

Received:
06 September 2019

Revised:
26 November 2019

Accepted:
27 November 2019

<https://doi.org/10.1259/bjr.20190770>

Cite this article as:

Monti CB, Codari M, De Cecco CN, Secchi F, Sardanelli F, Stillman AE. Novel imaging biomarkers: epicardial adipose tissue evaluation. *Br J Radiol* 2019; **92**: 20190770.

IMAGING PATIENTS WITH STABLE CHEST PAIN SPECIAL FEATURE: REVIEW ARTICLE

Novel imaging biomarkers: epicardial adipose tissue evaluation

¹CATERINA B. MONTI, MD, ²MARINA CODARI, MSc, PhD, ³CARLO NICOLA DE CECCO, MD, PhD, ^{1,4}FRANCESCO SECCHI, MD, PhD, ^{1,4}FRANCESCO SARDANELLI, MD and ³ARTHUR E. STILLMAN, MD, PhD

¹Department of Biomedical Sciences for Health, Università degli Studi di Milano, Milano, Italy

²Dipartimento di Elettronica, Informazione e Bioingegneria, Politecnico di Milano, Milano, Italy

³Division of Cardiothoracic Imaging, Department of Radiology and Imaging Sciences, Emory University Hospital, Atlanta, GA, USA

⁴Department of Radiology, IRCCS Policlinico San Donato, San Donato Milanese, Milano, Italy

Address correspondence to: Dr Carlo Nicola De Cecco
E-mail: carlo.dececco@emory.edu

ABSTRACT

Epicardial adipose tissue (EAT) is a metabolically activated beige adipose tissue, non-homogeneously surrounding the myocardium. Physiologically, EAT regulates toxic fatty acids, protects the coronary arteries against mechanical strain, regulates proinflammatory cytokines, stimulates the production of nitric oxide, reduces oxidative stress, and works as a thermogenic source against hypothermia. Conversely, EAT has pathologic paracrine interactions with the surrounded vessels, and might favour the onset of atrial fibrillation. In addition, initial atherosclerotic lesions can promote inflammation and trigger the EAT production of cytokines increasing vascular inflammation, which, in turn, may help the development of collateral vessels but also of self-stimulating, dysregulated inflammatory process, increasing coronary artery disease severity. Variations in EAT were also linked to metabolic syndrome. Echocardiography first estimated EAT measuring its thickness on the free wall of the right ventricle but does not allow accurate volumetric EAT estimates. Cardiac CT (CCT) and cardiac MR (CMR) allow for three-dimensional EAT estimates, the former showing higher spatial resolution and reproducibility but being limited by radiation exposure and long segmentation times, the latter being radiation-free but limited by lower spatial resolution and reproducibility, higher cost, and difficulties for obese patients. EAT radiodensity at CCT could be related to underlying metabolic processes. The correlation between EAT and response to certain pharmacological therapies has also been investigated, showing promising results. In the future, semi-automatic or fully automatic techniques, machine/deep-learning methods, if validated, will facilitate research for various EAT measures and may find a place in CCT/CMR reporting.

INTRODUCTION

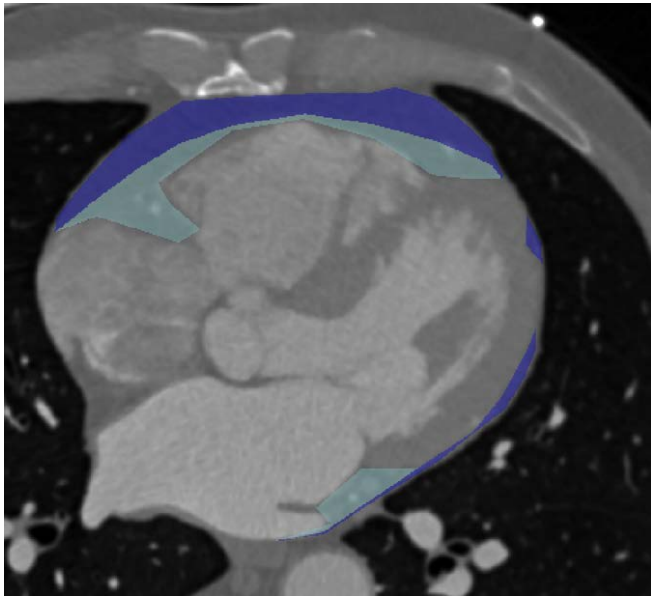
Epicardial adipose tissue (EAT) is the adipose tissue that lies immediately around the myocardium. EAT is in direct contact with both the myocardium itself and the coronary vessels, and it is surrounded by the visceral pericardium.¹

Since the early 2000s, EAT has become the object of many studies concerning biomarkers of cardiovascular risk, as its volume has been shown to be correlated to the incidence of pathologies such as metabolic syndrome, coronary atherosclerosis, or atrial fibrillation (AF).² In recent years, interest has shifted onto a precise EAT quantification, to potentially establish it as a reliable and non-invasive biomarker for cardiovascular risk.³

Epicardial adipose tissue anatomy and physiology

EAT is non-homogeneously distributed around the heart. Most of it lies at the cardiac base and apex, in the cardiac grooves and around the coronary arteries, and it is thicker around the right ventricle than around the left ventricle.⁴ Additional adipose tissue surrounding the heart, the so-called paracardial adipose tissue (PAT), can be found outside the parietal pericardium. The whole pericardial fat is defined as the sum of EAT and PAT.⁵ Figure 1 shows the respective localizations of these two fat deposits on a contrast-enhanced cardiac CT (CCT) scan.

Figure 1. Segmentation of EAT in light blue, and PAT in dark blue on a contrast-enhanced cardiac CT scan in a 64-year-old female patient. EAT, epicardial adipose tissue; PAT, paracardial adipose tissue.



