

This document is confidential and is proprietary to the American Chemical Society and its authors. Do not copy or disclose without written permission. If you have received this item in error, notify the sender and delete all copies.

An approach for magnetic halloysite nanocomposite with selective loading of SPION in the lumen

Journal:	<i>Inorganic Chemistry</i>
Manuscript ID	ic-2020-01039g.R2
Manuscript Type:	Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Hamza, Hady; University of Milan, chemistry Ferretti, Anna Maria; Consiglio Nazionale delle Ricerche, Istituto di Scienze e Tecnologie Molecolari, Laboratorio di Nanotecnologie Innocenti, Claudia; Università di Firenze and INSTM Research Unit, Dipartimento di Chimica Fidecka, Katarzyna; Università degli Studi di Milano, Dip. Chimica Licandro, Emanuela; University of Milan, Department of Organic and Industrial Chemistry Sangregorio, Claudio; CNR, ICCOM Maggioni, Daniela; Università degli Studi di Milano, Dip. Chimica

SCHOLARONE™
Manuscripts

An approach for magnetic halloysite nanocomposite with selective loading of SPION in the lumen

Hady Hamza,^a Anna Maria Ferretti,^b Claudia Innocenti,^{c,d,e} Katarzyna Fidecka,^a Emanuela Licandro,^a Claudio Sangregorio,^{c,d,e,*} Daniela Maggioni^{a,d,*}

^a Dipartimento di Chimica, Università degli Studi di Milano, Via Golgi 19, 20133 Milano, Italy

^b SCITEC-CNR, Sede Secondaria via G. Fantoli 16/15, 20138 Milano, Italy

^c ICCOM-CNR, via Madonna del Piano 10, 50019 Sesto Fiorentino, Italy

^d Consorzio INSTM, Via G. Giusti, 9, 50121, Firenze, Italy

^e Dipartimento di Chimica, Università degli Studi di Firenze, via della Lastruccia 3, 50019 Sesto Fiorentino, Italy

* Corresponding author: daniela.maggioni@unimi.it

Abstract

We present for the first time a method for the preparation of magnetic halloysite nanotubes (HNT) by loading of pre-formed superparamagnetic magnetite nanoparticles (SPION) of diameter size ~ 6 nm with a hydrodynamic diameter of ~ 10 nm into HNT. We found that the most effective route to reach this goal relies on the modification of the inner lumen of HNT by tetradecylphosphonic acid (TDP) to give HNT-TDP, followed by the loading with pre-formed oleic acid (OA) stabilized SPIONs. Transmission electron microscopy (TEM) evidenced the presence of high crystalline magnetic nanoparticles only in the lumen, partially ordered in chain-like structures. Conversely, attempts to obtain the same result by exploiting either the positive charge of the HNT inner lumen employing SPIONs covered with negatively charged capping agents or the *in situ* synthesis of SPIONs by thermal decomposition were not effective.

HNT-TDP were characterized by infrared spectroscopy (ATR-FTIR), thermogravimetric analysis (TGA) and ζ -potential, and all of the techniques confirmed the presence of TDP onto the HNT. Moreover, the inner localization of TDP was ascertained by the use of Nile Red, a molecule whose luminescence is very sensitive to the polarity of the environment. The free SPION@OA (as a colloidal suspension and as a powder) and SPION-in-HNT powder were magnetically characterized by measuring the ZFC-FC magnetization curves as well as the hysteresis cycles at 300 and 2.5 K, confirming that the superparamagnetic behavior and the main magnetic properties of the free SPIONs were preserved once embedded in SPION-in-HNT.

Keywords: Halloysites nanotubes, SPIONs, tetradecylphosphonic acid, magnetism

1. Introduction

Halloysite nanotubes (HNT) are unique natural nanomaterials composed by a double layered aluminosilicate with a hollow tubular structure (Figure 1a) in the micro range and an inner diameter usually ranging between 15 and 50 nm (Figure 1b).

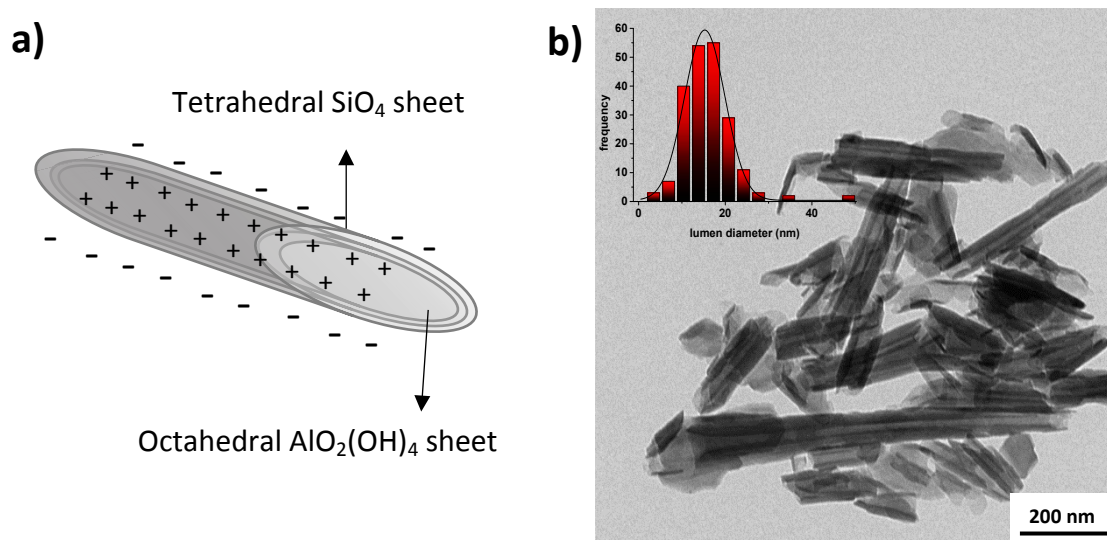


Figure 1. (a) Schematic representation of the structure of a halloysite with the indication of the molecular geometry around Al and Si centers specific for each layer and (b) TEM micrograph of a HNT sample (inset: HNT lumen diameter distribution).

The chemical composition of the HNT inner and outer parts is different. Specifically, the inner surface is made by alumina with exposed aluminum hydroxide groups (Al–OH), while the outer one is made by silica [siloxane groups (Si–O–Si)].¹ The structural formula of HNT is $\text{Al}_2(\text{OH})_4\text{Si}_2\text{O}_5 \cdot (\text{H}_2\text{O})_2$ when it is in its hydrated form.² Due to the different chemical composition of the outer and the inner parts, the HNT inner lumen results positively charged due to the protonation of exposed OH groups as well as the possible coordination vacancy at Al^{3+} sites.³ Instead, the outer part is negatively charged, due to the few OH groups only present in the structural defects of the siloxane layer and at the edges of the nanotubes.⁴ The potentiality for the industrial development of HNT-based derivatives is more than probable, due to the low price, eco-friendly properties and to the biocompatibility of this material, as well as facile and selective functionalization of the HNT two layers with different functional groups.^{5,6}

Thanks to these features, the HNT have been already employed as possible drug delivery vectors⁶⁻⁸ for proteins and small drugs,^{9,10} since negatively charged drugs can be easily retained by the positive lumen, which could act as an inorganic nanocapsule for controlled release.^{11,12} Conversely, long oligonucleotide strands, despite the negative charge, cannot be loaded in the HNT inner lumen possibly due to the big size. Nevertheless, they can be delivered by immobilizing them on the outer surface through different strategies.¹³⁻¹⁶ As regards HNT morphology, HNT can be considered as an alternative to multi-walled carbon

1
2
3 nanotubes for certain applications, with the advantage of being naturally available, durable, very cheap (HNT
4 4\$ per kg, while carbon nanotubes 500\$ per kg)¹⁷ and more biocompatible than carbon nanotubes.

5
6 HNT have been also proposed for the delivery of other active chemicals such as anti-corrosion agents and
7
8 flame retardants, as well as nano-reactors for enzymatic biocatalysts and water remediation agents for heavy
9
10 metals ions removal.^{7,18,19}

11
12 Superparamagnetic iron oxide nanoparticles (SPION) made of magnetite (Fe_3O_4) have been deeply studied
13
14 in the last decades especially for applications in the biomedical field.²⁰⁻²⁵ Indeed, their superparamagnetism
15
16 is essential for safe use *in vivo*, since their total magnetization is null in the absence of an applied external
17
18 magnetic field, thus preventing any aggregation event that could cause capillary occlusion. They possess
19
20 many favorable features for both imaging and therapy: first of all, they are biocompatible and
21
22 biodegradable,^{26,27} they can be exploited as contrast agents in magnetic resonance (MRI), as drug delivery
23
24 carriers, for various separating techniques, as heat mediators for magnetic fluid hyperthermia (MFH)
25
26 treatments, among others.²⁸ SPION have also been extensively used for triggering the drug release from
27
28 several different nanocomposites²⁹⁻³² or nanoporous systems.³³

29
30 Due to the biocompatibility of both HNT³ and SPION, many previous research studies focused on the
31
32 preparation of HNT-SPION nanocomposites, but in the most of the cases the SPION were anchored on the
33
34 outer surface of HNT,³⁴⁻⁴¹ or grown by coprecipitation in the HNT lumen.^{42,43} In this last case the crystallinity,
35
36 the size of the nanoparticles and their magnetic properties were hardly controllable. To the best of our
37
38 knowledge pre-formed magnetic NPs have never been selectively loaded in the inner lumen of HNT so far.

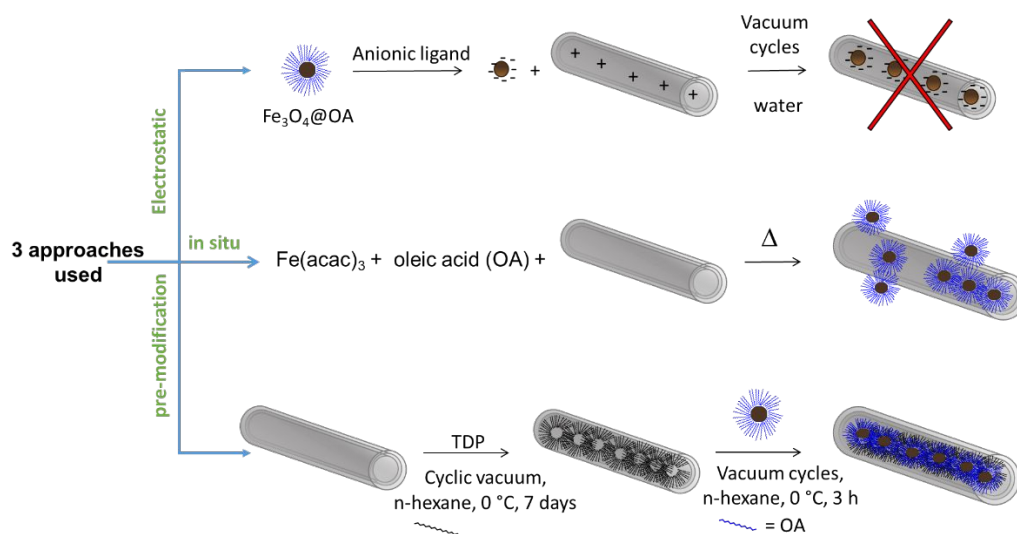
39
40 On the other hand, the loading of pre-formed SPION with shaped properties would be of interest for many
41
42 applications, such as MRI or MFH. Moreover, the inner loading could give rise to a higher oxidative resistance
43
44 of NPs compared to their anchoring on the HNT external surface. In fact, HNT could act as a protective barrier
45
46 slowing down the molecular oxygen action, as recently previewed for carbon nanotubes filled with SPION.⁴⁴

