Review

#### Unraveling the molecular mechanisms and the potential chemopreventive/therapeutic properties of natural compounds in melanoma

Fabrizio Fontana

Michela Raimondi

Alessandro Di Domizio<sup>a, l</sup>

Roberta M. Moretti

Marina Montagnani Marelli (The surname of Marina montagnani Marelli is: Montagnani Marelli (2 surnames), only 1 first name (Marina))

Patrizia Limonta<sup>a, 3</sup>

patrizia.limonta@unimi.it

<sup>a</sup>Department of Pharmacological and Biomolecular Sciences, University of Milano, Milano, Italy

<sup>b</sup>SPILLOproject, 20037, Paderno Dugnano, Milano, Italy<sup>1</sup>

\*Corresponding author at: Department of Pharmacological and Biomolecular Sciences, University of Milano, Via Balzaretti 9, 20133, Milano, Italy.

<sup>1</sup>(Website: www.spilloproject.com).

#### Abstract

Melanoma is the most fatal form of skin cancer. Current therapeutic approaches include surgical resection, chemotherapy, targeted therapy and immunotherapy. However, these treatment strategies are associated with development of drug resistance and severe side effects. In recent years, natural compounds have also been extensively studied for their anti-melanoma effects, including tumor growth inhibition, apoptosis induction, angiogenesis and metastasis suppression and cancer stem cell elimination. Moreover, a considerable number of studies reported the synergistic activity of phytochemicals and standard anti-melanoma agents, as well as the enhanced effectiveness of their synthetic derivatives and novel formulations. However, clinical data confirming these promising effects in patients are still scanty. This review emphasizes the anti-tumor mechanisms and potential application of the most studied natural products for melanoma prevention and treatment.

Keywords: Melanoma; Natural compounds; Phytochemicals; Chemoprevention; Novel therapeutic strategies

## 1 Introduction

Malignant melanoma represents the most aggressive form of skin cancer, with an increasing incidence worldwide. It derives from the transformation and uncontrolled growth of melanocytes, and it is characterized by different progression stages: early melanomas (stages 0 and 1) are localized and noninvasive; stage II tumors are larger and often present ulcerations, with a high risk to metastasize; advanced melanomas (stages III and IV) have already colonized other body tissues. The staging is used to determine treatment: the main options include surgical excision, chemotherapy, targeted therapy and immunotherapy [1,2]. Unfortunately, the currently available therapeutic strategies for metastatic melanoma have a relatively low success rate, due to the development of drug resistance generally associated with changes in drug pharmacokinetics, mutation/amplification of drug targets and enhanced efflux pump-mediated drug detoxification [3-5]. Moreover, the majority of both consolidated and emerging anti-melanoma treatments is characterized by severe adverse effects [6,7]. For these reasons, in the last decade the interest in natural compounds has increased, owing to their potent and selective anti-cancer activity. In fact, a great number of studies has consistently reported that phytochemicals can exert anti-proliferative, pro-apoptotic, anti-invasive and anti-angiogenic effects in melanoma cell lines and mouse models, without significant toxicity in the latter. In particular, there are different molecular mechanisms responsible for the anti-melanoma actions of these compounds, such as inhibition of tumor-promoting proteins and activation of tumor-suppressing cascades [8]. This review attempts to summarize the recent findings about the role of various natural products in melanoma prevention and treatment.

## 2 NATURAL COMPOUNDS TARGETING MELANOMA atural compounds targeting melanoma

Accumulating evidence has highlighted the ability of numerous natural compounds to specifically target different signaling molecules and pathways involved in tumorigenesis and in tumor progression. Several of these naturally occurring molecules have been tested in *in vitro*, pre-clinical and clinical studies, alone or in combination with standard anti-cancer therapies [9]. Among them, polyphenols (flavonoids, curcumin and resveratrol), organosulfur compounds (sulforaphane), terpenoids (artemisinin, oridonin and ursolic acid), saponins (ginsenosides), tocotrienols ( $\gamma$ - and  $\delta$ -isoforms), alkaloids (berberine, harmine and capsaicin) and hydroxycinnamic acids (caffeic acid and its phenethyl ester) have shown promise as anti-melanoma agents (Fig. 1).

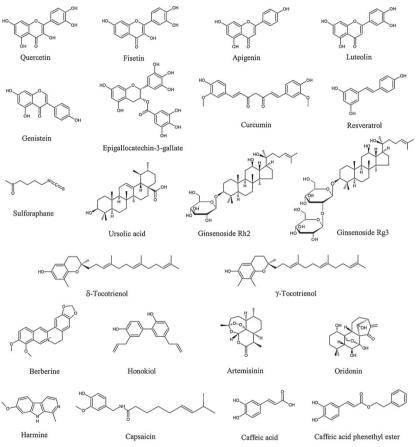


Fig. 1 Chemical structures of the main anti-melanoma natural compounds.

alt-text: Fig. 1

### 2.1 Quercetin

Quercetin is a penta-hydroxylated flavonol. It is particularly abundant in capers, tomatoes, apples, onions and tea, and because of this wide distribution in nature many studies have been conducted to investigate its chemopreventive and anti-tumor properties [10].

In different human melanoma cell lines quercetin was shown to reduce cell viability at low doses and to trigger apoptosis at high doses [11]. These anti-proliferative and pro-apoptotic effects were also confirmed *in vivo* [12,13], and were associated with: decrease in the B-cell lymphoma 2 (Bcl-2) expression [14]; nitric oxide (NO) production [15]; downregulation of protein kinase C-α (PKC-α) [16]; inhibition of the signal transducer and activator of transcription 3 (STAT3) pathway [17]. Interestingly, melanoma cells overexpressing tyrosinase, the rate-limiting enzyme in melanin synthesis, were more susceptible to the apoptosis induced by quercetin: in particular, the flavonol-mediated cell

death was associated with phosphorylation of p53, inhibition of glutathione reduction and reactive oxygen species (ROS) generation [18,19]. Finally, it has been recently demonstrated that this compound can severely alter the cellular bioenergetics in murine melanoma B164A5 cells, decreasing both the oxygen consumption and extracellular acidification rates [20].

The acquisition of invasive behavior is fundamental for the transformation of *in situ* melanoma into its most aggressive counterpart. Quercetin was found to suppress the metastatic potential of melanoma cells by inhibiting the matrix metalloproteinase 9 (MMP-9) activity [21], the hepatocyte growth factor (HGF)/c-MET signaling [22], the epithelial-to-mesenchymal transition (EMT) [23] and the interactions of the tumor cells with the endothelium [24].

The role of quercetin in UV protection and melanogenesis is still a matter of debate. Yin et al. have recently demonstrated that quercitrin, a glycosylated form of quercetin, can protect skin from UVB-induced oxidative damage [25]. On the other hand, Rafiq et al. reported that UVB-irradiated B16F10 melanoma cells subsequently treated with quercetin underwent a dose dependent reduction in cell viability and increased apoptotic cell death [26]. Moreover, quercetin exerted anti-melanogenic effects in UVA-exposed B16F10 cells through downregulation of NF-E2 p45-related factor 2 (Nrf2) activity [27] and reduced oxidative stress-induced and α-melanocyte stimulating hormone (α-MSH)-mediated melanogenesis in the same cell line [28–30], while enhancing melanin synthesis in HMVII melanoma cells [31].

It is now well established that calcitriol, the active form of vitamin D, is not only involved in the regulation of bone metabolism, but it also exerts significant antitumor effects as evidenced by *in vitro* and *in vivo* studies [32]. Specifically, melanoma cells express the vitamin D receptor (VDR), and vitamin D metabolites were shown to exert antiproliferative effects on melanoma cells [33-35]. In line with these observations, melanoma development has been shown to be correlated with vitamin D deficiency and dysfunctional VDR intracellular signaling pathways [36]. Quercetin was reported to enhance VDR activity in human colorectal adenocarcinoma cells (Caco-2) by altering cofactors recruitment [37] and upregulating VDR target genes [38]. Mechanistically, quercetin was shown to bind to the VDR receptor, as evidenced by *in silico* studies [39], thus triggering its antitumor effects. However, the possible interaction of quercetin with VDR is still a matter of debate [40]. So far, there are no studies reporting the possible interaction between quercetin and VDR signaling pathways in melanoma cells.

Synergistic anti-cancer activity was shown by quercetin when given in combination with either synthetic or natural compounds: the co-treatment with dacarbazine or temozolomide sensitizes melanoma cells to the anti-tumor effects of these chemotherapeutic agents through p53 activation [19,41]; the resistance of MeWo and WM164 cells to recombinant human tumor necrosis factor-related apoptosis-inducing ligand (rhTRAIL) was completely abrogated by addition of quercetin through upregulation of rhTRAIL-binding receptors DR4 and DR5 and increased degradation of the anti-apoptotic FLICE-like inhibitory protein (FLIP) [42]; a combination of quercetin and sulforaphane caused a significant additive effect in decreasing B16F10 cell proliferation and invasion *in vitro* and *in vivo* [43]; quercetin and curcumin synergistically inhibit melanoma cell viability by downregulation of Wnt/β-catenin signaling and apoptosis induction [44]; intravenous administration of quercetin and pterostilbene to mice suppressed the migration of melanoma cells to the liver [45].

Novel quercetin synthetic derivatives have been recently developed. In particular, Yamauchi et al. observed that methylquercetins can inhibit both the proliferation and migration of melanoma cells, also stimulating melanogenesis through modulation of microphthalmia-associated transcription factor (MITF) and p38 expression [46-48]. Quercetin glycosides were demonstrated to both induce [49,50] and suppress [51] melanogenesis.

Nanosized emulsions and lipid nanosystems containing quercetin exhibited important cytotoxic effects against B16F10 cells both *in vitro* and *in vivo*, with an increased solubility and oral bioavailability in mice with respect to the standard drug [52,53].

### 2.2 Fisetin

Fisetin is a flavonol, chemically referred to as 3,7,3',4'-tetrahydroxyflavone, which is commonly found in cucumbers, kiwi, onions, persimmons, apples and strawberries. It was observed to possess important neuroprotective effects, and recently its anti-cancer potential has also been investigated [54].

Different studies conducted by Syed et al. [55-57] demonstrated that fisetin can arrest melanoma cell proliferation through inhibition of the Wnt signaling pathway and direct binding to p70S6K and mTOR, and that it can activate both the intrinsic and extrinsic apoptotic pathways. Recent findings also suggest that fisetin can trigger mitochondrial apoptosis in uveal melanoma cells, while sparing normal retinal pigment epithelial cells [58], and that it can target Y-box binding protein 1 (YB-1)/ribosomal S6 kinase (RSK) axis in monolayer and 3D melanoma cultures [59].

Fisetin was reported to inhibit melanoma cell invasion through EMT reversion and suppression of mitogen-activated protein kinase (MAPK) and nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) signaling pathways [60].

 $\alpha$ -MSH- and 3-isobutyl-1-methylxanthine (IBMX)-induced melanosis in B16F10 melanoma cells was inhibited by fisetin treatment, which also activated the connective tissue growth factor (CTGF)/transforming growth factor  $\beta$  (TGF $\beta$ ) signaling pathway in human skin fibroblasts and 3T3-L1 cells, thus up-regulating skin fibril-related genes and decreasing adipogenesis, respectively [61]. On the contrary, 4'-O-methylfisetin significantly promoted melanogenesis in melanoma cells via activation of mammalian target of rapamycin complex 1 (mTORC1) [62].

In athymic nude mice subcutaneously implanted with BRAF-mutated melanoma cells, the combination therapy with fisetin and sorafenib, a small multi-kinase inhibitor that targets both the mutated and the wild-type BRAF

kinase, more effectively reduced the tumor growth when compared to the individual agents, via enhancement of apoptosis and inhibition of MAPK and phosphoinositide 3-kinase (PI3K) pathways [63]. Moreover, fisetin was shown to potentiate the anti-metastatic effects of sorafenib, leading to a decrease in N-cadherin, vimentin, fibronectin expression and to an increase in E-cadherin levels both *in vitro* and *in vitro* and *in vitro* [64].

The co-treatment with fisetin and melatonin also resulted in the synergistic activation of apoptosis, accompanied by suppression of cyclooxygenase-2 (COX-2), nitric oxide synthase (iNOS) and NF-κB signaling pathways [65].

## 2.3 Apigenin

Apigenin (4\*,5,7,-trihydroxyflavone) is a flavone present in many vegetables and fruits, particularly in chamomille, parsley, celery and oranges, and possesses radical-scavenging, anti-inflammatory and anti-carcinogenic properties [66].

The anti-tumor effects of apigenin have been investigated in different types of cancer, including melanoma. The main mechanisms of action are: G2/M cell cycle arrest and p-extracellular signal-regulated kinase 1/2 (p-ERK1/2), p-Akt and p-mTOR downregulation [67]; apoptosis activation, associated with ROS accumulation, cytochrome c release, DNA fragmentation, caspase-3 and poly (ADP-ribose) polymerase (PARP) cleavage [67,68]; suppression of cell migration through STAT3 and focal adhesion kinase (FAK) inhibition [69,70]; reduced vascular endothelial growth factor (VEGF) expression and secretion [71].

PD-1/PD-L1 checkpoint blockade-based immunotherapy has shown promising results in the treatment of melanoma. Interestingly, it has been recently demonstrated that apigenin downregulates the interferon gamma (IFN-γ)-induced PD-L1 expression in melanoma cells, by inhibiting STAT1 phosphorylation. Furthermore, apigenin enhanced the T cell-mediated melanoma killing *in vitro* and suppressed the melanoma xenograft growth by increasing CD4+ and CD8+T cell infiltration. Finally, apigenin boosted T cell immunity through downregulation of PD-L1 expression in dendritic cells [72].

Apigenin was found to overcome resistance to TRAIL and to increase TRAIL-mediated apoptosis in different cancer cell lines (such as breast cancer, colon cancer, hepatocellular carcinoma, pancreatic carcinoma cells), including melanoma cell lines [73].

Apigenin exhibited potent melanogenic activities by increasing the expression levels of MITF, tyrosinase, tyrosinase-related protein 1 (TRP-1) and TRP-2 and by activating the p38 pathway [74,75]. Similar results were obtained with apigenin-7-glucoside, which also suppressed melanoma cell proliferation [76].

Apigenin-loaded poly (lactic-co-glycolide) nanoparticles were reported to rapidly enter melanoma cancer cells, triggering mitochondrial apoptosis [77].

#### 2.4 Luteolin

Luteolin is a common flavone that exists in a variety of plants, such as celery, broccoli, parsley, thyme and rosemary, and a growing body of evidence has suggested that it possesses potent anti-inflammatory, neuroprotective and anti-tumor activity [78,79].

The anti-melanoma activities of luteolin were found to be correlated with anti-proliferative, anti-metastatic and anti-angiogenic effects, such as cyclin-dependent kinase 1 (CDK-1) and CDK-2 inhibition, PI3K/Akt pathway downregulation, reversion of  $\beta$ 3 integrin-mediated EMT and suppression of hypoxia-inducible factor 1-alpha (HIF1 $\alpha$ )/VEGF signaling [80-83].

The endoplasmic reticulum (ER) stress is a cellular process occurring in different physiological and pathological conditions, as well as after treatment with various synthetic and natural agents: the prolonged accumulation of unfolded and misfolded proteins in the ER lumen can activate a set of pro-death programs, such as the double-stranded RNA-dependent protein kinase PKR-like ER kinase (PERK)/eukaryotic initiation factor  $2\alpha$  (eIF2 $\alpha$ )/activating transcription factor 4 (ATF4)/C/EBP homologous protein (CHOP) pathway and the inositol-requiring enzyme  $1\alpha$  (IRE1 $\alpha$ )/c-Jun N-terminal kinase (JNK)/p38 MAPK cascade. Several phytochemicals were demonstrated to trigger ER stress-related death in tumor cells [84]: among them, luteolin specifically induced this pro-apoptotic pathway in melanoma cells [85].

Glutathione S-transferase (GST) is significantly involved in the metabolism and detoxification of drugs commonly used in melanoma therapy, thus affecting treatment effectiveness. Interestingly, Balyan et al. demonstrated that luteolin can selectively inhibit GST after tyrosinase-driven conversion in luteolin-quinone and luteolin-glutathione conjugate [86].

The anti-melanogenic effects of luteolin have been reported by various studies [87,88]. Improved melanogenesis inhibitory activity has been shown by different luteolin synthetic derivatives, including 3-prenyl-luteolin [89], luteolin 7-sulfate [90] and 7-O-substituted luteolin [91], as well as by the γ-irradiated compound [92].

### 2.5 Genistein

Genistein (4',5,7-trihydroxyisoflavone) is a phytoestrogen commonly found in soybeans that has been correlated with a decrease in the incidence of breast and prostate cancers [93,94].

Concerning melanoma, genistein treatment of different human and murine melanoma cell lines resulted in cell cycle arrest at G1/S and G2/M check points, accompanied by p21 upregulation, CDK1 and CDK2 inhibition and checkpoint kinase 2 (Chk2) activation [95–102]. Moreover, genistein was reported to promote morphological changes in melanoma cells, inducing a more dendritic and differentiated phenotype characterized by enhanced tyrosinase activity and melanin production [95–97].

It is well known that interleukin 8 (IL-8) synthesis can be stimulated by prostaglandin E2 (PGE2) in several pathologies, including cancer. Venza et al. demonstrated that PGE2 upregulates IL-8 expression in melanoma via the EP3 receptor and that genistein treatment can inactivate the latter, resulting in the reduction of IL-8 mRNA and protein levels and in the suppression of oral, uveal and cutaneous melanoma cell growth [103].

MicroRNAs (miRNAs) are endogenous, "22 nucleotide, non-coding RNAs implicated in RNA silencing and post-transcriptional control of gene expression. miRNAs may function as either oncogenes or tumor suppressors (oncomirs), depending on the specific cancer type. Sun et al. reported that genistein can inhibit human uveal melanoma cell growth via downregulation of miR-27a and of its target gene zinc finger and BTB domain containing 10 (ZBTB10) [104].

Genistein suppressed the invasive potential of melanoma cells via the FAK/paxillin pathway in vitro [105-107] and significantly decreased the number and size of liver and lung metastases in C57BL/6 mice [108-110].

