

From healthy aging to frailty: in search for the underlying mechanisms

Journal:	Current Medicinal Chemistry
Manuscript ID	Draft
Manuscript Type:	Review
Date Submitted by the Author:	n/a
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Keywords:	neuroplasticity, epigenetic, neuroinflammation, HPA axis, stress, aging

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Abstract

Population aging is accelerating rapidly worldwide, from 461 million people older than 65 years in 2004 to an estimated 2 billion people by 2050, leading to critical implications for the planning and delivery of health and social care.

The most problematic expression of population aging is the clinical condition of frailty, which is a state of increased vulnerability that develops as a consequence of the accumulation of microscopic damages in many physiological systems that leads to a striking and disproportionate change in health state, even after an apparently small insult.

Since little is known about the biology of frailty, an important perspective to understand this phenomenon is to establish how the alterations that physiologically occurs during a condition of healthy aging may instead promote cumulative decline with a subsequent depletion of homoeostatic reserve, and increased vulnerability also after minor stressor events.

In this context, the present review aims to provide a description of the molecular mechanisms that, by having a critical impact on behavior and neuronal function in aging, might be relevant for the development of frailty. Moreover, since these biological systems are also involved in the coping strategies set in motion to respond to environmental challenges, we propose a role for lifestyle stress as important player to drive frailty in aging.

1. From healthy aging to frailty

Life expectancy in the 20th century has greatly increased in the Western Countries. However, middle-aged and elderly people currently do not experience a parallel increase in health span due to the development of age-related pathologies. The most problematic expression of population aging is the clinical condition of frailty, a state of increased vulnerability to poor resolution of homoeostasis after a stressor event, which increases the risk of adverse outcome [1]. The high prevalence of chronic disabling conditions impact both on the quality of life of the individual as well as on the economic burden. On these bases it is crucial to understand the molecular mechanisms underlying frailty. It would be particularly important to identify the risk factors leading to frailty instead of healthy aging and their impact at molecular level.

Aging is associated with structural and physiological changes in the brain, the majority of which involve the synapses, the most important structures in neuronal network for maintaining accurate neuronal activity and normal brain functions. The maintenance of synaptic functionality requires preservation of proper synaptic structure, coordination of synaptic vesicle release and membrane excitability, and integration of retrograde signals from the postsynaptic terminal [2]. During aging these processes undergo a physiological decline [3]and structural changes in neurons and spines as well as alterations in neurotransmitters receptors expression and changes in electrophysiological properties occur [4-9]. Specifically, several morphological changes have been reported during aging, such as reduction in synaptic density, pruning of the dendritic tree, loss of dendritic spines, structural changes in the presynaptic active zone and alterations of receptors expression and trafficking [10]. Moreover, in both in human and rodents, loss of synapses in aging has been associated with cognitive impairment [11-13].

Spines represent key element of glutamatergic synapses, whose morphological and functional changes may affect synaptic events and excitatory synaptic signaling in aged neurons. Indeed, several lines of evidence indicate that aging is associated with a reduction of NMDA (N-methyl D-Aspartate) subunit expression and receptor activity in brain regions involved in higher brain function including synaptic plasticity, learning and memory [14-21].

Age-related changes can be detected in both excitatory and inhibitory synapses with a shift of the excitatory/inhibitory balance that can cause cognitive impairments [22], both in humans and in rodents[11-13]. GABAergic transmission is markedly impaired during aging, causing an increase in cortical excitability [23-25]. Indeed, many studies showed a reduction of GABA (γ -aminobutirric acid) synthesis [26] probably due also to the downregulation of the two isoforms of the enzyme responsible of GABA synthesis, namely GAD65 (Glutamic Acid Decarboxylase) and GAD67 [27] in different brain areas. Furthermore also the composition of GABA_{A-B} receptors changes during aging, contributing to changes in learning and memory processes [28]. For example, age related with an up-regulation of \square 1 subunits of GABA_AR in the rat hippocampus [29]and a reduction of GABA_BR1 in the same brain area is associated with with spatial learning impairment [28, 30].

It is not surprising that all these alterations in turn may impact the functions of other systems playing a critical role in the maintenance of synaptic function, as presented in the next section.

2. Molecular mechanisms altered in the aged brain

Based on the definition of frailty, this condition may be considered the expression of a loss in resilience – the ability to cope with adverse conditions- that may be the result of a progressive impairment in systems and mechanisms crucial for the proper synaptic function within the brain. Alterations of synaptic structure and/or function are in fact considered a potential risk factor or even a critical determinant for several neurodegenerative disorders associated with aging e.g. Alzheimer and Parkinson' diseases as well as for cognitive decline. These pathological conditions, which are indeed characterized by reductions in synapse numbers and alterations of genes encoding proteins involved in synapse formation and function can be considered as "synaptopathies", in which the dysfunction of the synapse is the central element [31].

Deficits of such systems and mechanisms would lead to increased vulnerability of the synapse to the impact of internal or external environment as well as to an enhanced risk to develop pathological conditions that characterize unhealthy aging.

Although we are aware that multiple molecular systems may contribute to alteration of synaptic function during aging, in this mini-review we will specifically focus on deficits of neuronal plasticity, dysfunction of

hypothalamic-pituitary-adrenal (HPA) axis, alterations of immune/inflammatory system as well as epigenetic modifications.

2.1 Deficits in neuronal plasticity and neurotrophic mechanisms

Among the mechanism that may contribute to age-related impairment, deficits of neuronal plasticity may play a crucial role. This term was used for the first time by Santiago Ramon y Cajal (1852-1934) [32] and now refers to "the ability to make adaptive changes related to the structure and function of the nervous system" [33]. According to this idea, even though brain plasticity is essentially a physical process, the term "neuronal plasticity" can stand not only for morphological modifications – for instance of the gray matter that can increase or narrow [34] - or for alteration in the connections between neurons that can be formed, shaped, modified or impaired. Indeed, it's important to consider that neuroplasticity is inherently connected and works together with the generation of new neurons (neurogenesis) and specific neurobiochemical changes in order to improve the brain's ability to adapt to its environment.

Historically, being plasticity essentially a physical and morphological phenomenon, it was thought that primary contributions to the decline of this brain feature were massive cell loss [35] as well as deterioration of dendritic branching [36]. However, we now know that the brain's modifications occurring during normal and successful aging are more subtle and selective than was once believed. Indeed, the plastic changes in structure and function, such as modifications in the neuronal morphology and alterations in the tissue density, that the brain goes through do not follow a homogenous and universal pattern within the different brain areas but are specific and peculiar [10]. Moreover, dendritic and spine densities and dynamics and functional interactions among different neurotransmitters are known to change differently in specific areas of the brain [37].

Among the brain regions affected by aging, the prefrontal cortex (PFC) and the hippocampus seem to be particularly vulnerable, but even within and between these regions the impact of aging on neuronal function can differ [10]. For instance, the morphology of neurons in the PFC is more susceptible to agerelated change, as these cells show a decrease in dendritic branching in rats [38] and humans [39], while in the hippocampus, subregion-specific changes in the expression of genes involved in the regulation of neuroplasticity have been observed [40]. Neurogenesis in the hippocampal dentate gyrus also decreases with age, which may contribute to the changes in brain's plasticity over the lifespan [41]. Moreover, induction and maintenance of long term potentiation, alongside alterations in calcium homeostasis, changes with age (Lister and Barnes, 2009). These alterations could be related to oxidation of proteins that are involved in cellular ion homeostasis. The resulting elevated calcium concentrations can cause neuronal degeneration and cell death, initiating several age-related impairments such as learning and memory deficits [42].

Among all the molecular regulators of neuronal plasticity, neurotrophic factors (NTFs), and in particular the neurotrophin family, may be considered key players. Indeed, besides their well-established activity in supporting the survival, differentiation and maintenance of neuronal functions, NTFs finely modulate all the crucial steps of network construction, from neuronal migration to experience-dependent refinement of local connections [43]. It is now well-known that NTFs are important mediators of neuronal plasticity not only during the development of the brain but also in adulthood where they continue to support and modulate axonal and dendritic growth, membrane receptor trafficking, neurotransmitter release, spine morphogenesis and neurogenesis [44, 45].

The neurotrophin family comprises of NGF, Brain-derived neurotrophic factor (BDNF), neurotrophin-3 (NT-3) and neurotrophin-4 (NT-4) that have all evolved from a common neurotrophin ancestor gene. Their actions are dependent on binding to transmembrane receptor systems – the tropomyosin receptor tyrosine kinase family and the p75 neurotrophin receptor [46]. Neurotrophins have preferential binding for specific receptors: NGF binds to TrkA, BDNF and NT-4 to TrkB, and NT-3 to TrkC, although there are also a number of promiscuous interactions [47]. All four neurotrophins can bind to the p75 receptor and the association of p75 with Trk receptors can regulate the affinity of Trk receptors for each respective neurotrophin, allowing for greater control of ligand–receptor interactions within this system [48].

BDNF is a key mediator of activity-dependent processes in the brain that have a major impact on neuronal development and plasticity. A central function of BDNF is to help neurons to adapt, survive and to speed up the brain's ability to make new connections.

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Neurotrophic factors seem to be very much relevant in the aging brain, indeed, a negative correlation between BDNF serum levels and age has been reported in a large cohort of healthy elderly individuals [49]. In preclinical studies, the gene expression of the total form of BDNF was reduced in the hippocampus and in the prefrontal cortex of 18-month-old aged rats and this reduction was paralleled by a significant decrease in the protein levels of the neurotrophin and of its receptor TrkB [50]. Moreover, in CA1 and CA3 sub-region of the hippocampus, aging caused reduction in total BDNF mRNA, in Exon IV-specific transcripts, and in transcripts with the long 3' UTRs, an effect that may be related to the cognitive decline observed in these animals [51]. In line with this idea, a clinical study on a cohort of 142 participants between 59 and 81 years of age has established a correlation between the reduction of serum BDNF levels and the shrinkage of the hippocampal volume with the decline in the spatial memory performance [52].

Interestingly, it has been shown that aging is also associated with an impaired responsiveness of BDNF expression and signaling to environmental stimuli [53]. For example, the abilities of challenges such as contextual learning [54], physical exercise [55] and brain lesion [56] to upregulate BDNF signaling are impaired in aged rodents.

