent developments in cancer immunotherapy, 696 with a focus on neoantigen vaccines. Journal of Cancer Metastasis and Treatment 697 [Internet]. 2018 Available Jan 18 [cited 2021 Apr 27];4. from: 698 https://jcmtjournal.com/article/view/2355
- 62. Schreiber H, Wu TH, Nachman J, Kast WM. Immunodominance and tumor escape. Semin
 Cancer Biol. 2002 Feb;12(1):25–31.
- 63. Giraldo NA, Sanchez-Salas R, Peske JD, Vano Y, Becht E, Petitprez F, et al. The clinical
 role of the TME in solid cancer. Br J Cancer. 2019 Jan;120(1):45–53.

- 64. Hegde PS, Chen DS. Top 10 Challenges in Cancer Immunotherapy. Immunity. 2020 Jan
 14;52(1):17–35.
- 65. Hu Y, An Q, Sheu K, Trejo B, Fan S, Guo Y. Single Cell Multi-Omics Technology:
 Methodology and Application. Front Cell Dev Biol [Internet]. 2018 [cited 2021 Apr 27];6.
- 707 Available from: https://www.frontiersin.org/articles/10.3389/fcell.2018.00028/full
- 710 67. Santegoets SJ, van Ham VJ, Ehsan I, Charoentong P, Duurland CL, van Unen V, et al.

The Anatomical Location Shapes the Immune Infiltrate in Tumors of Same Etiology and

712 Affects Survival. Clin Cancer Res. 2019 Jan 1;25(1):240–52.

711

- 68. Allam M, Cai S, Coskun AF. Multiplex bioimaging of single-cell spatial profiles for precision
 cancer diagnostics and therapeutics. npj Precision Oncology. 2020 May 1;4(1):1–14.
- 69. Gohil SH, lorgulescu JB, Braun DA, Keskin DB, Livak KJ. Applying high-dimensional
 single-cell technologies to the analysis of cancer immunotherapy. Nat Rev Clin Oncol.
 2021 Apr;18(4):244–56.
- 70. González-Silva L, Quevedo L, Varela I. Tumor Functional Heterogeneity Unraveled by
 scRNA-seq Technologies. Trends in Cancer. 2020 Jan;6(1):13–9.
- 720 71. Duan Q, Zhang H, Zheng J, Zhang L. Turning Cold into Hot: Firing up the Tumor
 721 Microenvironment. Trends in Cancer. 2020 Jul 1;6(7):605–18.
- 722 72. Chiocca E, Rabkin S. Oncolytic Viruses and Their Application to Cancer Immunotherapy.
 723 Cancer Immunol Res. 2014 Apr;2(4):295–300.
- 73. Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets:
 cancer as a paradigm. Nat Immunol. 2010 Oct;11(10):889–96.

726 74. Escobar G, Barbarossa L, Barbiera G, Norelli M, Genua M, Ranghetti A, et al. Interferon
727 gene therapy reprograms the leukemia microenvironment inducing protective immunity
728 to multiple tumor antigens. Nat Commun. 2018 24;9(1):2896.

729 75. Quail D, Joyce J. Microenvironmental regulation of tumor progression and metastasis. Nat
730 Med. 2013 Nov;19(11):1423–37.

731 76. Zhu L, Narloch JL, Onkar S, Joy M, Broadwater G, Luedke C, et al. Metastatic breast
732 cancers have reduced immune cell recruitment but harbor increased macrophages
733 relative to their matched primary tumors. J Immunother Cancer. 2019 Oct 18;7(1):265.

734 77. Bader JE, Voss K, Rathmell JC. Targeting Metabolism to Improve the Tumor
735 Microenvironment for Cancer Immunotherapy. Molecular Cell. 2020 Jun 18;78(6):1019–
736 33.

737 78. Pavlova NN, Thompson CB. THE EMERGING HALLMARKS OF CANCER
738 METABOLISM. Cell Metab. 2016 Jan 12;23(1):27–47.

739 79. Labadie BW, Bao R, Luke JJ. Reimagining IDO Pathway Inhibition in Cancer
740 Immunotherapy via Downstream Focus on the Tryptophan–Kynurenine–Aryl
741 Hydrocarbon Axis. Clin Cancer Res. 2019 Mar 1;25(5):1462–71.

742 80. Zhai L, Spranger S, Binder DC, Gritsina G, Lauing KL, Giles FJ, et al. Molecular Pathways:
743 Targeting IDO1 and Other Tryptophan Dioxygenases for Cancer Immunotherapy. Clin
744 Cancer Res. 2015 Dec 15;21(24):5427–33.

81. Naour JL, Galluzzi L, Zitvogel L, Kroemer G, Vacchelli E. Trial watch: IDO inhibitors in
cancer therapy. Oncolmmunology. 2020 Jan 1;9(1):1777625.

Redman JM, Steinberg SM, Gulley JL. Quick efficacy seeking trial (QuEST1): a novel
 combination immunotherapy study designed for rapid clinical signal assessment

749 metastatic castration-resistant prostate cancer. J Immunother Cancer. 2018 Dec750 1;6(1):91.

