## ANKRD11 variants: KBG syndrome and beyond

 Alma Kuechler ${ }^{1}$ | Emanuele Agolini ${ }^{5}$ ( ${ }^{\text {( }}$ | Beate Albrecht ${ }^{1}$<br>Carolina Baquero-Montoya ${ }^{6,7}$ | Axel Bohring ${ }^{8}$ | Nuria C. Bramswig ${ }^{1}$ | Andreas Busche ${ }^{8}$ | Andreas Dalski ${ }^{3}$ | Yiran Guo ${ }^{9}$ | Britta Hanker $^{3}$ | Yorck Hellenbroich ${ }^{3} \mid$ Denise Horn ${ }^{10}$ © $\mid$ A. Micheil Innes ${ }^{11}{ }^{(0)} \mid$ Chiara Leoni ${ }^{12}$ © $\mid$ Yun R. Li ${ }^{9,13}$ | Sally Ann Lynch ${ }^{14}$ © $\mid$ Milena Mariani ${ }^{15}$ © $\mid$ Livija Medne ${ }^{16,17}$ Barbara Mikat ${ }^{1}$ | Donatella Milani ${ }^{18}$ © | Roberta Onesimo ${ }^{12}$ © | Xilma Ortiz-Gonzalez ${ }^{19,20}$ © | Eva Christina Prott ${ }^{1,21}$ | Heiko Reutter ${ }^{22,23}$ ( ${ }^{\text {( }}$ | Eva Rossier ${ }^{24,25}$ | Angelo Selicorni ${ }^{15}$ | Peter Wieacker ${ }^{8}$ | Alisha Wilkens ${ }^{16}$ | Dagmar Wieczorek ${ }^{26}$ © | Elaine H. Zackai ${ }^{16,17}$ | Giuseppe Zampino ${ }^{12}$ Birgit Zirn ${ }^{25}$ | Hakon Hakonarson ${ }^{9,17}$ ( $\mid$ Matthew A. Deardorff ${ }^{27,28}$ | Gabriele Gillessen-Kaesbach ${ }^{3}$ | Frank J. Kaiser ${ }^{1,29}$<br>${ }^{1}$ Institut für Humangenetik, Universitätsklinikum Essen, Universität Duisburg-Essen, Essen, Germany<br>${ }^{2}$ Department of Internal Medicine, Thomas Jefferson University Hospital, Philadelphia, Pennsylvania, USA<br>${ }^{3}$ Institut für Humangenetik, Universität zu Lübeck, Lübeck, Germany<br>${ }^{4}$ Genetica Medica, Dipartimento di Scienze della Salute, Università degli Studi di Milano, Milan, Italy<br>${ }^{5}$ Laboratory of Medical Genetics, Ospedale Pediatrico Bambino Gesù, IRCCS, Rome, Italy<br>${ }^{6}$ Department of Pediatrics, Hospital Pablo Tobón Uribe, Medellín, Colombia<br>${ }^{7}$ Genetics Unit, Sura Ayudas Diagnosticas, Medellín, Colombia<br>${ }^{8}$ Institut für Humangenetik, Westfälische Wilhelms-Universität, Münster, Germany<br>${ }^{9}$ Center for Applied Genomics and Center for Data Driven Discovery in Biomedicine, Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA<br>${ }^{10}$ Institute of Medical and Human Genetics, Charité-Universitätsmedizin Berlin, Berlin, Germany<br>${ }^{11}$ Department of Medical Genetics and Alberta Children's Hospital Research Institute, University of Calgary, Calgary, Alberta, Canada<br>${ }^{12}$ Center for Rare Diseases and Birth Defects, Department of Woman and Child Health and Public Health, Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy<br>${ }^{13}$ Medical Scientist Training Program, University of Pennsylvania, Perelman School of Medicine, Philadelphia, Pennsylvania, USA<br>${ }^{14}$ Department of Clinical Genetics, Children's Health Ireland (CHI) at Crumlin, Dublin, Ireland<br>${ }^{15}$ Centro Fondazione Mariani per il Bambino Fragile ASST-Lariana Sant'Anna Hospital, Department of Pediatrics, San Fermo della Battaglia (Como), Italy<br>${ }^{16}$ Division of Human Genetics, Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA<br>${ }^{17}$ Department of Pediatrics, Children's Hospital of Philadelphia and Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA<br>${ }^{18}$ Fondazione IRCCS Cà Granda Ospedale Maggiore Policlinico Milano, Milan, Italy<br>${ }^{19}$ Department of Pediatrics, Division of Neurology, Epilepsy Neurogenetics Initiative, Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA<br>${ }^{20}$ Department of Neurology, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA<br>${ }^{21}$ Institut für Praenatale Medizin \& Humangenetik, Wuppertal, Germany<br>${ }^{22}$ Institute of Human Genetics, University Hospital of Bonn, Bonn, Germany

[^0]${ }^{23}$ Department of Neonatology and Pediatric Intensive Care, University Hospital of Bonn, Bonn, Germany<br>${ }^{24}$ Institut für Medizinische Genetik und Angewandte Genomik, Universität Tübingen, Tübingen, Germany<br>${ }^{25}$ Genetikum Stuttgart, Genetic Counselling and Diagnostics, Stuttgart, Germany<br>${ }^{26}$ Institute of Human Genetics, Medical Faculty and University Hospital Düsseldorf, Heinrich-Heine-University Düsseldorf, Düsseldorf, Germany<br>${ }^{27}$ Department of Pathology and Laboratory Medicine and Pediatrics, Children's Hospital Los Angeles, Los Angeles, California, USA<br>${ }^{28}$ Keck School of Medicine, University of Southern California, Los Angeles, California, USA<br>${ }^{29}$ Essener Zentrum für Seltene Erkrankungen (EZSE), Universitätsmedizin Essen, Essen, Germany

## Correspondence

Ilaria Parenti, Institut für Humangenetik, Universitätsklinikum Essen, Universität Duisburg-Essen, Hufelandstrasse 55, D-45122 Essen, Germany.
Email: ilaria.parenti@uk-essen.de

## Funding information

European Reference Network on Rare Congenital Malformations and Rare Intellectual Disability (ERN-ITHACA), Grant/Award Number: 3HP-HP-FPA ERN-01-2016/739516


#### Abstract

Mutations affecting the transcriptional regulator Ankyrin Repeat Domain 11 (ANKRD11) are mainly associated with the multisystem developmental disorder known as KBG syndrome, but have also been identified in individuals with Cornelia de Lange syndrome (CdLS) and other developmental disorders caused by variants affecting different chromatin regulators. The extensive functional overlap of these proteins results in shared phenotypical features, which complicate the assessment of the clinical diagnosis. Additionally, re-evaluation of individuals at a later age occasionally reveals that the initial phenotype has evolved toward clinical features more reminiscent of a developmental disorder different from the one that was initially diagnosed. For this reason, variants in ANKRD11 can be ascribed to a broader class of disorders that fall within the category of the so-called chromatinopathies. In this work, we report on the clinical characterization of 23 individuals with variants in ANKRD11. The subjects present primarily with developmental delay, intellectual disability and dysmorphic features, and all but two received an initial clinical diagnosis of either KBG syndrome or CdLS. The number and the severity of the clinical signs are overlapping but variable and result in a broad spectrum of phenotypes, which could be partially accounted for by the presence of additional molecular diagnoses and distinct pathogenic mechanisms.