Vasculogenic mimicry (VM) describes the ability of aggressive melanoma cells to form vascular-like structures in the absence of endothelial cells. Genistein not only exhibited anti-angiogenic properties in mouse models of melanoma [111] but also inhibited VM of uveal melanoma cells both *in vitro* and *in vivo* [112].

The effects of genistein on the immune system were evaluated by Guo et al. in adult female B6C3F1 mice injected with B16F10 cells. In particular, they observed that genistein could enhance host resistance to tumor formation, by increasing the activity of cytotoxic T and NK cells [113].

Synergistic anti-melanoma effects were shown by genistein when given in combination with standard chemotherapeutic agents, such as cyclophosphamide [114] and cisplatin [115]. Moreover, a recent study by Ji et al. has suggested that increasing the intracellular levels of ceramide can sensitize melanoma cells to the growth-suppressing activity of genistein [116].

5,7,4'-Trihydroxy-3'-methoxyisoflavone, obtained through biotransformation of genistein by two recombinant *Escherichia coli*strains, significantly reduced the proliferation of murine melanoma cells, without affecting the growth of normal murine fibroblasts [117].

In an interesting study conducted by Danciu et al., a lamellar lyotropic liquid crystal genistein-based formulation (LLC-Gen) was obtained in order to enhance the aqueous solubility of this compound. The formulation was applied locally, in a murine model of melanoma, with or without electroporation: the tumor volume, the amount of melanin and the degree of erythema were significantly reduced after 21 days of treatment, with an even better prognosis after electroporation [118].

### 2.6 Epigallocatechin-3-gallate

Epigallocatechin gallate (EGCG), the ester of epigallocatechin and gallic acid, belongs to the catechin subclass of flavonoids. It is the major component of green tea and is a potent free-radical scavenger and antioxidant. In addition, it is under study as a potential chemopreventive agent [119].

In melanoma cells, EGCG was found to exert both anti-proliferative and pro-apoptotic activities, inducing cyclin D1 and CDK2 downregulation, p16INK4a, p21CIP1/WAF1 and p27KIP1 activation, Bcl-2-associated X protein (Bax)/Bcl-2 ratio modulation and caspase-3, -7 and -9 cleavage [120]. Interestingly, these anti-cancer effects have been associated with inhibition of glucosidase II, a key enzyme involved in the glycoprotein synthesis in the ER, and with inflammasome downregulation, followed by reduced IL-1β secretion and NF-κB activity [121,122]. Furthermore, EGCG was demonstrated to act as an agonist of 67-kDa laminin receptor (67LR), a cell surface receptor highly expressed in melanoma cells, leading to mTOR pathway inhibition, merlin tumor suppressor activation and increased miRNA-let-7b expression [123,124].

EGCG treatment resulted in the suppression of melanoma cell migration and invasion, correlated with E-cadherin upregulation, HGF/SF-Met signaling dysregulation, MMP-2 inhibition and TNF receptor-associated factor 6 (TRAF6) inactivation [125-129].

In co-cultures of F10-OVA melanoma cells and tumor-specific CD3+T cells, EGCG reduced PD-L1 mRNA expression of 30% in the tumor cells and restored IL-2 mRNA expression in the lymphocytes, indicating that it can function as an immune checkpoint inhibitor [130].

EGCG was shown to possess anti-melanogenic activity, associated with decreased MITF production and tyrosinase expression [131,132].

Synergistic anti-melanoma effects of EGCG with different synthetic and natural anti-cancer agents have been observed: the co-treatment with dacarbazine significantly reduced the primary tumor growth and the number of

lung metastases in melanoma-bearing mice [133]; the addition of TRAIL to the EGCG treatment enhanced the apoptosis rate in human melanoma A375 cells [134]; EGCG sensitized melanoma cells to IFN-α-induced growth suppression [135]; with respect to monotherapy, the combination treatment with EGCG and vorinostat, a histone deacetylase (HDAC) inhibitor, resulted in significantly greater inhibition of cell proliferation and activation of apoptosis [136]; vitamin A increased the expression of the 67-kDa laminin receptor 67LR in B16 melanoma cells, potentiating the anti-proliferative activity of EGCG [137]. Moreover, EGCG was reported to overcome resistance to the BRAF inhibitor vemurafenib by activating 67LR-dependent protein phosphatase2A pathway in melanoma cells [138].

Novel EGCG synthetic derivatives have been recently developed: a 3,4,5-trimethoxybenzoyl ester analogue of EGCG was reported to bind to human dihydrofolate reductase and disrupt the folate cycle in melanoma cells, leading to cancer cell death [139]; although EGCG methylation generally reduces its anti-tumor properties, the anti-proliferative effects exerted by 7-OMe EGCGs on B16 cells were similar to those of EGCG [140]; 4-(S)-(2,4,6-trimethylthiobenzyl)-EGCG triggered apoptosis in melanoma cells via ROS-mediated autophagy induction [141].

Transferrin receptors are generally overexpressed in cancer cells and therefore can be exploited for the anti-tumor drug transport across cell membranes. Lemarié et al. reported that the intravenous administration of transferrin-bearing vesicles entrapping EGCG to mice bearing B16-F10 tumors successfully inhibited cancer progression [142].

Improved anti-melanoma efficacy was shown by EGCG when encapsulated in gold and chitosan nanoparticles [143,144], as well as in nanoethosomes [145], in in vitro and in vivo experiments.

### 2.7 Curcumin

Curcumin is a polyphenol obtained from *Curcuma longa*, commonly known as turmeric. It is nontoxic and characterized by many therapeutic properties, particularly by antioxidant, anti-inflammatory and anti-microbial activities [146,147].

As regards its anti-melanoma effects, curcumin was shown to arrest cell proliferation and to trigger both extrinsic and intrinsic apoptosis *in vitro* and *in vivo* [148-155]. In particular, it was reported to inhibit the NF-kB [156-158], STAT3 [159]. Akt/mTOR [160] and Wnt/8-catenin [44] signaling pathways, to induce ER stress [161], to activate mammalian Sterile 20-like kinase 1 (MST1)/INK cascade [162] and to enhance ROS production [163,164] in melanoma cells.

Curcumin exhibited anti-invasive properties, correlated with modulation of FAK and MMP-2 activity and of integrin receptor, nonmetastatic gene 23 (Nm23) and E-cadherin expression [165,166]. Moreover, curcumin selectively downregulated the levels of phosphatase of regenerating liver-3 (PRL-3), an oncogene involved in tumor metastasis, in B16F10 cells [167]. Interestingly, pulmonary administration of this compound resulted in a significant decrease in the number of lung metastatic nodules in melanoma mouse models [168].

Different curcumin-based combination therapies have been proposed for the treatment of melanoma: C6 ceramide and homoserine based C8-ceramide analogues promoted curcumin-mediated anti-proliferative and anti-angiogenic effects [169,170]; the co-treatment with tamoxifen and curcumin resulted in synergistic induction of apoptosis in G361 chemo-resistant cells [171]; fibroblast activation protein α (FAPα) vaccine combined with curcumin elicited the anti-cancer response to the natural compound via inhibition of indolamine-2,3-dioxygenase and of EMT [172]; 1-phenyl-2-decanoylamino-3-morpholino-1-propanol (PDMP) facilitated curcumin-induced apoptotic cell death by enhancing ceramide accumulation and c-Jun N-terminal kinase (JNK) phosphorylation whilst suppressing Akt/mTOR signaling pathway [173]; ABT-737, a novel Bcl-2 inhibitor, potentiated the pro-apoptotic activity of curcumin in WM-115 and B16 cell lines [174]; a combination of curcumin and NC10, a synthetic nitrosyl-iron complex, improved the anti-tumor efficacy of the monotherapy with the polyphenol [175]; borneol, a terpene derivative commonly used in Chinese medicine, effectively synergized with curcumin, by activating the caspase cascade in A375 cells [176]; the red united blue light irradiation greatly enhanced the oxidative stress-mediated cell death induced by curcumin [177].

Many recent studies have focused on the synthesis of anti-melanoma curcumin derivatives: the curcumin analog DM-1 triggered apoptotic cell death, alone or in combination with dacarbazine, both *in vitro* and *in vivo* [178-181]; the analog D6 significantly upregulated the p53 expression and inhibited the PI3K/Akt and NF-xB pathways, inducing mitochondrial apoptosis in different melanoma cell lines [182-184]; treatment with the FLLL32 and FLLL62 analogues resulted in STAT3 inhibition, ultimately leading to melanoma cell death [185,186]; the curcuminoid EF24 exhibited potent anti-proliferative, anti-metastatic and anti-angiogenic effects, also modulating miRNA-21 and miRNA-33b expression [187-189]; ferrocenyl curcuminoid derivatives showed improved anti-melanoma activity compared to the corresponding organic compounds [190]; different curcumin-like diarylpentanoid analogues were demonstrated to suppress the melanogenesis in B16 cells [191]; tetrahydricyrcumin, salicyl curcumin and curcumin-III greatly reduced the neo-vascularization in mouse melanoma models [192].

Micelles and liposomes are vesicles in which drugs can be trapped and administered more efficiently. Different curcumin-loaded micellar systems (chitosan-cholesterol-, poly(D,L-lactide)-, cholesterol-conjugated poly(D,L-lactide)-, PEG-, cholesterol- and vitamin E-conjugated PEG-based micelles) exhibited higher cytotoxicity with respect to the free compound in 2D and 3D melanoma cell cultures and in melanoma-bearing mice, successfully solubilizing and stabilizing the drug and promoting its controlled release to the tumor cells [193–199]; liposomal delivery of curcumin, alone or in combination with anti-cancer agents and molecules such as paclitaxel or STAT3 siRNA, resulted in growth-suppressing and anti-angiogenic effects in *in vitro* and *in vivo* melanoma models [200–204].

Nanoparticles and cyclodextrin-based carriers can also be used to encapsulate curcumin and its analogues, and have been recently proposed as promising new formulations for melanoma treatment [205-223].

An anti-Muc18 antibody-coupled curcumin more efficiently reduced the brain metastasis formation in mice inoculated with B16F10 cells [224].

#### 2.8 Resveratrol

Resveratrol (trans-3,4',5-trihydroxystilbene) is a grape-derived polyphenol that has been intensively studied for its chemopreventive potential [225].

In the case of melanoma, resveratrol was found to inhibit cell growth by inducing S phase arrest, with cyclins A, E, and B1 upregulation, dihydronicotinamide riboside quinone reductase 2 (NQO2)-mediated p53 overexpression, ERK1/2 pathway suppression, NF-κB inactivation and oncogenic miR-221 downregulation [226-230]. Moreover, it was shown to trigger mitochondrial apoptosis via Bax/Bcl-2 ratio modulation, caspase-3 cleavage, STAT3/β-catenin and survivin suppression, ROS production and ER stress activation [231-239]. Finally, it was observed to induce phenotype changes and to affect melanin synthesis in melanoma cells [240-243].

In addition to the above mentioned anti-cancer effects, resveratrol exhibited important anti-angiogenic and anti-invasive activities, associated with ανβ3 integrin inhibition, VEGF downregulation, increased thrombospondin 1 (TSP1) expression and Akt and FAK pathways inactivation [244–248]. Notably, in mice injected with B16 M cells resveratrol successfully prevented liver metastases by reducing IL-18-dependent expression of vascular cell adhesion protein 1 (VCAM-1) in tumor-activated hepatic sinusoidal endothelium, thus inhibiting melanoma cell adhesion to the microvasculature [249,250].

Cellular senescence is a tumor-suppressive mechanism generally associated with DNA damaging cancer therapies. In particular, it can be followed by deleterious effects in the tumor microenvironment, such as the acquisition of a senescence-associated secretory phenotype (SASP) which is responsible for the conversion of fibroblasts into pro-inflammatory cells capable of promoting cancer progression. In a recent study by Menicacci et al., chronic resveratrol treatment significantly inhibited MRC5 fibroblast SASP-related pro-tumor effects on melanoma cells, reducing the expression of EMT markers correlated with malignant features [251].

It is well documented that NO can participate to melanoma progression. Yang et al. demonstrated that NO can enhance melanoma metastatic potential via an apurinic/apyrimidinic endonuclease-1 (APE)/redox factor-1 (Ref-1)-driven feedback loop, which is suppressed by resveratrol [252].

Resveratrol was shown to potentiate the activity of different anti-melanoma agents: it sensitized melanoma cells to TRAIL pro-apoptotic effects [253,254]; the natural compound-mediated APE/Ref-1 inhibition increased the dacarbazine-induced cell death [255]; resveratrol addition rendered melanoma cells more sensitive to temozolomide treatment [255]; capsaicin and resveratrol synergistically triggered apoptosis through NO elevation in A375 cells [256]; melanoma sensitivity to cisplatin was enhanced by resveratrol via connexin 43 upregulation [257]; vemurafenib resistance was reversed by resveratrol treatment via Akt inactivation in BRAF-mutated melanoma cells [258]; synergistic anti-proliferative effects, correlated with AMP-activated protein kinase (AMPK), vasodilator-stimulated phosphoprotein (VASP) and VEGF modulation, were shown by a combination of resveratrol and 5-fluorouracil [259,260]; chloroquine synergized with the polyphenol to induce cytotoxicity in melanoma cells [261]; resveratrol improved the efficacy of high-dose IL-2 immunotherapy in B16F10 melanoma mouse models, also preventing the endothelial cell injury and inhibiting the development vascular leak syndrome [262]; resveratrol was demonstrated to act as radiotherapy sensitizer in radioresistant melanoma cell lines [263].

In the last decade, several resveratrol analogues have been produced in order to improve the pharmacokinetic properties and to increase the pharmacological potency of this compound for melanoma treatment [264-274].

## 2.9 Sulforaphane

Sulforaphane is an organic isothiocyanate with several health benefits. It is obtained from cruciferous plants, such as broccoli, cabbage and cauliflower [275].

In melanoma setting, sulforaphane was shown to trigger cell growth arrest and apoptosis, accompanied by the upregulation of early growth response protein 1 (EGR1), growth arrest and DNA-damage-inducible beta (GADD45B), ATF3 and CDKN1A, by the activation of caspase-3 and -9, Bax, p53, p53 upregulated modulator of apoptosis (PUMA), Fas and mouse double minute 2 homolog (MDM2) and by the downregulation of Bcl-2, BH3 interacting domain death agonist (Bid) and NF-κB [276-279]. Furthermore, it induced oxidative stress and modulated the expression of nerve growth factor receptors TrKA and p75NTR, shifting their ratio from pro-survival to pro-apoptotic [278-280].

Sulforaphane reduced the invasive potential of B16F10 melanoma cells by inhibiting MMPs activity, thereby suppressing lung metastases [281]. Moreover, it inhibited the spread of B16F10 cells through the stimulation of cell-mediated immune response, upregulation of IL-2 and IFN- $\gamma$  and downregulation of proinflammatory cytokines IL-1 $\beta$ , IL-6, TNF- $\alpha$  and granulocyte-macrophage colony-stimulating factor (GM-CSF) [282].

Cancer stem cells (CSCs) are widely recognized as a small subpopulation of cancer cells within the tumor mass responsible for the resistance to standard anti-cancer therapies. In particular, they possess self-renewal ability, as well as the capacity to give rise to the entire tumor cell bulk through differentiation, thus representing important therapeutic targets. Several phytochemicals were demonstrated to target CSCs, and sulforaphane was found to specifically eliminate the melanoma CSC subpopulation, by suppressing enhancer of zeste homolog 2 (Ezh2) function [283].

Sulforaphane-encapsulated microspheres exhibited potent anti-melanoma activity both in vitro and in vivo [284,285].

Tahata et al. investigated sulforaphane efficacy and safety in 17 patients with at least 2 atypical nevi and a prior history of melanoma. The patients were given three different oral doses of the compound (50, 100 or 200 µmol) daily for 28 days. Sulforaphane was well tolerated even at the higher dose and achieved dose-dependent levels in plasma and skin. Importantly, plasma levels of proinflammatory cytokines decreased, while the expression of the tumor suppressor decorin increased from day 1 to 28 [286].

#### 2.10 Ursolic acid

Ursolic acid is a triterpenoid exhibiting a wide spectrum of pharmacological properties, including anti-inflammatory and anti-microbial features. It is present in a variety of plants and herbs, such as thyme and rosemary, as well as in fruit peels [287].

Many studies have pointed out that ursolic acid possesses potent anti-melanoma activity, correlated with modulation of different pathways, including NF-κB, p53, Akt and ERK1/2 proteins, and with caspase cascade activation [288-292]. Notably, it has also been demonstrated that tyrosinase- and TRP-1-mediated melanogenesis and COX-2/PGE2 pathway are implicated in the resistance of melanoma cells to the ursolic acid cytotoxicity [293,294].

In addition to the above forenamed anti-melanoma effects, ursolic acid was observed to reduce the levels of VEGF, NO and proinflammatory cytokines in the serum of melanoma mouse models [295]. Furthermore, it significantly suppressed lung metastasis formation [296].

It should be noted that ursolic acid sensitized melanoma cells both to UV irradiation [297] and radiotherapy [298].

Novel derivatives and nanoformulations have been recently developed [299-302]; interestingly, the administration of low molecular weight heparin-conjugated ursolic acid and of inclusion complexes formed by the acid and cyclodextrins to melanoma-bearing mice resulted in a significant tumor growth inhibition [303-305].

### 2.11 Ginsenosides

Ginsenosides are triterpene saponins and are the major pharmacologically active components of ginseng root [306].

In melanoma cells, ginsenoside Rh2 was found to exert a G1 phase-specific suppressive effect on the Cdk2 activity and to induce caspase-3 and -8 dependent apoptosis [307,308]. Similar anti-proliferative and pro-apoptotic effects were also shown by other members of the ginsenoside group: ginsenoside Rg3 decreased HDAC3 expression, increased p53 acetylation, downregulated NF-κB-mediated fucosyltransferase 4 (FUT4) expression and inactivated epidermal growth factor receptor (EGFR)/MAPK pathway, leading to melanoma cell death both *in vitro* and *in vivo* [309-312]; upregulation of Fas, FasL and Bax protein expression and downregulation of procaspase-8, procaspase-3, mutant p53 and Bcl-2 protein expression were observed in SK-MEL-2 human melanoma cells after ginsenoside Rk1 treatment [313]; a major metabolite of the red ginseng ginsenoside Rb1, called 20-O-β-D-glucopyranosyl-20(S)-protopanaxadiol, suppressed melanoma cell growth by inducing autophagy and apoptosis via AMPK/JNK signaling activation [314].