- 83. Petroni G, Buqué A, Zitvogel L, Kroemer G, Galluzzi L. Immunomodulation by targeted
 anticancer agents. Cancer Cell. 2021 Mar 8;39(3):310–45.
- 84. Zhang J, Bu X, Wang H, Zhu Y, Geng Y, Nihira NT, et al. Cyclin D-CDK4 kinase
 destabilizes PD-L1 via cullin 3-SPOP to control cancer immune surveillance. Nature.
 2018 Jan 4;553(7686):91–5.
- 85. Olson B, Li Y, Lin Y, Liu ET, Patnaik A. Mouse Models for Cancer Immunotherapy
 Research. Cancer Discov. 2018 Nov;8(11):1358–65.
- 86. Bendell JC, Manji GA, Pant S, Lai DW, Colabella J, Berry W, et al. A phase I study to
 evaluate the safety and tolerability of AB680 combination therapy in participants with
 gastrointestinal malignancies. JCO. 2020 Feb 1;38(4_suppl):TPS788–TPS788.
- 761 87. Kim MP, Li X, Deng J, Zhang Y, Dai B, Allton KL, et al. Oncogenic KRAS recruits an
 762 expansive transcriptional network through mutant p53 to drive pancreatic cancer
 763 metastasis. Cancer Discov [Internet]. 2021 Jan 1 [cited 2021 Apr 18]; Available from:
 764 https://cancerdiscovery.aacrjournals.org/content/early/2021/03/31/2159-8290.CD-20-
- 765 1228
- 88. Calcinotto A, Brevi A, Chesi M, Ferrarese R, Garcia Perez L, Grioni M, et al. Microbiotadriven interleukin-17-producing cells and eosinophils synergize to accelerate multiple
 myeloma progression. Nat Commun. 2018 Dec 3;9(1):4832.
- 89. Chaput N, Lepage P, Coutzac C, Soularue E, Le Roux K, Monot C, et al. Baseline gut
 microbiota predicts clinical response and colitis in metastatic melanoma patients treated
 with ipilimumab. Ann Oncol. 2017 Jun 1;28(6):1368–79.

- 90. Lione L, Salvatori E, Petrazzuolo A, Massacci A, Maggio R, Confroti A, et al. Antitumor
 efficacy of a neoantigen cancer vaccine delivered by electroporation is influenced by
 microbiota composition. Oncoimmunology. 2021 Mar 23;10(1):1898832.
- 91. Hyeyoon K, Rira O, Sangjun P, Geun Eog J, Myeong SP, Sung-Eun K. Bifidobacterium
 longum RAPO enhances efficacy of anti-PD-1 immunotherapy in a mouse model of
 triple-negative breast cancer [Internet]. [cited 2021 Apr 27]. Available from:
 https://www.abstractsonline.com/pp8/#!/9325/presentation/1136
- 92. Pardi N, Hogan MJ, Porter FW, Weissman D. mRNA vaccines a new era in vaccinology.
 Nat Rev Drug Discov. 2018 Apr;17(4):261–79.
- 93. Van Hoecke L, Verbeke R, Dewitte H, Lentacker I, Vermaelen K, Breckpot K, et al. mRNA
 in cancer immunotherapy: beyond a source of antigen. Mol Cancer. 2021 Mar
 3;20(1):48.
- 94. Kauffman KJ, Webber MJ, Anderson DG. Materials for non-viral intracellular delivery of
 messenger RNA therapeutics. J Control Release. 2016 Oct 28;240:227–34.
- 786 95. Riley RS, June CH, Langer R, Mitchell MJ. Delivery technologies for cancer
 787 immunotherapy. Nature Reviews Drug Discovery. 2019 Mar;18(3):175–96.
- 96. Holtkamp S, Kreiter S, Selmi A, Simon P, Koslowski M, Huber C, et al. Modification of
 antigen-encoding RNA increases stability, translational efficacy, and T-cell stimulatory
 capacity of dendritic cells. Blood. 2006 Dec 15;108(13):4009–17.
- 97. Miao L, Zhang Y, Huang L. mRNA vaccine for cancer immunotherapy. Mol Cancer. 2021
 Feb 25;20(1):41.
- 98. Pardi N, Hogan MJ, Weissman D. Recent advances in mRNA vaccine technology. Curr
 Opin Immunol. 2020 Aug;65:14–20.

795	99. Krienke C, Kolb L, Diken E, Streuber M, Kirchhoff S, Bukur T, et al. A noninflammatory
796	mRNA vaccine for treatment of experimental autoimmune encephalomyelitis. Science.
797	2021 Jan 8;371(6525):145–53.

Patel MR, Bauer TM, Jimeno A, Wang D, LoRusso P, Do KT, et al. A phase I study of
mRNA-2752, a lipid nanoparticle encapsulating mRNAs encoding human OX40L, IL23, and IL-36γ, for intratumoral (iTu) injection alone and in combination with
durvalumab. JCO. 2020 May 20;38(15_suppl):3092–3092.

2 101. Zhang C, Zhou D. Adenoviral vector-based strategies against infectious disease and
 cancer. Human Vaccines & Immunotherapeutics. 2016 Aug 2;12(8):2064–74.

Rauch S, Jasny E, Schmidt KE, Petsch B. New Vaccine Technologies to Combat
Outbreak Situations. Front Immunol [Internet]. 2018 [cited 2021 Apr 28];9. Available
from: https://www.frontiersin.org/articles/10.3389/fimmu.2018.01963/full

807 103. Ramezanpour B, Haan I, Osterhaus A, Claassen E. Vector-based genetically modified
808 vaccines: Exploiting Jenner's legacy. Vaccine. 2016 Dec 7;34(50):6436–48.

809 104. Ertl HC. Viral vectors as vaccine carriers. Current Opinion in Virology. 2016 Dec 1;21:1–
810 8.

811 105. Afkhami S, Yao Y, Xing Z. Methods and clinical development of adenovirus-vectored
812 vaccines against mucosal pathogens. Mol Ther Methods Clin Dev. 2016;3:16030.

106. Logunov DY, Dolzhikova IV, Shcheblyakov DV, Tukhvatulin AI, Zubkova OV,
Dzharullaeva AS, et al. Safety and efficacy of an rAd26 and rAd5 vector-based
heterologous prime-boost COVID-19 vaccine: an interim analysis of a randomised
controlled phase 3 trial in Russia. Lancet. 2021 Feb 20;397(10275):671–81.

107. DiPaola RS, Plante M, Kaufman H, Petrylak DP, Israeli R, Lattime E, et al. A phase I
trial of pox PSA vaccines (PROSTVAC-VF) with B7-1, ICAM-1, and LFA-3 co-

stimulatory molecules (TRICOM) in patients with prostate cancer. J Transl Med. 2006Jan 3;4:1.

