

1 **PROGNOSTIC SIGNIFICANCE OF KI67 EVALUATED BY FLOW**
2 **CYTOMETRY IN DOGS WITH HIGH GRADE B-CELL LYMPHOMA**

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9
10 **Abstract**

11 Ki67 can discriminate between high- and low-grade canine lymphomas, but its prognostic role in
12 specific subtypes of the neoplasm is unknown. We assessed the prognostic significance of Ki67%
13 (percentage of Ki67-positive cells), evaluated via flow cytometry, in 40 dogs with high-grade B-cell
14 lymphoma, treated with a modified Wisconsin-Madison protocol (UW-25). The following variables
15 were investigated for association with lymphoma specific survival (LSS) and relapse free interval
16 (RFI): Ki67%, breed, sex, age, stage, substage, complete remission (CR). By multivariate analysis,
17 Ki67% (P=0.009) and achievement of CR (P=0.001) were independent prognostic factors for LSS.
18 Dogs with intermediate Ki67% (20.1-40%) presented longer LSS and RFI (median=866 and 428
19 days, respectively) than dogs with low (median=42 days, P<0.001; median=159 days, P=0.014) or
20 high (median=173 days, P=0.038; median=100 days, P=0.126) values. Determination of Ki67 is a
21 prognostic tool that improves the clinical usefulness of flow cytometric analysis in canine high-
22 grade B-cell lymphoma.

23

24 **Introduction**

25 Canine lymphoma represents a heterogeneous group of neoplasms arising from the malignant
26 transformation of lymphoid cells and is characterized by a broad range of clinical presentations and
27 potential outcomes.

28 Depending upon the grade of malignancy, lymphomas are cytologically grouped into two main
29 categories. The most commonly encountered forms are high grade lymphomas, clinically aggressive
30 and typically fatal within a short period of time when treatment is not instituted. Conversely low
31 grade lymphomas are rare and characterized by an indolent disease course.^{1,2}

32 Defining the immunophenotype is also reported to be important in predicting prognosis.³⁻⁷ In fact
33 multicentric T-cell forms, when compared with B-cell forms, seem to be associated with similar
34 initial response rates, but have significantly lower response durability, even following an
35 appropriate chemotherapy protocol.²

36 Moreover, although several prognostic factors that independently influence the response rate and the
37 survival time have been identified, the clinical outcome remains variable between identically treated
38 lymphomas.⁴ In fact, dogs with similar signalment, stage and substage of disease,
39 immunophenotype and tumor anatomic location may respond differently to the same treatment.⁸

40 In recent years, many studies have stressed the prognostic significance of the evaluation of tumor
41 biology and, in this context, the role of proliferative activity has received special attention. One of
42 the most frequently used methods to evaluate the growth fraction of neoplastic populations is the
43 detection of the Ki67 antigen.⁹ This proliferation-associated nuclear protein is expressed in all the
44 active phases of the cell cycle (G1, S, G2 and mitosis), but it is absent in resting cells (G0).^{10,11} The
45 exclusive expression in proliferating cells has made the antibodies raised against the Ki67 antigen
46 an invaluable diagnostic tool for grading, assessing clinical behavior and outcome in various human
47 malignancies.¹²⁻¹⁵

48 In particular, in non-Hodgkin's lymphoma, the human counterpart of canine lymphoma,¹⁶ Ki67 has
49 been found to be an independent prognostic factor.¹⁷⁻¹⁹ However, contradictory results have been
50 reported, mainly because of the heterogeneity within and among the different subtypes of the
51 disease.^{13,20,21}

52 The proliferative activity has also been evaluated in few studies on canine lymphoma and, while
53 Ki67 expression has shown a significant correlation with the grade of malignancy,^{22,23} its reliability
54 as prognostic marker is still unclear.^{24,25} In all these studies the determination of Ki67 has been
55 performed through immunohistochemistry in bioptic specimens. Moreover recently, our group has
56 demonstrated that flow cytometric detection of Ki67 is a powerful and non-invasive alternative
57 method able to discriminate between high and low grade canine lymphomas.²⁶

58 The aim of this study was to assess the prognostic significance of Ki67, evaluated by flow
59 cytometry, in dogs with high grade B-cell lymphoma being treated with the same multidrug
60 chemotherapy protocol.

