

From abundance-based to functional-based indicator species

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Abstract. Indicator species with high fidelity to a-priori defined groups of sites are a relevant tool to ecologically characterize plant or animal assemblages. The identification of indicator or diagnostic species is usually performed by summarizing the species abundances within each group of sites. Species with high concentration in a given group of sites are considered diagnostic of that particular group. Among the methods proposed for the determination of indicator species, only very few have considered the species functional traits. This is quite surprising, as species influence ecosystem processes via their traits. Therefore, the species functional traits should give a much better ecological characterization of a group of sites than the species abundances. The aim of this paper is thus to use the species functional characteristics to improve their diagnostic value. These characteristics include the species functional traits and all species-level indicators of environmental association. The proposed method consists of combining the species abundances and their functional characteristics into a single composite index, which can be interpreted as the species fuzzy degree of compatibility with each group of sites. The interpretation of this index in terms of fuzzy set theory allows to introduce a high degree of flexibility in the computation of the species diagnostic values. To show the behavior of the proposed index, two worked examples with data on Alpine vegetation in northern Italy and urban alien species in the city of Brussels (Belgium) are used.

KeyWords: Functional abundance; Functional centroid; Fuzzy sets; Permutation methods; Species occurrences.

1. Introduction

Indicator species with high fidelity to a particular group of sites are an important tool for the characterization of plant or animal assemblages (Tichý and Chytrý 2006). The detection of indicator or diagnostic species is typically performed by calculating the species occurrences or abundances in distinct groups of sites (or plots, relevés, sampling units, etc.). Species with high abundance concentration in a given group of sites compared to the other groups are considered diagnostic of that specific group (Tichý and Chytrý 2006; De Cáceres et al. 2010). The grouping may be based on compositional or environmental differences among sites or on other distinctive properties, such as different successional stages, land use types, or different levels of controlled experimental designs (Dufrêne and Legendre 1997; De Cáceres et al. 2012).

47 Many different approaches have been proposed to identify diagnostic species, (e.g. Dufrêne and
 48 Legendre 1997; Chytrý et al. 2002; Podani and Csányi 2010). The most widely used method for
 49 indicator species analysis is due to Dufrêne and Legendre (1997) who introduced a species-specific
 50 composite index called IndVal (Indicator Value). Given a community composition matrix containing
 51 the presence/absence or the abundance values x_{jn} of S species ($j = 1, 2, \dots, S$) in N plots ($n = 1, 2, \dots,$
 52 N), let the number of plots in group k ($k = 1, 2, \dots, G$) be N_k . IndVal is the product of two terms. The
 53 first term, which is called *specificity* (A_{jk} in the notation of Dufrêne and Legendre), is the ratio of the
 54 mean abundance of species j in group k and the sum of means of the same species over all groups.
 55 Specificity thus measures the probability that a given plot n that contains species j belongs to a target
 56 group k . Maximum specificity ($A_{jk} = 1$) is obtained if species j appears only in group k irrespective of
 57 its abundance. Minimum specificity ($A_{jk} = 0$) is obtained if species j is not contained in k .

58 The second component of IndVal is called *fidelity* (B_{jk}) and represents the number of presences of
 59 species j in group k compared to the total number of plots N_k in that group, thus measuring the
 60 probability that a given plot n that belongs to a target group k contains species j . Maximum fidelity
 61 ($B_{jk} = 1$) is obtained if species j occurs in all plots of group k , whereas minimum fidelity ($B_{jk} = 0$) is
 62 obtained if species j is not contained in k . For details, see Dufrêne and Legendre (1997).

63 Specificity and fidelity are then multiplied and scaled in the range $[0,100]$ to express the indicator
 64 value of species j for group k in terms of percentages:

$$65$$

$$66 \text{IndVal}_{jk} = A_{jk} \times B_{jk} \times 100 \quad (1)$$

$$67$$

68 Here, we assume that each of the S species is found at least in one plot, and that all N plots contain
 69 at least one species.

70 To determine the significance of the association of species j with group k we next have to compare
 71 the actual value of IndVal_{jk} with a null distribution obtained by randomly reassigning the abundances
 72 of j among the N plots. This operation generates a distribution of the test statistic under the null
 73 hypothesis that the occurrence of species j in a given plot n is due to chance alone (De Cáceres et al.
 74 2012).

75 Although indicator species analysis is generally used to ecologically characterize different groups
 76 of sites, to the best of our knowledge, only Ricotta et al. (2015) have proposed to include the
 77 functional traits of species, in addition to their occurrence and abundance, for the determination of
 78 indicator species. This is in spite of the fact that functional traits are routinely used in community
 79 ecology studies to inform on processes of community assembly (Wright et al. 2004; McGill et al.

80 2006; Adler et al. 2013; Kraft et al. 2015; Díaz et al. 2016), community responses to environmental
81 change (Voigt et al. 2007; Moretti and Legg 2009; Moles et al. 2014), and effects on ecosystem
82 functioning (Lavorel and Garnier 2002; Garnier et al. 2004; Díaz et al. 2007). Indeed, it is well known
83 that functional traits inform on the ecological strategies of species and, in turn, species influence
84 ecosystem processes via their traits (Mason and de Bello 2013). Hence, the dominant functional traits
85 in plant or animal assemblages should give a much better ecological characterization of a group of
86 sites, in terms of the local environmental conditions and ecosystem functioning, than the mere
87 occurrence of species.

