



Entomopathogenic nematodes and fungi to control *Hyalesthes obsoletus* (Hemiptera: Auchenorrhyncha: Cixiidae)

Abdelhameed Moussa · Michael Maixner · Dietrich Stephan · Giacomo Santoiemma · Alessandro Passera · Nicola Mori · Fabio Quaglino

Received: 20 August 2020 / Accepted: 28 December 2020
© The Author(s) 2021

Abstract *Hyalesthes obsoletus* Signoret (Hemiptera: Auchenorrhyncha: Cixiidae) is a univoltine, polyphagous planthopper that completes its life cycle, including the subterranean nymph cryptic stage, on herbaceous weeds. In vineyards, it can transmit ‘*Candidatus Phytoplasma solani*’, an obligate parasitic bacterium associated with bois noir (BN) disease of grapevine, from its host plants to grapevine when occasionally feeding on the latter. The main disease management strategies are based on vector(s) control.

Handling Editor: Ralf Ehlers.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10526-020-10076-1>.

A. Moussa · A. Passera · F. Quaglino
Department of Agricultural and Environmental Sciences - Production, Landscape, Agroenergy (DiSAA), University of Milan, Milan, Italy

A. Moussa
Pests and Plant Protection Department, Agricultural and Biological Research Division, National Research Centre, Dokki, Giza, Egypt

M. Maixner
Julius Kühn-Institute (JKI), Institute for Plant Protection in Fruit Crops and Viticulture, Siebeldingen, Germany

D. Stephan
Julius Kühn-Institute (JKI), Institute for Biological Control, Darmstadt, Germany

Insecticide treatments on grapevine canopy are completely inefficient on *H. obsoletus*, due to its life cycle. Consequently, control of this planthopper focuses on the nymphs living on the roots of their host plants. Such practices, based on herbicide application and/or weed management, can reduce vector density in the vineyard but can impact the environment or may not be applicable, highlighting the necessity for alternative strategies. In this study, the efficacy of entomopathogenic nematodes (EPNs; *Steinernema carpocapsae*, *S. feltiae*, *Heterorhabditis bacteriophora*) and fungi (EPFs; *Beauveria bassiana*, *Metarhizium anisopliae*, *Isaria fumosorosea*, *Lecanicillium muscarium*) against *H. obsoletus* nymphs (EPNs) and adults (EPNs and EPFs) was

G. Santoiemma
Department of Agronomy, Food, Natural resources, Animals and Environment (DAFNAE), University of Padova, Padova, Italy

N. Mori (✉)
Department of Biotechnology, University of Verona, Verona, Italy
e-mail: nicola.mori@univr.it

assessed under laboratory and greenhouse conditions. The majority of examined EPNs and EPFs were able to kill *H. obsoletus* exhibiting a range of effectiveness. *S. carpocapsae* (among EPNs) and *I. fumosorosea* (among EPFs) were found to be the most effective biocontrol agents in all trials carried out. Advantages and limitations of such promising biocontrol agents were discussed. Ecological competency and conditions that can impede or enhance the EPNs and EPFs performance should be investigated to optimize their performance under field conditions.

Keywords Planthoppers · Grapevine · Bois noir · Sustainability · Entomopathogens

Introduction

Hyalesthes obsoletus Signoret (Hemiptera: Auchenorrhyncha: Cixiidae) is a polyphagous planthopper able to transmit ‘*Candidatus* Phytoplasma solani’ (CaPsol) to plants (Quaglino et al. 2013), including *Vitis vinifera* L. (Maixner 1994). CaPsol, a plant obligate parasitic bacterium, is associated with bois noir (BN), a disease belonging to the complex of grapevine yellows which had high economic impact on viticulture in Europe in the last decades (Angelini et al. 2018). Although alternative insect vectors of CaPsol to grapevine were recently reported (Cvrković et al. 2014; Quaglino et al. 2019), BN epidemiology is principally determined by the life cycle of its main vector *H. obsoletus* (Mori et al. 2013).

H. obsoletus is a palaeartic, univoltine species that, in Europe, completes its life cycle mainly on bindweed (*Convolvulus arvensis* L.) and nettle (*Urtica dioica* L.) (Langer and Maixner 2004) but also on other host plants (Kosovac et al. 2019; Moussa et al. 2019). In summertime, females produce eggs on the root collar of host plants and after egg hatching the nymphs migrate into the soil to the roots from which they can acquire CaPsol. After a latency period, *H. obsoletus* becomes able to transmit CaPsol to plants for the duration of its life. Overwintering occurs as second-third instar nymphs in the soil. Fourth and fifth instar nymphs migrate to the soil surface in early spring. Adults emerge from end of May till end of June and they fly from beginning of July to end of August, based on climate, host plant, and region (Cargnus et al.

2012; Maixner and Johannesen 2014; Alma et al. 2015). During their flights, *H. obsoletus* adults can occasionally feed on grapevine and, if infected, transmit CaPsol. However, due to their limited feeding activity on grapevine and the short lifespan of the adult stage, they cannot transmit CaPsol from vine to vine. Grapevine is therefore a dead-end host for the pathogen (Bressan et al. 2007).

Since no effective control measures directly targeting phytoplasmas are available, the main strategies to manage the spreading of phytoplasma-associated diseases are based on preventive measures, including the control of vectors before their emergence from the ground (Bianco et al. 2019). Due to its cryptic life cycle and polyphagous feeding habit, insecticide treatments on grapevine canopy are completely inefficient against *H. obsoletus*. Thus, strategies for its control focus on depriving the nymphs of their feeding substrate, the host plant roots. Before adult emergence, bindweed and nettle can be suppressed by planting of ground covering rosette plants, repeated mowing or weeding (Maixner and Mori 2013; Mori et al. 2014a). Since *H. obsoletus* presence depends on the distribution of its natural plant hosts both within and outside the vineyards, such strategies are limited by restrictions on the use of herbicides in uncultivated areas, as well as mechanical weeding on ditches and embankments because of soil landslide. In Israel, *H. obsoletus* populations within vineyards are successfully limited by a push and pull strategy using chaste tree (*Vitex agnus-castus* L.) (Sharon et al. 2015), but such a strategy cannot be employed in Europe where this plant hosts both *H. obsoletus* and CaPsol (Moussa et al. 2019).

