

ORIGINAL ARTICLE

# Mesenchymal stromal cell-derived extracellular vesicles rescue radiation damage to murine marrow hematopoietic cells

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Mesenchymal stromal cells (MSCs) have been shown to reverse radiation damage to marrow stem cells. We have evaluated the capacity of MSC-derived extracellular vesicles (MSC-EVs) to mitigate radiation injury to marrow stem cells at 4 h to 7 days after irradiation. Significant restoration of marrow stem cell engraftment at 4, 24 and 168 h post irradiation by exposure to MSC-EVs was observed at 3 weeks to 9 months after transplant and further confirmed by secondary engraftment. Intravenous injection of MSC-EVs to 500cGy exposed mice led to partial recovery of peripheral blood counts and restoration of the engraftment of marrow. The murine hematopoietic cell line, FDC-P1 exposed to 500cGy, showed reversal of growth inhibition, DNA damage and apoptosis on exposure to murine or human MSC-EVs. Both murine and human MSC-EVs reverse radiation damage to murine marrow cells and stimulate normal murine marrow stem cell/progenitors to proliferate. A preparation with both exosomes and microvesicles was found to be superior to either microvesicles or exosomes alone. Biologic activity was seen in freshly isolated vesicles and in vesicles stored for up to 6 months in 10% dimethyl sulfoxide at  $-80^{\circ}\text{C}$ . These studies indicate that MSC-EVs can reverse radiation damage to bone marrow stem cells.

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## INTRODUCTION

Radiation exposure results in different levels of tissue injury depending on dose, including the immune system, the hematopoietic system, gastrointestinal tract, kidney, skin and lung.<sup>1,2</sup> Hematopoietic stem cells (HSCs) are sensitive to radiation and exposure can result in bone marrow failure. Three months after exposure to 100cGy whole-body irradiation, the engraftment capacity of murine marrow was reduced to 49% of the non-irradiated control marrow.<sup>3</sup> A number of radiation mitigators such as cytokines and growth factors have been described which improve hematopoietic recovery from irradiation damage.<sup>4–6</sup> The transplantation of marrow can restore hematopoiesis in lethally irradiated subjects;<sup>7</sup> however, aside from transplantation, the efficacy of these treatments is relatively limited and temporally constrained.

The mesenchymal stromal cells (MSCs) are multipotent and play a critical role in microenvironmental support of HSC.<sup>8,9</sup> The capacity of MSC for tissue repair has been reported in past decades. The repair mechanisms are believed to be related to either their differentiation capacity or paracrine effects.<sup>10,11</sup> Transplantation of MSC alone or with HSC has also been shown to enhance engraftment and improve bone marrow recovery from radiation injury.<sup>12–18</sup>

Extracellular vesicles (EVs) are the small spherical membrane particles released from cells, which contain mRNA, miRNA, noncoding RNA, protein, lipids and DNA. They have been shown to be involved in cell-to-cell communication and to affect the phenotype of target cells.<sup>19–25</sup> Recent studies have shown that MSC-EVs mediate reversal of different tissue injuries to kidney, brain and myocardium.<sup>26–28</sup> In this study, we evaluated whether marrow MSC-EVs could reverse irradiation damage to marrow stem/progenitor cells.

## MATERIALS AND METHODS

### Cell and culture medium and reagents

FDC-P1 cell line (ATCC, Manassas, VA, USA) was cultured in Dulbecco's modified Eagle's medium with 10% fetal bovine serum (FBS)/5% WEHI cell (ATCC) conditioned media. When preparing culture media for vesicle collection or vesicle-cell co-culture, vesicle-depleted FBS (overnight ultracentrifugation at 100 000 g) was used. Whole bone marrow cells (WBMCs) and lineage-negative cells were cultured in Dulbecco's modified Eagle's medium with 15% FBS/1% penicillin/streptomycin (PS) containing 50 ng/ml stem cell factor. Primary murine marrow-derived MSCs were cultured in minimum essential medium alpha with 10% FBS and 1% PS. All culture medium and related supplements were purchased from Life Technologies (Carlsbad, CA, USA). The antibodies against TER119 (#553669), B220 (#553083), Gr-1 (#553669), CD11b (#553307), CD4 (#553726), CD8 (#553026) and CD45 (#553076) were purchased from BD Bioscience (San Diego, CA, USA); the antibodies against CD73 (#12-0731-81), CD44 (#12-0441-82), CD29 (#12-029-82), CD105 (#12-1051-82), Sca-1 (#11-5981-82), Ia (#12-5321-82), CD3 (#112-0311-82), CD11b (#11-0112-82), CD45 (#11-045-82), CD34 (#11-0341-82), CD86 (#12-0861-82) and CD34 (#14-0341-85) were purchased from eBioscience (San Diego, CA, USA); ExoAb Antibody Kit (# EXOAB Kit-1) including antibodies against CD9, CD63 and CD81 were purchased from System Biosciences (Palo Alto, CA, USA).

### Experimental animals

Six- to eight-week-old male C57BL/6 or B6.SJL mice were purchased from Jackson Laboratory (Bar Harbor, ME, USA). All mouse studies were approved by the Institutional Animal Care and Use Committee at Rhode Island Hospital. The mice were killed by using CO<sub>2</sub> inhalation followed by cervical dislocation.

### Isolation of WBMCs

Cell preparation was performed as previously reported.<sup>29,30</sup> To harvest WBMCs, the marrow was flushed from tibiae, iliac crest and femurs into ice-

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cold phosphate-buffered saline (PBS)/5% heat-inactivated fetal calf serum/1% PS by a syringe with a 22-gauge needle. For isolation of lineage-negative cells, bones were crushed with ice-cold PBS/5% heat-inactivated fetal calf serum/1% PS by a mortar and pestle, followed by filtration through a 40 µm cell strainer (BD Biosciences). Mononuclear cells, were then isolated from WBM by density centrifugation using OptiPrep (Axis-Shield PoC, Oslo, Norway), and then depleted of lineage-positive (Lin+) cells using magnetic Dynabeads (Life Technologies) and anti-TER119, B220, Gr-1, CD11b, CD4 and CD8 antibodies.

### Culture of human/murine MSCs

Human marrow-derived MSCs (Donor #2002 I), purchased from the Texas A&M University System Health Science Center (Temple, TX, USA), were cultured in minimum essential medium alpha with 2–4 mM L-glutamine, 15% FBS and 1% PS according to the manufacturer's instructions.

The murine bone marrow-derived MSCs and bone-derived MSCs were isolated, cultured and characterized as per previous reports.<sup>31,32</sup> The MSCs were magnetically depleted of CD34+, CD45+ and CD11b+ cells. Cells were cultured for 7 days followed by vesicle collection. The 7-day conditioned medium from the murine bone-derived MSCs and murine bone marrow-derived MSCs were harvested and combined for vesicle isolation by differential ultracentrifugation.

The murine MSC phenotypes characterized by flow cytometry expressed CD44, CD29, CD105 and Sca-1 and did not express Ia, CD31, CD11b, CD45, CD34 and CD86 (Supplementary Figure S1A). The MSCs differentiated into osteogenic, adipogenic and chondrogenic cells when cultured in the appropriate differentiative media purchased from Life Technologies (Supplementary Figure S1B).

### EVs isolation and characterization

WBM or human/murine MSCs were cultured in medium with vesicle-depleted FBS for 7 days. Only less than eight passages of MSCs were used to produce MSC-EVs. The vesicles were isolated from culture medium using differential ultracentrifugation as previously described.<sup>20</sup> Unless otherwise noted, all vesicle separations in this study were by differential centrifugation at 300 *g* for 10 min, 2000 *g* for 30 min, 10 000 *g* for 1 h and 100 000 *g* for 1 h with collection of the 100 000 *g* pellet (exosomes). The vesicles were washed two times with PBS and either tested after storage for 1–7 days at 4 °C or resuspended in PBS with 10% dimethyl sulfoxide (DMSO) and stored at –80 °C. EVs were used within 1 week after harvested for the *in vivo* studies. EV functional effects *in vitro* were maintained for up to 6 months when stored in 10% DMSO at –80 °C.

Human and mouse marrow-derived MSC-EVs and WBMC-EVs were analyzed by electron microscopy as previously described.<sup>33</sup> The pictures are shown in Supplementary Figure S2. Surface epitope protein expression (CD9, CD63 and CD81) in human and mouse marrow-derived MSC-EVs and WBMC-EVs were analyzed by western blot (Supplementary Figure S3 and Supplementary Table S1). The number and size distribution of vesicles was determined on a NanoSight NS500 (Malvern Instruments, Malvern, UK) with a Syringe Pump (Supplementary Figure S4).