108. Cappuccini F, Bryant R, Pollock E, Carter L, Verrill C, Hollidge J, et al. Safety and
immunogenicity of novel 5T4 viral vectored vaccination regimens in early stage prostate
cancer: a phase I clinical trial. J Immunother Cancer. 2020 Jun;8(1).

Baden LR, Karita E, Mutua G, Bekker L-G, Gray G, Page-Shipp L, et al. Assessment of
the Safety and Immunogenicity of 2 Novel Vaccine Platforms for HIV-1 Prevention: A
Randomized Trial. Ann Intern Med. 2016 Mar 1;164(5):313–22.

Monath TP, Fast PE, Modjarrad K, Clarke DK, Martin BK, Fusco J, et al. rVSVΔGZEBOV-GP (also designated V920) recombinant vesicular stomatitis virus pseudotyped
with Ebola Zaire Glycoprotein: Standardized template with key considerations for a
risk/benefit assessment. Vaccine X. 2019 Apr 11;1:100009.

111. Andtbacka RHI, Collichio F, Harrington KJ, Middleton MR, Downey G, Öhrling K, et al.
Final analyses of OPTiM: a randomized phase III trial of talimogene laherparepvec
versus granulocyte-macrophage colony-stimulating factor in unresectable stage III–IV
melanoma. J Immunother Cancer [Internet]. 2019 Jun 6 [cited 2021 Apr 29];7. Available
from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6554874/

112. Ylösmäki E, Cerullo V. Design and application of oncolytic viruses for cancer
immunotherapy. Current Opinion in Biotechnology. 2020 Oct 1;65:25–36.

838 113. Friedman GK, Johnston JM, Bag AK, Bernstock JD, Li R, Aban I, et al. Oncolytic HSV839 1 G207 Immunovirotherapy for Pediatric High-Grade Gliomas. New England Journal of
840 Medicine. 2021 Apr 29;384(17):1613–22.

- 841 114. Brun J, McManus D, Lefebvre C, Hu K, Falls T, Atkins H, et al. Identification of
 842 Genetically Modified Maraba Virus as an Oncolytic Rhabdovirus. Mol Ther. 2010
 843 Aug;18(8):1440–9.
- 115. Ylösmäki E, Ylösmäki L, Fusciello M, Martins B, Ahokas P, Cojoc H, et al.
 Characterization of a novel OX40 ligand and CD40 ligand-expressing oncolytic
 adenovirus used in the PeptiCRAd cancer vaccine platform. Molecular Therapy Oncolytics. 2021 Mar 26;20:459–69.
- Tähtinen S, Feola S, Capasso C, Laustio N, Groeneveldt C, Ylösmäki EO, et al.
 Exploiting Preexisting Immunity to Enhance Oncolytic Cancer Immunotherapy. Cancer
 Res. 2020 Jun 15;80(12):2575–85.
- 117. Papachristofilou A, Hipp MM, Klinkhardt U, Früh M, Sebastian M, Weiss C, et al. Phase
 lb evaluation of a self-adjuvanted protamine formulated mRNA-based active cancer
 immunotherapy, BI1361849 (CV9202), combined with local radiation treatment in
 patients with stage IV non-small cell lung cancer. J Immunother Cancer. 2019 Feb
 855 8;7(1):38.
- 856 118. Rosa SS, Prazeres DMF, Azevedo AM, Marques MPC. mRNA vaccines manufacturing:
 857 Challenges and bottlenecks. Vaccine. 2021 Apr 15;39(16):2190–200.
- Anderson EJ, Rouphael NG, Widge AT, Jackson LA, Roberts PC, Makhene M, et al.
 Safety and Immunogenicity of SARS-CoV-2 mRNA-1273 Vaccine in Older Adults. N
 Engl J Med. 2020 Dec 17;383(25):2427–38.
- Monin L, Laing AG, Muñoz-Ruiz M, McKenzie DR, Barrio I del M del, Alaguthurai T, et
 al. Safety and immunogenicity of one versus two doses of the COVID-19 vaccine
 BNT162b2 for patients with cancer: interim analysis of a prospective observational
 study. The Lancet Oncology [Internet]. 2021 Apr 27 [cited 2021 Apr 28];0(0). Available

865 from: https://www.thelancet.com/journals/lanonc/article/PIIS1470-2045(21)00213866 8/abstract

- Yao C, Sun H-W, Lacey NE, Ji Y, Moseman EA, Shih H-Y, et al. Single-cell RNA-seq
 reveals TOX as a key regulator of CD8+ T cell persistence in chronic infection. Nat
 Immunol. 2019;20(7):890–901.
- Liu Y, Liu J, Xia H, Zhang X, Fontes-Garfias CR, Swanson KA, et al. Neutralizing Activity
 of BNT162b2-Elicited Serum. N Engl J Med. 2021 Mar 8;
- 123. Swaminathan G, Thoryk EA, Cox KS, Meschino S, Dubey SA, Vora KA, et al. A novel
 lipid nanoparticle adjuvant significantly enhances B cell and T cell responses to subunit vaccine antigens. Vaccine. 2016 Jan 2;34(1):110–9.
- McGrail DJ, Pilié PG, Rashid NU, Voorwerk L, Slagter M, Kok M, et al. High tumor
 mutation burden fails to predict immune checkpoint blockade response across all
 cancer types. Ann Oncol. 2021 May;32(5):661–72.
- 878 125. Saxena M, van der Burg SH, Melief CJM, Bhardwaj N. Therapeutic cancer vaccines.
 879 Nature Reviews Cancer. 2021 Apr 27;1–19.
- Minati R, Perreault C, Thibault P. A Roadmap Toward the Definition of Actionable
 Tumor-Specific Antigens. Front Immunol. 2020;11:583287.
- 127. De Mattos-Arruda L, Vazquez M, Finotello F, Lepore R, Porta E, Hundal J, et al.
 Neoantigen prediction and computational perspectives towards clinical benefit:
 recommendations from the ESMO Precision Medicine Working Group. Ann Oncol. 2020
 Aug;31(8):978–90.
- 886 128. Fritsch EF, Rajasagi M, Ott PA, Brusic V, Hacohen N, Wu CJ. HLA-binding properties
 887 of tumor neoepitopes in humans. Cancer Immunol Res. 2014 Jun;2(6):522–9.