EAT mass has been found to account for around 15–20% of the total cardiac mass, weighing around 50 g,⁶ for an average volume around 55 cm³. The correlation between an increased amount of EAT and visceral fat with obesity is yet under discussion.^{7,8} However, recent works have observed a causal link between obesity and accumulation and inflammation of EAT, potentially leading to coronary artery disease (CAD) or AF.⁹

As opposed to PAT, EAT originates from the splanchnopleuric mesoderm, as do mesenteric and omental fat.¹⁰ Physiologically, EAT has been shown to release free fatty acids, which are the main metabolites of cardiomyocytes, to keep toxic fatty acids at a healthy level, and it has a protective role for the coronary arteries against mechanical strain.⁴ In physiological conditions, EAT participates in the regulation of inflammation, regulating levels of pro-inflammatory cytokines, stimulates the production of nitric oxide, and reduces oxidative stress.¹¹

Moreover, EAT is composed of beige adipose tissue, which represents a conversion of white into brown adipose tissue following hormonal or temperature signals.^{12,13} A study by Sacks et al¹⁴ has revealed EAT as a thermogenic source for defending the myocardium and coronary vessels against hypothermia, using uncoupling protein-1 production as a marker. Indeed, uncoupling protein-1 is typically highly expressed in the beige adipose tissue, as it allows the production of heat by uncoupling electron transport from the synthesis of adenosine triphosphate, thus releasing the energy which would have been used for such process in the form of thermic energy.¹³ The beiging of white adipose tissue may be regarded as a metabolic activation, and thus as protective against metabolic disease.¹⁵ Moreover, an active thermogenic adipose tissue helps regulate circulating branched-chain amino acids, thus protecting individuals against obesity and diabetes.¹⁶

On the other hand, EAT, along with perivascular adipose tissue in general, has also been hypothesized to play a key role in the pathological development of cardiovascular disease, as it represents a metabolically active tissue with paracrine interactions with the surrounded vessels.¹⁷ Subjects with related pathologies such as hypertrophic cardiomyopathy or CAD have been found to have higher volumes of EAT than average.⁶ Increased EAT volume could be associated with a disruption of its physiological activity of regulating fatty acid levels around the heart.¹⁸ In fact, an excessive uptake of fatty acids by EAT might interfere with myocardial metabolism and with the functionality of the cardiac cycle, leading to arrhythmic complications as, for instance, the insurgence of AF.¹⁹

Moreover, EAT, as visceral adipose tissue, can be a source of proinflammatory cytokines. Even before cardiovascular pathology is overt and symptomatic, atherosclerotic lesions from CAD at an initial stage can promote inflammation and trigger the production from EAT of cytokines such as interleukin-1 β , interleukin-6, interleukin-6 soluble receptor and tumour necrosis factor α .²⁰ This process promotes an increase in EAT volume and an amplification of vascular inflammation. This, in turn, on one side may help neoangiogenesis and the development of collateral vessels, on the other side, may lead to a self-stimulating, dysregulated inflammatory process and the increase in severity of CAD.²¹ By means of the same mechanisms, an increase in EAT and its dysfunction may promote the onset of CAD.⁵ Both EAT volume and radiodensity were recently found to be associated with serum inflammatory markers, subclinical atherosclerosis and major cardiac events.²² EAT inflammation might play a role also in the development of AF.²³ Talman et al⁵ provide a comprehensive description and depiction of the pathophysiological role of EAT in their review.

Thus, an alteration in EAT, such as an increase in volume or thickness, or change in density, could be regarded as a marker of cardiovascular risk (a preclinical stage of cardiovascular disease), as also supported by the finding of modifications in EAT transcriptome in presence of severe CAD.²⁴

More generally, research has also linked variations in EAT with the presence of metabolic syndrome, a condition characterized by disruption of the physiological pathways regulating inflammation, given the similarities between its metabolic asset and that of visceral adipose tissue.⁸

IMAGING TECHNIQUES

Given the potential role of EAT in defining cardiovascular risk, different non-invasive techniques have been utilized in the attempts to provide a reliable quantification of its extent. The main methods for assessing EAT have so far been echocardiography, CCT, and cardiac MR (CMR).

At echocardiography, EAT appears as a hyperechoic area situated between the ventricular wall, more prominent around the free wall of the right ventricle, and the parietal pericardium.²⁵ Echocardiography has provided the first estimates of EAT by allowing to measure its thickness on the free wall of the right

Figure 2. Contrast-enhanced cardiac CT scan of a 64-year-old female patient depicting the thin pericardium (white arrowheads). The epicardial adipose tissue is immediately internal to the pericardium.



ventricle, where it is usually more prominent. EAT echocardiographic thickness measurements are usually performed perpendicularly to the myocardial wall in parasternal view, and long and short axis views.²⁶ Normal values are around 5 mm: Nelson et al²⁷ found a mean value of 4.7 ± 1.5 in asymptomatic subjects. In fact, some works suggest a cut-off value of 5 mm for EAT thickness to identify individuals at higher cardiovascular risk.^{28,29} Echocardiography is relatively inexpensive, widely available, and it does not bear limiting contraindications or radiation exposure. However, it presents important limitations such as the fact that two-dimensional echocardiography does not allow accurate volumetric EAT estimate, and its high dependence on the experience of the operator.^{30,31} Some authors have reported confusing pericardial effusion with EAT is sometimes possible.³¹

Alternatively, CCT is a volumetric technique, which allows for three-dimensional EAT estimates. EAT may be visible on CCT scans as a hypodense layer, with a density usually ranging -190 to -30 Hounsfield units,³ comprised between the myocardium and the visceral pericardium (Figure 2). EAT may be measured during CCT post-processing, not needing specific acquisitions. It can be quantified from different scans, most often either unenhanced scans for calcium scoring, or CCT angiographic scans, given that the EAT and pericardium are similarly visible in both, especially during the short time interval intercurrent between contrast administration and angiographic acquisition.^{32,33} Therefore, adding EAT volume estimate to the output of a CCT report could provide an additional risk estimate for cardiovascular disease, without adding to patient discomfort and radiation exposure.

Studies have reported a high reproducibility of EAT quantification on CCT scans.³⁴ However, standard electrocardiographically gated CCT, if used only for obtaining EAT estimates, could be questioned due to the ionizing radiation exposure, even though recently, Nagayama et al³⁵ proposed the use of non-gated CCT to quantify EAT volumes, achieving excellent concordance with

gated CCT with reduced radiation dose. Another limitation of EAT quantification by CCT angiography stems from long segmentation times, as a precise measurement is best performed on numerous small slice-thickness images, strongly limiting an integration into routine clinical practice. However, CCT allows the quantification of EAT X-ray radiodensity, which may be a biomarker of metabolic activity of EAT, possibly related to cardiovascular or metabolic disease.³⁶ EAT volume has also been assessed on positron emission tomography/CCT (PET/CCT), to associate anatomical data to functional information.³⁷ While to the best of our knowledge no meta-analysis has yet defined normal and pathological values for CCT-derived EAT volume and density, one study by Spearman et al reported values of CCT-derived EAT above 125 ml to be indicators of cardiac pathology.³⁸