Supporting the involvement of BDNF in age-related variations of brain plasticity, it was shown that the administration of exogenous BDNF in the frontal cortex of aged rats produced a marked increase of multi-threshold neurons of the locus coeruleus (LC), a major noradrenergic cell group with a remarkable capacity of remodeling, accompanied with a decrease in threshold current, suggesting that BDNF may contribute to functional changes in the presynaptic axon terminals of LC neurons in the aging brain [57].

Moreover, a clinical study involving 116 adults ranged in age from 20-93 investigated the effects of BDNF polymorphism on multiple indices of memory (prospective, associative, subjective complaints and item). The results suggest that genetic predisposition to the BDNF val66met polymorphism exerts an influence on multiple indices of episodic memory exacerbating age-related differences in memory across the lifespan [58]. This polymorphism caused by a valine (Val) to methionine (Met) base change at position 66 in the BDNF pro-domain [59] results in a decrease of regulated BDNF secretion and is known to contribute to the development of psychiatric disorders such as anxiety-related behavior [60] and depressive-like behavior [61]. Furthermore, this polymorphism also plays a role in age-related neurodegenerative diseases, such as

Alzheimer's disease and Parkinson's disease. Specifically, decreased expression of BDNF mRNA and protein has been reported in the hippocampus, neocortex and Meynert *nucleus basalis* of patients with AD [62-64], while a reduced BDNF level due to decreased *BDNF* gene transcription has been found in the *substantia nigra pars compacta* of patients with PD [65].

2.2 HPA axis dysfunctions

The hypothalamic–pituitary–adrenal (HPA) axis is an auto-regulating system that controls the circulating levels of glucocorticoids (cortisol in humans and corticosterone in rodents) (GCs).

In physiological condition, the paraventricular nucleus of the hypothalamus secretes corticotrophin releasing hormone (CRH) and vasopressin (VP), which are transported to the anterior pituitary where adrenocorticotropic hormone (ACTH) is released. ACTH then stimulates the adrenal cortex to secrete GCs. The activity of the HPA axis is regulated by a negative feedback mechanism: the increased levels of GCs in the blood turn off their own secretion by down-regulating the release of the CRH and of ACTH.

In the brain GCs bind two types of receptors: the mineralocorticoid (MRs) and the glucocorticoid (GRs). GRs are highly ubiquitous and expressed in most brain regions, whereas MRs are predominantly expressed in limbic regions. MR receptors have a high affinity for corticosteroids, so they are mostly occupied in basal conditions when circulating corticosteroid levels are low, while GRs have tenfold lower affinity and are more occupied as corticosteroid levels increase [66].

GCs can rapidly modulate neuronal activity through a non-genomic pathway (activation of membrane receptors that cause the release of neurotransmitters in the synaptic cleft) or a genomic pathway. In the latest, corticosteroid receptors translocate into the nucleus, where by binding the glucocorticoid responsive elements (GRE) can affect the transcriptional activity of target genes [67, 68].

The HPA axis exerts its effect following a U-shaped dose-response curve [69-72]. Indeed, low or high dose of hormone may have short term or long-term adverse consequences for the individuals, while the optimal response is achieved at intermediate doses.

For example, the exposure to an acute challenge increase the release of GCs in terms of minutes to hours with the enhancement of synaptic transmission, LTP, learning for self-preservation, while chronic stress

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exposure, lead to high levels of circulating GCs from months to years causing the suppression of synaptic functions and adaptive plasticity mechanisms leading to detrimental effect [71].

Several lines of clinical and preclinical evidence demonstrated that hyperactivity of the HPA axis contributes to neuronal and peripheral deterioration associated with aging [73-75] with a deregulation of basal HPA axis function and increased stress responsiveness [76].

In physiological conditions, cortisol is secreted over a 24-h period according to daily fluctuations [77]. In general, we can say that it rises early in the morning and decrease throughout the rest of the day [78] even if this profile may be also influenced by the wake-sleep, the light-dark cycles and the endogenous circadian rhythm [79]. In human, studies showing the correlation between changes in diurnal cortisol release and aging are variable: indeed, results are contradictory with some researcher showing a decline, others an increase probably due to individual's variability [80] and methodological flaws.

In old adults, altered morning-cortisol levels or a flatter morning-to-evening diurnal cortisol slope lead to HPA dysregulation and impaired neuroendocrine regulation, with increased vulnerability to stressors [78, 80-89]. These alterations may be involved in the clinical presentation observed in frail older subjects. Indeed, greater diurnal cortisol secretion and smaller decline in cortisol during morning hours have been associated with frailty [90, 91], whereas lower diurnal cortisol is correlated with longevity [92]. These increases occur both in male [93] and in female [94] even if the age-related changes in cortisol appear to be more clear for man, as compared to woman [86, 88].

HPA axis alterations during aging, both in humans and in rodents, are related to modification in glucocorticoid circadian rhythm [81, 95], with mild increases in morning levels and a failure to increase in the evening [81] and with changes in GC sensitivity that could lead to an impairment of the negative feedback, the so-called "glucocorticoid cascade" [96]. The impaired function of the glucocorticoid negative-feedback is associated with higher levels of circulating GCs that lead to the reduction of GR and altered GR signaling in the forebrain [97-101]. Dysfunction of negative feedback can produce higher stress–induced peaks in cortisol and prolonged stress response [102]. Moreover, aging-related elevation of glucocorticoids levels appears to threaten hippocampal neurons and may contribute to the loss of dendrite complexity in the rat hippocampus [103].

Aging is also associated with alteration of CRH and VP that may in turn account for the aberrant activity of the HPA axis. While some studies show progressive decreases of CRH expression [104, 105], the majority of the studies found increases of CRH production with advancing age [75, 83, 106-108]. Moreover, increased CRH levels during aging can be detrimental and may contribute to the development of age-related pathologies, such as depression, anxiety, neurodegeneration, immune and metabolic disorders[109].On the contrary, VP increases in parvocellular neurons during aging, while a decrease has been shown to occur in the suprachiasmatic nucleus [75].

Furthermore, the 11- beta hydroxyl steroid dehydrogenases (11 β HSD-1), the enzyme responsible of restoring active GCs is significantly increased in the hippocampus of aged mice, thus amplifying the steroid action and contributing to the aging process [110, 111].

GR down-regulation in hippocampus leads to memory impairment and HPA axis dysregulation, as well as deficits in the activation of GR-dependent transcription [99, 101, 112, 113]. Moreover, plasma glucocorticoids levels correlate with deficits in spatial learning and hippocampal degeneration in aged rats [114, 115] and with dysfunction of hippocampus-dependent memory tasks and reduced hippocampal volume in aged humans [82], while high salivary cortisol concentration predict declining performance on declarative memory tests [116]. Despite this, many older individuals have cognitive performance similar to younger ones [97, 117-120] and this can be due to the accumulation of a lifetime of experience that could alter the progression of brain aging, a phenomenon named "allostatic load" [121, 122].

2.3 Alterations of Immune/inflammatory

Another system altered in aging is the immune/inflammatory system. Historically, the brain was believed to be an immunologically privileged space, functioning fully isolated from the immune system. However, it is now accepted that there is a wide and constant bi-directional communication between the immune system and the CNS, across the relatively impenetrable blood- brain barrier (BBB) [123, 124]. As a consequence of this connection, neural activity is dramatically affected during and following an immune challenge that occurs in the periphery [125, 126], leading to the production of cytokine -signaling molecules possessing unique immunomodulatory functions- within the brain tissue, with activated microglia -resident immune

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cells of the CNS- driving the response [127]. Moreover, the discovery of the meningeal lymphatic system's capability of carrying fluids, immune cells, and macromolecules from the CNS to the draining lymph nodes [128, 129]corroborates the relevance of the signaling between the brain and the peripheral immune system.

As we age, our immune system undergoes several modifications of the inflammatory status with a decline of the adaptive immunity, called immunosenescence, coupled with a concomitant activation of the innate immune system, so-called "inflamm-aging", which may lead to a low-grade, chronic, systemic inflammation [130-132]. Immunosenescence of the acquired system has received a great attention in recent years. After birth, thymic productivity decreases with age, resulting in reduced T-cell repertoire in parallel with an increased growth of memory and effector-memory cells [133]. This imbalance culminates in a reduced ability to clear novel pathogens, prolonging infection duration as well as an increase in functionally distinct T-cell populations, which have an amplified pro-inflammatory phenotype [134]. Increases in numbers of antigen-specific cells with age are associated with an increment in the number of terminally differentiated 'senescent' cells, which occupy a large proportion of immune space. These cells, particularly CD8⁺, are extremely potent producers of inflammatory cytokines and cultures of senescent $CD8^+$ T cells show resistance to apoptosis, permanent loss of CD28 expression, altered cytokine profiles, reduced ability to respond to stress and various functional changes [135]. Moreover, with age these antigen-specific cells do not proliferate properly and after mitogenic/viral stimulation produce more IL-6 and TNF than their younger counterparts [136]. Therefore, T-cell immunosenescence is characterized by an increased proinflammatory phenotype and most probably contributes to inflamm-aging in a manner dependent on previous exposure to antigenic challenges and exhaustion of T-cell repertoire [137].

Inflamm-aging is thought to be a result of a cumulative lifetime exposure to antigenic stimuli caused by infections as well as exposure to non-infective antigens [138]. The consequent immune response, tissue injury and production of reactive oxygen species (ROS) that cause oxidative damage also triggers the release of additional cytokines, primarily from cells of the innate immune system [139]. This results in a vicious cycle, where the release of cytokines provokes a further damage, immune activation and remodeling of the tissue.

Within the innate immune system, microglia, as part of the myelomonocytic lineage, constitute the predominant cells and serve many functions including immune-surveillance of the microenvironment for pathogen invasion, danger signals, cellular debris, apoptotic cells, and alterations in neuronal phenotype [140]. In the young adult brain, normally quiescent microglia become activated in response to a threat [141] and undergo morphological changes, proliferation and production of pro-inflammatory cytokines [140]. Once the threat is resolved, alternatively activated microglia produce anti-inflammatory cytokines in addition to other signaling molecules that facilitate a return to resting state [141]. For example, alternatively activated microglial cells secrete growth factors such as BDNF, which promotes the growth, survival, differentiation of neurons and synapse formation[142]. In contrast, during normal aging, microglia undergo a process of senescence. Senescent microglia, which has been detected in aged rodents, exhibit telomere shortening when cultured [143], dissimilar brain distribution and decreased motility and phagocytic activity [144, 145], smaller dendritic arbors that look more variable in size, less circularly symmetric, and more elongated in shape [146, 147]. Moreover, histo-pathological evidences of aged human brains depict an increase in microglia expression of IL-1β and CD11b [148] and increased mRNA levels of IL-1 β were found in microglia of aged mice following inflammatory lipopolysaccharide stimulation, suggesting that an exaggerated inflammatory response occurs in healthy aging [149].