88 Ricotta et al. (2015) proposed a two-step procedure to include the species functional traits in the
89 evaluation of their diagnostic values. First, the indicator species that best characterize a given group
90 of plots are identified with the usual statistical tools based either on the species incidence or
91 abundance data. Next, the functional association between the abundance-based indicator species and
92 the target group of plots is tested by calculating the functional distance between the indicator species
93 and the functional centroids of all plots in that group. A species is considered diagnostic of a given
94 group if its mean functional distance from the plot centroids of the target group is significantly lower
95 than expected.

96 According to this approach, functional indicator analysis is limited to those species that are
97 considered diagnostic of a given group of plots in terms of traditional abundance-based methods. The
98 main reason invoked by Ricotta et al. (2015) for this restriction is that indicator species are generally
99 used for the *a-posteriori* ecological typification of one or more groups of plots. Therefore, “to
100 consider a species as functionally diagnostic of a particular habitat, the species should possess a
101 reasonable chance of being detected in the field” (Ricotta et al. 2015). On the other hand, this two-
102 step procedure prevents the species without significant diagnostic capacity in terms of species
103 abundances from being tested for their functional relevance. However, it may happen that a species
104 that is not diagnostic in terms of species abundances alone shows instead a strong functional
105 association with a target group of plots.

106 The aim of this paper is thus to develop a general method to include any measurable species
107 characteristic *sensu* Garnier et al. (2017) for the characterization of their diagnostic value. These
108 characteristics may include the species traits and all species-level indicators of environmental
109 association, such as Ellenberg’s indicator values (Ellenberg et al. 1991), or Grime’s primary adaptive
110 strategies (Grime 1977; Grime and Pierce 2012). Two worked examples on Alpine vegetation in
111 northern Italy and urban alien species in the city of Brussels (Belgium) are used to show the behavior
112 of the proposed method.

113 **2. Methods**

114 A promising line of attack to include the species characteristics into indicator species analysis may
115 consist in combining the species abundances and their functional or environmental association with
116 a given group of plots into a single composite index, in a similar way to that of Dufrêne and Legendre
117 (1997). Let x_{jn} be the presence/absence (0/1) or the abundance value of species j in plot n and d_{jk} be
118 the functional dissimilarity between species j and the functional centroid of all plots in group k . If d_{jk}
119 is measured in the range $[0,1]$, this quantity can be interpreted as the fuzzy degree of functional
120 distinctness of species j with respect to group k . Consequently, the similarity s_{jk} between species j and
121 the functional centroid of group k can be simply calculated as the complement of d_{jk} : $s_{jk} = 1 - d_{jk}$.
122 The functional association Φ_{jk} between species j and group k may then be expressed as the mean
123 abundance of species j in all plots belonging to group k multiplied by s_{jk} :

124

125
$$\Phi_{jk} = \frac{\sum_{n \in k} x_{jn} \times s_{jk}}{N_k} \quad (2)$$

126

127 where s_{jk} represents the fuzzy degree of functional compatibility between species j and group k , such
128 that the product $\pi_{jn} = x_{jn} \times s_{jk}$ may be interpreted as the ‘functional abundance’ of species j in plot n
129 (i.e., the fraction of abundance of species j that is functionally compatible with plot n) and Φ_{jk} as the
130 mean ‘functional abundance’ of species j in group k . Note that for presence and absence data
131 $0 \leq \Phi_{jk} \leq 1$. In this case, maximum functional association between species j and group k is obtained
132 if j occurs in all plots of group k with $s_{jk} = 1$. The significance of the association of a given species j
133 with a target group k can then be tested with the usual permutation methods by randomly reassigning
134 the functional abundances of j among the N plots.

135

136 **3. Worked examples**

137 **3.1. Alpine vegetation on a glacier foreland in northern Italy**

138 We used data on plant communities sampled along a primary succession on the foreland of the
139 Rutor Glacier (northern Italy). The data were sampled by Caccianiga et al. (2006). The original data
140 set can be found in Ricotta et al. (2016: Appendix S2) and contains the abundances of 45 Alpine
141 species collected in 59 vegetation plots of approximately 25 m². All species abundances were
142 measured with a five-point ordinal scale transformed to ranks. Based on the age of the moraine
143 deposits, the plots were classified into three successional stages: early succession (17 plots), mid

144 succession (32 plots), and late succession (10 plots); for additional details, see Caccianiga et al.
145 (2006).

146 Six quantitative traits, which provide a good representation of the species global spectrum of form
147 and function (Díaz et al. 2016) were selected: canopy height (CH; mm), leaf dry mass content
148 (LDMC; %), leaf dry weight (LDW; mg), specific leaf area (SLA; $\text{mm}^2 \times \text{mg}^{-1}$), leaf nitrogen content
149 (LNC; %), and leaf carbon content (LCC; %). All data are freely available in Caccianiga et al. (2006,
150 pp. 16-17).

151 To explore the behavior of the selected traits along the primary succession we performed a
152 principal component analysis (PCA) on the multivariate functional centroids of each plot, defined as
153 the mean of all trait values in each plot weighted by the total abundance of each species in that plot
154 (Garnier et al. 2004). Before calculations, all traits were standardized to zero mean and unit standard
155 deviation.

156 To assess the functional association of each species with the three successional stages, we next
157 performed a functional species indicator analysis. We first calculated the multivariate functional
158 centroid of each stage (i.e., the mean of all trait values in each stage weighted by the total abundance
159 of each species in that stage). We then computed the Euclidean distance between the trait values of
160 each species and the functional centroids of each successional stage. These distances were then
161 rescaled to the unit range by dividing each species-to-centroid distance by the maximum value in the
162 dataset. Finally, we calculated the functional association Φ_{jk} of all 45 species with each of the three
163 successional stages.

