

The multi-site *PeachRefPop* collection: a true cultural heritage and an invaluable international scientific tool for fruit trees

One sentence summary: design and realization of the *PeachRefPop*, the first international multi-site reference collection in peach (*P. persica*): an invaluable tool for scientific studies in perennial species

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Plant material: DB, SF, BQ, TB, TP, PA, MJA, IV, AL, DG; host institutions: DB, MC, SF, LR, PA, IE, JT, PD, IV; phytosanitary controls: AB; site characterization: WG; experimental design: MCAMB; phenotypic data analysis: AL, DG, CC; genotypic data analysis: IP, AC, MT, MJA, SM; core collection design and evaluation: SM, IV, MC; manuscript writing: MC, SM; manuscript revision: DB, LR, IV, SM and all authors for respective parts. Conception and coordination: LR and DB.

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Abstract

As sessile organisms, plants evolved a range of adaptive mechanisms to adjust their development and physiology to variable external conditions, particularly in perennial species subjected to long-term interplay with the environment. Exploiting the allelic diversity within available germplasm and leveraging the knowledge of the mechanisms regulating genotype interaction with the environment are crucial to address climatic challenges and assist the breeding of novel cultivars with improved resilience. The development of multisite collections is of utmost importance for the conservation and utilization of genetic materials and will greatly facilitate the dissection of genotype-by-environment interaction. Such resources are still lacking for perennial trees, facing with the intrinsic difficulties of successful propagations, materials exchange and living collection maintenance. This work describes the concept, design and realization of the first multi-site peach reference collection (*PeachRefPop: PRP*) located across different European countries and sharing the same experimental design. Other than an invaluable tool for scientific studies in perennial species, the *PRP* is configuring as the first milestone of an international collaborative project for the conservation and exploitation of European peach germplasm resources and, ultimately, as a true heritage for future generations.

1 Introduction

2 Since the Roman garden '*hortus*', fruit tree orchards have represented distinctive features of
3 the Mediterranean rural landscape, a synthesis of the interaction among genotype, environment and
4 human customs (**Biasi et al., 2009**). The diversity of pedo-climatic conditions and production
5 systems, along with plasticity of the genotype and human traditions has shaped the selection of a
6 multitude of local cultivars. These materials represent a cultural and genetic heritage of generations
7 of farmers and a 'common good' to preserve for present and future generations.

8 As sessile organisms, plants evolved a range of adaptive mechanisms to adjust their
9 development and physiology to variable external conditions, particularly in perennial species,
10 subjected to a long-term environmental exposure and interaction. Climate changes are impacting
11 cultivation environments, raising the need for more resilient cultivars able to maintain performances
12 across variable (and often unpredictable) weather conditions (**Varshney et al., 2011; Luedeling,**

13 **2012; Ramírez and Kallarackal, 2015**). Also, increasing the sustainability of fruit production
14 (particularly in terms of resource demands and disease management) requires leveraging
15 knowledge of the interactions between plants, soil, and environmental factors and how they affect
16 productivity and end-product quality (**Coakley et al., 1999; Singh et al., 2013; Parajuli et al., 2018**).
17 Peach [*Prunus persica* L. (Batsch)] originated in China (**Li et al., 2019**), later reaching Persia, the
18 Mediterranean Basin, Europe and the Americas, is now the third most cultivated fruit tree species in
19 temperate regions. Beside its importance as a crop, peach is a recognized model for genetic and
20 genomic studies in fruit trees, representing the ideal system for addressing two main challenges in
21 fruit tree breeding:

22 1) understanding and harnessing the allelic diversity within available gene pools; noteworthy for
23 peach, the intercompatibility with related species of the *Amygdalus* subgenus (almond, *P.*
24 *daurica*, *P. kansuensis*, etc.) has been long considered a source of natural variability,
25 particularly for the introgression of disease resistances (**Gradziel, 2002; Foulongne et al.,**
26 **2003**). However, interspecific hybrids have had poor applicability in current breeding programs
27 (**Cirilli et al., 2017**), although new genomic based strategies could change this trend (**Serra et**
28 **al., 2016**). Conversely, landraces and local ecotypes could be a source of resilience traits more
29 straightforward to introgress, making their preservation and exploitation a suitable strategy for
30 dealing with the changing climatic conditions.

31 2) systematic dissection of genotype-by-environment ($G \times E$) and/or by-management ($G \times E \times$
32 M) interactions as primary sources of variability for several important quantitative traits (**Bassi**
33 **et al., 2006; Myles, 2013; Chagné et al., 2014**). This is a critical point for genetic analyses of
34 complex traits, such as genome-wide association studies (GWAS) where germplasm collections
35 are characterized to identify quantitative trait loci (QTLs) across different environments, or
36 genome-wide selection (GS), used to predict genomic estimated breeding values.

37 The comprehension of genetic, epigenetic and physiological mechanisms as well as the estimation
38 of $G \times E$ and/or $G \times E \times M$ effects requires the development of multisite replicated collections and *ad*
39 *hoc* experimental designs. The availability of such type of resources is rapidly growing in annual
40 species, while it has not yet been implemented in perennial trees.

41 During the past century, peach orchard systems have changed dramatically following
42 innovations in orchard design, training systems and agronomic management (**Fideghelli et al.,**
43 **1988; Corelli-Grappadelli and Marini, 2008**), other than cultivar evolution. Noteworthy, the first
44 reported 'modern' orchard was a peach plantation established in Massa Lombarda (Ravenna, Italy)
45 at the end of the 19th century using the white fleshed local cultivar-population 'Buco Incavato'
46 (**Bellucci, 1908**). In the last decades, considerable breeding efforts have assisted the intensification
47 of cultivation techniques and the development of horticultural quality concepts with the introduction
48 of novel, fit-for-purpose cultivars (**Byrne et al., 2009**). In Europe, peach has a long cultivation history,
49 tracing back to the Ancient and Middle ages, and characterized by the isolation and propagation of

50 chance seedlings operated by farmers, through which each country has set its own pool of locally
51 adapted cultivars (**Bassi and Layne, 2009**). The paradigm shift to the modern controlled-crosses
52 approach in early US breeding programs has been the foundation of the dramatic varietal
53 improvement of the last century, beginning with the introduction of seedling materials from China in
54 the mid-19th century (e.g. 'Chinese Cling', progenitor of most modern cultivars) (**Faust and Timon,**
55 **1995; Byrne et al., 2009**). The worldwide spread of improved US materials, favored also by the
56 limited activities in other countries, has resulted in a rapid replacement of landraces and local
57 accessions, particularly in Europe. From the second half of the 20th century, however, novel
58 programs started in several European countries, although they were mostly based on US breeding
59 stocks with a marginal role for local cultivated germplasm. This led to a consequent loss of many
60 local cultivars, in parallel with a progressive narrowing of the genetic bases in modern cultivars
61 (**Aranzana et al., 2010; Verde et al., 2013**).

62 As awareness of genetic erosion in modern plant breeding increased (**Fu and Dong, 2015**)
63 the conservation and exploitation of genetic resources has become a fundamental aspect in crop
64 breeding (**Ford-Lloyd and Jackson, 1986**). Considerable efforts have been made in the collection
65 and characterization of many plant germplasms (including fruit tree species), along with the
66 development of approaches for their effective management and utilization (**Gepts, 2006**). The
67 concept of 'core collection', a subset of a germplasm collection of a species that captures most of
68 the genetic diversity while reducing redundancy, has represented an ideal solution for reducing costs
69 and increasing the efficiency of conservation programs (**Frankel and Brown, 1984**). Several
70 allocation methods have been developed for selecting core collections, attempting to maximize allelic
71 richness or allele coverage (MSTRAT, PowerCore, GenoCore), minimize or maximize genetic
72 distance (GDOpt, SimEli) or simultaneously accommodating for multiple criteria (Core Hunter)
73 (**Gouesnard et al., 2001, Kim et al., 2003; Thachuk et al., 2009; Odong et al., 2011; Krishnan et**
74 **al., 2014**). However, the effectiveness of the sampling strategies varied depending on the objective
75 of the core collection, the statistical approach for its definition and the measures for evaluating its
76 quality (**Odong et al., 2013**). Furthermore, beyond statistical considerations, other aspects are often
77 considered by the institutions hosting the collection, such as historical and socio-economic
78 importance, relevance for breeding activities, popularity among growers and consumers, or
79 distinctive phenotypic characteristics.

80 In *P. persica*, the absence of wild or feral populations makes *ex situ* collections the main
81 valuable reservoirs of allelic variability for many traits not yet exploited in current breeding programs.
82 Remarkable progress has been achieved in the phenotypic and genotypic characterization of peach
83 genetic resources (**Badenes et al., 2015; Cirilli et al., 2018; Yu et al., 2018**), taking advantage of
84 genome sequencing and the development of cutting-edge genotyping tools (**Verde et al., 2012;**
85 **Verde et al., 2017, Aranzana et al., 2019**). In the framework of the European collaborative project
86 FruitBreedomics (**Laurens et al., 2018**), a coordinated characterization of peach collections was

87 accomplished across relevant European repositories (**Micheletti et al., 2015; Hernandez-Mora et**
88 **al., 2017**), promoting increased utilization of resources and encouraging the sharing of conservation
89 responsibilities. For example, the *Prunus* Working Group within the Fruit Network in the European
90 Cooperative Programme on Plant Genetic Resources (ECPGR) is dealing with *Prunus*, including
91 peach (**Benediková and Giovannini, 2013**). Nevertheless, long-term maintenance of collections
92 remains particularly challenging due to intrinsic vulnerabilities (e.g. direct exposure to environmental
93 variables and pathogens) and costs for *in vivo* maintenance through vegetative propagation to
94 preserve the original genotypes. Moreover, compliance to phytosanitary requirements hampers the
95 sharing of resources among institutions, each having its own stock of materials, resulting in
96 redundancies or risk of loss for unique accessions.