These age-related abnormalities in structure, ramification and therefore function are called "microglia dystrophy" and are indicative of a senescent phenotype [150]. Recently, Bisht et al. coined the name "dark microglia" to identify a new cells phenotype rarely present in the adult brain, marked by condensation of cytoplasm and nucleoplasm, accompanied by endoplasmic reticulum dilation -the most well-characterized sign of oxidative stress- engulfed dendritic spines and axon terminals and nuclear chromatin remodeling [151, 152]. This phenotype becomes abundant not only during pathologic state, such as chronic stress or neurodegenerative diseases, but it was also observed during normal aging [151].

In summary, even during healthy aging individuals show a progressive increase in neuroinflammation characterized by increased glial activation and elevated steady-state levels of inflammatory cytokines. This altered inflammatory status may contribute to the cognitive decline typical of elderly people as well as to other pathological conditions as well as to a state of enhanced vulnerability.

2.4. Epigenetic modifications

During aging, insults or damages that normally accumulate with age (oxidation, advanced age glycation products, DNA damage, amyloid accumulation etc.) can interfere with normal epigenetic control.

Epigenetics refers to the heritable but reversible modifications on DNA that regulate the transcription of genes. These processes are modulated, throughout the lifespan, by environmental cues that can be "remembered" due to changes in the epigenome [153]. Epigenetic mechanisms include histone modifications, DNA methylationand microRNA (miRNA).

Recently, it has been shown that DNA methylation represents a critical biomarker for aging that may predict the age of human cell type and tissue [154-157]. In particular, DNA methylation status is close to zero in embryonic and iPS and also reflects cell passage number, supporting a strict link between epigenetic and aging process [154]. Moreover, DNA methylation pattern is significantly different between centenarians and newborn [157], with a more homogeneous profile in the latest. In particular, centenarians show loss of DNA methylation in promoters poor of CpG, while hypermethylation was observed in CpG island promoters [157].

Age-associated methylation occurs in a site and tissue-specific manner, with specific hypermethylation for example in human colon or liver [158, 159].

Furthermore, age is also associated with altered expression of DNMT1 and DNMT3, two key enzymes involved in DNA methylation, with a decreased expression of DNMT1 [160-162] and an increase in DNMT3 expression [160].Regarding histone modifications, the changesin histone methylation and acetylation observed during aging [163] is associated with an unbalance between histone acetyltransferases (HATs) and histone deacetylase (HDACs) with an increase in HDACs, negative regulators of memory acquisition and maintenance [164]. Accordingly, sirtuins (SIRT), which regulates histone acetylation, affect the aging process. For example, in yeast Sir2 regulates longevity and its abundance decline with aging [165]and SIRT-6 deficient mice have premature aging traits [166]. DNA methylation and histone modifications are strictly related since age-related hypomethylated regions are associated with various histone modifications, including H3K27ac, H3K4m1, H3K4m2, H3K4m3 and H3K9ac [167].

Taking together these results suggest that the lasting changes in DNA methylation may represent a form of molecular memory, even if it is currently unclear if age-associated changes in epigenetic pattern are merely an epiphenomenon or play a casual role in the development of age-related disease.

3. Role of stressful events

As previously described, alterations of different mechanisms contributing to synaptic function may occur during normal aging. However, when and how the impairment of these mechanisms leads to frailty and its pathological consequences is still unknown. We propose that, beside the physiological alteration of these molecular mechanisms, a critical impact on the switch from healthy aging to a pathological state may derive from the inability to cope with stressful and challenging events.

It is well known that stress represents a critical risk factor for the development of several pathologies, including those associated with aging. However, another important issue to consider is that the stress impact may have a different grade of severity with respect to the time in which this experience takes place. Accordingly, we can speculate that while the stress response in young, healthy individuals is adaptive, without a health burden, if such stimuli occur in old age or in unhealthy subjects, the outcome may be different, resulting in a number of alterations that might favor the insurgence of a pathological condition.

In line with this idea, we have previously demonstrated that the expression and function of the neurotrophin BDNF are reduced in animals exposed to chronic stress at different stages of life, an effect that is associated with pathological phenotype [168-171]. Since BDNF plays a critical role in resilience and a reduction of its expression is important for stress-coping strategies [172], the changes observed in aged rats may impair resilience thus contributing to functional deficits associated with aging.

Moreover, it is known that the HPA axis is the primary stress system in the body and it is sensitive to both acute and chronic stresses thus influencing aging process [173], an effect probably mediated by glucocorticoids circulating levels [174]. During aging, acute stress can prolong physiological recovery with a higher peak in cortisol in humans [175], but in rodents a contradictory effects were reported with corticosterone response reduced, exaggerated or prolonged in old compared to young rats [81]. On the contrary, chronic stress exposure impaired HPA sensitivity, probably by reducing GR concentration in

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hippocampus and hypothalamus [98] thus leading to less functional feedback and high levels of GCs circulating [176, 177]. Nevertheless, many older adults exhibit resilience to the HPA axis dysregulation [178], by compensating and adapting to the physiological deficits that accompany aging.

In elderly humans, the higher cortisol level is also associated with increase cognitive decline [116, 173, 179, 180]. Accordingly, a critical risk factor for the poor cognitive function associated with aging is life-long exposure to stressful events [71, 181] and both sustained GC basal levels and stress can drive hippocampal neuron loss and associated cognitive decline [182]. Moreover, environmental factors seem to contribute to age associated changes in DNA methylation [154, 183-185].

Indeed, cumulative lifetime stress correlates with accelerated aging brain in humans [186]. In addition, a high number of the genes interested by changed in methylations during aging contain a functional glucocorticoid responsive elements supporting the possibility that stress-induced acceleration of epigenetic aging is mediated by glucocorticoid signaling [186, 187].

Aging may also affect the inflammatory response to stress exposure. Indeed, the overall increase of proinflammatory mediators and the general activation of microglia observed after an adverse stimuli, are exacerbated when the adverse event occur in old age[188, 189]. For example, it has been reported that the activation of peripheral innate immune system induced by lipopolysaccaride injection in 24-month old aged mice results in an exaggerated neuroinflammatory response, with a significant up-regulation of the cerebral expression of the pro-inflammatory cytokines IL-1 β and IL-6, as compared to 3-6 month old mice [190]. Interestingly, these changes were associated with behavioral alterations and with a more severe sickness behavior syndrome in aged mice [190]. A similar result was found in aged mice following a peripheral immune-challenge with *E. coli*, where the inflammatory response was not only exacerbated but also prolonged [191]. In addition, it has been demonstrated an over-reactivity of aged microglia when exposed *ex-vivo* to lipopolysaccharide[192] as well as *in vivo* after an experimental injury [193], thus leading to extended chronic inflammation.

All in all, these observations clearly indicate that the response to stress exposure is affected during aging, which may lead to an exacerbated cellular and functional response that may contribute to the switch from healthy to frailty aging (Fig.1).

4. Conclusions

There is no doubt that everyone experiences adverse events during life that impact both health and quality of life. However, trajectories of health in aging can vary significantly depending on the individual.

According to our hypothesis, a reduction of resilience may underlie an increased risk to develop pathologic conditions during aging. In line with this idea, an elder fragility would be the consequence of reduced function and responsiveness of biological systems crucial for coping with stressful events.

It is therefore fundamental to identify and characterize the mechanisms that may contribute to the resilience or the vulnerability to stressful events, since such mechanisms may be significantly altered in the aged brain. However, a major gap in this field of research is that the studies are limited to assess molecular alterations under basal or resting conditions, without information on if and how these functional impairments result in lack of stress responsiveness.

The investigation of these mechanisms will be critical to identify genes and pathways whose changes may contribute to achieve a better understanding of the risk architecture for age-related vulnerability and may represent potential targets for the development of novel pharmacological intervention.

Legend

Figure 1: Normal brain aging is characterized by alterations of neuroplasticity, epigenetic mechanisms, neuroinflammation and HPA axis function. Stress by acting on the same systems, may drive the switch between health and frailty.

List of abbreviation

NMDA (N-methyl D-Aspartate); GABA (γ -aminobutirric acid); GAD65 (Glutamic Acid Decarboxylase); hypothalamic-pituitary-adrenal (HPA); PFC (prefrontal cortex); NTFs (neurotrophic factors); BDNF (Brainderived neurotrophic factor); NT-3 (neurotrophin-3); NT-4 (neurotrophin-4); LC (locus coeruleus); Val (valine); Met (methionine); GCs (glucocorticoids); CRH (corticotrophin releasing hormone); VP (vasopressin); ACTH (adrenocorticotropic hormone); MR (mineralocorticoid); GR (glucocorticoid); GRE (glucocorticoid responsive elements); 11 β HSD (11- beta hydroxyl steroid dehydrogenases); BBB (bloodbrain barrier); CNS (central nervous system); ROS (reactive oxygen species); HATs (histone acetyltransferases); HDACs (histone deacetylase); SIRT (sirtuins)

References

[1] Fried, L.P.; Tangen, C.M.; Walston, J.; Newman, A.B.; Hirsch, C.; Gottdiener, J.; Seeman, T.; Tracy, R.; Kop, W.J.; Burke, G.; McBurnie, M.A.; Cardiovascular Health Study Collaborative Research, G., Frailty in older adults: evidence for a phenotype. *J Gerontol A Biol Sci Med Sci*, **2001**, *56*, (3), M146-156.

[2] Azpurua, J.; Eaton, B.A., Neuronal epigenetics and the aging synapse. *Front Cell Neurosci*, **2015**, *9*,
208.

[3] Morrison, J.H.; Baxter, M.G., The ageing cortical synapse: hallmarks and implications for cognitive decline. *Nat Rev Neurosci*, **2012**, *13*, (4), 240-250.

[4] Nakamura, S.; Akiguchi, I.; Kameyama, M.; Mizuno, N., Age-related changes of pyramidal cell basal dendrites in layers III and V of human motor cortex: a quantitative Golgi study. *Acta Neuropathol*, **1985**, *65*, (3-4), 281-284.

[5] Barnes, C.A., Normal aging: regionally specific changes in hippocampal synaptic transmission. *Trends Neurosci*, **1994**, *17*, (1), 13-18.