164 To evaluate whether the functional association of each species with the three successional stages
165 was significantly higher than expected, we permuted the functional abundance values of each species
166 ϕ_{jk} among all 59 plots. The null hypothesis is that there is no difference in the value of Φ_{jk} among the
167 successional stages. P-values of positive functional association between a given species and each
168 group of plots were then calculated as the proportion of permutation-derived values of Φ_{jk} that are as
169 high or higher than the actual value (999 permutations, two-tailed test). All calculations were done
170 with a new R script available in Appendix 1.

171 Note that unlike in Ricotta et al. 2015, we calculated the functional abundances of each species
172 (π_{jn}) based on the mean distances from the group centroids and not from single plots. This because
173 the functional centroid of a given group of plots is likely a better indicator of the overall functioning
174 of that group compared to the centroids of the single plots in that group. Those who think it is better
175 to base the analysis on the centroids of single plots simply have to replace in Eq. (2) the functional
176 similarity s_{jk} between species j and the functional centroid of group k , with the functional similarity
177 s_{jn} between species j and the functional centroid of plot n .

178 We also ran a traditional indicator species analysis to test whether the species were significantly
179 associated with any of the three successional stages based only on presence/absence scores or on
180 abundance data. In this case, an indicator or diagnostic species is defined as a species that is more
181 common in a given group of plots than expected by chance alone. Hence, for species incidence or
182 abundance data, to assess whether a species is diagnostic of a given successional stage, the total
183 abundance (occurrence) of that species in each stage was compared with a random model in which
184 the species abundance (occurrence) values are randomly permuted within all plots (999 permutations;
185 two-tailed test), thus simulating the null condition whereby all plots have the same probability to host
186 each species, irrespective of their ecological preferences (De Cáceres and Legendre 2009; Ricotta et
187 al. 2015).

188

189 **3.2. Urban alien flora of Brussels**

190 Lososová et al. (2011) and Godefroid and Ricotta (2018) showed that different urban land use
191 types host different assemblages of alien plant species. Since alien species may represent a major
192 threat to ecosystems and biodiversity, analyzing their ecological preferences is of crucial importance,
193 particularly for urban areas where they represent a relevant portion of the local flora. Therefore, we
194 analyzed the environmental association of alien plant species for five urban land uses in the city of
195 Brussels.

196 The urban area of Brussels (161 km²; 1.2 million inhabitants) was divided in a grid of square cells
197 of 1 km². Within each cell, all spontaneous species of the vascular flora were sampled between 1992-
198 1994. Each species was then classified as alien or native according to Pyšek et al. (2004). Based on
199 the dominant land use type, each cell was associated to one of the following classes: densely built-up
200 urban areas (UD), open built-up areas (UO), urban forests (FOR), industrial areas (IND), and
201 agricultural areas (AGR). This land use classification was considered suitable for indicator species
202 analysis in the city of Brussels (see Godefroid and Ricotta 2018). Only the 159 grid cells that are
203 included in the administrative limits of the city for at least 75% of their area were used in this study.

204 To explore the response of plant species to urban soil and climatic conditions, we used the
205 Ellenberg indicator values (EIVs; Ellenberg et al. 1991) for soil nutrient availability (N), soil reaction
206 (R), soil moisture (F) and light (L). EIVs have been widely used in vegetation science for the
207 assessment of the species ecological niches (Diekmann 2003). For all EIVs, each species is given a
208 value on a 9-point ordinal scale based on expert knowledge, field observations, and partly on direct
209 measurements denoting the position at which plants reach peak abundance along environmental
210 gradients (Godefroid and Dana 2007, Bartelheimer and Poschold 2016). We used the Ellenberg
211 indicator values re-calibrated for the British Isles by Hill et al. (1999) because they are bioclimatically

212 closer to the study area compared to the original values estimated for Central European conditions.
213 First, the multivariate EIV centroid of each land use class was calculated as the weighted mean of the
214 four Ellenberg indicator values of all species (native and alien) present in that land use class
215 (Ellenberg et al. 1991). We next calculated the Euclidean distance between the indicator values of
216 each alien species and the centroids of each land use type. Only neophyte species introduced after
217 AD1500 with available EIVs (88 species) were considered in this study. Like in the previous example,
218 the Euclidean distances were linearly rescaled to the unit range by dividing each distance by the
219 maximum value in the dataset. By combining the species presence/absence scores with the species-
220 to-centroid distances, we calculated the functional association Φ_{jk} of the alien species with the urban
221 land uses. Finally, using indicator species analysis (999 permutations; two-tailed test), we identified
222 the alien species that best characterize each land use type in terms of both presence/absence scores
223 and functional association values.

224

225 **4. Results**

226 **4.1. Alpine vegetation on a glacier foreland**

227 The principal component analysis of the 59 vegetation plots sampled on the foreland of the Rutor
228 Glacier (Fig. 1) shows that the three successional stages are functionally well distinct in ordination
229 space. Along the primary succession, a significant increase of leaf dry weight and leaf carbon content
230 is observed together with a decrease of specific leaf area and leaf nitrogen content.

231 Using traditional indicator species analysis on presence/absence scores, we found 24 species
232 showing significant association ($p < 0.05$, two-tailed test) with one of the three successional stages
233 (Table 1). If the same analysis is performed on species abundances, the number of indicator species
234 increased to 28. Finally, if the indicator species analysis is performed on the functional association
235 Φ_{jk} of the alpine species with the three successional stages (thus considering both the species
236 abundances *and* their functioning), the number of indicator species increased to 31.