61

62 **Materials and methods**

63 **Case selection**

64 Dogs with multicentric high grade B-cell lymphoma diagnosed at the Veterinary Teaching Hospital
65 of the University of Turin between April 2011 and September 2014 were considered. The diagnosis
66 was based on clinical presentation (lymph node enlargement), cytological examination of lymph
67 nodes and flow cytometric analysis.

68 Inclusion criteria for the study were cytological diagnosis of high grade lymphoma according to the
69 updated Kiel classification,^{27,28} presence of flow cytometric B-cell immunophenotype, flow
70 cytometric Ki67 determination, treatment with a modified version of the University of Wisconsin-
71 Madison chemotherapy protocol (UW-25)²⁹ and the availability of follow up data. Dogs previously

72 treated with corticosteroid or chemotherapy agents were excluded. For each included dog,
73 signalment data (breed, sex and age), when available, were retrieved and clinical stage (I-V) and
74 substage (a or b) were assigned according to the World Health Organization (WHO) system.³⁰ In
75 particular, stage V was assigned when the neoplastic population, detected via flow cytometry, was
76 $\geq 3\%$ in peripheral blood and/or bone marrow.

77

78 **Flow cytometric immunophenotyping and Ki67 determination**

79 At time of initial staging, flow cytometric immunophenotyping was performed on lymph node fine-
80 needle aspirate biopsies (FNABs), peripheral blood samples and/or bone marrow aspirates within
81 24h from collection as previously reported.³¹

82 The following panel of monoclonal antibodies (mAbs) was used: CD45-Alexa647 (pan-leukocyte
83 marker; clone YKIX716.1, AbD Serotec, Oxford, UK), CD3-FITC (T-cells marker; clone
84 CA17.2A12, AbD Serotec), CD5-FITC (T-cells marker; clone YKIX322.3, AbD Serotec), CD4-
85 Alexa647 (T-helper marker; clone YKIX302.9, AbD Serotec), CD8-PE (T-cytotoxic/suppressor
86 maker; clone YCATE55.9, AbD Serotec), CD21-PE (B-cells marker; clone CA21D6, AbD Serotec),
87 CD79b-FITC (B-cells marker; clone AT107-2, AbD Serotec) and CD34-PE (precursor cells marker;
88 clone 1H6, Pharmingen Becton Dickinson, San Jose, CA, USA).

89 The proliferative activity was determined on the same lymph node FNABs used for
90 immunophenotyping. Cells were labelled with antiKi67-FITC monoclonal antibody (clone MIB-1,
91 DAKO, Glostrup, Copenhagen, Denmark) using a fixation and permeabilization method with
92 methanol, as described previously.²⁶

93 A minimum of 10.000 events were acquired both for immunophenotype and Ki67 determination on
94 BD Accuri C6 flow cytometer (Becton Dickinson). Data were analyzed using CFlow Plus software
95 (Beckton Dickinson). A gate of analysis was depicted on forward (FSC) versus side scatter (SSC)

96 plot in order to exclude debris and background. The proliferative activity was expressed as the
97 percentage of Ki67 positive cells (Ki67%) calculated on a SSC versus fluorescence intensity plot.

98

99 **Cytological evaluation**

100 Smears obtained by FNABs of enlarged lymph nodes were air-dried, fixed and stained with May-
101 Grünwald-Giemsa. Each case was classified according to the updated Kiel classification^{27,28} and
102 allocated to a specific grade of malignancy and cytological subtype.

103

104 **Follow up**

105 Information pertaining to the achievement of remission, occurrence of relapse, survival at the end of
106 chemotherapy protocol, lymphoma specific survival (LSS), relapse-free interval (RFI), date and
107 cause of death was collected.

108 Responses were classified as follows: complete remission (CR), which indicated reduction to
109 normal size of all measurable lymph nodes; partial remission (PR), which indicated more than 50%
110 but less than 100% reduction of all measurable lesions and stable disease (SD), which indicated less
111 than 50% reduction or no change in the size of all measurable lesions. Relapse was defined as
112 clinical reappearance and cytological evidence of lymphoma in any anatomical site in dogs having
113 experienced CR. RFI was defined as the time in days from when a dog achieved CR until relapse.
114 LSS was defined as the interval in days between the date on which chemotherapy was started and
115 the date of death due to lymphoma related causes.