CMR is, as of now, regarded as the easiest non-invasive approach for estimating the EAT volume mass.³⁹ Not unlike CCT, CMR is a volumetric technique that can provide highly reproducible, three-dimensional EAT measurements.⁴⁰ EAT can be visualized on images obtained with different CMR sequences, however it has been often quantified on cine sequences which are performed in almost all patients undergoing CMR⁴¹ (Figure 3). On cine bright-blood steady-state free-precession sequences, EAT may be visualized as a hyperintense, light grey area comprised between the hypointense myocardium and visceral pericardium (Figure 4). CMR is radiation-free and EAT estimation through CMR does not require the use of gadolinium-based contrast agents, whose safety has become increasingly debated during the last years.⁴²

Therefore, CMR can be considered a safe technique, provided that MRI contraindications are absent, such as marked obesity, claustrophobia or MR-unsafe devices.⁴³ In addition, we should consider drawbacks of CMR such as high cost, relatively long examination times, and limited availability. Obesity is important when EAT quantification for the estimation of cardiovascular risk is under consideration, as obese people are a growing population which could represent an interesting target in this context. Moreover, CMR scans generally have greater slice thicknesses than CCT scans, limiting the accuracy of EAT volume segmentation performed. In fact, to visualize the pericardium on the inferior slices of CMR scans is often challenging, as opposed to CCT scans.

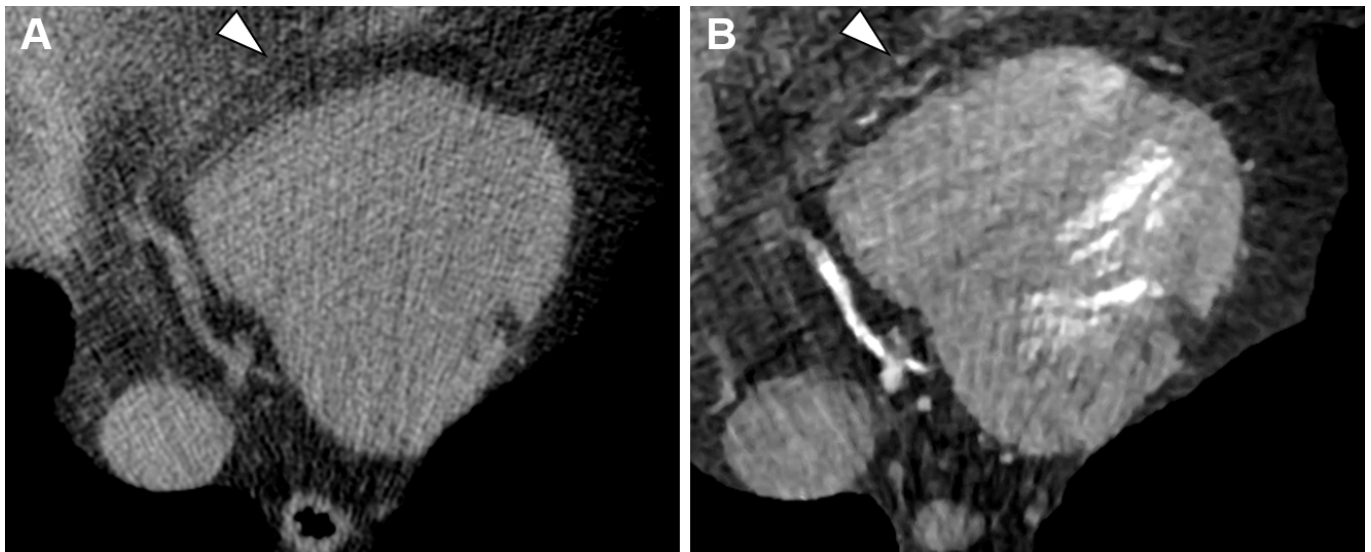
A comparison between the main imaging techniques for the evaluation of EAT is outlined in Table 1.

Epicardial adipose tissue as an imaging biomarker: actual evidence and open questions

Several studies have investigated the potential role of EAT as an imaging biomarker of various cardiovascular pathologies. EAT has been observed to be an independent risk factor for cardiovascular events in both healthy subjects and in patients with pre-existing diseases. It has been shown to directly correlate to the severity of cardiovascular pathologies, such as CAD, and to respond to metabolic changes originating from some pharmacological therapies.

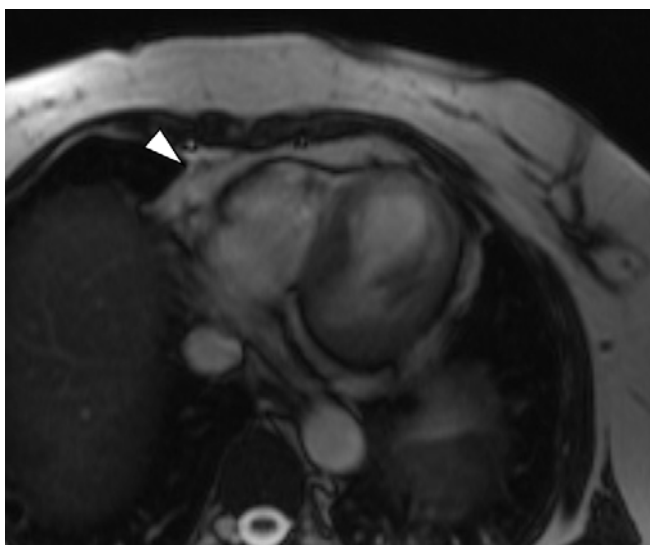
Longitudinal studies, with sample size ranging up to over 4000 subjects, have shown that EAT volume calculated from CCT or

Figure 3. Unenhanced (A) and arterial phase contrast-enhanced (B) cardiac computed tomography scans of a 52-year-old male patient showing the pericardium (white arrowheads), visible in both images as a thin, hyperdense line. This case is an example of the difficulties in segmenting the epicardial adipose tissue.



CMR may help predict the onset of arrhythmic complications such as AF. In 2010, a study by Thanassoulis et al,⁴⁴ part of the Framingham heart study, identified a strong positive independent association between EAT and prevalent AF, promoting a growing interest in EAT as imaging biomarker. In 2014, a study by Mahabadi et al,⁴⁵ part of the Heinz Nixdorf Recall study assessing novel cardiovascular risk factors in healthy subjects, also found a strong correlation between increased EAT and the incidence of AF. However, these authors also propose the hypothesis of the real predictor being the atrial size, as atrial size also bore the same correlation with AF incidence, and

Figure 4. Cardiac magnetic resonance true fast imaging with steady-state free precession sequence in a 64-year-old female patient, showing the pericardium (white arrowhead) as a thin, just visible, hypointense line surrounded by hyperintense adipose tissue on both sides.



indexing EAT for atrial size led to a loss of its risk prediction potential. In this regard, in 2016, a meta-analysis by Zhu et al⁴⁶ has divided patients for having paroxysmal or persistent AF, revealing that the role of EAT might differ with regards to the type of AF and underlying remodelling. This observation was also confirmed by other studies, investigating the characteristics of EAT in these two different patient groups.⁴⁷ Thus, further studies are needed to clarify the role of EAT as an independent predictor of the insurgence of different types of AF.