[6] Jacobs, B.; Schall, M.; Prather, M.; Kapler, E.; Driscoll, L.; Baca, S.; Jacobs, J.; Ford, K.; Wainwright,
 M.; Treml, M., Regional dendritic and spine variation in human cerebral cortex: a quantitative golgi study.
 Cereb Cortex, 2001, 11, (6), 558-571.

[7] Hof, P.R.; Duan, H.; Page, T.L.; Einstein, M.; Wicinski, B.; He, Y.; Erwin, J.M.; Morrison, J.H., Agerelated changes in GluR2 and NMDAR1 glutamate receptor subunit protein immunoreactivity in corticocortically projecting neurons in macaque and patas monkeys. *Brain Res*, **2002**, *928*, (1-2), 175-186.

[8] Duan, H.; Wearne, S.L.; Rocher, A.B.; Macedo, A.; Morrison, J.H.; Hof, P.R., Age-related dendritic and spine changes in corticocortically projecting neurons in macaque monkeys. *Cereb Cortex*, **2003**, *13*, (9), 950-961.

[9] Chang, Y.M.; Rosene, D.L.; Killiany, R.J.; Mangiamele, L.A.; Luebke, J.I., Increased action potential firing rates of layer 2/3 pyramidal cells in the prefrontal cortex are significantly related to cognitive performance in aged monkeys. *Cereb Cortex*, **2005**, *15*, (4), 409-418.

[10] Burke, S.N.; Barnes, C.A., Neural plasticity in the ageing brain. *Nat Rev Neurosci*, **2006**, *7*, (1), 30-40.

Current Medicinal Chemistry

[11] Coggan, J.S.; Grutzendler, J.; Bishop, D.L.; Cook, M.R.; Gan, W.; Heym, J.; Lichtman, J.W., Ageassociated synapse elimination in mouse parasympathetic ganglia. *J Neurobiol*, **2004**, *60*, (2), 214-226.

[12] Canas, P.M.; Duarte, J.M.; Rodrigues, R.J.; Kofalvi, A.; Cunha, R.A., Modification upon aging of the density of presynaptic modulation systems in the hippocampus. *Neurobiol Aging*, **2009**, *30*, (11), 1877-1884.

[13] Richard, M.B.; Taylor, S.R.; Greer, C.A., Age-induced disruption of selective olfactory bulb synaptic circuits. *Proc Natl Acad Sci U S A*, **2010**, *107*, (35), 15613-15618.

[14] Gonzales, R.A.; Brown, L.M.; Jones, T.W.; Trent, R.D.; Westbrook, S.L.; Leslie, S.W., N-methyl-Daspartate mediated responses decrease with age in Fischer 344 rat brain. *Neurobiol Aging*, **1991**, *12*, (3), 219-225.

[15] Pittaluga, A.; Fedele, E.; Risiglione, C.; Raiteri, M., Age-related decrease of the NMDA receptormediated noradrenaline release in rat hippocampus and partial restoration by D-cycloserine. *Eur J Pharmacol*, **1993**, *231*, (1), 129-134.

[16] Barnes, C.A.; Rao, G.; Shen, J., Age-related decrease in the N-methyl-D-aspartateR-mediated excitatory postsynaptic potential in hippocampal region CA1. *Neurobiol Aging*, **1997**, *18*, (4), 445-452.

[17] Magnusson, K.R., The aging of the NMDA receptor complex. *Front Biosci*, **1998**, *3*, e70-80.

[18] Eckles-Smith, K.; Clayton, D.; Bickford, P.; Browning, M.D., Caloric restriction prevents age-related deficits in LTP and in NMDA receptor expression. *Brain Res Mol Brain Res*, **2000**, *78*, (1-2), 154-162.

[19] Gore, A.C.; Oung, T.; Woller, M.J., Age-related changes in hypothalamic gonadotropin-releasing hormone and N-methyl-D-aspartate receptor gene expression, and their regulation by oestrogen, in the female rat. *J Neuroendocrinol*, **2002**, *14*, (4), 300-309.

[20] Liu, P.; Smith, P.F.; Darlington, C.L., Glutamate receptor subunits expression in memory-associated brain structures: regional variations and effects of aging. *Synapse*, **2008**, *62*, (11), 834-841.

[21] Zhao, X.; Rosenke, R.; Kronemann, D.; Brim, B.; Das, S.R.; Dunah, A.W.; Magnusson, K.R., The effects of aging on N-methyl-D-aspartate receptor subunits in the synaptic membrane and relationships to long-term spatial memory. *Neuroscience*, **2009**, *162*, (4), 933-945.

[22] Marin, O., Interneuron dysfunction in psychiatric disorders. *Nat Rev Neurosci*, **2012**, *13*, (2), 107120.

[23] Peinemann, A.; Lehner, C.; Conrad, B.; Siebner, H.R., Age-related decrease in paired-pulse intracortical inhibition in the human primary motor cortex. *Neurosci Lett*, **2001**, *313*, (1-2), 33-36.

[24] Pellicciari, M.C.; Miniussi, C.; Rossini, P.M.; De Gennaro, L., Increased cortical plasticity in the elderly: changes in the somatosensory cortex after paired associative stimulation. *Neuroscience*, 2009, 163, (1), 266-276.

[25] Huttunen, J.; Wikstrom, H.; Salonen, O.; Ilmoniemi, R.J., Human somatosensory cortical activation strengths: comparison between males and females and age-related changes. *Brain Res*, **1999**, *818*, (2), 196-203.

[26] Erlander, M.G.; Tillakaratne, N.J.; Feldblum, S.; Patel, N.; Tobin, A.J., Two genes encode distinct glutamate decarboxylases. *Neuron*, **1991**, *7*, (1), 91-100.

[27] Gold, J.R.; Bajo, V.M., Insult-induced adaptive plasticity of the auditory system. *Front Neurosci*, **2014**, *8*, 110.

[28] Rissman, R.A.; Mobley, W.C., Implications for treatment: GABAA receptors in aging, Down syndrome and Alzheimer's disease. *J Neurochem*, **2011**, *117*, (4), 613-622.

[29] Gutierrez, A.; Khan, Z.U.; Ruano, D.; Miralles, C.P.; Vitorica, J.; De Blas, A.L., Aging-related subunit expression changes of the GABAA receptor in the rat hippocampus. *Neuroscience*, **1996**, *74*, (2), 341-348.

[30] McQuail, J.A.; Banuelos, C.; LaSarge, C.L.; Nicolle, M.M.; Bizon, J.L., GABA(B) receptor GTP-binding is decreased in the prefrontal cortex but not the hippocampus of aged rats. *Neurobiol Aging*, **2012**, *33*, (6), 1124 e1121-1112.

[31] Calabrese, F.; Riva, M.A.; Molteni, R., Synaptic alterations associated with depression and schizophrenia: potential as a therapeutic target. *Expert Opin Ther Targets*, **2016**, *20*, (10), 1195-1207.

[32] Ramón y Cajal, S.R., Estudios sobre la degeneración y regeneracion del sistema nervioso. *Moya*, **1913-1914**.

[33] Fuchs, E.; Flugge, G., Adult neuroplasticity: more than 40 years of research. *Neural Plast*, **2014**, 2014, 541870.

Current Medicinal Chemistry

[34] Raz, N.; Lindenberger, U.; Rodrigue, K.M.; Kennedy, K.M.; Head, D.; Williamson, A.; Dahle, C.; Gerstorf, D.; Acker, J.D., Regional brain changes in aging healthy adults: general trends, individual differences and modifiers. *Cereb Cortex*, **2005**, *15*, (11), 1676-1689.

[35] Brody, H., Organization of the cerebral cortex. III. A study of aging in the human cerebral cortex. *J Comp Neurol*, **1955**, *102*, (2), 511-516.

[36] Scheibel, A.B., Dendritic changes in senile and presenile dementias. *Res Publ Assoc Res Nerv Ment Dis*, **1979**, *57*, 107-124.

[37] Hedden, T.; Gabrieli, J.D., Insights into the ageing mind: a view from cognitive neuroscience. *Nat Rev Neurosci*, **2004**, *5*, (2), 87-96.

[38] Grill, J.D.; Riddle, D.R., Age-related and laminar-specific dendritic changes in the medial frontal cortex of the rat. *Brain Res*, **2002**, *937*, (1-2), 8-21.

[39] Uylings, H.B.; de Brabander, J.M., Neuronal changes in normal human aging and Alzheimer's disease. *Brain Cogn*, **2002**, *49*, (3), 268-276.

[40] Burger, C., Region-specific genetic alterations in the aging hippocampus: implications for cognitive aging. *Front Aging Neurosci*, **2010**, *2*, 140.

[41] Olariu, A.; Cleaver, K.M.; Cameron, H.A., Decreased neurogenesis in aged rats results from loss of granule cell precursors without lengthening of the cell cycle. *J Comp Neurol*, **2007**, *501*, (4), 659-667.

[42] Kapogiannis, D.; Mattson, M.P., Disrupted energy metabolism and neuronal circuit dysfunction in cognitive impairment and Alzheimer's disease. *Lancet Neurol*, **2011**, *10*, (2), 187-198.

[43] Poo, M.M., Neurotrophins as synaptic modulators. *Nat Rev Neurosci*, **2001**, *2*, (1), 24-32.

[44] Lu, B., Pro-region of neurotrophins: role in synaptic modulation. *Neuron*, **2003**, *39*, (5), 735-738.

[45] Oliveira, S.L.; Pillat, M.M.; Cheffer, A.; Lameu, C.; Schwindt, T.T.; Ulrich, H., Functions of neurotrophins and growth factors in neurogenesis and brain repair. *Cytometry A*, **2013**, *83*, (1), 76-89.

[46] Chao, M.V.; Hempstead, B.L., p75 and Trk: a two-receptor system. *Trends Neurosci*, **1995**, *18*, (7), 321-326.

[47] Mitre, M.; Mariga, A.; Chao, M.V., Neurotrophin signalling: novel insights into mechanisms and pathophysiology. *Clin Sci (Lond)*, **2017**, *131*, (1), 13-23.

[48] Esposito, D.; Patel, P.; Stephens, R.M.; Perez, P.; Chao, M.V.; Kaplan, D.R.; Hempstead, B.L., The cytoplasmic and transmembrane domains of the p75 and Trk A receptors regulate high affinity binding to nerve growth factor. *J Biol Chem*, **2001**, *276*, (35), 32687-32695.