97 This article describes the concept, design and realization of the first multi-site peach
98 reference collection (named as *PeachRefPop: PRP*) across five locations in three European
99 countries (Italy, Spain and Greece). Other than an invaluable tool for scientific studies, the *PRP* is
100 configuring as the first milestone of an international collaborative project for the conservation and
101 exploitation of European peach germplasm resources and, ultimately, as a true heritage for future
102 generations.

103

104 **Results**

105 **1. Criteria for construction of a reference panel of peach accessions and progenies**

106 The *PRP* collection was built with the aim of selecting a reduced germplasm pool, reflecting the
107 original genetic and phenotypic diversity (**Figure 1**) and the cultural and socio-economic value of
108 peach cultivation, for its exploitation in future breeding programs. A four-step procedure was followed
109 (exemplified in **Figure 2**):

- 110 1. Definition of the collection domain
- 111 2. Establishment of the number of entries
- 112 3. Identification of the criteria for entries selection
- 113 4. Choice and allocation of the entries

114

115 **1.1 Definition of the *PeachRefPop* domain**

116 In order to build a resource representing the European peach diversity and breeding history, the
117 starting point was the genetic material characterized in the framework of the FP7 European project
118 FruitBreedomics (<http://fruitbreedomics.com/>) in a coordinated effort involving different
119 universities and research institutions across Europe and China. A total of 1,580 *Prunus* accessions
120 (comprising *P. persica* and its hybrids with *P. davidiana* and almond), were phenotyped and
121 genotyped with the IPSC 9k SNP array, as previously described (**Micheletti et al., 2015**). The
122 inclusion of only peach (including *P. ferganensis*, **Verde et al. 2012**) among all the available *Prunus*
123 accessions was the leading concept behind the definition of the *PRP* reference collection. Indeed,

124 as a consequence of many factors (genetic diversity, evolution history, mating system, geographical
125 distribution etc.), sampling strategies for the inclusion of wild relatives (e.g. species of *Amygdalus*
126 subgenus) may substantially differ from those for a cultivated species (e.g. peach) (Brown and
127 Marshall, 1995). Moreover, to avoid limitations on the exchange of plant material, the domain was
128 restricted to the European repositories. Based on these criteria, the starting panel for building the
129 PRP amounted to a total of 1,262 *P. persica* accessions. Besides accessions, progenies from
130 controlled crosses also represent a valuable source of informative materials for both genetic analysis
131 and breeding (or pre-breeding) activities. For this reason, 1,467 individuals from 18 progenies and
132 their parents (including an interspecific cross with a *P. davidiana* accession), also analyzed during
133 the FruitBreedomics project (Hernandez Mora et al., 2017), were considered in the construction
134 process.

135

136 1.2 Establishment of the *PeachRefPop* size

137 The definition of the size is one of the most critical decisions for the establishment of a reference
138 population. For fruit tree crops, the costs of *in vivo* maintenance are particularly onerous, and
139 together with long-term space availability in the field, the main limiting factor of running a germplasm
140 collection. In the perspective of analyzing the interactions between genotype and environment and/or
141 management practices, or performing genetic studies such as GWAS and GS, an adequate panel
142 size and experimental design are key factors for the power and reliability of statistical analyses. On
143 the other hand, for agrobiodiversity conservation purposes, the least number of accessions to include
144 in a core set depends on the level of genetic repetitiveness present in the original germplasm pool.
145 The first step towards the establishment of the PRP size was the assessment of the allelic richness
146 and redundancy observed at marker loci. Two series of core collections of incremental size were
147 generated, one based on the genetic diversity (Maximization method, OPT), the other through
148 random sampling (RAN). The maximization procedure (M strategy by Schoen and Brown, 1993), is
149 based on the sampling of the total allelic diversity observed at marker loci in the least number of
150 entries. By plotting the genetic diversity measured over the core size, a convex curve was obtained,
151 indicating the presence of redundancy across the European peach germplasm collection. The
152 inflection point, corresponding to a plateau in the increase of diversity, was observed at the level of
153 core 26. At this core size, 99.9% of the total genetic diversity was captured in the core obtained with
154 the M method in comparison to 93.5% with random sampling (Figure 3). The outperformance of the
155 optimized versus the random selection was observed across all the core sizes, indicating that the
156 Maximization strategy was more efficient and was to be preferred for conservation purposes in our
157 germplasm.

158 According to some recent works in peach (reviewed in Aranzana et al., 2019), a number of about
159 100 – 150 unrelated accessions usually provides a satisfactory resolution for identifying major loci
160 or developing prediction models. In light of all the above premises, an ideal target number of 400

161 entries was deemed adequate for allocating a minimum of 150 accessions and a maximum of 250
162 seedlings from progenies (including the parents) based on the outputs of selection criteria.

163

164 **1.3 Identification of the selection criteria**

165 In spite of the genetic redundancy observed and excluding the rare cases of synonymy, the vast
166 majority of the accessions are not overlapped across the various collections, being conserved for a
167 multitude of reasons and purposes, including scientific research, agrobiodiversity preservation or
168 support to breeding activities. To reconcile these reasons with the aim of creating a feasible, usable
169 and multi-purpose reference collection to be shared among European institutions, a mixed approach
170 has been considered for selecting the accessions. A subset of entries was sampled by using an
171 analytical strategy, based on the criteria of maximizing genetic (and phenotypic) diversity, also taking
172 into account the availability of whole genome re-sequencing data (WGRS); the remaining entries
173 were selected using an empirical strategy, leveraging the knowledge of an experts panel (e.g.
174 breeders, experienced researchers and curators of each repository) and considering the traditional
175 and historical value at national and/or regional levels, the relevance for breeders, growers and
176 consumers, and particular agronomical or pomological characteristics. Moreover, in order to
177 maintain a balanced representation of the genetic structure of the whole collection, the empirical
178 selection of accessions was partially supported by information on population structure (Structure and
179 PCA analysis available from **Micheletti et al., 2015**). Complementing the choice of accessions,
180 progenies were selected based on the availability of detailed genotypic and/or phenotypic
181 information, genetic background, scientific relevance and, above all, priority traits for breeding.

182

183 **1.4 Choice, evaluation and description of the *PeachRefPop* accessions**

184 Capturing the maximum amount of genetic diversity present in the entire collection while reducing
185 redundancy was the primary driver for sampling the first *PRP* subset (the core set). For this purpose,
186 the advanced M method, implemented in the software PowerCore (**Kim et al., 2007**) through a
187 modified heuristic algorithm, was used to select a core from the initial panel of accessions, based on
188 a set of 3,894 filtered SNPs previously described in **Micheletti et al., (2015)**. After superimposing
189 17 accessions with available whole-genome re-sequencing data, an ideal core of 69 accessions
190 (PwC_69) was extracted, representing a sampling size of 5.5% (**Supplemental File 1**). Considering
191 the many variables that could affect the actual availability of materials for grafting, a flexible approach
192 was further developed to rank each accession of the whole panel based on genotypic and phenotypic
193 diversity. Four different sets made up of 100 cores of 70 entries each were constructed with MSTRAT
194 by setting different combinations of genotypic (9 subsets of SNPs extracted approximately every 1.8
195 Mb in order to avoid linkage between them) and phenotypic data (7 qualitative and 10 quantitative
196 traits, following transformation of the latter into categories) (**Supplemental File 2**). Accessions were
197 ranked in groups according to the average frequency of inclusion across the four sets

198 (**Supplemental File 3**). Combining the core population extracted by PowerCore with the MSTRAT
199 ranking list resulted in a shortlist of 69 accessions (41 and 28, respectively, indicated as Core_69),
200 ensuring the inclusion of the maximum possible level of genetic diversity. For the completion of the
201 final *PRP* panel, the remaining 100 accessions (Priority_100) were empirically selected by experts,
202 following the above specified criteria.

203 Estimates of genetic diversity were used to compare the starting panel of 1,262 European
204 accessions (EU_1262), the core collection obtained by PowerCore (PwC_69) and the final *PRP*,
205 composed by joining Priority_100 and Core_69 subsets. In addition, Core Hunter software was used
206 to create additional core sets, either of 69 and 169 entries, based on the optimization of various
207 criteria, including allelic coverage (CV_169) and three distance-based algorithms A-NE (AN_69 and
208 AN_169), E-NE (EN_69 and EN_169) and E-E (EE_69 and EE_169). Concerning parameters
209 accounting for allelic diversity, all sets showed high and similar values for the allelic coverage (CV),
210 while the number of effective alleles (N_e) and expected heterozygosity (H_e) were slightly lower for
211 the Priority_100 subset (**Table 1**). The Shannon-Weaver diversity index (SH) was comparable
212 among the different subsets, ranging between 0.595 in EE_169 and 0.534 in Priority_100. SH
213 generally displays higher values in the presence of a reduced redundancy (**Peet, 1975**). In contrast,
214 values of observed heterozygosity (H_o) tended to be more variable, ranging from a minimum of 0.202
215 in PwC_69 to a maximum of 0.318 in AN_69. According to **Odong et al. (2013)**, distance-based
216 criteria were used for further evaluations, such as the minimization of A-NE distance, particularly
217 indicated for generalist collections (as the *PRP*), and maximization of either E-E or E-NE, both
218 suitable for core collection representing the extremes of the entire collection. A-NE distance
219 generally tends to decrease along with the increase of core size, being minimized in the AN_169
220 and AN_69 core sets (0.137 and 0.172, respectively), *a priori* optimized using this selection criterion.
221 Despite the relative low performance of both Priority_100 and Core_69 (0.188 and 0.195,
222 respectively), *PRP* sets showed satisfactory values for this index (0.165), most probably as a
223 consequence of the increased size. Regarding E-E and E-NE, *PRP* (as well as Priority_100) showed
224 lower values, particularly for E-NE distance, indicating the presence of a certain redundancy within
225 the panel.

