



A novel homozygous *VPS11* variant may cause generalized dystonia

Running head: *VPS11* variant associated with dystonia

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Abstract

In this work we describe the association of a novel homozygous *VPS11* variant with adult-onset generalized dystonia, providing a detailed clinical report and biological evidence of disease mechanism. Vps11 is a subunit of the HOPS complex, which promotes the fusion of late endosomes and autophagosomes with the lysosome. Functional studies on mutated fibroblasts showed marked lysosomal and autophagic abnormalities, which improved after overexpression of the wild type Vps11 protein. In conclusion, a deleterious *VPS11* variant, damaging the autophagic and lysosomal pathways, is the probable genetic cause of a novel form of generalized dystonia.

Introduction

Dystonia is a hyperkinetic movement disorder characterized by sustained or intermittent muscle contractions causing abnormal movements and/or postures. If the trunk and at least two other sites are involved, dystonia is defined as generalized¹. Inherited dystonias can be classified as isolated (dystonia is the only motor feature, except for tremor), combined (dystonia is associated with other movement disorders) or complex (dystonia co-occurs with other neurologic or systemic manifestations)².

Typically, isolated and combined dystonia have not characteristic features at brain MRI. On the other hand, complex dystonia often shows pathognomonic MRI changes. Indeed, complex dystonia is one of the most frequent clinical presentations of neurodegenerations with brain iron accumulation (NBIA), which is a group of genetic disorders displaying progressive iron accumulation in basal ganglia².

Vps11 aggregates with other Vps proteins (i.e., Vps16, Vps18, Vps33, Vps41, and Vps39) to form the “homotypic fusion and protein sorting complex (HOPS)”. The HOPS complex promotes the fusion of late endosomes and autophagosomes with lysosomes^{3,4}.

Homozygous *VPS11* mutations were associated with hypomyelinating leukodystrophy 12 (HLD12), characterized by appendicular spasticity, truncal hypotonia, opisthotonic posturing, and seizures. Brain MRIs of affected subjects present a thin corpus callosum and diffused hypomyelination. Two homozygous mutations were described so far (c.2536T>G p.C846G and c.1158_1184del p.L387-G395del) (Figure 1A)⁵⁻⁷.

Here we describe a novel homozygous *VPS11* variant probably causative of adult-onset generalized dystonia. We provide strong evidence of variant deleteriousness and demonstrate its highly damaging impact on the autophagy-lysosomal pathway.

Materials and methods

Clinical data

The subject underwent several neurological examinations, brain MRI and neurophysiological studies. Blood samples and a skin biopsy were collected. The Ethics Committee of the IRCCS Foundation Ca' Granda Ospedale Maggiore Policlinico (Milan, Italy) approved the study. Written informed consent for publication of clinical details, clinical images, and video recording was obtained from the patient.

Genetic analysis

The genomic DNA of the proband was analyzed by whole-exome sequencing (WES) using the Nextera Rapid Capture Exome Library kit (Illumina) on the Illumina NextSeq500 platform. Reads alignment and variant calling/annotation were performed using standard procedures. The candidate *VPS11* variant was validated by Sanger sequencing in the affected subject and his relatives. Homozygosity mapping was performed starting from WES data using the HomozygosityMapper online tool (<http://www.homozygositymapper.org/>)

Cell cultures and plasmid transfection

Skin fibroblasts of the patients and three controls were cultured in DMEM high glucose supplemented with 15% Fetal Bovine Serum (Euroclone), 1% penicillin/streptomycin (Sigma-Aldrich), and 1% of Amphotericin B (Sigma-Aldrich). Patient fibroblasts were transfected through lipofectamine with a wild-type *VPS11* untagged plasmid (HG21081-UT, Sinobiological).