## KEYWORDS

ANKRD11, chromatinopathies, Cornelia de Lange syndrome (CdLS), developmental disorders, KBG syndrome (KBGS)

## 1 | INTRODUCTION

Transcriptional regulators are key players in numerous biological processes. Ankyrin Repeat Domain 11 (ANKRD11) is an important coregulator able to induce changes in gene expression by recruiting chromatin remodelers to target genes upon interaction with specific transcriptional repressors or activators. ${ }^{1,2}$ The corresponding gene (OMIM *611192) is located at 16q24.3 and encodes a 298 kDa protein of 2663 amino acids containing five ankyrin repeats (amino acids 162-284), two repression domains (amino acids 318-611 and 23692663) and one activation domain (amino acids 1851-2145). ${ }^{1}$ Due to its unique structure, ANKRD11 is believed to mediate both transcriptional activation and repression. ${ }^{1,3}$ ANKRD11 is best characterized for its function as a co-regulator in the developing brain, where it plays a critical role for the proliferation of neural progenitors, for the genesis and positioning of newborn neurons, ${ }^{4}$ for neuronal plasticity ${ }^{5}$ and for dendritic differentiation. ${ }^{6}$

ANKRD11 was first associated with human disease when deletions at 16q24.3 were identified in individuals with autism spectrum disorder (ASD). ${ }^{7}$ Two years later, Willemsen and colleagues provided evidence for a novel microdeletion syndrome by describing four patients characterized by ASD, variable levels of intellectual disability and dysmorphic features carrying interstitial deletions at $16 q 24.3$. ${ }^{8}$ Subsequent reports of individuals with intellectual disability, facial dysmorphism, and ASD allowed the narrowing of the minimal common region of overlap of this 16q24.3 microdeletion syndrome to ANKRD11 only, suggesting a role of ANKRD11 in neurodevelopment. ${ }^{9,10}$

The first point mutations in ANKRD11 were identified in seven individuals with KBG syndrome (KBGS, OMIM \#148050). ${ }^{5}$ This is a rare disorder named after the initials of the first three affected individuals and characterized by intellectual disability, global developmental delay, short stature, skeletal anomalies, distinctive facial features, and macrodontia of the upper central incisors. ${ }^{11}$ Since the first description by Sirmaci and colleagues, ${ }^{5}$ additional individuals with KBGS have
been reported to carry point mutations, duplications or microdeletions involving ANKRD11, thus pointing to ANKRD11 as the main gene responsible for this syndrome. ${ }^{12-27}$ Importantly, a marked interfamilial and intra-familial phenotypical variability has been reported in association with KBGS, indicating variable expressivity and penetrance. ${ }^{11,20}$ With the falling cost and increasing accessibility of next generation sequencing technologies and microarrays, variants in ANKRD11 have also been reported in association with neurodevelopmental syndromes other than KBGS. Specifically, an individual with an initial clinical diagnosis of Silver-Russell syndrome was found to harbor a 348 kb microdeletion at 16q24.3 encompassing ANKRD11 and SPG7. ${ }^{28}$ Point mutations in ANKRD11 were also identified in subjects with phenotypes reminiscent of Cornelia de Lange syndrome (CdLS) (OMIM \#122470). ${ }^{29-32}$ Loss-of-function variants in ANKRD11 were similarly described in association with Coffin-Siris syndrome (CSS) (OMIM \#135900). ${ }^{23}$ Importantly, CdLS and CSS clinically overlap to some extent with KBGS. The shared clinical features include a variable degree of developmental delay and intellectual disability, growth retardation, limb anomalies and characteristic facial dysmorphism. ${ }^{23,29,30}$ These findings suggest that variants in ANKRD11 are not necessarily associated with KBGS only, but that they are rather linked to a larger spectrum of neurodevelopmental syndromes. Accordingly, ANKRD11 has been described as one of the most frequently mutated genes in individuals with severe developmental disorders. ${ }^{33,34}$

In this work we discuss the clinical and molecular findings of 23 individuals with variants in ANKRD11 and describe a wide spectrum of phenotypes associated with mutations in this gene.

## 2 | MATERIALS AND METHODS

## 2.1 | Cohort

Individuals herein described were recruited thanks to a large international cooperation that includes Germany, Italy, Ireland, Colombia, Canada, and the United States.

Procedures including subjects were initially approved by the Ethical Committee of the University of Lübeck (approval number for human studies HLO7-158) and the Ethical Committees of the respective institutions. All procedures were performed in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. Informed consent was obtained from all individuals included in this study. An additional informed consent was collected for the publication of subjects' photographs.

Individuals were analyzed by means of exome sequencing, gene panels or microarrays at their respective institutions. Referring physicians provided detailed developmental, neurological, and behavioral history of the subjects. Variants were described on the ANKRD11 NM_013275.6 RefSeq transcript using HGVS recommendations. ${ }^{35}$ All variants have been submitted to the ClinVar database and have been assigned the following accession numbers: SCV001478030-SCV001478045.

## 2.2 | Facial dysmorphology novel analysis

The Facial Dysmorphology Novel Analysis (FDNA Inc., Boston, MA) technology combines facial recognition software with biological knowledge. This technology allows to detect dysmorphic features and recognizable patterns of facial malformations from 2D facial photographs. Face2Gene (FDNA) was used as a tool for computer analysis of subjects' photographs (https://face2gene.com). ${ }^{36}$

## 3 | RESULTS

## 3.1 | Individuals

The group of individuals described herein is composed of 11 females and 12 males, with an age range extending from four to 23 years. The median age of the initial clinical diagnosis was 6 years and 5 months, whereas the median age of the last clinical examination was 9 years and 3 months. Phenotypical appearance of the individuals can be found in Figure 1.