Ginsenosides have been recently proposed as skin-whitening agents, owing to their ability to affect melanin synthesis via tyrosinase inhibition [315-318]. Notably, Kim et al. have also suggested that ginsenoside F1 may attenuate B16F10 melanoma cell hyperpigmentation via Ras homologous (Rho) signaling activation and subsequent dendrite retraction [319]. However, the role of ginsenosides in altering melanoma cell morphology is still unclear, since a recent study by Jiang et al. has demonstrated that ginsenoside Rh2 can induce dendrite formation by changing the physical properties of cholesterol-regulated membrane lipid bilayers [320].

Promising anti-angiogenic and anti-metastatic potential in melanoma treatment was shown by ginsenosides Rb2, Rg3 and Rp1 [321-325].

Ginsenoside Rh2 treatment enhanced the anti-cancer immunological response in melanoma mouse models, by increasing T-lymphocyte infiltration in the tumor and by triggering cytotoxicity in spleen lymphocytes [326].

Similarly, ginsenoside Rg3 induced immunogenic cell death in B16F10 melanoma cells, as evidenced by upregulated surface expression of calreticulin and heat shock proteins; moreover, the proportion of dendritic CRT<sup>+</sup> CD11c<sup>+</sup> cells was increased in the Rg3-treated group, which also secreted IFN-γ, an effector molecule for anti-tumor activity in T cells [327].

Oral administration of ginsenoside Rh2 to C57BL/6 mice bearing B16 melanoma synergistically enhanced the anti-tumor activity of cyclophosphamide in a dose-dependent manner. Furthermore, it decreased the micronucleus formation in polychromatic erythrocytes and DNA strand breaks in white blood cells, suggesting that it not only increases the anti-melanoma efficacy of cyclophosphamide but also reduces its genotoxic effects [328].

New synthetic ginsenoside derivatives have been recently obtained, showing considerable cytotoxicity against melanoma cells [329]. Interestingly, Cui et al. cloned and characterized a novel ginsenoside-transforming β-glucosidase (BglG167b) derived from *Microbacterium* sp. Gsoil 167 that can efficiently hydrolyze gypenoside XVII, a deglycosylated product of major ginsenoside Rb1, into gypenoside LXXV, which displayed an enhanced anti-melanoma effect compared to the original compound [330].

In a study by Zare-Zardini et al., novel drug delivery systems based on ginsenoside Rh2-treated highly porous graphene were produced in order to improve the compound cytotoxic effects on different cancer cell lines, including

melanoma A375 cells [331].

#### 2.12 Tocotrienols

Tocotrienols (TTs) are hydrophobic compounds which belong to the vitamin E family. They are composed of a chromanol ring linked to an unsaturated isoprenoid side chain; the number and the position of methyl substitutes on the chromanol ring identify  $\alpha$ -,  $\beta$ -,  $\gamma$ - and  $\delta$ -TT isoforms. Natural sources containing high levels of tocotrienols are annatto (*Bixa orellana* L.) seeds, red palm oil, rice bran and other vegetable oils. In several studies TTs were reported to exert health promoting effects based on their neuroprotective, cardioprotective, antioxidant, anti-inflammatory and cholesterol-lowering properties [332]. In addition, anti-proliferative, pro-apoptotic and anti-metastatic activities have been demonstrated in a wide range of cancer cells *in vitro* and *in vitro* [333–337].

We demonstrated that δ-TT exerts pro-apoptotic effects in human melanoma A375 and BLM cell lines, by triggering the activation of the PERK/p-eIF2α/ATF4/CHOP, IRE1α and caspase-4 ER stress-related pathways [338,339]. We also showed that, unlike vemurafenib, δ-TT specifically targets the ABCG2-positive CSCs subpopulation in the A375 cell line, preventing the formation of melanospheres and inducing their disaggregation [340].

In malignant melanoma cells, γ-TT-induced cell death was found to be associated with the cleavage of procaspases, the activation of JNK signaling pathway and the suppression of NF-κB, EGFR and inhibitor of DNA-binding (Id) family proteins. Moreover, γ-TT significantly suppressed melanoma cell invasion capability, by downregulating mesenchymal markers and restoring E- and γ-cadherin expression [341].

A synergistic tumor-suppressing activity was observed by combining TTs with standard anti-cancer agents or other natural compounds. In particular, TTs were reported to potentiate lovastatin-mediated growth inhibition of murine melanoma B16 cells, by dysregulating HMG-CoA reductase activity both *in vitro* and *in vivo* [342]. Moreover, in the same cell line γ-TT was found to upregulate the aryl hydrocarbon receptor (AhR) expression, enhancing the anti-proliferative activity of baicalein, a flavone isolated from the roots of *Scutellaria baicalensis* and *Scutellaria lateriflor* [343].

Despite the many anti-cancer properties, TTs fail to reach the tumor mass after intravenous administration, and melanoma-targeted vesicles bearing transferrin have been proposed to overcome bioavailability limitations [344].

#### 2.13 Berberine

Berberine is a benzylisoquinoline alkaloid extracted from various plants, particularly from those belonging to the genus *Berberis*. Ongoing experimental and clinical studies have pointed out great potential of this compound in the regulation of glucose and lipid homeostasis, inflammation and cancer growth [345].

Serafim et al. demonstrated that in melanoma cells berberine is concentrated in mitochondria at low doses, promoting G1 phase arrest, while it accumulates in the cytoplasm and in the nucleus at higher doses, inducing G2 arrest [346]. Importantly, berberine was also shown to trigger apoptosis [347] and necrosis [348] and to inhibit melanin synthesis [349] in different melanoma cell lines.

Melanoma cell migration was significantly affected by berberine treatment through modulation of PI3K/Akt, FAK and NF-kB pathways *in vitro* [350,351]. Furthermore, the alkaloid successfully enhanced the survival of melanoma-bearing mice, by reducing pulmonary metastases via MMP downregulation and AMPK-mediated suppression of ERK activity and COX-2 expression [352,353].

Berberine exhibited antiangiogenic activity associated with the inhibition of HIF, VEGF, NO and proinflammatory cytokines [354].

Finally, it should be underlined that increased anti-melanoma effects were shown by berberine when given in combination with doxorubicin [355].

### 2.14 Other compounds

Honokiol is a lignan occurring in several species of the genus *Magnolia*. It exerted cytostatic and cytotoxic effects in melanoma cells, by reducing cyclin D1, CDK1 and CDK2 levels, attenuating Akt/mTOR signaling, suppressing HIF-1 pathway, disrupting mitochondrial electron transport chain (ETC) function and inducing caspase/PARP cleavage [356-360]. In addition, it activated ER stress via direct binding to GRP78 ATPase domain and Calpain-10 and CHOP/GADD153 cascade [361,362]. Furthermore, it affected the migration ability of Hs294t and SK-Mel28 cells through inhibition of NADPH oxidase 1 (Nox1) and blockade of the interactions between the enzyme subunits p22 and p47 [363]. It also specifically targeted and eliminated melanoma CSCs via AMPK activation and Notch-2 downregulation [364,365], while its bis-dichloroacetate derivative demonstrated enhanced activity in vemurafenib-resistant melanoma *in vivo* [366].

Artemisinin and its derivatives, such as artenusate, artemison and dihydoartemisinin, are well known antimalarial drugs. Interestingly, in melanoma cell lines they were shown to exert potent anti-proliferative and pro-apoptotic effects, associated with Wnt/β-catenin signaling inhibition and oxidative stress induction, and to suppress cell migration through downregulation of MMP-2 and ανβ3 integrin expression [367–372].

Oridonin, a diterpenoid isolated from Rabdosia rubescens, was found to induce G2/M phase arrest and differentiation of melanoma cells, and to activate p53 and ERK pathways, to suppress IGF-1R signaling and fatty acid

synthase (FAS) activity, to modulate Bax/Bcl-xL ratio and Bim expression and to promote cytochrome c release from mitochondria, ultimately triggering apoptosis [373-377]. Moreover, it was reported to affect melanoma cell invasive potential and TGF-B1-mediated EMT via PI3K/Akt downregulation [378].

Harmine is a β-carboline alkaloid from the plant *Peganum harmala*. Hamsa et al. reported that in B16F10 cells it can activate both intrinsic and extrinsic apoptosis, suppress angiogenesis through modulation of VEGF, NO and pro-inflammatory cytokines, and inhibit invasion via MMP-9 and ERK downregulation [379–381]. Notably, harmine analogues have also shown promising anti-melanoma effects both *in vitro* and *in vivo* [382,383].

Capsaicin is an alkaloid derived from chili peppers of the genus *Capsicum*. Morré et al. correlated its anti-melanoma activity with the inhibition of cell surface NADH oxidase [384]. More recently, in melanoma cells capsaicin was demonstrated to inhibit NF-κB-driven proliferation [385], to trigger Bcl-2 dependent apoptosis [386] and to suppress PI3K/Akt signaling-mediated migration [387].

Caffeic acid and its phenetyl ester (CAPE) are bioactive compounds from the propolis extract. They were observed to reduce p-Akt, p-mTOR and X-linked inhibitor of apoptosis protein (XIAP) levels [388] and to trigger ROS formation, GSH depletion and mitochondrial apoptosis [389-392], as well as to inhibit VEGFR-2-driven neovascularization [393] and tyrosinase dependent melanin synthesis [394,395] in melanoma cells and mouse models.

The molecular mechanisms underlying the anti-melanoma effects of the natural compounds here discussed are summarized in Fig. 2.

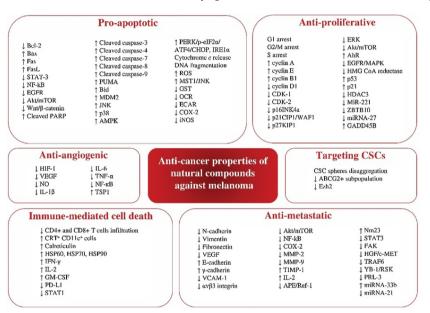


Fig. 2 Molecular mechanisms underlying the anti-melanoma effects of natural compounds. Phytochemicals modulate several key pathways involved in apoptosis, proliferation, metastasis, angiogenesis, cancer stemness and immune response.

alt-text: Fig. 2

## 3 CONCLUDING REMARKS oncluding remarks

The present article gives an overview of recent evidence about the anti-melanoma effects of various natural products.

Nutraceuticals present numerous advantages since they are usually nontoxic and their cost is highly affordable around the world. It is now well established that they can exert anti-melanoma properties, and many in vitro and pre-clinical studies have been conducted to clarify the molecular mechanisms underlying these activities. Moreover, recent studies focused on the development of new synthetic derivatives, formulations and combinations with standard drugs, have clearly suggested that these compounds hold promise for melanoma prevention and treatment. However, the relevance of these findings still needs to be confirmed in patients, and clinical trials aimed at validating nutraceuticals effectiveness are urgently required.

# **Declaration of Competing Interest**

The authors declare that there are no conflicts of interest.

## **ACKNOWLEDGEMENTS**cknowledgements

P. Limonta's laboratory is supported by MIUR (Departments of Excellence Italian Law n. 232, 11 thth December 2016). We thank Comitato Emme Rouge Onlus for providing financial support for our research.

### References

- [1] G.C. Leonardi, L. Falzone, R. Salemi, A. Zanghì, D.A. Spandidos, J.A. Mccubrey, S. Candido and M. Libra, Cutaneous melanoma: From pathogenesis to therapy (Review), *Int. J. Oncol.* 52, 2018, 1071-1080, https://doi.org/10.3892/ijo.2018.4287.
- [2] B. Domingues, J. Lopes, P. Soares and H. Populo, Melanoma treatment in review, Immuno Targets Ther. 7, 2018, 35-49, https://doi.org/10.2147/itt.s134842.
- [3] B.S. Kalal, D. Upadhya and V.R. Pai, Chemotherapy resistance mechanisms in advanced skin cancer, Oncol. Rev. 11, 2017, 326, https://doi.org/10.4081/oncol.2017.326.
- [4] M. Winder and A. Virós, Mechanisms of Drug Resistance in Mdrug resistance in melanoma, Handb. Exp. Pharmacol. 249, 2018, 91-108, https://doi.org/10.1007/164 2017 17.
- [5] I. Kozar, C. Margue, S. Rothengatter, C. Haan and S. Kreis, Many ways to resistance: How melanoma cells evade targeted therapies Biochim. Biophys. Acta. Rev. Cancer.how melanoma cells evade targeted therapies, Biochim. Biophys. Acta. Rev. Cancer.how melanoma cells evade targeted therapies, Biochim. Biophys. Acta. Rev. Cancer.how melanoma cells evade targeted therapies, Biochim. Biophys. Acta. Rev. Cancer.how melanoma cells evade targeted therapies, Biochim. Biophys. Acta. Rev. Cancer.how melanoma cells evade targeted therapies.
- [6] C. Luther, U. Swami, J. Zhang, M. Milhem and Y. Zakharia, Advanced stage melanoma therapies: Detailing the present and exploring the future, Crit. Rev. Oncol. Hematol. 133, 2019, 99-111, https://doi.org/10.1016/j.critrevonc.2018.11.002.
- [7] A. Boada, C. Carrera, S. Segura, H. Collgros, P. Pasquali, D. Bodet, S. Puig and J. Malvehy, Cutaneous toxicities of new treatments for melanoma, Clin. Transl. Oncol. 20, 2018, 1373–1384, https://doi.org/10.1007/s12094-018-1891-7.
- [8] T.N. Chinembiri, L.H. du Plessis, M. Gerber, J.H. Hamman and J. du Plessis, Review of natural compounds for potential skin cancer treatment, *Molecules.* 19, 2014, 11679-11721, https://doi.org/10.3390/molecules190811679.
- [9] M.K. Pandey, S.C. Gupta, A. Nabavizadeh and B.B. Aggarwal, Regulation of cell signaling pathways by dietary agents for cancer prevention and treatment, Semin. Cancer Biol. 46, 2017, 158-181, https://doi.org/10.1016/j.semcancer.2017.07.002.
- [10] A. Rauf, M. Imran, I.A. Khan, M.- Ur-Rehman, S.A. Gilani, Z. Mehmood and M.S. Mubarak, Anticancer potential of quercetin: Aa comprehensive review, *Phyther. Res.* 32, 2018, 2109–2130, https://doi.org/10.1002/ptr.6155.
- [11] K. Rpsner, C. Röpke, V. Pless and G.L. Skovgaard, Late Type Apoptosis and Apoptosis Free Lethal Effect of Quercetin in Human Mtype apoptosis and apoptosis free lethal effect of quercetin in human melanoma, Biosci. Biotechnol. Biochem. 70, 2006, 2169-2177, https://doi.org/10.1271/bbb.60129.
- [12] S. Caltagirone, C. Rossi, A. Poggi, F.O. Ranelletti, P.G. Natali, M. Brunetti, F.B. Aiello and M. Piantelli, Flavonoids apigenin and quercetin inhibit melanoma growth and metastatic potential, *Int. J. Cancer* 87, 2000, 595-600, https://doi.org/10.1002/1097-0215(20000815)87:4<595::AID-IJC21>3.0.CO;2-5.
- [13] R. Kale, M. Saraf, A. Juvekar and P. Tayade, Decreased B16F10 melanoma growth and impaired tumour vascularization in BDF1 mice with quercetin-cyclodextrin binary system, *J. Pharm. Pharmacol.* 58, 2006, 1351-1358.
- [14] X. Zhang, Q. Xu and I. Saiki, Quercetin inhibits the invasion and mobility of murine melanoma B16-BL6 cells through inducing apoptosis via decreasing Bcl-2 expression, Clin. Exp. Metastasis 18, 2000, 415-421, https://doi.org/10.1023/A:1010960615370.
- [15] P. Ferrer, M. Asensi, S. Priego, M. Benlloch, S. Mena, A. Ortega, E. Obrador, J.M. Esteve and J.M. Estrela, Nitric oxide mediates natural polyphenol-induced Bcl-2 down-regulation and activation of cell death in metastatic B16 melanoma, J. Biell. Biol. Chem. 282, 2007, 2880-2890, https://doi.org/10.1074/jbc.M605934200.
- [16] X.M. Zhang, J. Chen, Y.G. Xia and Q. Xu, Apoptosis of murine melanoma B16-BL6 cells induced by quercetin targeting mitochondria, inhibiting expression of PKC-a and translocating PKC-6, Cancer Chemother.