### Transplantation

C57BL/6J or B6.SJL recipient mice received 200, 500 or 950 cGy based on experimental design 2–4 h prior to cell infusion (Gammacell 40 Exactor, Cesium 137 source irradiator, Best Theratronics Ltd., Ottawa, ON, Canada, 0.94–0.96 Gy/min). A 300–500 µl volume of WBM cells or lineage-negative stem cells were injected by tail vein. Donor chimerism in peripheral blood or marrow was determined by four-color flow cytometry (BD LSR II flow cytometer; BD Biosciences) using a cocktail of fluorescently tagged antibodies against CD45.1, CD45.2, B220, CD3, CD11b and GR-1. The percentage of engraftment was calculated as the ratio of CD45.1 (donor) cells to CD45.1 plus CD45.2. For secondary transplantation, WBM was collected from primary recipient mice, and infused into lethally irradiated mice with subsequent determination of ratios of original donor and recipient cells. For competitive engraftment, WBMCs collected from donor mice were competed with an equal number of host WBMCs into lethally irradiated mice.

Two basic models for engraftment of marrow stem cells were utilized. In one, we competed donor marrow versus residual host marrow in sublethally irradiated (200 cGy) mice. This is a model we have previously validated.<sup>3,34</sup> In the other model, we competed donor cells (B6.SJL CD45.1) versus host cells (C57BL/6J CD45.2) in equal ratios.

### EVs labeling procedure

EVs were directly labeled with 1 µM Vybrant Cell Tracers DiO or DiD (Life Technologies) by incubation for 30 min at 37 °C and then washed twice by ultracentrifugation at 100 000 *g* for 1 h in 1 × PBS.

### Fluorescence molecular tomography

Fluorescence molecular tomography was used to evaluate the EVs biodistribution in tissues of live animals. Mice were injected with 2 × 10<sup>9</sup> of DiD-labeled human MSC-EVs by tail vein injection, after 24 h post 500cGy irradiation. The mice were killed and the organs were dissected for fluorescent signal scanning by the FMT-4000 scanner (PerkinElmer, Waltham, MA, USA) according to the manufacturer's recommended procedure at 6 h post EVs injection. The quantification of the fluorescence data was performed with TrueQuant software (version 3.0; PerkinElmer). The EVs' fluorescence intensity in dissected tissue was determined using region of interest analysis.

### Apoptosis assay

The histone-associated DNA fragmentation was detected in FDC-P1 cells by using Cell Death Detection ELISAPlus kit (Roche Molecular Biochemicals, Indianapolis, IN, USA), according to the manufacturer's instructions.

### Western blot assay

Cells were harvested and lysed in RIPA lysis buffer (Thermo Scientific, Waltham, MA, USA). Twenty micrograms of protein samples were separated on sodium dodecyl sulfate–polyacrylamide gel electrophoresis and transferred onto a polyscreen PVDF membrane. PARP-specific antibody and Phospho-H2A.X (Ser139) Antibody (Cell Signaling Technology, Danvers, MA, USA) were used for immunoblotting. Amersham ECL Advance Western Blotting Detection Kit was used for detection of protein (GE Healthcare, Piscataway, NJ, USA).

### Cell proliferation assay and colony assay

FDC-P1 cells were seeded in a 96-well plate at 1000–1500 cells/200 µl/well and co-cultured with vesicles for 10–14 days. Cell proliferation was based on measurement of cellular DNA content via fluorescent dye binding using a CyQuant proliferation assay kit (Life Technologies) according to the manufacturer's protocol or direct analysis by counting colony formation in each well of a 96-well plate using an inverted microscopy with ×2.5 objectives. Methylcellulose-based reagents were used for murine bone marrow colony-forming cell assay according to the manufacturer's protocol (Stemcell Technologies Inc., Vancouver, BC, Canada).

### miRNA reverse transcriptase-PCR analysis

RNA was isolated from WBMCs with miRNeasy Micro kit (Qiagen, Valencia, CA, USA) according to the manufacturer's protocol. RNA was measured by Nanodrop ND-1000 spectrophotometry (Fisher Scientific Inc., Chelmsford, MA, USA). Fifty nanograms of RNA was converted into cDNA using RT Primers and TaqMan microRNA RT Kit (Applied Biosystems, Foster City, CA, USA), followed by cDNA preamplified using RT PreAmp Primers and Taqman PreAmp Master Mix (Applied Biosystems) according to the manufacturer's protocol. About 0.25 µl diluted PreAmp product was mixed with TaqMan Universal PCR Master Mix and miRNA primer and run using an Applied Biosystems 7900HT real-time PCR instrument (Applied Biosystems). The primers of miR125a-5p, miR210 and *RNU6B* were purchased from Applied Biosystems. *RNU6B* gene was used as the endogenous control. Results were analyzed using the  $\delta\delta C_t$  method, Vancouver, BC, Canada.

### miRNA transfection

We purchased miRNA Mimics from Qiagen, including miR221, miR451a, miR654-3p, miR486-5p, miR142-5p, miR-466i-5p, miR106b, miR125a-5p, miR106b, miR210-5p, miR199, miR21-5p and miR29a-3a. For miRNA overexpression experiments, 100 nM mimic RNA or control miRNA (control) in 10 µl 'R buffer' was transfected into FDC-P1 cells using the Neon electroporation transfection system (Life Technologies) with an optimal program at 1400 V with two 20 ms pulses.

Statistics

Non-parametric Mann–Whitney *U*-tests and analysis of variance with multi-comparison tests were used to determine statistical significance among the groups shown in each experiment (GraphPad Prism; Graphpad Software Inc., La Jolla, CA, USA) and the level of statistical significance was set at 0.05. All *P*-values are two-sided.

RESULTS

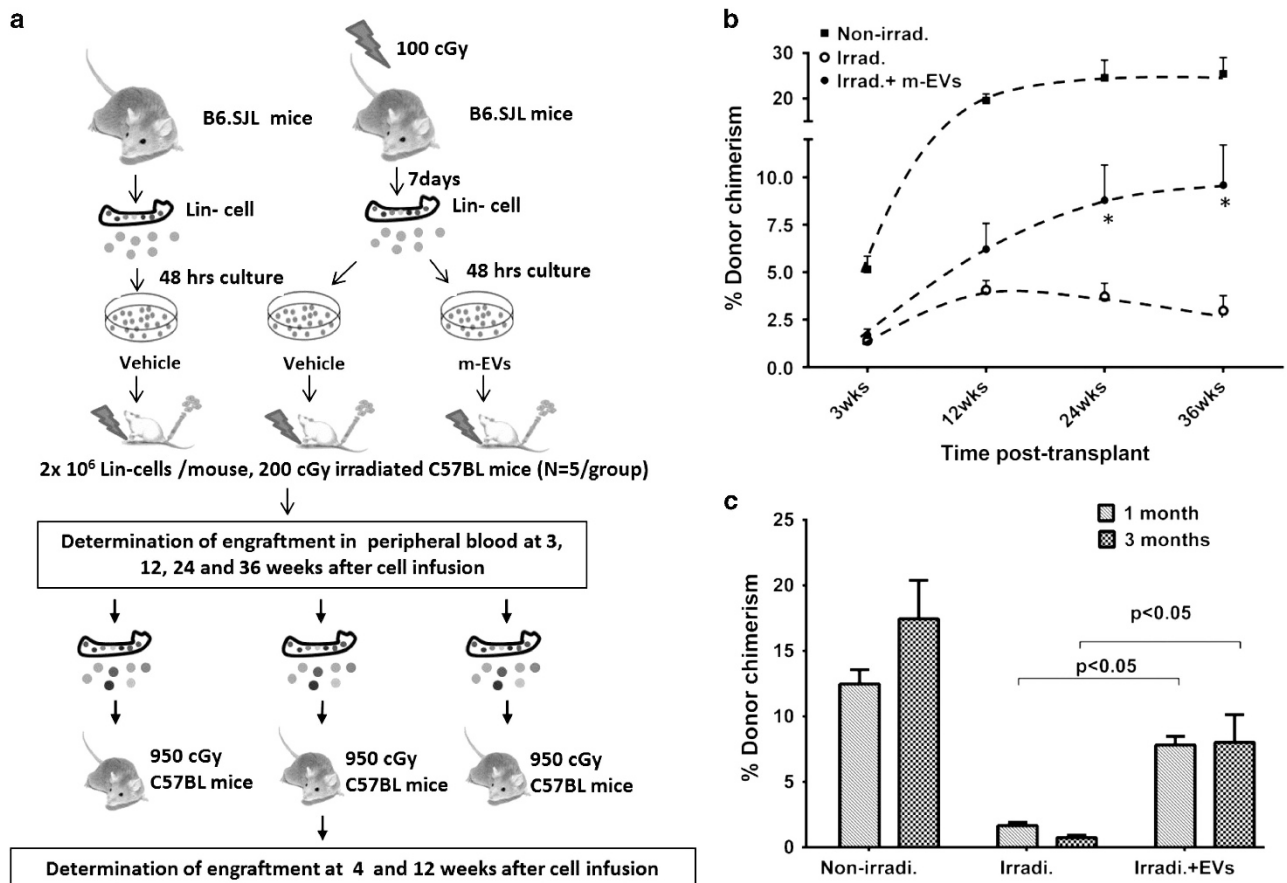
Murine and human marrow MSC-EVs reduce radiation damage to marrow stem cells at 4 h to 7 days after irradiation