237 Apart from *Veronica bellidioides*, all species that were identified as diagnostic of one or more
238 successional stages in terms of presence/absence scores are also diagnostic of the same successional
239 stages in terms of species abundances (see Table 1). Likewise, all abundance-based diagnostic species
240 represent a subset of the functionally-based diagnostic species. Therefore, traditional indicator
241 species analysis and functional indicator species analysis are not in contrast with each other. For some
242 species, occurrence-based or distance-based analysis is powerful enough to highlight their diagnostic
243 values. For some other species, we also need to consider their functional traits. In this sense, at least
244 in our case, functional indicator species analysis enables to highlight the diagnostic value of a larger
245 pool of species.

246 4.2. Urban alien flora of Brussels

247 Godefroid and Ricotta (2018) highlighted a significant relationship between land use composition
248 and EIVs, such that 1 km² cells with similar land use composition tend to be similar also in terms of
249 Ellenberg indicator values. Compared to less human impacted land uses, densely urbanized areas
250 have on average lower EIVs for soil moisture and higher EIVs for light, soil reaction (pH) and
251 nitrogen availability.

252 Traditional indicator species analysis on presence/absence scores identified 24 alien species
253 showing significant association ($p < 0.05$, two-tailed test) with certain land use classes (Table 2).
254 Therefore, it seems that a relevant portion of alien species has rather narrow ecological requirements
255 allowing them to differentiate between more or less impacted land use types (for details, see
256 Godefroid and Ricotta 2018). The largest number of diagnostic species is associated to open built-up
257 areas (13 species), whereas for presence/absence scores, only one single species, *Sisymbrium*
258 *altissimum*, is diagnostic of the industrial areas.

259 If the indicator species analysis is performed on the functional association Φ_{jk} of the alien species
260 with the urban land uses (taking into account both the species presence/absence scores *and* their
261 EIVs), the number of species that are significantly associated with at least one land use class increases
262 to 35. As in the previous example, all species that were diagnostic of one or more land use class in
263 terms of presence/absence scores are also diagnostic of the same land use classes in terms of EIVs.

264

265 5. Discussion

266 Plant functional traits have been increasingly used to explore patterns of co-occurrence in plant
267 communities (e.g. McGill et al. 2006, Adler et al. 2013, Nathan et al. 2015), but this approach has
268 been overlooked in indicator species analyses. Although diagnostic species are generally used as
269 ecological indicators of community or habitat types (De Cáceres et al. 2010), most of the methods
270 used for their determination do not take into account their functional traits. In this paper we thus
271 developed a method for indicator species analysis which combines the species abundances with their
272 functional traits.

273 Tested on an urban alien flora, this method has demonstrated its effectiveness. First, by allowing
274 more indicator species to be detected (35 instead of 24 with the traditional method). Second, because
275 the 11 newly identified indicators make ecological sense. Among these, there are for example several
276 species escaped from gardens, such as *Asparagus officinalis*, *Aster lanceolatum* or *Syringa vulgaris*,
277 logically identified as indicators of open built-up areas. Also, species that are typically recorded in
278 highly urbanized areas (e.g. wasteland and pavements), like *Coronopus didymus* and *Hirschfeldia*

279 *incana*, were rightly detected as indicators of densely built-up areas, while the classical approach was
280 unable to identify them as such.

281 Likewise, the early-successional stage of the Rutor glacier foreland is comprised of fast-growing
282 species with strongly ruderal characteristics *sensu* Grime (1977), which highlight the influence of
283 disturbance on pioneer communities. By contrast, the mid and late-successional stages are
284 progressively characterized by an increasing number of stress-tolerator species that are adapted to
285 substantial periods of low temperature (Caccianiga et al. 2006). As shown in Figure 1, this shift from
286 ruderal to stress-tolerator communities along the primary succession correlates strongly with changes
287 in resource-use traits, such as increasing leaf dry matter content (LDMC) and decreasing specific leaf
288 area (SLA). In this view, the three newly identified indicators, *Agrostis rupestris*, *Luzula spicata* and
289 *Veronica bellidioides* are all stress-tolerator species with high LDMC and low SLA (see Caccianiga
290 et al. 2006, Table 2). Therefore, while the usual abundance-based approach was unable to identify
291 them as diagnostic, their association with the mid-successional stage (*Agrostis rupestris* and *Luzula*
292 *spicata*) and the late-successional stage (*Veronica bellidioides*) is supported by their functional
293 characteristics.

294 Like in traditional indicator species analysis which randomizes the species occurrences or
295 abundances among plots, functional indicator species analysis randomizes the functional abundances
296 π_{jn} of species among plots. As a result, there is no ecological inconsistency between the abundance-
297 based and functional-based methods for identifying diagnostic species. If the functional
298 characteristics of a target species are strongly correlated with specific ecosystem properties such that
299 the species is exclusive or highly preferential of a given community, occurrence-based analysis is
300 powerful enough to highlight its diagnostic value. Some more ubiquitous species show significant
301 differences among communities in terms of their abundances, while for a third group of species, a
302 functional fine-tuning to the specific biotic and abiotic conditions of their habitat is needed to
303 highlight their diagnostic value. Therefore, by applying a ‘cascade’ of different methods, we can
304 identify step by step groups of species with different diagnostic power, thus improving the flexibility
305 of the species association with a target group of plots.