- Pharmacol. 55, 2005, 251-262, https://doi.org/10.1007/s00280-004-0863-5.
- [17] H.H. Cao, A.K.W. Tse, H.Y. Kwan, H. Yu, C.Y. Cheng, T. Su, W.F. Fong and Z.L. Yu, Quercetin exerts anti-melanoma activities and inhibits STAT3 signaling, *Biochem. Pharmacol.* 87, 2014, 424-434, https://doi.org/10.1016/j.bcp.2013.11.008.
- [18] T. Thangasamy, S. Sittadjody, S. Lanza-Jacoby, P.R. Wachsberger, K.H. Limesand and R. Burd, Quercetin selectively inhibits bioreduction and enhances apoptosis in melanoma cells that overexpress tyrosinase, *Nutr. Cancer* 59, 2007, 258-268, https://doi.org/10.1080/01635580701499545.
- [19] T. Thangasamy, S. Sittadjody, K.H. Limesand and R. Burd, Tyrosinase overexpression promotes ATM-dependent p53 phosphorylation by quercetin and sensitizes melanoma cells to dacarbazine, *Cell. Oncol.* 30, 2008, 371–387. https://doi.org/10.3233/CLO-2008-0441.
- [20] A. Sturza, I. Pavel, S. Ancusa, C. Danciu, C. Dehelean, O. Duicu and D. Muntean, Quercetin exerts an inhibitory effect on cellular bioenergetics of the B164A5 murine melanoma cell line, *Mol. Cell. Biochem.* 447, 2018, 103–109, https://doi.org/10.1007/s11010-018-3296-x.
- [21] X.-M. Zhang, S.-P. Huang and Q. Xu, Quercetin inhibits the invasion of murine melanoma B16-BL6 cells by decreasing pro-MMP-9 via the PKC pathway, *Cancer Chemother. Pharmacol.* 53, 2004, 82-88, https://doi.org/10.1007/s00280-003-0702-0.
- [22] H.-H. Cao, C.-Y. Cheng, T. Su, X.-Q. Fu, H. Guo, T. Li, A.K.-W. Tse, H.-Y. Kwan, H. Yu and Z.-L. Yu, Quercetin inhibits HGF/c-Met signaling and HGF-stimulated melanoma cell migration and invasion, Mol. Cancer 14, 2015, 103, https://doi.org/10.1186/s12943-015-0367-4.
- [23] D. Patel and N. Sharma, Inhibitory effect of quercetin on epithelial to mesenchymal transition in SK-MEL-28 human melanoma cells defined by in vitro analysis on 3D collagen gels, *Onco. Targets*, *Ther.* 9, 2016, 6445-6459, https://doi.org/10.2147/OTT.S109253.
- [24] M. Piantelli, C. Rossi, M. Iezzi, R. La Sorda, S. Iacobelli, S. Alberti and P.G. Natali, Flavonoids inhibit melanoma lung metastasis by impairing tumor cells endothelium interactions, *J. Cell. Physiol.* 207, 2006, 23-29, https://doi.org/10.1002/jcp.20510.
- [25] Y. Yin, W. Li, Y.O. Son, L. Sun, J. Lu, D. Kim, X. Wang, H. Yao, L. Wang, P. Pratheeshkumar, A.J. Hitron, J. Luo, N. Gao, X. Shi and Z. Zhang, Quercitrin protects skin from UVB-induced oxidative damage, *Toxicol. Appl. Pharmacol.* 269, 2013, 89–99, https://doi.org/10.1016/j.taap.2013.03.015.
- [26] R.A. Rafiq, A. Quadri, L.A. Nazir, K. Peerzada, B.A. Ganai and S.A. Tasduq, A potent inhibitor of phosphoinositide 3-kinase (PI3K) and mitogen activated protein (MAP) kinase signalling, Quercetin (3, 3', 4', 5, 7-pentahydroxyflavone) promotes cell death in Ultraviolet (UV)-B-irradiated B16F10 melanoma cells. *PLoS One* 10, 2015, e0131253, https://doi.org/10.1371/journal.pone.0131253.
- [27] A. Chaiprasongsuk, T. Onkoksoong, T. Pluemsamran, S. Limsaengurai and U. Panich, Photoprotection by dietary phenolics against melanogenesis induced by UVA through Nrf2-dependent antioxidant responses, *Redox Biol.* 8, 2016, 79-90, https://doi.org/10.1016/j.redox.2015.12.006.
- [28] Y.I. Kim, Hyperin and Quercetin Modulate Oxidative Stress-Induced Melanogenesis, Biol. Pharm. Bull. 35, 2012, 2023-2027, https://doi.org/10.1248/bpb.b12-00592.
- [29] Y.M. Yang, Y.O. Son, S.A. Lee, Y.M. Jeon and J.C. Lee, Quercetin inhibits \alpha-MSH-stimulated melanogenesis in B16F10 melanoma cells, Phyther. Res. 25, 2011, 1166-1173, https://doi.org/10.1002/ptr.3417.
- [30] T. Fujii and M. Saito, Inhibitory Effect of Quercetin Isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanogenesis by Mouse Melanoma Ceffect of quercetin isolated from Rose Hip (Rosa canina L.) against Melanoma Ceffect of
- [31] H. Nagata, S. Takekoshi, R. Takeyama, T. Homma and R.Y. Osamura, Quercetin Enhances Melanogenesis by Increasing the Activity and Synthesis of Tyrosinase in Human Melanoma Cells and in Normal Human Melanocytes Pigmentenhances melanogenesis by increasing the activity and synthesis of tyrosinase in human melanoma cells and in normal human melanocytes, Pigm. Cell Res. 17, 2004, 66-73, https://doi.org/10.1046/j.1600-0749.2003.00113.x.
- [32] S.J. Weinstein, M.P. Purdue, S.A. Smith-Warner, A.M. Mondul, A. Black, J. Ahn, W.Y. Huang, R.L. Horst, W. Kopp, H. Rager, R.G. Ziegler and D. Albanes, Serum 25-hydroxyvitamin D, vitamin D binding protein and risk of colorectal cancer in the Prostate, Lung, Colorectal and Ovarian Cancer Screening TrialInt J Cancer 136, 2015, E654-E664,

- https://doi.org/10.1002/ijc.29157.
- [33] E.M. Burns, C.A. Elmets and N. Yusuf, Invited Review Vitamin D and skin cancer Photochem. review vitamin d and skin cancer, Photochem 91, 2015, 201-209, https://doi.org/10.1111/php.12382.
- [34] M.N. Ombra, P. Paliogiannis, V. Doneddu, M.C. Sini, M. Colombino, C. Rozzo, I. Stanganelli, F. Tanda, A. Cossu and G. Palmieri, Vitamin D status and risk for malignant cutaneous melanoma: recent advances, Eur. J. Cancer Prev. 26, 2017, 532-541, https://doi.org/10.1097/CEJ.0000000000000334.
- [35] F. Pandolfi, L. Franza, C. Mandolini and P. Conti, Immune Modulation by Vitamin D: Special Emphasis on Its Role in Prevention and Treatment of Cancer Clim Modulation by Vitamin d: special emphasis on its role in prevention and treatment of Cancer, Clim. Ther. 39, 2017, 884-893, https://doi.org/10.1016/j.clinthera.2017.03.012.
- [36] A. Piotrowska, J. Wierzbicka and M.A. Żmijewski, Vitamin D in the skin physiology and pathology, Acta Biochim, Pol. 63, 2016, 17-29, https://doi.org/10.18388/abp.2015 1104.
- [37] J. Inoue, J.M. Choi, T. Yoshidomi, T. Yashiro and R. Sato, Quercetin enhances VDR activity, leading to stimulation of its target gene expression in Caco-2 cells, J. Nutr. Sci. Vitaminol. (Tokyo). Nutr. Sci. Vitaminol. (Tokyo). 56, 2010 326-330, https://doi.org/10.3177/insv.56.326.
- [38] Y.J. Chae, K.H. Cho, I.S. Yoon, C.K. Noh, H.J. Lee, Y. Park, E. Ji, M.D. Seo and H.J. Maeng, Vitamin Deceptor Mediated Upregulation of CYP3A4 and MDR1 by Qd receptor-mediated upregulation of CYP3A4 and MDR1 by quercetin in Caco-2 cells, Planta Med. 82, 2016, 121-130, https://doi.org/10.1055/s-0035-1557898.
- [39] K.Y. Lee, H.S. Choi, H.S. Choi, K.Y. Chung, B.J. Lee, H.J. Maeng and M.D. Seo, Quercetin Directly Interacts with Vitamin D Receptor (VDR): Structural Implication of VDR Activation by QuercetinBiomol Ther (Seoul).directly interacts with vitamin d receptor (VDR): structural implication of VDR activation by quercetin, Biomol. Ther. (Seoul) 24, 2016, 191-198, https://doi.org/10.4062/biomolther.2015.122.
- [40] A.J. Lau, R. Politi, G. Yang and T.K. Chang, Cell-based and in silico evidence against quercetin and structurally-related flavonols as activators of vitamin D receptor, J-Steroid Biochem Mol. Biol. 163, 2016, 59-67, https://doi.org/10.1016/j.jsbmb.2016.03.039.
- [41] T. Thangasamy, S. Sittadjody, G.C. Mitchell, E.E. Mendoza, V.M. Radhakrishnan, K.H. Limesand and R. Burd, Quercetin abrogates chemoresistance in melanoma cells by modulating ΔNp73, BMC Cancer 10, 2010, 282, https://doi.org/10.1186/1471-2407-10-282.
- [42] K.A. Turner, J.M. Manouchehri and M. Kalafatis, Sensitization of recombinant human tumor necrosis factor-related apoptosis-inducing ligand-resistant malignant melanomas by quercetin, *Melanoma Res.* 2018, 1, https://doi.org/10.1097/CMR.0000000000000447.
- [43] S.J. Pradhan, R. Mishra, P. Sharma and G.C. Kundu, Quercetin and sulforaphane in combination suppress the progression of melanoma through the down-regulation of matrix metalloproteinase-9, Exp. Ther. Med. 1, 2010, 915-920, https://doi.org/10.3892/etm.2010.144.
- [44] N.S. Srivastava and R.A.K. Srivastava, Curcumin and quercetin synergistically inhibit cancer cell proliferation in multiple cancer cells and modulate Wnt/β-catenin signaling and apoptotic pathways in A375 cells, *Phytomedicine*. **52**, 2019, 117-128, https://doi.org/10.1016/j.phymed.2018.09.224.
- [45] P. Ferrer, M. Asensi, R. Segarra, A. Ortega, M. Benlloch, E. Obrador, M.T. Varea, G. Asensio, L. Jordá and J.M. Estrela, Association between Pterostilbene and Quercetin Inhibits Metastatic Activity of B16 melanoma, Neoplasia 7, 2005, 37-47, https://doi.org/10.1593/neo.04337.
- [46] K. Yamauchi, T. Mitsunaga, M. Inagaki and T. Suzuki, Synthesized quercetin derivatives stimulate melanogenesis in B16 melanoma cells by influencing the expression of melanin biosynthesis proteins MITF and p38 MAPK, Bioorganic Med. Chem. Med. Chem. Lett. 22, 2014, 3331-3340, https://doi.org/10.1016/j.bmc.2014.04.053.
- [47] K. Yamauchi, T. Mitsunaga, S.H. Afroze and M.N. Uddin, Structure Activity Relationships of Methylquercetin on Anti-migration and Anti-proliferation Activity in B16 Melanoma C\_activity relationships of Methylquercetin on anti-migration and anti-proliferation activity in B16 melanoma cells, Anticancer Res. 37, 2017, 1575–1579, https://doi.org/10.21873/anticanres.11487.
- [48] K. Yamauchi and T. Mitsunaga, Methylquercetins stimulate melanin biosynthesis in a three-dimensional skin model, J. Nat. Med. 72, 2018, 563-569, https://doi.org/10.1007/s11418-018-1175-0.
- [49] K. Yamauchi, T. Mitsunaga and I. Batubara, Novel guercetin glucosides from Helminthostachys zeylanica root and acceleratory activity of melanin biosynthesis, J. Nat. Med. 67, 2013, 369-374,

- https://doi.org/10.1007/s11418-012-0672-9.
- [50] K. Yamauchi, T. Mitsunaga and I. Batubara, Synthesis of quercetin glycosides and their melanogenesis stimulatory activity in B16 melanoma cells, *Bioorganic Med. Chem. Med. Chem. Lett.* 22, 2014, 937-944, https://doi.org/10.1016/j.bmc.2013.12.062.
- [51] H.G. Jung, H.H. Kim, S. Paul, J.Y. Jang, Y.H. Cho, H.J. Kim, J.M. Yu, E.S. Lee, B.J. An, S.C. Kang and B.H. Bang, Quercetin-3-O-β-d-glucopyranosyl-(1→6)-β-d-glucopyranoside suppresses melanin synthesis by augmenting p38 MAPK and CREB signaling pathways and subsequent cAMP down-regulation in murine melanoma cells, Saudi J. Biol. Sci. 22, 2015, 706-713, https://doi.org/10.1016/j.sjbs.2015.03.009.
- [52] A.S. Jain, S.M. Shah, M.S. Nagarsenker, Y. Nikam, R.P. Gude, F. Steiniger, J. Thamm and A. Fahr, Lipid Colloidal Carriers for Improvement of Anticancer Activity of Orally Delivered Quercetin: Formulation, Characterization and Establishing In Vitro In Vivo Acolloidal carriers for improvement of anticancer activity of orally delivered quercetin: formulation, characterization and establishing in vitro In vivo advantage, J. Biomed. Nanotechnol. 9, 2013, 1230–1240, https://doi.org/10.1166/jbn.2013.1636.
- [53] C.L. Dora, L.F. Costa Silva, L. Mazzarino, J.M. Siqueira, D. Fernandes, L.K. Pacheco, M.F. Maioral, M.C. Santos-Silva, A.L. Muccillo Baisch, J. Assreuy and E. Lemos-Senna, Oral Delivery of a High Quercetin Payload Nanosized Emulsion: In Vitro and In Vivo Activity Against B16-F10 Melanoma, J. Nanosci. Nanotechnol. 16, 2016, 1275-1281, https://doi.org/10.1166/jnn.2016.11675.
- [54] H.C. Pal, R.L. Pearlman and F. Afaq, Fisetin and Its Role in Chronic Dits role in chronic diseases, Adv. Exp. Med. Biol. 928, 2016, 213-244, https://doi.org/10.1007/978-3-319-41334-1 10.
- [55] D.N. Syed, F. Afaq, N. Maddodi, J.J. Johnson, S. Sarfaraz, A. Ahmad, V. Setaluri and H. Mukhtar, Inhibition of Human Melanoma Cell Growth by the Dietary Flavonoid Fisetin Is Associated with Disruption of Wnt/ Catenin Signaling and decreased Mitf-Lhuman melanoma cell growth by the dietary flavonoid fisetin is associated with disruption of wnt/β-Catenin signaling and decreased mitf-levels, J. Invest. Dermatol. 131, 2011, 1291-1299, https://doi.org/10.1038/jid.2011.6.
- [56] D.N. Syed, J.C. Chamcheu, M.I. Khan, M. Sechi, R.K. Lall, V.M. Adhami and H. Mukhtar, Fisetin inhibits human melanoma cell growth through direct binding to p70S6K and mTOR: Findings from 3-D melanoma skin equivalents and computational modeling, *Biochem. Pharmacol.* 89, 2014, 349–360, https://doi.org/10.1016/j.bcp.2014.03.007.
- [57] D.N. Syed, R.K. Lall, J.C. Chamcheu, O. Haidar and H. Mukhtar, Involvement of ER stress and activation of apoptotic pathways in fisetin induced cytotoxicity in human melanoma, *Arch. Biochem. Biophys.* 563, 2014, 108–117, https://doi.org/10.1016/j.abb.2014.06.034.
- [58] K. Wang, D.-N. Hu, H.-W. Lin, W.-E. Yang, Y.-H. Hsieh, H.-W. Chien and S.-F. Yang, Fisetin induces apoptosis through mitochondrial apoptosis pathway in human uveal melanoma cells, *Environ. Toxicol.* 33, 2018, 527–534, https://doi.org/10.1002/tox.22538.
- [59] M. Sechi, R.K. Lall, S.O. Afolabi, A. Singh, D.C. Joshi, S.-Y. Chiu, H. Mukhtar and D.N. Syed, Fisetin targets YB-1/RSK axis independent of its effect on ERK signaling: insights from in vitro and in vivo melanoma models, *Sci. Rep.* 8, 2018, 15726, https://doi.org/10.1038/s41598-018-33879-w.
- [60] H.C. Pal, S. Sharma, L.R. Strickland, S.K. Katiyar, M.E. Ballestas, M. Athar, C.A. Elmets and F. Afaq, Fisetin inhibits human melanoma cell invasion through promotion of mesenchymal to epithelial transition and by targeting MAPK and NFκB signaling pathways, *PLoS One* 9, 2014, e86338https://doi.org/10.1371/journal.pone.0086338.
- [61] M.-S. Shon, R.-H. Kim, O.J. Kwon, S.-S. Roh and G.-N. Kim, Beneficial role and function of fisetin in skin health via regulation of the CCN2/TGF-β signaling pathway, Food Sci. Biotechnol. 25, 2016, 133–141, https://doi.org/10.1007/s10068-016-0110-y.
- [62] A. Kumagai, N. Horike, Y. Satoh, T. Uebi, T. Sasaki, Y. Itoh, Y. Hirata, K. Uchio-Yamada, K. Kitagawa, S. Uesato, H. Kawahara, H. Takemori and Y. Nagaoka, A potent inhibitor of sik2, 3, 3′, 7-trihydroxy-4′-methoxyflavon (4′-o-methylfisetin), promotes melanogenesis in b16f10 melanoma cells, *PLoS One* 6, 2011, e26148, https://doi.org/10.1371/journal.pone.0026148.
- [63] H.C. Pal, R.D. Baxter, K.M. Hunt, J. Agarwal, C.A. Elmets, M. Athar and F. Afaq, Fisetin, a phytochemical, potentiates sorafenib-induced apoptosis and abrogates tumor growth in athymic nude mice implanted with BRAF-mutated melanoma cells, Oncotarget 6, 2015, 28296-28311, https://doi.org/10.18632/oncotarget.5064.
- [64] H.C. Pal, A.C. Diamond, L.R. Strickland, J.C. Kappes, S.K. Katiyar, C.A. Elmets, M. Athar and F. Afaq, Fisetin, a dietary flavonoid, augments the anti-invasive and anti-metastatic potential of sorafenib in melanoma,