47
48 In the literature, there are many examples of *in-situ* formed metallic, quantum dot, oxide NPs, selectively
49
50 loaded onto or into HNT,⁴⁵ while only very few examples report about loading of pre-formed nanoparticles
51
52 (NPs) in the HNT lumen, and, all of them deal with non-magnetic NPs. Specifically, one concerned silver NPs
53
54 of 2 nm diameter stabilized by citrate anions loaded by exploiting the opposite charge of the positive HNT
55
56 lumen and negative NPs.⁴⁶ The same procedure was recently exploited for the loading of palladium NPs.⁴⁷ In
57
58 another example, 2 nm carbon nanodots were loaded into the lumen by exploiting the vacuum pouring
59
60 technique.⁴⁸ In all of the other studies the NPs were made grow in the presence of HNT,⁴⁹ but in many cases,
the growing NPs interacted with the outer surface instead of the inner one. Only few examples describing
the selective inner growth have been reported up to now: the first one showed the ability of Au NPs to
selectively form in the HNT lumen;⁵⁰ the same was achieved for Ru-based metallic clusters,⁵¹ while the last
reported about the possibility to grow iron oxide NPs made by a coprecipitation method.⁴³

The goal of this work was to find a reliable and reproducible method to selectively fill the HNT lumen with pre-formed SPION possessing well-defined and possibly good magnetic properties. The final aim of this study was to obtain HNT-SPION as a "building block" for further developments of suitable nanocomposites. The nanocomposites based on SPION-in-HNT could be useful for different applications, such as i) in biomedical field, as new theranostic agents for MRI, MFH and controlled release of a drug by an external magnetic stimulus; ii) in catalysis, with the double advantage to recover the system due to the magnetic NPs in the inner lumen leaving the external HNT surface available for further decoration with catalytic organometallic compounds or other types of nanoparticles; iii) in water remediation, by exploiting a capturing agent or a photo-reactive organometallic compound anchored on the external surface while maintaining the ability to magnetically recover the nanocomposite; iv) in tissue engineering for the development of 3D scaffolds able to align cells of an anisotropic growing tissue.

2. Results and discussion

In order to fill the HNT with magnetic NPs, we investigated three distinct approaches (depicted in Scheme 1): the electrostatic interaction, the *in situ* formation of SPION, and the pre-modification approach. All these three strategies are described in the next paragraphs, even though only the third one gave us the desired HNT-SPION adduct.



Scheme 1. Schematic depiction of the three distinct approaches followed in this study to obtain HNT-SPION nanocomposite with the selective loading of SPION in the inner part.

2.1 Attempts to prepare SPION-in-HNT using water-dispersible negatively charged SPION: the electrostatic approach. The first method tried to exploit the charge difference between the inner and the outer part of the HNT. Indeed, the inner layer of alumina remains positively charged up to pH 8.5, whilst the external silica layer is negatively charged for almost all the pH range above 1.5.³ This charge difference has been extensively

1
2
3 exploited for the loading of small negatively charged molecules into HNT inner lumen and their forward
4 sustain release at intended site.⁵² Hence, despite the overall negative charge of HNT at physiological pH, the
5 inner alumina remains positively charged to a certain extent. As already mentioned in the Introduction, just
6 few examples were reported on the loading of pre-formed NPs in the HNT lumen.⁴⁶⁻⁴⁸ One of them was
7 carried out using small negative Ag NPs.⁴⁶ In that case, the prepared NPs were tiny (ca 2.6 nm), the loading
8 procedure was extremely simple and involved the use of neither sonication nor vacuum cycles to induce the
9 NPs to enter into the HNT lumen, so that it can be concluded the diffusion by Brownian motions was effective
10 enough to fill the HNT lumen with NPs.

11
12 We tried to follow the same approach but applying vacuum/N₂ cycles in order to exploit the strong capillary
13 pressures affecting the HNT lumen, and taking into account that the HNT inner diameter measured by TEM
14 was $\sim 15.3 \pm 0.3$ nm. Hence, we prepared negatively charged SPION of a suitably small size, ranging from 5.1
15 to 6.9 nm. This size was judged to be a good compromise between the need of employing NPs small enough
16 to enter the lumen but, at the same time, big enough to maintain good magnetic properties. We adopted
17 different approaches for the SPION synthesis (resumed in Table 1), passing from a simple coprecipitation
18 method⁵³ to thermal decomposition syntheses.^{54,55} Indeed, one of the main goals of this work was the
19 development of a strategy to fill HNT with the best NPs in terms of magnetic properties. It is well known that,
20 for SPION, the best magnetic properties are achievable through thermal decomposition syntheses, able to
21 modulate size and crystallinity.⁵⁶

22
23 In Table 1 we reported the data of the several essays we made with the electrostatic attraction approach.
24 These implied a slight variation of the SPION dimension, but most of all, the variation of the surface capping
25 ligands that in vain were changed, trying to encourage the entry of negatively charged SPION into the
26 nanotube lumen avoiding early SPION aggregation. Despite all the efforts made, none of the attempted
27 procedures was successful. All the experimental details, a related discussion and figures (Figures S1-S8), are
28 reported in the Supporting Information.

29
30 We therefore concluded that the electrostatic attraction, useful for small molecules loading in HNT lumen,
31 ^{10,11, 52} as well as for tiny non-magnetic NPs,⁴⁶⁻⁴⁸ was not effective in the case of these magnetic nanoparticles,
32 regardless of the SPION size or coating. This could be due to the SPION mutual attraction in water, that can
33 be reinforced by additional attractive forces such as hydrogen bonds between different particles, especially
34 when they are in the restricted space volume at the entrance of the halloysite lumen in conditions of reduced
35 pressure.

Table 1. Mean size (diameter, nm) of synthesized SPION as measured by TEM and DLS on water suspensions. The synthetic method used is indicated together with the relative reference (DMSA = dimercaptosuccinic acid; GA = gallic acid; TMAOH = tetramethylammonium hydroxide; PA = protocatechuic acid; OA = oleic acid).

Sample ID	Ref	NPs Synthesis Method	NP Stabilizer	TEM/nm	DLS/nm (hexane) ^b	DLS/nm (water)
SPION1@DMSA	53	Coprecipitation with NaOH	DMSA ⁵⁷	5.3 ± 1.0	13.1 ± 3.0	13.5 ± 2.4
SPION2@DMSA	53	Coprecipitation without NaOH	DMSA ⁵⁷	--- ^a	10.6 ± 4.0	---
SPION3@DMSA ^c	54	Thermal decomposition using Fe(acac) ₃	DMSA ⁵⁷	5.1 ± 1.6	8.0 ± 1.9	---
SPION4@GA	55	Thermal decomposition using iron oleate	GA ²²			13.0 ± 2.5
SPION4@TMAOH	55	Thermal decomposition using iron oleate	TMAOH ⁵⁸	6.9 ± 1.0	10.7 ± 2.3	10.0 ± 3.0
SPION4@PA	55	Thermal decomposition using iron oleate	PA			13.0 ± 4.0
SPION3@OA ^c	54	Thermal decomposition using Fe(acac) ₃	OA	6.1 ± 1.3	10.9 ± 3.0	---

^a The TEM images were not acquired on this sample.

^b The DLS measurement was performed in n-hexane suspension on the as-prepared SPION@OA.

^c The synthetic procedure for the preparation of SPION@OA was the same, but repeated twice, leading to slightly different.

2.2 Attempts of SPION-in-HNT preparation using thermal decomposition in the presence of pristine HNT.

In this case we tried to carry out an *in situ* thermal decomposition synthesis of SPIONs in the presence of HNT as reported in the literature⁵⁷ (Scheme 1). The chosen method for the synthesis of SPIONs was in principle able to lead to highly magnetized and monodispersed SPIONs into HNT. Thus, the iron precursor and the other reactants were made diffusing into the lumen of HNT reducing the pressure in the vessel for some minutes before starting the reflux, to assure the diffusion into the lumen of the reaction mixture. HNT-SPION adduct was washed thoroughly using ethanol by centrifugation. The light brown powder recovered was magnetic, being attracted by a neodymium-iron-boron magnet and was dispersible in water. Unfortunately, TEM analysis of the obtained product showed that, even if some SPION were localized inside the HNT (highlighted in Figure S2 by red arrows), they were also found attached to the external surface, showing that the growth of NPs was not selectively confined to the HNT lumen. Moreover, SPION were not highly monodispersed and, owing to the effect of high temperature, the outer surface of HNT was partly etched.

2.3 SPION-in-HNT by functionalization of the inner lumen of HNT with tetradecylphosphonic acid (TDP).

The failed attempts obtained by pursuing the two previous paths prompted us to verify whether the SPION could be inserted in their native state, that is with the apolar capping agent (oleate) still on the surface, by properly modifying the lumen polarity of HNT. From the literature, it is known that it is possible to selectively functionalize the lumen of HNT using a phosphonic acid, which preferentially reacts with alumina compared to silica.^{60,61} The inner surface of HNT was successfully modified with tetradecylphosphonic acid (TDP).⁶¹ To ascertain whether the reaction had taken place, we employed several analytical techniques. As a first qualitative assay, we used Nile Red dye. Indeed, this molecule is very sensitive to the environment in which it is dispersed, in that both the absorption and the emission are heavily perturbed.⁶² Nile Red is highly emissive in hydrophobic environments, as the one present in the functionalized lumen with TDP, while it is

1
2
3 completely quenched in water. After the loading of Nile Red (see Experimental Part for details) the HNT-TDP
4 turned purple, hence they were thoroughly washed with water to remove all the possible externally
5 interacting dye. The still wet purple HNT-TDP sample, observed under UV light irradiation, exhibited a red
6 luminescence. On the contrary, the pristine HNT treated in the same way with Nile Red and washed with
7 water turned purple as well, but they did not show any emission under the UV light irradiation, as, on the
8 contrary, the HNT-TDP did. This behavior was ascribed to the different microenvironment in which the dye
9 is lying: in the case of HNT-TDP sample, the Nile Red is likely intercalated into the apolar long aliphatic chains,
10 thus preserving Nile Red from the quenching provoked by its direct interaction with water (Figure S10).

11
12
13 ATR-FTIR spectroscopy (Figure 2a) and thermogravimetric analysis (TGA, Figure 2b) confirmed the
14 successful functionalization of halloysites. The ATR-FTIR spectrum of HNT-TDP (Figure 2a, top), as well as the
15 one of pristine HNT (Figure 2a, middle), showed the characteristic sharp bands related to Al-OH stretching of
16 lumen and interlayer alumina peaking at 3692 and 3624 cm^{-1} , respectively, and of the OH stretching of
17 hydrogen-bonded water present in the interlayers (3551 cm^{-1}).⁶⁰ The spectrum of HNT-TDP showed the
18 stretching bands of the aliphatic chain in the region between 3000-2800 cm^{-1} (2959, 2919 and 2851 cm^{-1}),
19 together with the deformation (scissoring) of CH_2 groups (1468 cm^{-1}), while the bands due to the C-P-O
20 stretching in the region 1100 – 800 cm^{-1} overlapped with the very intense bands of HNT, hampering their
21 visualization and attribution. Finally, the broad P-O-H band for the unbound TDP visible in the TDP spectrum
22 (bottom) at $\sim 2300 \text{ cm}^{-1}$ as well as the complete disappearing of the P=O band vibration at 1211 cm^{-1} suggest
23 that TDP is bonded to the halloysites, and in particular to the alumina lumen layer, in a deprotonated
24 form.^{60,61}