[49] Ziegenhorn, A.A.; Schulte-Herbruggen, O.; Danker-Hopfe, H.; Malbranc, M.; Hartung, H.D.; Anders,
D.; Lang, U.E.; Steinhagen-Thiessen, E.; Schaub, R.T.; Hellweg, R., Serum neurotrophins--a study on the time
course and influencing factors in a large old age sample. *Neurobiol Aging*, 2007, 28, (9), 1436-1445.

[50] Calabrese, F.; Guidotti, G.; Racagni, G.; Riva, M.A., Reduced neuroplasticity in aged rats: a role for the neurotrophin brain-derived neurotrophic factor. *Neurobiol Aging*, **2013**, *34*, (12), 2768-2776.

[51] Chapman, T.R.; Barrientos, R.M.; Ahrendsen, J.T.; Hoover, J.M.; Maier, S.F.; Patterson, S.L., Aging and infection reduce expression of specific brain-derived neurotrophic factor mRNAs in hippocampus. *Neurobiol Aging*, **2012**, *33*, (4), 832 e831-814.

[52] Erickson, K.I.; Prakash, R.S.; Voss, M.W.; Chaddock, L.; Heo, S.; McLaren, M.; Pence, B.D.; Martin, S.A.; Vieira, V.J.; Woods, J.A.; McAuley, E.; Kramer, A.F., Brain-derived neurotrophic factor is associated with age-related decline in hippocampal volume. *J Neurosci*, **2010**, *30*, (15), 5368-5375.

[53] Mattson, M.P.; Magnus, T., Ageing and neuronal vulnerability. *Nat Rev Neurosci*, 2006, 7, (4), 278294.

[54] Monti, B.; Berteotti, C.; Contestabile, A., Dysregulation of memory-related proteins in the hippocampus of aged rats and their relation with cognitive impairment. *Hippocampus*, **2005**, *15*, (8), 1041-1049.

[55] Adlard, P.A.; Perreau, V.M.; Cotman, C.W., The exercise-induced expression of BDNF within the hippocampus varies across life-span. *Neurobiol Aging*, **2005**, *26*, (4), 511-520.

[56] Yurek, D.M.; Fletcher-Turner, A., Lesion-induced increase of BDNF is greater in the striatum of young versus old rat brain. *Exp Neurol*, **2000**, *161*, (1), 392-396.

[57] Nakai, S.; Matsunaga, W.; Ishida, Y.; Isobe, K.; Shirokawa, T., Effects of BDNF infusion on the axon terminals of locus coeruleus neurons of aging rats. *Neurosci Res*, **2006**, *54*, (3), 213-219.

Current Medicinal Chemistry

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[58] Kennedy, K.M.; Reese, E.D.; Horn, M.M.; Sizemore, A.N.; Unni, A.K.; Meerbrey, M.E.; Kalich, A.G.,
 Jr.; Rodrigue, K.M., BDNF val66met polymorphism affects aging of multiple types of memory. *Brain Res,* 2015, 1612, 104-117.

[59] Shimizu, E.; Hashimoto, K.; Iyo, M., Ethnic difference of the BDNF 196G/A (val66met) polymorphism frequencies: the possibility to explain ethnic mental traits. *Am J Med Genet B Neuropsychiatr Genet*, **2004**, *126B*, (1), 122-123.

[60] Chen, Z.Y.; Jing, D.; Bath, K.G.; Ieraci, A.; Khan, T.; Siao, C.J.; Herrera, D.G.; Toth, M.; Yang, C.; McEwen, B.S.; Hempstead, B.L.; Lee, F.S., Genetic variant BDNF (Val66Met) polymorphism alters anxiety-related behavior. *Science*, **2006**, *314*, (5796), 140-143.

[61] Yu, H.; Wang, D.D.; Wang, Y.; Liu, T.; Lee, F.S.; Chen, Z.Y., Variant brain-derived neurotrophic factor Val66Met polymorphism alters vulnerability to stress and response to antidepressants. *J Neurosci*, **2012**, *32*, (12), 4092-4101.

[62] Phillips, H.S.; Hains, J.M.; Armanini, M.; Laramee, G.R.; Johnson, S.A.; Winslow, J.W., BDNF mRNA is decreased in the hippocampus of individuals with Alzheimer's disease. *Neuron*, **1991**, *7*, (5), 695-702.

[63] Peng, S.; Wuu, J.; Mufson, E.J.; Fahnestock, M., Precursor form of brain-derived neurotrophic factor and mature brain-derived neurotrophic factor are decreased in the pre-clinical stages of Alzheimer's disease. *J Neurochem*, **2005**, *93*, (6), 1412-1421.

[64] Lee, J.; Fukumoto, H.; Orne, J.; Klucken, J.; Raju, S.; Vanderburg, C.R.; Irizarry, M.C.; Hyman, B.T.; Ingelsson, M., Decreased levels of BDNF protein in Alzheimer temporal cortex are independent of BDNF polymorphisms. *Exp Neurol*, **2005**, *194*, (1), 91-96.

[65] Murer, M.G.; Yan, Q.; Raisman-Vozari, R., Brain-derived neurotrophic factor in the control human brain, and in Alzheimer's disease and Parkinson's disease. *Prog Neurobiol*, **2001**, *63*, (1), 71-124.

[66] Joels, M., Functional actions of corticosteroids in the hippocampus. *Eur J Pharmacol*, **2008**, *583*, (2-3), 312-321.

[67] Lu, N.Z.; Wardell, S.E.; Burnstein, K.L.; Defranco, D.; Fuller, P.J.; Giguere, V.; Hochberg, R.B.; McKay,
 L.; Renoir, J.M.; Weigel, N.L.; Wilson, E.M.; McDonnell, D.P.; Cidlowski, J.A., International Union of
 Pharmacology. LXV. The pharmacology and classification of the nuclear receptor superfamily:

glucocorticoid, mineralocorticoid, progesterone, and androgen receptors. *Pharmacol Rev*, **2006**, *58*, (4), 782-797.

[68] Revest, J.M.; Kaouane, N.; Mondin, M.; Le Roux, A.; Rouge-Pont, F.; Vallee, M.; Barik, J.; Tronche, F.; Desmedt, A.; Piazza, P.V., The enhancement of stress-related memory by glucocorticoids depends on synapsin-la/lb. *Mol Psychiatry*, **2010**, *15*, (12), 1125, 1140-1151.

[69] Chrousos, G.P., Stress and disorders of the stress system. *Nat Rev Endocrinol*, **2009**, *5*, (7), 374-381.

[70] Chrousos, G.P.; Kino, T., Intracellular glucocorticoid signaling: a formerly simple system turns stochastic. *Sci STKE*, **2005**, (304), pe48.

[71] McEwen, B.S.; Bowles, N.P.; Gray, J.D.; Hill, M.N.; Hunter, R.G.; Karatsoreos, I.N.; Nasca, C., Mechanisms of stress in the brain. *Nat Neurosci*, **2015**, *18*, (10), 1353-1363.

[72] Nicolaides, N.C.; Kyratzi, E.; Lamprokostopoulou, A.; Chrousos, G.P.; Charmandari, E., Stress, the stress system and the role of glucocorticoids. *Neuroimmunomodulation*, **2015**, *22*, (1-2), 6-19.

[73] Ferrari, E.; Magri, F., Role of neuroendocrine pathways in cognitive decline during aging. *Ageing Res Rev*, **2008**, *7*, (3), 225-233.

[74] Sapolsky, R.M., Glucocorticoids, stress, and their adverse neurological effects: relevance to aging. *Exp Gerontol*, **1999**, *34*, (6), 721-732.

[75] Swaab, D.F.; Bao, A.M.; Lucassen, P.J., The stress system in the human brain in depression and neurodegeneration. *Ageing Res Rev*, **2005**, *4*, (2), 141-194.

[76] Sapolsky, R.M.; Altmann, J., Incidence of hypercortisolism and dexamethasone resistance increases with age among wild baboons. *Biol Psychiatry*, **1991**, *30*, (10), 1008-1016.

[77] Otte, C.; Hart, S.; Neylan, T.C.; Marmar, C.R.; Yaffe, K.; Mohr, D.C., A meta-analysis of cortisol response to challenge in human aging: importance of gender. *Psychoneuroendocrinology*, **2005**, *30*, (1), 80-91.

[78] Fries, E.; Dettenborn, L.; Kirschbaum, C., The cortisol awakening response (CAR): facts and future directions. *Int J Psychophysiol*, **2009**, *72*, (1), 67-73.

[79] Van Cauter, E., Diurnal and ultradian rhythms in human endocrine function: a minireview. *Horm Res*, **1990**, *34*, (2), 45-53.

Current Medicinal Chemistry

[80] Almeida, D.M.; McGonagle, K.; King, H., Assessing daily stress processes in social surveys by combining stressor exposure and salivary cortisol. *Biodemography Soc Biol*, **2009**, *55*, (2), 219-237.

[81] Lightman, S.L.; Windle, R.J.; Julian, M.D.; Harbuz, M.S.; Shanks, N.; Wood, S.A.; Kershaw, Y.M.; Ingram, C.D., Significance of pulsatility in the HPA axis. *Novartis Found Symp*, **2000**, *227*, 244-257; discussion 257-260.

[82] Lupien, S.J.; de Leon, M.; de Santi, S.; Convit, A.; Tarshish, C.; Nair, N.P.; Thakur, M.; McEwen, B.S.;
Hauger, R.L.; Meaney, M.J., Cortisol levels during human aging predict hippocampal atrophy and memory deficits. *Nat Neurosci*, **1998**, *1*, (1), 69-73.

[83] Tizabi, Y.; Aguilera, G.; Gilad, G.M., Age-related reduction in pituitary corticotropin-releasing hormone receptors in two rat strains. *Neurobiol Aging*, **1992**, *13*, (2), 227-230.

[84] Purnell, J.Q.; Brandon, D.D.; Isabelle, L.M.; Loriaux, D.L.; Samuels, M.H., Association of 24-hour cortisol production rates, cortisol-binding globulin, and plasma-free cortisol levels with body composition, leptin levels, and aging in adult men and women. *J Clin Endocrinol Metab*, **2004**, *89*, (1), 281-287.

[85] Seeman, T.E.; Singer, B.; Wilkinson, C.W.; McEwen, B., Gender differences in age-related changes in HPA axis reactivity. *Psychoneuroendocrinology*, **2001**, *26*, (3), 225-240.