- Oncotarget 7, 2016, https://doi.org/10.18632/oncotarget.6237.
- [65] C. Yi, Y. Zhang, Z. Yu, Y. Xiao, J. Wang, H. Qiu, W. Yu, R. Tang, Y. Yuan, W. Guo and W. Deng, Melatonin Enhances the Anti Tumor Effect of Fisetin by Inhibiting COX 2/iNOS and NF-κβ/p300 Signaling PathwaysPLoS One.enhances the anti-tumor effect of Fisetin by inhibiting COX-2/iNOS and NF-κβ/p300 Signaling pathways, PLoS One 9, 2014, e99943https://doi.org/10.1371/journal.pone.0099943.
- [66] F. Ali, F. Rahul, S. Naz and Y.H. Jyoti, Siddique, Health functionality of apigenin: Aa review, Int. J. Food Prop. 20, 2017, 1197-1238, https://doi.org/10.1080/10942912.2016.1207188.
- [67] G. Zhao, X. Han, W. Cheng, J. Ni, Y. Zhang, J. Lin and Z. Song, Apigenin inhibits proliferation and invasion, and induces apoptosis and cell cycle arrest in human melanoma cells, *Oncol. Rep.* 37, 2017, 2277–2285, https://doi.org/10.3892/or.2017.5450.
- [68] S. Das, J. Das, A. Samadder, N. Boujedaini and A.R. Khuda-Bukhsh, Apigenin-induced apoptosis in A375 and A549 cells through selective action and dysfunction of mitochondria, *Exp. Biol. Med.* 237, 2012, 1433–1448, https://doi.org/10.1258/ebm.2012.012148.
- [69] M.A. Hasnat, M. Pervin, J.H. Lim and B.O. Lim, Apigenin attenuates melanoma cell migration by inducing anoikis through integrin and focal adhesion kinase inhibition, *Molecules* 20, 2015, 21157–21166, https://doi.org/10.3390/molecules201219752.
- [70] H.H. Cao, J.H. Chu, H.Y. Kwan, T. Su, H. Yu, C.Y. Cheng, X.Q. Fu, H. Guo, T. Li, A.K.W. Tse, G.X. Chou, H.B. Mo and Z.L. Yu, Inhibition of the STAT3 signaling pathway contributes to apigenin-mediated anti-metastatic effect in melanoma, *Sci. Rep.* 6, 2016, 21731, https://doi.org/10.1038/srep21731.
- [71] S.-C. Chao, S.-C. Huang, D.-N. Hu and H.-Y. Lin, Subtoxic levels of apigenin inhibit expression and secretion of VEGF by uveal melanoma cells via suppression of ERK1/2 and PI3K/Akt pathways, evidence-based complement, Altern Med. 2013, 2013, 817674https://doi.org/10.1155/2013/817674.
- [72] L. Xu, Y. Zhang, K. Tian, X. Chen, R. Zhang, X. Mu, Y. Wu, D. Wang, S. Wang, F. Liu, T. Wang, J. Zhang, S. Liu, Y. Zhang, C. Tu and H. Liu, Apigenin suppresses PD-L1 expression in melanoma and host dendritic cells to elicit synergistic therapeutic effects, J. Exp. Clin. Cancer Res. 37, 2018, 261, https://doi.org/10.1186/s13046-018-0929-6.
- [73] J. Ding, G. Polier, R. Köhler, M. Giaisi, P.H. Krammer and M. Li-Weber, Wogonin and related natural flavones overcome tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) protein resistance of tumors by down-regulation of c-FLIP protein and up-regulation of TRAIL receptor 2 expression, J. Biol. Chem. 287, 2012, 641-649, https://doi.org/10.1074/jbc.M111.286526.
- [74] Y. Ye, G.-X. Chou, H. Wang, J.-H. Chu and Z.-L. Yu, Flavonoids, apigenin and icariin exert potent melanogenic activities in murine B16 melanoma cells, *Phytomedicine* 18, 2010, 32–35, https://doi.org/10.1016/j.phymed.2010.06.004.
- [75] Y. Ye, H. Wang, J.H. Chu, G.X. Chou and Z.L. Yu, Activation of p38 MAPK pathway contributes to the melanogenic property of apigenin in B16 cells, *Exp. Dermatol.* 20, 2011, 755-757, https://doi.org/10.1111/j.1600-0625.2011.01297.x.
- [76] N. Nasr Bouzaiene, F. Chaabane, A. Sassi, L. Chekir-Ghedira and K. Ghedira, Effect of apigenin-7-glucoside, genkwanin and naringenin on tyrosinase activity and melanin synthesis in B16F10 melanoma cells, *Life Sci.* 144, 2016, 80–85, https://doi.org/10.1016/j.lfs.2015.11.030.
- [77] S. Das, J. Das, A. Samadder, A. Paul and A.R. Khuda-Bukhsh, Strategic formulation of apigenin-loaded PLGA nanoparticles for intracellular trafficking, DNA targeting and improved therapeutic effects in skin melanoma in vitro, *Toxicol. Lett.* 223, 2013, 124-138, https://doi.org/10.1016/j.toxlet.2013.09.012.
- [78] S.F. Nabavi, N. Braidy, O. Gortzi, E. Sobarzo-Sanchez, M. Daglia, K. Skalicka-Woźniak and S.M. Nabavi, Luteolin as an anti-inflammatory and neuroprotective agent: Aa brief review, *Brain Res. Bull.* 119, 2015, 1-11, https://doi.org/10.1016/j.brainresbull.2015.09.002.
- [79] M. Imran, A. Rauf, T. Abu-Izneid, M. Nadeem, M.A. Shariati, I.A. Khan, A. Imran, I.E. Orhan, M. Rizwan, M. Atif, T.A. Gondal and M.S. Mubarak, Luteolin, a flavonoid, as an anticancer agent: Aa review, *Biomed. Pharmacothe.*112, 2019, , 108612https://doi.org/10.1016/j.biopha.2019.108612.
- [80] F. Casagrande and J.M. Darbon, Effects of structurally related flavonoids on cell cycle progression of human melanoma cells: Regulation of cyclin-dependent kinases CDK2 and CDK1, *Biochem. Pharmacol.* 61, 2001, 1205–1215, https://doi.org/10.1016/S0006-2952(01)00583-4.

- [81] J.S. Ruan, Y.P. Liu, L. Zhang, L.G. Yan, F.T. Fan, C.S. Shen, A.Y. Wang, S.Z. Zheng, S.M. Wang and Y. Lu, Luteolin reduces the invasive potential of malignant melanoma cells by targeting β3 integrin and the epithelial-mesenchymal transition, *Acta Pharmacol. Sin.* 33, 2012, 1325–1331, https://doi.org/10.1038/aps.2012.93.
- [82] X. Yao, W. Jiang, D. Yu and Z. Yan, Luteolin inhibits proliferation and induces apoptosis of human melanoma cells in vivo and in vitro by suppressing MMP-2 and MMP-9 through the PI3K/AKT pathway, Food Funct.

  10, 2019, 703-712, https://doi.org/10.1039/c8fo02013b.
- [83] C. Li, Q. Wang, S. Shen, X. Wei and G. Li, HIF-1α/VEGF signaling-mediated epithelial-mesenchymal transition and angiogenesis is critically involved in anti-metastasis effect of luteolin in melanoma cells, *Phyther. Res* 33, 2019, 798-807, https://doi.org/10.1002/ptr.6273.
- [84] P. Limonta, R.M. Moretti, M. Marzagalli, F. Fontana, M. Raimondi and M. Montagnani Marelli, Role of Endoplasmic Reticulum Stress in the Anticancer Activity of Natural Compounds, Int. J. Mol. Sci. 20, 2019, https://doi.org/10.3390/ijms20040961.
- [85] J.K. Kim, K.A. Kang, Y.S. Ryu, M.J. Piao, X. Han, M.C. Oh, S.J. Boo, S.U. Jeong, Y.J. Jeong, S. Chae, S.Y. Na and J.W. Hyun, Induction of Endoplasmic Reticulum Stress via Reactive Oxygen Species Mediated by Luteolin in Melanoma Cells, Anticancer Res. 36, 2016, 2281-2289.
- [86] R. Balyan, S.K. Kudugunti, H.A. Hamad, M.S. Yousef and M.Y. Moridani, Bioactivation of luteolin by tyrosinase selectively inhibits glutathione S-transferase, *Chem. Biol. Interact.* 240, 2015, 208–218, https://doi.org/10.1016/j.cbi.2015.08.011.
- [87] S.M. An, H.J. Kim, J.-E. Kim and Y.C. Boo, Flavonoids, taxifolin and luteolin attenuate cellular melanogenesis despite increasing tyrosinase protein levels, *Phyther. Res.* 22, 2008, 1200-1207, https://doi.org/10.1002/ptr.2435.
- [88] M.Y. Choi, H.S. Song, H.S. Hur and S.S. Sim, Whitening activity of luteolin related to the inhibition of cAMP pathway in α-MSH-stimulated B16 melanoma cells, *Arch. Pharm. Res.* 31, 2008, 1166-1171, https://doi.org/10.1007/s12272-001-1284-4.
- [89] E.T. Arung, K. Shimizu, H. Tanaka and R. Kondo, 3-Prenyl luteolin, a new prenylated flavone with melanin biosynthesis inhibitory activity from wood of Artocarpus heterophyllus, *Fitoterapia* 81, 2010, 640-643, https://doi.org/10.1016/j.fitote.2010.03.011.
- [90] J.Y. Kwak, J.K. Seok, H.J. Suh, Y.H. Choi, S.S. Hong, D.S. Kim and Y.C. Boo, Antimelanogenic effects of luteolin 7-sulfate isolated from Phyllospadix iwatensis Makino, *Br. J. Dermatol.* 175, 2016, 501-511, https://doi.org/10.1111/bjd.14496.
- [91] K. Yamauchi, A. Fujieda and T. Mitsunaga, Selective synthesis of 7-O-substituted luteolin derivatives and their melanonenesis and proliferation inhibitory activity in B16 melanoma cells, *Bioorganie\_Med. Chem. Lett.*28, 2018, 2518-2522, https://doi.org/10.1016/j.bmcl.2018.05.051.
- [92] E.-B. Byun, H.-Y. Song, S. Mushtaq, H.-M. Kim, J.A. Kang, M.-S. Yang, N.-Y. Sung, B.-S. Jang and E.-H. Byun, Gamma-Irradiated Luteolin Inhibits 3-Isobutyl-1-Methylxanthine-Induced Melanogenesis Through the Regulation of CREB/MITF, PI3K/Akt, and ERK Pathways in B16BL6 Melanoma Cells J. Med. Food. irradiated luteolin inhibits 3-Isobutyl-1-Methylxanthine-Induced melanogenesis through the regulation of CREB/MITF, PI3K/Akt, and ERK pathways in B16BL6 melanoma cells, J. Med. Food 20, 2017, 812-819, https://doi.org/10.1089/jmf.2016.3890.
- [93] S. Ziaei, R. Halaby and Dietary Isoflavones, Breast Cancer RiskMedicines.risk, Medicines 4, 2017, https://doi.org/10.3390/medicines4020018.
- [94] M.K. Sivoňová, P. Kaplán, Z. Tatarková, L. Lichardusová, R. Dušenka and J. Jurečeková, Androgen receptor and soy isoflavones in prostate cancer, *Mol. Clin. Oncol.* 10, 2019, 191-204, https://doi.org/10.3892/mco.2018.1792.
- [95] S. Rauth, J. Kichina and A. Green, Inhibition of growth and induction of differentiation of metastatic melanoma cells in vitro by genistein: chemosensitivity is regulated by cellular p53, *Br. J. Cancer* 75, 1997, 1559–1566 http://www.ncbi.nlm.nih.gov/pubmed/9184169.
- [96] R.R. Hartmann, D. Rimoldi, F.J. Lejeune and S. Carrel, Cell differentiation and cell-cycle alterations by tyrosine kinase inhibitors in human melanoma cells, *Melanoma Res.* 7 (Suppl. 2), 1997, S27-S33 http://www.ncbi.nlm.nih.gov/pubmed/9578414.

- [97] I.R. Record, J.L. Broadbent, R.A. King, I.E. Dreosti, R.J. Head and A.L. Tonkin, Genistein inhibits growth of B16 melanoma cells in vivo and in vitro and promotes differentiation in vitro, *Int. J. Cancer* 72, 1997, 860-864, https://doi.org/10.1002/(SICI)1097-0215(19970904)72:5<860::AID-I]C24>3.0.CO;2-B.
- [98] T. Kuzumaki, T. Kobayashi and K. Ishikawa, Genistein induces p21(Cip1/WAF1) expression and blocks the G1 to S phase transition in mouse fibroblast and melanoma cells, *Biochem. Biophys. Res. Commun.* 251, 1998, 291–295, https://doi.org/10.1006/bbrc.1998.9462.
- [99] C.-H. Yan, X.-G. Chen, V. Li and R. Han, Effects of Genistein, A Soybean Derived Isoflavone, on Proliferation and Differentiation of B16-BL6 Mouse Melanoma Cgenistein, a soybean-derived isoflavone, on proliferation and differentiation of B16-BL6 mouse melanoma cells, J. Asian Nat. Prod. Res. 1, 1999, 285-299, https://doi.org/10.1080/10286029908039877.
- [100] J.M. Darbon, M. Penary, N. Escalas, F. Casagrande, F. Goubin-Gramatica, C. Baudouin and B. Ducommun, Distinct Chk2 activation pathways are triggered by genistein and DNA- damaging agents in human melanoma cells, J. Biol. Chem. 275, 2000, 15363-15369, https://doi.org/10.1074/jbc.275.20.15363.
- [101] F. Casagrande and J.M. Darbon, p21(CIP1) is dispensable for the G2 arrest caused by genistein in human melanoma cells, Exp. Cell Res. 258, 2000, 101-108, https://doi.org/10.1006/excr.2000.4914.
- [102] H.Z. Wang, Y. Zhang, L.P. Xie, X.Y. Yu and R.Q. Zhang, Effects of genistein and daidzein on the cell growth, cell cycle, and differentiation of human and murine melanoma cells, J. Nutr. Biochem. 13, 2002, 421-426, https://doi.org/10.1016/S0955-2863(02)00184-5.
- [103] I. Venza, M. Visalli, R. Oteri, C. Beninati, D. Teti and M. Venza, Genistein reduces proliferation of EP3-expressing melanoma cells through inhibition of PGE2-induced IL-8 expression, *Int. Immunopharmacol.* 62, 2018, 86-95, https://doi.org/10.1016/j.intimp.2018.06.009.
- [104] Q. Sun, R. Cong, H. Yan, H. Gu, Y. Zeng, N. Liu, J. Chen and B. Wang, Genistein inhibits growth of human uveal melanoma cells and affects microRNA-27a and target gene expression, *Oncol. Rep.* 22, 2009, 563–567, https://doi.org/10.3892/or 00000472.
- [105] C. Yan and R. Han, Suppression of adhesion-induced protein tyrosine phosphorylation decreases invasive and metastatic potentials of B16-BL6 melanoma cells by protein tyrosine kinase inhibitor genistein,

  \*Invasion Metastasis\*\* 17, 1997, 189-198 http://www.ncbi.nlm.nih.gov/pubmed/9778591.
- [106] C. Yan and R. Han, Genistein suppresses adhesion-induced protein tyrosine phosphorylation and invasion of B16-BL6 melanoma cells, *Cancer Lett.* 129, 1998, 117-124, https://doi.org/10.1016/S0304-3835(98)00093-7.
- [107] S. Cui, J. Wang, Q. Wu, J. Qian, C. Yang and P. Bo, Genistein inhibits the growth and regulates the migration and invasion abilities of melanoma cells via the FAK/paxillin and MAPK pathways, *Oncotarget* 8, 2017, 21674–21691, https://doi.org/10.18632/oncotarget.15535.
- [108] L.G. Menon, R. Kuttan, M.G. Nair, Y.C. Chang and G. Kuttan, Effect of isoflavones genistein and daidzein in the inhibition of lung metastasis in mice induced by B16F-10 melanoma cells, *Nutr. Cancer* 30, 1998, 74-77 https://doi.org/10.1080/01635589809514644.
- [109] D. Li, J.A. Yee, M.H. McGuire, P.A. Murphy and L. Yan, Soybean isoflavones reduce experimental metastasis in mice, J. Nutr. 129, 1999, 1075-1078.
- [110] C. Danciu, F. Borcan, F. Bojin, I. Zupko and C. Dehelean, Effect of the isoflavone genistein on tumor size, metastasis potential and melanization in a B16 mouse model of murine melanoma, Nat. Prod. Prod. Commun. 8, 2013, 343-346.
- [111] H.G. Farina, M. Pomies, D.F. Alonso and D.E. Gomez, Antitumor and antiangiogenic activity of soy isoflavone genistein in mouse models of melanoma and breast cancer, *Oncol. Rep.* 16, 2006, 885-891 <a href="http://www.ncbi.nlm.nih.gov/pubmed/16969510">http://www.ncbi.nlm.nih.gov/pubmed/16969510</a>.
- [112] R. Cong, Q. Sun, L. Yang, H. Gu, Y. Zeng and B. Wang, Effect of Genistein on vasculogenic mimicry formation by human uveal melanoma cells, *J. Exp. Clin. Cancer Res.* 28, 2009, 124, https://doi.org/10.1186/1756-9966-28-124.
- [113] T.L. Guo, J.A. McCay, L.X. Zhang, R.D. Brown, L. You, N.A. Karrow, D.R. Germolec and K.L. White, Genistein modulates immune responses and increases host resistance to B16F10 tumor in adult female B6C3F1 mice, Nutr. 131, 2001, 3251-3258, https://doi.org/10.1093/in/131.12.3251.