Protein blotting and biochemical studies

Western blot analyses were performed in triplicate on whole-protein lysates from cultured fibroblasts using the following primary antibodies: Vps11 (HPA039020, Sigma-Aldrich); Actin (A2066, Sigma-Aldrich), Lysosome-associated membrane glycoprotein 1 - LAMP1 (ab25630, Abcam), β -glucocerebrosidase - GBA (EPR5143(3), Abcam), α -glucosidase - GAA (02D05, Genzyme), Acid ceramidase - ASAH1 (sc-28486, Santa Cruz Biotechnology), p62 (MABN130, Sigma-Aldrich), LC3A/B (ab58610, Abcam), Beclin-1 (D40C5, Cell Signaling Technology). Enzyme activities of lysosomal and plasma membrane glycohydrolases were determined in total cell lysates using already published methods⁸. A two-tailed Student's t-test was performed to assess the statistical significance of the protein amount and lysosomal activities differences in the patient compared to the fibroblasts of three controls.

Electron microscopy

Approximately 5×10^6 fibroblasts were incubated with the fixing solution (glutaraldehyde 3%). Samples were dehydrated through graded alcohols, infiltrated, and embedded in Spurr's resin. Ultrathin sections (60 nm) were cut with Ultratome Nova (LKB). Observations were performed using a EM 109 transmission electron microscope (Zeiss).

Results

Clinical features

The family of the proband lived in a very small and geographically isolated village of Southern Italy for many generations. Family members considered very plausible the distant consanguinity of the parents (I.1 and I.2). The father of the proband died at the age of 42 because of a cerebral hemorrhage; the mother died at the age of 51 because of the complications of asthma due to bacterial pneumonia. Two of proband's brothers (II.2 and II.6) died within the first year of life for unknown reasons. All the siblings, which had by far passed their brother's age at onset, were reported healthy. All available siblings were neurologically examined and none of them displayed any sign of dystonia or neurological involvement (Figure 1B).

The proband was born at term after an uneventful pregnancy and had normal psychomotor development. From the age of 30 years, the proband developed progressive dystonic postures, initially affecting the left upper limb. In the following 5 years, dystonia progressively became generalized, involving the trunk, limbs, neck, and larynx. Moreover, progressive dysarthria and dysphagia appeared.

The proband reached our clinic at 40 years of age presenting with severe generalized dystonia affecting the trunk, face, neck (torticollis), and limbs (left > right), accompanied by moderate dysphagia and complete anarthria. Moderate lower limb hyperreflexia was also observed (Video). The proband did not display any evident major cognitive impairment; however, due to the limitations related to complete anarthria and severe motor impairment an in-depth neuropsychological assessment was not performed.

Brain MRI displayed mild brain atrophy with more conspicuous involvement of basal ganglia. Interestingly, substantia nigra, red nucleus, and globus pallidus presented a marked symmetrical hypointensity in T2-weighted sequences. The T2*-weighted sequence showed symmetrical bipallidal hypointensity, which may be suggestive of iron accumulation (Figure 1C). Neurophysiological studies were unremarkable.

Therapeutic challenges with levodopa and anticholinergics did not bring any benefit. Botulinum toxin injections brought a slight improvement of the dystonic posture of the neck and left limbs. At the age of 50, the patient underwent bilateral deep brain stimulation of the internal globus pallidus (GPi DBS). No significant improvement of the dystonic features was observed after DBS (Video).

Genetic analysis

The suspected consanguinity of the parents suggested a homozygous mutation as the cause of the disease. WES was performed on the proband. No pathogenic mutations were found in known disease genes for inherited dystonia and NBIA. A filtering analysis for rare (allele frequency <0.1%) homozygous variants with protein impact was performed. Only the c.136C>T, p.P46S variant in *VPS11* gene was identified. Rare biallelic variants were identified either in obvious non-candidate genes or were carried also by healthy siblings. The *VPS11* variant lies on chromosome 11 in one of the two homozygous regions identified by homozygosity mapping (GRCh37: Chr 6: 30993880-31238851 and Chr11: 117985545-123454978) (Figure 1E). Sanger sequencing confirmed the variant to be homozygous in the proband (II.7) and heterozygous in subjects II.3, II.4, and II.8 (Figure 1B and 1E).