- Abelin JG, Keskin DB, Sarkizova S, Hartigan CR, Zhang W, Sidney J, et al. Mass
 Spectrometry Profiling of HLA-Associated Peptidomes in Mono-allelic Cells Enables
 More Accurate Epitope Prediction. Immunity. 2017 Feb 21;46(2):315–26.
- 130. Pak H, Michaux J, Huber F, Chong C, Stevenson BJ, Müller M, et al. Sensitive
 immunopeptidomics by leveraging available large-scale multi-HLA spectral libraries,
 data-independent acquisition and MS/MS prediction. Mol Cell Proteomics. 2021 Apr
 9;100080.
- 895 131. Bulik-Sullivan B, Busby J, Palmer CD, Davis MJ, Murphy T, Clark A, et al. Deep learning
 896 using tumor HLA peptide mass spectrometry datasets improves neoantigen
 897 identification. Nat Biotechnol. 2018 Dec 17;
- 132. Gartner JJ, Parkhurst MR, Gros A, Tran E, Jafferji MS, Copeland A, et al. A machine
 learning model for ranking candidate HLA class I neoantigens based on known
 neoepitopes from multiple human tumor types. Nature Cancer. 2021 May 3;1–12.
- 901 133. Ong E, Wong MU, Huffman A, He Y. COVID-19 coronavirus vaccine design using
 902 reverse vaccinology and machine learning. bioRxiv [Internet]. 2020 Mar 21 [cited 2021
 903 Apr 17]; Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7239068/
- 134. Keshavarzi Arshadi A, Webb J, Salem M, Cruz E, Calad-Thomson S, Ghadirian N, et
 al. Artificial Intelligence for COVID-19 Drug Discovery and Vaccine Development. Front
 Artif Intell [Internet]. 2020 Aug 18 [cited 2021 Apr 17];3. Available from:
 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7861281/
- 908 135. Gupta RG, Li F, Roszik J, Lizée G. Exploiting Tumor Neoantigens to Target Cancer
 909 Evolution: Current Challenges and Promising Therapeutic Approaches. Cancer Discov.
 910 2021 Mar 15;

- 911 136. Marcu A, Bichmann L, Kuchenbecker L, Kowalewski DJ, Freudenmann LK, Backert L,
- et al. HLA Ligand Atlas: a benign reference of HLA-presented peptides to improve Tcell-based cancer immunotherapy. J Immunother Cancer. 2021 Apr;9(4).
- 914 137. Yadav M, Jhunjhunwala S, Phung QT, Lupardus P, Tanguay J, Bumbaca S, et al.
 915 Predicting immunogenic tumour mutations by combining mass spectrometry and exome
 916 sequencing. Nature. 2014 Nov 27;515(7528):572–6.
- 138. Laumont CM, Vincent K, Hesnard L, Audemard É, Bonneil É, Laverdure J-P, et al.
 Noncoding regions are the main source of targetable tumor-specific antigens. Sci Transl
 Med. 2018 Dec 5;10(470).
- 920 139. Jeyanathan M, Afkhami S, Smaill F, Miller MS, Lichty BD, Xing Z. Immunological
 921 considerations for COVID-19 vaccine strategies. Nat Rev Immunol. 2020
 922 Oct;20(10):615–32.
- 140. Corti C, Crimini E, Tarantino P, Pravettoni G, Eggermont AMM, Delaloge S, et al.
 Current Perspectives: SARS-CoV-2 vaccines for cancer patients: a call to action.
 European Journal of Cancer [Internet]. 2021 Feb 24 [cited 2021 Mar 15];0(0). Available
 from: https://www.ejcancer.com/article/S0959-8049(21)00082-4/abstract
- 927 141. Sahin U, Oehm P, Derhovanessian E, Jabulowsky RA, Vormehr M, Gold M, et al. An
 928 RNA vaccine drives immunity in checkpoint-inhibitor-treated melanoma. Nature. 2020
 929 Sep;585(7823):107–12.
- 142. Kranz LM, Diken M, Haas H, Kreiter S, Loquai C, Reuter KC, et al. Systemic RNA
 delivery to dendritic cells exploits antiviral defence for cancer immunotherapy. Nature.
 2016 Jun 16;534(7607):396–401.
- 143. Larocca C, Schlom J. Viral vector-based therapeutic cancer vaccines. Cancer J. 2011
 Oct;17(5):359–71.