Considering these limitations, a promising approach to control the vector populations could be based on the utilization of biocontrol agents such as entomopathogenic nematodes (EPNs) and fungi (EPFs). In particular, several *Steinernema* and *Heterorhabditis* EPNs have been reported as effective biocontrol agents against a broad range of insects with a cryptic life cycle like *H. obsoletus* (Grewal et al. 2005; Lacey and Georgis 2012; Guerrero and Pardey 2019). EPNs efficacy depends on their survival for a long time without their host targets in the soil, and their ability to find the hosts by ambush (i.e., *Steinernema carpocapsae*) or cruising (i.e., *Heterorhabditis bacteriophora*) strategy (Kaya et al. 1993; Grewal et al. 1994). Concerning EPFs, they are

reported as important antagonists of soil-dwelling insect pests adapted to live in agricultural soils, such as the grapevine phylloxera in vineyards (Kirchmair et al. 2004). Interestingly, the EPF *Metarhizium anisopliae* showed a great efficacy against *H. obsoletus* adults under laboratory conditions (Langer et al. 2005). In this study, the efficacy of different EPNs and EPFs against *H. obsoletus* nymphs and adults under laboratory and greenhouse conditions were assessed to develop effective and innovative approaches to control the main vector of CaPsol.

Materials and methods

Hyalesthes obsoletus collection

Nymphs of *H. obsoletus* were collected on two dates immediately before the different experiments. The collection for laboratory bioassay was done in the middle of May 2019. Nymphs were obtained from pots of stinging nettle plants at the rearing facility in Julius Kühn-Institute (JKI) in Siebeldingen, Germany. Collection for the greenhouse trials was performed in late May 2019 from roots of nettle plants growing at the borders of a highly BN-affected vineyard in Mosel area (49.9198°N, 7.0627°E), Germany. Nymphs were placed in falcon tubes filled with the same soil in which they were collected and sent to the laboratory for trials. Nymphs identity was confirmed based on the taxonomic key by Stöckmann et al. (2013).

Adults of *H. obsoletus* were collected from bindweed and stinging nettle plants using a sweep net and mouth aspirator from mid-June till the end of July 2019, based on the trial requirements, in the vicinity of highly BN-affected vineyards in Mosel area (49.9198°N, 7.0627°E, and 49.1928°N, 8.0830°E), Germany. Collected adults were placed in collapsible insect mesh cages with shoots of nettle plants as food source, transferred to the laboratory, and identified by taxonomic key (Bertin et al. 2010). *H. obsoletus* adults were subject to immediate use in bioassays and greenhouse efficacy trials.

Entomopathogenic nematodes

Three EPNs were applied in both bioassays and greenhouse trials against *H. obsoletus* nymphs and adults. In detail, the utilized EPNs (*Steinernema*

carpocapsae, *S. feltiae*, *Heterorhabditis bacteriophora*, and a combination (1:1) of *S. feltiae* and *H. bacteriophora*) were purchased from E-nema® Company (Schwentental, Germany) (Table 1) and maintained at 4 °C. Immediately before using, each EPN (supplied in powder) was suspended in tap water and tested for its viability by counting the infective juveniles (IJs) under stereomicroscope. IJs without a response to stimulators were considered dead (Lacey 1997). All EPNs showed a viability higher than 95%. Before the application, each EPN was serially diluted in tap water to reach the required concentration for bioassay (200 IJs ml⁻¹) and greenhouse trials (400 IJs ml⁻¹) (Guerrero and Pardey 2019).

Entomopathogenic fungi

Three distinct EPF isolates of *Beauveria bassiana*, *Metarhizium anisopliae*, and *Isaria fumosorosea* (provided by Dr. Dietrich Stephan - Institute for Biological Control, JKI, Darmstadt, Germany), were applied in preliminary screening against *H. obsoletus* adults. In addition to these isolates, one commercial isolate of *M. anisopliae* and one of *Lecanicillium muscarium* were purchased from Koppert Biological Systems Company (Verona, Italy) and applied in greenhouse trials against *H. obsoletus* adults (Table 1). EPFs were cultivated in Petri dishes on malt extract peptone agar (30 g of malt extract, 3 g of peptone, 15 g of agar in 1 l of distilled water) previously autoclaved at 121 °C for 10 min. After inoculation, EPFs were grown for five days at room temperature (22 °C) and for six days at 4 °C. For each EPF isolate, fungal conidia were recovered from the mycelium, suspended in distilled water and counted (three times per isolate) at 400× magnification through a haemocytometer. For each EPF isolate, stock solutions of conidial suspension were prepared at the concentrations of 10⁶, 10⁸ and 10¹⁰ conidia ml⁻¹ and stored at 4 °C until use.

Entomopathogenic nematodes: laboratory bioassays

Bioassay to evaluate the efficacy of EPNs against *H. obsoletus* nymphs was conducted in 12-well cell culture plates with lids. According to the protocols of Kaya and Stock (1997) and Glazer and Lewis (2000), in each well, filled with 1 g of autoclaved sand, 1 ml of

Table 1 Entomopathogenic nematodes (EPNs) and fungi (EPFs) used against *Hyalesthes obsoletus* adults and nymphs

Agent	Scientific name	Strain acronym	Source	<i>H. obsoletus</i> target stage
EPNs	<i>Steinernema carpocapsae</i>	Sc	E-nema®	Nymphs and adults
	<i>Heterorhabditis bacteriophora</i>	Hb		
	<i>Steinernema feltiae</i>	Sf		
	<i>H. bacteriophora</i> & <i>S. feltiae</i>	Hb + Sf		
EPFs	<i>Beauveria bassiana</i>	JKI-BI-1124	Julius Kühn-Institut	Adults
		JKI-BI-1125		
		JKI-BI-1126		
	<i>Metarhizium anisopliae</i>	JKI-BI-1428		
		JKI-BI-1429		
		JKI-BI-1430		
	<i>Isaria fumosorosea</i>	JKI-BI-1497		
		JKI-BI-1499		
		JKI-BI-1500		
	<i>Metarhizium anisopliae</i>	1111	Koppert biological systems company	
	<i>Lecanicillium muscarium</i>	2222	Company	

EPN suspension (200 IJs ml⁻¹) was applied and one nymph was placed. Control plates were treated with distilled water before placing the nymphs. Bioassay to evaluate the efficacy of EPNs against *H. obsoletus* adults was conducted in plastic Petri dishes (9 cm diameter) padded with filter paper. Following Glazer and Lewis (2000) procedures, in each dish, fresh shoots of stinging nettle (5 cm) were placed as a food source together with 12 adults of *H. obsoletus*, and 1 ml of EPN suspension (200 IJs ml⁻¹) was applied using a hand-held sprayer. Control dishes were sprayed with distilled water before placing the adults. The edges of Petri dishes were dried with tissue paper to prevent the adults sticking to water droplets. Three replicates of each plate (nymphs)/Petri dish (adults) were made per EPN as well as the control. All plates/dishes were placed in a controlled chamber (25 °C, 72% RH, L:D 16:8) for six days. Mortality readings were taken daily for six consecutive days. To confirm that insect mortality was caused by the activity of the EPNs, the presence of EPN was evaluated by dissecting nymphs and adult insect bodies under a 40× magnifying stereomicroscope. This was done after rinsing single dead nymphs and adults in a conical flask filled with 20 ml of distilled water, for removing nematodes from their surface. Nymphs and adults were then placed on a moist filter paper padded plastic

Petri dish and maintained at 25 °C for three days according to Glazer and Lewis (2000).