The p.P46S *VPS11* variant is novel and absent from public databases (i.e., 1000G and gnomAD). The Proline at position 46 is highly conserved in mammalian orthologues (Figure 1F) and is localized in the Vps11 N-terminal β -propeller domain (Figure 1A), which is known to be essential for protein-protein interactions⁹. Several in silico tools predict the *VPS11* variant to be deleterious (Polyphen 2: Possibly Damaging 0.495; MutationTaster: Disease Causing 0.999; CADD-PHRED: Deleterious 23.4). Moreover, the Pro > Ser substitution is non-conservative and the peculiar properties of Proline residues (e.g., their rigid conformation with low flexibility) are frequently important to stabilize protein structure. For these reasons, *VPS11* was selected as the candidate etiological gene.

Functional studies

Immunoblot studies on total lysates from cultured fibroblasts showed a slight but statistically significant increase of Vps11 protein amount in mutated cells compared to controls. In addition, patient fibroblasts showed a striking increase in autophagic proteins p62, LC3A, and LC3B, with normal Beclin-1 levels, and augmented lysosomal proteins and hydrolases (LAMP1, β -glucocerebrosidase, acid α -glucosidase, and acid ceramidase), suggesting a significant accumulation of the late-endosomal and lysosomal components (Figure 2A).

Transient overexpression of wild-type Vps11 protein in patient fibroblasts significantly improved the pathological phenotype at the protein level, demonstrating the deleterious effect of the Vps11 p.P46S variant (Figure 2A).

Lysosomal enzymatic activities in patient fibroblasts (i.e., β -glucocerebrosidase, β -galactosidase, and β -hexosaminidase) were three to four times higher than in controls. The plasma membrane enzymatic activities were also significantly increased, of about two to three times compared to controls (Figure 2B).

Electron microscopy of patient fibroblasts showed large clear vacuolar structures, which were completely absent in control fibroblasts (Figure 3). This result appears to be consistent with a morphological alteration of the autophagic and/or endo-lysosomal compartments, in line with the results of protein studies.

Discussion

The identification of a *VPS11* deleterious variant in a patient with adult-onset generalized dystonia and neurodegeneration confirms the important role of the lysosomal and autophagic pathways in the pathogenesis of neurodegenerative disorders ⁷.

In a previous study, knockdown of *VPS11* gene in HeLa cells caused an increase of autophagy markers (i.e., p62, LC3A, and LC3B). Moreover, the accumulation of immature autophagosomes and the reduction of autolysosomes were observed. These data suggest that *VPS11* knockdown severely impairs autophagy flux ⁶.

In the present study, patient fibroblasts presented an over-expression of autophagic proteins p62 and LC3B without alteration of Beclin-1 levels, suggesting an accumulation of autophagosomes without autophagy

induction. Moreover, an increased amount and activity of lysosomal hydrolases suggested the accumulation of late endo-lysosomes. In addition, considering the increased plasma membrane lysosomal activities, these stored vesicles are probably partly exocytosed. The transfection of patient-derived fibroblasts with the wild-type *VPS11* plasmid and consequent Vps11 overexpression ameliorated the pathological phenotype at the protein level, suggesting a pathogenic role of the identified *VPS11* variant. These results are in line with previous *VPS11* knockdown models, indicating that the p.P46S variant is probably a loss-of-function mutation (Figure 2C).

Hörtnagel *et al.* conducted a microscope analysis on tissue samples derived from subjects affected with HLD12 carrying a homozygous *VPS11* mutation (p.C846G). In these patients, electron microscopy on skin biopsies showed several intra-cellular clear vacuoles⁷. In our study, electron microscopy analysis on patient fibroblasts showed a strikingly consistent result, suggesting that the cytoplasmic accumulation of clear vacuoles, consistent with an alteration of the endo-lysosomal compartment, is the pathological hallmark of *VPS11*-related disease.