To determine whether murine marrow MSC-EVs could reverse hematopoietic radiation damage by *in vitro* exposure of hematopoietic cells to EVs, we investigated the reversal of radiation injury to marrow stem cells 7 days after irradiation (Figure 1a). Murine lineage-negative cells, harvested from B6.SJL mice 7 days post 100cGy whole-body irradiation, were cultured with  $2 \times 10^9$ /ml murine MSC-EVs or vehicle for 48 h (unless otherwise noted, PBS served as vehicle controls in the all experiments). The cells were then tail vein injected into 200cGy exposed C57BL/6J mice and engraftment was analyzed at 3, 12, 24 and 36 weeks post-transplant. This is an engraftment model in which infused cells compete with residual host marrow cells.<sup>3,34</sup> Vesicle exposure led to a statistically significant ( $P < 0.05$ ) increase in engraftment by the irradiated cells at 24 and 36 weeks, with the average percent donor engraftment equal to  $8.8 \pm 1.9$  and  $9.6 \pm 2.1\%$  for the vesicle-treated groups compared with  $3.7 \pm 0.7$  and  $3.0 \pm 0.8\%$  for the non-vesicle-treated groups at 24 and 36 weeks, respectively (Figure 1b).

We evaluated the capacity of marrow cells from the primary transplants to give rise to marrow repopulation in secondary transplants: WBMCs were harvested 36 weeks post-transplant and these cells were transplanted into lethally irradiated B6.SJL mice. There was a significant increase ( $P < 0.05\%$ ) in the engraftment rate by the irradiated cells treated with vesicles at 1 and 3 months after a secondary transplantation with  $7.8 \pm 0.7$  and  $8.0 \pm 2.1\%$  of donor chimerism compared with  $1.7 \pm 0.2$  and  $0.7 \pm 0.2\%$  for non-vesicle-treated groups, respectively (Figure 1c).

In a similar experiment (Supplementary Figure S5A), murine lineage-negative cells were harvested 24 h after 100cGy whole-body irradiation, and cultured with  $2 \times 10^9$ /ml of vesicles or vehicle for 24 h. There was a statistically significant increase in donor engraftment by the lineage-negative cells incubated with murine MSC-EVs at 6 and 8 months with engraftment rates at  $3.0 \pm 0.8$  and  $3.2 \pm 0.9\%$  for the vesicle-treated groups compared with  $0.7 \pm 0.4$  and  $0.7 \pm 0.4\%$  for the non-vesicle-treated groups, respectively (Supplementary Figure S5B). Again, there was persistent enhanced engraftment with vesicle exposure in secondary transplantation with an approximately fivefold higher engraftment rate in vesicle-treated cells compared with non-vesicle-treated cells (Supplementary Figure S5C).

We further investigated whether human marrow MSC-EVs could rescue murine hematopoietic radiation damage *in vitro* (Supplementary Figure S6A). Murine WBMCs, harvested 4 h after 100cGy whole-body irradiation, were cultured with human MSC-EVs at  $2 \times 10^8$ ,  $1 \times 10^9$  and  $2 \times 10^9$ /ml, respectively. Following 24 h of co-culture, the cells were then competitively engrafted into 950cGy-exposed C57BL/6J mice and chimerism was measured up



**Figure 1.** Mesenchymal stem cell-derived EVs reverse radiation damage to marrow stem cells (a) Schematic diagram of the reversal of radiation injury to marrow stem cells 7 days after irradiation. (b) Effect of murine MSC-EVs on engraftment capacity of irradiated mouse marrow cells over time post cell infusion. The percentage of engraftment (donor chimerism) was calculated as the ratio of CD45.1 (donor) cells to CD45.1 plus CD45.2. Data are expressed as mean  $\pm$  s.e.m.,  $N = 5$ /group. (c) Vesicle effects on secondary engraftment. Data are expressed as mean  $\pm$  s.e.m.,  $N = 6$ /group.

to 5 months post-transplant. There was a trend of increased engraftment by cells treated with all three doses of vesicles compared with non-vesicle-treated cells (Supplementary Figure S6B). There was a statistically significant increase in engraftment when the three vesicle-treated groups were pooled together as the vesicle-treated group compared with the non-vesicle-treated group (Supplementary Figure S6C).

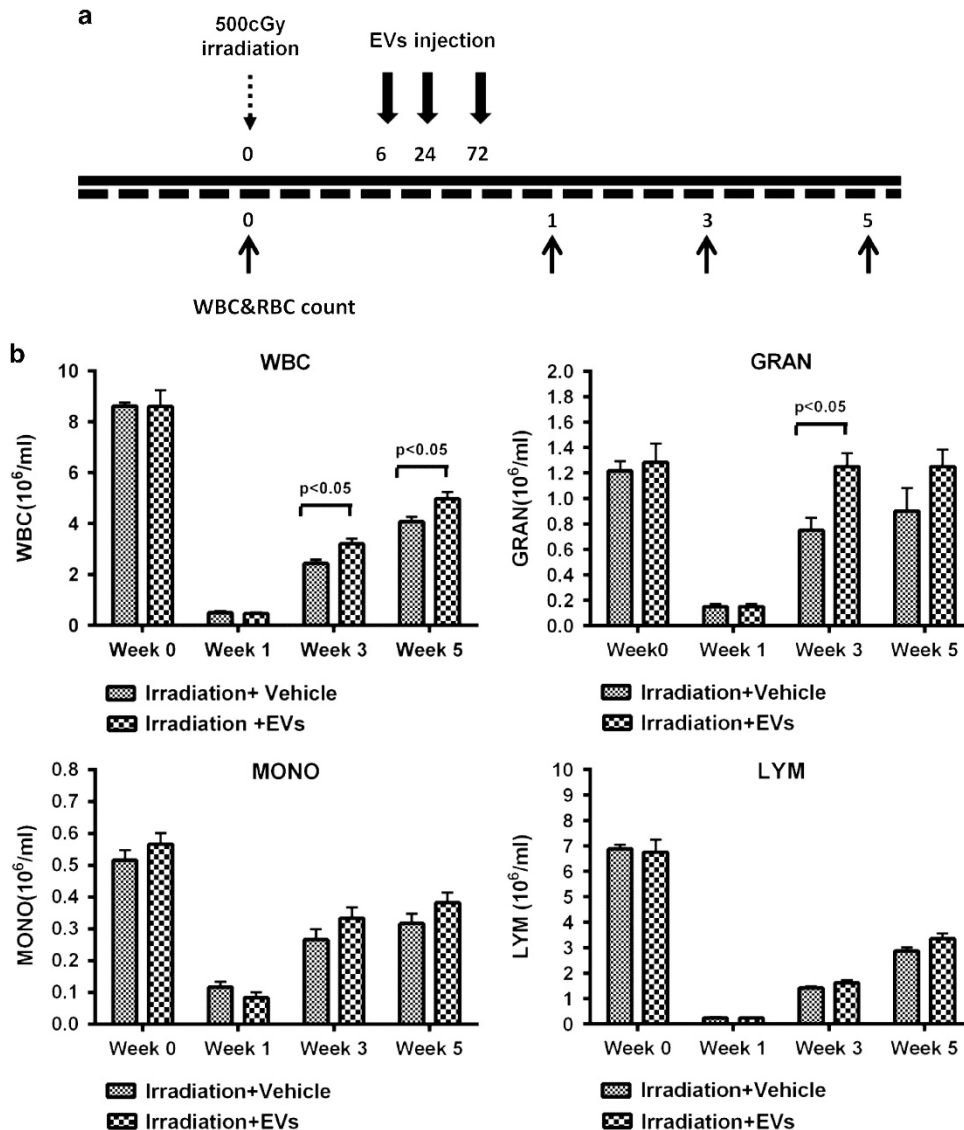
Thus, our data indicate that MSC-EVs can reverse radiation damage to marrow when administered at 4 h to 7 days after irradiation.