Concerning CAD, one study by Bastarrika et al⁴⁸ has observed that EAT, especially when quantified from CCT as opposed to two-dimensional measurements obtained from echocardiography, bears a positive correlation to the extent of CAD, assessed by both calcium scoring and stenosis severity. A study by Alexopoulos et al⁴⁹ from 2010 found increased EAT in patients with obstructive CAD or non-calcified plaques, correlating EAT to high-risk CAD. In 2014, a study by Mahabadi et al,⁵⁰ also part of the Heinz Nixdorf Recall study, found that the volume of EAT was associated to the progression of calcium scores, especially in younger subjects with low calcium scores, thus presenting it as a potential early predictor of CAD. In 2017, Fuller et al⁵¹ identified EAT as a potential risk predictor for sudden death related to CAD, from the finding that subjects with CAD who died from cardiac-related sudden death had significantly higher EAT volumes than those who died from other causes. One systematic review by Nerlekar et al⁵² reported that an increased EAT volume can be associated to the presence of high-risk plaques, though the *primum movens* of this process has yet to be clarified. Another review by Spearman et al³⁸ suggested that EAT provided incremental value for risk assessment over calcium scoring alone. Overall, the role of EAT as a predictor of CAD-related risk, being also fully supported by studies conducted on the Framingham cohort,⁵³ is more recognized than that related to AF.

Table 1. Comparison among the main imaging techniques for the evaluation of epicardial adipose tissue.

	Advantages	Disadvantages
Echocardiography	Readily available, inexpensive, quick	Highly operator-dependent, no volumetric measurements possible, difficult in patients who have motion or breathing issues
CCT	High definition, volumetric measurements possible, best visibility of the pericardium	Long segmentation times at low slice thickness when analysing contrast-enhanced series, ionizing radiations
CMR	Absence of ionizing radiations, volumetric measurements possible, allows quantification of attenuation	Expensive, not always available, long scan times, pericardium hard to visualize especially at lower sections of the heart, contraindicated in some patients

CCT, cardiac CT; CMR, cardiac MR.

Another important topic is the possible use of the radiodensity of EAT at CCT as a potential biomarker. Indeed, EAT radiodensity might differ according to underlying inflammation or due to the different levels of metabolic activity of adipose tissue at different browning degrees.³⁶ In 2017, one study by Antonopoulos et al⁵⁴ quantified fat radiodensity in epicardial, thoracic, and subcutaneous adipose tissue, observing that radiodensity could help in the investigation of the characteristics of different adipose tissues. Browning of EAT is an alternative explanation and is supported by a recent study showing decreased brown fat related genes in patients with acute coronary syndrome compared with stable CAD or non-CAD.⁵⁵ Marwan et al⁵⁶ found a lower radiodensity of pericoronary adipose tissue in normal segments compared to pathologic ones, however, their finding might also have been affected by the presence of contrast media, which may have a higher distribution in the pericoronary fat that surrounds atherosclerotic segments. Another possible explanation for the variations in EAT radiodensity in presence of cardiac pathology could be found in the higher metabolic activity of segments with CAD, due to underlying inflammation.⁵⁷ Conversely, Franssens et al⁵⁸ found a lower EAT radiodensity in patients at higher risk of cardiovascular disease, and Hell et al⁵⁹ observed decreased EAT density in patients with higher EAT volumes, body mass index, and a family history positive for CAD. Another study by Balcer et al⁶⁰ did not find EAT radiodensity differences between segments with or without atherosclerotic lesions in patients with acute myocardial infarction. In a recent study, Goeller et al²² found a strong association between EAT radiodensity and major adverse cardiac events. An increase in EAT radiodensity has also been linked to the presence of myocardial infarction with nonobstructive coronary arteries, and Tako-Tsubo syndrome.⁶¹

The correlation between EAT and response to certain pharmacological therapies has also been investigated. One study by Kang et al⁶² found an increased EAT thickness in CAD patients as a predictor of the insurgence of statin-associated diabetes mellitus. Moreover, statins have been observed to reduce EAT radiodensity, and this reduction can be associated to a decrease in EAT metabolic activity.³⁶ This process, which may be found also in other brown adipose tissue deposits, might potentially help explain the pathway between statin therapy and the development of diabetes, as statins inhibit the browning of fat, and have been linked to EAT regression.^{63–65} These findings are well in agreement with the fact that treatment with statins lowers risk of CAD, albeit bearing the risk of insurgence of diabetes.⁶⁶ One work by Ko et al,⁶⁷ conducted in patients undergoing haemodialysis treated with different drugs,

found a lack of EAT increase during the treatment with sevelamer, possibly due to its anti-inflammatory effect.

The association of metabolic information to anatomical data stemming from PET/CCT has led to yet debateable findings. A higher metabolic activity has been observed by Mazurek et al⁶⁸ in EAT as opposed to other fat deposits, in particular white adipose tissue. While one possible interpretation for this finding is that the increased uptake was secondary to inflammation in adipose tissue,⁶⁹ the absence of significant differences in EAT between patients with or without CAD contradicts this explanation. The higher uptake of EAT compared to other deposits could be due to the higher metabolic activity of brown adipose tissue compared to white.³⁶

FUTURE PERSPECTIVES

Methods allowing an automatic and reliable EAT segmentation gained recently a growing attention, as they would permit to integration of EAT estimation in routine practice. In this regard, the target technique for EAT segmentation would be unenhanced, ECG-gated CCT, which is already performed in subjects for the evaluation of calcium scoring, so to theoretically obtain two independent risk biomarkers from only one scan.^{70,71} CMR would find its main use on patients who already undergo examinations for clinical indications, that however do not represent the population at higher cardiovascular risk related to CAD. Especially on newer scanners, radiation exposure from scans for calcium scoring have proven to be extremely low, thus overcoming the main limitation of CCT.⁷² However, manual quantification of EAT on thin, CCT angiography slices may be highly time-consuming, and would at present not be feasible in routine clinical practice.

Different studies have so far proposed innovative approaches to accelerate EAT segmentation at CCT, ranging from semi-automatic techniques to fully automatic, deep-learning methods. In addition to quickening segmentation times, making EAT quantification fit for clinical practice, the adoption of fully automated segmentation methods would also provide a high reproducibility for EAT volumes.