306 Like for any other ecological indicator that is based on the analysis of the functional traits of
307 species, the most relevant decisions to be taken include which traits to use, the choice of the
308 dissimilarity coefficient and how to calculate the functional centroid (Anderson 2006, Lavorel et al.
309 2008). Note that a relevant aspect of the proposed method is that for the calculation of the multivariate
310 functional centroid of a target group of plots, abundant species contribute more than rare species. The
311 biological justification for this differential weighting of rare and abundant species is that the extent
312 to which plant species affect a wide variety of ecosystem functions, such as carbon balance or nutrient

313 dynamics, is largely determined by their abundance. This ‘mass-ratio’ effect is determined by the
 314 observation that, at least for plants, a larger body mass implies major contribution to syntheses,
 315 resource fluxes and degradative processes (see Grime 1998). Alternative methods to calculate the
 316 functional centroid of a target group of plots may consist in weighting all species equally, irrespective
 317 of their abundance, or in iteratively excluding the target species from the calculation of the
 318 multivariate centroid by means of leave-one out methods.

319 Finally note that, from a more technical viewpoint, the interpretation of the functional dissimilarity
 320 d_{jk} as a fuzzy degree of functional distinctness between a given species j and a target group of plots k
 321 opens the way for introducing a high degree of flexibility in the computation of the species functional
 322 abundances $\pi_{jn} = x_{jn} \times s_{jk}$.

323 Fuzzy set theory has been already described elsewhere (Klir and Yuan 1995; Klir and Wierman
 324 1999), and the reader is referred to these papers for details. For our purposes, it is sufficient to observe
 325 that if we define the quantity d_{jk} in terms of fuzzy set theory, then functional similarity becomes its
 326 fuzzy complement $s_{jk} = C(d_{jk})$. Based on this fuzzy characterization of the relationship between d_{jk}
 327 and s_{jk} , we can use a wide range of generalized complement operators for defining a biologically
 328 meaningful measure of fuzzy compatibility between a given species and a target group of plots.

329 Besides the standard fuzzy complement operation $C(d_{jk}) = 1 - d_{jk}$ there exists a broad class of
 330 functions $C(d_{jk}): [0,1] \rightarrow [0,1]$ that allow to assign a value $C(d_{jk})$ in the range $[0,1]$ to each
 331 dissimilarity value d_{jk} . These functions need to conform to a set of requirements that make them
 332 suitable as fuzzy generalizations of the standard complement operation (see Klir and Wierman 1999).
 333 The resulting values can then be used to calculate the species functional abundances in each plot as
 334 $\pi_{jn} = x_{jn} \times C(d_{jk})$. For example, a fuzzy complement operation that might be used in the context of
 335 indicator species analysis is the threshold function (Klir and Yuan 1995). For a threshold value t in
 336 the range $[0,1]$, this function is defined as:

337

$$338 \quad C(d_{jk}) = \begin{cases} 1 & \text{for } d_{jk} < t \\ 0 & \text{for } d_{jk} \geq t \end{cases} \quad (3)$$

339

340 According to Eq. (3), the species occurrences or abundances in a given group of plots are
 341 considered only if their functional dissimilarity to the functional centroid of that group of plots is
 342 below an a-priori defined threshold.

343 Two additional complement operators that are commonly used in multivariate analysis for
344 computing a measure of similarity in the range $[0,1]$ from the corresponding distances are:

345 $C(d_{jk}) = \sqrt{1-d_{jk}}$ or $C(d_{jk}) = \sqrt{1-d_{jk}^2}$ (see Legendre and Legendre 2012). All these functions
346 enable the practitioner to change the sensitivity of the complement operator to high or low values of
347 d_{jk} according to its specific requirements. For additional aspects on the ecological applications of
348 fuzzy complement functions, see Ricotta (2008).

349 Overall, we see the flexibility associated to fuzzy complement operators as a great potential
350 advantage for ecologists as it allows to compute relevant aspects of the species diagnostic power from
351 different viewpoints and motivations. Therefore, we hope that the use of functional indicator species
352 analysis will help improve the ecological characterization of plant and animal assemblages.

353

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355 **References**

- 356 Adler, P.B., Fajardo, A., Kleinhesselink, A.R., Kraft, N.J.B. (2013) Trait-based tests of co-existence
357 mechanisms. *Ecology Letters* 16: 1294–1306.
- 358 Anderson, M.J. (2006) Distance-based tests for homogeneity of multivariate dispersions. *Biometrics*
359 62: 245–253.
- 360 Bartelheimer, M., Poschold, P. (2016) Functional characterizations of Ellenberg indicator values – a
361 review on ecophysiological determinants. *Functional Ecology* 30: 506–516.
- 362 Caccianiga, M., Luzzaro, A., Pierce, S., Ceriani, R.M., Cerabolini, B. (2006) The functional basis of
363 a primary succession resolved by CSR classification. *Oikos* 112: 10–20.
- 364 Chytrý, M., Tichý, L., Holt, J., Botta-Dukát, Z. (2002) Determination of diagnostic species with
365 statistical fidelity measures. *Journal of Vegetation Science* 13: 79–90.
- 366 Díaz, S., Kattge, J., Cornelissen, J.H.C., Wright, I.J., Lavorel, S. et al. (2016) The global spectrum of
367 plant form and function. *Nature* 529: 167–171.
- 368 Díaz, S., Lavorel, S., de Bello, F., Quétier, F., Grigulis, K., Robson, T.M. (2007) Incorporating plant
369 functional diversity effects in ecosystem service assessments. *Proceedings of the National*
370 *Academy of Sciences* 104: 20684–20689.
- 371 De Cáceres, M., Legendre, P. (2009) Associations between species and groups of sites: indices and
372 statistical inference. *Ecology* 90: 3566–3574.
- 373 De Cáceres, M., Legendre, P., Moretti, M. (2010) Improving indicator species analysis by combining
374 groups of sites. *Oikos* 119: 1674–1684.
- 375 De Cáceres, M., Legendre, P., Wisser, S.K., Brotons, L. (2012) Using species combinations in
376 indicator value analyses. *Methods in Ecology and Evolution* 3: 973–982.
- 377 Diekmann, M. (2003) Species indicator values as an important tool in applied plant ecology - a
378 review. *Basic and Applied Ecology* 4: 493–506.
- 379 Dufrêne, M., Legendre, P. (1997) Species assemblages and indicator species: the need for a flexible
380 asymmetrical approach. *Ecological Monographs* 67: 345–366.
- 381 Ellenberg, H., Weber, H.E., Düll, R., Wirth, V., Werner, W., Paulissen, D. (1991) Zeigerwerte von
382 Pflanzen in Mitteleuropa. *Scripta Geobotanica* 18: 1–48.
- 383 Garnier, E., Cortez, J., Billes, G., Navas, M.L., Roumet, C., Debussche, M., Laurent, G., Blanchard,
384 A., Aubry, D., Bellmann, A., Neill, C., Toussaint, J.P. (2004) Plant functional markers capture
385 ecosystem properties during secondary succession. *Ecology* 85: 2630–2637.