Very interestingly, mutations of *VPS16* and *VPS41* genes were recently found to cause complex dystonia, often associated with brain abnormalities compatible with brain iron accumulation¹⁰⁻¹². Vps16 and Vps41 associate with Vps11 to form the HOPS complex. Therefore, our finding supports the hypothesis that the disruption of HOPS normal function is crucial in the pathogenesis of this group of complex dystonias.

Our findings suggest that *VPS11* mutations are associated with at least two different phenotypes: the infantile-onset hypomyelinating leukodystrophy 12 (HLD12) and the later onset *VPS11*-associated neurodegeneration. The identification of additional cases will help to disentangle the genotype-phenotype correlation in *VPS11*-related disorders. Phenotypic heterogeneity associated with mutations of a single gene is not a novelty in the field of inherited dystonias; for example, *PLA2G6* mutations can cause both Infantile Neuroaxonal Dystrophy (INAD) and the later onset *PLA2G6*-related dystonia-parkinsonism.

Intriguingly, lysosomal impairment was already associated with brain iron accumulation, such as in the case of *WDR45* and *ATP13A2* gene mutations, which are directly involved in autophagic and lysosomal functions, respectively. Moreover, lysosomes and autophagy play a central role in the cellular recycling of iron from aged organelles, such as damaged mitochondria¹³. One can speculate that the abnormality of lysosomal-

autophagic function described here probably adversely affects the handling of intra-cellular iron and could contribute to the process of neurodegeneration with brain iron accumulation. However, more studies are needed to comprehend the link between lysosomal-autophagic dysfunction and neurodegeneration in *VPS11*-associated neurodegeneration. In addition, neuropathological evidence would be necessary to definitively establish the nature of MRI abnormalities of this disorder.

In conclusion, this work strongly suggests the association of a recessive deleterious variant of *VPS11* gene with a novel form of generalized dystonia and provides in vitro evidence for a crucial role of the lysosomal-autophagic pathway in the pathogenesis of this neurodegenerative disorder.

Acknowledgments

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Author Contributions

EM and ADF contributed to the conception and design of the study; EM, FC, LB, SS, LS, FB, GF, MM, MG, CE, MA and SC contributed to the acquisition and analysis of data; EM, SD, GPC, NB and ADF contributed to drafting the text and preparing the figures.

Potential Conflicts of Interest

None.

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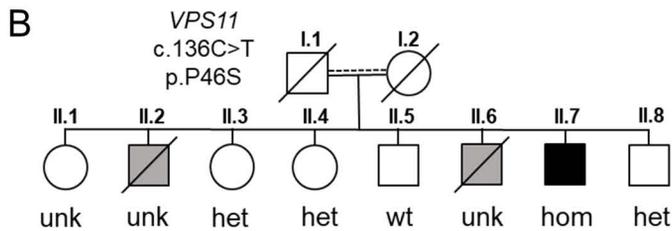
Figure legends

Figure 1: Genetic analysis, family pedigree, and brain MRI: A) Vps11 protein domains with the positions of the pathogenic mutations already described and the deleterious variant presented here (underlined); B) Pedigree of the family under study. Black symbol denotes the affected individual. Grey symbol indicates unknown status; C) Brain MRI of the subject II.7. Axial T2 (1-3) and coronal FLAIR (4-6) display bilateral T2 hypointensity of substantia nigra, red nucleus, and globus pallidus. Axial FFE (7) shows bilateral pallidal hypointensity; D) Homozygosity mapping plot displays two homozygosity peaks on chromosomes 6 and 11 in the proband. E) Electropherograms of the *VPS11* wild-type (upper) and the c.136C>T homozygous variant (lower) of the proband (II.7). F) Alignment of Vps11 protein homologs shows the conservation of the mutated amino acid (Proline46).