226 The population structure of peach germplasm is well represented in the *PRP*, in agreement
227 with the presence of clusters of breeding-derived accessions (further separated in peach- and
228 nectarine-type groups), Occidental traditional and admixed entries with prevalent Oriental origins
229 (**Figure 4A**). Structure is also preserved in the other core sets, except for that selected through the
230 E-E distance algorithm, tending to oversample the admixed group (**Supplemental Figure 1**). PCA
231 was also run to check the distribution of the *PRP* with respect to the other sets, the first two
232 components explaining 15.9 and 8.4%, respectively, of the total variance detected. In the scatter
233 plot, 95% confidence ellipses show almost overlapping areas (except for EE_169), confirming that
234 the *PRP* panel is well distributed to represent the structure of the starting germplasm (**Figure 4B**).

235 Finally, Neighbor-joining (NJ) tree, based on the dissimilarity matrix between the whole panel of
236 1,262 accessions, was also built to assess the distribution of *PRP* accessions (Figure 4C).

237 A number of accessions of historical and regional importance, mostly belonging to the
238 Occidental traditional cluster were included. For example, French cultivars dating from late Middle
239 Age ('Grosse Mignonne', 'Millecoton de Septembre', 'Reine des Verges', 'Brugnon Violet') (Okie et
240 al., 2008), traditional non-melting Spanish cultivars ('Amarillo de Agosto 1', 'Calante', 'Campiel',
241 'Jesca', 'Groc Abel', 'Groc Alto') (Badenes et al., 1998; Wünsch et al., 2006) and the Italian
242 'Crasiommo Rosso' (a white fleshed nectarine belonging to the 'Sbergie' type) and 'Poppa di
243 Venere', firstly reported at the end of eighteenth century (Majoli, 1790 - 1810). The richness of the
244 Italian peach germplasm is also widely represented by materials from several regions, including
245 Sicilia ('Imera', 'Tardiva di Ficarazzi', 'Settembrina di Bivona', 'Gialla di Moavero') (Marchese et al
246 2005), Campania ('Zingara Nera'), Puglia ('Percoco di Turi'), Liguria ('Michelini'), Emilia-Romagna
247 ('Buco Incavato', 'San Varano 2' and 'San Varano 3', 'Rosa del West', this last used for the
248 preparation of the famous cocktail 'Bellini') and Tuscany ('Regina di Londa') (Gallesio, 2003; Monte
249 et al., 2006; Liverani and Giovannini, 2016). Early breeding materials, mainly from US programs
250 and funders of most of the currently cultivated materials are also included, along with commercial
251 cultivars of worldwide diffusion (Supplemental File 4).

252 Finally, *PRP* accessions embrace a wide range of phenotypic variability for traits related to
253 fruit quality, resistance or tolerance against major diseases (brown rot, powdery mildew, leaf curl,
254 aphids and Sharka disease), tree habitus and phenology (Figure 5 and Supplemental File 4).

255

256 1.5 Choice and description of the *PeachRefPop* progenies

257 Seedlings from 15 cross populations from the research and breeding activities of some European
258 universities and institutions were also added. Most of these accessions were already described in
259 depth (Hernandez-Mora et al., 2017). The leading criterion for the choice of breeding materials was
260 the effective segregation of priority traits in peach, mainly related to phenology (fruit developmental
261 period, maturity date), fruit quality (fresh weight, soluble solid content, titratable acidity, texture and
262 aroma) and disease resistance (brown rot, powdery mildew, green peach aphids and Plum Pox
263 Virus) (Table 2). A range of breeding materials was considered, such as F1, F2, BC1 populations
264 as well as hybrids with *P. davidiana*, particularly interesting as a source of PPV resistance
265 (Decroocq et al., 2005). Within each selected cross-population (except for Sf x G and CREA-Forli
266 progenies), a distance matrix estimated from IPSC 9K SNP data was used to support the choice of
267 the seedlings (data not shown).

268

269 2. Experimental design and orchard sites description

270 Accessions and seedlings were propagated through grafting on a common 'GF677' rootstock by the
271 same nursery. All accessions and seedlings were grafted on the same year (August 2015) to obtain

272 trees with the same age. In order to ensure an adequate compromise between the number of
273 replicate trees and sustainable costs of maintenance, an augmented design with replicated checks
274 was adopted (**Figure 6A**). Accessions and seedlings were assigned to 8 subgroups (i.e. 'a1' to 'a4'
275 for accessions and 's1' to 's4' for seedlings) and arranged in two blocks:

- 276 - the M1 block, which includes two copies of the entire collection of 169 accessions plus 20
277 cross parents (the 'a' subgroup) and 214 seedlings (the 's' subgroup). Replicates are
278 randomly arranged in two separate sub-blocks. The composition of the M1 block is the same
279 across all sites but the order is randomized.
- 280 - the M2 block, which includes two partial replicates of the collection (85 accessions, 10 cross-
281 parents and 112 seedlings). Partial replicates are randomly arranged in two separate sub-
282 blocks, as for the M1 one. According to a pair wise scheme, one 'a' and one 's' subgroup is
283 common to at least two sites.
- 284 - the accessions 'Big Top', 'Springcrest' and 'Nectaross' were set as checks, randomly
285 distributed over M1 (10 replicates each) and M2 (7 replicate each) blocks.

286 A full copy of the *PRP* was planted in orchards of 4 institutions from 3 countries (Greece, Italy and
287 Spain) in addition to a partial one hosted by CREA-Roma (Italy) (**Figure 6B**), and including only the
288 'a' subgroup without block randomization (**Figure 6B**).

289 The site of the Institute of Plant Breeding and Genetic Resources (IPB&GR) in Naoussa
290 (Imathia region, Greece) is located at geographical coordinates 40°37' N, 22°06' E and 119 m
291 altitude. The area is characterized by dry summers and enough chilling hours. The average annual
292 temperature is 15.5° C, with an average minimum and maximum of 10.8° and 21.6° C, respectively.
293 The average annual precipitation is 724 mm (mostly concentrated in autumn-winter period). The soil
294 is sandy-loam, neutral reaction (pH 6.8), low carbonate content and 2.50% of organic matter
295 (average measures from soil depth until 45 cm).

296 The site of the Institute of Agrifood Research and Technology (IRTA) in Gimenezs (Catalonia
297 region, Spain) is located at geographical coordinates 41°65' N, 0°39' E and 259 m altitude. The
298 average annual temperature is 14.3° C, with an average minimum and maximum of 2.3° and 27.4°
299 C, respectively. The average annual precipitation is 349 mm. The soil is sandy-loam, sub-alkaline
300 reaction (pH 7.7), very high carbonate content and 2.65% of organic matter.

301 The site of the Instituto Murciano de Investigacion y Desarrollo Agrario y Alimentario (IMIDA)
302 in Mula (Murcia region, Spain) is located at geographical coordinates 38°3'55.595 N, 1°25'42.931 O
303 and 278 m altitude. The average annual temperature is 17.8° C, with an average minimum and
304 maximum temperatures of 3.2° and 29.7° C, respectively. The average annual precipitation is 308
305 mm. The soil is clay, sub-alkaline reaction (pH 7.8), very high carbonate content and 2.65% of
306 organic matter.

307 The site of the Centro di Ricerca per le Produzioni Vegetali (CRPV) in Imola (Emilia-
308 Romagna region, Italy) is located at geographical coordinates 44°33' N, 12°33' E and 53 m altitude.

309 The average annual temperature is 13.9° C, with an average minimum and maximum of 12.5° and
310 25.1° C, respectively. The average annual precipitation is 766 mm. The soil is silty-loam, neutral
311 reaction (pH 7.2), moderate carbonate content and 1.47% of organic matter.

312 The site of the Research Centre for Olive, Citrus and Tree Fruit of Rome (CREA, Italy), is
313 located at 41°47'48.0"N 12°33'58.1"E and 79 m altitude. The average annual temperature is 16.8°
314 C, with an average minimum and maximum of 12.1° and 21.8° C, respectively. The average annual
315 precipitation is 792 mm. The soil is sandy-loam, sub-alkaline reaction (pH 7.7), no carbonate content
316 and 1.9% of organic matter.