Entomopathogenic nematodes: greenhouse trials

To evaluate the efficacy of EPNs against *H. obsoletus* nymphs and adults, greenhouse trials were conducted on potted stinging nettle (*Urtica dioica* L.) and faba bean (*Vicia faba* L.) plants, respectively. The nettle plants (diameter about 0.1 m and height approximately 0.4 m) were grown in 3.0 l pots, while the bean plants (diameter about 0.1 m and height approximately 0.25 m) in 1.0 l pots and placed in transparent plastic ventilated cages. All plants were in good vegetative condition and did not show symptoms of biotic and abiotic stresses. In each pot with nettle, 5 ml of EPN suspension (400 IJs ml⁻¹) was applied to the soil using a hand-held sprayer, and a total of 20 nymphs were placed after treatment. The control potted plants were treated with 5 ml of distilled water before placing the nymphs. On each caged bean plant, 10 ml of EPN suspension (400 IJs ml⁻¹, with 0.02% Tween 80) was applied using a hand-held sprayer, and 15 *H. obsoletus* adults were released. The control caged plants were treated with 10 ml of distilled water with 0.02% Tween 80 before releasing insect adults. Three replicates, arranged in randomized blocks, were

made per EPN as well as the control. Potted plants were kept in a controlled chamber (25 °C, 72% RH, L:D 16:8). Mortality was recorded six days after treatment. As described above for the bioassay, mortality due to nematodes infection was confirmed by dissecting the dead nymph and adult insect bodies under a 40× magnifying stereomicroscope.

Entomopathogenic fungi: laboratory bioassays

Initially, the nine EPF isolates provided by the Institute for Biological Control JKI (Table 1) were screened for their entomopathogenic activity against *H. obsoletus* adults in plastic Petri dishes (8.5 cm diameter), filled to a depth of 5 mm with a mixture of plaster of Paris and charcoal (10:1) and moistened with distilled water (Green 1964; Langer et al. 2005). In each Petri dish, fresh bindweed (*Convolvulus arvensis* L.) shoots (5 cm) were placed as a food source, 1 ml of EPF conidia suspension (10^8 conidia ml⁻¹) was applied using a hand-held sprayer, and 12 adults were released. The control dishes were treated with distilled water before the release of insects. Three replicates were conducted per EPF isolate as well as the control. Petri dishes were kept in a controlled chamber (25 °C, 72% RH, L:D 16:8). Mortality readings were taken after six days. Single dead insects from each EPF isolate were placed in malt extract peptone agar plates kept for three days at room temperature (22 °C) and for five days at 4 °C. Mortality due to EPF infection was confirmed through the observation of EPF mycelium growth on the insect body. Subsequently, EPF isolates, found active against *H. obsoletus* in the initial screening test, were employed in a bioassay applying conidial suspension at different concentrations (10^6 , 10^8 , 10^{10} conidia ml⁻¹). The bioassays were conducted in plastic Petri dishes as described above for the initial screening test. Three replicates were conducted per concentration per EPF isolate as well as the control. Petri dishes were kept in controlled chamber (25 °C, 72% RH, L:D 16:8) for six days. Mortality readings were taken daily for six consecutive days. Single dead insects from each bioassay were placed in malt extract peptone agar plates and checked to confirm EPF-related mortality as described above.

Entomopathogenic fungi: greenhouse trials

Greenhouse trials were conducted on potted stinging nettle plants, placed singly in transparent plastic ventilated cages, to evaluate the efficacy against *H. obsoletus* adults of the previously selected JKI EPF isolates along with two commercial EPFs (*M. anisopliae* and *L. muscarium*). On each caged plant, 1 ml of EPF suspension (10^8 conidia ml⁻¹) was applied on the potted plants using a hand-held sprayer, and 15 *H. obsoletus* adults were released. The control caged plants were treated with 1 ml of distilled water before the release of insects. Three replicates were made per EPF as well as the control. Caged potted plants were kept in a controlled chamber (25 °C, 72% RH, L:D 16:8) for six days. Mortality readings were taken after six days. EPF-related mortality was confirmed by observing the EPFs mycelium growth directly on insect bodies on stinging nettle or placing single dead insects in malt extract peptone agar plates as described above.

Statistical analysis

In the bioassays, the median lethal time (LT₅₀) for EPNs against nymphs and adults was calculated from daily mortality data. The median lethal concentration (LC₅₀) of EPFs against adults was log₁₀-transformed and calculated from the data obtained on the fourth day post-inoculation (dpi). LT₅₀ and LC₅₀ results with their fiducial confidence limits were obtained by R statistical package “ecotox” (Hlina 2020) based on the probit analysis (Finney 1971; Wheeler et al. 2006; Robertson et al. 2007). In the screening (EPFs) and greenhouse trials (EPNs and EPFs), mortality data of nymphs and adults were tested for normality and equality of variance with the Shapiro-Wilk test followed by Levene’s test. Greenhouse trials data were subjected to ANOVA followed by Tukey’s honestly significant difference (HSD) post-hoc test for multiple comparisons. Graphical representations of the statistical analysis results were produced using R statistical package “ggplot2” (Wickham et al. 2020). All the analyses were done using R (version 3.6.2) (R Core Team 2019).