25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

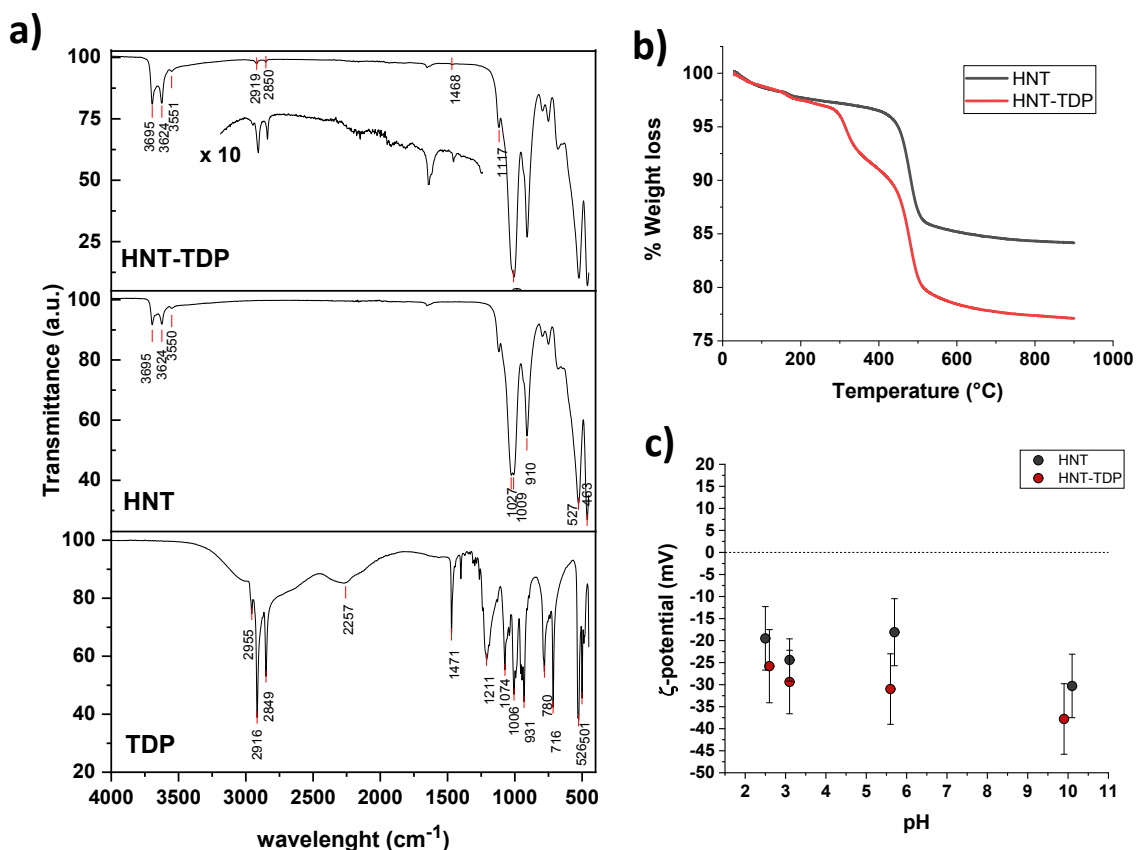


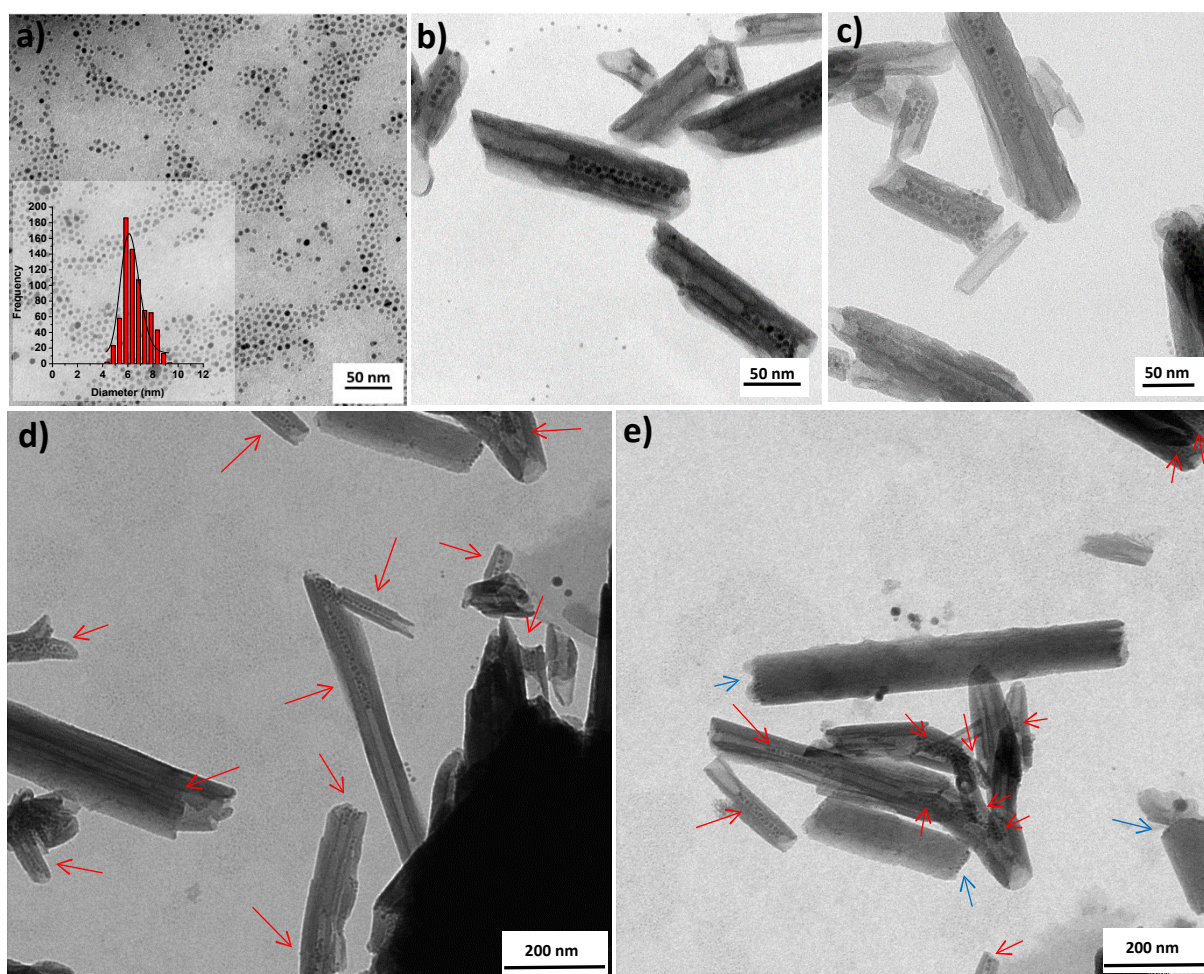
Figure 2. Characterization of the HNT-TDP (a) FTIR spectrum of HNT-TDP (top; the inset shows the magnified region of the spectrum containing signals of TDP) compared with the FTIR spectra of HNT (middle) and TDP (bottom); (b) thermogravimetric analysis (TGA) of pristine HNT and HNT-TDP derivative; (c) ζ -potential analyses of suspensions of HNT and HNT-TDP.

TGA analysis of HNT (black trace of Figure 2b) showed a mass loss step (ca. 2.5 %) corresponding to the loss of adsorbed water on surface (onset $T = 45$ °C) and into interlayer (onset $T = 163$ °C), and a second mass loss step (ca. 14.6%, onset $T = 450$ °C, inflection point at 480 °C) assigned to the dehydroxylation of structural AlOH groups of halloysites.⁶³ Differently, the TGA profile of HNT-TDP (red trace of Figure 2b) showed an extra mass loss step (ca. 5.4 %, onset $T = 300$ °C, inflection point at 320 °C) attributed to the degradation of TDP.

We carried out also the analysis of ζ -potential before and after the treatment with TDP and the obtained values as a function of pH are shown in Figure 2c. The observation of a more negative value for HNT-TDP is not only a further confirmation of the interaction of the HNT with the TDP but, indirectly, it also suggests the TDP localization is in the inner part of the nanotube. Indeed, if the phosphonic acid mostly interacts with the positively charged inner part, it neutralizes a portion of these charges, thus making slightly more negative the whole nanotube, as experimentally observed.

The preparation of SPION-in-HNT nanocomposite was achieved using SPION3@OA nanoparticles, obtained by the same thermal decomposition method⁵⁵ used previously (see § 2.1), that this time afforded magnetic

1
2
3 NPs with 6.1 ± 1.3 nm mean diameter as shown by transmission electron microscope (Figure 3a) and a
4 hydrodynamic diameter of 10.9 ± 3.0 nm (Figure S11).
5
6
7



39 **Figure 3.** TEM micrographs of (a) as-synthesized SPION@OA (in the inset the SPION diameter distribution); (b-e)
40 SPION-in-HNT prepared by pre-functionalization of HNT with TDP in the lumen. Panels (b) and (c) show images
41 taken at higher magnification compared to (d) and (e). Red arrows mark the HNT containing SPION. Blue arrows
42 indicate SPION at the edge of few HNT.
43

44 The as-synthesized SPION@OA were effectively loaded into the HNT lumen by using repeated vacuum/N₂
45 cycles under stirring while keeping the temperature at 0 °C with an ice bath to avoid the heavy evaporation
46 of the volatile solvent. The procedure was repeated until the light brown color of the suspension (due to
47 SPION@OA) turned transparent, and conversely the deposited HNT turned from white to brown. The SPION
48 loading was selectively directed into the inner part of HNT, without any relevant interaction with the HNT
49 outer surface, as clearly shown by TEM analysis (Figure 3b-e). This result shows that the size and the nature
50 of coating of SPION have a major role in successful loading into the inner lumen of HNT. Despite the fact that
51 SPIONs used in the electrostatic approach (see above § 2.1) were in the size range 5.1-6.9 nm, which is less
52 than the half of the inner lumen diameter, no loading was observed when they were coated with DMSA or
53 the other polar molecules. On the contrary, SPION@OA were able to reach the inner lumen probably mostly
54
55
56
57
58
59
60

1
2
3 due to the apolar weak interactions between the aliphatic chains on NPs (OA) and in the inner lumen of the
4 nanotubes (TDP). The TDP functionalization is also mandatory to observe the effective loading of SPION.
5 Indeed, treating SPION3@OA with pristine HNT resulted in just sporadic loading into HNT, as clearly showed
6 by TEM images (Figure S12).
7
8

9
10 Unfortunately, the commercial pristine HNT used in this work were of low quality if compared with other
11 HNT deriving from other mines. In some images we evidenced the presence of unrolled HNT or kaolin-like
12 sheets (see Figure S13). In Figure 3e there is a marked difference between the usual HNT, filled with SPION,
13 that presents a lumen and walls clearly distinguishable (indicated with red arrows), and some peculiar HNT
14 (highlighted with light blue arrows) that have a much larger diameter and do not show clearly the lumen, as
15 if they are not empty, thus hampering the entrance of SPION. In this case some SPION interacted
16 preferentially with the HNT edges rather than with the lumen. Moreover, when we tried to completely fill all
17 the HNT by doubling the SPION amount, we noticed by TEM analysis (Figure S14) that only a part concurred
18 to increase the amount of SPION into HNT, the remaining been captured by the kaolin-like sheets.
19
20
21
22
23
24

25 **2.4 Australian HNT with a larger lumen filled with apolar SPION@OA.**

26
27
28 Although very promising, the results obtained so far are not totally satisfying in terms of the amount of
29 SPION embedded in the HNT. We ascribed this behavior to the poor quality of the commercial HNT batch,
30 containing some large and apparently filled HNT. In order to verify this hypothesis, we considered a new
31 sample of HNT derived from the Camel Lake mine (Australia). These HNT are characterized by a higher
32 regularity than Aldrich HNT (Figure S15), with a larger lumen (23.8 ± 6.0 nm), even though they are on average
33 longer (770 ± 300 nm). As in the case of the commercial HNT, we made apolar the inner lumen of Australian
34 HNT (A-HNT from now on) by a selective functionalization with TDP. Also in this case the SPION@OA were
35 loaded selectively within the A-HNT@TDP, but only when a maximum amount of 0.4 mg SPION per 10 mg of
36 HNT was used (Figure 4). As for the commercial HNT, the SPION did not fill completely the lumen.
37 Nevertheless, when we tried to increase the SPION / HNT mass ratio, we found that SPION were massively
38 localized also outside the lumen (Figure S16). At the same time, some semi-full or apparently empty HNT
39 were visible, although the lumen of the A-HNT was on average wider, such as not to prevent the entrance of
40 NPs of 5-6 nm diameter. We think that the ability of air to escape from the HNT is essential to fully fill a
41 nanotube. Indeed, in some TEM images air bubbles seem to be present within the inner lumen where SPION
42 were not localized (see Figure 4, blue arrows). Moreover, to be more effective in the removal of air contained
43 in the HNT lumen, we applied a pre-vacuum cycle to the HNT before adding the SPION. In that case, we did
44 not find a significant number of particles in the lumen, but all external to the HNT (Figure S17). This evidence
45 made us conclude that the entry of SPION in the lumen takes place immediately after the break of the vacuum
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

and the restoration of the ambient pressure in the reaction vessel.⁴⁸ Obviously, the proper concentration of NPs must be present in order to make the loading as effective as possible.