317

318 **Discussion and Perspectives**

319 The concept of the *PRP* arises from the growing awareness about current and common
320 issues on *ex situ* peach conservation across European institutions. Fluctuations in funds availability
321 and intrinsic constraints of living orchard collections threaten the long-term preservation of diversity
322 resources, causing a progressive loss of valuable materials. Reference or core collections have been
323 designed for several fruit tree species, for example olive (**Khadari et al., 2003; El Bakkali et al.,**
324 **2012; Belaj et al., 2012**), grape (**Laucou, et al., 2011**), cherry (**Campoy et al., 2016**), apple (**Gross**
325 **et al., 2013; Lassois et al., 2016**), apricot (**Krichen et al., 2012**). Nevertheless, they have mainly
326 been created for improving resource allocation in the context of a single institution or repository. The
327 development of a trans-national and shared strategy is configuring as the most promising opportunity
328 in the conservation approach. Actual establishment of the *PRP* has required huge coordination
329 efforts, facing with the effective availability of materials, the difficulties of their exchange and the
330 success of clonal propagations (particularly for old, often unique, accessions). The sampling strategy
331 for *PRP* has been defined to accommodate multiple purposes while maintaining the maximum
332 possible diversity compared to the starting panel. The final panel was assembled by the combination
333 of two different subsets: the first (Core_69), ensuring the preservation of the total allele number with
334 the minimum number of accessions, was extracted by widely adopted maximization strategies, either
335 using a class coverage criterion (PowerCore) or Shannon-Wheaver Index (MSTRAT), this last
336 penalizing redundancy. The second subset accommodating for other scopes (Priority_100) was
337 chosen by experts, with a robust knowledge on the genetic structure in peach providing a reliable
338 criterion for assisting selection. As a whole, genetic analysis supports that *PRP* composition is highly
339 representative of the diversity of peach germplasms present in European collections, as it retains all
340 the allelic variability present within the starting panel, specifically targets defined genetic clusters
341 according to the genetic structure and includes most relevant phenotypic traits. Indeed, differences
342 among the various sampling strategies were negligible for allelic coverage (CV), expected
343 heterozygosity and SH index, revealing a buffer effect towards optimization criteria. Such effect could
344 be expected, since peach has experienced a severe domestication bottleneck with a reduction of
345 genetic diversity, followed by a strong artificial selection during domestication and modern

346 improvement (**Verde et al., 2013; Yu et al., 2018; Li et al., 2019**). This is also reflected in the narrow
347 genetic bases of peach germplasm available across main European repositories. Thus, the high
348 level of allelic redundancy allows selecting many different subpopulations able to retain the same
349 amount of genetic variation. In spite of this, a preliminary validation using distance-based criterion
350 not used in the selection stage provides satisfactory results for the A-NE index, the most indicative
351 for evaluating the quality of multipurpose collection (**Odong et al., 2013**). Conversely, E-E and,
352 particularly, E-NE indices resulted less optimized, due to a certain redundancy on the Priority_100
353 subset (i.e. a higher number of genotypes providing unique alleles). This was mainly due to the
354 inclusion of accessions of traditional and breeding values, respectively belonging to the Occidental
355 Traditional and Occidental breeding clusters, characterized by a very narrow genetic background.
356 Clearly, the inclusion of these materials is crucial in the overall perspective of balancing diversity and
357 usefulness, as they integrated various fundamental qualities such as popularity, prestige, tradition
358 and breeding. A similar mixed strategy was also recently optimized for creating a core collection for
359 Swiss pear germplasm (**Urrestarazu et al., 2019**).

360 Climate challenges in peach growing areas increase the need for resilient cultivars, able to
361 maintain productivity while showing an increased capacity for adaptation to sub-optimal conditions.
362 Nevertheless, resilience and adaptive traits often have a complex inheritance and a strong
363 interaction with the environment or cultivation practice. The partitioning of phenotypic variation into
364 genotypic, environmental and their interaction components involves *ad hoc* experimental designs
365 and integration of field data on a common set of genetic materials under a range of different
366 environmental/management conditions. Multi-environment trials (METs) have been extensively used
367 to study GxExM interactions, carry out GWAS and develop GS models for complex traits in annual
368 crops (**Malosetti et al., 2013; Gutierrez et al., 2015; Zhu et al., 2018; Bustos-Korts et al., 2019**).
369 In contrast, such experimental designs are lagging in fruit trees, largely because of the need for large
370 and diverse germplasm sets for quantitative genetics analyses and the above-mentioned difficulties
371 in material propagation and exchange. The *PRP* aims to fill this gap, as the replicated design and
372 the different pedo-climatic conditions across sites are particularly indicated for the dissection of
373 interactions between genotype and environment and/or management practice. Also, the inclusion of
374 both accessions and progenies allows development and testing of novel statistical approaches for
375 genomics-assisted breeding, such as joint linkage-association analysis (**Yu et al., 2008; Lu et al.,**
376 **2010**) and genome-wide selection (**Resende et al., 2012; van Nocker and Gardiner, 2014**).

377 In perspective, the *PRP* should fulfill several purposes, from scientific research to education
378 and traineeship of young breeders. A better understanding of diversity is expected to encourage the
379 use of broad-ranging germplasm (maybe also in other existing *ex situ* collections) in breeding
380 programs. In the last decades, the mission of many agriculture-oriented institutions has shifted from
381 the traditional focus of establishing horticultural collections to a wider target of preserving germplasm
382 resources and agricultural heritage (**Hammer et al., 2003; Havens et al., 2006**). This objective is of

383 utmost importance for fruit tree species of ancient cultivation history, such as peach. For these
384 reasons, a number of traditional and local cultivars (either old or relatively modern) has been included
385 in the *PRP*, as a safeguard of an integral part of the rural landscape and collective memory. Since
386 information and descriptions about local germplasms are scarce and often restricted to cultivation
387 areas, their choice has been directly handled by curators of each repository, with the aid of
388 experienced breeders.

389

390 **Materials and Methods**

391 **Dataset**

392 A set of 1,262 accessions was selected as representative of the *Prunus* germplasm maintained in
393 collections of four different European countries. The complete list of institutions providing plant
394 materials, SNP genotyping and phenotypic data for seven monogenic traits have been previously
395 described (Micheletti et al., 2015). All data, including phenotyping of 10 quantitative traits, were
396 retrieved from FruitBreedomics database available at
397 <http://bioinformatics.tecnoparco.org/fruitbreedomics/> website.

398 **Construction of core subsets**

399 The advanced M (maximization) strategy implemented in PowerCore v. 1.0 (Kim et al., 2007) using
400 3,894 SNP markers, was carried out to extract a core subset able to capture all the alleles observed
401 in the entire collection. The size of the final core collection depends on the level of variability and
402 redundancy present in the whole panel and cannot be set *a priori*. Seventeen kernel accessions with
403 available whole-genome re-sequencing data were superimposed through the 'preferential selection'
404 tool, which retains the accessions defined by the user without validation. The standard M strategy
405 implemented in MSTRAT (Gouesnard et al., 2001) was also applied. MSTRAT algorithm selects a
406 subset of n accessions from the N accessions of the entire collection by maximizing the number of
407 alleles (and/or trait classes) at each locus. The sampling size estimated with PowerCore was set as
408 default parameter and four sets of 100 core collections were constructed by using different
409 combinations of genotypic and phenotypic data. Due to the restraints in the number of variables
410 MSTRAT is able to manage, different subsets of approximately 100 SNPs each were obtained
411 through an *ad hoc* developed Perl script program, by extracting 1 SNP every 1,800 Kbp,
412 corresponding to the max boundary for LD found in some subpopulations of the original plant
413 material (Micheletti et al 2015). Seven qualitative and 10 quantitative traits (these last transformed
414 into qualitative categories) were used as phenotypic data. For each run, the core size was set to 70
415 and 100 independent replicates with 100 iterations were generated. The Shannon-Weaver diversity
416 index was used as a second criterion to classify core subsets. Redundancy was assayed through
417 the 'Redundancy' tool implemented in MSTRAT, which samples two different sets of core collections
418 of increasing size, as defined by the user, through the application of the maximization strategy or
419 random sampling. For this analysis a subset of 445 SNP markers was pruned from the whole set of

420 4271 using Plink v1.07 with a window size of 50, a shift of 7 and a variance inflation factor (VIF) of
421 2. Redundancy was assayed in the whole panel of accessions with a step of 5 in the first 100, 5
422 repetitions and 50 iterations. The Mixed Replica search algorithm implemented in the Core Hunter II
423 software (De Beukelaer et al., 2012) was used to generate a core collection of fixed size (either of
424 69 and 169 entries) based on the optimization of the Modified Rogers' (MR) distance measure
425 (Wright, 1978), with a weight of 1.0. For the evaluation of the quality of the different core subsets,
426 genetic distance-based criteria were considered: the average genetic distance between all the
427 entries of each core collection (E-E); the average distance between each entry and the nearest
428 neighboring entry for each core collection (N-E); the average distance between each genotype of
429 the entire collection and the nearest entry in each core collection (A-NE). The quality of each
430 collection increase for lower value of A-NE (the maximum representation is obtained for AN = 0,
431 when each accession is represented by itself or by an identical duplicate), and higher value both for
432 E-NE (maximizes the average distance between each selected individual and the closest other
433 selected item in the core) and E-E (maximizes the average distance between each pair of selected
434 individuals in the core.).

435 **Genetic diversity and population analyses**

436 Genetic diversity measures were performed using GenAlex 6.41 (Peakall et al., 2006) and include:
437 number of effective alleles (N_e , the number of equally frequent alleles required to give the observed
438 level of heterozygosity), levels of observed (H_o) and expected (H_e) heterozygosity, and the Shannon-
439 Weaver index (I_s). Allelic coverage was calculated by the function CV implemented in Core Hunter
440 II software. Population structure was inferred using a model-based clustering algorithm ADMIXTURE
441 v1.22 (Alexander et al., 2009). From SNP data, the software identifies K a priori genetic clusters
442 provided by the user and for each individual it estimates the probability of membership to each
443 cluster. A preliminary analysis was performed by inputting successive values of K from 2 to 6. The
444 value of K that maximized the predictive accuracy was chosen based on a 10-fold cross-validation
445 procedure with 10 different fixed initial seeds (Supplemental Figure 2). Data of Principal
446 Component Analysis (PCA) were retrieved from a previous work (Micheletti et al., 2015). The 95%
447 confidence ellipses in the scatter plot were estimated using PAST software (Hammer et al., 2001).
448 Phylogenetic tree was built from a pairwise genetic distance matrix between individuals clustered
449 with NJ method in TASSEL (Bradbury et al., 2007). Bootstrap replicate and tree reconstruction were
450 performed in MEGA7 software (Kumar et al., 2016).

451

452 **Supplemental Materials**

453 **Supplemental File 1.** PowerCore output.

454 **Supplemental File 2.** MSTRAT outputs for the four settings.

455 **Supplemental File 3.** Accession ranking by MSTRAT frequencies.