*In vivo* reversal of irradiation damage to marrow stem cells by human marrow MSC-EVs

We next evaluated the capacity of human MSC-EVs to reverse marrow radiation damage by *in vivo* administration. C57BL/6 mice were injected intravenously with  $4 \times 10^9$  human MSC-EVs at 6, 24 and 72 h after exposure to 500cGy whole-body irradiation (Figure 2a). We determined WBC, granulocyte, lymphocyte and monocyte counts by HemaTure Analyze (Heska Corporation,

Loveland, CO, USA) at 1 day before irradiation, and 1, 3 and 5 weeks after irradiation (Figure 2b). There was a significant WBC restoration with total WBC counts of  $3.2 \pm 0.2 \times 10^6/\text{ml}$  in the vesicle-treated group and  $2.4 \pm 0.2 \times 10^6/\text{ml}$  in the non-vesicle-treated group at 3 weeks after irradiation ( $P < 0.05$ ). The significant restoration was also seen at 5 weeks after irradiation with WBC counts of  $5.0 \pm 0.3 \times 10^6$  vs  $4.1 \pm 0.2 \times 10^6/\text{ml}$  ( $P < 0.05$ ). Granulocyte level was restored to  $1.3 \pm 0.1 \times 10^6/\text{ml}$  at 3 weeks post irradiation in the vesicle-treated group while in the non-vesicle-treated group was  $0.8 \pm 0.1 \times 10^6/\text{ml}$  ( $P < 0.05$ ). There were no other significant changes in peripheral blood cell types. Our data suggest that MSC-EVs can rescue radiation damage in bone marrow cells *in vivo*.

The impact of murine MSC-EVs or WBMC-EVs on the *in vitro* growth of murine bone marrow or FDC-P1 hematopoietic cells To determine if exposure of irradiated hematopoietic cells to vesicles could promote bone marrow stem cell proliferation, we next investigated the effect of murine MSC-EVs on murine WBM cell recovery from radiation damage. WBM cells, harvested from 0 or



**Figure 2.** Mesenchymal stem cell-derived EVs promote WBC and granulocyte recovery after 500cGy irradiation. (a) Schematic diagram of experimental design. (b) The effect of human MSC-EVs on peripheral leukocyte counts after radiation injury. The WBC, granulocyte, lymphocyte and monocyte counts were used as indices of recovery (mean  $\pm$  s.e.m.,  $N = 6/\text{group}$ ).

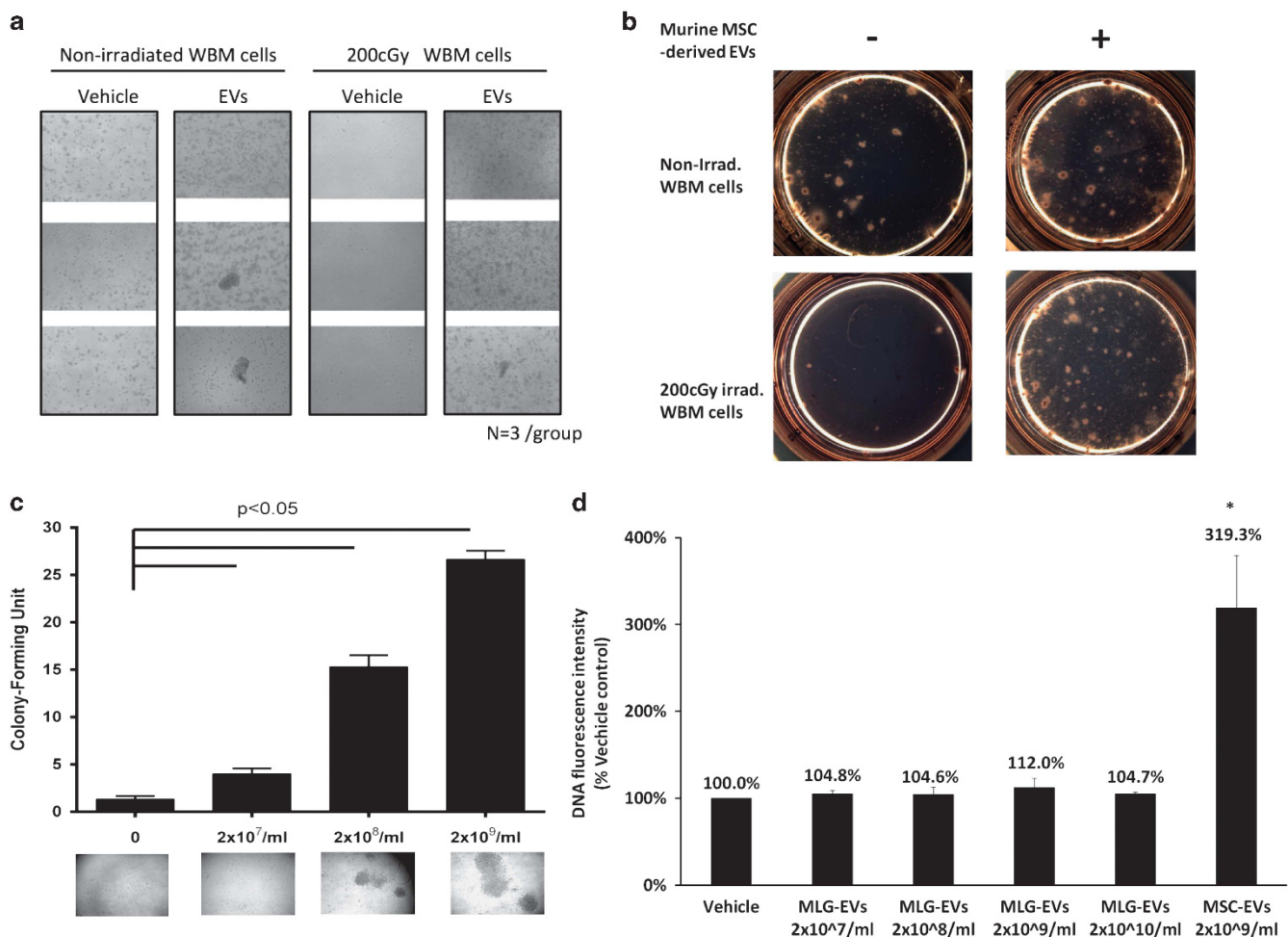
200cGy whole-body irradiated mice, were cultured in Dulbecco's modified Eagle's medium with 50 ng/ml stem cell factor and 15% FBS with the presence or absence of  $2 \times 10^9$  vesicles/ml murine MSC-EVs for 10 days and then cultured in methylcellulose medium as per the manufacturer's instructions for another 10 days. Vesicle exposure induced a significant increase proliferation (Figure 3a) and colony formation (Figure 3b) in irradiated WBM cells.

We also evaluated the effect of adding MSC-EVs and vesicles isolated directly from murine marrow to FDC-P1 murine hematopoietic cells *in vitro*. FDC-P1 cells were exposed to 500cGy irradiation then cultured with murine MSC-EVs or WBM-EVs for 10 days. As expected, MSC-EVs exposure induced a significant increase proliferation in irradiated FDC-P1 cells (data not shown). Interestingly, WBM-EVs exposure led to a dose-dependent restoration of cell growth (Figure 3c). To evaluate the specific capacity of MSC-EVs on recovery from radiation damage, four doses of EVs isolated from mouse lung fibroblast cells were added to 500cGy irradiated FDC-P1 cells for 10 days and there were not significant improvement on cell proliferation (Figure 3d). This indicates that the restoration of proliferation after irradiation is specific for MSC-EVs.