Thirteen individuals received a clinical diagnosis of KBGS (Individuals 1, 8, 10, 11, 14, 15, 17, 18, 19, 20, 21, 22, and 23). Three individuals received an initial clinical diagnosis of CdLS or atypical CdLS that was reconfirmed at a later re-evaluation (Individuals 2, 5, and 6), while five individuals were diagnosed as CdLS during early childhood but were reclassified as KBGS after a re-evaluation later in life (Individuals 3, 4, 7, 9, and 12). Two individuals presented with nonspecific syndromic intellectual disability and developmental delay (Individuals 13 and 16). A schematic representation of the diagnostic evaluation of the individuals of the present series is provided in the Figure S1.

An additional clinical analysis was carried out with the Face2Gene database for all individuals for whom photographic material was available (all except Individuals 9 and 14) (Table 1). The software assigned a likely diagnosis of KBGS to 17 out of 21 individuals (Individuals 1, 2, $3,4,6,7,8,10,11,12,13,16,17,18,19,21$, and 22). CdLS was considered a possible diagnosis with medium-low probability for Individuals $2,4,6,7,11$, and 12 . No obvious diagnosis was assigned to Individuals 5, 15, 20, and 23. Additional syndromes with medium-low similarity scores that were contemplated as differential diagnosis in at least five individuals of our cohort include Wiedemann-Steiner syndrome, fetal alcohol syndrome, PMM2-related disorder and WilliamsBeuren syndrome.

## 3.2 | Clinical features

An overview of the clinical features of each individual is listed in Table 1.

Milestones in motor and verbal development were found to be delayed for all individuals but one: sitting independently was achieved at a median age of 12 months and walking independently at 24 months. The median age of pronouncing the first words was


FIGURE 1 Phenotypical appearance of the following individuals: (A) Individual 1, age 13; (B) Individual 2, age 8; (C) Individual 3, age 7.5; (D) Individual 5, age 17; (E) Individual 6, age 4; (F) Individual 8, age 10; (G) Individual 10, age 9; (H) Individual 11, age 3; (I) Mother of Individual 13, age 21; (J) Individual 15, age 8; (K) Individual 16, age 13; (L) Individual 17, age 9; (M) Individual 18, age 9; (N) Individual 19, age 7;
(O) Individual 20, age 6; (P) Individual 21, age 4; (Q) Individual 22, age 5; (R) Individual 23, age 14

24 months. Individuals 11, 13, and 21 are currently still non-verbal at an age of 6,4 , and 15 years, respectively.

Intellectual disability and behavioral problems were also detected in the large majority of the individuals ( $85 \%$ and $68 \%$ of subjects, respectively). The level of intellectual disability could be assessed in four individuals and appeared moderate in one and mild in three subjects. Behavioral problems ranged from shyness or inability to recognize and respect personal boundaries to aggressiveness, autistic features and attention deficit hyperactivity disorder.

The most frequent phenotypical features found in our cohort comprise a characteristic face wide at the zygoma (70\%), low anterior hairline (65\%), synophrys (65\%), thick eyebrows (70\%), long eyelashes (78\%), anteverted nostrils (78\%), broad nasal tip (70\%), thick alae nasi (65\%), long philtrum (83\%), macrodontia of central incisors (65\%),
delayed bone age (67\%), short fifth finger (61\%), and clinodactyly of the fifth finger (70\%).

Additional features commonly observed include arched eyebrows (48\%), smooth philtrum (48\%), thin upper vermilion (52\%), brachydactyly (48\%), small hands and feet (52\%), proximally set thumbs (48\%), visual problems (50\%), and delayed dentition (50\%).

## 3.3 | Clinical re-evaluation and age-dependent phenotypical evolution

Five individuals of the present cohort were diagnosed as CdLS during early childhood but were reclassified as KBGS after phenotypical re-evaluation (Subjects 3, 4, 7, 9, and 12). Table 2 provides an


| TABLE 1 | (Continued) |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Mouth | Large mouth | - | - | - | + | - | + | - | - | + | - | + | - |
|  | Downturned corners of the mouth | - | + | - | - | - | + | - | + | - | - | + | - |
|  | Thin upper vermilion | - | + | + | - | + | + | + | + | - | - | + | - |
|  | Thick lower vermilion | - | + | - | - | - | - | - | + | + | + | - | + mild |
|  | Macrodontia of central incisors | + | NA | + | + | - | + | - | + | + and Talon cusps | + | - | + |
|  | Delayed dentition | NA | NA | NA | NA | NA | + | - | NA | - | - | - | NA |
| Skin | Hypertrichosis | - | - | + | - | + | + | + | - | NA | - | - | + |
|  | Hairy elbows | - | - | - | - | NA | + | - | - | - | - | - | - |
| Skeleal | Small hands/feet | - | + | + | + | + | - | - | - | + | Small toes, normal lenght of hands and feet | - | - |
|  | Brachydatyly | - | + | + | + | + | - | - | Mildly broad and short terminal phalanges | + | - | - | + mild |
|  | Proximally set thumbs | - | - | - | + | + | - | + | - | . | + slighty | - | + |
|  | Clinodactryly fith finger | - | + mild | - | + | - | - | - | + mild | + | + | + | - |
|  | Short fifth finger | + | + mild | + | + | + | - | - | + mild | + | + | + | - |
|  | A/Hypoplasia of the distal phalanges of the fifth finger | - | - | - | + | + | - | - | - | - | Short middle phalanx | - | - |
|  | A/Hypoplastic nails | - | - | - | - | + | + | - | - | - | - | - | - |
|  | Syndactyly of toes | - | - | + | NA | - | - | - | - | . | Mild basal syndactyly II/III hand and feet | - | - |
|  | Prominent interphalangeal joints | - | - | - | - | + | - | - | - | $\cdot$ | - | - | - |
|  | Joint laxity | NA | NA | - | NA | - | - (pes planum) | - | NA | - | - | - | - |
|  | Delayed bone age | NA | NA | NA | NA | NA | NA | NA | NA | + | NA | NA | + |
| Other | Cardiac anomalies | - | + peripheral pulmonary stenosis (normalized) | - | - | - | - | - | - | - | + ASD (subclinical) / VSD (spontaneously closed) | - | - |
|  | Genitourinary anomalies | - | - | - | - | - | - | - | - | + crptorchidism | - | - | NA |
|  | CNS anomalies | - | - | - | - | - | + | NA | MRI: small cerebellar vermis, arachnoidal cyst | + | no MR1 performed | - | - (mildly enlarged outer CNF spaces frontotemporal) |
|  | Feeding problems | - | + | - | - | - | - | - | - | - | - | + | + in infancy |
|  | Hearing problems | + | - | - | NA | - | - | - | - | - | - | - | - |
|  | Seizures | - | - | - | - | - | - | - | One seizure, EEG intermittent deceleration right parieto-occipital (no medication) | + | - | - | + (for 2 years starting at age 4 years 6 months. Valproate until age 10 now seizure-free without medication but EEG changes) |
|  | Frequent infections | - | - | - | - | - | - | - | - | $\cdot$ | - | - | - (but bronchial problems due to prematurity) |
| Cognitive development | ID | + | - (1Q:104) | + | + | - | + (10:39) | + (mild) | - | + | $\begin{aligned} & \text { + (IQ-Test 2014: 78; IQ- } \\ & \text { Test 2016: 58) } \end{aligned}$ | + | + (1Q: 50-75) |
|  | Behavioral problems | + ASD-ike | + | + | + | NA | + | - | - | + | +aggressiveness, trouble in social groups | - | + ADHD |
| Motor development | It Age of sitting independenty | NA | 17 months | NA | NA | NA | 12 months | 9 months | NA | 18 months | 13-14 months | 3 years | NA |
|  | Age of walking independently | after 18 months | 22 months | NA | NA | 18 months | 15 months | 24 months | 22 months | 24 months | 24 months | 3.5 years | 18-19 months |
| Verbal development | Age of first words | after 12 months | 2 years | Delayed | NA | 24 months | 18 months | 16 months | 18 months | 4 years | 24 months | No speech development | 30 months |
| Other |  | Hypopigmentation | Epicanthus, upslanting palpebral fissures, large protruding ears, rhinophonia aperta, velopharyngeal incompetence (surgically corrected). muscular hypotonia |  |  | Congenital hip dysplasia |  |  | Muscular hypotonia, protruding ears |  | High palate, maxillary prognatism, vermillion of upper lip tended |  | Pulmunary displasia, cerebral hemorrage |