[86] Adam, E.K.; Hawkley, L.C.; Kudielka, B.M.; Cacioppo, J.T., Day-to-day dynamics of experience-cortisol associations in a population-based sample of older adults. *Proc Natl Acad Sci U S A*, **2006**, *103*, (45), 17058-17063.

[87] Dmitrieva, N.O.; Almeida, D.M.; Dmitrieva, J.; Loken, E.; Pieper, C.F., A day-centered approach to modeling cortisol: diurnal cortisol profiles and their associations among U.S. adults. *Psychoneuroendocrinology*, **2013**, *38*, (10), 2354-2365.

[88] Karlamangla, A.S.; Friedman, E.M.; Seeman, T.E.; Stawksi, R.S.; Almeida, D.M., Daytime trajectories of cortisol: demographic and socioeconomic differences--findings from the National Study of Daily Experiences. *Psychoneuroendocrinology*, **2013**, *38*, (11), 2585-2597.

[89] Nater, U.M.; Hoppmann, C.A.; Scott, S.B., Diurnal profiles of salivary cortisol and alpha-amylase change across the adult lifespan: evidence from repeated daily life assessments. *Psychoneuroendocrinology*, **2013**, *38*, (12), 3167-3171.

[90] Johar, H.; Emeny, R.T.; Bidlingmaier, M.; Reincke, M.; Thorand, B.; Peters, A.; Heier, M.; Ladwig, K.H., Blunted diurnal cortisol pattern is associated with frailty: a cross-sectional study of 745 participants aged 65 to 90 years. *J Clin Endocrinol Metab*, **2014**, *99*, (3), E464-468.

[91] Varadhan, R.; Walston, J.; Cappola, A.R.; Carlson, M.C.; Wand, G.S.; Fried, L.P., Higher levels and blunted diurnal variation of cortisol in frail older women. *J Gerontol A Biol Sci Med Sci*, **2008**, *63*, (2), 190-195.

[92] Noordam, R.; Jansen, S.W.; Akintola, A.A.; Oei, N.Y.; Maier, A.B.; Pijl, H.; Slagboom, P.E.; Westendorp, R.G.; van der Grond, J.; de Craen, A.J.; van Heemst, D.; Leiden Longevity Study, g., Familial longevity is marked by lower diurnal salivary cortisol levels: the Leiden Longevity Study. *PLoS One*, **2012**, *7*, (2), e31166.

[93] Belanger, A.; Candas, B.; Dupont, A.; Cusan, L.; Diamond, P.; Gomez, J.L.; Labrie, F., Changes in serum concentrations of conjugated and unconjugated steroids in 40- to 80-year-old men. *J Clin Endocrinol Metab*, **1994**, *79*, (4), 1086-1090.

[94] Murakami, K.; Nakagawa, T.; Shozu, M.; Uchide, K.; Koike, K.; Inoue, M., Changes with aging of steroidal levels in the cerebrospinal fluid of women. *Maturitas*, **1999**, *33*, (1), 71-80.

[95] Linkowski, P.; Van Onderbergen, A.; Kerkhofs, M.; Bosson, D.; Mendlewicz, J.; Van Cauter, E., Twin study of the 24-h cortisol profile: evidence for genetic control of the human circadian clock. *Am J Physiol*, 1993, *264*, (2 Pt 1), E173-181.

[96] Sapolsky, R.M.; Krey, L.C.; McEwen, B.S., The neuroendocrinology of stress and aging: the glucocorticoid cascade hypothesis. *Endocr Rev*, **1986**, *7*, (3), 284-301.

[97] Issa, A.M.; Rowe, W.; Gauthier, S.; Meaney, M.J., Hypothalamic-pituitary-adrenal activity in aged, cognitively impaired and cognitively unimpaired rats. *J Neurosci*, **1990**, *10*, (10), 3247-3254.

[98] Ferrari, E.; Magri, F.; Dori, D.; Migliorati, G.; Nescis, T.; Molla, G.; Fioravanti, M.; Solerte, S.B., Neuroendocrine correlates of the aging brain in humans. *Neuroendocrinology*, **1995**, *61*, (4), 464-470.

[99] Bizon, J.L.; Helm, K.A.; Han, J.S.; Chun, H.J.; Pucilowska, J.; Lund, P.K.; Gallagher, M., Hypothalamicpituitary-adrenal axis function and corticosterone receptor expression in behaviourally characterized young and aged Long-Evans rats. *Eur J Neurosci*, **2001**, *14*, (10), 1739-1751.

Current Medicinal Chemistry

[100] Lund, P.K.; Hoyt, E.C.; Bizon, J.; Smith, D.R.; Haberman, R.; Helm, K.; Gallagher, M., Transcriptional mechanisms of hippocampal aging. *Exp Gerontol*, **2004**, *39*, (11-12), 1613-1622.

[101] Lee, S.Y.; Hwang, Y.K.; Yun, H.S.; Han, J.S., Decreased levels of nuclear glucocorticoid receptor protein in the hippocampus of aged Long-Evans rats with cognitive impairment. *Brain Res*, **2012**, *1478*, 48-54.

[102] Sapolsky, R., Sick of poverty. *Sci Am*, **2005**, *293*, (6), 92-99.

[103] Hibberd, C.; Yau, J.L.; Seckl, J.R., Glucocorticoids and the ageing hippocampus. J Anat, 2000, 197 Pt4, 553-562.

[104] Cizza, G.; Calogero, A.E.; Brady, L.S.; Bagdy, G.; Bergamini, E.; Blackman, M.R.; Chrousos, G.P.; Gold, P.W., Male Fischer 344/N rats show a progressive central impairment of the hypothalamic-pituitary-adrenal axis with advancing age. *Endocrinology*, **1994**, *134*, (4), 1611-1620.

[105] Kasckow, J.W.; Regmi, A.; Mulchahey, J.J.; Plotsky, P.M.; Hauger, R.L., Changes in brain corticotropin-releasing factor messenger RNA expression in aged Fischer 344 rats. *Brain Res*, 1999, *822*, (1-2), 228-230.

[106] Hauger, R.L.; Thrivikraman, K.V.; Plotsky, P.M., Age-related alterations of hypothalamic-pituitaryadrenal axis function in male Fischer 344 rats. *Endocrinology*, **1994**, *134*, (3), 1528-1536.

[107] Ceccatelli, S.; Calza, L.; Giardino, L., Age-related changes in the expression of corticotropin-releasing hormone receptor mRNA in the rat pituitary. *Brain Res Mol Brain Res*, **1996**, *37*, (1-2), 175-180.

[108] Meijer, O.C.; Topic, B.; Steenbergen, P.J.; Jocham, G.; Huston, J.P.; Oitzl, M.S., Correlations between hypothalamus-pituitary-adrenal axis parameters depend on age and learning capacity. *Endocrinology*, **2005**, *146*, (3), 1372-1381.

[109] Aguilera, G., HPA axis responsiveness to stress: implications for healthy aging. *Exp Gerontol*, **2011**, 46, (2-3), 90-95.

[110] Cooper, M.S., 11beta-Hydroxysteroid dehydrogenase: a regulator of glucocorticoid response in osteoporosis. *J Endocrinol Invest*, **2008**, *31*, (7 Suppl), 16-21.

[111] Holmes, M.C.; Carter, R.N.; Noble, J.; Chitnis, S.; Dutia, A.; Paterson, J.M.; Mullins, J.J.; Seckl, J.R.; Yau, J.L., 11beta-hydroxysteroid dehydrogenase type 1 expression is increased in the aged mouse hippocampus and parietal cortex and causes memory impairments. *J Neurosci*, **2010**, *30*, (20), 6916-6920.

[112] Sapolsky, R.M.; Krey, L.C.; McEwen, B.S., Corticosterone receptors decline in a site-specific manner in the aged rat brain. *Brain Res*, **1983**, *289*, (1-2), 235-240.

[113] Murphy, E.K.; Spencer, R.L.; Sipe, K.J.; Herman, J.P., Decrements in nuclear glucocorticoid receptor (GR) protein levels and DNA binding in aged rat hippocampus. *Endocrinology*, **2002**, *143*, (4), 1362-1370.

[114] Landfield, P.W.; Baskin, R.K.; Pitler, T.A., Brain aging correlates: retardation by hormonalpharmacological treatments. *Science*, **1981**, *214*, (4520), 581-584.

[115] Landfield, P.W.; Waymire, J.C.; Lynch, G., Hippocampal aging and adrenocorticoids: quantitative correlations. *Science*, **1978**, *202*, (4372), 1098-1102.

[116] Li, G.; Cherrier, M.M.; Tsuang, D.W.; Petrie, E.C.; Colasurdo, E.A.; Craft, S.; Schellenberg, G.D.;
 Peskind, E.R.; Raskind, M.A.; Wilkinson, C.W., Salivary cortisol and memory function in human aging.
 Neurobiol Aging, 2006, 27, (11), 1705-1714.

[117] Gage, F.H.; Dunnett, S.B.; Bjorklund, A., Spatial learning and motor deficits in aged rats. *Neurobiol Aging*, **1984**, *5*, (1), 43-48.

[118] Gallagher, M.; Pelleymounter, M.A., An age-related spatial learning deficit: choline uptake distinguishes "impaired" and "unimpaired" rats. *Neurobiol Aging*, **1988**, *9*, (4), 363-369.

[119] Markowska, A.L.; Stone, W.S.; Ingram, D.K.; Reynolds, J.; Gold, P.E.; Conti, L.H.; Pontecorvo, M.J.;
 Wenk, G.L.; Olton, D.S., Individual differences in aging: behavioral and neurobiological correlates. *Neurobiol Aging*, **1989**, *10*, (1), 31-43.

[120] Matzel, L.D.; Grossman, H.; Light, K.; Townsend, D.; Kolata, S., Age-related declines in general cognitive abilities of Balb/C mice are associated with disparities in working memory, body weight, and general activity. *Learn Mem*, **2008**, *15*, (10), 733-746.

[121] McEwen, B.S., Allostasis, allostatic load, and the aging nervous system: role of excitatory amino acids and excitotoxicity. *Neurochem Res*, **2000**, *25*, (9-10), 1219-1231.

Current Medicinal Chemistry

[122] Stewart, J.A., The detrimental effects of allostasis: allostatic load as a measure of cumulative stress. *J Physiol Anthropol*, **2006**, *25*, (1), 133-145.

[123] Banks, W.A., The blood-brain barrier in neuroimmunology: Tales of separation and assimilation. *Brain Behav Immun*, **2015**, *44*, 1-8.