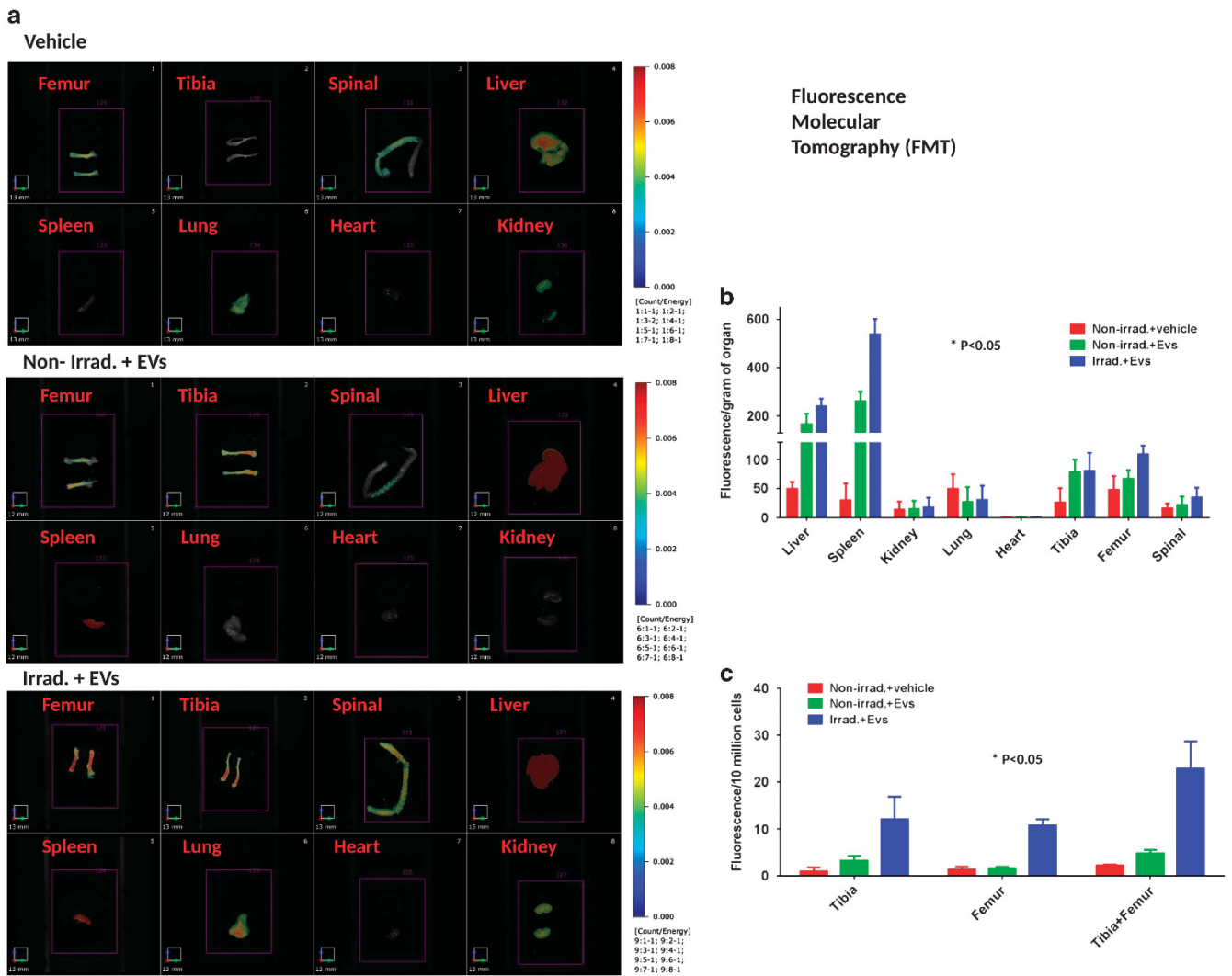
EVs biodistribution in dissected organs and EVs intracellular uptake

To evaluate the biodistribution of EVs after injection, mice were injected with  $2 \times 10^9$  of DiD-labeled human MSC-EVs by tail vein, after 24 h post 500cGy irradiation. At 6 h post EVs injection, the mice were killed and the organs, including heart, lung, spleen, kidney, liver, tibia, femur and spinal were dissected for fluorescent signal scanning by an FMT-4000 scanner (Figure 4a). There was a significantly strong fluorescent signal in the liver and spleen, followed by a moderate signal in the bone marrow from legs and a mild signal in spinal, but no signal in heart, kidney and lung was detected, compared with PBS-treated group (Figure 4b). The fluorescence intensity in the spleen and bone marrow from legs with radiation exposure was significantly higher when compared with those organs from non-irradiation exposed mice after DiD-labeled EVs injection ( $P < 0.05$ ; Figures 4b and c). This result suggests a specific accumulation of EVs at the site of injury.

We next examined the uptake rate of EVs by murine WBMCs by flow cytometry. The fresh WBMCs harvested from the tibia and femur were cultured with DiD-labeled human MSC-EVs for



**Figure 3.** Restoration of growth on hematopoietic cells by *in vitro* exposure to MSC/WBM-EVs. **(a)** Exposure to murine MSC-EVs led to a significant expansion of bone marrow cell ( $n = 3$ /group). Cell growth images were taken under a Zeiss Observer Z1 microscope (Carl Zeiss AS, Oslo, Norway) with  $\times 2.5$  objectives. **(b)** Murine MSC-EVs promoted colony formation on irradiated bone marrow cells. **(c)** Reversal of radiation toxicity to FDC-P1 cells by murine WBM-EVs. 500cGy irradiated FDC-P1 cells were treated with  $2 \times 10^7$ ,  $2 \times 10^8$  and  $2 \times 10^9$ /ml of murine MSC-EVs for 10 days. Colony formation in the well of 96-well plate was determined by counting using a Zeiss Observer Z1 microscope (Carl Zeiss AS) and  $\times 2.5$  objectives.  $N = 3$ / group; colony:  $> 50$  cells per cluster. **(d)** Effect of MLG-EVs on recovery of growth on radiation damage FDC-P1 cells. 500cGy irradiated FDC-P1 cells were treated with  $2 \times 10^7$ ,  $2 \times 10^8$ ,  $2 \times 10^9$  and  $2 \times 10^{10}$ /ml of MLG-EVs or  $2 \times 10^9$ /ml murine MSC-EVs for 10 days. The proliferation of radiation-damaged FDC-P1 cells was determined by using CyQUANT NF Cell Proliferation Assay, with values normalized to the levels of vehicle control (mean  $\pm$  s.d.,  $n = 3$ /group). \* $P < 0.05$  compared with vehicle control. MLG, mouse lung fibroblast.



**Figure 4.** EVs biodistribution in dissected organs from mice. **(a)** Typical images of dissected organs from mice with 0 or 500cGy radiation exposure at 6 h after DiD-EVs injection.  $N=3$  mice/group. **(b)** EVs distribution quantitated with fluorescence molecular tomography. Fluorescence intensity of organs was normalized by organ weight. Fluorescence intensity is expressed as the average of fluorescence intensity (count/energy)  $\pm$  the standard error of the mean (s.e.m.). ANOVA with multi-comparison test was performed.  $*P < 0.05$ , Irrad.+EVs vs Non-irrad.+EVs. **(c)** Bone marrow fluorescence quantification. The bone marrow cells were flushed out from two of tibia and two of femur and counted by a hemocytometer after detection of fluorescence by fluorescence molecular tomography. Then the intensity of fluorescence was normalized by 10 million bone marrow cells, expressed as the average of fluorescence intensity (count/energy)  $\pm$  s.e.m., The  $t$ -test was performed.  $*P < 0.05$ , fluorescence intensity of femur, Irrad.+EVs vs Non-irrad.+EVs.

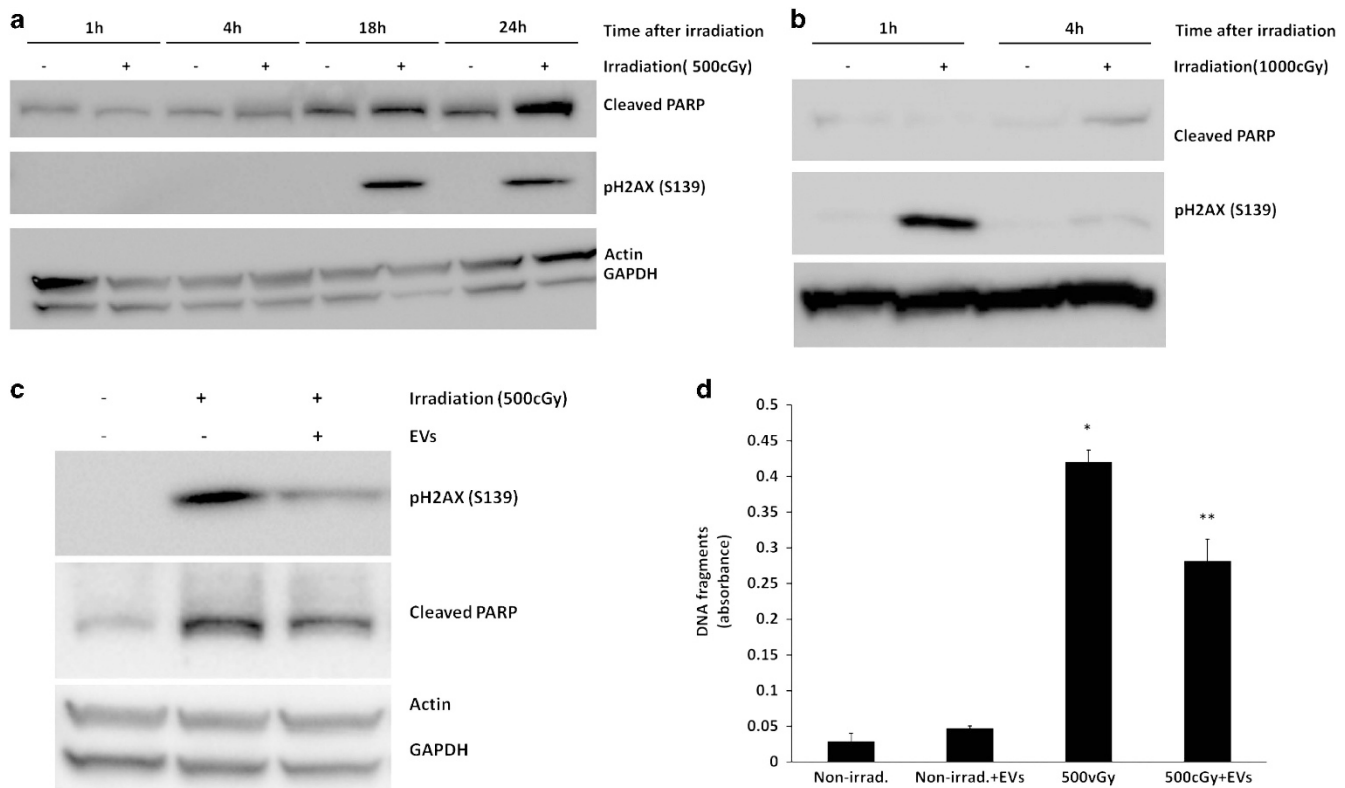
24 h. There was a  $31.73 \pm 1.55\%$  uptake rate of WBMCs and this was further confirmed by confocal microscopy (Supplementary Figure S7A). A similar study also showed MSC-EVs intracellularization in FDC-P1 cells (Supplementary Figure S7B).