- 935 144. Capone S, Raggioli A, Gentile M, Battella S, Lahm A, Sommella A, et al.
 936 Immunogenicity of a new gorilla adenovirus vaccine candidate for COVID-19. Molecular
 937 Therapy. 2021 Apr;S1525001621002100.
- 938 145. D'Alise AM, Leoni G, Cotugno G, Troise F, Langone F, Fichera I, et al. Adenoviral
 939 vaccine targeting multiple neoantigens as strategy to eradicate large tumors combined
 940 with checkpoint blockade. Nat Commun. 2019 Jun 19;10(1):2688.
- 941 146. Bilusic M, McMahon S, Madan RA, Karzai F, Tsai Y-T, Donahue RN, et al. Phase I
 942 study of a multitargeted recombinant Ad5 PSA/MUC-1/brachyury-based
 943 immunotherapy vaccine in patients with metastatic castration-resistant prostate cancer
 944 (mCRPC). J Immunother Cancer. 2021 Mar 1;9(3):e002374.
- 945 147. Barrett JA, Cai H, Miao J, Khare PD, Gonzalez P, Dalsing-Hernandez J, et al. Regulated
 946 intratumoral expression of IL-12 using a RheoSwitch Therapeutic System® (RTS®)
 947 gene switch as gene therapy for the treatment of glioma. Cancer Gene Ther. 2018
 948 Jun;25(5–6):106–16.
- 148. Sasso E, D'Alise AM, Zambrano N, Scarselli E, Folgori A, Nicosia A. New viral vectors
 for infectious diseases and cancer. Semin Immunol. 2020 Aug;50:101430.
- 149. Lang FF, Conrad C, Gomez-Manzano C, Yung WKA, Sawaya R, Weinberg JS, et al.
 Phase I Study of DNX-2401 (Delta-24-RGD) Oncolytic Adenovirus: Replication and
 Immunotherapeutic Effects in Recurrent Malignant Glioma. J Clin Oncol. 2018 May
 10;36(14):1419–27.
- 955 150. Calvo Tardón M, Allard M, Dutoit V, Dietrich P-Y, Walker PR. Peptides as cancer
 956 vaccines. Curr Opin Pharmacol. 2019 Aug;47:20–6.

957	151.	Aucouturier J, Dupuis L, Deville S, Ascarateil S, Ganne V. Montanide ISA 720 and 51:
958		a new generation of water in oil emulsions as adjuvants for human vaccines. Expert
959		Rev Vaccines. 2002 Jun;1(1):111–8.

Som GG, Filippov DV, van der Marel GA, Overkleeft HS, Melief CJ, Ossendorp F. Two
in one: improving synthetic long peptide vaccines by combining antigen and adjuvant in
one molecule. Oncoimmunology [Internet]. 2014 Jul 3 [cited 2021 Apr 29];3(7).
Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4292239/

153. Maynard SK, Marshall JD, MacGill RS, Yu L, Cann JA, Cheng LI, et al. Vaccination with 964 965 synthetic long peptide formulated with CpG in an oil-in-water emulsion induces robust 966 E7-specific CD8 T cell responses and TC-1 tumor eradication. BMC Cancer [Internet]. 967 2019 Jun 6 [cited 2021 Apr 29];19. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6555006/ 968

969 154. Melief CJM, van der Burg SH. Immunotherapy of established (pre)malignant disease
970 by synthetic long peptide vaccines. Nature Reviews Cancer. 2008 May;8(5):351–60.

971 155. Tanaka Y, Wada H, Goto R, Osada T, Yamamura K, Fukaya S, et al. TAS0314, a novel
972 multi-epitope long peptide vaccine, showed synergistic antitumor immunity with PD973 1/PD-L1 blockade in HLA-A*2402 mice. Scientific Reports. 2020 Oct 14;10(1):17284.

974 156. Kondo S, Shimizu T, Koyama T, Sato J, Iwasa S, Yonemori K, et al. First-in-human
975 study of the cancer peptide vaccine TAS0313 in patients with advanced solid tumors.
976 Cancer Science. 2021;112(4):1514–23.

977 157. Keskin DB, Anandappa AJ, Sun J, Tirosh I, Mathewson ND, Li S, et al. Neoantigen
978 vaccine generates intratumoral T cell responses in phase Ib glioblastoma trial. Nature.
979 2019 Jan;565(7738):234–9.

- 980 158. Ott PA, Hu-Lieskovan S, Chmielowski B, Govindan R, Naing A, Bhardwaj N, et al. A
 981 Phase Ib Trial of Personalized Neoantigen Therapy Plus Anti-PD-1 in Patients with
 982 Advanced Melanoma, Non-small Cell Lung Cancer, or Bladder Cancer. Cell. 2020 Oct
 983 15;183(2):347-362.e24.
- 984 159. Awad MM, Govindan R, Spigel DR, Garon EB, Kohler V, Vyasamneni R, et al. A
 985 personal neoantigen vaccine NEO-PV-01 in combination with chemotherapy and
 986 pembrolizumab induces broad de novo immune responses in first-line non-squamous
 987 NSCLC: Associations with clinical outcomes [Internet]. [cited 2021 Apr 28]. Available
 988 from: https://www.abstractsonline.com/pp8/#!/9325/presentation/1131
- 989 160. Hu Z, Leet DE, Allesøe RL, Oliveira G, Li S, Luoma AM, et al. Personal neoantigen
 990 vaccines induce persistent memory T cell responses and epitope spreading in patients
 991 with melanoma. Nat Med. 2021 Mar;27(3):515–25.
- 992 161. Jackson NAC, Kester KE, Casimiro D, Gurunathan S, DeRosa F. The promise of mRNA
 993 vaccines: a biotech and industrial perspective. NPJ Vaccines. 2020 Feb 4;5(1):11.
- 162. Tatematsu M, Funami K, Seya T, Matsumoto M. Extracellular RNA Sensing by Pattern
 Recognition Receptors. J Innate Immun. 2018;10(5–6):398–406.
- 996 163. Doria-Rose N, Suthar MS, Makowski M, O'Connell S, McDermott AB, Flach B, et al.
 997 Antibody Persistence through 6 Months after the Second Dose of mRNA-1273 Vaccine
 998 for Covid-19. N Engl J Med. 2021 Apr 6;
- 999 164. Gomez PL, Robinson JM. Vaccine Manufacturing. Plotkin's Vaccines. 2018;51-60.e1.
- 1000 165. Robert-Guroff M. Replicating and non-replicating viral vectors for vaccine development.
 1001 Curr Opin Biotechnol. 2007 Dec;18(6):546–56.
- 1002 166. Diamond MS, Pierson TC. The Challenges of Vaccine Development against a New
 1003 Virus during a Pandemic. Cell Host Microbe. 2020 May 13;27(5):699–703.