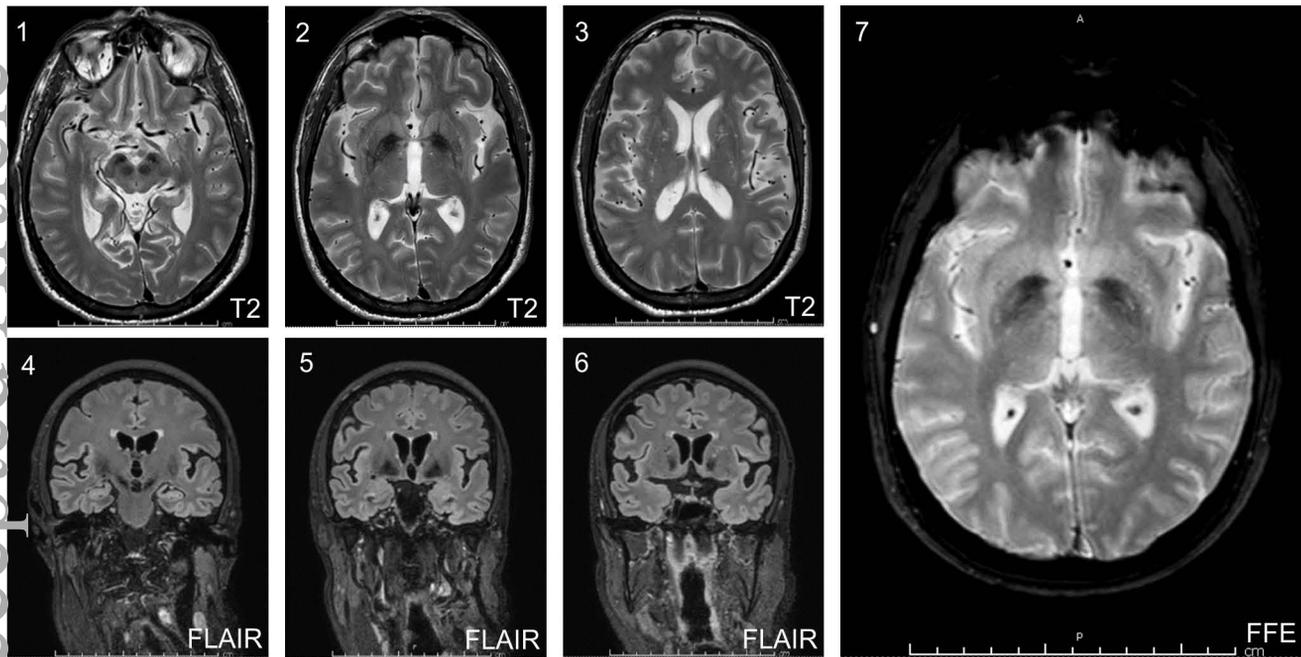
Figure 2: Immunoblot of autophagic and lysosomal proteins and lysosomal enzymatic activities: A) Immunoblots of fibroblast lysates display a statistically significant increase of autophagic and lysosomal proteins in proband cells (II.7) compared to controls (CTR). The overexpression of wild-type Vps11 protein in patient fibroblasts (II.7 + Vps11) ameliorates the abnormal phenotype. * = $p < 0.05$, ** = $p < 0.02$. B) Lysosomal and plasmatic membrane enzymatic activities in patient fibroblasts (II.7) are significantly higher than in controls (CTR). GCase = β -glucocerebrosidase, Gal = β -galactosidase, Hex = β -hexosaminidase, PM = plasma membrane, ** = $p < 0.01$, **** = $p < 0.0001$. C) Cartoon model of the pathological autophagic and lysosomal functions associated with the identified *VPS11* deleterious variant. The homozygous *VPS11* loss-of-function variant causes a dysfunction of the HOPS complex leading to a defect of the fusion of lysosomes with autophagosomes. Consequently, the accumulation of abnormal lysosomal and autophagic vesicles

develops. Adapted from the template “Mutation of HOPS Complex Subunits”, by BioRender.com (2020), and from Steel D et al. *Ann Neurol* 2020.