116

117 **Statistical analysis**

118 LSS and RFI for all dogs were estimated using the Kaplan-Meier product limit method.
119 Contingency tables were prepared for each of following variables: Ki67% (low, intermediate, high),

120 breed (purebred or crossbred), sex (male or female), age ($<$ or \geq 10 years), stage (I-IV or V),
121 substage (a or b), CR (yes or no). Pearson's χ^2 with z-test for column proportion comparisons and
122 Bonferroni adjustment for multiple comparisons were calculated to test the association between
123 each variable with the achievement of CR and survival at the end of first chemotherapy protocol
124 (UW-25). Dogs that died for causes other than lymphoma and dogs that had not yet completed the
125 protocol and did not meet the event (CR or death) were excluded from contingency tables. Ki67%
126 cut-off values were defined rounding the thresholds of 25th and 75th percentiles to 20 and 40%,
127 respectively, and thus generating the following groups: low if $Ki67 \leq 20\%$, intermediate if Ki67
128 between 20.1% and 40%, high if $Ki67 > 40\%$.

129 To evaluate the prognostic significance of each variable, univariate logistic regression for LSS and
130 RFI was first used and variables with a P value < 0.3 were then included in a multivariate Cox
131 proportional hazards model progression analysis with a backward step selection. Kaplan-Meier
132 curves were drawn for Ki67% groups and compared by log-rank test to assess the survival analysis.
133 Dogs that were alive at the end of the study, lost to follow-up or dead due to causes other than
134 lymphoma were censored for survival analysis. Differences were considered significant with $P <$
135 0.05. Statistical analyses were performed using SPSS software (IBM SPSS Statistics, IBM
136 Corporation, Chicago, IL, USA).

137

138 **Results**

139 **Lymphoma cases**

140 Forty cases met inclusion criteria and were enrolled in the study.

141 Data about the identification of breed were reported for 39 cases. There were 2 (66.7%) purebred
142 dogs (3 Labrador retrievers, 2 German shepherds, 2 Dobermans, 2 Bloodhounds, 2 Pitt bull terriers
143 and 1 each of Italian Mastiff, Great Dane, Poodle, Dachshund, Beagle, Bernese mountain dog,

144 Cavalier King Charles Spaniel, Golden retriever, Jack Russell, Rottweiler, White Swiss shepherd,
145 Cocker Spaniel, English bulldog, Lagotto romagnolo, American Staffordshire terrier) and 13
146 (33.3%) crossbred dogs. Sixteen dogs (42.1%) were males (1 castrated) and 22 (57.9%) were
147 females (9 spayed), while in 2 cases the sex was unknown. The age was only reported for 37 dogs
148 and the median age was 9 years (range, 4-15 years).

149 The included lymphomas were cytologically classified as follows: 8 (20%) centroblastic
150 monomorphic, 24 (60%) centroblastic polymorphic predominantly large cell, 5 (12.5%)
151 immunoblastic, 2 (5%) lymphoblastic and 1 (2.5%) plasmacytoid.

152 At time of diagnosis 27 dogs (67.5%) were in stage IV (10 substage a, 16 substage b and 1
153 unknown) and 13 (32.5%) in stage V (all substage b).

154

155 **Response to treatment**

156 CR was achieved in 25 (62.5%) dogs. Twelve out of these 25 (48%) relapsed (median RFI=180
157 days; range 28-530), 10 (40%) were still in CR at the end of the study (median follow up
158 period=321 days; range 60-1005) and 3 (12%) died of causes unrelated to lymphoma after 34, 210
159 and 240 days from the beginning of chemotherapy, with lymphoma remaining in CR.

160 Relapses were treated with a second UW-25 or with other rescue protocols (DMAC; L-
161 asparaginase+lomustine), depending on when the relapse occurred and owner compliance. At the
162 end of the study, 8 out of 12 relapsed dogs (66.7%) were dead because of progressive disease
163 (median LSS = 390 days; range 150-866), 3 (25%) were in PR (follow up period of 515, 800 and
164 1108 days) and 1 (8.3%) was in SD (follow up period = 295 days).

165 Among 15 dogs that did not achieve CR, 11 (73.3%) died because of PD (median LSS=42 days;
166 range 15-1100), 3 (20%) were in PR at the end of the study (follow up period of 28, 157 and 653
167 days) and 1 (6.7%) died of causes unrelated to lymphoma after 45 days. Estimated median RFI and

168 LSS for all dogs were 414 days (95% CI range 228-600 days) and 442 days (95% CI range 236-648
169 days), respectively.