Nichols et al³² quantified EAT volume, although calling it “pericardial adipose tissue”, with a semi-automatic method on coronary CCT angiographic scans. Their results show a very strong correlation between EAT values obtained by two different readers, albeit with a slight negative bias. A subsequent study by Barbosa et al⁷³ proposed a semi-automatic method on unenhanced CCT scans,

that bore a maximum interoperator error of around 10%. Goeller et al²² also used a semi-automatic method, a software named QFAT, to quantify EAT volume and radiodensity when studying such parameters in relation with CAD.

The first fully automatic EAT segmentation method was presented in 2014 by Ding et al⁷⁴ who used CCT examinations with 2.5–3 mm of slice thickness, each data set amounting to 50–60 slices. Automatically segmented EAT displayed a very strong correlation ($r = 0.96$) with EAT volume manually quantified by an expert operator; the segmentation time was under 60s on a standard computer. However, it also presented difficulties in recognizing the pericardium, especially when segmenting the highest or lowest slices, where the pericardial sac is not always clearly visible. In 2018, Commandeur et al⁷¹ proposed a novel segmentation method employing deep learning techniques. So far, they achieved a high correlation between automatic and manual EAT measurements ($R = 0.945$), with a non-significant bias (0.12cm^3) at Bland–Altman

analysis. However, their method still needs validation on larger cohorts.

CONCLUSIONS

EAT has emerged as an imaging biomarker especially useful for predicting CAD and its complications. This, along with the recent advances in the development of partially or fully automated methods for EAT segmentation may open road to the possibility of using EAT as a risk predictor in a clinical setting, adding a novel, independent risk biomarker to routine practice and facilitate research which presently is cumbersome to perform.

CONFLICTS OF INTEREST

Carlo N. De Cecco receives institutional research support and/or honorarium as speaker from Siemens and Bayer. Francesco Sardanelli received institutional research support and honorarium as speaker from Bracco, Bayer and General Electric. The other authors have no conflict of interest to disclose.

REFERENCES

- Gaborit B, Sengenès C, Ancel P, Jacquier A, Dutour A. Role of epicardial adipose tissue in health and disease: a matter of fat? *Compr Physiol* 2017; **7**: 1051–82. doi: <https://doi.org/10.1002/cphy.c160034>
- Antonopoulos AS, Antoniadis C. The role of epicardial adipose tissue in cardiac biology: classic concepts and emerging roles. *J Physiol* 2017; **595**: 3907–17. doi: <https://doi.org/10.1113/JP273049>
- Mancio J, Azevedo D, Saraiva F, Azevedo AI, Pires-Morais G, Leite-Moreira A, et al. Epicardial adipose tissue volume assessed by computed tomography and coronary artery disease: a systematic review and meta-analysis. *Eur Heart J Cardiovasc Imaging* 2018; **19**: 490–7. doi: <https://doi.org/10.1093/ehjci/jex314>
- Sacks HS, Fain JN. Human epicardial adipose tissue: a review. *Am Heart J* 2007; **153**: 907–17. doi: <https://doi.org/10.1016/j.ahj.2007.03.019>
- Talman AH, Psaltis PJ, Cameron JD, Meredith IT, Seneviratne SK, Wong DTL. Epicardial adipose tissue: far more than a fat depot. *Cardiovasc Diagn Ther* 2014; **4**: 416–29. doi: <https://doi.org/10.3978/j.issn.2223-3652.2014.11.05>
- Corradi D, Maestri R, Callegari S, Pastori P, Goldoni M, Luong TV, et al. The ventricular epicardial fat is related to the myocardial mass in normal, ischemic and hypertrophic hearts. *Cardiovasc Pathol* 2004; **13**: 313–6. doi: <https://doi.org/10.1016/j.carpath.2004.08.005>
- Marchington JM, Mattacks CA, Pond CM. Adipose tissue in the mammalian heart and pericardium: structure, foetal development and biochemical properties. *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry* 1989; **94**: 225–32. doi: [https://doi.org/10.1016/0305-0491\(89\)90337-4](https://doi.org/10.1016/0305-0491(89)90337-4)
- Malavazos AE, Ermetici F, Coman C, Corsi MM, Morricone L, Ambrosi B. Influence of epicardial adipose tissue and adipocytokine levels on cardiac abnormalities in visceral obesity. *Int J Cardiol* 2007; **121**: 132–4. doi: <https://doi.org/10.1016/j.ijcard.2006.08.061>
- Packer M. Epicardial adipose tissue may mediate deleterious effects of obesity and inflammation on the myocardium. *J Am Coll Cardiol* 2018; **71**: 2360–72. doi: <https://doi.org/10.1016/j.jacc.2018.03.509>
- Ho E, Shimada Y. Formation of the epicardium studied with the scanning electron microscope. *Dev Biol* 1978; **66**: 579–85. doi: [https://doi.org/10.1016/0012-1606\(78\)90263-4](https://doi.org/10.1016/0012-1606(78)90263-4)
- Li R, Wang W-Q, Zhang H, Yang X, Fan Q, Christopher TA, et al. Adiponectin improves endothelial function in hyperlipidemic rats by reducing oxidative/nitrative stress and differential regulation of eNOS/iNOS activity. *Am J Physiol Endocrinol Metab* 2007; **293**: E1703–8. doi: <https://doi.org/10.1152/ajpendo.00462.2007>
- Chechi K, Voisine P, Mathieu P, Laplante M, Bonnet S, Picard F, et al. Functional characterization of the Ucp1-associated oxidative phenotype of human epicardial adipose tissue. *Sci Rep* 2017; **7**: 15566. doi: <https://doi.org/10.1038/s41598-017-15501-7>
- XY Q, SL Q, Xiong WH, Rom O, Chang L, Jiang ZS, et al. Pvat in atherosclerosis: a double-edged sword. *Cardiovasc Diabetol* 2018; **17**: 134. doi: <https://doi.org/10.1186/s12933-018-0777-x>
- Sacks HS, Fain JN, Holman B, Cheema P, Chary A, Parks F, et al. Uncoupling protein-1 and related messenger ribonucleic acids in human epicardial and other adipose tissues: epicardial fat functioning as brown fat. *J Clin Endocrinol Metab* 2009; **94**: 3611–5. doi: <https://doi.org/10.1210/jc.2009-0571>
- van den Berg SM, van Dam AD, Rensen PCN, de Winther MPJ, Lutgens E. Immune Modulation of Brown(ing) Adipose Tissue in Obesity. *Endocr Rev* 2017; **38**: 46–68. doi: <https://doi.org/10.1210/er.2016-1066>
- Yoneshiro T, Wang Q, Tajima K, Matsushita M, Maki H, Igarashi K, et al. Bcaa catabolism in brown fat controls energy homeostasis through SLC25A44. *Nature* 2019; **572**: 614–9. doi: <https://doi.org/10.1038/s41586-019-1503-x>
- Akoumianakis I, Tarun A, Antoniadis C. Perivascular adipose tissue as a regulator of vascular disease pathogenesis: identifying novel therapeutic targets. *Br J Pharmacol* 2017; **174**: 3411–24. doi: <https://doi.org/10.1111/bph.13666>
- Iacobellis G, Corradi D, Sharma AM. Epicardial adipose tissue: anatomic, biomolecular and clinical relationships with the heart. *Nat Clin Pract Cardiovasc Med* 2005; **2**: 536–43. doi: <https://doi.org/10.1038/nccardio0319>
- Manzella D, Grella R, Marfella R, Giugliano D, Paolisso G. Elevated post-prandial free fatty acids are associated with cardiac sympathetic overactivity in type II diabetic