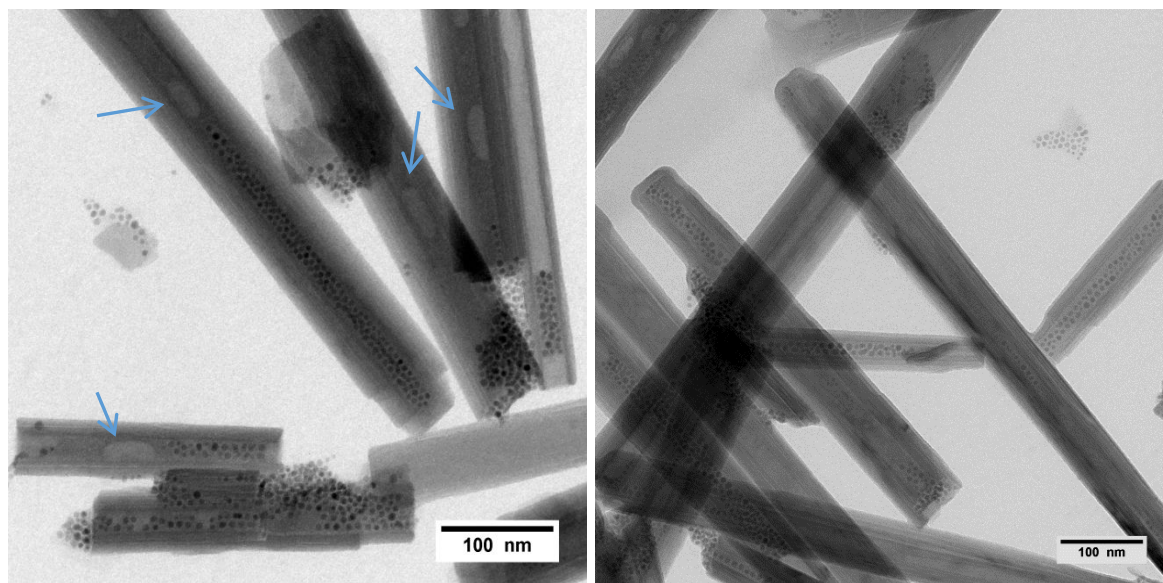


Figure 4. TEM micrographs of SPION-in-A-HNT (A-HNT= Australian Camel lake halloysites). Blue arrows mark the HNT containing possible air bubbles.

2.5 Magnetic properties characterization of SPION-in-HNT nanocomposite. Magnetic measurements performed on SPION and SPION-in-HNT demonstrated the nanoparticles display the typical superparamagnetic behavior expected for a set of nanoparticles of this size and that the main magnetic properties have been maintained after the SPION embedding in HNT. The pristine SPION were measured as a hexane solution (SPION-sol) and dried powder (SPION-pow). This last measurement was also used to estimate and remove the diamagnetic contribution of the solvent, thus obtaining reliable magnetization saturation (M_s) value. The M_s of SPIONs was estimated as the magnetic moment recorded at the highest experimental field applied (50 kOe) divided by the iron content of SPION-sol evaluated by AAS analysis (2.2 % w/w). As expected for systems of reduced size, the M_s value of the SPIONs (72 emu/g Fe at 300 K and 82 emu/g Fe at 2.5 K) is lower than the bulk value of magnetite, but it is quite high for 6 nm nanoparticles, where, normally a considerable ratio of disordered spins at the surface to ordered spins in the core is observed.⁶⁴ The ZFC/FC magnetization curves (Figure 5a) evidenced the superparamagnetic behavior of the samples, characterized by a maximum in the ZFC curve, the position of which determines, as a rough approximation the blocking temperature (T_b) of the system, and by a thermal irreversibility at low temperature. It is interesting to note how the T_b of the pristine SPIONs increases from solution (18.8 K) to powder (32.3 K), while it reaches an intermediate value for SPION-in-HNT (29.3 K). The increase of T_b for a nanoparticle set measured in different conditions is generally ascribed to the enhanced inter-particle interactions.⁶⁵ In the case of SPION-sol and SPION-pow, for instance, the dipolar interactions are stronger in

the powder sample as the NPs are much closer each other with respect to the solution sample. Assuming that the embedding procedure of SPIONs in HNT induced only negligible modification in the magnetic anisotropy of the pristine SPIONs (see Figure 5a), the value of T_B observed for HNT-SPION suggests that the embedded nanoparticles are moderately interacting, as it is between that of SPION-sol (low interactions) and SPION-pow (strong interactions), closer to the latter. This result is consistent with the expected confinement of the SPIONs in the HNT lumen, that implies a reduction of the mean distance among the SPIONs with respect to a diluted solution.

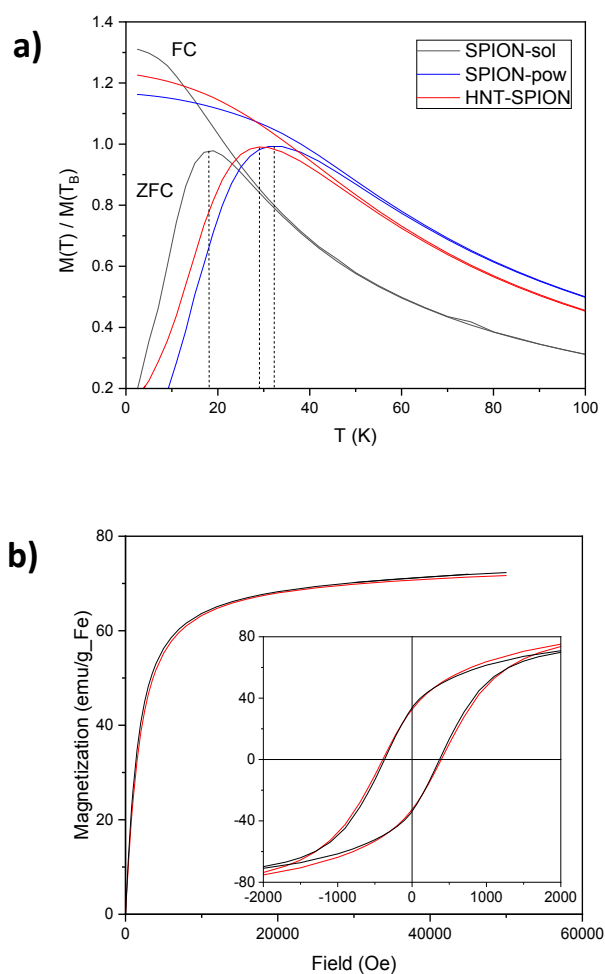


Figure 5. a) ZFC (lower curve) and FC (upper curve) magnetizations acquired with a 50 Oe field. The vertical dashed lines remark the position of the ZFC maxima corresponding to the blocking temperatures (T_B). For a better presentation, each curve was normalized to the magnetization value at the corresponding T_B ; **b)** Magnetization curves at high (300 K, main panel) and low (2.5 K, inset) temperature for SPIONs (black line) and SPION-in-HNT (red line).

As expected for a superparamagnetic system, the samples present magnetic hysteresis only for temperature below T_B , as shown in Figure 5b, where the magnetization curves, M vs H , are reported. At low

1
2
3 temperature (2.5 K), both samples exhibit similar coercivity ($H_c = 370$ Oe for SPIONs and $H_c = 395$ Oe for
4 SPION-in-HNT) and remanence (34 and 32 emu/g Fe, for SPIONs and SPION-in-HNT, respectively), that reduce
5 to zero at high temperature (300 K). The similarity of the curve shape acquired before and after HNT
6 embedding confirms that the loading procedure does not alter significantly the magnetic properties of the
7 pristine SPIONs. The maintenance of the magnetic properties allowed us to accurately estimate the
8 concentration of SPIONs in the SPION-in-HNT simply by the ratio of the saturation magnetization of the two
9 samples. Actually, in our case, the comparison is very accurate, as the same ratio is found at all the magnetic
10 fields because the whole normalized magnetization curves are perfectly superimposable.⁶⁶ The iron
11 concentration obtained by the superimposition of the curves at high temperature is 5.3 % w/w, which is very
12 close to the concentration that would be achieved if all the SPIONs in the preparation solution were loaded
13 in the HNT lumen (5.5 % w/w), corresponding to an encapsulation yield of 96 % and to a Fe_3O_4 % w/w of 7.3
14 %. Furthermore, the total magnetization of the SPION-in-HNT is enough (3.82 emu/g) to guarantee a
15 magnetic response to an appropriate magnetic gradient, allowing the use of the compound in several
16 practical applications. This result underlines the efficacy of the proposed NP embedding procedure.
17
18
19
20
21
22
23
24
25
26
27

28 **3. Conclusions**

29
30
31 In summary, we have successfully prepared a novel magnetic halloysite nanocomposite with apolar SPIONs
32 selectively loaded in the inner lumen of pre-functionalized HNT, exploiting vacuum- N_2 cycles. The new SPION-
33 in-HNT nanocomposite has been morphologically characterized by TEM as well as from the magnetic point
34 of view. SPION@OA in the inner lumen of HNT did not change their magnetic properties, retaining the
35 superparamagnetic character at 300 K with a moderate inter-particle interaction once charged into the
36 lumen of the nanotubes, and a sufficient total magnetization such that the SPION-in-HNT obtained with this
37 embedding procedure can respond to an external magnetic stimulus. The SPION-in-HNT nanocomposite was
38 successfully obtained also using HNT of different source (A-HNT) endowed with a bigger lumen diameter.
39
40
41
42
43

44 Despite this more favorable characteristic for the NP loading, even in this case the NPs did not completely
45 filled all the HNT, suggesting that, rather than the size of the nanoparticles, it is the ability to release the
46 initial air contained in the nanotubes to play a fundamental role in allowing the NP to enter.
47
48

49 Attempts to load pre-formed negatively charged SPIONs, exploiting the opposite charges of the HNT lumen,
50 failed due to an early aggregation of the single NPs once subjected to an external force, which pushed them
51 at the entrance of the lumen.
52
53

54 Finally, preliminary results on the inclusion of preformed Au nanoparticles of ≈ 5 nm coated with oleylamine
55 (data not shown), suggest the procedure here proposed could be used as a general method for the loading
56 of other kinds of inorganic nanoparticles stabilized with a hydrophobic layer. Depending on the nature of the
57
58
59
60

loaded NPs the applications could span from catalysis to drug delivery or to waste water treatment, just to mention a few.

4. Experimental Part

4.1 Materials and instruments. Anhydrous FeCl_3 and $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$ (Merck Germany), ethanol 99.8% (Merck UK), oleic acid 90% (Alfa Aesar), n-hexane 96% and acetone (Scharlau), iron acetyl acetonate 97%, 1,2-hexadecandiol 90%, Tetradecylphosphonic acid 97%, DMSA 98%, triethylamine 99%, 16-Phosphonohexadecanoic acid 97%, Protocatechuic acid 97%, Gallic acid >97.5 %, Tetramethylammonium hydroxide pentahydrate 97%, Sodium hydroxide pellets 98%, Oleylamine 70%, diphenyl ether 99%, benzyl ether 98% and halloysite nanotubes HNT (Sigma Aldrich), Nile red >99% (Abcr) were all used as received without further purification. Toluene (99.5%, Sigma Aldrich) was distilled from sodium under nitrogen prior to use. Ultrapure milli-Q water (Millipore, resistivity=18M $\Omega \text{ cm}^{-2}$) was used for the preparation of the aqueous solutions.