- 1004 167. Bolles M, Deming D, Long K, Agnihothram S, Whitmore A, Ferris M, et al. A double inactivated severe acute respiratory syndrome coronavirus vaccine provides incomplete
 protection in mice and induces increased eosinophilic proinflammatory pulmonary
 response upon challenge. J Virol. 2011 Dec;85(23):12201–15.
- 1008 168. Chandler RE. Optimizing safety surveillance for COVID-19 vaccines. Nat Rev Immunol.
 2020 Aug;20(8):451–2.
- 1010 169. Tate JG, Bamford S, Jubb HC, Sondka Z, Beare DM, Bindal N, et al. COSMIC: the
 1011 Catalogue Of Somatic Mutations In Cancer. Nucleic Acids Res. 2019 Jan
 1012 8;47(D1):D941–7.
- 1013 170. Lee EY, Kulkarni RP. Circulating biomarkers predictive of tumor response to cancer
 1014 immunotherapy. Expert Rev Mol Diagn. 2019 Oct;19(10):895–904.
- 1015 171. Litchfield K, Reading JL, Puttick C, Thakkar K, Abbosh C, Bentham R, et al. Meta1016 analysis of tumor- and T cell-intrinsic mechanisms of sensitization to checkpoint
 1017 inhibition. Cell. 2021 Feb 4;184(3):596-614.e14.
- 1018 172. Bedard PL, Hyman DM, Davids MS, Siu LL. Small molecules, big impact: 20 years of
 1019 targeted therapy in oncology. Lancet. 2020 Mar 28;395(10229):1078–88.
- 1020 173. Jeselsohn R, Buchwalter G, De Angelis C, Brown M, Schiff R. ESR1 mutations—a
 1021 mechanism for acquired endocrine resistance in breast cancer. Nat Rev Clin Oncol.
 1022 2015 Oct;12(10):573–83.
- 1023 174. Punta M, Jennings VA, Melcher AA, Lise S. The Immunogenic Potential of Recurrent
 1024 Cancer Drug Resistance Mutations: An In Silico Study. Front Immunol.
 1025 2020;11:524968.

- 1026 175. Sanchez-Vega F, Mina M, Armenia J, Chatila WK, Luna A, La KC, et al. Oncogenic
 1027 Signaling Pathways in The Cancer Genome Atlas. Cell. 2018 Apr 5;173(2):3211028 337.e10.
- 1029 176. Sever R, Brugge JS. Signal Transduction in Cancer. Cold Spring Harb Perspect Med
 1030 [Internet]. 2015 Apr [cited 2021 Apr 27];5(4). Available from:
 1031 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4382731/
- 1032 177. Syeda MM, Wiggins JM, Corless BC, Long GV, Flaherty KT, Schadendorf D, et al.
 1033 Circulating tumour DNA in patients with advanced melanoma treated with dabrafenib or
 1034 dabrafenib plus trametinib: a clinical validation study. The Lancet Oncology. 2021 Mar
 1035 1;22(3):370–80.
- 1036 178. Ococks E, Frankell AM, Soler NM, Grehan N, Northrop A, Coles H, et al. Longitudinal
 1037 tracking of 97 esophageal adenocarcinomas using liquid biopsy sampling. Annals of
 1038 Oncology. 2021 Apr 1;32(4):522–32.
- 1039 179. Melief CJM, Hall T van, Arens R, Ossendorp F, Burg SH van der. Therapeutic cancer
 1040 vaccines. J Clin Invest. 2015 Sep 1;125(9):3401–12.
- 1041 180. Berger MF, Mardis ER. The emerging clinical relevance of genomics in cancer
 1042 medicine. Nat Rev Clin Oncol. 2018 Jun;15(6):353–65.
- 1043 181. Robinson J, Barker DJ, Georgiou X, Cooper MA, Flicek P, Marsh SGE. IPD-IMGT/HLA
 1044 Database. Nucleic Acids Res. 2020 Jan 8;48(D1):D948–55.
- 1045 182. Peters B, Nielsen M, Sette A. T Cell Epitope Predictions. Annu Rev Immunol. 2020 Apr
 1046 26;38:123–45.
- 1047 183. Sarkizova S, Klaeger S, Le PM, Li LW, Oliveira G, Keshishian H, et al. A large
 1048 peptidome dataset improves HLA class I epitope prediction across most of the human
 1049 population. Nat Biotechnol. 2020 Feb;38(2):199–209.

Figure 1. Main vaccines formulations developed for cancer therapy. Four types of vaccine
platforms have been developed for therapeutic purposes: viral/bacterial-based, gene-based,
peptide-based and cell-based vaccines (27). Examples of each different strategies are
depicted. Abbreviations: T-VEC, Talimogene laherparepvec; HSV-1, Herpes simplex virus 1;
DC, dendritic cell; DNA, Deoxyribonucleic acid; mRNA, messenger ribonucleic acid; APC,
antigen-presenting cell; IL-2, Interleukin-2; TNF, Tumor necrosis factor; IFNγ, Interferon
gamma.

1057

1058 Figure 2. Targets for tumor vaccines fall into two general classes: tumor-associated 1059 antigens (TAAs) and tumor-specific antigens (TSAs). TAAs are self-antigens that are 1060 either preferentially or abnormally expressed in tumor cells but may be expressed at some 1061 level in normal cells, as well. T cells that bind with high affinity to TAAs are typically deleted 1062 from the immune repertoire by central and peripheral tolerance mechanisms. TSAs, 1063 comprised of oncoviral antigens and neoantigens, are tumor-specific. Consequently, they are 1064 generally highly immunogenic, due to lack of central tolerance. TSAs associated to oncogenic 1065 viruses have been identified in virus-induced cancers such as human papillomavirus (HPV)-1066 associated cervical cancer, hepatitis B virus-associated hepatocellular carcinoma and human 1067 herpesvirus 8-associated Kaposi sarcoma. Tumor neoantigens are products of somatic mutations acquired during carcinogenesis. NeoAg encoded by oncogenic driver mutations 1068 1069 may be prevalent across patients and tumor types, so they are referred to as shared 1070 neoantigens. However, the majority of NeoAg are unique to individual patients' tumors (private neoantigens). To date, through integration of tumor sequencing with the prediction of MHC-1071 1072 binding epitopes, it is possible to tailor tumor NeoAg selection on the single patient level (27). 1073 Tumor specificity "optimal" (antigen present only in cancer cells) "good" (antigen preferentially 1074 expressed in cancer cells) or "variable" (antigen overexpressed/shared with healthy tissues). 1075 Central tolerance "high" (antigen physiologically expressed in healthy tissues), "low" (central 1076 tolerance present but antigens restricted to immune-privileged sites) or "none" (no evidence 1077 of immunological tolerance) (18) (22).