- patients. *Diabetologia* 2002; **45**: 1737–8. doi: <https://doi.org/10.1007/s00125-002-0965-8>
20. Laine P, Kaartinen M, Penttilä A, Panula P, Paaonon T, Kovanen PT. Association between myocardial infarction and the mast cells in the adventitia of the infarct-related coronary artery. *Circulation* 1999; **99**: 361–9. doi: <https://doi.org/10.1161/01.CIR.99.3.361>
 21. Mazurek T, Zhang L, Zalewski A, Mannion JD, Diehl JT, Arafat H, et al. Human epicardial adipose tissue is a source of inflammatory mediators. *Circulation* 2003; **108**: 2460–6. doi: <https://doi.org/10.1161/01.CIR.0000099542.57313.C5>
 22. Goeller M, Achenbach S, Marwan M, Doris MK, Cadet S, Commandeur F, et al. Epicardial adipose tissue density and volume are related to subclinical atherosclerosis, inflammation and major adverse cardiac events in asymptomatic subjects. *J Cardiovasc Comput Tomogr* 2018; **12**: 67–73. doi: <https://doi.org/10.1016/j.jcct.2017.11.007>
 23. Kusayama T, Furusho H, Kashiwagi H, Kato T, Murai H, Usui S, et al. Inflammation of left atrial epicardial adipose tissue is associated with paroxysmal atrial fibrillation. *J Cardiol* 2016; **68**: 406–11. doi: <https://doi.org/10.1016/j.jcct.2015.11.005>
 24. McAninch EA, Fonseca TL, Poggioli R, Panos AL, Salerno TA, Deng Y, et al. Epicardial adipose tissue has a unique transcriptome modified in severe coronary artery disease. *Obesity* 2015; **23**: 1267–78. doi: <https://doi.org/10.1002/oby.21059>
 25. Eroglu S. How do we measure epicardial adipose tissue thickness by transthoracic echocardiography? *Anatol J Cardiol* 2015; **15**: 416–9. doi: <https://doi.org/10.5152/akd.2015.5991>
 26. Singh N, Singh H, Khanijoun HK, Iacobellis G. Echocardiographic assessment of epicardial adipose tissue—a marker of visceral adiposity. *McGill J Med* 2007; **10**: 26–30.
 27. Nelson MR, Mookadam F, Thota V, Emani U, Al Harthi M, Lester SJ, et al. Epicardial fat: an additional measurement for subclinical atherosclerosis and cardiovascular risk stratification? *J Am Soc Echocardiogr* 2011; **24**: 339–45. doi: <https://doi.org/10.1016/j.jecho.2010.11.008>
 28. Bertaso AG, Bertol D, Duncan BB, Foppa M. Epicardial fat: definition, measurements and systematic review of main outcomes. *Arq Bras Cardiol* 2013; **101**: e18–28. doi: <https://doi.org/10.5935/abc.20130138>
 29. Lima-Martínez MM, Paoli M, Donis JH, Odreman R, Torres C, Iacobellis G, et al. Cut-Off point of epicardial adipose tissue thickness for predicting metabolic syndrome in Venezuelan population. *Endocrinología y Nutrición* 2013; **60**: 570–6. doi: <https://doi.org/10.1016/j.endoen.2013.12.007>
 30. Patel VB, Shah S, Verma S, Oudit GY. Epicardial adipose tissue as a metabolic transducer: role in heart failure and coronary artery disease. *Heart Fail Rev* 2017; **22**: 889–902. doi: <https://doi.org/10.1007/s10741-017-9644-1>
 31. Iacobellis G, Assael F, Ribaud MC, Zappaterreno A, Alessi G, Di Mario U, et al. Epicardial fat from echocardiography: a new method for visceral adipose tissue prediction. *Obes Res* 2003; **11**: 304–10. doi: <https://doi.org/10.1038/oby.2003.45>
 32. Nichols JH, Samy B, Nasir K, Fox CS, Schulze PC, Bamberg F, et al. Volumetric measurement of pericardial adipose tissue from contrast-enhanced coronary computed tomography angiography: a reproducibility study. *J Cardiovasc Comput Tomogr* 2008; **2**: 288–95. doi: <https://doi.org/10.1016/j.jcct.2008.08.008>
 33. La Grutta L, Toia P, Farruggia A, Albano D, Grassedonio E, Palmeri A, et al. Quantification of epicardial adipose tissue in coronary calcium score and CT coronary angiography image data sets: comparison of attenuation values, thickness and volumes. *Br J Radiol* 2016; **89**: 20150773. doi: <https://doi.org/10.1259/bjr.20150773>