- [114] J. Wietrzyk, J. Boratynski, G. Grynkiewicz, A. Ryczynski, C. Radzikowski and A. Opolski, Antiangiogenic and antitumour effects in vivo of genistein applied alone or combined with cyclophosphamide, *Anticancer Res.* 21, 2001, 3893–3896 http://www.ncbi.nlm.nih.gov/pubmed/11911265.
- [115] S. Tamura, T. Bito, M. Ichihashi and M. Ueda, Genistein enhances the cisplatin-induced inhibition of cell growth and apoptosis in human malignant melanoma cells, *Pigment. Cell Res.* 16, 2003, 470-476, https://doi.org/10.1034/j.1600-0749.2003.00068.x.
- [116] C. Ji, Y.L. Yang, L. He, B. Gu, J.P. Xia, W.L. Sun, Z.L. Su, B. Chen and Z.G. Bi, Increasing ceramides sensitizes genistein-induced melanoma cell apoptosis and growth inhibition, *Biochem. Biophys. Res. Commun.* 421, 2012, 462–467, https://doi.org/10.1016/j.bbrc.2012.04.012.
- [117] C.M. Chiang, Y.J. Chang, J.Y. Wu and T.S. Chang, Production and anti-melanoma activity of methoxyisoflavones from the biotransformation of genistein by two recombinant Escherichia coli strains, *Molecules* 22, 2017, https://doi.org/10.3390/molecules22010087.
- [118] C. Danciu, S. Berkó, G. Varju, B. Balázs, L. Kemény, I.B. Németh, A. Cioca, A. Petrus, C. Dehelean, C.I. Cosmin, E. Amaricai and C.C. Toma, The effect of electroporation of a lyotroic liquid crystal genistein-based formulation in the recovery of murine melanoma lesions, *Int. J. Mol. Sci.* 16, 2015, 15425–15441, https://doi.org/10.3390/ijms160715425.
- [119] C. Chu, J. Deng, Y. Man and Y. Qu, Green Tea Extracts Epigallocatechin 3-gallate for Different Ttea extracts Epigallocatechin-3-gallate for different treatments, Biomed Res. Int. 2017, 2017, 5615647https://doi.org/10.1155/2017/5615647.
- [120] M. Nihal, N. Ahmad, H. Mukhtar and G.S. Wood, Anti-proliferative and proapoptotic effects of (-)-epigallocatechin-3- gallate on human melanoma: Possible implications for the chemoprevention of melanoma, Int. J. Cancer 114, 2005, 513-521, https://doi.org/10.1002/ijc.20785.
- [121] L.Z. Ellis, W. Liu, Y. Luo, M. Okamoto, D. Qu, J.H. Dunn and M. Fujita, Green tea polyphenol epigallocatechin-3-gallate suppresses melanoma growth by inhibiting inflammasome and IL-1β secretion, *Biochem. Biophys. Res. Commun.* 414, 2011, 551–556, https://doi.org/10.1016/j.bbrc.2011.09.115.
- [122] L. Konta, P. Száraz, J.É. Magyar, K. Révész, G. Bánhegyi, J. Mandl and M. Csala, Inhibition of glycoprotein synthesis in the endoplasmic reticulum as a novel anticancer mechanism of (-)-epigallocatechin-3-gallate, *BioFactors* 37, 2011, 468-476, https://doi.org/10.1002/biof.189.
- [123] S. Tsukamoto, Y. Huang, D. Umeda, S. Yamada, S. Yamashita, M. Kumazoe, Y. Kim, M. Murata, K. Yamada and H. Tachibana, 67-kDa laminin receptor-dependent Protein Phosphatase 2A (PP2A) activation elicits melanoma-specific antitumor activity overcoming drug resistance, J. Biol. Chem. 289, 2014, 32671–32681, https://doi.org/10.1074/jbc.M114.604983.
- [124] S. Yamada, S. Tsukamoto, Y. Huang, A. Makio, M. Kumazoe, S. Yamashita and H. Tachibana, Epigallocatechin-3-O-gallate up-regulates microRNA-let-7b expression by activating 67-kDa laminin receptor signaling in melanoma cells, Sci. Rep. 6, 2016, 19225, https://doi.org/10.1038/srep19225.
- [125] Y. Wu, Y. Lin, H. Liu and J. Li, Inhibition of invasion and up-regulation of E-cadherin expression in human malignant melanoma cell line A375 by (-)-epigallocatechin-3-gallate, J. Huazhong Univ Sci. Technol. [Medical]

  Huazhong Univ Sci. Technol. Med. Sci. 28, 2008, 356-359, https://doi.org/10.1007/s11596-008-0330-3.
- [126] I.H. Kwak, Y.H. Shin, M. Kim, H.Y. Cha, H.J. Nam, B.S. Lee, S.C. Chaudhary, K.S. Pai and J.H. Lee, Epigallocatechin-3-gallate inhibits paracrine and autocrine hepatocyte growth factor/scatter factor-induced tumor cell migration and invasion, Exp. Mol. Med. 43, 2011, 111-120, https://doi.org/10.3858/emm.2011.43.2.013.
- [127] T. Watanabe, H. Kuramochi, A. Takahashi, K. Imai, N. Katsuta, T. Nakayama, H. Fujiki and M. Suganuma, Higher cell stiffness indicating lower metastatic potential in B16 melanoma cell variants and in (2)-epigallocatechin gallate-treated cells, *J. Cancer Res. Clin. Oncol.* 138, 2012, 859-866, https://doi.org/10.1007/s00432-012-1159-5.
- [128] C.-W. Chang, Y.-H. Hsieh, W.-E. Yang, S.-F. Yang, Y. Chen and D.-N. Hu, Epigallocatechingallate inhibits migration of human uveal melanoma cells via downregulation of matrix Metalloproteinase-2 activity and ERK1/2 pathway Biomed Res. Int. 2014, 2014, 1-9, https://doi.org/10.1155/2014/141582.
- [129] J. Zhang, Z. Lei, Z. Huang, X. Zhang, Y. Zhou, Z. Luo, W. Zeng, J. Su, C. Peng and X. Chen, Epigallocatechin-3-gallate(EGCG) suppresses melanoma cell growth and metastasis by targeting TRAF6 activity, Oncotarget 7 2016, 79557-79571, https://doi.org/10.18632/oncotarget.12836.

- [130] A. Rawangkan, P. Wongsirisin, K. Namiki, K. Iida, Y. Kobayashi, Y. Shimizu, H. Fujiki and M. Suganuma, Green tea catechin is an alternative immune checkpoint inhibitor that inhibits PD-l1 expression and lung tumor growth, *Molecules* 23, 2018, https://doi.org/10.3390/molecules23082071.
- [131] D.S. Kim, S.H. Park, S.B. Kwon, K. Li, S.W. Youn and K.C. Park, (-)-Epigallocatechin-3-gallate and hinokitiol reduce melanin synthesis via decreased MITF production, *Arch. Pharm. Res.* 27, 2004, 334–339, https://doi.org/10.1007/BF02980069.
- [132] K. Sato and M. Toriyama, Depigmenting Effect of Catechins Molecules: effect of catechins, Molecules 14, 2009, 4425-4432, https://doi.org/10.3390/molecules14114425.
- [133] J.D. Liu, S.H. Chen, C.L. Lin, S.H. Tsai and Y.C. Liang, Inhibition of melanoma growth and metastasis by combination with (-)-epigallocatechin-3-gallate and dacarbazine in mice, *J. Cell. Biochem.* 83, 2001, 631-642, https://doi.org/10.1002/jcb.1261.
- [134] Q. Shen, F. Tian, P. Jiang, Y. Li, L. Zhang, J. Lu and J. Li, EGCG enhances TRAIL-mediated apoptosis in human melanoma A375 cell line, *J. Huazhong Univ. Sci. Technol.* [Medical Med. Sci. 29, 2009, 771-775, https://doi.org/10.1007/s11596-009-0620-4.
- [135] M. Nihal, H. Ahsan, I.A. Siddiqui, H. Mukhtar, N. Ahmad and G.S. Wood, (-)-Epigallocatechin-3-gallate (EGCG) sensitizes melanoma cells to interferon induced growth inhibition in a mouse model of human melanoma *Cell Cycle* 3, 2009, 2057–2063, https://doi.org/10.4161/cc.8.13.8862.
- [136] M. Nihal, C.T. Roelke and G.S. Wood, Anti-melanoma effects of vorinostat in combination with polyphenolic antioxidant (-)-Epigallocatechin-3-Gallate (EGCG), *Pharm. Res.* 27, 2010, 1103–1114, https://doi.org/10.1007/s11095-010-0054-5.
- [137] J.H. Lee, M. Kishikawa, M. Kumazoe, K. Yamada and H. Tachibana, Vitamin A enhances antitumor effect of a green tea polyphenol on melanoma by upregulating the polyphenol sensing molecule 67-kDa laminin receptor, *PLoS One* 5, 2010, e11051https://doi.org/10.1371/journal.pone.0011051.
- [138] S. Tsukamoto, Y. Huang, D. Umeda, S. Yamashita, M. Kumazoe, Y. Kim, M. Murata, K. Yamada and H. Tachibana, 67-kDa laminin receptor-dependent protein phosphatase 2A (PP2A) activation elicits melanoma-specific antitumor activity overcoming drug resistance, Biol. Chem. 289, 2014, 32671-32681, https://doi.org/10.1074/jbc.M114.604983.
- [139] L. Sánchez-del-Campo and J.N. Rodríguez-López, Targeting the methionine cycle for melanoma therapy with 3-O-(3,4,5-trimethoxybenzoyl)-(-)-epicatechin, *Int. J. Cancer* 123, 2008, 2446-2455, https://doi.org/10.1002/jic.23813.
- [140] H. Tanaka, M. Yamanouchi, H. Miyoshi, K. Hirotsu, H. Tachibana and T. Takahashi, Solid-phase synthesis of a combinatorial methylated (±)- epigallocatechin gallate library and the growth-inhibitory effects of these compounds on melanoma B16 cells, Chem.—An Asian J. 5, 2010, 2231-2248, https://doi.org/10.1002/asia.201000372.
- [141] J. Xie, J.-P. Yun, Y.-N. Yang, F. Hua, X.-W. Zhang, H. Lin, X.-X. Lv, K. Li, P.-C. Zhang and Z.-W. Hu, A novel ECG analog 4-(S)-(2,4,6-trimethylthiobenzyl)-epigallocatechin gallate selectively induces apoptosis of B16-F10 melanoma via activation of autophagy and ROS, Sci. Rep. 7, 2017, 42194, https://doi.org/10.1038/srep42194.
- [142] F. Lemarié, C.W. Chang, D.R. Blatchford, R. Amor, G. Norris, L. Tetley, G. Mcconnell and C. Dufès, Antitumor activity of the tea polyphenol epigallocatechin-3-gallate encapsulated in targeted vesicles after intravenous administration, *Nanomedicine* 3, 2013, 181-192, https://doi.org/10.2217/nnm.12.83.
- [143] C.-C. Chen, D.-S. Hsieh, K.-J. Huang, Y.-L. Chan, P.-D. Hong, M.-K. Yeh and C.-J. Wu, Improving anticancer efficacy of (-)-epigallocatechin-3-gallate gold nanoparticles in murine B16F10 melanoma cells, *Drug Des. Dev*.

  Ther. 8, 2014, 459-474, https://doi.org/10.2147/DDDT.S58414.
- [144] I.A. Siddiqui, D.J. Bharali, M. Nihal, V.M. Adhami, N. Khan, J.C. Chamcheu, M.I. Khan, S. Shabana, S.A. Mousa and H. Mukhtar, Excellent anti-proliferative and pro-apoptotic effects of (-)-epigallocatechin-3-gallate encapsulated in chitosan nanoparticles on human melanoma cell growth both in vitro and in vivo, *Nanomeo* in vivo, *Nanomeo* in vivo, *Nanomeo* in vivo, *Nanotechnology, Nanotechnol.* Biol. Med. 10, 2014, 1619-1626, https://doi.org/10.1016/j.nano.2014.05.007.
- [145] B. Liao, H. Ying, C. Yu, Z. Fan, W. Zhang, J. Shi, H. Ying, N. Ravichandran, Y. Xu, J. Yin, Y. Jiang and Q. Du, (–)-Epigallocatechin gallate (EGCG)-nanoethosomes as a transdermal delivery system for docetaxel to treat implanted human melanoma cell tumors in mice, *Int. J. Pharm.* 512, 2016, 22–31, https://doi.org/10.1016/j.ijpharm.2016.08.038.

- [146] M. Pulido-Moran, J. Moreno-Fernandez, C. Ramirez-Tortosa and M.C. Ramirez-Tortosa, Curcumin and health, Molecules 21, 2016, 264, https://doi.org/10.3390/molecules21030264.
- [147] A.B. Kunnumakkara, D. Bordoloi, G. Padmavathi, J. Monisha, N.K. Roy, S. Prasad and B.B. Aggarwal, Curcumin, the golden nutraceutical: multitargeting for multiple chronic diseases, *Br. J. Pharmacol.* 174, 2017, 1325–1348, https://doi.org/10.1111/bph.13621.
- [148] J.A. Bush, K.J.J. Cheung and G. Li, Curcumin induces apoptosis in human melanoma cells through a Fas receptor/caspase-8 pathway independent of p53, Exp. Cell Res. 271, 2001, 305-314, https://doi.org/10.1006/excr.2001.5381.
- [149] J. Odot, P. Albert, A. Carlier, M. Tarpin, J. Devy and C. Madoulet, In vitro and in vivo anti-tumoral effect of curcumin against melanoma cells, Int. J. Cancer 111, 2004, 381-387, https://doi.org/10.1002/ijc.20160.
- [150] M.A. Bill, C. Bakan, D.M. Benson, J. Fuchs, G. Young and G.B. Lesinski, Curcumin induces proapoptotic effects against human melanoma cells and modulates the cellular response to immunotherapeutic cytokines, *Mol. Cancer Ther.* 8, 2009, 2726–2735, https://doi.org/10.1158/1535-7163.MCT-09-0377.
- [151] C. Lu, E. Song, D.N. Hu, M. Chen, C. Xue, R. Rosen and S.A. McCormick, Curcumin induces cell death in human uveal melanoma cells through mitochondrial pathway, *Curr. Eye Res.* 35, 2010, 352-360, https://doi.org/10.3109/02713680903521944.
- [152] A. Abusnina, T. Keravis, I. Yougbaré, C. Bronner and C. Lugnier, Anti-proliferative effect of curcumin on melanoma cells is mediated by PDE1A inhibition that regulates the epigenetic integrator UHRF1, Mol. Nutr. Food Res. 55, 2011, 1677-1689, https://doi.org/10.1002/mnfr.201100307.
- [153] L.-X. Chen, Y.-J. He, S.-Z. Zhao, J.-G. Wu, J.-T. Wang, L.-M. Zhu, T.-T. Lin, B.-C. Sun and X.-R. Li, Inhibition of tumor growth and vasculogenic mimicry by curcumin through down-regulation of the EphA2/PI3K/MMP pathway in a murine choroidal melanoma model, *Cancer Biol. Ther.* 11, 2011, 229–235, https://doi.org/10.4161/cbt.11.2.13842.
- [154] Y. Qiu, T. Yu, W. Wang, K. Pan, D. Shi and H. Sun, Curcumin-induced melanoma cell death is associated with mitochondrial permeability transition pore (mPTP) opening, *Biochem. Biophys. Res. Commun.* 448, 2014, 15–21, https://doi.org/10.1016/j.bbrc.2014.04.024.
- [155] A.J. Jiang, G. Jiang, L.T. Li and J.N. Zheng, Curcumin induces apoptosis through mitochondrial pathway and caspases activation in human melanoma cells, *Mol. Biol. Rep.* 42, 2015, 267-275, https://doi.org/10.1007/s11033-014-3769-2.
- [156] M. Zheng, S. Ekmekcioglu, E.T. Walch, C.-H. Tang and E.A. Grimm, Inhibition of nuclear factor-κB and nitric oxide by curcumin induces G2/M cell cycle arrest and apoptosis in human melanoma cells, *Melanoma Res.* 14, 2004, 165–171, https://doi.org/10.1097/01.cmr.0000129374.76399.19.
- [157] D.R. Siwak, S. Shishodia, B.B. Aggarwal and R. Kurzrock, Curcumin-induced antiproliferative and proapoptotic effects in melanoma cells are associated with suppression of IkB kinase and nuclear factor kB activity and are independent of the B-Raf/mitogen-activated/ extracellular signal-regulated protein kinase pathway and the Akt pathway, Cancer 104, 2005, 879-890, https://doi.org/10.1002/cncr.21216.
- [158] Y.E. Marín, B.A. Wall, S. Wang, J. Namkoong, J.J. Martino, J. Suh, H.J. Lee, A.B. Rabson, C.S. Yang, S. Chen and J.H. Ryu, Curcumin downregulates the constitutive activity of NF-kB and induces apoptosis in novel mouse melanoma cells, *Melanoma Res.* 17, 2007, 274–283, https://doi.org/10.1097/CMR.0b013e3282ed3d0e.
- [159] Y.P. Zhang, Y.Q. Li, Y.T. Lv and J.M. Wang, Effect of curcumin on the proliferation, apoptosis, migration, and invasion of human melanoma A375 cells, *Genet. Mol. Res.* 14, 2015, 1056-1067, https://doi.org/10.4238/2015.February.6.9.
- [160] G. Zhao, X. Han, S. Zheng, Z. Li, Y. Sha, J. Ni, Z. Sun, S. Qiao and Z. Song, Curcumin induces autophagy, inhibits proliferation and invasion by downregulating AKT/mTOR signaling pathway in human melanoma cells, Oncol. Rep. 35, 2016, 1065–1074, https://doi.org/10.3892/or.2015.4413.
- [161] J. Bakhshi, L. Weinstein, K.S. Poksay, B. Nishinaga, D.E. Bredesen and R.V. Rao, Coupling endoplasmic reticulum stress to the cell death program in mouse melanoma cells: effect of curcumin, *Apoptosis* 13, 2008, 904–914, https://doi.org/10.1007/s10495-008-0221-x.
- [162] T. Yu, J. Ji and Y.L. Guo, MST1 activation by curcumin mediates JNK activation, Foxo3a nuclear translocation and apoptosis in melanoma cells, *Biochem. Biophys. Res. Commun.* 441, 2013, 53–58, https://doi.org/10.1016/j.bbrc.2013.10.008.