| 22 | 23 |
| :---: | :---: |
| USA | USA |
| f | m |
| KBG | KBG |
| KBG (high) | Neurofibromatosis type 1 (medium) |
| Wiedemann-Steiner (mediuum) | Noonan syndrom (low), Turner syndrome (low), Sotos syndrome (low) |
| ANKRD11 | ANKRD11 |
| 9 | 4-12 |
| c.1381_1384delGAAA | chr16:89,335,426-89,371,803del |
| p.(Gu461GInts ${ }^{\text {s }}$ - ${ }^{\text {/ }}$ |  |
| NA | denovo |
| NA | NA |
| NA | NA |
| NA | NA |
| NA | NA |
| 4 years 3 months | 13 years 7 months |
| $92.2 \mathrm{~cm}(-2.45 \mathrm{SD})$ | $158 \mathrm{~cm}(-0.37 \mathrm{SD})$ |
| $12.6 \mathrm{~kg}(-2.3 \mathrm{sD})$ | $40.9 \mathrm{~kg}(-1.0 \mathrm{SD})$ |
| $46 \mathrm{~cm}(-3.0 \mathrm{SD})$ | $54.9 \mathrm{~cm}(-0.19$ SD) |




| Other | Cardica anomalies |
| :---: | :---: |
|  | Genitourinary anomalies |
|  | CNS anomalies |
|  | Feeding problems |
|  | Hearing problems <br> Seizures |
|  | Frequent infections |
| Cognitive development |  |
| Motor development | Age of sitting independenty Age of walking independently |
| Vertal <br> development | Age of first words |
| Other |  |

TABLE 2 Comparison of the main clinical features of CdLS and KBG

|  |  | CdLS |  | Individual 3 <br> Single evaluation <br> 7 years <br> 6 months | Individual 4 <br> Single <br> evaluation <br> 9 years | Individual 7 |  | Individual 9 |  | Individual 12 |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | KBG | First evaluation 18 months |  |  | Last evaluation 6 years 10 months | First evaluation 3 years 3 months | Last evaluation 10 years <br> 4 months | First evaluation 5 years 2 months | Last evaluation <br> 11 years <br> 6 months |
| Growth | IUGR |  | + | - | - | NA | - |  | + |  | - |  |
|  | Postnatal short stature | + | + | - | + | $\pm$ | - | + | (-)(growth hormone treatment) | - | - |
|  | Microcephaly | + | - | - | - | - | - | + | NA | - | - |
| Craniofacial features | Brachycephaly | + | + | - | - | + | + | + | - | - | - |
|  | Low anterior hairline | + | $\pm$ | + | NA | + | + | + | + | + | + |
|  | Frontal bossing | - | + | - | - | - | + | - | - | - | - |
|  | Triangular face | - | + | + | + | - | $\pm$ | - | + | + | + |
|  | Prominent cheekbones | - | + | + | + | - | + | - | NA | + | + |
|  | Thick eyebrows | + | + | + | + | - | + | - | + | + | + |
|  | Synophrys | + | $\pm$ | + | + | $\pm$ | + | + | NA | - | - |
|  | Long eyelashes | + | + | + | - | $\pm$ | + | + | + | Not long, but prominent | Not long, but prominent |
|  | Depressed nasal bridge | + | - | - | - | $\pm$ | - | - | - | - | - |
|  | Prominent nasal bridge | - | + | + | - | - | $\pm$ | - | NA | mild | mild |
|  | Anteverted nostrils | + | + | + | + | + | - | + | + | + | + |
|  | Bulbous nasal tip | - | + | - | - | - | $\pm$ | - | + | (+) | (+) |
|  | Long smooth philtrum | + | + | + | Long | Long | + | - | + | + | + |
|  | Thin upper lip | + | + | + | - | + | $\pm$ | - | - | + | + |
|  | Downturned corners of the mouth | + | - | - | - | - | - | - | NA | - | - |
|  | Widely spaced teeth | + | - | + | - | NA | - | - | NA | - | - |
|  | Macrodontia | - | + | - | + | - | - | - | + | + | + |
|  | Micrognathia | + | - | - | - | $\pm$ | - | - | NA | Mild | Mild |

Abbreviations: CdLS, Cornelia de Lange syndrome; NA, not assessed; IUGR, intrauterine growth retardation.