456 **Supplemental File 4.** *PeachRefPop* accessions description.

457 **Supplemental Figure 1.** Population structure estimated in the core sets AN_169, EE_169, EN_169
458 and CV_169.

459 **Supplemental Figure 2.** Predictive accuracy (cross-validation error) of population stratification in
460 both Refpop_169 and EU_1262 as determined by Admixture software.

461

462 **Acknowledgments**

463 We wish to thank Claudio Buscaroli and Martina Lama for field assistance, Remo Chiozzotto for lab
464 assistance and Michela Troggio (Fondazione Edmund Mach) for genotypic analyses. We thank the
465 INRA's '*Prunus* Genetic Resources Center' of INRA-Nouvelle Aquitaine-Bordeaux for preserving and
466 managing the peach collections.

Table 1. Genetic analysis and parameters for the different core subsets. N_e : number of effective alleles); SH: Shannon-Weaver diversity index; H_o : observed Heterozygosity; H_e : expected Heterozygosity; CV: percentage allelic coverage; MR distance: average Modified Rogers genetic distance; E-E: average entry to entry distance; A-NE: average distance between each genotype of the collection and the nearest entry, E-NE: average distance between each entry and the nearest entry.

Set name	N_e	SH	H_o	H_e	CV	MR distance		
						EE	EE	EE
EU_1262	1.621	0.547	0.292	0.367	0.995	0.285	-	0.131
PwC_69	1.675	0.574	0.202	0.39	0.987	0.318	0.203	0.237
EE_69	1.705	0.587	0.234	0.401	0.992	0.347	0.209	0.229
AN_69	1.645	0.560	0.318	0.378	0.977	0.286	0.172	0.210
EN_69	1.704	0.587	0.269	0.402	0.991	0.334	0.207	0.275
CV_169	1.638	0.556	0.285	0.375	0.995	0.302	0.163	0.212
EE_169	1.721	0.595	0.224	0.408	0.994	0.330	0.183	0.191
AN_169	1.643	0.559	0.300	0.377	0.987	0.290	0.137	0.203
EN_169	1.683	0.578	0.277	0.394	0.993	0.315	0.175	0.256
Core_69	1.713	0.593	0.247	0.406	0.988	0.303	0.195	0.212
Priority_100	1.597	0.534	0.283	0.356	0.979	0.277	0.188	0.179
<i>PeachRefPop</i>	1.647	0.563	0.270	0.379	0.988	0.290	0.165	0.180

Table 2. Description of the progenies used for establishing the *PeachRefPop* collection. Traits abbreviation: FD, flowering date; MD, maturity date; SSC, soluble solid content; FW, fruit weight; BR, brown rot; TA, titratable acidity; SwS, slow-softening texture; PM, powdery mildew; PPV, Plum Pox Virus; GPA, green peach aphid; SH, stony hard texture.

Cross (parents)	Acronym	Institution	Type of Progeny	Seedlings #	Trait(s)
'Bolero' × 'Oro A'	B × O	UMIL - Milan	F1	9	MD, SSC, FW, skin overcolor, aroma
'Contender' × 'Elegant Lady'	C × EL	UMIL - Milan	F1	14	BR, MD
'Max 10' × 'Rebus 028'	M × R	UMIL - Milan	F1	9	MD, TA, SSC, FW, SwS
'Sweetfire' × 'Garcica'	Sf × G	UMIL - Milan	F1	15	MD, TA, SSC, FW, SwS
'Belbinette' × 'Nectalady'	Bb × NI	IRTA - Lleida	F1	20	FD, MD, TA, SSC, FW
'Big Top' × 'Nectaross'	Bt × Nr	IRTA - Lleida	F1	19	FD, MD, TA, SSC, FW
'Big Top' × 'Armking'	Bt × Ak	IRTA - Lleida	F1	18	FD, MD, TA, SSC, FW
'Subirana' × 'Feraude'	PN643	IRTA - Lleida	F1	7	Fruit shape
'Summergrand' × 'P. davidiana P1908'	SD	INRA - Avignon	F1	6	PM, PPV
'Zephyr' × [(['Summergrand' (S) × 'P. davidiana P1908']) × S]	BC2	INRA - Avignon	BC2	13	FD, PM, PPV, TA, SSC, FW
'Pamirskij 5' × 'Rubira'	P × R	INRA - Avignon	F2	13	PM, GPA, foliage colour
FRF 1495 × FRF 1148 (Ma 16-03-059)	POP1376	CREA - Forli	F1	17	PM; fruit pubescence
IFF 983 × Ma 25-01-042	POP1115	CREA - Forli	F1	17	TA, SwS and aroma
FRF 1695 × FRF 1681	POP1095	CREA - Forli	F1	19	SH
FRF 813 × FRF 691	POP1039	CREA - Forli	F1	18	skin overcolor

467 Figure legends

468 **Figure 1.** Overview of the range of phenotypic diversity in the *PeachRefPop*.

469 **Figure 2.** Graphical summary of the overall scheme followed for selecting the *PeachRefPop*
470 collection.

471 **Figure 3.** Redundancy in starting panel (EU_1262) estimated with MSTRAT software using a set of
472 445 SNPs. Datapoints represent averaged values over 5 independent repetitions for each size.

473 **Figure 4.** A) Population structure estimated in the whole panel (EU_1262) and *PeachRefPop*, as
474 estimated for K (number of a priori cluster) equal to 4; B) PCA analysis of the subsets with a core
475 size of 169 entries. Scores for each accession were obtained from the work of **Micheletti et al.,**
476 **2015**. The 95% confidence ellipses in the scatter plot were estimated using PAST software. C) NJ
477 phylogenetic tree. Blue navy squared indicated accessions with traditional and historical value, violet
478 circle indicates the other *PeachRefPop* accessions, while colors reflect the population structure.

479 **Figure 5.** Distribution of main phenotypic traits in the *PeachRefPop* collection.

480 **Figure 6.** A) Experimental design and B) Google maps satellite imageries of the established
481 *PeachRefPop* orchards across the different European sites.

482

483 **References**

484 Alexander DH, Novembre J, Lange K. (2009) Fast model-based estimation of ancestry in unrelated
485 individuals. *Genome Res.*; 19:1655–64

486 Aranzana, M. J., Decroocq, V., Dirlwanger, E., Eduardo, I., Gao, Z. S., Gasic, K. et al. (2019). *Prunus*
487 genetics and applications after de novo genome sequencing: achievements and prospects. *Horticulture*
488 research, 6.

489 Badenes, M.L., Martínez-Calvo, J. and Llácer, G. (1998a) Analysis of peach germplasm from Spain.
490 *Acta Horticulturae* 465, 243–250.

491 Belaj, A., del Carmen Dominguez-García, M., Atienza, S. G., Urdíroz, N. M., De la Rosa, R., Satovic,
492 Z., ... & Del Río, C. (2012). Developing a core collection of olive (*Olea europaea* L.) based on molecular
493 markers (DARs, SSRs, SNPs) and agronomic traits. *Tree Genetics & Genomes*, 8(2), 365-378.

494 Benediková, D. and Giovannini, D. (2013). Review on genetic resources in the ECPGR *Prunus*
495 Working Group. *Acta Horticulturae* 981:43-51.

496 Biasi R, Botti F, Cullotta S, Barbera G (2012a) The role of Mediterranean fruit tree orchards and
497 vineyards in maintaining the traditional agricultural landscapes. *Acta Hort* 940:79–88

498 Bradbury, P. J., Zhang, Z., Kroon, D. E., Casstevens, T. M., Ramdoss, Y., & Buckler, E. S. (2007).
499 TASSEL: software for association mapping of complex traits in diverse samples. *Bioinformatics*,23(19), 2633-
500 2635.

501 Campoy, J. A., Lerigoleur-Balsemin, E., Christmann, H., Beauvieux, R., Girollet, N., Quero-García, J.,
502 ... & Barreneche, T. (2016). Genetic diversity, linkage disequilibrium, population structure and construction of
503 a core collection of *Prunus avium* L. landraces and bred cultivars. *BMC plant biology*, 16(1), 49.

504 Chagné, D., Dayatilake, R. Diack, M. Oliver, H. Ireland, A. Watson, S.E. Gardiner, J.W. Johnston,
505 R.J. Schaffer and S. Tustin (2014) Genetic and environmental control of fruit maturation, dry matter and
506 firmness in apple (*Malus × domestica* Borkh.). *Hortic. Res.* 1: 14046.

507 Cirilli, M., Geuna, F., Babini, A. R., Bozhkova, V., Catalano, L., Cavagna, B., ... & Ilardi, V. (2016).
508 Fighting Sharka in peach: current limitations and future perspectives. *Frontiers in plant science*, 7, 1290.

509 Cirilli, M., Rossini, L., Geuna, F., Palmisano, F., Minafra, A., Castrignanò, T., ... & Bassi, D. (2017).
510 Genetic dissection of Sharka disease tolerance in peach (*P. persica* L. Batsch). *BMC Plant Biology*,17(1), 192.

511 Cirilli, M., Flati, T., Gioiosa, S., Tagliaferri, I., Ciacciulli, A., Gao, Z., ... & Rossini, L. (2018). PeachVar-
512 DB: a curated collection of genetic variations for the interactive analysis of peach genome data. *Plant and Cell*
513 *Physiology*, 59(1), e2-e2.

514 Coakley, S. M., Scherm, H., & Chakraborty, S. (1999). Climate change and plant disease
515 management. *Annual review of phytopathology*, 37(1), 399-426.

516 Corelli-Grappadelli, L., and Marini, R. P. (2008) Orchard Planting Systems. In: Layne DR, Bassi D,
517 editors. *The Peach: Botany, Production and Uses*. Wallingford: CABI. pp. 264–288.