170

171 **Proliferative activity**

172 The mean Ki67% was 33.8% (SD=14.2%) and the median was 30.7% (range 10-67%). Six cases
173 presented low Ki67% ($\leq 20\%$), 24 were in the intermediate group (20.1-40%) and 10 were in the
174 high group ($>40\%$).

175

176 **Survival at the end of chemotherapy protocol and achievement of CR**

177 Survival at the end of chemotherapy was significantly associated with the achievement of CR
178 ($P=0.001$). In fact, 91.7% of the dogs that achieved CR were alived compared with 33.3% of dogs
179 that did not reach CR (Table 1).

180 Ki67% showed near-significant association with both survival ($P=0.063$) and achievement of CR
181 ($P=0.075$) at the end of chemotherapy protocol. In fact, percentages of both survival and CR were
182 higher for dogs with intermediate Ki67% (85.7% and 81%, respectively) compared with dogs with
183 low (50% and 33%) and high Ki67% (50% and 60%) (Table 1).

184

185 **Prognostic factors for LSS and RFI**

186 Ki67% ($P=0.007$) and achievement of CR ($P=0.001$) significantly influenced LSS on univariate
187 analysis and were confirmed to be independent prognostic factors for LSS ($P=0.009$ and $P=0.001$
188 respectively) in the multivariate analysis (Table 2).

189 None of the variables significantly influenced RFI in the univariate analysis and none were of
190 prognostic significance for RFI in the multivariate analysis (Table 2).

191 The Kaplan-Meier analysis showed that dogs with intermediate Ki67% have significantly longer
192 LSS (median=866 days) than dogs with low (median=42 days; $P<0.001$) and high Ki67%
193 (median=173 days; $P=0.038$) (Fig. 1).

194 Intermediate Ki67% wa a significant predictor also for 1 year and 2 years survival ($P=0.001$ and
195 $P=0.004$ versus low and high Ki67%, respectively, at both time points) (Fig. 1). Dogs with
196 intermediate Ki67% reported also longer RFI (median =428 days) than dogs with low (median=159
197 days, $P=0.014$) and high Ki67% (median=100 days, $P=0.126$), although the difference with the high
198 Ki67% group did not reach statistical significance (Fig. 2).

199

200 **DISCUSSION**

201 Ki67 is one of the most widely used markers of cell proliferation. Although it is considered an
202 important factor for grading neoplasms and predicting their biological behavior,^{12,14} its clinical
203 relevance is still being debated both in human and canine lymphomas. In a previous work,²⁶ we
204 assessed the feasibility of flow cytometric determination of Ki67 in canine lymphoma and we
205 demonstrated its association with malignancy grade, regardless of phenotype and morphology.

206 In this study we investigated the prognostic significance of Ki67, as evaluated by flow cytometry in
207 dogs with high grade B cell lymphoma treated with the UW-25 chemotherapy protocol.

208 We focused on the most common type of canine lymphoma to limit heterogeneity with regards to
209 some clinical prognostic features, such as malignancy grade and immunophenotype.⁴ Likewise in
210 our case series, all dogs were treated with the same chemotherapeutic protocol to avoid treatment
211 bias in the response, although the LSS of relapsed dogs may have been influenced by receiving
212 multiple reinduction or rescue protocols.

213 The achievement of CR and the intermediate Ki67% values were associated with the survival at the
214 end of chemotherapy protocol, suggesting their prognostic role, even though the association with the
215 Ki67 did not reach a statistical significance.

216 Based on multivariate analysis, Ki67% and CR were found to be independent prognostic factors for
217 LSS, while none of the investigated variables had prognostic significance for RFI. Moreover, the
218 Kaplan-Meier analysis confirmed that an intermediate Ki67% was associated with a better prognosis
219 with longer LSS and RFI compared with dogs with low or high Ki67%.