[124] Ellwardt, E.; Walsh, J.T.; Kipnis, J.; Zipp, F., Understanding the Role of T Cells in CNS Homeostasis. *Trends Immunol*, **2016**, *37*, (2), 154-165.

[125] Maier, S.F.; Watkins, L.R., Cytokines for psychologists: implications of bidirectional immune-to-brain communication for understanding behavior, mood, and cognition. *Psychol Rev*, **1998**, *105*, (1), 83-107.

[126] Maier, S.F., Bi-directional immune-brain communication: Implications for understanding stress, pain, and cognition. *Brain Behav Immun*, **2003**, *17*, (2), 69-85.

[127] Turrin, N.P.; Gayle, D.; Ilyin, S.E.; Flynn, M.C.; Langhans, W.; Schwartz, G.J.; Plata-Salaman, C.R., Proinflammatory and anti-inflammatory cytokine mRNA induction in the periphery and brain following intraperitoneal administration of bacterial lipopolysaccharide. *Brain Res Bull*, **2001**, *54*, (4), 443-453.

[128] Louveau, A.; Smirnov, I.; Keyes, T.J.; Eccles, J.D.; Rouhani, S.J.; Peske, J.D.; Derecki, N.C.; Castle, D.;
 Mandell, J.W.; Lee, K.S.; Harris, T.H.; Kipnis, J., Structural and functional features of central nervous system
 lymphatic vessels. *Nature*, **2015**, *523*, (7560), 337-341.

[129] Raper, D.; Louveau, A.; Kipnis, J., How Do Meningeal Lymphatic Vessels Drain the CNS? *Trends Neurosci*, **2016**, *39*, (9), 581-586.

[130] Xia, S.; Zhang, X.; Zheng, S.; Khanabdali, R.; Kalionis, B.; Wu, J.; Wan, W.; Tai, X., An Update on Inflamm-Aging: Mechanisms, Prevention, and Treatment. *J Immunol Res*, **2016**, *2016*, 8426874.

[131] Franceschi, C.; Bonafe, M.; Valensin, S.; Olivieri, F.; De Luca, M.; Ottaviani, E.; De Benedictis, G., Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann N Y Acad Sci*, **2000**, *908*, 244-254.

[132] Franceschi, C.; Campisi, J., Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *J Gerontol A Biol Sci Med Sci*, **2014**, *69 Suppl 1*, S4-9.

[133] Pawelec, G., Hallmarks of human "immunosenescence": adaptation or dysregulation? *Immun Ageing*, **2012**, *9*, (1), 15.

[134] Fagiolo, U.; Cossarizza, A.; Scala, E.; Fanales-Belasio, E.; Ortolani, C.; Cozzi, E.; Monti, D.; Franceschi,

C.; Paganelli, R., Increased cytokine production in mononuclear cells of healthy elderly people. *Eur J Immunol*, **1993**, *23*, (9), 2375-2378.

[135] Effros, R.B.; Dagarag, M.; Spaulding, C.; Man, J., The role of CD8+ T-cell replicative senescence in human aging. *Immunol Rev*, **2005**, *205*, 147-157.

[136] Vescovini, R.; Biasini, C.; Fagnoni, F.F.; Telera, A.R.; Zanlari, L.; Pedrazzoni, M.; Bucci, L.; Monti, D.;
 Medici, M.C.; Chezzi, C.; Franceschi, C.; Sansoni, P., Massive load of functional effector CD4+ and CD8+ T
 cells against cytomegalovirus in very old subjects. *J Immunol*, **2007**, *179*, (6), 4283-4291.

[137] Baylis, D.; Bartlett, D.B.; Patel, H.P.; Roberts, H.C., Understanding how we age: insights into inflammaging. *Longev Healthspan*, **2013**, *2*, (1), 8.

[138] De Martinis, M.; Franceschi, C.; Monti, D.; Ginaldi, L., Inflamm-ageing and lifelong antigenic load as major determinants of ageing rate and longevity. *FEBS Lett*, **2005**, *579*, (10), 2035-2039.

[139] Cannizzo, E.S.; Clement, C.C.; Sahu, R.; Follo, C.; Santambrogio, L., Oxidative stress, inflamm-aging and immunosenescence. *J Proteomics*, **2011**, *74*, (11), 2313-2323.

[140] Kreutzberg, G.W., Microglia: a sensor for pathological events in the CNS. *Trends Neurosci*, **1996**, *19*, (8), 312-318.

[141] Colton, C.A., Heterogeneity of microglial activation in the innate immune response in the brain. *J Neuroimmune Pharmacol*, **2009**, *4*, (4), 399-418.

[142] Parkhurst, C.N.; Yang, G.; Ninan, I.; Savas, J.N.; Yates, J.R., 3rd; Lafaille, J.J.; Hempstead, B.L.; Littman, D.R.; Gan, W.B., Microglia promote learning-dependent synapse formation through brain-derived neurotrophic factor. *Cell*, **2013**, *155*, (7), 1596-1609.

[143] Flanary, B.E.; Streit, W.J., Telomeres shorten with age in rat cerebellum and cortex in vivo. *J Anti Aging Med*, **2003**, *6*, (4), 299-308.

[144] Flanary, B.E.; Sammons, N.W.; Nguyen, C.; Walker, D.; Streit, W.J., Evidence that aging and amyloid promote microglial cell senescence. *Rejuvenation Res*, **2007**, *10*, (1), 61-74.

[145] Hefendehl, J.K.; Neher, J.J.; Suhs, R.B.; Kohsaka, S.; Skodras, A.; Jucker, M., Homeostatic and injuryinduced microglia behavior in the aging brain. *Aging Cell*, **2014**, *13*, (1), 60-69.

Current Medicinal Chemistry

[146] Sierra, A.; Gottfried-Blackmore, A.C.; McEwen, B.S.; Bulloch, K., Microglia derived from aging mice exhibit an altered inflammatory profile. *Glia*, **2007**, *55*, (4), 412-424.

[147] Tremblay, M.E.; Zettel, M.L.; Ison, J.R.; Allen, P.D.; Majewska, A.K., Effects of aging and sensory loss on glial cells in mouse visual and auditory cortices. *Glia*, **2012**, *60*, (4), 541-558.

[148] Sheng, J.G.; Mrak, R.E.; Griffin, W.S., Enlarged and phagocytic, but not primed, interleukin-1 alphaimmunoreactive microglia increase with age in normal human brain. *Acta Neuropathol*, **1998**, *95*, (3), 229-234.

[149] Henry, C.J.; Huang, Y.; Wynne, A.M.; Godbout, J.P., Peripheral lipopolysaccharide (LPS) challenge promotes microglial hyperactivity in aged mice that is associated with exaggerated induction of both proinflammatory IL-1beta and anti-inflammatory IL-10 cytokines. *Brain Behav Immun*, **2009**, *23*, (3), 309-317.

[150] Koellhoffer, E.C.; McCullough, L.D.; Ritzel, R.M., Old Maids: Aging and Its Impact on Microglia Function. *Int J Mol Sci*, **2017**, *18*, (4).

[151] Bisht, K.; Sharma, K.P.; Lecours, C.; Sanchez, M.G.; El Hajj, H.; Milior, G.; Olmos-Alonso, A.; Gomez-Nicola, D.; Luheshi, G.; Vallieres, L.; Branchi, I.; Maggi, L.; Limatola, C.; Butovsky, O.; Tremblay, M.E., Dark microglia: A new phenotype predominantly associated with pathological states. *Glia*, **2016**, *64*, (5), 826-839.
[152] Schonthal, A.H., Endoplasmic reticulum stress: its role in disease and novel prospects for therapy. *Scientifica (Cairo)*, **2012**, *2012*, 857516.

[153] Boyd-Kirkup, J.D.; Green, C.D.; Wu, G.; Wang, D.; Han, J.D., Epigenomics and the regulation of aging. *Epigenomics*, **2013**, *5*, (2), 205-227.

[154] Horvath, S., DNA methylation age of human tissues and cell types. *Genome Biol*, **2013**, *14*, (10), R115.

[155] Horvath, S.; Zhang, Y.; Langfelder, P.; Kahn, R.S.; Boks, M.P.; van Eijk, K.; van den Berg, L.H.; Ophoff,
R.A., Aging effects on DNA methylation modules in human brain and blood tissue. *Genome Biol*, **2012**, *13*,
(10), R97.

[156] Florath, I.; Butterbach, K.; Muller, H.; Bewerunge-Hudler, M.; Brenner, H., Cross-sectional and longitudinal changes in DNA methylation with age: an epigenome-wide analysis revealing over 60 novel age-associated CpG sites. *Hum Mol Genet*, **2014**, *23*, (5), 1186-1201.

[157] Heyn, H.; Li, N.; Ferreira, H.J.; Moran, S.; Pisano, D.G.; Gomez, A.; Diez, J.; Sanchez-Mut, J.V.; Setien,
F.; Carmona, F.J.; Puca, A.A.; Sayols, S.; Pujana, M.A.; Serra-Musach, J.; Iglesias-Platas, I.; Formiga, F.;
Fernandez, A.F.; Fraga, M.F.; Heath, S.C.; Valencia, A.; Gut, I.G.; Wang, J.; Esteller, M., Distinct DNA
methylomes of newborns and centenarians. *Proc Natl Acad Sci U S A*, **2012**, *109*, (26), 10522-10527.

[158] Issa, J.P.; Ottaviano, Y.L.; Celano, P.; Hamilton, S.R.; Davidson, N.E.; Baylin, S.B., Methylation of the oestrogen receptor CpG island links ageing and neoplasia in human colon. *Nat Genet*, **1994**, *7*, (4), 536-540.

[159] Choi, E.K.; Uyeno, S.; Nishida, N.; Okumoto, T.; Fujimura, S.; Aoki, Y.; Nata, M.; Sagisaka, K.; Fukuda,

Y.; Nakao, K.; Yoshimoto, T.; Kim, Y.S.; Ono, T., Alterations of c-fos gene methylation in the processes of aging and tumorigenesis in human liver. *Mutat Res*, **1996**, *354*, (1), 123-128.

[160] Casillas, M.A., Jr.; Lopatina, N.; Andrews, L.G.; Tollefsbol, T.O., Transcriptional control of the DNA methyltransferases is altered in aging and neoplastically-transformed human fibroblasts. *Mol Cell Biochem*, 2003, 252, (1-2), 33-43.

[161] Zhang, Z.; Deng, C.; Lu, Q.; Richardson, B., Age-dependent DNA methylation changes in the ITGAL
 (CD11a) promoter. *Mech Ageing Dev*, 2002, *123*, (9), 1257-1268.