518 Decroocq, V., et al. "Analogues of virus resistance genes map to QTLs for resistance to sharka disease
519 in *Prunus davidiana*." *Molecular genetics and genomics* 272.6 (2005): 680-689.

520 El Bakkali, A., Haouane, H., Moukhli, A., Costes, E., Van Damme, P., & Khadari, B. (2013).
521 Construction of core collections suitable for association mapping to optimize use of Mediterranean olive (*Olea*
522 *europaea* L.) genetic resources. *PLoS One*, 8(5), e61265.

523 Faust, M. and Timon, B. (1995) Origin and dissemination of peach. *Horticultural Reviews* 17, 331–
524 379.

525 Fideghelli, C., Monastra, F. and De Salvador, R.F. (1988) Evoluzione delle forme d'allevamento e delle
526 densità d'impianto in peschicoltura. In: Sansavini, S. (ed.) *Proceedings of XVIII Convegno Peschicolo*. Grafi
527 che MDM, Forlì, Italy, pp. 63–81.

528 Ford-Lloyd, B. and Jackson, M (1986) *Plant Genetic Resources: An Introduction to Their Conservation*
529 *and Use*, Edward Arnold, London.

530 Franco J (2006) Sampling strategies for conserving maize diversity when forming core subsets using
531 genetic markers. *Crop Sci*46(2):854–864

532 Frankel, OH., and Brown, A.H.D (1984) Current plant genetic resources – a critical appraisal. In:
533 *Genetics: New Frontiers (vol IV)*. New Delhi, India: Oxford and IBH Publishing)

534 Fu, Y. B., & Dong, Y. B. (2015). Genetic erosion under modern plant breeding: case studies in
535 Canadian crop gene pools. In *Genetic Diversity and Erosion in Plants* (pp. 89-104). Springer, Cham.

536 Gallesio, G. (2003) Il trattato del pesco di Giorgio Gallesio. In: Baldini, E. (ed.) *Gli inediti trattati del*
537 *pesco e del ciliegio. Complementi scientifici della 'Pomona Italiana' di Giorgio Gallesio*. Accademia dei
538 *Georgofili*, Florence, Italy, pp. 9–146.

539 Gepts, P. (2006). Plant genetic resources conservation and utilization. *Crop Science*, 46(5), 2278-
540 2292.

541 Gross, B. L., Volk, G. M., Richards, C. M., Reeves, P. A., Henk, A. D., Forsline, P. L., ... & Chao, C.
542 T. (2013). Diversity captured in the USDA-ARS national plant germplasm system apple core collection. *Journal*
543 *of the American Society for Horticultural Science*, 138(5), 375-381.

544 Khadari, B., Breton, C., Moutier, N., Roger, J., Besnard, G., Bervillé, A., & Dosba, F. (2003). The use
545 of molecular markers for germplasm management in a French olive collection. *Theoretical and Applied*
546 *Genetics*, 106(3), 521-529.

547 Kim, K. W., Chung, H. K., Cho, G. T., Ma, K. H., Chandrabalan, D., Gwag, J. G., ... & Park, Y. J.
548 (2007). PowerCore: a program applying the advanced M strategy with a heuristic search for establishing core
549 sets. *Bioinformatics*, 23(16), 2155-2162.

550 Krishnan RR, Sumathy R, Ramesh S, Bindroo B, Naik GV. SimEli: Similarity elimination method for
551 sampling distant entries in development of core collections. *Crop Sci*. 2014; 54(3):1070–8.

552 Krichen, L., Audergon, J. M., & Trifi-Farah, N. (2012). Relative efficiency of morphological characters
553 and molecular markers in the establishment of an apricot core collection. *Hereditas*, 149(5), 163-172.

554 Hammer, Ø., Harper, D. A., & Ryan, P. D. (2001). PAST: paleontological statistics software package
555 for education and data analysis. *Palaeontologia electronica*, 4(1), 9.

556 Laucou, V., Lacombe, T., Dechesne, F., Siret, R., Bruno, J. P., Dessup, M., ... & Santoni, S. (2011).
557 High throughput analysis of grape genetic diversity as a tool for germplasm collection management.
558 *Theoretical and Applied Genetics*, 122(6), 1233-1245.

559 Malosetti, Marcos, Jean-Marcel Ribaut, and Fred A. van Eeuwijk. "The statistical analysis of multi-
560 environment data: modeling genotype-by-environment interaction and its genetic basis. *Frontiers in*
561 *physiology*, 4 (2013): 44.

562 Marchese A et al (2005) Molecular characterisation of Sicilian *Prunus persica* cultivars using
563 microsatellites. *The Journal of Horticultural Science and Biotechnology* 80: 121-129

564 Mora, J. R. H., Micheletti, D., Bink, M., Van de Weg, E., Cantín, C., Nazzicari, N., ... & Campoy, J. A.
565 (2017). Integrated QTL detection for key breeding traits in multiple peach progenies. *BMC genomics*, 18(1),
566 404.

567 Li Y, Cao K, Zhu G, Fang W, Chen C, Wang X, Zhao PP, Guo J, Ding T, Guan L, Zhang Q, Guo W,
568 Fei Z, Wang L (2019). Genomic analyses of an extensive collection of wild and cultivated accessions provide
569 new insights into peach breeding history. *Genome Biology* 20: 36

570 Liverani, A. and Giovannini, D. Pesco. In: Fideghelli, C. *Atlante dei fruttiferi autoctoni italiani*, volume
571 III. ISBN 978-88-99595-35-7 (Italian)

572 Foulongne M, Pascal T, Arus P, Kervella J. The potential of *Prunus davidiana* for introgression into
573 peach [*Prunus persica* (L.) Batsch] assessed by comparative mapping. *Theoret Appl Genet* 2003; 107: 227–
574 238.

575 Gradziel TM (2002) Almond species as sources of new germplasm for peach improvement. *Acta Hort*
576 592:81–88

577 Gouesnard, B., Bataillon, T. M., Decoux, G., Rozale, C., Schoen, D. J., & David, J. L. (2001). MSTRAT:
578 An algorithm for building germ plasm core collections by maximizing allelic or phenotypic richness. *Journal of*
579 *heredity*, 92(1), 93-94.

580 Gutiérrez, L., Germán, S., Pereyra, S., Hayes, P. M., Pérez, C. A., Capettini, F., ... & Fros, D. (2015).
581 Multi-environment multi-QTL association mapping identifies disease resistance QTL in barley germplasm from
582 Latin America. *Theoretical and applied genetics*, 128(3), 501-516.

583 Laurens, F., Aranzana, M. J., Arus, P., Bassi, D., Bink, M., Bonany, J., ... & Mauroux, J. B. (2018). An
584 integrated approach for increasing breeding efficiency in apple and peach in Europe. *Horticulture research*,
585 5(1), 11.

586 Li, Y., Cao, K., Zhu, G., Fang, W., Chen, C., Wang, X., ... & Zhang, Q. (2019). Genomic analyses of
587 an extensive collection of wild and cultivated accessions provide new insights into peach breeding history.
588 *Genome biology*, 20(1), 36.

589 Luedeling, E. (2012). Climate change impacts on winter chill for temperate fruit and nut production: a
590 review. *Scientia Horticulturae*, 144, 218-229.

591 Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: molecular evolutionary genetics analysis
592 version 7.0 for bigger datasets. *Molecular biology and evolution*, 33(7), 1870-1874.

593 Myles, S. (2013) Improving fruit and wine: what does genomics have to offer? *Trends Genet.* 29: 190–
594 196.

595 Majoli C. (1790-1810). *Plantarum collectio juxta Linnaeanum systema a Lectore Caesare Majolio*
596 *Hieronimino digesta et depicta*. Forli.

597 Micheletti, D. et al. Whole-genome analysis of diversity and SNP-major gene association in peach
598 germplasm. *PLoS ONE* 10, e0136803 (2015).

599 Moing A, Poessel JL, Svanella-Dumas L, Loonis M, Kervella J. (2003) Biochemical basis of low fruit
600 quality of *Prunus davidiana*, a pest and disease resistance donor for peach breeding. *J Am Soc Hortic Sci*; 128:
601 55–62.

602 Monte, M., F. Sottile, E. Barone, T. Caruso, and A. Bazzoni. 2006. The Sicilian Peach (*Prunus persica*
603 L. Batsch) Germplasm: Horticultural Characteristics and Sanitary Status. *Acta Hort.* 713: 57-60

604 Odong T, van Heerwaarden J, Jansen J, van Hintum TJ, van Eeuwijk F. Statistical techniques for
605 defining reference sets of accessions and microsatellite markers. *Crop Sci.* 2011; 51(6):2401–11.

606 Okie, WR, Bacon, T and Bassi, D (2008) Fresh Market Cultivar Development. In: Layne DR, Bassi D,
607 editors. *The Peach: Botany, Production and Uses*. Wallingford: CABI. pp. 264–288.

608 Parajuli, R., Thoma, G., and Matlock, M. D. (2018). Environmental sustainability of fruit and vegetable
609 production supply chains in the face of climate change: A review. *Science of The Total Environment*.

610 Peakall, R. O. D., and Smouse, P. E. (2006). GENALEX 6: genetic analysis in Excel. Population
611 genetic software for teaching and research. *Molecular ecology notes*, 6(1), 288-295.

612 Peet, R. K. (1975). Relative diversity indices. *Ecology*, 56(2), 496-498.

613 Purcell S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A. R., Bender, D. et al. (2007). PLINK:
614 a tool set for whole-genome association and population-based linkage analyses. *The American Journal of*
615 *Human Genetics*, 81(3), 559-575.