- 386 Garnier, E., Stahl, U., Laporte, M.-A., Kattge, J., Mougnot, I., et al. 2017. Towards a thesaurus of
387 plant characteristics: an ecological contribution. *Journal of Ecology* 105: 298–309.
- 388 Godefroid, S., Dana, E.D. (2007) Can Ellenberg's indicator values for Mediterranean plants be used
389 outside their region of definition? *Journal of Biogeography* 34: 62–68.
- 390 Godefroid, S., Ricotta, C. (2018) Alien plant species do have a clear preference for different land uses
391 within urban environments, *Urban Ecosystems* 21: 1189–1198.
- 392 Grime, J.P. (1977) Evidence for the existence of three primary strategies in plants and its relevance
393 to ecological and evolutionary theory. *American Naturalist* 111: 1169–1194.
- 394 Grime, J.P. (1998) Benefits of plant diversity to ecosystems: immediate, filter and founder effects.
395 *Journal of Ecology* 86: 902–910.
- 396 Grime, J.P., Pierce, S. (2012) *The evolutionary strategies that shape ecosystems*. Wiley-Blackwell,
397 Chichester, UK.
- 398 Hill, M.O., Mountford, J.O., Roy, D.B., Bunce, R.G.H. (1999) Ellenberg's indicator values for British
399 plants. ECOFACT Vol. 2, Technical Annex. Institute of Terrestrial Ecology, Huntingdon, UK.
- 400 Klir, G., Yuan, B. (1995) *Fuzzy Sets and Fuzzy Logic: Theory and Applications*. Prentice Hall, Upper
401 Saddle River, New Jersey.
- 402 Klir, G.J., Wierman, M.J. (1999) *Uncertainty-Based Information*. Physica-Verlag, Heidelberg.
- 403 Kraft, N.J.B., Godoy, O., Levine, J.M. (2015) Plant functional traits and the multidimensional nature
404 of species coexistence. *Proceedings of the National Academy of Sciences of the USA* 12 : 797-
405 802.
- 406 Lambinon, J., Delvosalle, L., Duvigneaud, J. (2012) *Nouvelle Flore de la Belgique, du Grand-Duché
407 de Luxembourg, du Nord de la France et des Régions voisines*. Jardin botanique national de
408 Belgique, Meise.
- 409 Lavorel, S., Grigulis, K., McIntyre, S., Williams, N.S.G., Garden, D., Dorrough, J., Berman, S.,
410 Quéfier, F., Thébault, A., Bonis, A. (2008) Assessing functional diversity in the field –
411 methodology matters! *Functional Ecology* 22: 134–147.
- 412 Lavorel, S., Garnier, E. (2002) Predicting changes in community composition and ecosystem
413 functioning from plant traits: revisiting the Holy Grail. *Functional ecology* 16: 545–556.
- 414 Legendre, P., Legendre, L. (2012) *Numerical Ecology*. Elsevier, Amsterdam.
- 415 Lososová, Z., Horsák, M., Chytrý, M., Čejka, T., Danihelka, J., Fajmon, K., Hájek, O., Juříčková, L.,
416 Kintrová, K., Láníková, D., Otýpková, Z., Řehořek, V., Tichý, L. (2011) Diversity of Central
417 European urban biota: effects of human-made habitat types on plants and land snails. *Journal of
418 Biogeography* 38: 1152–1163.
- 419 Mason, N.W.H., de Bello, F. (2013) Functional diversity: a tool for answering challenging ecological
420 questions. *Journal of Vegetation Science* 24: 777–780.
- 421 McGill, B.J., Enquist, B.J., Weiher, E., Westoby, M. (2006) Rebuilding community ecology from
422 functional traits. *Trends in Ecology and Evolution* 21: 178–185.
- 423 Moles, A.T., Perkins, S.E., Laffan, S.W., Flores-Moreno, H., Awasthy, M. et al. (2014). Which is a
424 better predictor of plant traits: temperature or precipitation?. *Journal of Vegetation Science* 25:
425 1167–1180.
- 426 Moretti, M., Legg, C. (2009) Combining plant and animal traits to assess community functional
427 responses to disturbance. *Ecography* 32: 299–309.
- 428 Pignatti, S. (1982) *Flora d'Italia*. Edagricole, Bologna.
- 429 Podani, J., Csányi, B. (2010) Detecting indicator species: some extensions of the IndVal measure.
430 *Ecological Indicators* 10: 1119–1124.
- 431 Pyšek, P., Richardson, D.M., Rejmánek, M., Webster, G., Williamson, M., Kirschner, J. (2004) Alien
432 plants in checklists and floras: towards better communication between taxonomists and ecologists.
433 *Taxon* 53:131–143.
- 434 Ricotta, C., Carboni, M., Acosta, A.T.R. (2015) Let the concept of indicator species be functional!
435 *Journal of Vegetation Science* 26: 839–847.