FIGURE 2 Distribution of the ANKRD11 variants at the protein and DNA level. (A) Schematic representation of the ANKRD11 protein and its domains, with relative position of the identified variants (generated with PROSITE MyDomains ${ }^{37}$ ). The ankyrin repeats are shown in blue, the repression domains in green and the activation domain in orange. Loss-of-function mutations are depicted in red, whereas the in-frame deletion is depicted in gray. (B) Schematic representation of the coding ANKRD11 exons in scale with the above shown protein. The ankyrin domain is encoded by exons 6-8, the activation and first repression domains by exon 9, and the second repression domain by exons 9-13
overview of the clinical features of these individuals as well as of the differences and commonalities between "classic" manifestations of CdLS and KBGS, with a particular focus on facial dysmorphism.

Individuals 3 and 4 were examined only once by the referring physicians. For both, the facial features at a first evaluation were considered reminiscent of CdLS and, consequently, a clinical diagnosis of CdLS was assigned. After the identification of the ANKRD11 variants, the clinical features of the individuals were re-evaluated and found compatible with ANKRD11-associated KBGS. Individual 12 was examined twice (before and after the molecular diagnosis). No significant phenotypical evolution was observed for this individual but, similarly to Individuals 3 and 4, the re-evaluation of the clinical features appeared consistent with the recently disclosed molecular diagnosis.

Individuals 7 and 9 received a clinical diagnosis of CdLS at an early age. The diagnosis of Individual 7 was mainly driven by the facial features, which included brachycephaly, low anterior hairline, synophrys, long eyelashes, depressed nasal bridge, anteverted nostrils, long philtrum, thin upper lip, and micrognathia. Individual 9 was characterized by intrauterine growth retardation, microcephaly, brachycephaly, low anterior hairline, synophrys, long eyelashes, and anteverted nares. At the latest evaluation, an evolution of the phenotype was observed, primarily related to the shape of the face and the nose. Both subjects developed a triangular shaped face and a bulbous nasal tip. In addition, Individual 7 presented with frontal bossing and prominent cheekbones, while the permanent dentition of Individual 9 featured macrodontia.

## 3.4 | Molecular findings

Seventeen distinct ANKRD11 variants were identified in our cohort composed of 23 individuals (Figure 2), including seven out-of-frame deletions (c.915delA, p.(Pro306Hisfs*62); c.1711_1723del, p.(Thr571Alafs*15); c.2398_2401del, p.(Glu800Asnfs*62); c.2408_2412del, p.(Lys803Argfs*5);
c.1903_1907del, p.(Lys635Glnfs*26); c.1381_1384del, p.(Glu461Glnfs*48); c.3591_3594del, p.(Lys1198Argfs*119)), two out-of-frame duplications (c.7356dupC, p.(Lys2453Glnfs*79); c.3888dupC, p.(Asn1297GInfs*3)), five nonsense variants (c.1977C>A, p.(Tyr659*); c.2692C>T, p.(Arg898*); c.4218C>A, p.(Tyr1406*); c.4087C>T, p.(Arg1363*); c.5123C>A, p. (Ser1708*), one in-frame deletion (c.7411_7422del, p.(Thr2471_ Gly2474del)), one splicing variant (c. $7470+2 \mathrm{~T}>\mathrm{C}$ ) and one deletion encompassing multiple exons (chr16:89,335,426-89,371,803del). All point variants primarily involve exon 9 of ANKRD11, hence confirming the role of this exon as mutational hotspot of ANKRD11. ${ }^{17,27}$ The four amino acids deletion (Individual 8; c.7411_7422del; p.(Thr2471_Gly2474del)) is located in the highly conserved C -terminal repression domain, which is important for proteasome-mediated degradation of ANKRD11. ${ }^{18}$ This variant might therefore impair the functional activity of the protein and/or trigger a dominant negative effect upon dimerization with wild type ANKRD11. The remaining loss-of-function variants are instead predicted to activate nonsense-mediated mRNA decay, thereby resulting in ANKRD11 haploinsufficiency.

Of the mutations herein described, 11 are novel and six have been previously described, namely p.(Glu461Glnfs*48), p. (Lys635Glufs*26), p.(Tyr659*), p.(Glu800Asnfs*62), p.(Lys803Argfs*5), and $p$.(Arg1363*). The p.(Lys635Glufs*26) variant appears to be particularly frequent in the population of individuals with developmental disorders, as it was already reported in the literature in 10 different families and is also shared by four unrelated individuals of our cohort (Individuals 9, 14, 18, and 19). ${ }^{19,20,23,24,27,33,38-42}$

## 3.5 | Familial cases

The inheritance of the variants could be verified in 18 individuals. The mutation occurred de novo in 15 individuals and was maternally inherited in Individuals 4, 12, and 13. Detailed clinical data of the mother of Individual 12 are currently not available. The mother of Individual 4 was reported as mildly affected. She presents with short
stature, low weight, deep-set eyes, depressed nasal bridge, large mouth, proximally set thumbs, clinodactyly of the fifth finger and incomplete prono-supination of the elbow. The mother of Individual 13 (Figure 1(I)) received a clinical diagnosis of KBGS and displayed a low anterior hairline, arched and thick eyebrows with synophrys, long eyelashes, myopia, anteverted nostrils, thick alae nasi, a large mouth with thin upper vermilion and thick lower vermilion, macrodontia of central incisors and mild intellectual disability.

## 4 | DISCUSSION

ANKRD11 plays a pivotal role in the pathogenesis of KBGS and related disorders. Herein we report on 23 individuals carrying 17 distinct variants in ANKRD11, thereby expanding the cohort of individuals with mutations in this gene. Reasons for referral of index cases were growth retardation, facial dysmorphism and a variable degree of developmental delay and intellectual disability. All individuals underwent complete physical and dysmorphological evaluation from expert clinical geneticists and all but two (Individuals 13 and 16) received a clinical diagnosis of either CdLS or KBGS. Photographic material was submitted to Face2Gene for an additional clinical evaluation. This database includes an unprecedented amount of phenotypic and genotypic information associated with more than 10000 diseases and has proven to be a valuable tool for the interpretation of facial features. ${ }^{36,43,44}$ Face2Gene assigned a diagnosis of KBGS with high/ medium similarity scores to 17 out of 21 individuals for whom photographs were available. The remaining four individuals (Individuals $5,15,20$, and 23) were not associated to any syndrome with a high probability. Previous studies have proven that the Facial Dysmorphology Novel Analysis technology could match the capabilities of expert clinicians and in some cases also outperform them. ${ }^{36,43,44}$ Diagnosis-aiding tools are particularly important for syndromes like KBGS, for which some of the typical and most recognizable clinical features (i.e., macrodontia, delayed bone age, and a bulbous nasal tip/broad nasal bridge) might appear only later in life. Our data confirm the importance of facial analysis technologies as a tool to assist geneticists to assess the most appropriate clinical diagnosis in order to facilitate management and treatment of patients.