Results

Biocontrol efficacy of entomopathogenic nematodes against *H. obsoletus* nymphs and adults

In the bioassay, *S. carpocapsae* was the most effective with a LT_{50} value of 3.24 and 3.69 days post-inoculation (dpi) for *H. obsoletus* nymphs and adults, respectively (Fig. 1a, b). *S. feltiae* was the second most effective with a LT_{50} of 3.49 and 4.05 dpi for nymphs and adults, respectively (Fig. 1c, d). The mixture of *H. bacteriophora* and *S. feltiae* showed a LT_{50} of 3.65 and 4.61 dpi for nymphs and adults, respectively (Fig. 1e, f). *H. bacteriophora* (Hb) was the least effective with a LT_{50} of 4.34 and 5.06 dpi for nymphs for adults (Fig. 1g, h) (Supplementary Table S1). In the greenhouse trials, results of ANOVA [nymphs ($F_{4,10} = 82.55$; $p < 0.001$); adults ($F_{4,10} = 87.96$; $p < 0.001$)], followed by Tukey's HSD test, revealed that the average mortality rate of *H. obsoletus* nymphs and adults treated with the different EPNs were significantly higher than the non-treated control. Among EPNs, *S. carpocapsae* showed the highest efficacy against *H. obsoletus* nymphs (average

mortality rate 86.67%) and adults (81.67%), while *H. bacteriophora* the lowest (56.67% against nymphs; 55.00% against adults) (Fig. 2a, b; Supplementary Table S2).

Biocontrol efficacy of entomopathogenic fungi against *H. obsoletus* adults

In the initial screening, results of ANOVA ($F_{9,20} = 20.39$; $p < 0.001$), followed by Tukey's HSD test, revealed that the average mortality rate of *H. obsoletus* adults were significantly higher than the non-treated control in all EPFs treatments, except *B. bassiana* strain 1124 and *M. anisopliae* 1428 (Fig. 2c). In the bioassay, the seven effective EPFs were employed to define their proper concentration leading to 50% of mortality of *H. obsoletus* adults. *I. fumosorosea* strains 1497 and 1499 were found to be the most virulent against *H. obsoletus* adults with a LC_{50} (\log_{10} concentration) of 6.07 and 6.46 conidia ml^{-1} , respectively. *M. anisopliae* strains 1429 and 1430 showed a LC_{50} of 8.21 and 8.51 conidia ml^{-1} , respectively. *B. bassiana* strains 1125 and 1126 were the least virulent with a LC_{50} of 8.89 and 9.23 conidia

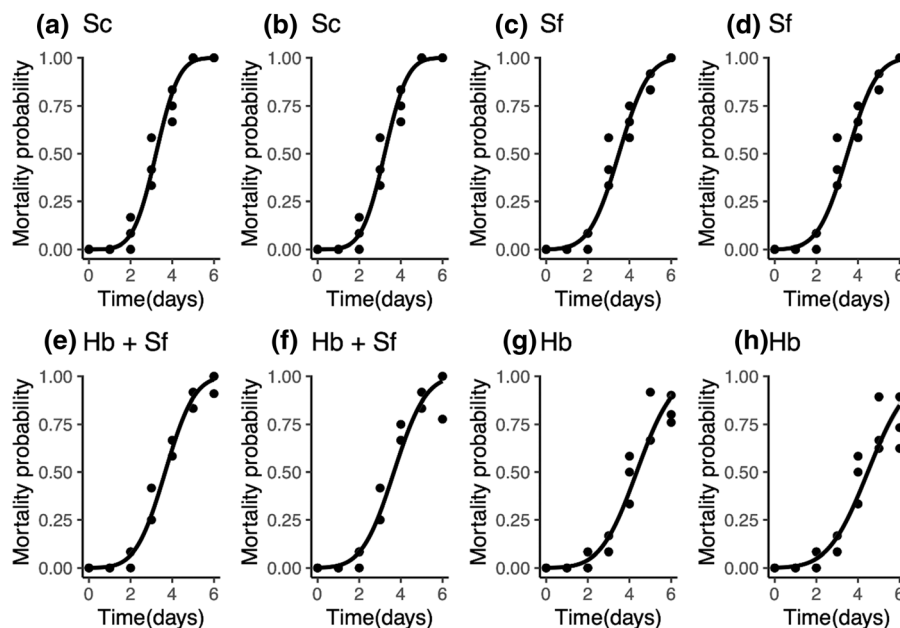


Fig. 1 Time-to-death curve of different EPNs against nymphs and adults of *H. obsoletus*. Black dots represent the observations. Black curves were computed within R statistical package “ecotox” using a glm smoothing method based on a binomial distribution with a probit link function. *S. carpocapsae* (Sc)

against *H. obsoletus* nymphs (a) and adults (b); *S. feltiae* (Sf) against nymphs (c) and adults (d); *H. bacteriophora* + *S. feltiae* (Hb + Sf) against nymphs (e) and adults (f); *H. bacteriophora* (Hb) against nymphs (g) and adults (h)