220 These findings are discordant with the results of few previous studies that evaluated the prognostic
221 significance of Ki67 in dogs with lymphoma. In the work by Kiupel et al.²⁵ Ki67 expression showed
222 no prognostic value, while Phillips et al.²⁴ reported that Ki67 was predictive for duration of first RFI
223 but not overall survival. Differences in inclusion criteria and method of Ki67 determination could
224 account for these discrepancies. In fact, in the previous studies,^{24,25} both high and low grade
225 lymphomas and both B and T-cell immunophenotype were included. Moreover, Ki67 detection was
226 carried out through immunohistochemistry on biptic specimens, while we used flow cytometric
227 analysis of FNABs. Furthermore the different selection of the cutoff values to define groups may
228 have influenced the results. In our work, we used an approach similar to that of Phillips et al.²⁴,
229 using median and the 75th percentile to differentiate two prognostic groups and we get near
230 significant results with the latter (data not shown). Observing that the longest survival times were
231 associated with intermediate Ki67% values, we assessed the prognostic significance of Ki67%
232 dividing cases in three different groups using quartiles. Furthermore, we rounded the quartile cutoff
233 values to 20% and 40% in order to simplify the use in clinical practice. Unfortunately, a direct
234 comparison of our cut-off values with those assessed by Phillips et al is not possible because they
235 did not report the actual percentages that define quartiles in their caseload. However, this
236 comparison, although interesting, would presumably be limited by the different method use to

237 measure Ki67 expression. In this regard, Kiupel et al.²⁵ did not get significant results despite the
238 application of thresholds similar to ours (Ki67<20%; 21-40%; 41-60%; >60%).

239 Contradictory results on prognostic significance of Ki67 have also been reported in human non-
240 Hodgkin's lymphoma, because of the heterogeneity of the different subtypes of this disease.¹⁹ Many
241 studies have determined Ki67 in aggressive diffuse large B-cell lymphomas, with a wide range of
242 expression.²¹ In accordance with our results, in the largest one Jerkeman et al.³² found that patients
243 with either low (<60%) or high Ki67 (>90%) expression demonstrated a trend toward overall lower
244 survival than patients with moderate expression (60-90%). Moreover, a low proliferation index was
245 associated with a low failure-free survival compared with moderate or high indexes. This behavior
246 is likely because lymphomas with a low proliferation rate exhibit resistance to cycle specific
247 cytotoxic chemotherapy, given that the majority of cells are resting in the G0 phase of the cell cycle.
248 Conversely, in cases with high proliferation rates, treatment failure may be caused through regrowth
249 or by the increasing likelihood of further mutations.

250 In addition to the proliferative activity, we also found that achievement of CR was an independent
251 prognostic factor for LSS, as reported in previous studies, where obtaining CR led to prolonged
252 survival for dogs with aggressive lymphoma.^{7,33,34} Stage and substage did not shown prognostic
253 significance for LSS or RFI, in contrast with some authors^{8,35,36} but, in accordance with others.^{37,38}
254 These differences may be because of the inclusion of different types of lymphoma, different
255 therapeutic strategies and to the different methods and cut-offs used to stage the disease.

256 Major limits of this work are its retrospective nature and the limited number of cases. Prospective
257 studies considering a large number of lymphomas are required to confirm the clinical usefulness of
258 a Ki67-based stratification of patients.

259 In conclusion, flow cytometric determination of Ki67 was found to be an independent predictor for
260 LSS in treated high grade B cell lymphomas; intermediate values were associated with the best

261 prognosis. We previously demonstrated that this determination is useful in discriminating between
262 low and high grade lymphomas. Thus, we suggest the introduction of Ki67 in the routine panel of
263 labeling in order to add diagnostic and prognostic value to the flow cytometric analysis.

264

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269 **REFERENCES**

- 270 1. Teske E. Canine malignant lymphoma: a review and comparison with human non-
271 Hondgkin's lymphomas. *Vet Quat* 1994;16:209–219.
- 272 2. Vail DM, Pinkerton ME, Young KM. Canine lymphoma and lymphoid leukemia. In
273 Withrow and MacEwen's Small Animal Clinical Oncology. (5th edn), Withrow SJ, Vail DM
274 (eds.). WB Saunders: St. Louis, 2013;608–638.
- 275 3. *Greenlee PG, Filippa DA, Quimby FW, Patnaik AK, Calvano SE, Matus RE et al.*
276 *Lymphomas in dogs. A morphologic, immunologic and clinical study. Cancer* 1990;66:480–
277 490.
- 278 4. Teske E, van Heerde P, Rutteman GR, Kurzman ID, Moore PF and MacEwen EG.
279 Prognostic factors of treatment of malignant lymphoma in dogs. *Am J Vet Med Assoc*
280 1994;205:1722–1728.
- 281 5. Ruslander DA, Gebhard DH, Tompkins MB, Grindem CB and Page RL. Immunophenotypic
282 characterization of canine lymphoproliferative disorders. *In Vivo* 1997;112:169–172.