The most frequent features reported in our cohort and in the literature comprise intellectual disability, delayed or absent speech, motor delay, behavioral problems and a characteristic facial gestalt. Limb anomalies, delayed bone age, feeding difficulty and visual problems are also frequently observed. ${ }^{19,20}$ Number and severity of each of these clinical signs vary within our cohort. The specific combination of features can therefore lead to a broad spectrum of clinical phenotypes, ranging from mild to severe. Previous publications have shown that the severity of the phenotype does not depend on the type or position of the ANKRD11 variant. ${ }^{18-20}$ Comparison of clinical signs of all reported cases of recurrent mutations (in our cohort and in previously reported individuals) as well as of familial cases has confirmed the absence of a linear genotype-phenotype correlation and highlighted the existence of variable penetrance and intra-familial
variability. ${ }^{20}$ Importantly, the possibility of multiple molecular diagnoses should also be taken into account for an appropriate evaluation of these phenotypes, as they can influence the phenotypical appearance. ${ }^{45}$ The majority of our individuals was analyzed by targeted gene panel and we are therefore unable to exclude the presence of additional variants.

Variants in ANKRD11 have been mainly described in association with KBGS. ${ }^{5,19,20,27}$ Accordingly, based on a recent review, 171 out of 228 individuals with ANKRD11 variants reported in 38 different studies were formally diagnosed as KBGS. ${ }^{34}$ However, variants in this gene have also been identified in individuals with neurodevelopmental disorders other than KBGS, namely CdLS and CSS. ${ }^{23,29-32}$ These syndromes share some overlapping clinical features that may be difficult to discern from one another. ${ }^{11,46,47}$ Eight of the individuals reported here received an initial clinical diagnosis of mild CdLS or atypical CdLS. However, after re-evaluation later in life, a KBGS diagnosis was assigned to five of these individuals. Notably, for two of them, the primary phenotypical differences between the first and the last clinical evaluation were related to the shape of the face and the region of the nose, a finding that is compatible with the physiological progression into adolescence/adulthood. Together with a previously reported CdLS subject with a variant in ANKRD11, ${ }^{30}$ our cohort points to the existence of an age-dependent phenotypical evolution from CdLS to KBGS from infancy to adolescence, mainly concerning nose and facial contour. Therefore, clinical follow ups are crucial for the assessment of the proper clinical diagnosis.

Importantly, the protein complexes so far associated with CdLS, CSS, and KBGS are all involved in the regulation of transcription and chromatin structure. Epigenetic modifications and transcriptional dysregulation are therefore considered a key molecular feature of these disorders. ${ }^{6,48}$ ANKRD11 can control chromatin accessibility and mediate transcriptional regulation upon interaction with histone deacetylases and nuclear receptors. ${ }^{4,49}$ The cohesin complex, responsible for the onset of CdLS, is important for gene expression, DNA repair and long-range interactions between distant genomic regions. ${ }^{50}$ The CSS-associated SWItch/Sucrose Non-Fermentable (SWI/SNF) complex is well known for chromatin remodeling. ${ }^{47}$ Notably, the SWI/SNF complex is known to directly interact with the cohesin loader. In yeast, the SWI/SNF complex recruits the cohesin loader to nucleosome-free regions that the cohesin loader subsequently helps to maintain. ${ }^{51}$ A direct interaction between ANKRD11 and these two protein complexes has instead not been reported yet. The substantial functional overlap that characterizes these proteins as well as the direct physical interactions that have been reported for some of them could be found accountable for the shared clinical features observed in individuals with different neurodevelopmental disorders. The increasing accessibility of next generation sequencing technologies will allow the identification of additional variants in ANKRD11 in individuals with clinical diagnoses different from KBGS. Correspondingly, several variants in ANKRD11 were identified in individuals with severe undiagnosed developmental disorders and/or intellectual disability. ${ }^{33,34,52}$ Also in this cohort, variants in ANKRD11 have been reported in KBGS subjects but also in individuals with nonspecific
intellectual disability or with CdLS. For this reason, variants in this gene should be ascribed to a more ample group of neurodevelopmental disorders that fall within the category of chromatinopathies rather than to KBGS per se. ${ }^{53,54}$ In line with the recently proposed dyadic approach for the description of diagnostic entities, ${ }^{55}$ the disease phenotypes observed in association with variants in ANKRD11 could be defined as "ANKRD11-related chromatinopathies" or "ANKRD11-related neurodevelopmental disorders."

The growing number of variants in ANKRD11 and the varying severity of behavioral and developmental phenotypes associated with these variants make the understanding of the causative mechanisms particularly important. Since the levels of ANKRD11 are tightly regulated during the cell cycle, the most likely pathogenic mechanism is haploinsufficiency. ${ }^{18}$ However, analysis of ANKRD11 transcript levels in cell lines of patients has revealed escape from nonsense-mediated mRNA decay to some extent. ${ }^{15,18}$ Furthermore, some variants might also lead to a dominant negative pathogenic mechanism due to a lack of proteasome-mediated degradation of the truncated protein. This proposed mechanism depends on the dimerization between the N terminal repression domains of the wild type and mutant ANKRD11 when the degradation of the mutant protein is impaired by the disruption of the C-terminal degradation signals. ${ }^{18}$ The potential that a greater understanding of the molecular mechanism may lead to eventual therapeutic insights represents an exciting prospect.

## ACKNOWLEDGEMENTS

The authors are grateful to the individuals and their families for generously donating samples and clinical information. This work has been generated within the European Reference Network on Rare Congenital Malformations and Rare Intellectual Disability (ERN-ITHACA) (EU Framework Partnership Agreement ID: 3HP-HP-FPA ERN-01-2016/739516).

## CONFLICT OF INTEREST

The authors declare no conflict of interest.

## PEER REVIEW

The peer review history for this article is available at https://publons. com/publon/10.1111/cge. 13977.