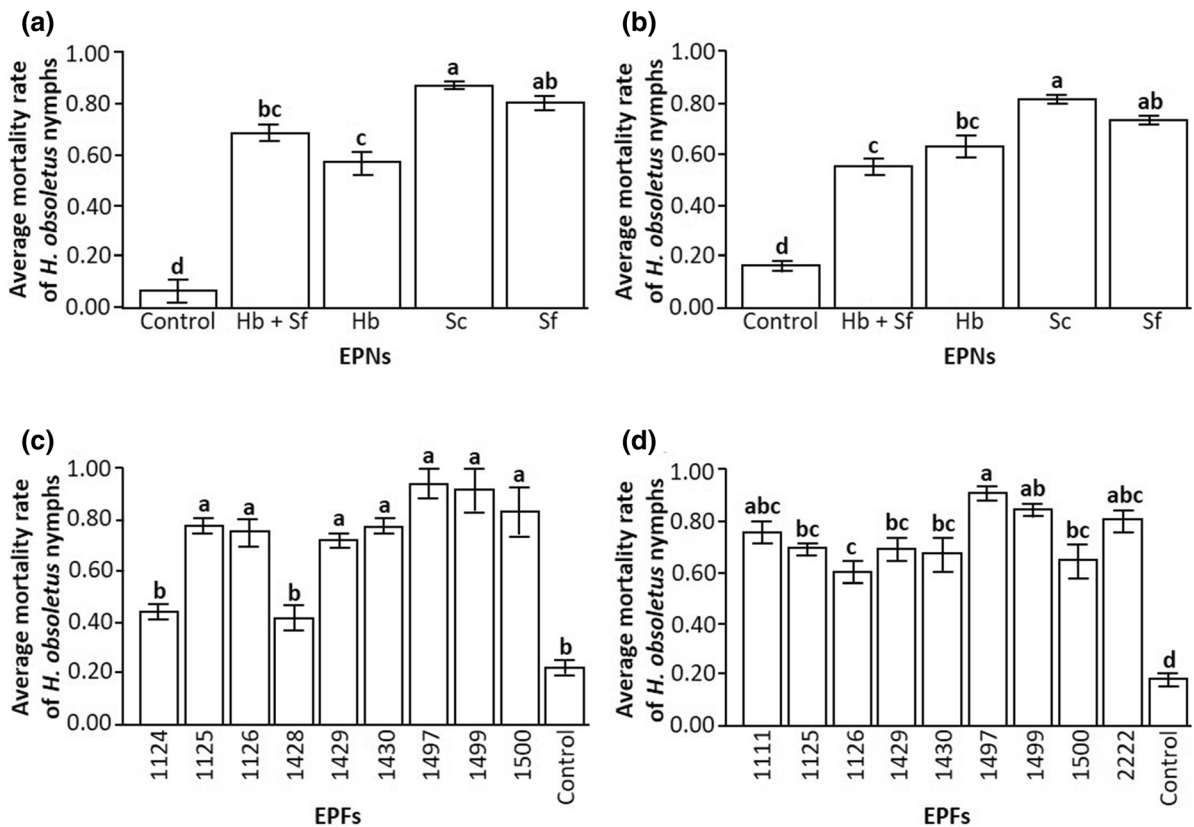


Fig. 2 Average (\pm SE) mortality rate of *Hyalesthes obsoletus* nymphs and adults treated with EPNs and EPFs. EPNs greenhouse trials against *H. obsoletus* nymphs (a) and adults (b); EPFs initial screening test against *H. obsoletus* adults (c); EPFs greenhouse trials against *H. obsoletus* adults (d). On each bar: letters (a-d) indicate significant differences ($p < 0.05$). Acronyms in (a) and (b): Hb + Sf (*H. bacteriophora* + *S.*

feltiae); Hb (*H. bacteriophora*); Sc (*S. carpocapsae*); Sf (*S. feltiae*). Acronyms in (c) and (d): 1124, 1125, 1126 (*B. bassiana* strains JKI-BI-1124, JKI-BI-1125, JKI-BI-1126); 1428, 1429, 1430 (*M. anisopliae* strains JKI-BI-1428, JKI-BI-1429, JKI-BI-1430); 1497, 1499, 1500 (*I. fumosorosea* strains JKI-BI-1497, JKI-BI-1499, JKI-BI-1500); 1111 (*M. anisopliae* strain 1111); 2222 (*L. muscarium* strain 2222)

ml⁻¹, respectively (Fig. 3; Supplementary Table S3). In the greenhouse trials, results of ANOVA ($F_{9,20} = 23.87$, $p < 0.001$), followed by Tukey's HSD test, revealed that the average mortality rate of *H. obsoletus* adults treated with the different EPFs were significantly higher than the non-treated control. Among EPFs, *I. fumosorosea* strain 1497 showed the highest efficacy against *H. obsoletus* adults (average mortality rate 91.1%). This percentage was not significantly different in comparison to that obtained by *I. fumosorosea* strain 1499 (84.45%), *M. anisopliae* strain 1111 (75.56%), and *L. muscarium* strain 2222 (80.00%). On the contrary, it was significantly different in comparison to that obtained by *M. anisopliae* strains 1429 (68.89%) and 1430 (66.67%), and *B. bassiana* strains 1125 (68.89%). *B. bassiana* strain

1126 showed the lowest efficacy (60.00%) (Fig. 2d; Supplementary Table S4).

Discussion

Due to the complex life cycle of *Hyalesthes obsoletus*, most strategies to control its populations in the vineyard agro-ecosystem are not effective or can impact the environment (Maixner and Mori 2013). In the last years, biocontrol has been proposed and frequently utilized as sustainable strategy to control plant pathogen insect vectors (Kumar 2016; Abdel-Razek et al. 2017; Abd El-Ghany et al. 2018). Entomopathogenic nematodes (EPNs) and fungi (EPFs) have been largely employed as effective

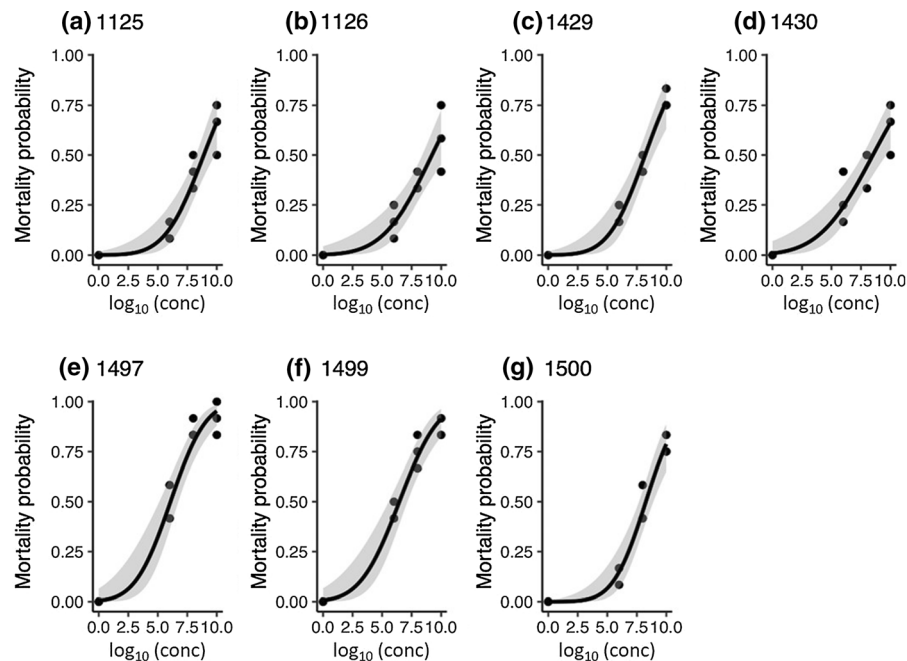


Fig. 3 Dose-response curve of EPFs against *H. obsoletus* adults. Black dots represent the observations. Black curves were computed within R statistical package “ecotox” using a glm smoothing method based on a binomial distribution with a probit link function. Shaded grey areas represent the 95% confidence

intervals. *B. bassiana* strains JKI-BI-1125 (1125) (a), JKI-BI-1126 (1126) (b); *M. anisopliae* strains JKI-BI-1429 (1429) (c), JKI-BI-1430 (1430) (d); *I. fumosorosea* strains JKI-BI-1497 (1497) (e), JKI-BI-1499 (1499) (f), JKI-BI-1500 (1500) (g)

biocontrol agents against insects with a cryptic life cycle, including phytoplasma vectors (Grewal et al. 2005; Lacey and Georgis 2012; Guerrero and Pardey 2019), making this approach promising also for the main vector of ‘*Candidatus Phytoplasma solani*’ to grapevine, *H. obsoletus*.