- 436 Ricotta, C., de Bello, F., Moretti, M., Caccianiga, M., Cerabolini, B., Pavoine, S. (2016) Measuring
437 the functional redundancy of biological communities: a quantitative guide. *Methods in Ecology*
438 and *Evolution* 7: 1386–1395.
- 439 Ricotta, C. (2008) Computing additive -diversity from presence and absence scores: A critique and
440 alternative parameters. *Theoretical Population Biology* 73: 244–249.
- 441 Tichý, L., Chytrý, M. (2006) Statistical determination of diagnostic species for site groups of unequal
442 size. *Journal of Vegetation Science* 17: 809–818.
- 443 Voigt, W., Perner, J., Hefin Jones, T. (2007) Using functional groups to investigate community
444 response to environmental changes: two grassland case studies. *Global Change Biology* 13: 1710–
445 1721.
- 446 Wright, I.J., Reich, P.B., Westoby, M., Ackerly, D.D., Baruch, Z. et al. (2004) The worldwide leaf
447 economics spectrum. *Nature* 428: 821–827.

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451 **Supplementary Materials**

452 **Appendix 1.** R script for the calculation of functional indicator species.

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455 **Authors Contribution statement.** C.R. conceived the idea; All authors developed the methodology and
456 analyzed the data; M.CAC. and B.C. collected the data for Italy and S.G. for Belgium; C.R. took the
457 lead in writing the main text and M.CAR. in writing the R script. All authors revised the manuscript
458 critically and approved the final version.

459

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461

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463 **Table 1.** Alpine species with significant diagnostic power for the three successional stages on the
 464 glacier foreland in terms of presence/absence scores (P/A), species abundances (AB) and functional
 465 association values Φ_{jk} ($p < 0.05$, 999 randomizations, two-tailed test). The functional association
 466 values are obtained by combining the species abundances with their functional traits. Nomenclature
 467 according to Pignatti (1982).
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Indicator Species	Early Succession			Mid Succession			Late Succession		
	P/A	AB	Φ_{jk}	P/A	AB	Φ_{jk}	P/A	AB	Φ_{jk}
<i>Achillea moschata</i> Wulfen				x	x	x			
<i>Adenostyles leucophylla</i> (Willd.) Rchb.	x	x	x						
<i>Agrostis rupestris</i> All.						x			
<i>Anthoxanthum odoratum</i> L.				x	x	x			
<i>Arabis alpina</i> L.	x	x	x						
<i>Avenula versicolor</i> (Vill.) M. Láinz							x	x	x
<i>Carex curvula</i> All.							x	x	x
<i>Carex sempervirens</i> Vill.							x	x	x
<i>Cerastium uniflorum</i> Clairv.	x	x	x						
<i>Erigeron uniflorus</i> L.				x	x	x			
<i>Festuca halleri</i> All.					x	x			
<i>Gnaphalium supinum</i> L.	x	x	x						
<i>Hieracium angustifolium</i> Hoppe				x	x	x			
<i>Homogyne alpina</i> (L.) Cass.							x	x	x
<i>Juncus trifidus</i> L.							x	x	x
<i>Leucanthemopsis alpina</i> (L.) Heywood	x	x	x						
<i>Linaria alpina</i> (L.) Mill.	x	x	x						
<i>Luzula lutea</i> (All.) DC.							x	x	x
<i>Luzula spicata</i> (L.) DC.						x			
<i>Minuartia recurva</i> (All.) Schinz & Thell.				x	x	x			
<i>Myosotis alpestris</i> F. W. Schmidt				x	x	x			
<i>Oxyria digyna</i> (L.) Hill	x	x	x						
<i>Phleum rhaeticum</i> (Humphries) Rauschert		x	x						
<i>Poa alpina</i> L.				x	x	x			
<i>Saxifraga aizoides</i> L.	x	x	x						
<i>Saxifraga bryoides</i> L.					x	x			
<i>Silene acaulis</i> (L.) Jacq.				x	x	x			
<i>Trifolium badium</i> Schreb.		x	x						
<i>Trifolium pallescens</i> Schreb.				x	x	x			
<i>Tussilago farfara</i> L.	x	x	x						
<i>Veronica bellidioides</i> L.							x		x