ζ -potential measurements were carried out using a Malvern Zetasizer nano ZS instrument equipped with a 633 nm solid state He–Ne laser at a scattering angle of 173° , typically dissolving samples at a concentration of 1 mg/mL or less at 25°C . The measurements were averaged on at least three repeated runs.

Iron content on SPION was determined by different methods. AAS analysis was carried out on a Perkin-Elmer Pinaacle 900 instrument for SPION6@OA. For all the other preparations, a spectrophotometric method was employed on an Agilent model 8543 spectrophotometer at room temperature and using disposable cuvettes with 1.0 cm path length. For Fe determination by AAS, few microliters of the particle suspension were digested with 1 mL aqua regia/HCl overnight at room temperature in a 10 mL volumetric flask, and subsequently filled up with MilliQ water. For spectrophotometric analyses few microliters of the particle suspensions were digested with 1 mL aqua regia/HCl at room temperature in a 10 mL volumetric flask. Subsequently, they were filled up with (i) 0.1 mL NH_2OH solution (10% w/w), (ii) 6 mL acetate buffer solution (pH 4.6, 0.15M), (iii) 0.2 mL 1,10-phenantroline solution (0.6% w/w, $\text{H}_2\text{O}/\text{MeOH}$ 10 : 1) (iv) milliQ water until the volume of 10 mL was reached. The absorbance of the band centered at 510 nm due to $\text{Fe}(\text{phen})_3^{2+}$ complex was then measured and the concentration of Fe was finally obtained, acquiring a calibration curve, which was obtained with standards prepared with the same procedure starting from a commercial AAS standard solution.

Thermogravimetric analysis (TGA) was carried out in air atmosphere and in the temperature range $50\text{--}800^\circ\text{C}$ with a heating rate of $5^\circ\text{C}\cdot\text{min}^{-1}$, using a Mettler-Toledo thermogravimetric balance (TGA/DSC 2 Star[®] System) on ca 10 mg of lyophilized samples.

ATR-FTIR spectra were acquired on a PerkinElmer Frontier instrument equipped with an ATR accessory with a diamond/ZnSe crystal. The IR spectra were registered between 4000 and 400 cm^{-1} .

1
2
3 Transmission Electron Microscopy (TEM) TEM micrographs were collected using a Zeiss Libra 200 FE
4 instrument equipped with an in column pre-aligned omega filter that improves the contrast and a Schottky
5 field emission gun at 200 kV. Alternatively, TEM images were collected using an EFTEM LEO 912AB (Zeiss) at
6 100 kV. Samples were prepared by dropping a dilute solution of the samples onto 200 mesh carbon-coated
7 copper grids, and after blotting the excess of water, the samples were let drying at least for 24 h in air. The
8 nanoparticle size was measured by Pebbles and Pebbles-Juggler⁶⁸ or by Image-J free software. Imaging
9 Platform software (Olympus). For each sample the measured NPs were around 400.

10
11
12
13
14
15 Magnetic measurements were carried out by a SQUID magnetometer from Quantum Design Ltd. Powder
16 samples (SPION-pow and SPION-in-HNT) were prepared by enclosing a small amount of powder in a Teflon
17 tape; solution sample (SPION-sol) was measured in a gel cap. The diamagnetic contribution of Teflon was
18 found negligible, while that of the cap and solvent was evaluated as the magnetization component, linear
19 with the magnetic field, needed to be added to the SPION-sol magnetization in order to obtain the same
20 slope of the SPION-pow curve in the high field region. The magnetization curves were obtained by acquiring
21 the magnetic moment of the sample as a function of the applied magnetic field ranging from 0 to ± 50 kOe,
22 at low (2.5 K) and room temperature (300 K). The Zero Field Cooled (ZFC) and Field Cooled (FC)
23 magnetizations were acquired as a function of the temperature applying a 50 Oe probe field after cooling the
24 sample in the absence (ZFC) and presence (FC) of the probe field.

25
26
27
28
29
30
31
32
33 **4.2 Synthesis of magnetic nanoparticles and attempts for their loading in the HNT lumen.** SPION@OA and
34 their derivatives after ligand exchange with DMSA, GA, PA and TMAOH are described in the Supporting
35 Information, together with the loading attempts of these negatively charged SPION in HNT and the one-pot
36 procedure with thermal decomposition synthesis of SPION in the presence of HNT.

37
38
39
40
41 **4.3 Modification of Halloysite Lumen with Tetradecylphosphonic acid (HNT-TDP).** Halloysites (250 mg)
42 were added under stirring to a solution of TDP (0.27837 g, 1 mmol) in 250 mL of 4:1 EtOH: H₂O. The EtOH:
43 H₂O solution was adjusted to pH 4 with HCl 0.1 M. The halloysite suspension was transferred to a 500 mL
44 flask, which was then evacuated using a vacuum-pump. The fizzing of the suspension indicated that air was
45 removed from the halloysite lumen and replaced with TDP solution. The vacuum/nitrogen cycles were
46 repeated 3 times in order to maximize TDP loading. After stirring for a week at room temperature, the
47 modified halloysites were rinsed with EtOH:H₂O for 5 times and recovered by centrifugation, and finally dried
48 at 100 °C overnight under vacuum. The TDP excess was recovered from the gathered supernatants and stored
49 for the next functionalization process. After the functionalization process, colloidal stability of HNT-TDP
50 aqueous suspension was similar to that of untreated HNT at the same pH, indicating that the outermost
51 surface of the clay nanotubes was not hydrophobized. Recovered HNT-TDP ca 250 mg.

1
2
3 **4.4 Qualitative assay for lumen hydrophobicity by Nile Red dye.** Nile Red (1 mg, 3.1×10^{-3} mmol) was
4 dissolved in 10 mL ethanol and left under stirring until the compound was completely dissolved. Then, HNT-
5 TDP (20 mg) were added to the alcohol dye solution and the flask was cooled at 0 °C before reducing the
6 pressure by a mechanical pump and leaving the mixture for 2 h. The halloysites were recovered by
7 centrifuging at 4226 rcf for 10 min. The blue-violet pellet was separated from the supernatant and washed
8 with water and recovered by centrifugation. After HNT were dried, the recovered pellet appeared blue under
9 visible light and red-emissive under UV-lamp irradiation. The very same procedure was followed for the
10 loading of Nile Red in pristine HNT. After the washing of the treated HNT with water, the color under visible
11 light irradiation was blue-violet, whilst under UV light irradiation no light emission was detected.
12
13
14
15
16
17
18

19 **4.5 Preparation of HNT-TDP with SPION selectively loaded in the HNT lumen (SPION@OA-in-HNT-TDP).**
20 In a Schlenk tube SPION3@OA (3.75 mL, 2.2 mg/mL) were mixed with 150 mg HNT-TDP in 150 mL n-hexane.
21 The suspension was shaken for 1 min by a vortex, then the suspension was cooled at 0 °C, and while standing
22 in the ice bath, it was placed under reduced pressure and magnetic stirring until the solvent was visibly
23 reduced. Then, another 150 mL n-hexane were added. The color of the supernatant turned clearer with
24 respect to the beginning (from light brown to beige). The cycle of reduced pressure under stirring was
25 repeated once more, until the supernatant appeared completely clear (about 3 h). The HNT-TDP/SPION were
26 recovered by centrifugation (3 min, 470 rcf), the supernatant removed and the precipitate washed with 5 mL
27 n-hexane.
28
29
30
31
32
33
34

35 **4.6 Pre-treatment of the HNT mineral to obtain purified HNT in powder.** A piece of mineral clay was cut
36 in thin slices, keeping the part of clay as white as possible. Slices were dried at 50 °C for 1 h. Then, the slices
37 were gently hand-grinded in a mortar. To remove the soluble impurities, 2 g of powder was dispersed in 20
38 mL distilled water, followed by sonication for 20 min, 20 min stirring, then centrifuge 117 rcf for 20 min.
39 Then, the slurry was re-dispersed in 500 mL distilled water after the pH was adjusted at ca. 7 to achieve a
40 good suspension. The suspension was left to settle down the non-dissolved impurities, then the supernatant
41 containing HNT was transferred in another vial. The adjustment of pH at slightly acidic values ($\approx 4-5$) caused
42 the flocculation of HNT. The supernatant was discarded and HNT slurry dried at 60-80 °C, and grinded in a
43 mortar to give a white powder ready for further uses.
44
45
46
47
48
49

50 **5. Acknowledgments**

51
52
53 DM and HH are grateful to prof. John L. Keeling from Geological Survey of South Australia, Department for
54 Energy and Mining for the kind gift of a sample of HNT mineral from Camel Lake (Australia).
55
56
57
58
59
60

Part of this work was carried out at NOLIMITS, an advanced imaging facility established by the Università degli Studi di Milano. C.S. and C.I. gratefully acknowledge EC COST Action TD1402 (RADIOMAG) and INFN HADROMAG projects for partially funding the work.

6. Supporting Information

Experimental procedures, DLS and ζ -potential measurements and TEM images for the various HNT samples obtained with the electrostatic procedure. TEM images for the HNT sample prepared by in-situ procedure. Digital picture of HNT samples treated with Nile Red. DLS measurement and TEM images of many assays carried out on HNT-TDP samples by using the pre-modification procedure. This material is available free of charge via the Internet at <http://pubs.acs.org>.

7. References

- (1) Yuan, P.; Tan, D.; Annabi-Bergaya, F. Properties and Applications of Halloysite Nanotubes: Recent Research Advances and Future Prospects. *Appl. Clay Sci.* **2015**, *112–113*, 75–93.
- (2) Joussein, E.; Petit, S.; Churchman, J.; Theng, B.; Righi, D.; Delvaux, B. Halloysite Clay Minerals — a Review. *Clay Miner.* **2005**, *40* (4), 383–426.
- (3) Vergaro, V.; Abdullayev, E.; Lvov, Y. M.; Zeitoun, A.; Cingolani, R.; Rinaldi, R.; Leporatti, S. Cytocompatibility and Uptake of Halloysite Clay Nanotubes. *Biomacromolecules* **2010**, *11* (3), 820–826.
- (4) Szpilska, K.; Czaja, K.; Kudła, S. Halloysite Nanotubes as Polyolefin Fillers. *Polimery/Polymers* **2015**, *60* (6), 359–371.
- (5) Zhang, H. Selective Modification of Inner Surface of Halloysite Nanotubes: A Review. *Nanotechnol. Rev.* **2017**, *6* (6), 573–581.
- (6) Massaro, M.; Cavallaro, G.; Colletti, C. G.; Lazzara, G.; Milioto, S.; Noto, R.; Riela, S. Chemical Modification of Halloysite Nanotubes for Controlled Loading and Release. *J. Mater. Chem. B* **2018**, *6* (21), 3415–3433.
- (7) Lvov, Y.; Wang, W.; Zhang, L.; Fakhrullin, R. Halloysite Clay Nanotubes for Loading and Sustained Release of Functional Compounds. *Adv. Mater.* **2016**, *28* (6), 1227–1250.
- (8) Satish, S.; Tharmavaram, M.; Rawtani, D. Halloysite Nanotubes as a Nature 's Boon for Biomedical Applications. **2019**, *6*, 1–16.
- (9) Lee, Y.; Jung, G.; Cho, S. J.; Geckeler, K. E.; Fuchs, H. Cellular Interactions of Doxorubicin-Loaded DNA-Modified Halloysite Nanotubes. *Nanoscale* **2013**, *5* (18), 8577–8585.
- (10) Tully, J.; Yendluri, R.; Lvov, Y. Halloysite Clay Nanotubes for Enzyme Immobilization. *Biomacromolecules* **2016**, *17* (2), 615–621.