1078

1079 Figure 3. Potential Strategies to heat up the TME. (A) Targeting cellular metabolism and 1080 certain metabolites within the TME to reduce immunosuppressive regulatory T cells (Tregs), myeloid derived suppressor cells (MDSCs), tumor-associated macrophages (TAMs), or to 1081 1082 generate metabolically fit T cells with better mitochondrial activity to protect against the tumor. Image captions: °, immune cell; ^ stroma cell; * cancer cell. (B) Targeting epigenetic 1083 1084 modulators to either promote immunogenicity of tumor cells or to re-educate TAMs, MDSCs, 1085 or Tregs for the support of T-cell effector functions (71). (C) Induced activation of the innate 1086 immune sensing system with stimulator of STING agonists or boosting cross presenting DCs 1087 to promote tumor Ag-specific T-cell trafficking or function within the TME. (D) Creating an 1088 inflamed TME via OVs or nanoparticle delivery of key immunomodulatory factors (71). In this 1089 regard, OVs deserve a special mention, as they are capable of tumor-specific replication, 1090 which can provide a therapeutic opportunity (72). Briefly, OVs are naturally occurring or 1091 genetically engineered viral species, able to selectively kill cancer cells without damaging 1092 healthy tissue (72). Their mechanism of action is multimodal, as the injection of OVs in 1093 primary/accessible tumors induces immunogenic cell death (ICD) of tumor cells, promoting 1094 the build-up of an inflamed TME (71). In fact, OVs support Natural Killer (NK)-cell and T-cell 1095 immune responses, ultimately improving the lysis of OV-infected cancer cells. Moreover, the 1096 activation of antiviral innate immunity, such as type I IFNs and IFN-stimulated genes (ISGs), 1097 promotes the release of damage- and pathogen-associated molecular patterns (DAMPs and 1098 PAMPs), the exposure of viral/tumor Ag as well as the polarization of TAMs towards anti-tumor 1099 M1 phenotype within the TME (73) (74). The consequent OVs-mediated upregulation of 1100 immune checkpoints (i.e. Programmed death-ligand 1 and Programmed death-ligand 2, PD-1101 L1 and PD-L2, respectively), provides a rationale for combination immunotherapy of OVs plus 1102 ICB. Abbreviations: STING, stimulator of interferon genes; RNA Pol II, RNA polymerase II; 1103 TNF, tumor necrosis factor; TF, transcription factor; IncRNA, long non-coding ribonucleic acid; 1104 miRNA, microRNA; HAT, histone acetyltransferase; KDM, histone lysine demethylase; SAM, 1105 S-adenosyl methionine; DC, dendritic cell; IFN, interferon; MDSC, myeloid-derived suppressor 1106 cell; NK, natural killer; TAM, tumor-associated macrophage; M1, classically activated

- 1107 macrophages; M2, alternatively activated macrophages; ILC1/2, Innate Lymphoid Cells 1/2;
- 1108 IL, interleukin; N1, antitumorigenic neutrophil; N2, pro-tumorigenic neutrophil; Treg, regulatory
- 1109 T cell; Th, T helper cell; TGF- β , Transforming Growth Factor- β ; OV, oncolytic virus; TLR, Toll-
- 1110 like receptor; LPS, Lipopolysaccharides; CCL28, Chemokine (C-C motif) ligand 28.
- 1111
- 1112

Figure 4. Proposed Ag-based cancer vaccination strategies. Different mutations/biological
dependencies may instruct various vaccination strategies and combinatorial agents.
Abbreviations: NeoAg, neoantigens; TAA, tumor-associated antigens; NGS, Next-Generation
sequencing; #, multiple; TME, tumor microenvironment; TMB, tumor mutation burden.

- Figure 5. Key issues to boost applicability and improve clinical efficacy of future cancer vaccines programs. Main areas of development for cancer vaccinology are the rapid introduction of technological advancements, the identification of clear populations that could benefit from CV programs, efforts to allow for comparability of different clinical trials and the establishment of a global workforce able to sustain possible demand and supply-chain. Abbreviations: COSMIC, Catalogue of Somatic Mutations In Cancer; LNP, lipid nanoparticle; LPX, lipoplexes; TME, tumor microenvironment.
- 1125