We further evaluated if the miRNAs enriched in human MSC-EVs could be transferred or unregulated in murine bone marrow cells. Several miRNA candidates that were shown to be enriched in human MSC-EV miRNA profile through deep sequencing analysis (data not shown) were examined in murine WBMCs after treatment with human MSC-EVs. We found that EVs treatment caused a  $3.7 \pm 1.3$  and  $3.3 \pm 1.6$ -fold increase in miR210 and miR125a-5p expression (Supplementary Figure S7C), suggesting that an upregulation of miRNA expression in target cells after EVs treatment might be due to horizontal transfer of the miRNA from EVs to target cells.

Our data indicate that EVs can accumulate in the injured bone marrow and alter the miRNA expression in bone marrow after EVs injection to mice.

MSC-EVs reverse radiation-induced DNA damage and apoptosis in FDC-P1 cells

We further investigated the expression of apoptosis, cleaved PARP and DNA damage, phosphorylated H2AX<sup>35</sup> by western blot analysis in FDC-P1 cells from 1 to 24 h after 500cGy irradiation. There was a significant increase of phosphorylated H2AX and cleaved PARP 18 h after radiation. However, a significant increase of phosphorylated H2AX was detected at 1 h after 1000cGy irradiation and then phosphorylated H2AX rapidly decreased in 4 h post irradiation (Figures 5a and b). Our data indicated a time- and dose-dependent radiation damage in FDC-P1 cells. We next investigated if vesicle exposure could reverse radiation-induced DNA damage and apoptosis. FDC-P1 cells were exposed to 500cGy irradiation then cultured with or without murine MSC-EVs ( $2 \times 10^9$ /ml) for 18 h. There was a significant increase in cleaved PARP and phosphorylated H2AX after radiation exposure; however, vesicle treatment significantly reduced the level of cleaved PARP and H2AX phosphorylation (Figure 5c),



**Figure 5.** Reversal of DNA damage and apoptosis of irradiated FDC-P1 cells *in vitro* by exposure to murine MSC-EVs. **(a, b)** Phosphorylated H2AX and cleaved PARP levels in FDC-P1 cell. **(a)** Cells were harvested at 1, 4, 18 and 24 h after 500cGy irradiation. **(b)** Cells were harvested at 1 and 4 after 1000cGy irradiation. Protein expression was assayed by western blot analysis. **(c)** 500cGy irradiated FDC-P1 cells were treated with  $2 \times 10^9$ /ml of murine MSC-EVs for 18 h. Phosphorylated H2AX and cleaved PARP were measured by western blot analysis. Actin or GAPDH was used as an internal control. The experiments were repeated three times. **(d)** DNA fragmentation in FDC-P1 cells 18 h post 500cGy irradiation with murine MSC-EVs treatment was evaluated using a Cell Death Detection ELISA kit. The data are expressed as mean  $\pm$  s.e.m. of three separate experiments. \* $P < 0.05$  compared with control cells, \*\* $P < 0.05$  compared with the 500cGy irradiated group.

indicating that apoptosis in irradiated cells was reversed by exposure to vesicles. Similar results were also seen with human MSC-EVs or murine WBM-EVs. In addition, DNA fragmentation, an indicator of apoptosis, was determined by the Cell Death Detection ELISA and further confirmed that murine MSC-EVs reverse radiation-induced apoptosis in FDC-P1 cells (Figure 5d).

The effects of different EV populations derived from murine or human MSC on proliferation normal or irradiated FDC-P1 hematopoietic cells

Vesicles were isolated using differential ultracentrifugation steps (300, 10 000 and then 100 000 *g*). This is a classic method for preparation of exosomes. However, this separation isolates the smaller vesicles, discarding larger vesicles (microvesicles). We, therefore, investigated the effects of three different preparations of vesicles on cell proliferation among the 10k pellet (large vesicles, microvesicles), the 100-10k pellet (small vesicles, exosomes) and the 100k pellet (no 10k spin, both small and large vesicles, exosomes and microvesicles).

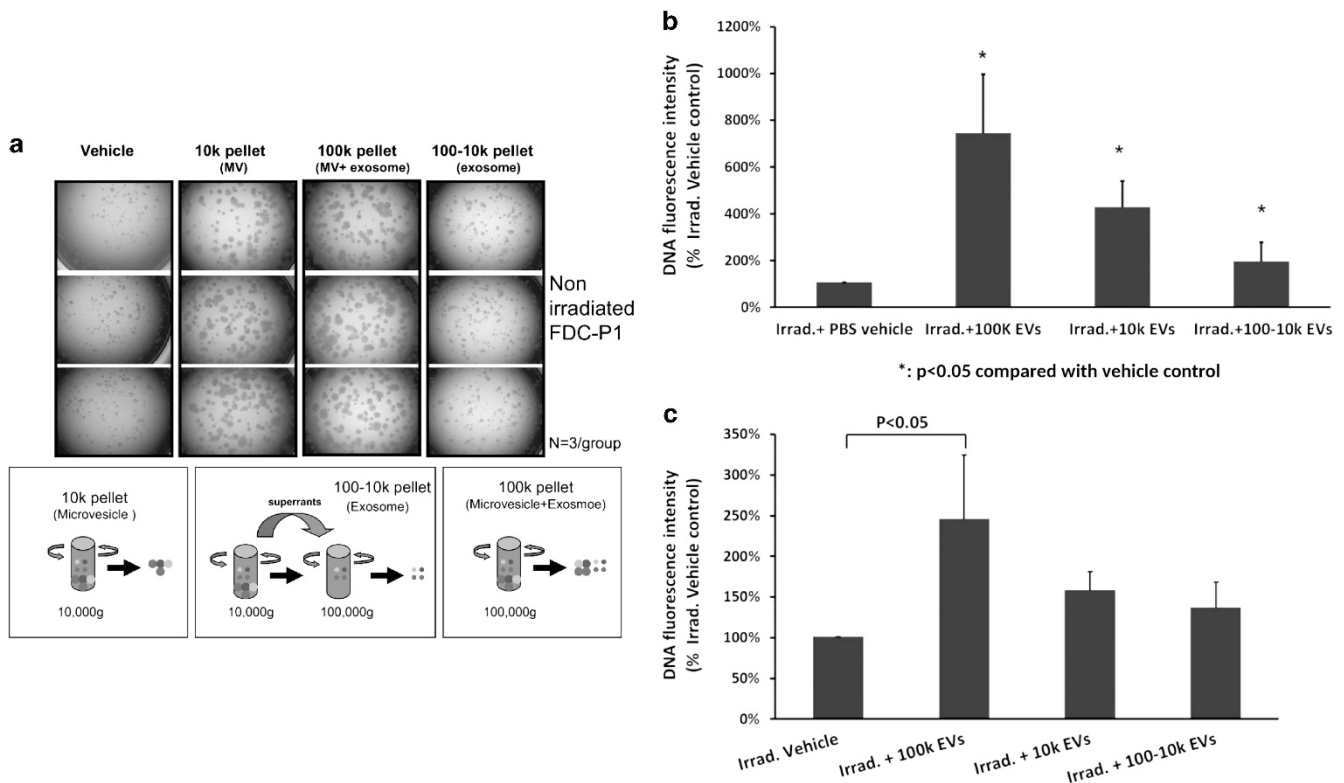
Normal FDC-P1 cells were seeded in a 96-well plate at 1500 cells/well and cultured with three fractions of murine WBM-EVs ( $2 \times 10^9$ /ml) for 7 days, followed by microscopic analysis of cell proliferation. With respect to inducing cellular proliferation, exosomes were clearly inferior compared with microvesicles and the combined exosome and microvesicle population (100k pellet) was superior (Figure 6a).