- 1
2
3 (11) Lun, H.; Ouyang, J.; Yang, H. Natural Halloysite Nanotubes Modified as an Aspirin Carrier. *RSC Adv.* **2014**, *4*
4 (83), 44197–44202.
5
6
7 (12) Kirimlioğlu, G. Y.; Yazan, Y. Development, Characterization and in Vitro Release Characteristics of Rabeprazole
8 Sodium in Halloysite Nanotubes. *Eur Int J Sci Technol* **2016**, *5* (4), 99–109.
9
10 (13) Long, Z.; Zhang, J.; Shen, Y.; Zhou, C.; Liu, M. Polyethyleneimine Grafted Short Halloysite Nanotubes for Gene
11 Delivery. *Mater. Sci. Eng. C* **2017**, *81* (May), 224–235.
12
13
14 (14) Shi, Y.-F.; Tian, Z.; Zhang, Y.; Shen, H.-B.; Jia, N.-Q. Functionalized Halloysite Nanotube-Based Carrier for
15 Intracellular Delivery of Antisense Oligonucleotides. *Nanoscale Res. Lett.* **2011**, *6* (1), 608.
16
17
18 (15) Liu, J.; Zhang, Y.; Zeng, Q.; Zeng, H.; Liu, X.; Wu, P.; Xie, H.; He, L.; Long, Z.; Lu, X.; Xiao, M.; Zhu, Y.; Bo, H.; Cao,
19 K. Delivery of RIPK4 Small Interfering RNA for Bladder Cancer Therapy Using Natural Halloysite Nanotubes. *Sci.*
20 *Adv.* **2019**, *5* (9), eaaw6499.
21
22
23 (16) Lee, Y.; Jung, G.-E.; Cho, S. J.; Geckeler, K. E.; Fuchs, H. Cellular Interactions of Doxorubicin-Loaded DNA-
24 Modified Halloysite Nanotubes. *Nanoscale* **2013**, *5* (18), 8577–8585.
25
26
27 (17) Abdullayev, E.; Lvov, Y. Halloysite Clay Nanotubes for Controlled Release of Protective Agents. *J. Nanosci.*
28 *Nanotechnol.* **2011**, *11* (11), 10007–10026.
29
30
31 (18) Koteja, A.; Matusik, J. Di- and Triethanolamine Grafted Kaolinites of Different Structural Order as Adsorbents
32 of Heavy Metals. *J. Colloid Interface Sci.* **2015**, *455*, 83–92.
33
34
35 (19) Du, M.; Guo, B.; Jia, D. Newly Emerging Applications of Halloysite Nanotubes: A Review. *Polym. Int.* **2010**, *59*
36 (5), 574–582.
37
38 (20) El-Boubbou, K. Magnetic Iron Oxide Nanoparticles as Drug Carriers: Clinical Relevance. *Nanomedicine* **2018**, *13*
39 (8), 953–971.
40
41
42 (21) Ruiz, A.; Morais, P. C.; Bentes de Azevedo, R.; Lacava, Z. G. M.; Villanueva, A.; del Puerto Morales, M. Magnetic
43 Nanoparticles Coated with Dimercaptosuccinic Acid: Development, Characterization, and Application in
44 Biomedicine. *J. Nanoparticle Res.* **2014**, *16* (11), 2589–2609.
45
46
47 (22) Hao, R.; Yu, J.; Ge, Z.; Zhao, L.; Sheng, F.; Xu, L.; Li, G.; Hou, Y. Developing Fe₃O₄ Nanoparticles into an Efficient
48 Multimodality Imaging and Therapeutic Probe. *Nanoscale* **2013**, *5* (23), 11954–11963.
49
50
51 (23) Huh, Y.-M.; Jun, Y.; Song, H.-T.; Kim, S.; Choi, J.; Lee, J.-H.; Yoon, S.; Kim, K.-S.; Shin, J.-S.; Suh, J.-S.; Cheon, J. In
52 Vivo Magnetic Resonance Detection of Cancer by Using Multifunctional Magnetic Nanocrystals. *J. Am. Chem.*
53 *Soc.* **2005**, *127* (35), 12387–12391.
54
55
56 (24) Grillone, A.; Ciofani, G. Magnetic Nanotransducers in Biomedicine. *Chem. - A Eur. J.* **2017**, *23* (64), 16109–
57 16114.
58
59
60

- 1
2
3 (25) Zhang, L.; Dong, W.-F.; Sun, H.-B. Multifunctional Superparamagnetic Iron Oxide Nanoparticles: Design,
4 Synthesis and Biomedical Photonic Applications. *Nanoscale* **2013**, *5* (17), 7664.
5
6
7 (26) Mazuel, F.; Espinosa, A.; Luciani, N.; Reffay, M.; Le Borgne, R.; Motte, L.; Desboeufs, K.; Michel, A.; Pellegrino,
8 T.; Lalatonne, Y.; Wilhelm, C. Massive Intracellular Biodegradation of Iron Oxide Nanoparticles Evidenced
9 Magnetically at Single-Endosome and Tissue Levels. *ACS Nano* **2016**, *10* (8), 7627–7638.
10
11
12 (27) Nosrati, H.; Salehiabar, M.; Fridoni, M.; Abdollahifar, M.-A.; Kheiri Manjili, H.; Davaran, S.; Danafar, H. New
13 Insight about Biocompatibility and Biodegradability of Iron Oxide Magnetic Nanoparticles: Stereological and In
14 Vivo MRI Monitor. *Sci. Rep.* **2019**, *9* (1), 7173.
15
16
17 (28) Ulbrich, K.; Holá, K.; Šubr, V.; Bakandritsos, A.; Tuček, J.; Zbořil, R. Targeted Drug Delivery with Polymers and
18 Magnetic Nanoparticles: Covalent and Noncovalent Approaches, Release Control, and Clinical Studies. *Chem.*
19 *Rev.* **2016**, *116* (9), 5338–5431.
20
21
22 (29) Guo, Y.; Zhang, Y.; Ma, J.; Li, Q.; Li, Y.; Zhou, X.; Zhao, D.; Song, H.; Chen, Q.; Zhu, X. Light/Magnetic
23 Hyperthermia Triggered Drug Released from Multi-Functional Thermo-Sensitive Magnetoliposomes for Precise
24 Cancer Synergetic Theranostics. *J. Control. Release* **2018**, *272* (November 2016), 145–158.
25
26
27 (30) Sun, Q.; Cheng, D.; Yu, X.; Zhang, Z.; Dai, J.; Li, H.; Liang, B.; Shuai, X. A PH-Sensitive Polymeric Nanovesicle
28 Based on Biodegradable Poly(Ethylene Glycol)-b-Poly(2-(Diisopropylamino)Ethyl Aspartate) as a MRI-Visible
29 Drug Delivery System. *J. Mater. Chem.* **2011**, *21* (39), 15316–15326.
30
31
32 (31) Kim, D. H.; Vitol, E. A.; Liu, J.; Balasubramanian, S.; Gosztola, D. J.; Cohen, E. E.; Novosad, V.; Rozhkova, E. A.
33 Stimuli-Responsive Magnetic Nanomicelles as Multifunctional Heat and Cargo Delivery Vehicles. *Langmuir*
34 **2013**, *29* (24), 7425–7432.
35
36
37 (32) Riedinger, A.; Guardia, P.; Curcio, A.; Garcia, M. A.; Cingolani, R.; Manna, L.; Pellegrino, T. Subnanometer Local
38 Temperature Probing and Remotely Controlled Drug Release Based on Azo-Functionalized Iron Oxide
39 Nanoparticles. *Nano Lett.* **2013**, *13* (6), 2399–2406.
40
41
42 (33) Thirunavukkarasu, G. K.; Cherukula, K.; Lee, H.; Jeong, Y. Y.; Park, I.-K.; Lee, J. Y. Magnetic Field-Inducible Drug-
43 Eluting Nanoparticles for Image-Guided Thermo-Chemotherapy. *Biomaterials* **2018**, *180*, 240–252.
44
45
46 (34) Fizir, M.; Dramou, P.; Zhang, K.; Sun, C.; Pham-Huy, C.; He, H. Polymer Grafted-Magnetic Halloysite Nanotube
47 for Controlled and Sustained Release of Cationic Drug. *J. Colloid Interface Sci.* **2017**, *505*, 476–488.
48
49
50 (35) Wei, Q.; Shi, R.; Lu, D.; Lei, Z. In Situ Formation of Gold Nanoparticles on Magnetic Halloysite Nanotubes via
51 Polydopamine Chemistry for Highly Effective and Recyclable Catalysis. *RSC Adv.* **2016**, *6* (35), 29245–29253.
52
53
54 (36) Dai, J.; Wei, X.; Cao, Z.; Zhou, Z.; Yu, P.; Pan, J.; Zou, T.; Li, C.; Yan, Y. Highly-Controllable Imprinted Polymer
55 Nanoshell at the Surface of Magnetic Halloysite Nanotubes for Selective Recognition and Rapid Adsorption of
56 Tetracycline. *RSC Adv.* **2014**, *4* (16), 7967–7978.
57
58
59
60

- 1
2
3 (37) Xie, Y.; Qian, D.; Wu, D.; Ma, X. Magnetic Halloysite Nanotubes/Iron Oxide Composites for the Adsorption of
4 Dyes. *Chem. Eng. J.* **2011**, *168* (2), 959–963.
5
6
7 (38) Arshadi, M.; Eskandarloo, H.; Enayati, M.; Godec, M.; Abbaspourrad, A. Highly Water-Dispersible and
8 Antibacterial Magnetic Clay Nanotubes Functionalized with Polyelectrolyte Brushes: High Adsorption Capacity
9 and Selectivity toward Heparin in Batch and Continuous System. *Green Chem.* **2018**, *20* (24), 5491–5508.
10
11
12 (39) Maleki, A.; Hajizadeh, Z. Magnetic Aluminosilicate Nanoclay: A Natural and Efficient Nanocatalyst for the
13 Green Synthesis of 4H-Pyran Derivatives. *Silicon* **2019**, *11* (6), 2789–2798.
14
15
16 (40) Kadam, A. A.; Jang, J.; Lee, D. S. Supermagnetically Tuned Halloysite Nanotubes Functionalized with
17 Aminosilane for Covalent Laccase Immobilization. *ACS Appl. Mater. Interfaces* **2017**, *9* (18), 15492–15501.
18
19
20 (41) Wan, X.; Zhan, Y.; Long, Z.; Zeng, G.; He, Y. Core@double-Shell Structured Magnetic Halloysite Nanotube
21 Nano-Hybrid as Efficient Recyclable Adsorbent for Methylene Blue Removal. *Chem. Eng. J.* **2017**, *330* (July),
22 491–504.
23
24
25 (42) Duan, J.; Liu, R.; Chen, T.; Zhang, B.; Liu, J. Halloysite Nanotube-Fe₃O₄ Composite for Removal of Methyl Violet
26 from Aqueous Solutions. *Desalination* **2012**, *293*, 46–52.
27
28
29 (43) Zheng, P.; Du, Y.; Ma, X. Selective Fabrication of Iron Oxide Particles in Halloysite Lumen. *Mater. Chem. Phys.*
30 **2015**, *151*, 14–17.
31
32
33 (44) Baaziz, W.; Liu, X.; Florea, I.; Begin-Colin, S.; Pichon, B. P.; Ulhaq, C.; Ersen, O.; Soria-Sánchez, M.; Zafeiratos, S.;
34 Janowska, I.; Beginb, D.; Pham-Huu, C. Carbon Nanotube Channels Selectively Filled with Monodispersed Fe₃-
35 xO₄ Nanoparticles. *J. Mater. Chem. A* **2013**, *1* (44), 13853–13861.
36
37
38 (45) Vinokurov, V. A.; Stavitskaya, A. V.; Glotov, A. P.; Novikov, A. A.; Zolotukhina, A. V.; Kotelev, M. S.; Gushchin, P.
39 A.; Ivanov, E. V.; Darrat, Y.; Lvov, Y. M. Nanoparticles Formed onto/into Halloysite Clay Tubules: Architectural
40 Synthesis and Applications. *Chem. Rec.* **2018**, *18* (7–8), 858–867.
41
42
43 (46) Zeng, X.; Wang, Q.; Wang, H.; Yang, Y. Catalytically Active Silver Nanoparticles Loaded in the Lumen of
44 Halloysite Nanotubes via Electrostatic Interactions. *J. Mater. Sci.* **2017**, *52* (14), 8391–8400.
45
46
47 (47) Hamdi, J.; Blanco, A. A.; Diehl, B.; Wiley, J. B.; Trudell, M. L. Room-Temperature Aqueous Suzuki–Miyaura
48 Cross-Coupling Reactions Catalyzed via a Recyclable Palladium@Halloysite Nanocomposite. *Org. Lett.* **2019**, *21*
49 (10), 3471–3475.
50
51
52 (48) Wu, S.; Qiu, M.; Guo, B.; Zhang, L.; Lvov, Y. Nanodot-Loaded Clay Nanotubes as Green and Sustained Radical
53 Scavengers for Elastomer. *ACS Sustain. Chem. Eng.* **2017**, *5* (2), 1775–1783.
54
55
56 (49) Gao, X.; Tang, F.; Jin, Z. Pt–Cu Bimetallic Nanoparticles Loaded in the Lumen of Halloysite Nanotubes.
57 *Langmuir* **2019**, *35* (45), 14651–14658.
58
59
60 (50) Rostamzadeh, T.; Islam Khan, M. S.; Riche', K.; Lvov, Y. M.; Stavitskaya, A. V.; Wiley, J. B. Rapid and Controlled