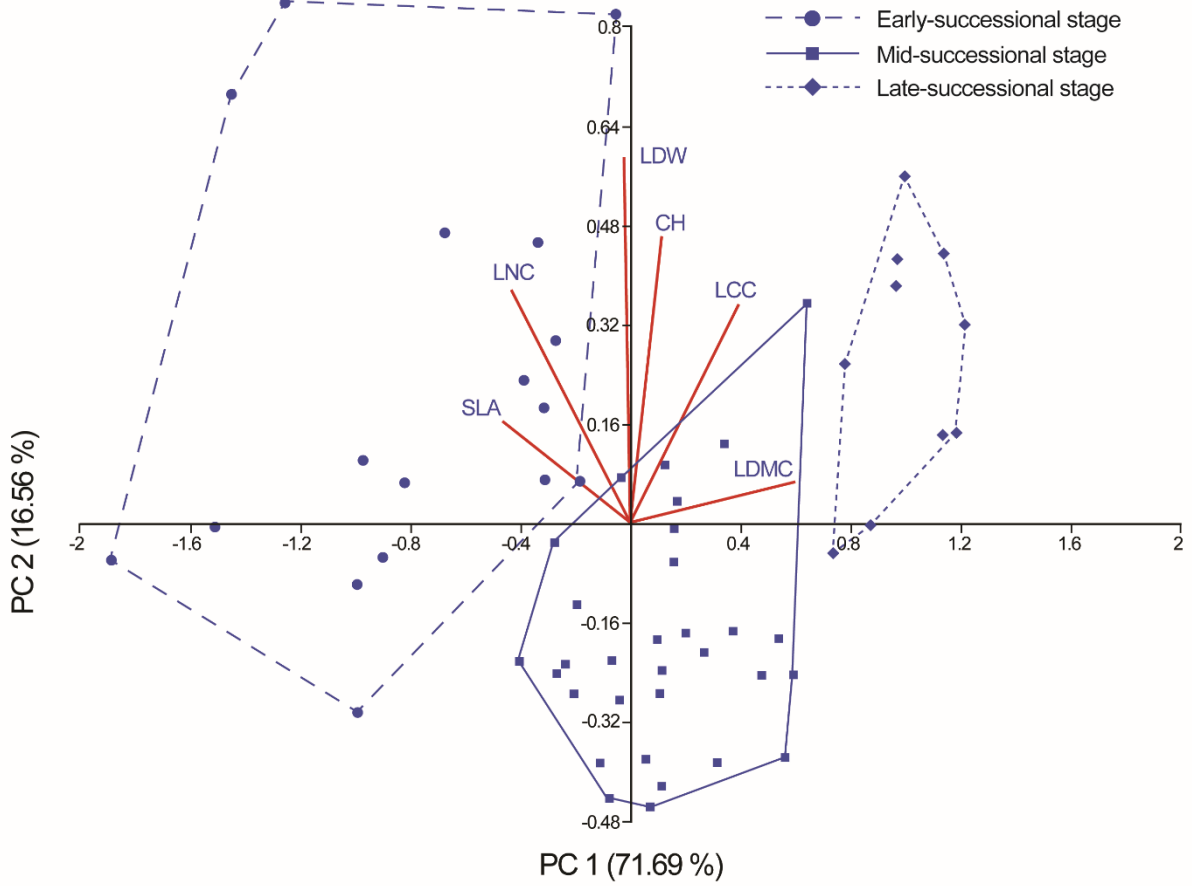
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472 **Table 2.** Alien species with significant diagnostic power for the selected urban land use classes of
 473 Brussels in terms of presence/absence scores (P/A) and functional association values Φ_{jk} ($p < 0.05$,
 474 999 randomizations, two-tailed test). UD = densely built-up urban areas, UO = open built-up areas,
 475 FOR = urban forests, IND = industrial areas, AGR = agricultural areas. Nomenclature according to
 476 Lambinon et al. (2012).
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Indicator Species	UD		UO		FOR		IND		AGR	
	P/A	Φ_{jk}	P/A	Φ_{jk}	P/A	Φ_{jk}	P/A	Φ_{jk}	P/A	Φ_{jk}
<i>Acer platanoides</i> L.			x	x						
<i>Acer pseudoplatanus</i> L.				x		x				
<i>Allium schoenoprasum</i> L.			x	x						
<i>Alnus incana</i> (L.) Moench					x	x				
<i>Asparagus officinalis</i> L.				x						
<i>Aster lanceolatus</i> Willd.				x						
<i>Barbarea intermedia</i> Boreau				x						
<i>Buddleja davidii</i> Franch.	x	x						x		
<i>Castanea sativa</i> Mill.			x	x	x	x				
<i>Coronopus didymus</i> (L.) Smith		x								
<i>Cymbalaria muralis</i> P. Gaertn., B. Mey. et Scherb.			x	x						
<i>Fallopia japonica</i> (Houtt.) Ronse Decraene				x						
<i>Fallopia sachalinensis</i> (F. Schmidt Petrop.) Ronse Decraene			x	x						
<i>Galinsoga quadriradiata</i> Ruiz et Pav.	x	x								
<i>Hirschfeldia incana</i> (L.) Lagrèze-Fossat		x								
<i>Impatiens glandulifera</i> Royle				x						x
<i>Impatiens parviflora</i> DC.					x	x				
<i>Juncus tenuis</i> Willd.					x	x				
<i>Ligustrum ovalifolium</i> Hassk.									x	x
<i>Mahonia aquifolium</i> (Pursh) Nutt.	x	x								
<i>Matricaria discoidea</i> DC.	x	x								
<i>Phalaris canariensis</i> L.	x	x								
<i>Pseudofumaria lutea</i> (L.) Borkh.			x	x						
<i>Robinia pseudoacacia</i> L.			x	x						
<i>Saponaria officinalis</i> L.								x		
<i>Sisymbrium altissimum</i> L.							x	x		
<i>Solidago canadensis</i> L.			x	x						
<i>Solidago gigantea</i> Ait.			x	x						
<i>Symphoricarpos albus</i> (L.) S.F. Blake			x	x						x
<i>Syringa vulgaris</i> L.				x						
<i>Taxus baccata</i> L.			x	x		x				
<i>Trifolium hybridum</i> L.									x	x
<i>Veronica filiformis</i> Smith			x	x						
<i>Veronica persica</i> Poiret			x	x					x	x
<i>Vinca major</i> L.										x

480

481 **Figure 1.**
 482 Biplot of the principal component analysis of the 59 plots of Alpine vegetation with the convex hulls
 483 of the three stages identified along the primary succession. The amount of variance explained by the
 484 first two axes is shown in brackets. CH = canopy height, LDMC = leaf dry mass content, LDW = leaf
 485 dry weight, SLA = specific leaf area, LNC = leaf nitrogen content, LCC = leaf carbon content.
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