Journal Prevention

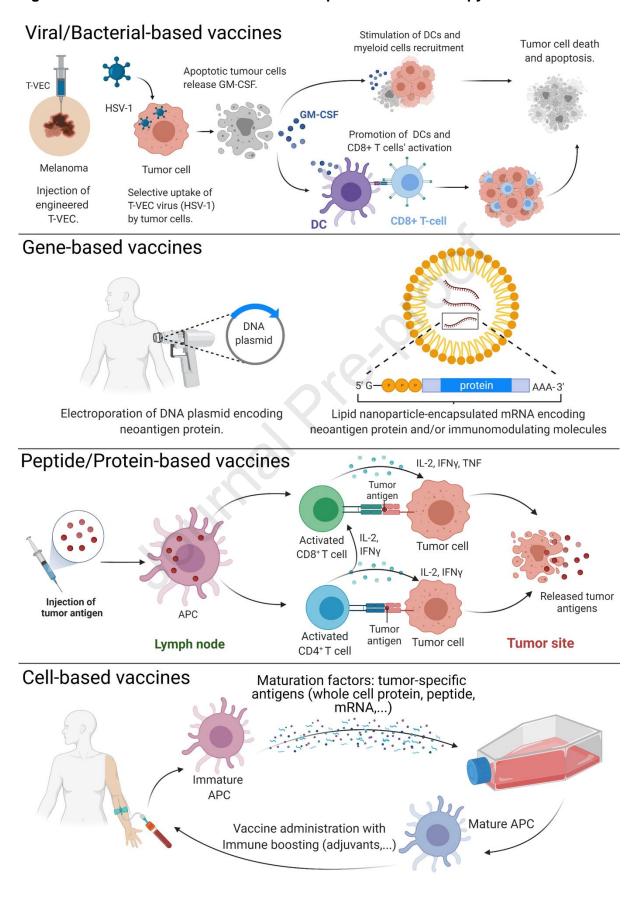
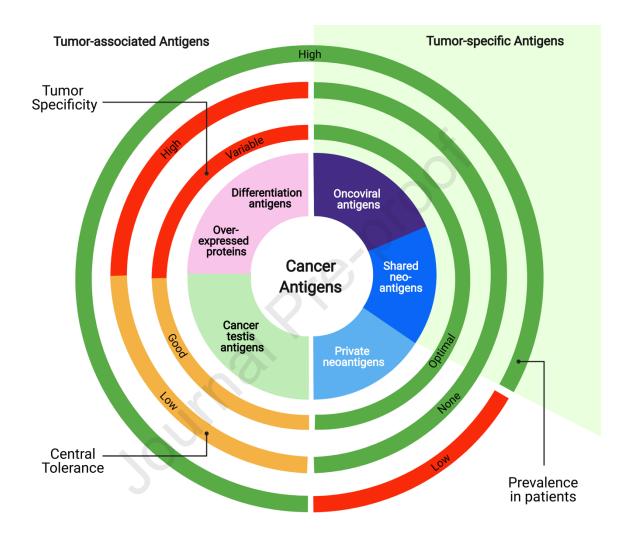




Figure 2. Targets for tumor vaccines fall into two general classes: tumor-associated antigens (TAAs) and tumor-specific antigens (TSAs).





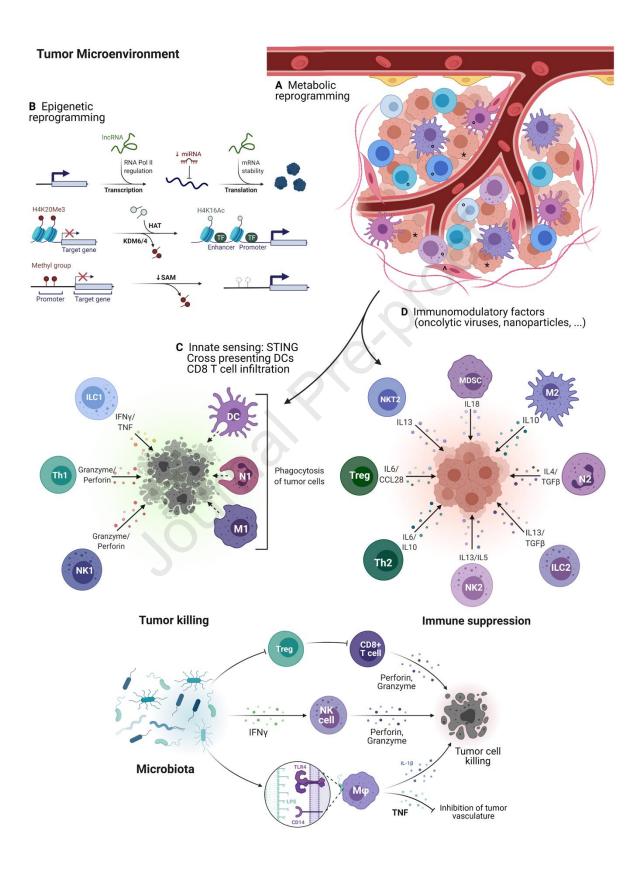


Figure 4. Proposed Ag-based cancer vaccination strategies.

Platform	Personalized (NeoAg)	Shared (NeoAg/TAA)	
Mutation Classification	Nonsynonimous Mut	Driver Mut/TAA	Resistance Mut/Oncoviral
Vaccination Strategy	Therapeutic (established disease)	Therapeutic (precursor stages)	Prophylactic (adjuvant, precursor stages
Target Selection	Patient-specific NGS	Reference Datasets	
Intra-/Inter-Tumor Heterogeneity	High	Low	
# Targets	# mutations	1 or few mutations	# mutations
Combination Therapy	Immunotherapies, TME-altering drugs	Immunotherapies, TME-altering drugs	Small molecules, Endocrine therapy, Others
Best Predicted Application	TMB-high Tumors	TMB-low Tumors	
Scalability / Cost	Low / High	High / Low	High / Medium*-Low *due to concomitant treatme

High / Low

Figure 5. Key issues to boost applicability and improve clinical efficacy of future cancer

vaccines programs.

•

•

•

Research & Technology Clinical Scenario Patient & Disease Selection Open-Source Data (i.e. COSMIC, • HLAseq population studies) **Combination/Concomitant Therapies** • (Neo-)Adjuvant Setting vs Advanced Antigen Discovery Delivery Platforms (i.e. LNP, LPX) TME modifying agents **Preclinical Validation Models Novel Biomarkers Combination/Concomitant Therapies** (predictive & prognostic) **Trial Comparability Global Preparedness**

- Harmonize in silico epitope prediction models
- Homogeneous Data Collection (Efficacy, Immunogenicity, Patient Selection)
- Sufficiently-large Trials
- Different Endpoints

- Scale-up Production and Distribution •
- Shared, Updated Escape Variants • Database
- Streamlined Processes from patient selection to vaccine administration
- Keep in place integrated global workforces and infrastructures