616 Ramírez, F., and Kallarackal, J. (2015). Responses of fruit trees to global climate change. Springer.

617 Schoen, D. J., and Brown, A. H. D. (1995). Maximising genetic diversity in core collections of wild
618 relatives of crop species. *Core collections genetic resources* (T. Hodgkin, AHD Brown, Th. JL van Hintum and
619 EAV Morales, eds.). John Wiley & Sons, Chichester, UK, 55-77.

620 Serra O, Donoso JM, Picañol R, Batlle I, Howad W, Eduardo I, Arús P (2016) Marker-assisted
621 introgression (MAI) of almond genes into the peach background: a fast method to mine and integrate novel
622 variation from exotic sources in long intergeneration species. *Tree Genetics & Genomes* 12: 96

623 Singh, H. C. P., Rao, N. K. S., and Shivashankar, K. S. (Eds.). (2013). *Climate-Resilient Horticulture:*
624 *Adaptation and Mitigation Strategies* (pp. 81-88). Springer India.

625 Thachuk, C., Crossa, J., Franco, J., Dreisigacker, S., Warburton, M., and Davenport, G. F. (2009).
626 Core Hunter: an algorithm for sampling genetic resources based on multiple genetic measures. *BMC*
627 *bioinformatics*, 10(1), 243.

628 Urrestarazu, J., Kägi, C., Bühlmann, A., Gassmann, J., Santesteban, L. G., Frey, J. E. et al. (2019).
629 Integration of expert knowledge in the definition of Swiss pear core collection. *Scientific Reports*, 9(1), 8934.

630 Varshney, R. K., Bansal, K. C., Aggarwal, P. K., Datta, S. K., and Craufurd, P. Q. (2011). Agricultural
631 biotechnology for crop improvement in a variable climate: hope or hype? *Trends in plant science*, 16(7), 363-
632 371.

633 Verde, I. et al. Development and evaluation of a 9K SNP array for peach by internationally coordinated
634 SNP detection and validation in breeding germplasm. *PLoS ONE* 7, e35668 (2012).

635 Verde, I. et al. The high-quality draft genome of peach (*Prunus persica*) identifies unique patterns of
636 genetic diversity, domestication and genome evolution. *Nat. Genet.* 45, 487–U47 (2013).

637 Verde, I. et al. The Peach v2.0 release: high-resolution linkage mapping and deep resequencing
638 improve chromosome-scale assembly and contiguity. *BMC Genomics* 18, 225 (2017).

639 Yu, Y. et al. Genome re-sequencing reveals the evolutionary history of peach fruit edibility. Nat.
640 Commun. 9, 5404 (2018).
641 Zhu, X. M., Shao, X. Y., Pei, Y. H., Guo, X. M., Li, J., Song, X. Y., & Zhao, M. A. (2018). Genetic
642 diversity and genome-wide association study of major ear quantitative traits using high-density SNPs in
643 maize. *Frontiers in plant science*, 9.



Figure 1. Overview of the range of phenotypic diversity in the *PeachRefPop*.

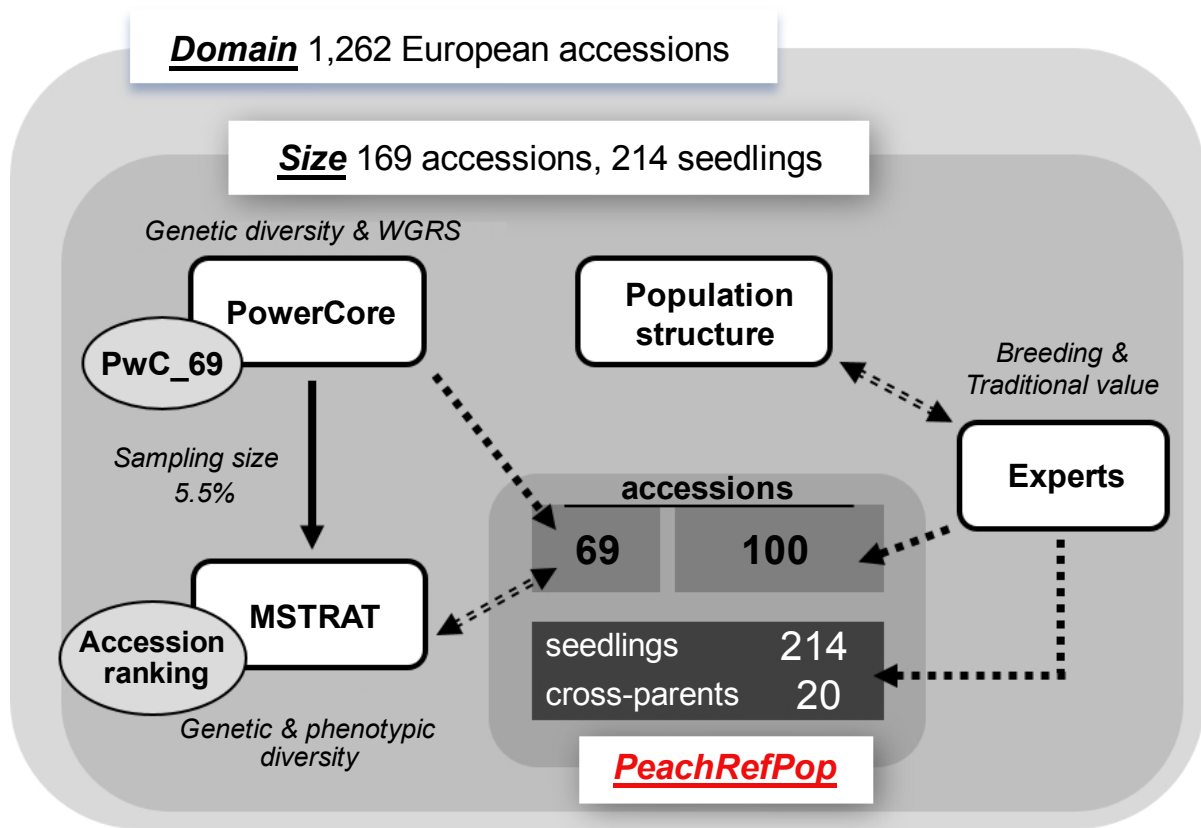


Figure 2. Graphical summary of the overall scheme followed for selecting the *PeachRefPop* collection.

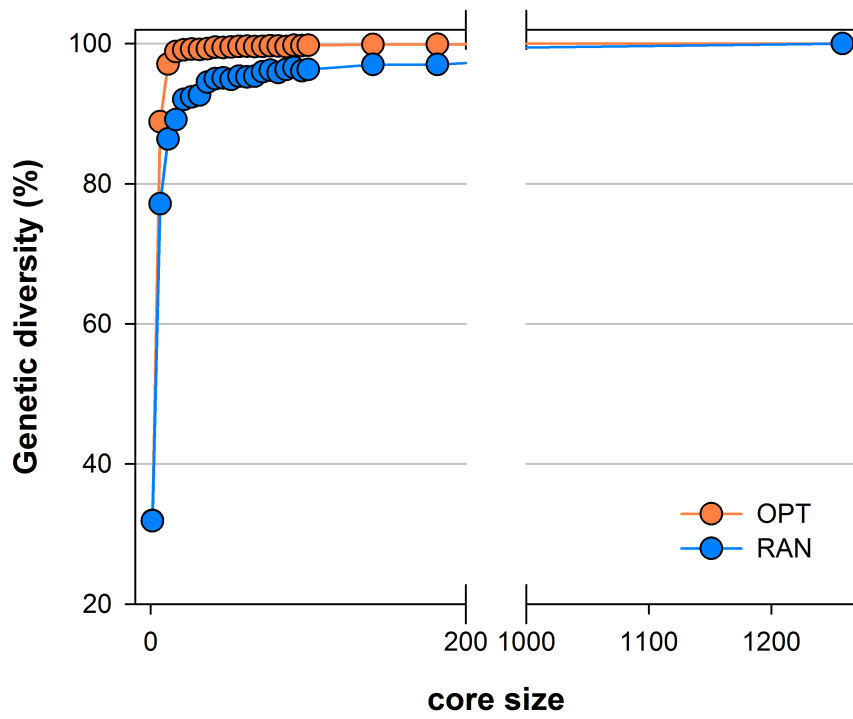


Figure 3. Redundancy in starting panel (EU_1262) estimated with MSTRAT software using a set of 445 SNPs. Datapoints represent averaged values over 5 independent repetitions for each size.