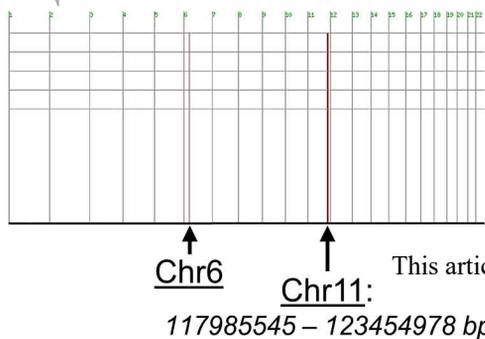
Figure 3: Electron microscopy analysis: Patient fibroblasts (D, E, F) show large clear vacuolar structures in the cytoplasm (black arrows), which are absent in control fibroblasts (A, B, C), derived from three healthy subjects. This abnormality is consistent with an alteration of the autophagic and/or endo-lysosomal compartments.



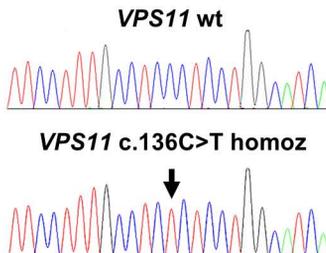
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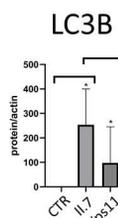
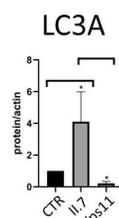
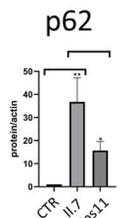
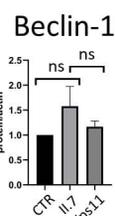
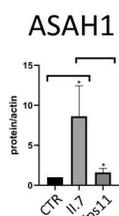
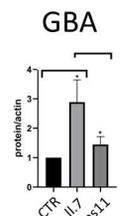
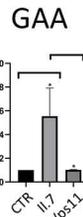
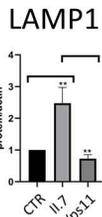
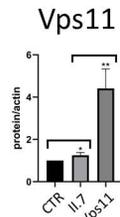
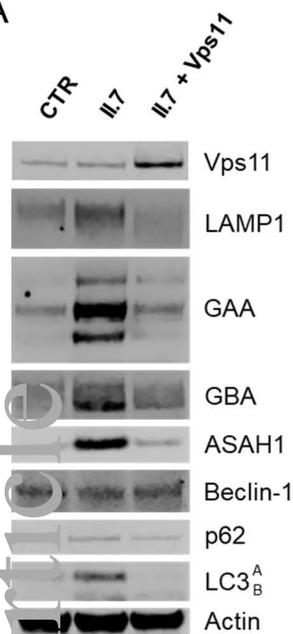