The results obtained in this study demonstrated that all the examined EPNs are able to kill *H. obsoletus* nymphs and adults and that the EPFs, except *Beauveria bassiana* strain 1124 and *Metharizium anisopliae* strain 1428, are able to control the adults in both laboratory bioassays and greenhouse trials, exhibiting a range of effectiveness related to their virulence against the target insect. In all conducted trials, *Steinernema carpocapsae* and *Isaria fumosorosea* were found to be the most effective biocontrol agents of *H. obsoletus* among the examined EPNs and EPFs, respectively.

Concerning *Steinernema* spp., our findings are in agreement with Le Vieux and Malan (2013) showing that, in laboratory bioassay performed against the vine mealybug *Planococcus ficus*, the EPN *Steinernema yirgalemense* moved 15 cm vertically downward, and

infected its insect target inducing a mortality of 95%. Another study demonstrated that the combination of *S. yirgalemense* with specific adjuvants increased its biocontrol activity against the vine mealy bug on grapevine leaves in both laboratory and semi-field conditions (Platt et al. 2019). Such evidence fortifies the possibility of applying *Steinernema* spp. in the open field against both subterranean forms and adults of *H. obsoletus*. Among tested EPNs, also *Heterorhabditis bacteriophora* showed a high efficacy in *H. obsoletus* biocontrol. Interestingly, this EPN was reported to be effective against the nymphs of *Haplaxius crudus*, the insect vector of ‘*Candidatus Phytoplasma palmae*’ associated with Palm Lethal Yellowing disease in Florida, USA (Guerrero and Pardey 2019), and of *Aeneolamia* spp., a putative vector of genetically distinct phytoplasmas (Pérez Miliàn et al. 2018). Moreover, *H. bacteriophora* strongly reduced the survival of the root-form of grapevine phylloxera (English-Loeb et al. 1999). This evidence highlighted that, in vineyard agroecosystems, treatments based on the application of *H. bacteriophora* could be effective against multiple

insect pests. Based on all these evidences, it would be interesting to apply a combination of *S. carpocapsae*, found here as more effective against *H. obsoletus* nymphs and adults, and *H. bacteriophora*, reported in previous studies as the most effective EPN against phytoplasma insect vectors and grapevine insect pests with a cryptic life stage.

Concerning *Isaria fumosorosea*, found here as the most effective EPF against *H. obsoletus*, previous studies showed its biocontrol activity against various nymphal stages of the green leafhopper *Empoasca decipiens* Paoli under laboratory and greenhouse conditions (Tounou et al. 2003; Kodjo et al. 2011). Similar efficacy was found by treatments with *Metarhizium anisopliae* (strain Ma43) and *Beauveria bassiana* (strain Bba113) (Tounou et al. 2003; Kodjo et al. 2011). For all these EPFs, percentage of mortality and LC₅₀ values reported against *E. decipiens* were comparable to those observed in this study against *H. obsoletus* adults. Moreover, promising results obtained in the present study with two strains of *M. anisopliae* confirmed its entomopathogenic activity against *H. obsoletus* adults under laboratory conditions (Langer et al. 2005). Interestingly, *Beauveria bassiana*, two strains of which showed a great biocontrol activity against *H. obsoletus* adults in the present work, was found naturally infecting and causing visual symptoms on *H. obsoletus* adults in Georgia (Caucasus region) (Chkhaidze et al. 2017). Moreover, *B. bassiana* showed an efficacy in biocontrol of young stages and adults of *Scaphoideus titanus* Ball, the insect vector of flavescence dorée phytoplasma, in semi-field and field trials (Mori et al. 2014b). All these evidences underlined that *B. bassiana* can control both the insect vectors of phytoplasmas associated with the main grapevine yellows diseases. Thus, *B. bassiana* strains represent really promising EPFs for application in vineyards.

Effectiveness of EPNs and EPFs, as well as other living organisms used as biocontrol agents, depends on a range of climatic and environmental parameters allowing their liveliness and entomopathogenic activity. In particular, it is crucial that the target insect stage is present when climatic parameters are optimal for EPNs and/or EPFs (Lacey and Georgis 2012; Wang and Wang 2017). In the case of *H. obsoletus*, it is known that the duration of the cryptic (subterranean) phase of its life cycle, involving the nymph stages, is dependent on the degree day units that can be

estimated based on forecasting models measuring the accumulated heat units (Maixner and Mori 2013). Such models allow understanding of the life cycle of the insect as well as narrowing the spraying window of products for plant protection, including EPNs and EPFs. In particular, the spraying window should prioritize two important aspects: (1) the ecological competency of EPNs as well as EPFs; (2) the proper timing for application against the different stages and instars of *H. obsoletus*. In Europe and the Mediterranean area, considering the life cycle of *H. obsoletus* and the environmental conditions suitable for EPNs and EPFs utilized in the present study, it should be recommended to apply EPNs and EPFs on *H. obsoletus* host plants in the open field from mid-September to October and/or in early spring to optimize the activity of each biocontrol agent and avoid resistance in the insect target populations. Moreover, given their ability to colonize the soil after their inoculation (Meyling and Eilenberg 2006; Denno et al. 2008), EPNs and EPFs could reduce the *H. obsoletus* population density for a long time. According to our result EPFs could be applied also with foliar application from end of May till end of June against newly hatched adults before grapevine infestation. Optimized application of entomopathogenic nematodes (on the soil) and fungi (on the plants) can increase the control of *H. obsoletus* nymphs and adults, respectively.

In conclusion, the majority of EPNs and EPFs utilized in the present study showed a considerable biocontrol activity against *H. obsoletus* nymphs and adults in laboratory bioassays and greenhouse trials. The ecological competency of both EPNs and EPFs, the conditions that can impede or enhance their performance, the barriers that can block infection from taking place on the target host, and the possible actions on non-target species should be carefully investigated for a better understanding of their potential performance under field conditions.

Acknowledgements The authors thank Prof Itamar Glazer of ARO, Volcani Center, ISRAEL for his suggestions and text revision.

Authors' contribution Abdelhameed Moussa: Conceptualization, data curation, formal analysis, investigation, methodology, validation, writing - original draft. Michael Maixner: conceptualization, methodology, resources, supervision, writing - review and editing. Dietrich Stephan:

methodology, resources, writing - review and editing. Giacomo Santoiemma: data curation, formal analysis, validation. Alessandro Passera: data curation, formal analysis, validation. Nicola Mori: conceptualization, project administration, resources, supervision, Writing - review and editing. Fabio Quaglino: conceptualization, project administration, supervision, writing - review and editing.