[162] Li, Y.; Liu, Y.; Strickland, F.M.; Richardson, B., Age-dependent decreases in DNA methyltransferase levels and low transmethylation micronutrient levels synergize to promote overexpression of genes implicated in autoimmunity and acute coronary syndromes. *Exp Gerontol*, **2010**, *45*, (4), 312-322.

[163] Calvanese, V.; Lara, E.; Kahn, A.; Fraga, M.F., The role of epigenetics in aging and age-related diseases. *Ageing Res Rev*, **2009**, *8*, (4), 268-276.

[164] McQuown, S.C.; Barrett, R.M.; Matheos, D.P.; Post, R.J.; Rogge, G.A.; Alenghat, T.; Mullican, S.E.; Jones, S.; Rusche, J.R.; Lazar, M.A.; Wood, M.A., HDAC3 is a critical negative regulator of long-term memory formation. *J Neurosci*, **2011**, *31*, (2), 764-774.

[165] Russell, S.J.; Kahn, C.R., Endocrine regulation of ageing. *Nat Rev Mol Cell Biol*, **2007**, *8*, (9), 681-691.

[166] Mostoslavsky, R.; Chua, K.F.; Lombard, D.B.; Pang, W.W.; Fischer, M.R.; Gellon, L.; Liu, P.;
Mostoslavsky, G.; Franco, S.; Murphy, M.M.; Mills, K.D.; Patel, P.; Hsu, J.T.; Hong, A.L.; Ford, E.; Cheng, H.L.;
Kennedy, C.; Nunez, N.; Bronson, R.; Frendewey, D.; Auerbach, W.; Valenzuela, D.; Karow, M.; Hottiger,
M.O.; Hursting, S.; Barrett, J.C.; Guarente, L.; Mulligan, R.; Demple, B.; Yancopoulos, G.D.; Alt, F.W.,

Current Medicinal Chemistry

Genomic instability and aging-like phenotype in the absence of mammalian SIRT6. *Cell*, **2006**, *124*, (2), 315-329.

[167] McClay, J.L.; Aberg, K.A.; Clark, S.L.; Nerella, S.; Kumar, G.; Xie, L.Y.; Hudson, A.D.; Harada, A.; Hultman, C.M.; Magnusson, P.K.; Sullivan, P.F.; Van Den Oord, E.J., A methylome-wide study of aging using massively parallel sequencing of the methyl-CpG-enriched genomic fraction from blood in over 700 subjects. *Hum Mol Genet*, **2014**, *23*, (5), 1175-1185.

[168] Luoni, A.; Berry, A.; Calabrese, F.; Capoccia, S.; Bellisario, V.; Gass, P.; Cirulli, F.; Riva, M.A., Delayed BDNF alterations in the prefrontal cortex of rats exposed to prenatal stress: preventive effect of lurasidone treatment during adolescence. *Eur Neuropsychopharmacol*, **2014**, *24*, (6), 986-995.

[169] Luoni, A.; Macchi, F.; Papp, M.; Molteni, R.; Riva, M.A., Lurasidone exerts antidepressant properties in the chronic mild stress model through the regulation of synaptic and neuroplastic mechanisms in the rat prefrontal cortex. *Int J Neuropsychopharmacol*, **2014**, *18*, (4).

[170] Roceri, M.; Cirulli, F.; Pessina, C.; Peretto, P.; Racagni, G.; Riva, M.A., Postnatal repeated maternal deprivation produces age-dependent changes of brain-derived neurotrophic factor expression in selected rat brain regions. *Biol Psychiatry*, **2004**, *55*, (7), 708-714.

[171] Roceri, M.; Hendriks, W.; Racagni, G.; Ellenbroek, B.A.; Riva, M.A., Early maternal deprivation reduces the expression of BDNF and NMDA receptor subunits in rat hippocampus. *Mol Psychiatry*, 2002, *7*, (6), 609-616.

[172] Taliaz, D.; Loya, A.; Gersner, R.; Haramati, S.; Chen, A.; Zangen, A., Resilience to chronic stress is mediated by hippocampal brain-derived neurotrophic factor. *J Neurosci*, **2011**, *31*, (12), 4475-4483.

[173] Pardon, M.C., Stress and ageing interactions: a paradox in the context of shared etiological and physiopathological processes. *Brain Res Rev*, **2007**, *54*, (2), 251-273.

[174] Gaffey, A.E.; Bergeman, C.S.; Clark, L.A.; Wirth, M.M., Aging and the HPA axis: Stress and resilience in older adults. *Neurosci Biobehav Rev*, **2016**, *68*, 928-945.

[175] Van Cauter, E.; Leproult, R.; Kupfer, D.J., Effects of gender and age on the levels and circadian rhythmicity of plasma cortisol. *J Clin Endocrinol Metab*, **1996**, *81*, (7), 2468-2473.

[176] Seeman, T.E.; Robbins, R.J., Aging and hypothalamic-pituitary-adrenal response to challenge in humans. *Endocr Rev*, **1994**, *15*, (2), 233-260.

[177] Wilkinson, C.W.; Peskind, E.R.; Raskind, M.A., Decreased hypothalamic-pituitary-adrenal axis sensitivity to cortisol feedback inhibition in human aging. *Neuroendocrinology*, **1997**, *65*, (1), 79-90.

[178] Hildon, Z.; Smith, G.; Netuveli, G.; Blane, D., Understanding adversity and resilience at older ages. *Sociol Health Illn*, **2008**, *30*, (5), 726-740.

[179] Lupien, S.J.; Fiocco, A.; Wan, N.; Maheu, F.; Lord, C.; Schramek, T.; Tu, M.T., Stress hormones and human memory function across the lifespan. *Psychoneuroendocrinology*, **2005**, *30*, (3), 225-242.

[180] Meaney, M.J.; O'Donnell, D.; Rowe, W.; Tannenbaum, B.; Steverman, A.; Walker, M.; Nair, N.P.; Lupien, S., Individual differences in hypothalamic-pituitary-adrenal activity in later life and hippocampal aging. *Exp Gerontol*, **1995**, *30*, (3-4), 229-251.

[181] Pardon, M.C.; Rattray, I., What do we know about the long-term consequences of stress on ageing and the progression of age-related neurodegenerative disorders? *Neurosci Biobehav Rev*, **2008**, *32*, (6), 1103-1120.

[182] Pedersen, W.A.; Wan, R.; Mattson, M.P., Impact of aging on stress-responsive neuroendocrine systems. *Mech Ageing Dev*, **2001**, *122*, (9), 963-983.

[183] Fraga, M.F.; Ballestar, E.; Paz, M.F.; Ropero, S.; Setien, F.; Ballestar, M.L.; Heine-Suner, D.; Cigudosa, J.C.; Urioste, M.; Benitez, J.; Boix-Chornet, M.; Sanchez-Aguilera, A.; Ling, C.; Carlsson, E.; Poulsen, P.; Vaag, A.; Stephan, Z.; Spector, T.D.; Wu, Y.Z.; Plass, C.; Esteller, M., Epigenetic differences arise during the lifetime of monozygotic twins. *Proc Natl Acad Sci U S A*, **2005**, *102*, (30), 10604-10609.

[184] Gronniger, E.; Weber, B.; Heil, O.; Peters, N.; Stab, F.; Wenck, H.; Korn, B.; Winnefeld, M.; Lyko, F., Aging and chronic sun exposure cause distinct epigenetic changes in human skin. *PLoS Genet*, **2010**, *6*, (5), e1000971.

[185] Hannum, G.; Guinney, J.; Zhao, L.; Zhang, L.; Hughes, G.; Sadda, S.; Klotzle, B.; Bibikova, M.; Fan, J.B.; Gao, Y.; Deconde, R.; Chen, M.; Rajapakse, I.; Friend, S.; Ideker, T.; Zhang, K., Genome-wide methylation profiles reveal quantitative views of human aging rates. *Mol Cell*, **2013**, *49*, (2), 359-367.

Current Medicinal Chemistry

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[186] Zannas, A.S.; Arloth, J.; Carrillo-Roa, T.; Iurato, S.; Roh, S.; Ressler, K.J.; Nemeroff, C.B.; Smith, A.K.;
Bradley, B.; Heim, C.; Menke, A.; Lange, J.F.; Bruckl, T.; Ising, M.; Wray, N.R.; Erhardt, A.; Binder, E.B.;
Mehta, D., Lifetime stress accelerates epigenetic aging in an urban, African American cohort: relevance of glucocorticoid signaling. *Genome Biol*, **2015**, *16*, 266.

[187] Zannas, A.S.; Chrousos, G.P., Epigenetic programming by stress and glucocorticoids along the human lifespan. *Mol Psychiatry*, **2017**, *22*, (5), 640-646.

[188] Calcia, M.A.; Bonsall, D.R.; Bloomfield, P.S.; Selvaraj, S.; Barichello, T.; Howes, O.D., Stress and neuroinflammation: a systematic review of the effects of stress on microglia and the implications for mental illness. *Psychopharmacology (Berl)*, **2016**, *233*, (9), 1637-1650.

[189] Liu, Y.Z.; Wang, Y.X.; Jiang, C.L., Inflammation: The Common Pathway of Stress-Related Diseases. *Front Hum Neurosci*, **2017**, *11*, 316.

[190] Godbout, J.P.; Chen, J.; Abraham, J.; Richwine, A.F.; Berg, B.M.; Kelley, K.W.; Johnson, R.W., Exaggerated neuroinflammation and sickness behavior in aged mice following activation of the peripheral innate immune system. *FASEB J*, **2005**, *19*, (10), 1329-1331.

[191] Barrientos, R.M.; Watkins, L.R.; Rudy, J.W.; Maier, S.F., Characterization of the sickness response in young and aging rats following E. coli infection. *Brain Behav Immun*, **2009**, *23*, (4), 450-454.

[192] Frank, M.G.; Barrientos, R.M.; Watkins, L.R.; Maier, S.F., Aging sensitizes rapidly isolated hippocampal microglia to LPS ex vivo. *J Neuroimmunol*, **2010**, *226*, (1-2), 181-184.

[193] Norden, D.M.; Muccigrosso, M.M.; Godbout, J.P., Microglial priming and enhanced reactivity to secondary insult in aging, and traumatic CNS injury, and neurodegenerative disease. *Neuropharmacology*, 2015, *96*, (Pt A), 29-41.