## DATA AVAILABILITY STATEMENT <br> Not applicable.

## ORCID

Ilaria Parenti (D) https://orcid.org/0000-0002-1825-6237 Cristina Gervasini (D) https://orcid.org/0000-0002-1165-7935 Emanuele Agolini (D) https://orcid.org/0000-0001-6543-6225 Denise Horn (iD https://orcid.org/0000-0003-0870-8911 A. Micheil Innes (D) https://orcid.org/0000-0001-9881-5467

Chiara Leoni (D) https://orcid.org/0000-0002-4089-637X
Sally Ann Lynch (D) https://orcid.org/0000-0003-3540-1333
Milena Mariani (i) https://orcid.org/0000-0002-7446-7418

Donatella Milani (DD https://orcid.org/0000-0002-3087-8514
Roberta Onesimo (iD https://orcid.org/0000-0003-3128-6657
Xilma Ortiz-Gonzalez (D) https://orcid.org/0000-0002-1710-4696
Heiko Reutter (D) https://orcid.org/0000-0002-3591-5265
Dagmar Wieczorek (iD https://orcid.org/0000-0003-2812-6492
Hakon Hakonarson (iD https://orcid.org/0000-0003-2814-7461

## REFERENCES

1. Zhang A, Li C-W, Chen JD. Characterization of transcriptional regulatory domains of ankyrin repeat cofactor-1. Biochem Biophys Res Commun. 2007;358:1034-1040.
2. Li C-W, Dinh GK, Zhang A, Chen JD. Ankyrin repeats-containing cofactors interact with ADA3 and modulate its co-activator function. Biochem J. 2008;413:349-357.
3. Neilsen PM, Cheney KM, Li C-W, et al. Identification of ANKRD11 as a p53 coactivator. J Cell Sci. 2008;121:3541-3552.
4. Gallagher D, Voronova A, Zander MA, et al. Ankrd11 is a chromatin regulator involved in autism that is essential for neural development. Dev Cell. 2015;32:31-42.
5. Sirmaci A, Spiliopoulos M, Brancati F, et al. Mutations in ANKRD11 cause KBG syndrome, characterized by intellectual disability, skeletal malformations, and Macrodontia. Am J Hum Genet. 2011;89:289-294.
6. Ka M, Kim W-Y. ANKRD11 associated with intellectual disability and autism regulates dendrite differentiation via the BDNF/TrkB signaling pathway. Neurobiol Dis. 2018;111:138-152.
7. Marshall CR, Noor A, Vincent JB, et al. Structural variation of chromosomes in autism Spectrum disorder. Am J Hum Genet. 2008;82:477-488.
8. Willemsen MH, Fernandez BA, Bacino CA, et al. Identification of ANKRD11 and ZNF778 as candidate genes for autism and variable cognitive impairment in the novel 16q24.3 microdeletion syndrome. Eur J Hum Genet. 2010;18:429-435.
9. Youngs EL, Hellings JA, Butler MG. ANKRD11 gene deletion in a 17-year-old male. Clin Dysmorphol. 2011;20:170-171.
10. Isrie $M$, Hendriks Y , Gielissen N , et al. Haploinsufficiency of ANKRD11 causes mild cognitive impairment, short stature and minor dysmorphisms. Eur J Hum Genet. 2012;20:131-133.
11. Brancati F, Sarkozy A, Dallapiccola B. KBG syndrome. Orphanet J Rare Dis. 2006;1:50.
12. Sacharow S, Li D, Fan YS, Tekin M. Familial 16q24.3 microdeletion involving ANKRD11 causes a KBG-like syndrome. Am J Med Genet A. 2012;158A:547-552.
13. Khalifa M, Stein J, Grau L, et al. Partial deletion of ANKRD11 results in the KBG phenotype distinct from the $16 q 24.3$ microdeletion syndrome. Am J Med Genet A. 2013;161:835-840.
14. Lo-Castro A, Brancati F, Digilio MC, et al. Neurobehavioral phenotype observed in KBG syndrome caused by ANKRD11 mutations. Am J Med Genet B Neuropsychiatr Genet. 2013;162:17-23.
15. Crippa M, Rusconi D, Castronovo C, et al. Familial intragenic duplication of ANKRD11 underlying three patients of KBG syndrome. Mol Cytogenet. 2015;8:20.
16. Kim HJ, Cho E, Park JB, Im WY, Kim HJ. A Korean family with KBG syndrome identified by ANKRD11 mutation, and phenotypic comparison of ANKRD11 mutation and 16q24.3 microdeletion. Eur J Med Genet. 2015;58:86-94.
17. Ockeloen CW, Willemsen MH, de Munnik S, et al. Further delineation of the KBG syndrome phenotype caused by ANKRD11 aberrations. Eur J Hum Genet. 2015;23:1176-1185.
18. Walz K, Cohen D, Neilsen PM, et al. Characterization of ANKRD11 mutations in humans and mice related to KBG syndrome. Hum Genet. 2015;134:181-190.
19. Goldenberg A, Riccardi F, Tessier A, et al. Clinical and molecular findings in 39 patients with KBG syndrome caused by deletion or mutation of ANKRD11. Am J Med Genet A. 2016;170:2847-2859.
20. Low K, Ashraf T, Canham N, et al. Clinical and genetic aspects of KBG syndrome. Am J Med Genet A. 2016;170:2835-2846.
21. Low KJ, Hills A, Williams M, Duff-Farrier C, McKee S, Smithson SF. A splice-site variant in ANKRD11 associated with classical KBG syndrome. Am J Med Genet A. 2017;173:2844-2846.
22. Bianchi PM, Bianchi A, Digilio MC, Tucci FM, Sitzia E, De Vincentiis GC. Audiological findings in a de novo mutation of ANKRD11 gene in KBG syndrome: report of a case and review of the literature. Int J Pediatr Otorhinolaryngol. 2017;103:109-112.
23. Miyatake S, Okamoto N, Stark Z, et al. ANKRD11 variants cause variable clinical features associated with KBG syndrome and coffin-Sirislike syndrome. J Hum Genet. 2017;62:741-746.
24. Popp B, Ekici AB, Thiel CT, et al. Exome Pool-Seq in neurodevelopmental disorders. Eur J Hum Genet. 2017;25:13641376.
25. De Bernardi ML, Ivanovski I, Caraffi SG, et al. Prominent and elongated coccyx, a new manifestation of KBG syndrome associated with novel mutation in ANKRD11. Am J Med Genet A. 2018;176:19911995.
26. Scarano E, Tassone M, Graziano C, et al. Novel mutations and unreported clinical features in KBG syndrome. Mol Syndromol. 2019; 10:130-138.