Funding Open Access funding provided by Università degli Studi di Verona, Italy.

Compliance with ethical standards

Conflict of interest The authors declare that there are no conflicts of interest associated with this publication.

Ethical approval This research did not involve any studies with human participants or animals (vertebrates) performed by any of the authors.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Abd El-Ghany NM, Abdel-Razek AS, Djelouah K, Moussa A (2018) Efficacy of bio-rational insecticides against *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) on tomatoes. *Biosci Res* 15:28–40
- Abdel-Razek AS, Abd El-Ghany NM, Djelouah K, Moussa A (2017) An evaluation of some eco-friendly biopesticides against *Bemisia tabaci* on two greenhouse tomato varieties in Egypt. *J Plant Prot Res* 57:9–17
- Alma A, Tedeschi R, Lessio F, Picciau L, Gonella E, Ferracini C (2015) Insect vectors of plant pathogenic mollicutes in the Euro-Mediterranean region. *Phytopath Moll* 5:53–73
- Angelini E, Constable F, Duduk B, Fiore N, Quaglino F, Bertaccini A (2018) Grapevine phytoplasmas. In: Rao GP, Bertaccini A, Fiore N, Lieferting LW (eds) *Characterisation and epidemiology of phytoplasma-associated diseases. Phytoplasmas: plant pathogenic bacteria–I*. Springer, Singapore, pp 123–152
- Bertin S, Picciau L, Acs Z, Alma A, Bosco D (2010) Molecular differentiation of four *Reptalus species* (Hemiptera: Cixiidae). *Bull Entomol Res* 100:551–558
- Bianco PA, Romanazzi G, Mori N, Myrie W, Bertaccini A (2019) Integrated management of phytoplasma diseases. In: Bertaccini A, Weintraub GP, Rao GP, Mori N (eds) *Transmission and management of phytoplasma-associated diseases. Phytoplasmas: plant pathogenic bacteria–II*. Springer, Singapore, pp 208–237
- Bressan A, Turata R, Maixner M, Spiazzi S, Boudon-Padieu E, Girolami V (2007) Vector activity of *Hyalesthes obsoletus* living on nettles and transmitting a stolbur phytoplasma to grapevines: a case study. *Ann Appl Biol* 150:331–339
- Cargnù E, Pavan F, Mori N, Martini M (2012) Identification and phenology of *Hyalesthes obsoletus* (Hemiptera: Cixiidae) nymphal instars. *Bull Entomol Res* 102:504–514
- Chkhaidze N, Burjanadze M, Lobghanidze M, Koridze K (2017) First report of entomopathogenic fungi on the insect vector *Hyalesthes obsoletus* Signoret (Hemiptera: Cixiidae) in Georgia. *IOBC/WPRS Bull* 129:43–46
- Cvrković T, Jović J, Mitrović M, Krstić O, Toševski I (2014) Experimental and molecular evidence of *Reptalus panzeri* as a natural vector of bois noir. *Plant Pathol* 63:42–53
- Denno RF, Gruner DS, Kaplan I (2008) Potential for entomopathogenic nematodes in biological control: a meta-analytical synthesis and insights from trophic cascade theory. *J Nematol* 40:61–72
- English-Loeb G, Villani M, Martinson T, Forsline A, Consolie N (1999) Use of entomopathogenic nematodes for control of grape phylloxera (Homoptera: Phylloxeridae): a laboratory evaluation. *Environ Entomol* 28:890–894
- Finney DJ (1971) *Probit analysis*. Cambridge University Press, Cambridge
- Glazer I, Lewis EE (2000) Bioassays for entomopathogenic nematodes. In: Navon A, Ascher K (eds) *Bioassays of entomopathogenic microbes and nematodes*. CABI, pp 229–247
- Green CD (1964) The effect of crowding upon the fecundity of *Folsomia candida* (William) var. *Distincta* (Bagnall)(Collembola). *Entomol Exp Appl* 7:62–70
- Grewal P, Lewis E, Gaugler R, Campbell J (1994) Host finding behaviour as a predictor of foraging strategy in entomopathogenic nematodes. *Parasitology* 108:207–215
- Grewal PS, Ehlers RU, Shapiro-Ilan DI (2005) *Nematodes as biocontrol agents*. CABI Publishing, Wallingford
- Guerrero MR, Pardey AEB (2019) Selection of entomopathogenic nematodes to control nymphs of *Haplaxius crudus* (Van Duzee) (Hemiptera: Cixiidae). *Am J Entomol* 3:24–29
- Hlina BL (2020) *Ecotox: Analysis of ecotoxicology*. R package version 1.4.2
- Kaya HK, Burlando T, Thurston GS (1993) Two entomopathogenic nematode species with different search strategies for insect suppression. *Environ Entomol* 22:859–864
- Kaya HK, Stock SP (1997) Chapter VI - techniques in insect nematology. In: Lacey LA (ed) *Manual of techniques in insect pathology*. Academic, London, pp 281–324
- Kirchmair M, Huber L, Porten M, Rainer J, Strasser H (2004) *Metarhizium anisopliae*, a potential agent for the control of grape phylloxera. *Biol Control* 49:295–303
- Kodjo T, Hans-Michael P, Komi A, Katharina R, Gilbert Z, Christian B (2011) A laboratory assessment of the potential of selected entomopathogenic fungi to control the green