To evaluate the capacity of murine marrow-EVs to reverse radiation damage, FDC-P1 cells were co-cultured with  $2 \times 10^9$ /ml of WBM-EVs after 500cGy exposure. The 100k pellet showed the largest effect on the reversal of radiation damage at  $745 \pm 252\%$  of

the non-vesicle-treated control group by using the CyQUANT proliferation assay, while, the 10k pellet treated group was determined to be  $429 \pm 111\%$  and the 100-10k pellet was at  $198 \pm 80\%$  of non-vesicle-treated group ( $P < 0.05$ ) (Figure 6b). Again in the same experimental design, the combined fractions of human MSC-EVs were superior to the 10k and 100-10k fractions in the recovery of radiation damage in FDC-P1 cells (Figure 6c).

The effect of *in vivo* injection of human MSC-EVs on murine engraftable stem cells

We next tested the capacity of human MSC-EVs isolated from different fractions on reversal of murine bone marrow damage *in vivo* (Figure 7a).  $4 \times 10^9$  vesicles of the three different fractions (10k, 100k-10k and 100k combined fractions) of human MSC-EVs were injected intravenously into B6.SJL mice 6, 24 and 72 h after 500cGy whole-body irradiation. The mice were killed to harvest WBM cells at week 5 post irradiation. Two million of the WBM cells were competitively engrafted with the same amount of C57BL/6J WBM cells into lethally irradiated C57BL/6J mice and engraftment was analyzed at 1, 3 and 6 months post-transplant. The trend of increased engraftment in mice treated with three different fractions of vesicles compared with non-vesicle-treated mice was shown at 1, 3 and 6 months post-transplant time points (data not shown). The 100k (combined fractions) fractions showed a significant increase in donor chimerism in bone marrow at 6 months post-transplant with five times the level of engraftment compared with the irradiation vehicle control. The 10k and 100-10k fractions showed intermediate recoveries (Figure 7b).



**Figure 6.** *In vitro* stimulation of the growth of FDC-P1 cells by different fractions of vesicles. **(a)** The effects of different separate approaches on proliferation of non-irradiated FDC-P1 cells. Cell proliferation images were taken under microscopy with x2.5 objective ( $N = 3/\text{group}$ ). **(b)** Different fractions of WBM-EVs promoted radiated FDC-P1 cell proliferation. The proliferation of radiation-damaged FDC-P1 cells was determined by using CyQUANT NF Cell Proliferation Assay, with values normalized to the levels of untreated cells. The cells were treated with three fractions of vesicles for 10 days (mean  $\pm$  s.d.,  $n = 3/\text{group}$ ). \* $P < 0.05$ , compared with Irrad.+PBS vehicle control. **(c)** Effect of different human MSC-EVs fractions on irradiated FDC-P1 growth *in vitro*. FDC-P1 cells were co-cultured with  $2 \times 10^9/\text{ml}$  of EVs after 500cGy exposure. The proliferation of cells was determined by using CyQUANT NF Cell Proliferation Assay after 10 days of MSC-EVs treatment (mean  $\pm$  s.d.,  $n = 3/\text{group}$ ).

Human MSC-EV associated miRNAs enhance reversal of radiation damage in FDC-P1 cells

A differential miRNA expression profile among 10k pellet (microvesicles), 100-10k pellet (exosomes) and 100k pellet (both exosomes and microvesicles) was determined using deep sequencing (data not shown). We focused on several of the most enriched miRNAs in 100k EVs, including miR221, miR451a, miR654-3p, miR486-5p, miR142-5p, miR106b-3p, miR125a-5p, miR155-5p, miR210-5p, miR199a-3p, miR21-5p and miR29a-3p to further investigate the reversal of radiation damage in FDC-P1 cells. After exposure to 500cGy, miRNAs were transfected into FDC-P1 cells using Neon electroporation transfection system. Cell proliferation assay by CyQuant proliferation assay and DNA fragmentation assay by the Cell Death Detection ELISA were performed 48 h post-miRNA transfection. We found that overexpression of miR221, miR451a and miR654-3p in 500cGy exposure FDC-P1 cells showed an increase growth effect of radiated cells at  $124.8 \pm 2.8$ ,  $134.2 \pm 6.4$  and  $139.7 \pm 2.5\%$  of the scramble miRNA-treated control group, respectively ( $P < 0.05$ ; Figure 8a). Overexpression of miR106b-3p, miR155-5p and miR210-5p showed inhibition of radiation-induced apoptosis (DNA fragmentation formation) from  $5.7 \pm 0.6$ -fold increase to  $3.5 \pm 0.2$ ,  $3.7 \pm 0.6$  and  $3.0 \pm 0.8$ -fold change, respectively, compared with non-radiation exposed cells ( $P < 0.05$ ; Figure 8b).

## DISCUSSION

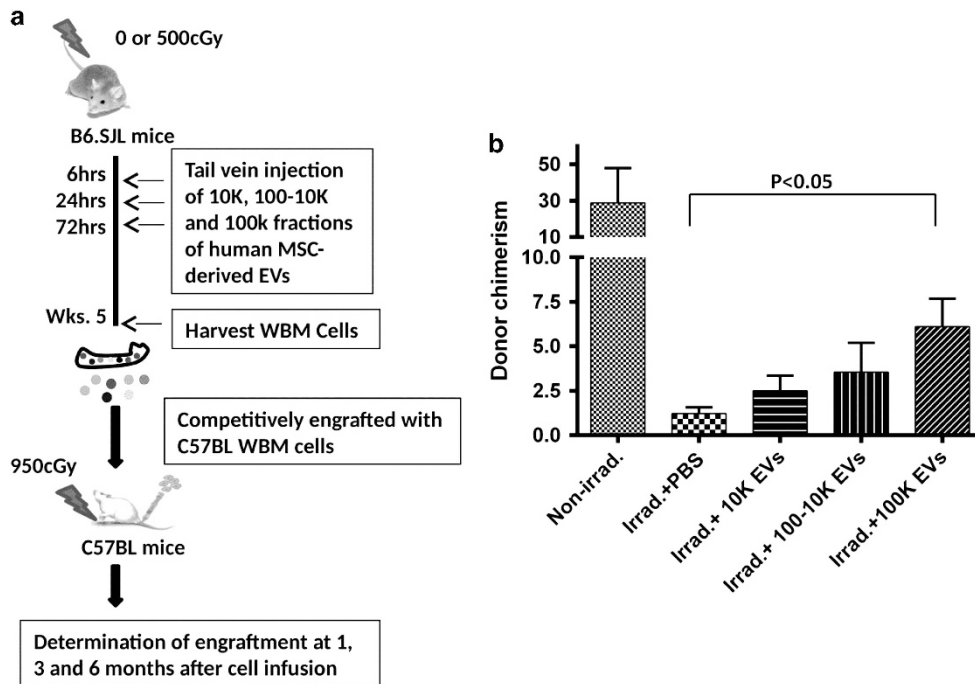
In this study, we have demonstrated that vesicles derived from murine or human marrow MSCs or from murine WBMcs are able to reverse radiation injury to murine bone marrow *in vivo* and *in vitro*. In addition, our study showed that vesicles stimulate

proliferation and reverse radiation-induced DNA damage and apoptosis in FDC-P1 cells.