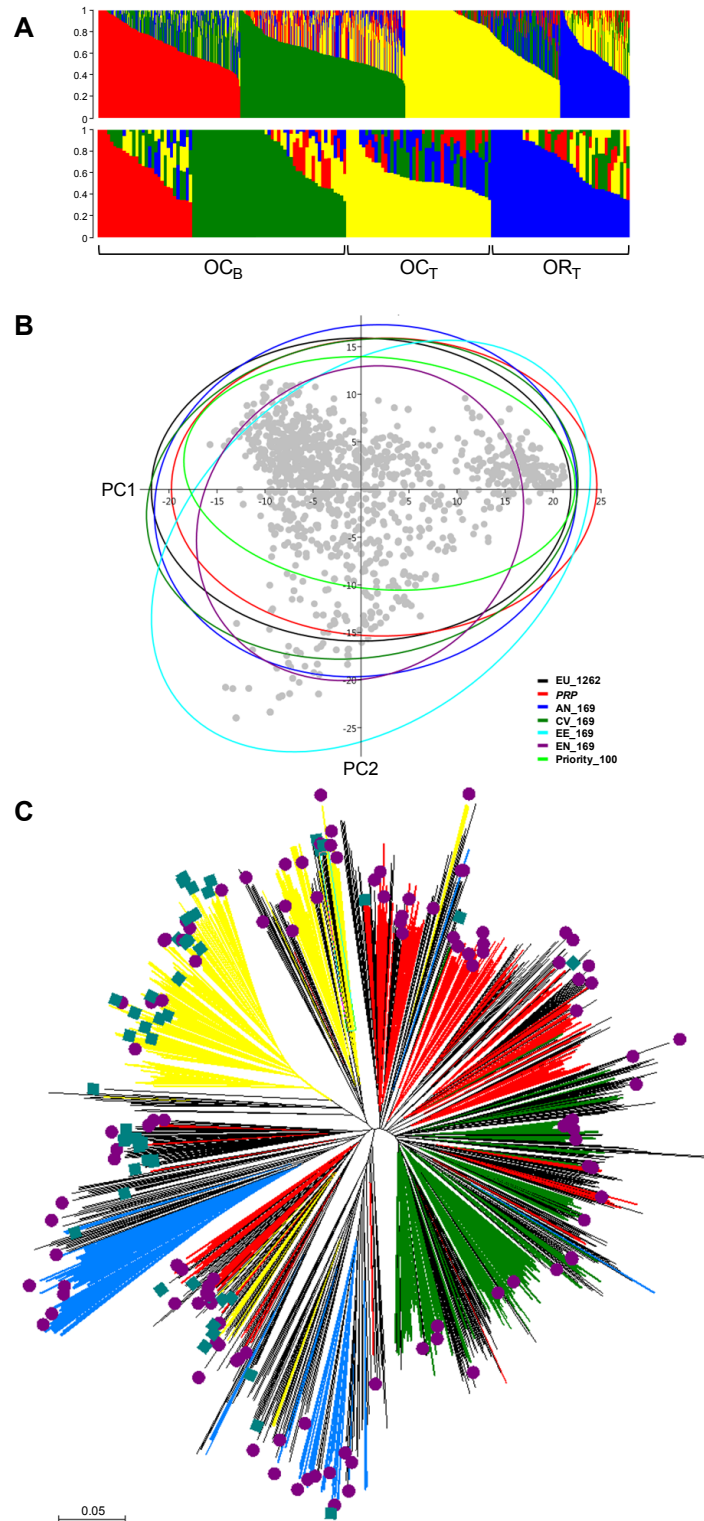


Figure 4. A) Population structure estimated in the whole panel (EU_1262) and *PeachRefPop*, as estimated for K (number of a priori cluster) equal to 4; B) PCA analysis of the subsets with a core size of 169 entries. Scores for each accession were obtained from the work of [Micheletti et al., 2015](#). The 95% confidence ellipses in the scatter plot were estimated using PAST software. C) NJ phylogenetic tree. Blue navy squared indicated accessions with traditional and historical value, violet circle indicates the other *PeachRefPop* accessions, while colors reflect the population structure.

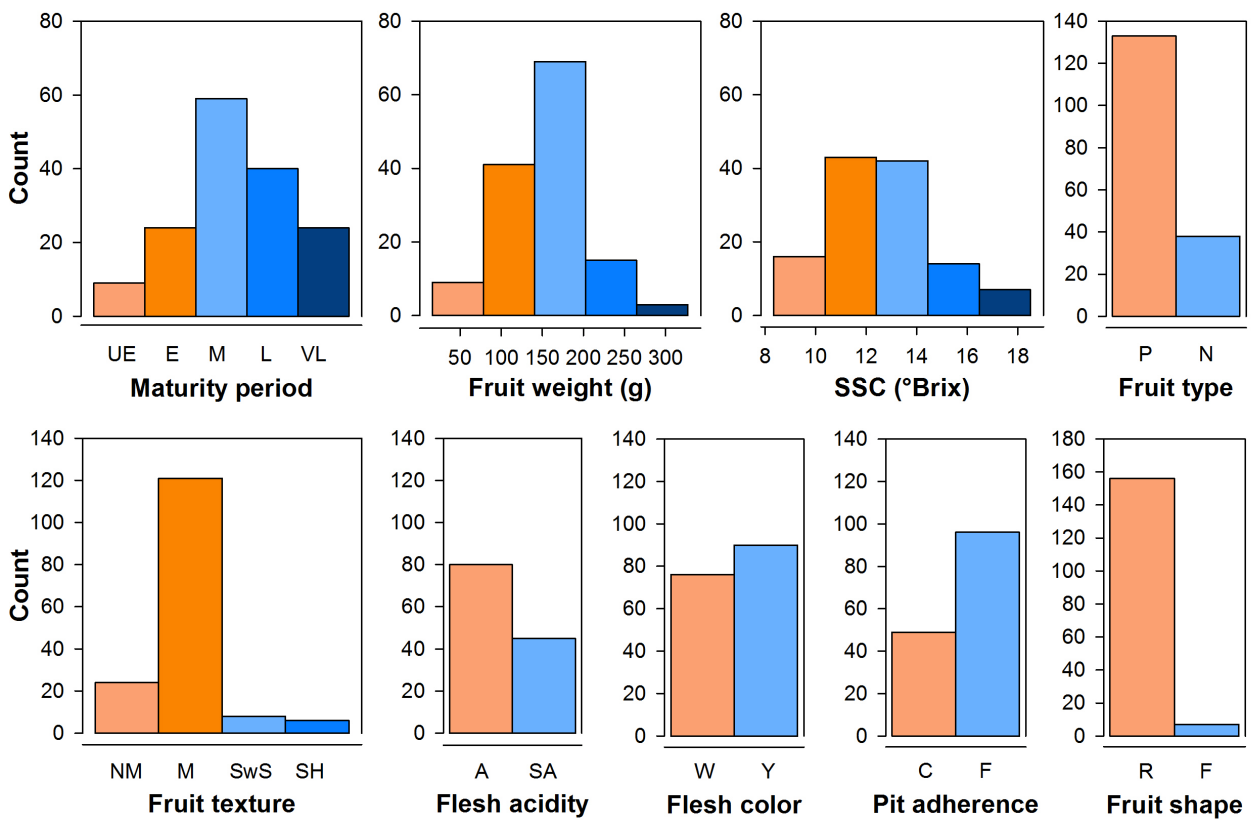


Figure 5. Distribution of main phenotypic traits in the *PeachRefPop* collection.

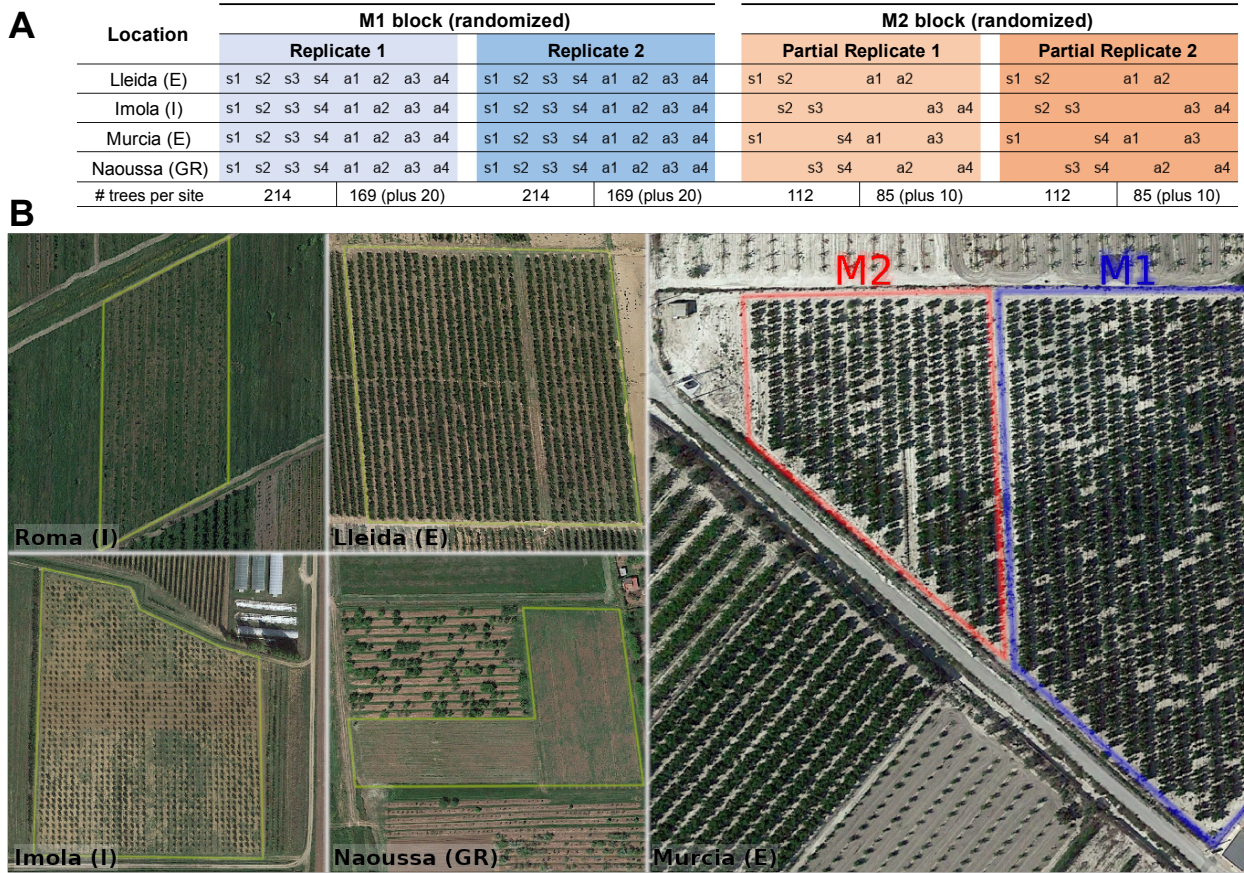


Figure 6. A) Experimental design and B) Google maps satellite imageries of the established *PeachRefPop* orchards across the different European sites.