- [163] W. Liao, W. Xiang, F.F. Wang, R. Wang and Y. Ding, Curcumin inhibited growth of human melanoma A375 cells via inciting oxidative stress, *Biomed. Pharmacother.* 95, 2017, 1177-1186, https://doi.org/10.1016/j.biopha.2017.09.026.
- [164] A. Kocyigit and E.M. Guler, Curcumin induce DNA damage and apoptosis through generation of reactive oxygen species and reducing mitochondrial membrane potential in melanoma cancer cells, *Cell. Mol. Biol.* (Noisy-Le-Grand). 63, 2017, 97-105, https://doi.org/10.14715/cmb/2017.63.11.17.
- [165] S. Ray, N. Chattopadhyay, A. Mitra, M. Siddiqi and A. Chatterjee, Curcumin exhibits antimetastatic properties by modulating integrin receptors, collagenase activity, and expression of Nm23 and E-cadherin, J. Environ. Pathol. Toxicol. Oncol. 22, 2003, 49-58 http://www.ncbi.nlm.nih.gov/pubmed/12678405.
- [166] A. Banerji, J. Chakrabarti, A. Mitra and A. Chatterjee, Effect of curcumin on gelatinase a (MMP-2) activity in B16F10 melanoma cells, Cancer Lett. 211, 2004, 234-242, https://doi.org/10.1016/j.canlet.2004.02.007.
- [167] L. Wang, Y. Shen, R. Song, Y. Sun, J. Xu and Q. Xu, An Anticancer Effect of Curcumin Mediated by Down-Regulating Phosphatase of Regenerating Liver-3 Expression on Highly Metastatic Melanoma Canticancer effect of Curcumin mediated by down-regulating phosphatase of regenerating Liver-3 expression on highly metastatic melanoma cells, Mol. Pharmacol. 76, 2009, 1238-1245, https://doi.org/10.1124/mol.109.059105.
- [168] K. Shimada, K. Ushijima, C. Suzuki, M. Horiguchi, H. Ando, T. Akita, M. Shimamura, J. Fujii, C. Yamashita and A. Fujimura, Pulmonary administration of curcumin inhibits B16F10 melanoma lung metastasis and invasion in mice, Cancer Chemother. Pharmacol. 82, 2018, 265-273, https://doi.org/10.1007/s00280-018-3616-6.
- [169] T. Yu, J. Li and H. Sun, C6 ceramide potentiates curcumin-induced cell death and apoptosis in melanoma cell lines in vitro, Cancer Chemother. Pharmacol. 66, 2010, 999-1003, https://doi.org/10.1007/s00280-010-1374-1.
- [170] S. Barui, S. Saha, V. Yakati and A. Chaudhuri, Systemic Codelivery of a Homoserine Derived Ceramide Analogue and Curcumin to Tumor Vasculature Inhibits Mouse Tumor Godelivery of a homoserine derived ceramide analogue and curcumin to tumor vasculature inhibits mouse tumor growth, Mol. Pharm. 13, 2016, 404-419, https://doi.org/10.1021/acs.molpharmaceut.5b00644.
- [171] S.J. Chatterjee and S. Pandey, Chemo-resistant melanoma sensitized by tamoxifen to low dose curcumin treatment through induction of apoptosis and autophagy, *Cancer Biol. Ther.* 11, 2011, 216-228, https://doi.org/10.4161/cbt.11.2.13798.
- [172] G.-M. Jiang, W.-Y. Xie, H.-S. Wang, J. Du, B.-P. Wu, W. Xu, H.-F. Liu, P. Xiao, Z.-G. Liu, H.-Y. Li, S.-Q. Liu, W.-J. Yin, Q.-G. Zhang, J.-P. Liang and H.-J. Huang, Curcumin combined with FAPαc vaccine elicits effective antitumor response by targeting indolamine-2,3-dioxygenase and inhibiting EMT induced by TNF-α in melanoma, *Oncotarget* -6, 2015, 25932-25942, https://doi.org/10.18632/oncotarget.4577.
- [173] T. Yu, J. Li, Y. Qiu and H. Sun, 1-phenyl-2-decanoylamino-3-morpholino-1-propanol (PDMP) facilitates curcumin-induced melanoma cell apoptosis by enhancing ceramide accumulation, JNK activation, and inhibiting PI3K/AKT activation, Mol. Cell. Biochem. 361, 2012, 47-54, https://doi.org/10.1007/s11010-011-1086-9.
- [174] T. Yu, C. Chen, Y. Sun, H. Sun, T.H. Li, J. Meng and X. Shi, ABT-737 sensitizes curcumin-induced anti-melanoma cell activity through facilitating mPTP death pathway, *Biochem. Biophys. Res. Commun.* 464, 2015, 286–291, https://doi.org/10.1016/j.bbrc.2015.06.144.
- [175] R.-J. Shiau, J.-Y. Wu, S.-J. Chiou and Y.-D. Wen, Effects of curcumin on nitrosyl-iron complex-mediated DNA cleavage and cytotoxicity, Planta Med. 78, 2012, 1342-1350, https://doi.org/10.1055/s-0032-1315020.
- [176] J. Chen, L. Li, J. Su, B. Li, T. Chen and Y.S. Wong, Synergistic apoptosis-inducing effects on A375 human melanoma cells of natural borneol and curcumin, *PLoS One* 9, 2014, , e101277https://doi.org/10.1371/journal.pone.0101277.
- [177] T. Niu, Y. Tian, Z. Mei and G. Guo, Inhibition of autophagy enhances curcumin united light irradiation-induced oxidative stress and tumor growth suppression in human melanoma cells, Sci. Rep. 6, 2016, 31383, https://doi.org/10.1038/srep31383.
- [178] F. Faião-Flores, J.A.Q. Suarez, S.S. Maria-Engler, V. Soto-Cerrato, R. Pérez-Tomás and D.A. Maria, The curcumin analog DM-1 induces apoptotic cell death in melanoma, *Tumor Biol.* 34, 2013, 1119-1129, https://doi.org/10.1007/s13277-013-0653-y.
- [179] F. Faião-Flores, J.A.Q. Suarez, V. Soto-Cerrato, M. Espona-Fiedler, R. Pérez-Tomás and D.A. Maria, Bcl-2 family proteins and cytoskeleton changes involved in DM-1 cytotoxic effect on melanoma cells, *Tumor Biol.* 34, 2013, 1235–1243, https://doi.org/10.1007/s13277-013-0666-6.

- [180] F. Faião-Flores, J.A.Q. Suarez, A.C. Fruet, S.S. Maria-Engler, P.C. Pardi and D.A. Maria, Curcumin analog DM-1 in monotherapy or combinatory treatment with dacarbazine as a strategy to inhibit in vivo melanoma progression, *PLoS One* 10, 2015, , e0118702https://doi.org/10.1371/journal.pone.0118702.
- [181] É.A. de Oliveira, D.S. de Lima, L.E. Cardozo, G.F. de Souza, N. de Souza, D.K. Alves-Fernandes, F. Faião-Flores, J.A.P. Quincoces, S.B. de M. Barros, H.I. Nakaya, G. Monteiro and S.S. Maria-Engler, Toxicogenomic and bioinformatics platforms to identify key molecular mechanisms of a curcumin-analogue DM-1 toxicity in melanoma cells, *Pharmacol. Res.* 125, 2017, 178-187, https://doi.org/10.1016/j.phrs.2017.08.018.
- [182] M. Pisano, G. Pagnan, M.A. Dettori, S. Cossu, I. Caffa, I. Sassu, L. Emionite, D. Fabbri, M. Cilli, F. Pastorino, G. Palmieri, G. Delogu, M. Ponzoni and C. Rozzo, Enhanced anti-tumor activity of a new curcumin-related compound against melanoma and neuroblastoma cells, *Mol. Cancer* 9, 2010, 137, https://doi.org/10.1186/1476-4598-9-137.
- [183] C. Rozzo, M. Fanciulli, C. Fraumene, A. Corrias, T. Cubeddu, I. Sassu, S. Cossu, V. Nieddu, G. Galleri, E. Azara, M.A. Dettori, D. Fabbri, G. Palmieri and M. Pisano, Molecular changes induced by the curcumin analogue D6 in human melanoma cells, Mol. Cancer 12, 2013, 37, https://doi.org/10.1186/1476-4598-12-37.
- [184] M. Pisano, A. Palomba, A. Tanca, D. Pagnozzi, S. Uzzau, M.F. Addis, M.A. Dettori, D. Fabbri, G. Palmieri and C. Rozzo, Protein expression changes induced in a malignant melanoma cell line by the curcumin analogue compound D6, BMC Cancer 16, 2016, 317, https://doi.org/10.1186/s12885-016-2362-6.
- [185] M.A. Bill, J.R. Fuchs, C. Li, J. Yui, C. Bakan, D.M. Benson, E.B. Schwartz, D. Abdelhamid, J. Lin, D.G. Hoyt, S.L. Fossey, G.S. Young, W.E. Carson, P.K. Li and G.B. Lesinski, The small molecule curcumin analog FLLL32 induces apoptosis in melanoma cells via STAT3 inhibition and retains the cellular response to cytokines with anti-tumor activity, *Mol. Cancer* 9, 2010, 165, https://doi.org/10.1186/1476-4598-9-165.
- [186] M.A. Bill, C. Nicholas, T.A. Mace, J.P. Etter, C. Li, E.B. Schwartz, J.R. Fuchs, G.S. Young, L. Lin, J. Lin, L. He, M. Phelps, P.K. Li and G.B. Lesinski, Structurally modified curcumin analogs inhibit STAT3 phosphorylation and promote apoptosis of human renal cell carcinoma and melanoma cell lines, *PLoS One* 7, 2012, , e40724https://doi.org/10.1371/journal.pone.0040724.
- [187] C.H. Yang, J. Yue, M. Sims and L.M. Pfeffer, The Curcumin Analog EF24 Tcurcumin analog EF24 targets NF-kB and miRNA-21, and Has Potent Anticancer Activity In Vitro and In VivoPLoS One. has potent anticancer activity in vitro and in vivo, PLoS One 8, 2013, , e71130https://doi.org/10.1371/journal.pone.0071130.
- [188] P. Zhang, H. Bai, G. Liu, H. Wang, F. Chen, B. Zhang, P. Zeng, C. Wu, C. Peng, C. Huang, Y. Song and E. Song, MicroRNA-33b, upregulated by EF24, a curcumin analog, suppresses the epithelial-to-mesenchymal transition (EMT) and migratory potential of melanoma cells by targeting HMGA2, *Toxicol. Lett.* 234, 2015, 151-161, https://doi.org/10.1016/j.toxlet.2015.02.018.
- [189] F. Schmitt, M. Gold, G. Begemann, I. Andronache, B. Biersack and R. Schobert, Fluoro and pentafluorothio analogs of the antitumoral curcuminoid EF24 with superior antiangiogenic and vascular-disruptive effects, Bioorganie, Med. Chem. 25, 2017, 4894-4903, https://doi.org/10.1016/j.bmc.2017.07.039.
- [190] A. Arezki, G.G. Chabot, L. Quentin, D. Scherman, G. Jaouen and E. Brulé, Synthesis and biological evaluation of novel ferrocenyl curcuminoid derivatives, *Medchemcomm* 2, 2011, 190-195, https://doi.org/10.1039/c0md00231c.
- [191] T. Hosoya, A. Nakata, F. Yamasaki, F. Abas, K. Shaari, N.H. Lajis and H. Morita, Curcumin-like diarylpentanoid analogues as melanogenesis inhibitors, *J. Nat. Med.* 66, 2012, 166–176, https://doi.org/10.1007/s11418-011-0568-0.
- [192] P.V. Leyon and G. Kuttan, Studies on the role of some synthetic curcuminoid derivatives in the inhibition of tumour specific angiogenesis,, *J. Exp. Clin. Cancer Res.* 22, 2003, 77-83 http://www.ncbi.nlm.nih.gov/pubmed/12725326.
- [193] Z. Ma, A. Haddadi, O. Molavi, A. Lavasanifar, R. Lai and J. Samuel, Micelles of poly(ethylene oxide)-b -poly(ε-caprolactone) as vehicles for the solubilization, stabilization, and controlled delivery of curcumin, *J. Biomed Mater. Res. Part A* 36A, 2008, 300–310, https://doi.org/10.1002/jbm.a.31584.
- [194] L. Zhao, C. Yang, J. Dou, Y. Xi, H. Lou and G. Zhai, Development of RGD-functionalized PEG-PLA micelles for delivery of curcumin, J. Biomed. Nanotechnol. 11, 2015, 436-446, https://doi.org/10.1166/jbn.2015.1919.
- [195] Y. Lu, L. Miao, Y. Wang, Z. Xu, Y. Zhao, Y. Shen, G. Xiang and L. Huang, Curcumin micelles remodel tumor microenvironment and enhance vaccine activity in an advanced melanoma model, *Mol. Ther.* 24, 2016, 364–374, https://doi.org/10.1038/mt.2015.165.

- [196] P. Kumari, O.S. Muddineti, S.V.K. Rompicharla, P. Ghanta, A.K. B B N, B. Ghosh and S. Biswas, Cholesterol-conjugated poly(D, L-lactide)-based micelles as a nanocarrier system for effective delivery of curcumin in cancer therapy, *Drug Deliv.* 24, 2017, 209-223, https://doi.org/10.1080/10717544.2016.1245365.
- [197] B. Wang, X. Liu, Y. Teng, T. Yu, J. Chen, Y. Hu, N. Liu, L. Zhang and Y. Shen, Improving anti-melanoma effect of curcumin by biodegradable nanoparticles, *Oncotarget* -8, 2017, 108624–108642, https://doi.org/10.18632/oncotarget.20585.
- [198] O.S. Muddineti, P. Kumari, E. Ray, B. Ghosh and S. Biswas, Curcumin-loaded chitosan-cholesterol micelles: Evaluation in monolayers and 3D cancer spheroid model, Nanomedicine 2017, https://doi.org/10.2217/nnm-2017-0036.
- [199] O.S. Muddineti, A. Vanaparthi, S.V.K. Rompicharla, P. Kumari, B. Ghosh and S. Biswas, Cholesterol and vitamin E-conjugated PEGylated polymeric micelles for efficient delivery and enhanced anticancer activity of curcumin: evaluation in 2D monolayers and 3D spheroids, *Artif. Cells* Nanomedicine Nanomed. Biotechnol. 2018, 1-14, https://doi.org/10.1080/21691401.2018.1435551.
- [200] Y. Chen, Q. Wu, Z. Zhang, L. Yuan, X. Liu and L. Zhou, Preparation of curcumin-loaded liposomes and evaluation of their skin permeation and pharmacodynamics, *Molecules* 17, 2012, 5972-5987, https://doi.org/10.3390/molecules17055972.
- [201] A. Karewicz, D. Bielska, A. Loboda, B. Gzyl-Malcher, J. Bednar, A. Jozkowicz, J. Dulak and M. Nowakowska, Curcumin-containing liposomes stabilized by thin layers of chitosan derivatives, *Colloids Surfaces B Biointerfaces* 109, 2013, 307–316, https://doi.org/10.1016/j.colsurfb.2013.03.059.
- [202] G. Mondal, S. Barui, S. Saha and A. Chaudhuri, Tumor growth inhibition through targeting liposomally bound curcumin to tumor vasculature, *J. Control. Release* 172, 2013, 832-840, https://doi.org/10.1016/j.jconrel.2013.08.302.
- [203] H.B. Ruttala and Y.T. Ko, Liposomal co-delivery of curcumin and albumin/paclitaxel nanoparticle for enhanced synergistic antitumor efficacy, *Colloids Surf. B. Biointerfaces. Bioin*
- [204] A. Jose, K.M. Ninave, S.K. Gade, S. Labala and V.V.K. Venuganti, Effective Skin Cancer Treatment by Topical Co-delivery of Curcumin and STAT3 siRNA Using Cationic LipesomesAAPS PharmSciTech.skin Cancer treatment by topical Co-delivery of curcumin and STAT3 siRNA using cationic liposomes, AAPS PharmSciTech 19, 2017, 166-175, https://doi.org/10.1208/s12249-017-0833-y.
- [205] L. Mazzarino, L.F.C. Silva, J.C. Curta, M.A. Licínio, A. Costa, L.K. Pacheco, J.M. Siqueira, J. Montanari, E. Romero, J. Assreuy, M.C. Santos-Silva and E. Lemos-Senna, Curcumin-Loaded Lipid and Polymeric Nanocapsules

  Stabilized by Nonionic Surfactants: An In Vitro and In Vivo Antitumor Activity on B16-F10 Melanoma and Macrophage Uptake Comparative Sloaded lipid and polymeric nanocapsules stabilized by nonionic surfactants: an in vitro
  and in vivo antitumor activity on B16-F10 melanoma and macrophage uptake comparative study, J. Biomed. Nanotechnol. 7, 2011, 406-414, https://doi.org/10.1166/jbn.2011.1296.
- [206] F.F. de Souza, M.C. dos Santos, D.C.S. dos Passos, E.C. de Oliveira Lima and L.A. Guillo, Curcumin Associated Magnetite Nanoparticles Inhibit In Vitro Melanoma Cell Gassociated magnetite nanoparticles inhibit in vitro melanoma cell growth, J. Nanosci. Nanotechnol. 11, 2011, 7603-7610, https://doi.org/10.1166/jnn.2011.5124.
- [207] V. Paunovic, B. Ristic, Z. Markovic, B. Todorovic-Markovic, M. Kosic, J. Prekodravac, T. Kravic-Stevovic, T. Martinovic, M. Micusik, Z. Spitalsky, V. Trajkovic and L. Harhaji-Trajkovic, c-Jun N-terminal kinase-dependent apoptotic photocytotoxicity of solvent exchange-prepared curcumin nanoparticles, *Biomed. Microdevices.* 18, 2016, 37, https://doi.org/10.1007/s10544-016-0062-2.
- [208] O.S. Muddineti, P. Kumari, S. Ajjarapu, P.M. Lakhani, R. Bahl, B. Ghosh and S. Biswas, Xanthan gum stabilized PEGylated gold nanoparticles for improved delivery of curcumin in cancer, *Nanotechnology* -27, 2016, , 325101https://doi.org/10.1088/0957-4484/27/32/325101.
- [209] K. Das, S. Nimushakavi, A. Chaudhuri and P.K. Das, An Integrin Targeting RGDK-Tagged Nanocarrier: Anticancer Efficacy of Loaded CurcuminChemMedChem. integrin-targeting RGDK-Tagged nanocarrier: anticancer efficacy of Loaded curcumin, ChemMedChem 12, 2017, 738-750, https://doi.org/10.1002/cmdc.201700085.
- [210] S.P. Singh, S.B. Alvi, D.B. Pemmaraju, A.D. Singh, S.V. Manda, R. Srivastava and A.K. Rengan, NIR triggered liposome gold nanoparticles entrapping curcumin as in situ adjuvant for photothermal treatment of skin cancer, *Int. J. Biol. Macromol.* 110, 2018, 375–382, https://doi.org/10.1016/j.ijbiomac.2017.11.163.
- [211] F. Tavakoli, R. Jahanban-Esfahlan, K. Seidi, M. Jabbari, R. Behzadi, Y. Pilehvar-Soltanahmadi and N. Zarghami, Effects of nano-encapsulated curcumin-chrysin on telomerase, MMPs and TIMPs gene expression in mouse