- 283 6. Dobson JM, Blackwood LB, McInnes EF, Bostock DE, Nicholls P, Hoather TM, Tom BD.
284 Prognostic variables in canine multicentric lymphosarcoma. *J Small Anim Pract*
285 2001;42:377–384.
- 286 7. Simon D, Moreno SN, Hirschberger J, Moritz A, Kohn B, Neumann S et al. Efficacy of a
287 continuous, multiagent chemotherapeutic protocol versus a short-term single-agent protocol
288 in dogs with lymphoma. *Am J Vet Med Assoc* 2008; 232:879–885.
- 289 8. Marconato L, Stefanello D, Valenti P, Bonfanti U, Comazzi S, Roccabianca P et al.
290 Predictors of long-term survival in dogs with high-grade multicentric lymphoma. *Am J Vet*
291 *Med Assoc* 2011; 238:480–485.
- 292 9. Schlüter C, Duchrow M, Wohlenberg C, Becker MH, Key G, Flad HD, Gerdes J. The cell
293 proliferation-associated antigen of antibody Ki-67: a very large, ubiquitous nuclear protein
294 with numerous repeated elements, representing a new kind of cell cycle-maintaining
295 proteins. *J Cell Biol* 1993; 123:513–522.
- 296 10. Gerdes J, Lemke H, Baisch H, Wacker HH, Schwab U and Stein H. Cell cycle analysis of a
297 cell proliferation associated human nuclear antigen defined by the monoclonal antibody Ki-
298 67. *J Immunol* 1984;133: 1710–1715.
- 299 11. Bruno S, Darzynkiewicz Z. Cell cycle dependent expression and stability of the nuclear
300 protein detected by Ki-67 antibody in HL-60 cells. *Cell Prolif.* 1992; 25:31–40.
- 301 12. Brown DC, Gatter KC. Monoclonal antibody Ki-67: its use in histopathology.
302 *Histopathology* 1990; 17:489–503.
- 303 13. Brown DC, Gatter KC. Ki67 protein: the immaculate deception? *Histopathology* 2000;
304 40:2–11.
- 305 14. Scholzen T, Gerdes J. The Ki-67 protein: From the known and the unknown. *J Cell Physiol*
306 2000;182:311–322.

- 307 15. Endl E, Gerdes J. The Ki-67 Protein: Fascinating Forms and an Unknown Function. *Exp*
308 *Cell Res* 2000; 257:231–237.
- 309 16. MacEwen EG. Spontaneous tumors in dogs and cats: Models for the study of cancer biology
310 and treatment. *Cancer Metastasis Rev* 1990; 9:125–36.
- 311 17. Grogan TM, Lippman SM, Spier CM, Slymen DJ, Rybski JA, Rangel CS et al. Independent
312 prognostic significance of a nuclear proliferation antigen in diffuse large cell lymphomas as
313 determined by the monoclonal antibody Ki-67. *Blood* 1988; 71: 1157-1160.
- 314 18. Miller TP, Grogan TM, Dahlberg S, Spier CM, Braziel RM, Banks PM et al. Prognostic
315 Significance of the Ki-67 Associated Proliferative Antigen in Aggressive Non-Hodgkin's
316 Lymphomas: A Prospective Southwest Oncology Group Trial. *Blood* 1994; 83:1460–1466.
- 317 19. Broyde A, Boycov O, Strenov Y, Okon E, Shpilberg O and Bairey O. Role and prognostic
318 significance of the Ki-67 index in non-Hodgkin's lymphoma. *Am J Hematol* 2009; 84:338–
319 343.
- 320 20. Palestro G, Pich A, Chiusa L, Geuna M, Ponti R, Kerim S et al. Biological Heterogeneity of
321 Diffuse Mixed Small and Large Cell Non-Hodgkin's Lymphomas Assessed by DNA Flow
322 Cytometry and Ki67. *Leukemia and Lymphoma* 1995; 19:461–472.
- 323 21. Bairey O. Role and Prognostic Significance of the Ki-67 Index in Non-Hodgkin's
324 Lymphoma. *EJCMO* 2010;2:113–9.
- 325 22. Fournel-Fleury C, Magnol JP, Chabanne L, Ghernati I, Marchal T, Bonnefond C et al.
326 Growth fractions in canine non-Hodgkin's lymphomas as determined in situ by the
327 expression of the Ki-67 antigen. *J Comp Pathol* 1997;117:61–72.
- 328 23. Patruno R, Zizzo N, Zito AF, Catalano V, Valerio P, Pellecchia V et al. Microvascular
329 density and endothelial area correlate with Ki-67 proliferative rate in the canine non-
330 Hodgkin's lymphoma spontaneous model. *Leukemia & Lymphoma* 2006;47:1138–1143.