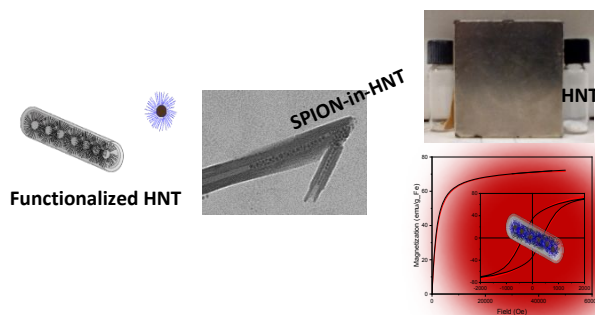
1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

In Situ Growth of Noble Metal Nanostructures within Halloysite Clay Nanotubes. *Langmuir* **2017**, *33* (45), 13051–13059.

- (51) Vinokurov, V. A.; Stavitskaya, A. V.; Chudakov, Y. A.; Ivanov, E. V.; Shrestha, L. K.; Ariga, K.; Darrat, Y. A.; Lvov, Y. M. Formation of Metal Clusters in Halloysite Clay Nanotubes. *Sci. Technol. Adv. Mater.* **2017**, *18* (1), 147–151.
- (52) Veerabadran, N. G.; Price, R. R.; Lvov, Y. M. Clay Nanotubes for Encapsulation and Sustained Release of Drugs. *NANO Br. Reports Rev.* **2007**, *2* (2), 115–120.
- (53) Jiang, W.; Wu, Y.; He, B.; Zeng, X.; Lai, K.; Gu, Z. Effect of Sodium Oleate as a Buffer on the Synthesis of Superparamagnetic Magnetite Colloids. *J. Colloid Interface Sci.* **2010**, *347* (1), 1–7.
- (54) Kim, B. H.; Lee, N.; Kim, H.; An, K.; Park, Y. I.; Choi, Y.; Shin, K.; Lee, Y.; Kwon, S. G.; Na, H. B.; Park, J.-G.; Ahn, T.-Y.; Kim, Y.-W.; Moon, W. K.; Choi, S. H.; Hyeon, T. Large-Scale Synthesis of Uniform and Extremely Small-Sized Iron Oxide Nanoparticles for High-Resolution T 1 Magnetic Resonance Imaging Contrast Agents. *J. Am. Chem. Soc.* **2011**, *133* (32), 12624–12631.
- (55) Sun, S.; Zeng, H.; Robinson, D. B.; Raoux, S.; Rice, P. M.; Wang, S. X.; Li, G. Monodisperse MFe₂O₄ (M = Fe, Co, Mn) Nanoparticles. *J. Am. Chem. Soc.* **2004**, *126* (1), 273–279.
- (56) Ansari, S. A. M. K.; Ficiarà, E.; Ruffinatti, F. A.; Stura, I.; Argenziano, M.; Abollino, O.; Cavalli, R.; Guiot, C.; D'Agata, F. Magnetic Iron Oxide Nanoparticles: Synthesis, Characterization and Functionalization for Biomedical Applications in the Central Nervous System. *Materials*. February 2, 2019, pp 465–489.
- (57) Lvov, Y.; Panchal, A.; Fu, Y.; Fakhrullin, R.; Kryuchkova, M.; Batasheva, S.; Stavitskaya, A.; Glotov, A.; Vinokurov, V. Interfacial Self-Assembly in Halloysite Nanotube Composites. *Langmuir* **2019**, *35* (26), 8646–8657.
- (58) Galli, M.; Guerrini, A.; Cauteruccio, S.; Thakare, P.; Dova, D.; Orsini, F.; Arosio, P.; Carrara, C.; Sangregorio, C.; Lascialfari, A.; Maggioni, D.; Licandro, E. Superparamagnetic Iron Oxide Nanoparticles Functionalized by Peptide Nucleic Acids. *RSC Adv.* **2017**, *7* (25), 15500–15512.
- (59) Galli, M.; Rossotti, B.; Arosio, P.; Ferretti, A. M.; Panigati, M.; Ranucci, E.; Ferruti, P.; Salvati, A.; Maggioni, D. A New Catechol-Functionalized Polyamidoamine as an Effective SPION Stabilizer. *Colloids Surfaces B Biointerfaces* **2019**, *174* (October 2018), 260–269.
- (60) Taroni, T.; Meroni, D.; Fidecka, K.; Maggioni, D.; Longhi, M.; Ardizzone, S. Halloysite Nanotubes Functionalization with Phosphonic Acids: Role of Surface Charge on Molecule Localization and Reversibility. *Appl. Surf. Sci.* **2019**, *486* (April), 466–473.
- (61) Yah, W. O.; Takahara, A.; Lvov, Y. M. Selective Modification of Halloysite Lumen with Octadecylphosphonic Acid: New Inorganic Tubular Micelle. *J. Am. Chem. Soc.* **2012**, *134* (3), 1853–1859.
- (62) Malo De Molina, P.; Appavou, M. S.; Gradzielski, M. Oil-in-Water Microemulsion Droplets of TDMAO/Decane

- 1
2
3 Interconnected by the Telechelic C18-EO150-C18: Clustering and Network Formation. *Soft Matter* **2014**, *10*
4 (28), 5072–5084.
5
6
7 (63) Chen, Y.; Zhang, Y.; Liu, J.; Zhang, H.; Wang, K. Preparation and Antibacterial Property of Polyethersulfone
8 Ultrafiltration Hybrid Membrane Containing Halloysite Nanotubes Loaded with Copper Ions. *Chem. Eng. J.*
9 **2012**, *210*, 298–308.
10
11
12 (64) Kim, T.; Shima, M. Reduced Magnetization in Magnetic Oxide Nanoparticles. *J. Appl. Phys.* **2007**, *101* (9),
13 09M516.
14
15
16 (65) Mørup, S. Superparamagnetism and Spin Glass Ordering in Magnetic Nanocomposites. *Europhys. Lett.* **1994**,
17 *28*, 671–676.
18
19
20 (66) Arosio, P.; Baldi, G.; Chiellini, F.; Corti, M.; Dessy, A.; Galinetto, P.; Gazzarri, M.; Grandi, M. S.; Innocenti, C.;
21 Lascialfari, A.; Lorenzi, G.; Orsini, F.; Piras, A. M.; Ravagli, C.; Sangregorio C. Magnetism and spin dynamics of
22 novel encapsulated iron oxide superparamagnetic nanoparticles. *Dalton Trans.* **2013**, *42*, 10282–10291.
23
24
25 (67) Roca, A. G.; Veintemillas-verdaguer, S.; Port, M.; Robic, C.; Serna, C. J.; Morales, M. P. Effects of Nanoparticle
26 and Aggregate Size on the Relaxometric Properties of MR Contrast Agents Based on High Quality Magnetite
27 Nanoparticles. *J. Phys. Chem. B* **2009**, *113*, 7033–7039.
28
29
30 (68) Mondini, S.; Ferretti, A. M.; Puglisi, A.; Ponti, A. Pebbles and PebbleJuggler: Software for Accurate, Unbiased,
31 and Fast Measurement and Analysis of Nanoparticle Morphology from Transmission Electron Microscopy
32 (TEM) Micrographs. *Nanoscale* **2012**, *4* (17), 5356–5372.
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table of Contents



SPION nanoparticles are selectively loaded into halloysite lumen, keeping their superparamagnetic character.

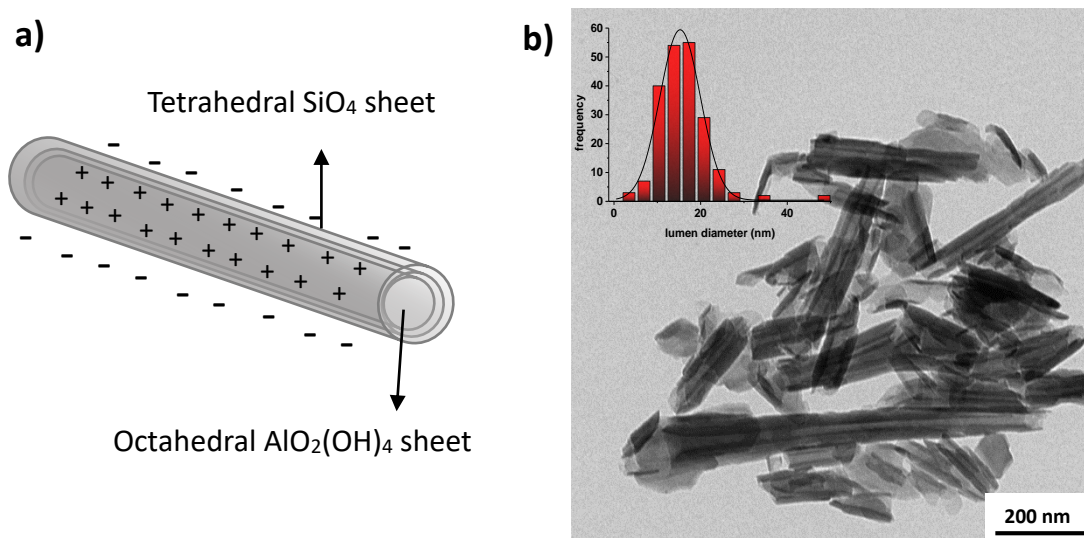


Figure 1. (a) Schematic representation of the structure of a halloysite with the indication of the molecular geometry around Al and Si centers specific for each layer and (b) TEM micrograph of a HNT sample (inset: HNT lumen diameter distribution).