27. Gnazzo M, Lepri FR, Dentici ML, et al. KBG syndrome: common and uncommon clinical features based on 31 new patients. Am J Med Genet. 2020;182:1073-1083.
28. Spengler S, Oehl-Jaschkowitz B, Begemann M, Hennes P, Zerres K, Eggermann T. Haploinsufficiency of ANKRD11 (16q24.3) is not obligatorily associated with cognitive impairment but shows a clinical overlap with silver-Russell syndrome. Mol Syndromol. 2013;4: 246-249.
29. Ansari M, Poke G, Ferry Q, et al. Genetic heterogeneity in Cornelia de Lange syndrome (CdLS) and CdLS-like phenotypes with observed and predicted levels of mosaicism. J Med Genet. 2014;51:659-668.
30. Parenti I, Gervasini C, Pozojevic J, et al. Broadening of cohesinopathies: exome sequencing identifies mutations in ANKRD11 in two patients with Cornelia de Lange-overlapping phenotype: broadening of cohesinopathies. Clin Genet. 2016;89:74-81.
31. Aoi H, Mizuguchi T, Ceroni JR, et al. Comprehensive genetic analysis of 57 families with clinically suspected Cornelia de Lange syndrome. J Hum Genet. 2019;64:967-978.
32. Cucco F, Sarogni P, Rossato S, et al. Pathogenic variants in EP300 and ANKRD11 in patients with phenotypes overlapping Cornelia de Lange syndrome. Am J Med Genet. 2020;182:1690-1696.
33. Deciphering Developmental Disorders Study. Prevalence and architecture of de novo mutations in developmental disorders. Nature. 2017;542:433-438.
34. Hanly C, Shah H, Au PYB, Murias K. Description of neurodevelopmental phenotypes associated with 10 genetic neurodevelopmental disorders: a scoping review. Clin Genet. 2021;99: 335-346.
35. den Dunnen JT, Dalgleish R, Maglott DR, et al. HGVS recommendations for the description of sequence variants: 2016 update. Hum Mutat. 2016;37(6):564-569.
36. Gurovich Y, Hanani Y, Bar O, et al. Identifying facial phenotypes of genetic disorders using deep learning. Nat Med. 2019;25:60-64.
37. Hulo N, Bairoch A, Bulliard V, et al. The 20 years of PROSITE. Nucleic Acids Res. 2007;36:D245-D249.
38. Iossifov I, O'Roak BJ, Sanders SJ, et al. The contribution of de novo coding mutations to autism spectrum disorder. Nature. 2014;515: 216-221.
39. Reynaert N, Ockeloen CW, Sävendahl L, et al. Short stature in KBG syndrome: first responses to growth hormone treatment. Horm Res Paediatr. 2015;83:361-364.
40. Deciphering Developmental Disorders Study. Large-scale discovery of novel genetic causes of developmental disorders. Nature. 2015; 519:223-228.
41. Kosmicki JA, Samocha KE, Howrigan DP, et al. Refining the role of de novo protein truncating variants in neurodevelopmental disorders using population reference samples. Nat Genet. 2017;49(4):504-510.
42. Murray N, Burgess B, Hay R, et al. KBG syndrome: an Australian experience. Am J Med Genet. 2017;173:1866-1877.
43. Basel-Vanagaite L, Wolf L, Orin M, et al. Recognition of the Cornelia de Lange syndrome phenotype with facial dysmorphology novel analysis: recognition of the CdLS phenotype with FDNA. Clin Genet. 2016;89:557-563.
44. Latorre-Pellicer A, Ascaso Á, Trujillano L, et al. Evaluating Face2Gene as a tool to identify Cornelia de Lange syndrome by facial phenotypes. IJMS. 2020;21:1042.
45. Posey JE, Harel T, Liu P, et al. Resolution of disease phenotypes resulting from multilocus genomic variation. N Engl J Med. 2017;376:21-31.
46. Kline AD, Krantz ID, Sommer A, et al. Cornelia de Lange syndrome: clinical review, diagnostic and scoring systems, and anticipatory guidance. Am J Med Genet A. 2007;143A:1287-1296.
47. Santen GWE, Kriek M, van Attikum H. SWI/SNF complex in disorder: SWItching from malignancies to intellectual disability. Epigenetics. 2012;7:1219-1224.
48. Yuan B, Pehlivan D, Karaca E, et al. Global transcriptional disturbances underlie Cornelia de Lange syndrome and related phenotypes. J Clin Invest. 2015;125:636-651.
49. Zhang A, Yeung PL, Li C-W, et al. Identification of a novel family of Ankyrin repeats containing cofactors for p160 nuclear receptor Coactivators. J Biol Chem. 2004;279:33799-33805.
50. Mehta GD, Kumar R, Srivastava S, Ghosh SK. Cohesin: functions beyond sister chromatid cohesion. FEBS Lett. 2013;587:2299-2312.
51. Lopez-Serra L, Kelly G, Patel H, Stewart A, Uhlmann F. The Scc2-Scc4 complex acts in sister chromatid cohesion and transcriptional regulation by maintaining nucleosome-free regions. Nat Genet. 2014;46:1147-1151.
52. Yan H, Shi Z, Wu Y, et al. Targeted next generation sequencing in 112 Chinese patients with intellectual disability/developmental delay: novel mutations and candidate gene. BMC Med Genet. 2019;20:80.
53. Parenti I, Teresa-Rodrigo ME, Pozojevic J, et al. Mutations in chromatin regulators functionally link Cornelia de Lange syndrome and clinically overlapping phenotypes. Hum Genet. 2017;136:307-320.
54. Avagliano L, Parenti I, Grazioli P, et al. Chromatinopathies: a focus on Cornelia de Lange syndrome. Clin Genet. 2020;97:3-11.
55. Biesecker LG, Adam MP, Alkuraya FS, et al. A dyadic approach to the delineation of diagnostic entities in clinical genomics. Am J Hum Genet. 2021;108:8-15.

## SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of this article.

How to cite this article: Parenti I, Mallozzi MB, Hüning I, et al. ANKRD11 variants: KBG syndrome and beyond. Clinical Genetics. 2021;1-14. https://doi.org/10.1111/cge. 13977


[^0]:    This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.
    © 2021 The Authors. Clinical Genetics published by John Wiley \& Sons Ltd.