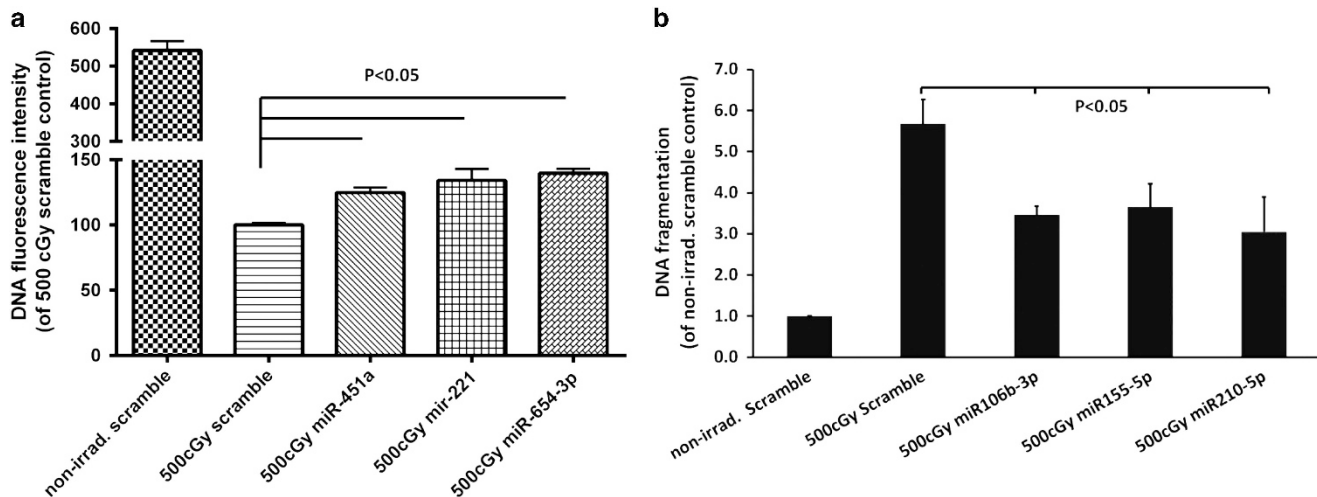
Many studies have demonstrated that the administration of MSCs can protect and reverse radiation damage to bone marrow<sup>15-17</sup> or other tissues.<sup>36-42</sup> However, the mechanisms underlying these beneficial effects are largely unknown and controversial. Yang *et al.*<sup>15</sup> and Lange *et al.*<sup>17</sup> reported that after intravenous infusion of MSC, there was only a small number of MSC found in the target organ, but there was a significant increase of hematopoietic recovery after irradiation. These data are consistent with studies on MSC reversal of kidney injury where MSC transiently accumulated in the injured kidney following intravenous infusion, but only few of these permanently engrafted within the kidney.<sup>43-46</sup> Recent studies showed that the conditioned medium or EVs from MSCs had a similar effect on reversal of tissue injury,<sup>26,28,47,48</sup> indicating that MSC engraftment or repopulation of target cells by MSCs might not be required. In our study, we demonstrated that MSC-EVs clearly reverse the radiation damage to marrow hematopoietic cells both *in vivo* and *in vitro*, which supports the hypothesis that MSC acts as a paracrine mediator to repair injured target cells.<sup>10,11,49</sup>

EVs have the capacity to transfer biological information from parent cells to target cells. Several studies have provided evidence that vesicles could transfer mRNA to target cells.<sup>21,24,25,50</sup> It was also demonstrated that vesicles from injured lung enter target marrow cells and induce lung-specific genes expression via both transfer of mRNA and a transcriptional modulator.<sup>51</sup> Work using mouse hybrid co-culture with reverse transcriptase-PCR primers specific for rat or mouse surfactant B and C indicated that if rat lung was cultured across from mouse marrow both rat and mouse surfactant B and C mRNA were detected early but only mouse





**Figure 7.** Evaluation of different fractions of human MSC-EVs for facilitating recovery from radiation injury *in vivo*. **(a)** Schematic diagram of experimental design. **(b)** Bone marrow was collected at 6 months post-transplant and analyzed for chimerism by FACS analysis. Each bar represents the average donor chimerism (mean ± s.e.m., 6 mice/group). \* $P < 0.05$  (Irradiation+PBS vs Irradiation+100 K by non-parametric Mann-Whitney *U*-test).



**Figure 8.** Overexpression of miRNA in FDC-P1 cells enhanced recovery from radiation. **(a)** miRNA enhances radiation-injured FDC-P1 cell proliferation. **(b)** miRNA prevents radiation-induced apoptosis in FDC-P1 cell (mean ± s.d.,  $n = 3$ /group).

mRNA persisted in cytokine supported cultures. Thus a stable epigenetic change was induced by lung vesicles in the target marrow cells. These effects appear to be variably RNase sensitive and more recently, non-specific miRNA depletion of vesicles from Drosha knockdown-MSC was shown to abrogate the protective effect of vesicles in a kidney injury model,<sup>52</sup> indicating that the miRNA in the vesicles may play a critical transcriptional role in the healing capacity of MSC. Our data showed that overexpression of EV-enriched miRNA in FDC-P1 cells could partially reverse the radiation damage. We found that overexpression of miR221, miR451 and miR654-3p stimulated cell growth and overexpression of miR210-5p, miR106b-3p and miR155-5p prevented radiation-induced apoptosis. Interestingly, Grosso *et al.*<sup>53</sup> indicated that overexpression of miR210 increases radioresistance and promotes a

more efficient DNA repair. This suggests that vesicles mediate tissue repair via a noncoding RNA-induced epigenetic alteration in injured tissue. The underlying mechanisms need further investigation.

We demonstrated that different MSC-EV populations have varied effects on reversal of radiation damage on murine HSC. The combined fractions (microvesicles and exosomes) showed the greatest reversal effects. Microvesicles and exosomes are the two major types of EVs. However, there is large overlap between these vesicle populations: this led to the suggestion that it is appropriate to simply designate them as EVs and then define them further by the cell of origin and the context of specific experiments. The diversity of proteins and nucleotides between exosomes and microvesicles has been reported.<sup>54,55</sup> We have determined vesicular protein or RNA per particle of vesicle from different subsets of

human MSC-EVs. The amount of protein in the particle is variable among microvesicle and exosome, and the combination of these two subsets is  $4.6 \pm 3.2$ ,  $1.9 \pm 1.0$  and  $2.8 \pm 2.3 \times 10^{-6}$  ng per particle, respectively. The amount of RNA is  $9.1 \pm 4.1$ ,  $4.8 \pm 2.9$  and  $5.4 \pm 3.4 \times 10^{-9}$  ng per particle, respectively. This is not related to huge difference of volume of EVs. Our data showed that different fractions of MSC-EVs facilitated recovery from radiation injury. This suggested that different populations of vesicles carry different information packets that might be selected and transferred to specific target cells.

Apoptosis and cell proliferation are critical for the maintenance of homeostasis in the hematopoietic system. DNA damage and apoptosis is an important cause of bone marrow irradiation injury.<sup>56–58</sup> The low-dose radiation exposure is able to result in a long time injury of HSC.<sup>3,57</sup> The immediate cellular response to DNA damage from radiation is cell cycle arrest. Here, we found that EVs from MSC or WBM could downgrade phosphorylation of H2AX after radiation exposure, indicating an acceleration of DNA repair efficiency or inhibition of DNA damage. We also showed that EV treatment could inhibit apoptosis and induce proliferation after radiation exposure *in vitro*. These are consistent with findings that MSC-derived EVs caused significant decrease of apoptosis in tubular cells of an acute kidney injury mice model<sup>59</sup> and increased proliferation.<sup>10</sup> Thus, the mechanisms underlying recovery of hematopoietic cells by vesicles are involved not only in attenuating DNA damage and apoptosis but also in stimulating proliferation.

Transplantation of MSC alone or with HSC has been shown to partially enhance engraftment and improve bone marrow recovery from radiation injury.<sup>12–18</sup> But it requires a period of time for preparation including a donor search before transplantation, therefore limiting its clinical application, especially in the case of a radiation emergency. Our data show that MSC-EVs partially reverse radiation damage to marrow stem cells. This EV-based, cell-free therapeutic strategy provides a convenient and safe therapeutic potential for rescuing marrow damage in patients treated with marrow toxic agents compared with MSC transplant. Early intervention is recommended in order to obtain the better long-term benefit. Some reports have shown that the best time to rescue the radiation injury is during the first 24–52 h.<sup>60</sup> Here, we showed that MSC-EVs reverse the radiation damage from 4 h to 1 week post-radiation exposure. Vesicles thus represent an interesting therapeutic strategy for the possible use in a radiation accident.

In summary, MSC-EVs reverse bone marrow cell radiation damage by accelerated HSC proliferation and differentiation and inhibition of DNA damage and apoptosis. EV reversal of marrow stem cell radiation damage suggests a unique new approach to radiation mitigation.

## CONFLICT OF INTEREST

The authors declare no conflict of interest.

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## AUTHOR CONTRIBUTIONS

This study was designed, supervised and coordinated by PQ. The manuscript was written by SW, revised by PQ and commented on by all authors. SW designed, performed experiments, collected, analyzed and interpreted data. MD designed and coordinated study. YC and CS performed experiments. EP and YD performed flow cytometry analysis. MDT, MP and AC conducted bleeding animals for engraftment analysis. MD, LG and JA contributed to mice

transplantation, DC, SB and FC provided technical advice, and GC gave conceptual advice and GC and DC edited the manuscript.

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Supplementary Information accompanies this paper on the Leukemia website (<http://www.nature.com/leu>)