 34. Nakazato R, Shmilovich H, Tamarappoo BK, Cheng VY, Slomka PJ, Berman DS, et al. Interscan reproducibility of computer-aided epicardial and thoracic fat measurement from noncontrast cardiac CT. *J Cardiovasc Comput Tomogr* 2011; **5**: 172–9. doi: <https://doi.org/10.1016/j.jcct.2011.03.009>
 35. Nagayama Y, Nakamura N, Itatani R, Oda S, Kusunoki S, Takahashi H, et al. Epicardial fat volume measured on nongated chest CT is a predictor of coronary artery disease. *Eur Radiol* 2019; **29**: 3638–46. doi: <https://doi.org/10.1007/s00330-019-06079-x>
 36. Raggi P, Gadiyaram V, Zhang C, Chen Z, Lopaschuk G, Stillman AE. Statins reduce epicardial adipose tissue attenuation independent of lipid lowering: a potential pleiotropic effect. *J Am Heart Assoc* 2019; **8**: e013104. doi: <https://doi.org/10.1161/JAHA.119.013104>
 37. Janik M, Hartlage G, Alexopoulos N, Mirzoyev Z, McLean DS, Arepalli CD, et al. Epicardial adipose tissue volume and coronary artery calcium to predict myocardial ischemia on positron emission tomography-computed tomography studies. *J Nucl. Cardiol* 2010; **17**: 841–7. doi: <https://doi.org/10.1007/s12350-010-9235-1>
 38. Spearman JV, Renker M, Schoepf UJ, Krazinski AW, Herbert TL, De Cecco CN, et al. Prognostic value of epicardial fat volume measurements by computed tomography: a systematic review of the literature. *Eur Radiol* 2015; **25**: 3372–81. doi: <https://doi.org/10.1007/s00330-015-3765-5>
 39. Kessels K, Cramer M-JM, Velthuis B. Epicardial adipose tissue imaged by magnetic resonance imaging: an important risk marker of cardiovascular disease. *Heart* 2006; **92**: 962. doi: <https://doi.org/10.1136/hrt.2005.074872>
 40. Flüchter S, Haghi D, Dinter D, Heberlein W, Kühl HP, Neff W, et al. Volumetric assessment of epicardial adipose tissue with cardiovascular magnetic resonance imaging. *Obesity* 2007; **15**: 870–8. doi: <https://doi.org/10.1038/oby.2007.591>
 41. Petrini M, Ali M, Cannà PM, Zambelli D, Cozzi A, Codari M, et al. Epicardial adipose tissue volume in patients with coronary artery disease or non-ischaemic dilated cardiomyopathy: evaluation with cardiac magnetic resonance imaging. *Clin Radiol* 2019; **74**: 81.e1–81.e7. doi: <https://doi.org/10.1016/j.crad.2018.09.006>
 42. Fraum TJ, Ludwig DR, Bashir MR, Fowler KJ. Gadolinium-Based contrast agents: a comprehensive risk assessment. *J Magn Reson Imaging* 2017; **46**: 338–53. doi: <https://doi.org/10.1002/jmri.25625>
 43. Shellock FG. Reference manual for magnetic resonance safety, implants. *and Devices: Edition 2019: Biomedical Research Publishing Group* 2019;.
 44. Thanassoulis G, Massaro JM, O'Donnell CJ, Hoffmann U, Levy D, Ellinor PT, et al. Pericardial fat is associated with prevalent atrial fibrillation: the Framingham heart study. *Circ Arrhythm Electrophysiol* 2010; **3**: 345–50. doi: <https://doi.org/10.1161/CIRCEP.109.912055>
 45. Mahabadi AA, Lehmann N, Kalsch H, Bauer M, Dykun I, Kara K, et al. Association of epicardial adipose tissue and left atrial size on non-contrast CT with atrial fibrillation: the heinz Nixdorf recall study. *Eur Heart J Cardiovasc Imaging* 2014; **15**: 863–9. doi: <https://doi.org/10.1093/ehjci/jeu006>
 46. Zhu W, Zhang H, Guo L, Hong K. Relationship between epicardial adipose tissue volume and atrial fibrillation : A systematic review and meta-analysis. *Herz* 2016; **41**: 421–7. doi: <https://doi.org/10.1007/s00059-015-4387-z>
 47. Psychari SN, Tsoukalas D, Varvarousis D, Papaspyropoulos A, Gkika E, Kotsakis A, et al. Opposite relations of epicardial adipose tissue to left atrial size in paroxysmal and permanent atrial fibrillation. *SAGE Open Medicine* 2018; **6**: 205031211879990. doi: <https://doi.org/10.1177/2050312118799908>
 48. Bastarrika G, Broncano J, Schoepf UJ, Schwarz F, Lee YS, Abro JA, et al. Relationship between coronary artery disease and epicardial adipose