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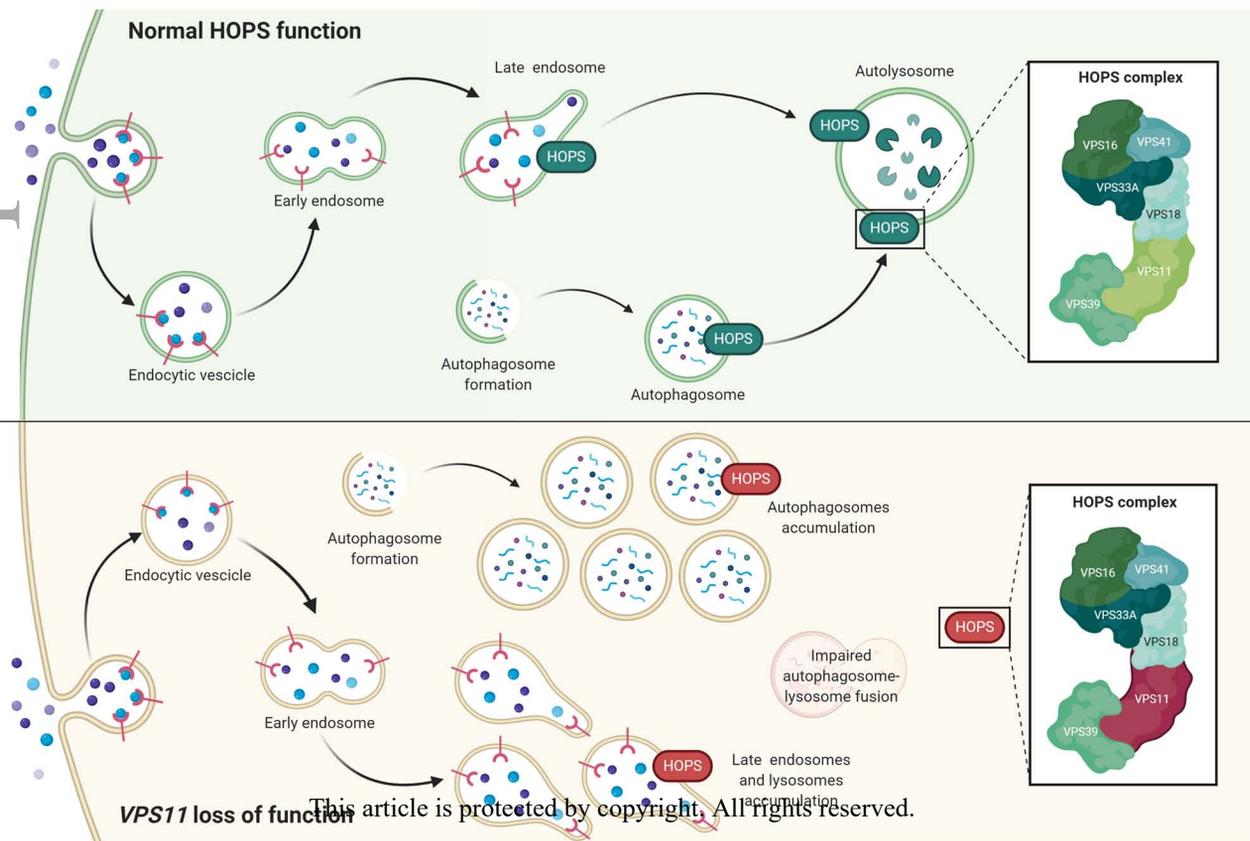
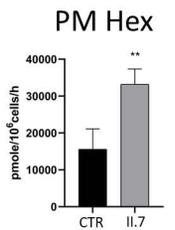
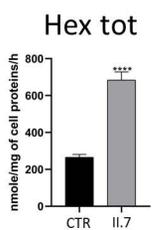
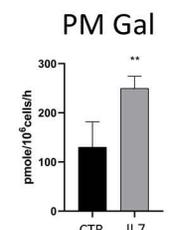
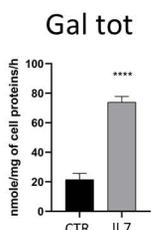
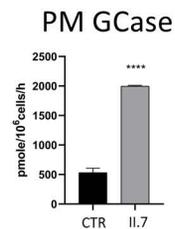
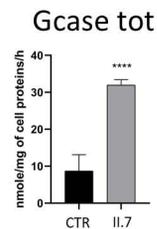
Vps11
p.P46S

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| <i>P. troglodytes</i> | ATPASGSAASKFCLCLPP-GILYQLKQHNILASV |
| <i>M. mulatta</i> | ATPASGSAASKFCLCLPP-GILYQLKQHNILASV |
| <i>C. lupus</i> | AAPASGPAASKFCLCLPP-GILYQLKQHNILASV |
| <i>B. taurus</i> | AAPASGPAASKFCLCLPP-GILYQLKQHNILASV |
| <i>M. musculus</i> | AAPVSGSAASKFCLCLPP-GILYQLKQHNILASV |
| <i>R. norvegicus</i> | -----LYQLKQHNILASV |
| <i>G. gallus</i> | -----AGGKPFALPP-GILYQLKQHSILVSV |
| | -----SENGKNFQLEM-GILYQLKQHNILVSV |

A



B



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