- leafhopper *Empoasca decipiens* Paoli (Homoptera: Cicadellidae). *J Appl Biosci* 47:3195–3204
- Kosovac A, Jakovljević M, Krstić O, Cvrković T, Mitrović M, Toševski I, Jović J (2019) Role of plant-specialized *Hyalesthes obsoletus* associated with *Convolvulus arvensis* and *Crepis foetida* in the transmission of ‘*Candidatus Phytoplasma solani*’-inflicted bois noir disease of grapevine in Serbia. *Eur J Plant Pathol* 153:183–195
- Kumar OB (2016) Biocontrol of insect pests. In: Omkar (ed) Ecofriendly pest management for food security. Elsevier, San Diego, pp 25–61
- Lacey LA (1997) Manual of techniques in insect pathology. Academic, London
- Lacey LA, Georgis R (2012) Entomopathogenic nematodes for control of insect pests above and below ground with comments on commercial production. *J Nematol* 44:218–225
- Langer M, Maixner M (2004) Molecular characterisation of grapevine yellows associated phytoplasmas of the stolbur-group based on RFLP-analysis of non-ribosomal DNA. *Vitis* 43:191–199
- Langer M, Maixner M, Kirchmair M, Huber L (2005) Efficacy of *Metarhizium anisopliae* against *Hyalesthes obsoletus* (Auchenorrhyncha: Cixiidae). *Vitis* 44:99–100
- Le Vieux PD, Malan AP (2013) The potential use of entomopathogenic nematodes to control *Planococcus ficus* (Signoret)(Hemiptera: Pseudococcidae). *South Afric J Enol Viticul* 34:296–306
- Maixner M (1994) Transmission of German grapevine yellows (Vergilbungskrankheit) by the planthopper *Hyalesthes obsoletus* (Auchenorrhyncha: Cixiidae). *Vitis* 33:103–104
- Maixner M, Johannesen J (2014) Optimized monitoring of host populations of the bois noir vector, *Hyalesthes obsoletus*, based on flight phenology observations. *IOBC/WPRS Bull* 105:151–157
- Maixner M, Mori N (2013) Management of bois noir through vector control. *Phytopath Moll* 3:4–46
- Meyling NV, Eilenberg J (2006) Occurrence and distribution of soil borne entomopathogenic fungi within a single organic agroecosystem. *Agric Ecosyst Environ* 113:336–341
- Mori N, Mitrović J, Smiljković M, Duduk N, Paltrinieri S, Bertaccini A, Duduk B (2013) *Hyalesthes obsoletus* in Serbia and its role in the epidemiology of corn reddening. *Bull Insectol* 66:245–250
- Mori N, Pavan F, Maixner M (2014a) Control of *Hyalesthes obsoletus* nymphs based on chemical weeding and insecticides applied on *Urtica dioica*. *Vitis* 53:103–109
- Mori N, Tonello D, Posenato G, Pozzebon A, Duso C (2014b) Efficacy of biopesticides against *Scaphoideus titanus* ball in different experimental conditions. *IOBC/WPRS Bull* 105:45–48
- Moussa A, Mori N, Faccincani M, Pavan F, Bianco PA, Quaglino F (2019) *Vitex agnus-castus* cannot be used as trap plant for the vector *Hyalesthes obsoletus* to prevent infections by ‘*Candidatus Phytoplasma solani*’ in northern Italian vineyards: experimental evidence. *Ann Appl Biol* 175:302–312
- Pérez Milián JR, Pérez Pérez Y, Álvarez JF, Ruano Rossil JM (2018) Biological control of sugarcane froghopper, *Aeneolamia* spp., by the nematode *Heterorhabditis bacteriophora* and entomopathogenic fungi *Metarhizium anisopliae* and *Beauveria bassiana* as economically sustainable option. *CEIBA* 55:21–27
- Platt T, Stokwe NF, Malan AP (2019) Foliar application of *Steinernema yirgalemense* to control *Planococcus ficus*: assessing adjuvants to improve efficacy. *South Afr J Enol Viticul* 40:1–7
- Quaglino F, Sanna F, Moussa A, Faccincani M, Passera A, Casati P, Bianco PA, Mori N (2019) Identification and ecology of alternative insect vectors of ‘*Candidatus Phytoplasma solani*’ to grapevine. *Sci Rep* 9:19522
- Quaglino F, Zhao Y, Casati P, Bulgari D, Bianco PA, Wei W, Davis RE (2013) ‘*Candidatus Phytoplasma solani*’, a novel taxon associated with stolbur-and bois noir-related diseases of plants. *Int J Syst Evol Microbiol* 63:2879–2894
- R Core Team (2019) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria (<https://www.r-project.org/>)
- Robertson JL, Savin NE, Russell RM, Preisler HK (2007) Bioassays with arthropods. CRC Press
- Sharon R, Tomer M, Sokolsky T, Sofer-Arad C, Zahavi T (2015) Trap plants reduces grapevine yellows disease incidence in commercial vineyards. *Phytopath Moll* 5:S107–S108
- Stöckmann M, Biedermann R, Nickel H, Niedringhaus R (2013) The nymphs of the planthoppers and leafhoppers of Germany. WABV Fründ
- Tounou AK, Agboka K, Poehling HM (2003) Evaluation of the entomopathogenic fungi *Metarhizium anisopliae* and *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) for control of the green leafhopper *Empoasca decipiens* (Homoptera: Cicadellidae) and potential side effects on the egg parasitoid *Anagrus atomus* (Hymenoptera: Mymaridae). *Biocontrol Sci Tech* 13:715–728
- Wang C, Wang S (2017) Insect pathogenic fungi: genomics, molecular interactions, and genetic improvements. *Annu Rev Entomol* 62:73–90
- Wheeler MW, Park RM, Bailey AJ (2006) Comparing median lethal concentration values using confidence interval overlap or ratio tests. *Environ Toxic Chem* 25:1441–1444
- Wickham H, Chang W, Henry L, Pedersen TL, Takahashi K, Wilke C, Woo K, Yutani H, Dunnington D (2020) ggplot2: Create elegant data visualisations using the grammar of graphics. R package version 3.3.2

Abdelhameed Moussa is a doctoral candidate at Department of Agricultural and Environmental Sciences, University of Milan, Italy. His PhD project focuses on developing integrated sustainable strategies for the control of grapevine bois noir disease.

Michael Maixner is deputy head of the Institute for Plant Protection in Fruit Crops and Viticulture, Julius Kühn-Institut, Siebeldingen, Germany. His research activity focuses on etiology, epidemiology, and control of phytoplasma diseases and other vector transmissible diseases of grapevine.

Dietrich Stephan is head of the Microbiology and Bioengineering laboratory and leads the working group Entomopathogenic fungi at Institute for Biological Control, Julius Kühn-Institut, Darmstadt, Germany. His research focuses on

production, formulation, and application of micro-organisms for plant protection.

Giacomo Santoemma has a post-doc position at Department of Agronomy, Food, Natural resources, Animals and Environment, University of Padova, Italy. His research focuses on spatial ecology and environmental data analysis of insect pests and vectors.

Alessandro Passera has a post-doc position at Department of Agricultural and Environmental Sciences, University of Milan, Italy. His research interests are beneficial endophytic bacteria, biocontrol agents, and microbiota in relation to plant health.

Nicola Mori is professor of general and applied entomology at Department of Biotechnology, University of Verona, Italy. His research focuses on biology and ecology of insect pests and phytoplasma vectors, and their control through sustainable strategies.

Fabio Quaglino is professor of plant pathology at Department of Agricultural and Environmental Sciences, University of Milan, Italy. His research focuses on taxonomy and ecology of phytoplasmas, and epidemiology and control of the associated diseases.