- tissue quantification at cardiac CT: comparison between automatic volumetric measurement and manual bidimensional estimation. *Acad Radiol* 2010; **17**: 727–34.
49. Alexopoulos N, McLean DS, Janik M, Arepalli CD, Stillman AE, Raggi P. Epicardial adipose tissue and coronary artery plaque characteristics. *Atherosclerosis* 2010; **210**: 150–4. doi: <https://doi.org/10.1016/j.atherosclerosis.2009.11.020>
 50. Mahabadi AA, Lehmann N, Kälsch H, Robens T, Bauer M, Dykun I, et al. Association of epicardial adipose tissue with progression of coronary artery calcification is more pronounced in the early phase of atherosclerosis. *JACC: Cardiovascular Imaging* 2014; **7**: 909–16. doi: <https://doi.org/10.1016/j.jcmg.2014.07.002>
 51. Fuller B, Garland J, Anne S, Beh R, McNeven D, Tse R. Increased epicardial fat thickness in sudden death from stable coronary artery atherosclerosis. *Am J Forensic Med Pathol* 2017; **38**: 162–6. doi: <https://doi.org/10.1097/PAF.0000000000000310>
 52. Nerlekar N, Brown AJ, Muthalaly RG, Talman A, Hettige T, Cameron JD, et al. Association of epicardial adipose tissue and High-Risk plaque characteristics: a systematic review and Meta-Analysis. *J Am Heart Assoc* 2017; **6**: doi: <https://doi.org/10.1161/JAHA.117.006379>
 53. Mahabadi AA, Massaro JM, Rosito GA, Levy D, Murabito JM, Wolf PA, et al. Association of pericardial fat, intrathoracic fat, and visceral abdominal fat with cardiovascular disease burden: the Framingham heart study. *Eur Heart J* 2009; **30**: 850–6. doi: <https://doi.org/10.1093/eurheartj/ehn573>
 54. Antonopoulos AS, Sanna F, Sabharwal N, Thomas S, Oikonomou EK, Herdman L, et al. Detecting human coronary inflammation by imaging perivascular fat. *Sci Transl Med* 2017; **9**: eaal2658. doi: <https://doi.org/10.1126/scitranslmed.aal2658>
 55. Moreno-Santos I, Macías-González M, Porras-Martín C, Castellano-Castillo D, Sánchez-Espín G, Gómez-Doblas JJ, et al. Role of epicardial adipose tissue NPR-C in acute coronary syndrome. *Atherosclerosis* 2019; **286**: 79–87. doi: <https://doi.org/10.1016/j.atherosclerosis.2019.05.010>
 56. Marwan M, Hell M, Schubbäck A, Gauss S, Bittner D, Pflederer T, et al. Ct attenuation of Pericoronary adipose tissue in normal versus atherosclerotic coronary segments as defined by intravascular ultrasound. *J Comput Assist Tomogr* 2017; **41**: 762–7. doi: <https://doi.org/10.1097/RCT.0000000000000589>
 57. Mazurek T, Kochman J, Kobylecka M, Wilimski R, Filipiak KJ, Królicki L, et al. Inflammatory activity of pericoronary adipose tissue may affect plaque composition in patients with acute coronary syndrome without persistent ST-segment elevation: preliminary results. *Kardiol Pol* 2014; **72**: 410–6. doi: <https://doi.org/10.5603/KPa.2013.0320>
 58. Franssens BT, Nathoe HM, Visseren FLJ, van der Graaf Y, Leiner T, Algra A, Group SS, et al. Relation of epicardial adipose tissue Radiodensity to coronary artery calcium on cardiac computed tomography in patients at high risk for cardiovascular disease. *Am J Cardiol* 2017; **119**: 1359–65. doi: <https://doi.org/10.1016/j.amjcard.2017.01.031>
 59. Hell MM, Achenbach S, Schubbäck A, Klinghammer L, May MS, Marwan M. Ct-Based analysis of pericoronary adipose tissue density: relation to cardiovascular risk factors and epicardial adipose tissue volume. *J Cardiovasc Comput Tomogr* 2016; **10**: 52–60. doi: <https://doi.org/10.1016/j.jcct.2015.07.011>
 60. Balcer B, Dykun I, Schlosser T, Forsting M, Rassaf T, Mahabadi AA. Pericoronary fat volume but not attenuation differentiates culprit lesions in patients with myocardial infarction. *Atherosclerosis* 2018; **276**: 182–8. doi: <https://doi.org/10.1016/j.atherosclerosis.2018.05.035>
 61. Gaibazzi N, Martini C, Botti A, Pinazzi A, Bottazzi B, Palumbo AA. Coronary inflammation by computed tomography Pericoronary fat attenuation in MINOCA and Tako-Tsubo syndrome. *J Am Heart Assoc* 2019; **8**: e013235. doi: <https://doi.org/10.1161/JAHA.119.013235>
 62. Kang J, Kim Y-C, Park JJ, Kim S, Kang S-H, Cho YJ, et al. Increased epicardial adipose tissue thickness is a predictor of new-onset diabetes mellitus in patients with coronary artery disease treated with high-intensity statins. *Cardiovasc Diabetol* 2018; **17**: 10. doi: <https://doi.org/10.1186/s12933-017-0650-3>
 63. Alexopoulos N, Melek BH, Arepalli CD, Hartlage G-R, Chen Z, Kim S, et al. Effect of intensive versus moderate lipid-lowering therapy on epicardial adipose tissue in hyperlipidemic post-menopausal women: a substudy of the BELLES trial (beyond endorsed lipid lowering with EBT scanning). *J Am Coll Cardiol* 2013; **61**: 1956–61. doi: <https://doi.org/10.1016/j.jacc.2012.12.051>
 64. Balaz M, Becker AS, Balazova L, Straub L, Müller J, Gashi G, et al. Inhibition of mevalonate pathway prevents adipocyte browning in mice and men by affecting protein prenylation. *Cell Metab* 2019; **29**: 901–16. doi: <https://doi.org/10.1016/j.cmet.2018.11.017>
 65. Aiman U, Najmi A, Khan R. Statin induced diabetes and its clinical implications. *J Pharmacol Pharmacother* 2014; **5**: 181–5. doi: <https://doi.org/10.4103/0976-500X.136097>
 66. Stone NJ, Robinson JG, Lichtenstein AH, Bairey Merz CN, Blum CB, Eckel RH, et al. ACC/AHA guideline on the treatment of blood cholesterol to reduce atherosclerotic cardiovascular risk in adults. *Journal of the American College of Cardiology* 2013; **2014**: 2889–934.
 67. Ko SM, Zhang C, Chen Z, D'Marco L, Bellasi A, Stillman AE, et al. Epicardial adipose tissue volume increase in hemodialysis patients treated with sevelamer or calcium-based phosphate binders: a substudy of the Renegal in new dialysis trial. *J Nephrol* 2016; **29**: 683–90. doi: <https://doi.org/10.1007/s40620-016-0310-9>
 68. Mazurek T, Kobylecka M, Zielenkiewicz M, Kurek A, Kochman J, Filipiak KJ, et al. Pet/Ct evaluation of 18F-FDG uptake in pericoronary adipose tissue in patients with stable coronary artery disease: independent predictor of atherosclerotic lesions' formation? *J. Nucl. Cardiol.* 2017; **24**: 1075–84. doi: <https://doi.org/10.1007/s12350-015-0370-6>
 69. Madonna R, Massaro M, Scoditti E, Pescetelli I, De Caterina R. The epicardial adipose tissue and the coronary arteries: dangerous liaisons. *Cardiovasc Res* 2019; **115**: 1013–25. doi: <https://doi.org/10.1093/cvr/cvz062>
 70. Callister TQ, Cooil B, Raya SP, Lippolis NJ, Russo DJ, Raggi P. Coronary artery disease: improved reproducibility of calcium scoring with an electron-beam CT volumetric method. *Radiology* 1998; **208**: 807–14. doi: <https://doi.org/10.1148/radiology.208.3.9722864>
 71. Commandeur F, Goeller M, Betancur J, Cadet S, Doris M, Chen X, et al. Deep learning for quantification of epicardial and thoracic adipose tissue from Non-Contrast CT. *IEEE Trans Med Imaging* 2018; **37**: 2804799: 1835: 46. doi: <https://doi.org/10.1109/TMI.2018.2804799>
 72. Marwan M, Achenbach S, Korosoglou G, Schermund A, Schneider S, Bruder O, et al. German cardiac CT registry: indications, procedural data and clinical consequences in 7061 patients undergoing cardiac computed tomography. *Int J Cardiovasc Imaging* 2018; **34**: 807–19. doi: <https://doi.org/10.1007/s10554-017-1282-0>
 73. Barbosa JG, Figueiredo B, Bettencourt N, Tavares JMRS. Towards automatic quantification of the epicardial fat in non-contrasted CT images. *Comput Methods Biomech Biomed Engin* 2011; **14**: 905–14. doi: <https://doi.org/10.1080/10255842.2010.499871>
 74. Ding X, Terzopoulos D, Diaz-Zamudio M, Berman DS, Slomka PJ, Dey D. Automated epicardial fat volume quantification from non-contrast CT. *Medical Imaging 2014: Image Processing* 2014.