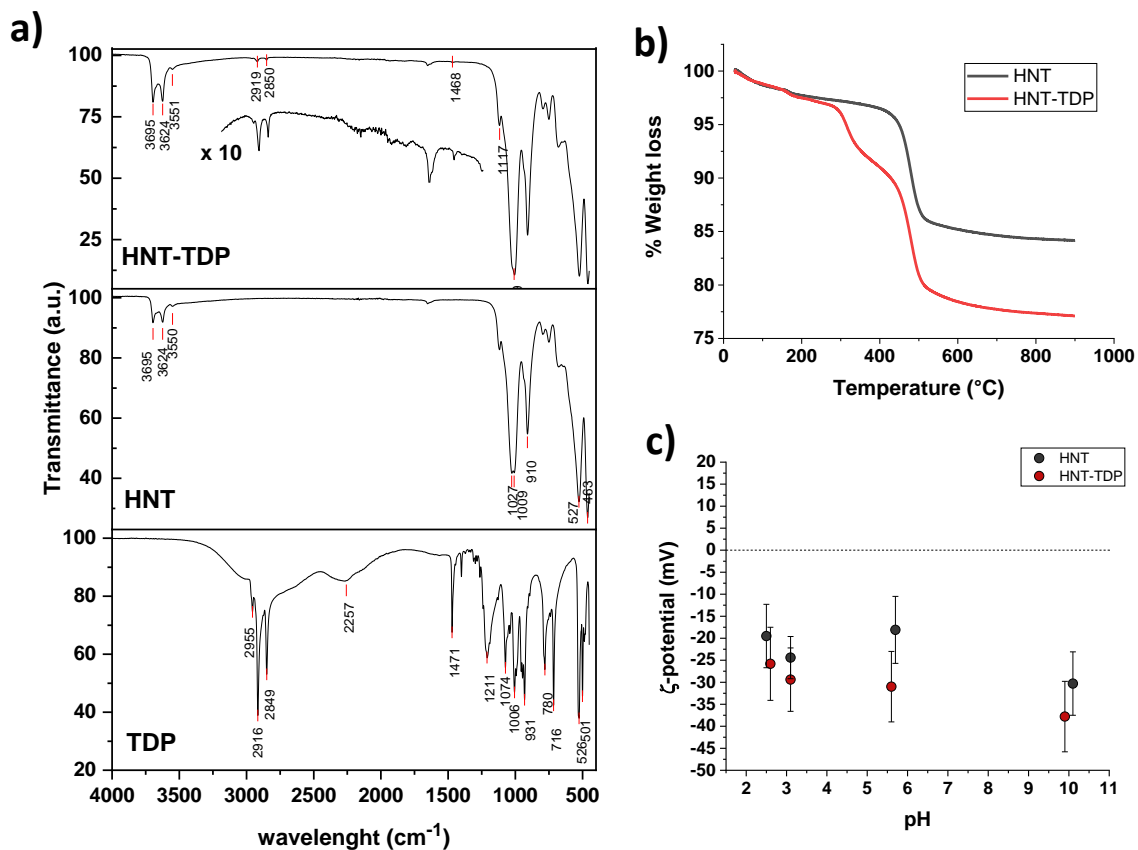


Figure 2. Characterization of the HNT-TDP (a) FTIR spectrum of HNT-TDP (top; the inset shows the magnified region of the spectrum containing signals of TDP) compared with the FTIR spectra of HNT (middle) and TDP (bottom); (b) thermogravimetric analysis (TGA) of pristine HNT and HNT-TDP derivative; (c) ζ -potential analyses of suspensions of HNT and HNT-TDP.

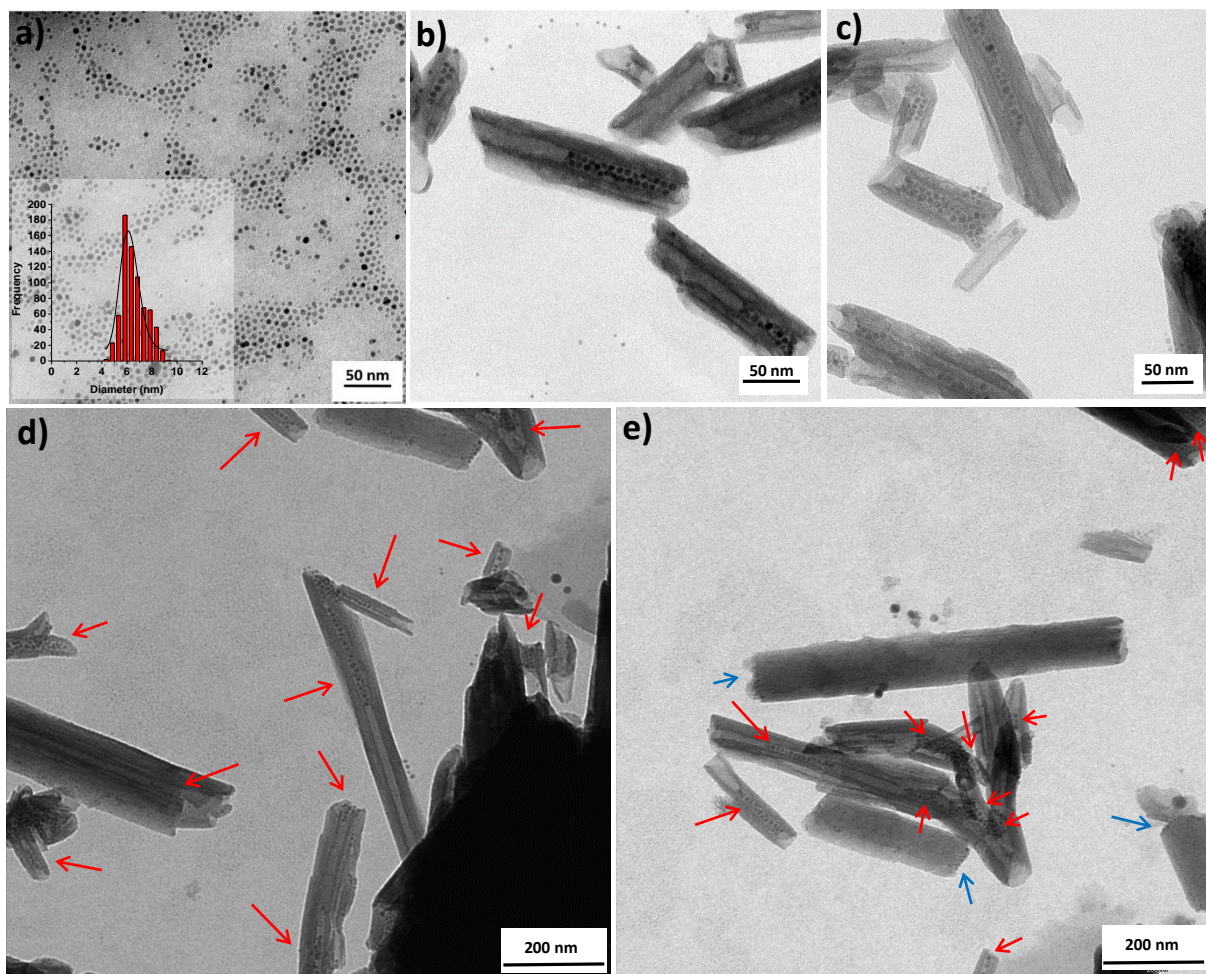


Figure 3. TEM micrographs of **(a)** as-synthesized SPION@OA (in the inset the SPION diameter distribution); **(b-e)** SPION-in-HNT prepared by pre-functionalization of HNT with TDP in the lumen. Panels **(b)** and **(c)** show images taken at higher magnification compared to **(d)** and **(e)**. Red arrows mark the HNT containing SPION. Blue arrows indicate SPION at the edge of few HNT.

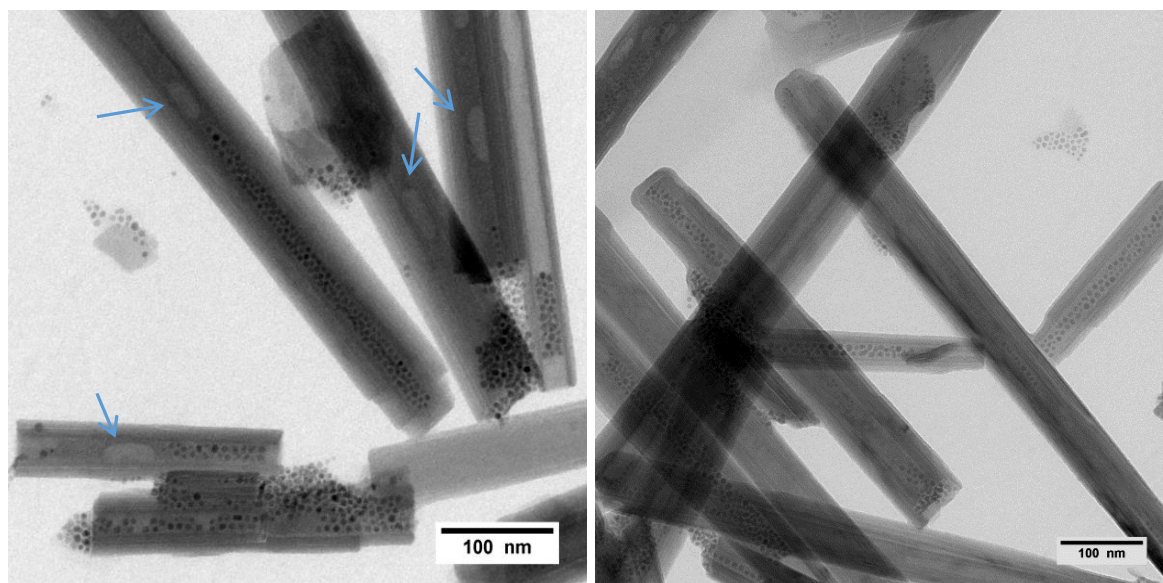


Figure 4. TEM micrographs of SPION-in-A-HNT (A-HNT= Australian Camel lake halloysites). Blue arrows mark the HNT containing possible air bubbles.

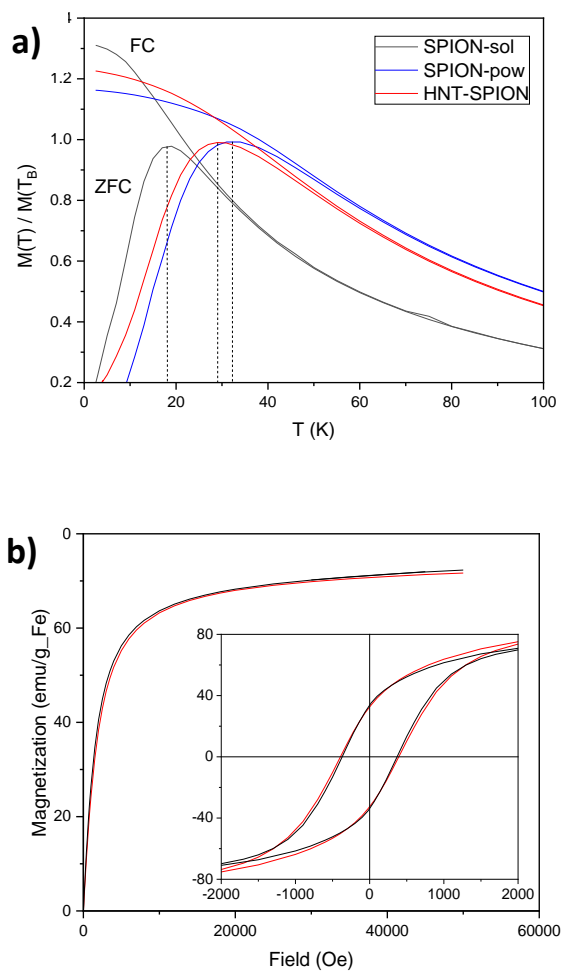


Figure 5. a) ZFC (lower curve) and FC (upper curve) magnetizations acquired with a 50 Oe field. The vertical dashed lines remark the position of the ZFC maxima corresponding to the blocking temperatures (T_B). For a better presentation, each curve was normalized to the magnetization value at the corresponding T_B ; b) Magnetization curves at high (300 K, main panel) and low (2.5 K, inset) temperature for SPIONs (black line) and SPION-in-HNT (red line).