- 331 24. Phillips BS, Kass PH, Naydan DK, Winthrop MD, Griffey SM and Madewell BR. Apoptotic
332 and proliferation indexes in canine lymphoma. *J Vet Diagn Invest* 2000;12:111–117.
- 333 25. Kiupel M, Teske E, Bostock D. Prognostic Factors for Treated Canine Malignant
334 Lymphoma. *Vet Pathol* 1999;36:292–300.
- 335 26. Poggi A, Miniscalco B, Morello E, Comazzi S, Gelain ME, Aresu L, Riondato F. Flow
336 cytometric evaluation of ki67 for the determination of malignancy grade in canine
337 lymphoma. *Vet Comp Onc* 2013; doi:10.1111/vco.12078.
- 338 27. Fournel-Fleury C, Magnol JP, Bricaire P, Marchal T, Chabanne L, Delverdier A et al.
339 Cytohistological and immunological classification of canine malignant lymphomas:
340 Comparison with human non-Hodgkin's lymphomas. *J Comp Pathol* 1997;117:35–59.
- 341 28. Ponce F, Marchal T, Magnol JP, Turinelli V, Ledieu D, Bonnefont C et al. A morphological
342 study of 608 cases of canine malignant lymphoma in France with a focus on comparative
343 similarities between canine and human lymphoma morphology. *Vet Pathol* 2010;47:414–
344 433.
- 345 29. Garrett L.D, Thamm DH, Chun R, Dudley R and Vail DM. Evaluation of a 6 month
346 chemotherapy protocol with no maintenance therapy for dogs with lymphoma. *J Vet Intern*
347 *Med* 2002;16,704–709.
- 348 30. Owen L. TNM Classification of Tumors in Domestic Animals. World Health Organization,
349 Geneva, Switzerland 1980;46–47.
- 350 31. Gelain ME, Mazzilli M, Riondato F, Marconato L and Comazzi S. Aberrant phenotypes and
351 quantitative antigen expression in different subtypes of canine lymphoma by flow cytometry.
352 *Vet Immunol Immunopathol* 2008;12(3–4):179–188.
- 353 32. Jerkeman M, Anderson H, Dictor M, Kvaloy S, Akerman M and Cavallin-Stahl E.
354 Assessment of biological prognostic factors provides clinically relevant information in

- 355 patients with diffuse large B-cell lymphoma. A Nordic Lymphoma Group study. *Ann*
356 *Hematol* 2004;83:414–419.
- 357 33. Ettinger SN. Principles of Treatment for Canine Lymphoma. *Clin Tech Small Anim Pract.*
358 2003; 18:92–97.
- 359 34. Lurie DM, Gordon IK, Thèon AP, Rodriguez CO, Suter SE and Kent MS. Sequential low-
360 dose rate half-body irradiation and chemotherapy for the treatment of canine multicentric
361 lymphoma. *J Vet Intern Med* 2009; 23:1064–1070.
- 362 35. Jagielski D, Lechowski R, Hoffmann-Jagielska M, and Winiarczyk S. A Retrospective Study
363 of the Incidence and Prognostic Factors of Multicentric Lymphoma in Dogs (1998–2000). *J*
364 *Vet Med* 2002;49:419–424.
- 365 36. Lana SE, Jackson TL, Burnett RC, Morley PS and Avery AC. Utility of polymerase chain
366 reaction for analysis of antigen receptor rearrangement in staging and predicting prognosis in
367 dogs with lymphoma. *J Vet Inter Med* 2006;20:329–334.
- 368 37. Morrison-Collister KE, Rassnick KM, Northrup NC, Kristal O, Chretin JD, Williams Le et
369 al. A combination chemotherapy protocol with MOPP and CCNU consolidation (Tufts
370 VELCAP-SC) for the treatment of canine lymphoma. *Vet Comp Oncol* 2003;1:180–190.
- 371 38. Flory AB, Rassnick KM, Stokol T, Scrivani PV and Erb HN. Stage migration in dogs with
372 lymphoma. *J Vet Intern Med* 2007;21,1041–1047.

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374