- B16F10 melanoma tumour model, Artif. Cells Nanomedicine Nanomed. Biotechnol. 46, 2018, 75-86, https://doi.org/10.1080/21691401.2018.1452021.
- [212] F. Rahimi-Moghaddam, N. Azarpira and N. Sattarahmady, Evaluation of a nanocomposite of PEG-curcumin-gold nanoparticles as a near-infrared photothermal agent: an in vitro and animal model investigation, *Laser Med. Sci.* 33, 2018, 1769–1779, https://doi.org/10.1007/s10103-018-2538-1.
- [213] L.E.A. de Camargo, D. Brustolin Ludwig, T.T. Tominaga, B. Carletto, G.M. Favero, R.M. Mainardes and N.M. Khalil, Bovine serum albumin nanoparticles improve the antitumour activity of curcumin in a murine melanoma model, J. Microencapsul. 35, 2018, 467-474, https://doi.org/10.1080/02652048.2018.1526340.
- [214] S. Datta, S.K. Misra, M.L. Saha, N. Lahiri, J. Louie, D. Pan and P.J. Stang, Orthogonal self-assembly of an organoplatinum(II) metallacycle and cucurbit[8]uril that delivers curcumin to cancer cells, *Proc. Natl. Acad. Sci.* 115, 2018, 8087-8092, https://doi.org/10.1073/pnas.1803800115.
- [215] S. Guerrero, M. Inostroza-Riquelme, P. Contreras-Orellana, V. Diaz-Garcia, P. Lara, A. Vivanco-Palma, A. Cárdenas, V. Miranda, P. Robert, L. Leyton, M.J. Kogan, A.F.G. Quest and F. Oyarzun-Ampuero, Curcumin-loaded nanoemulsion: a new safe and effective formulation to prevent tumor reincidence and metastasis, *Nanoscale* 10, 2018, 22612–22622, https://doi.org/10.1039/C8NR06173D.
- [216] M. Mimeault and S.K. Batra, Potential applications of curcumin and its novel synthetic analogs and nanotechnology-based formulations in cancer prevention and therapy, *Chin. Med.* 6, 2011, 31, https://doi.org/10.1186/1749-8546-6-31.
- [217] S. Mangalathillam, N.S. Rejinold, A. Nair, V.K. Lakshmanan, S.V. Nair and R. Jayakumar, Curcumin loaded chitin nanogels for skin cancer treatment via the transdermal route, *Nanoscale* 4, 2012, 239-250, https://doi.org/10.1039/c1nr11271f.
- [218] D. Michel, J.M. Chitanda, R. Balogh, P. Yang, J. Singh, U. Das, A. El-Aneed, J. Dimmock, R. Verrall and I. Badea, Design and evaluation of cyclodextrin-based delivery systems to incorporate poorly soluble curcumin analogs for the treatment of melanoma, *Eur. J. Pharm. Biopharm.* 81, 2012, 548-556, https://doi.org/10.1016/j.ejpb.2012.03.016.
- [219] S. Alam, J.J. Panda and V.S. Chauhan, Novel dipeptide nanoparticles for effective curcumin delivery, Int. J. Nanomedicine. 7, 2012, 4207-4222, https://doi.org/10.2147/IJN.S33015.
- [220] M. Cui, D.J. Naczynski, M. Zevon, C.K. Griffith, L. Sheihet, I. Poventud-Fuentes, S. Chen, C.M. Roth and P.V. Moghe, Multifunctional albumin nanoparticles as combination drug carriers for intra-tumoral chemotherapy, *Adv. Healthc. Mater.* 2, 2013, 1236–1245, https://doi.org/10.1002/adhm.201200467.
- [221] Y. Sun, L. Du, Y. Liu, X. Li, M. Li, Y. Jin and X. Qian, Transdermal delivery of the in situ hydrogels of curcumin and its inclusion complexes of hydroxypropyl-β-cyclodextrin for melanoma treatment, *Int. J. Pharm.* 469, 2014, 31–39, https://doi.org/10.1016/j.ijpharm.2014.04.039.
- [222] M. Poorghorban, U. Das, O. Alaidi, J.M. Chitanda, D. Michel, J. Dimmock, R. Verrall, P. Grochulski and I. Badea, Characterization of the host-guest complex of a curcumin analog with β-cyclodextrin and β-cyclodextrin-gemini surfactant and evaluation of its anticancer activity, *Int. J. Nanomedicine*. 10, 2015, 503-515, https://doi.org/10.2147/IJN.S70828.
- [223] G. Loch-Neckel, L. Santos-Bubniak, L. Mazzarino, A.V. Jacques, B. Moccelin, M.C. Santos-Silva and E. Lemos-Senna, Orally Administered Chitosan Coated Polycaprolactone Nanoparticles Containing Curcumin Attenuate

  Metastatic Melanoma in the Ladministered chitosan-coated polycaprolactone nanoparticles containing curcumin attenuate metastatic melanoma in the lungs, J. Pharm. Sci. 104, 2015, 3524–3534,

  https://doi.org/10.1002/jps.24548.
- [224] P. Langone, P.R. Debata, S. Dolai, G.M. Curcio, J.D.R. Inigo, K. Raja and P. Banerjee, Coupling to a cancer cell-specific antibody potentiates tumoricidal properties of curcumin, *Int. J. Cancer* 131, 2012, E569-E578, https://doi.org/10.1002/jic.26479.
- [225] A. Rauf, M. Imran, M.S. Butt, M. Nadeem, D.G. Peters and M.S. Mubarak, Resveratrol as an anti-cancer agent: Aa review, *Crit. Rev. Food Sci. Nutr.* 58, 2018, 1428-1447, https://doi.org/10.1080/10408398.2016.1263597.
- [226] M. Larrosa, F.A. Tomás-Barberán and J.C. Espín, Grape polyphenol resveratrol and the related molecule 4-hydroxystilbene induce growth inhibition, apoptosis, S-phase arrest, and upregulation of cyclins A, E, and B1 in human SK-Mel-28 melanoma cells, *J. Agric. Food Chem.* 51, 2003, 4576-4584, https://doi.org/10.1021/jf030073c.
- [227] T. Hsieh, Z. Wang, C.V. Hamby and J.M. Wu, Inhibition of melanoma cell proliferation by resveratrol is correlated with upregulation of quinone reductase 2 and p53, Biochem. Biophys. Res. Commun. 334, 2005,

- 223-230, https://doi.org/10.1016/j.bbrc.2005.06.073.
- [228] G.W. Osmond, E.M. Masko, D.S. Tyler, S.J. Freedland and S. Pizzo, In vitro and in vivo evaluation of resveratrol and 3,5-dihydroxy-4′- acetoxy-trans-stilbene in the treatment of human prostate carcinoma and melanoma, J. Surg. Res. 179, 2013, e141-e148, https://doi.org/10.1016/j.jss.2012.02.057.
- [229] M. Lei, Y. Dong, C. Sun and X. Zhang, Resveratrol inhibits proliferation, promotes differentiation and melanogenesis in HT-144 melanoma cells through inhibition of MEK/ERK kinase pathway, *Microb. Pathog.* 111, 2017, 410-413, https://doi.org/10.1016/j.micpath.2017.09.029.
- [230] F. Wu and L. Cui, Resveratrol suppresses melanoma by inhibiting NF-kB/miR-221 and inducing TFG expression, Arch. Dermatol. Res. 309, 2017, 823-831, https://doi.org/10.1007/s00403-017-1784-6.
- [231] R.M. Niles, M. McFarland, M.B. Weimer, A. Redkar, Y.M. Fu and G.G. Meadows, Resveratrol is a potent inducer of apoptosis in human melanoma cells, *Cancer Lett.* 190, 2003, 157-163, https://doi.org/10.1016/S0304-3835(02)00676-6.
- [232] M.P. Fuggetta, S. D'Atri, G. Lanzilli, M. Tricarico, E. Cannavò, G. Zambruno, R. Falchetti and G. Ravagnan, In vitro antitumour activity of resveratrol in human melanoma cells sensitive or resistant to temozolomide, Melanoma Res. 14, 2004, 189-196, https://doi.org/10.1097/01.cmr.0000130007.54508.b2.
- [233] P.R. Van Ginkel, S.R. Darjatmoko, D. Sareen, L. Subramanian, S. Bhattacharya, M.J. Lindstrom, D.M. Albert and A.S. Polans, Resveratrol inhibits uveal melanoma tumor growth via early mitochondrial dysfunction, *Investig. Ophthalmol. Vis. Sci.* 49, 2008, 1299–1306, https://doi.org/10.1167/iovs.07-1233.
- [234] G. Gatouillat, E. Balasse, D. Joseph-Pietras, H. Morjani and C. Madoulet, Resveratrol induces cell-cycle disruption and apoptosis in chemoresistant B16 melanoma, *J. Cell. Biochem.* 110, 2010, 893-902, https://doi.org/10.1002/jcb.22601.
- [235] S. Habibie, S. Yokoyama, S. Abdelhamed, H. Awale, Y. Sakurai and I. Hayakawa, Saiki, Survivin suppression through STAT3/β-catenin is essential for resveratrol-induced melanoma apoptosis, *Int. J. Oncol.* 45, 2014, 895–901, https://doi.org/10.3892/ijo.2014.2480.
- [236] M. Wang, T. Yu, C. Zhu, H. Sun, Y. Qiu, X. Zhu and J. Li, Resveratrol triggers protective autophagy through the ceramide/Akt/mTOR pathway in melanoma B16 cells, *Nutr. Cancer* 66, 2014, 435-440, https://doi.org/10.1080/01635581.2013.878738.
- [237] Z. Wu, B. Liu, E. Cailing, J. Liu, Q. Zhang, J. Liu, N. Chen, R. Chen and R. Zhu, Resveratrol inhibits the proliferation of human melanoma cells by inducing G1/S cell cycle arrest and apoptosis, *Mol. Med. Rep.* 11, 2015, 400-404, https://doi.org/10.3892/mmr.2014.2716.
- [238] J.-R. Heo, S.-M. Kim, K.-A. Hwang, J.-H. Kang and K.-C. Choi, Resveratrol induced reactive oxygen species and endoplasmic reticulum stress-mediated apoptosis, and cell cycle arrest in the A375SM malignant melanoma cell line, Int. J. Mol. Med. 42, 2018, 1427–1435, https://doi.org/10.3892/ijmm.2018.3732.
- [239] H. Zhao, L. Han, Y. Jian, Y. Ma, W. Yan, X. Chen, H. Xu and L. Li, Resveratrol induces apoptosis in human melanoma cell through negatively regulating Erk/PKM2/Bcl-2 axis, Onco. Targets. Ther. 11, 2018, 8995-9006, https://doi.org/10.2147/OTT.S186247.
- [240] S. Yang and F.L.J. Meyskens, Alterations in activating protein 1 composition correlate with phenotypic differentiation changes induced by resveratrol in human melanoma, *Mol. Pharmacol.* 67, 2005, 298-308, https://doi.org/10.1124/mol.104.006023.
- [241] H. Satooka and I. Kubo, Resveratrol as a kcat type inhibitor for tyrosinase: Potentiated melanogenesis inhibitor, Bioorg. Med. Chem. 20, 2012, 1090-1099, https://doi.org/10.1016/j.bmc.2011.11.030.
- [242] Y.-J. Chen, Y.-Y. Chen, Y.-F. Lin, H.-Y. Hu and H.-F. Liao, Resveratrol Inhibits Alpha Melanocyte Stimulating Hormone Signaling, Viability, and Invasiveness in Melanoma Cells, Evidence Based Cinhibits alpha-melanocyte-Stimulating hormone signaling, viability, and invasiveness in melanoma cells, evidence-based complement, Altern. Med. 2013, 2013, 1-8, https://doi.org/10.1155/2013/632121.
- [243] J. Park, J.H. Park, H.-J. Suh, I.C. Lee, J. Koh and Y.C. Boo, Effects of resveratrol, oxyresveratrol, and their acetylated derivatives on cellular melanogenesis, *Arch. Dermatol. Res.* 306, 2014, 475-487, https://doi.org/10.1007/s00403-014-1440-3.

- [244] M. Belleri, D. Ribatti, M. Savio, L.A. Stivala, L. Forti, E. Tanghetti, P. Alessi, D. Coltrini, A. Bugatti, S. Mitola, S. Nicoli, V. Vannini and M. Presta, alphavbeta3 Integrin-dependent antiangiogenic activity of resveratrol stereoisomers, *Mol. Cancer Ther.* 7, 2008, 3761–3770, https://doi.org/10.1158/1535-7163.MCT-07-2351.
- [245] V. Trapp, B. Parmakhtiar, V. Papazian, L. Willmott and J.P. Fruehauf, Anti-angiogenic effects of resveratrol mediated by decreased VEGF and increased TSP1 expression in melanoma-endothelial cell co-culture, Angiogenesis 13, 2010, 305–315, https://doi.org/10.1007/s10456-010-9187-8.
- [246] S. Bhattacharya, S.R. Darjatmoko and A.S. Polans, Resveratrol modulates the malignant properties of cutaneous melanoma through changes in the activation and attenuation of the antiapoptotic protooncogenic protein Akt/PKB, *Melanoma Res.* 21, 2011, 180-187, https://doi.org/10.1097/CMR.0b013e3283456dfc.
- [247] M.-C. Chen, W.-W. Chang, Y.-D. Kuan, S.-T. Lin, H.-C. Hsu and C.-H. Lee, Resveratrol inhibits LPS-induced epithelial-mesenchymal transition in mouse melanoma model, *Innate Immun.* 18, 2012, 685-693, https://doi.org/10.1177/1753425912436589.
- [248] X. Chen, W. Li, C. Xu, J. Wang, B. Zhu, Q. Huang, D. Chen, J. Sheng, Y. Zou, Y.M. Lee, R. Tan, P. Shen, Y.K. Wong, Q. Lin, J. Wang and Z. Hua, Comparative profiling of analog targets: A case study on resveratrol for mouse melanoma metastasis suppression, Theranostics 8, 2018, 3504-3516, https://doi.org/10.7150/thno.24336.
- [249] M. Asensi, I. Medina, A. Ortega, J. Carretero, M.C. Baño, E. Obrador and J.M. Estrela, Inhibition of cancer growth by resveratrol is related to its low bioavailability, *Free Radic. Biol. Med.* 33, 2002, 387-398, https://doi.org/10.1016/S0891-5849(02)00911-5.
- [250] C. Salado, E. Olaso, N. Gallot, M. Valcarcel, E. Egilegor, L. Mendoza and F. Vidal-Vanaclocha, Resveratrol prevents inflammation-dependent hepatic melanoma metastasis by inhibiting the secretion and effects of interleukin-18, *J. Transl. Med.* 9, 2011, 59, https://doi.org/10.1186/1479-5876-9-59.
- [251] B. Menicacci, A. Laurenzana, A. Chillà, F. Margheri, S. Peppicelli, E. Tanganelli, G. Fibbi, L. Giovannelli, M. Del Rosso and A. Mocali, Chronic Resveratrol Treatment Inhibits MRC5 Fibroblast SASP Related Protumoral Effects on melanoma cells, J. Gerontol. A Biol. Sci. 72, 2017, 1187-1195, https://doi.org/10.1093/gerona/glw336.
- [252] S. Yang, F.L. Meyskens, F. Liu, R. Chiu, Z. Yang and B.J. Misner, Nitric oxide initiates progression of human melanoma via a feedback loop mediated by apurinic/apyrimidinic endonuclease-1/redox factor-1, which is inhibited by resveratrol, *Mol. Cancer Ther.* 7, 2008, 3751–3760, https://doi.org/10.1158/1535-7163.mct-08-0562.
- [253] S. Fulda and K.M. Debatin, Sensitization for Tumor Necrosis Factor Related Apoptosis Inducing Ligand-Induced Apoptosis by the Chemopreventive Agent Rumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis by the chemopreventive agent resveratrol, Cancer Res. 64, 2004, 337-346, https://doi.org/10.1158/0008-5472.CAN-03-1656.
- [254] V.N. Ivanov, M.A. Partridge, G.E. Johnson, S.X.L. Huang, H. Zhou and T.K. Hei, Resveratrol sensitizes melanomas to TRAIL through modulation of antiapoptotic gene expression, *Exp. Cell Res.* 314, 2008, 1163–1176, https://doi.org/10.1016/j.yexcr.2007.12.012.
- [255] G.W. Osmond, C.K. Augustine, P.A. Zipfel, J. Padussis and D.S. Tyler, Enhancing melanoma treatment with resveratrol, J. Surg. Res. 172, 2012, 109-115, https://doi.org/10.1016/j.jss.2010.07.033.
- [256] M.Y. Kim, Nitric oxide triggers apoptosis in A375 human melanoma cells treated with capsaicin and resveratrol, Mol. Med. Rep. 5, 2012, 585-591, https://doi.org/10.3892/mmr.2011.688.
- [257] Y.J. Cheng, M.Y. Chang, W.W. Chang, W.K. Wang, C.F. Liu, S.T. Lin and C.H. Lee, Resveratrol Enhances Chemosensitivity in Mouse Melanoma Model Through Connexin 43 Uenhances chemosensitivity in mouse melanoma model through connexin 43 upregulation, Environ. Toxicol. 30, 2015, 877-886, https://doi.org/10.1002/tox.21952.
- [258] H. Luo, M. Umebayashi, K. Doi, T. Morisaki, S. Shirasawa and T. Tsunoda, Resveratrol Overcomes Cellular Resistance to Vemurafenib Through Dephosphorylation of AKT in BRAF-mutated Melanoma Covercomes cellular resistance to vemurafenib through dephosphorylation of AKT in BRAF-mutated melanoma cells, Anticancer Res. 36, 2016, 3585–3589 http://www.ncbi.nlm.nih.gov/pubmed/27354627.
- [259] D. Cosco, D. Paolino, J. Maiuolo, L. Di Marzio, M. Carafa, C.A. Ventura and M. Fresta, Ultradeformable liposomes as multidrug carrier of resveratrol and 5-fluorouracil for their topical delivery, *Int. J. Pharm.* 489, 2015, 1-10, https://doi.org/10.1016/j.ijpharm.2015.04.056.
- [260] S.Hw. Lee, B.Seon. Koo, S.Yi. Park and Y.Mi. Kim, Anti-angiogenic effects of resveratrol in combination with 5-fluorouracil on B16 murine melanoma cells, Mol. Med. Rep. 12, 2015, 2777-2783, https://doi.org/10.3892/mmr.2015.3675.

- [261] J.J. Junco, A. Mancha-Ramirez, G. Malik, S.J. Wei, D.J. Kim, H. Liang and T.J. Slaga, Ursolic acid and resveratrol synergize with chloroquine to reduce melanoma cell viability, *Melanoma Res.* 25, 2015, 103-112, https://doi.org/10.1097/CMR.0000000000000137.
- [262] H. Guan, N.P. Singh, U.P. Singh, P.S. Nagarkatti and M. Nagarkatti, Resveratrol prevents endothelial cells injury in high-dose interleukin-2 therapy against melanoma, *PLoS One* 7, 2012, , e35650https://doi.org/10.1371/journal.pone.0035650.
- [263] Y. Fang, M.J. Bradley, K.M. Cook, E.J. Herrick and M.B. Nicholl, A potential role for resveratrol as a radiation sensitizer for melanoma treatment, *J. Surg. Res.* 183, 2013, 645-653, https://doi.org/10.1016/j.jss.2013.02.037.
- [264] Y. Wong, G. Osmond, K.I. Brewer, D.S. Tyler and M.B. Andrus, Synthesis of 4'-ester analogs of resveratrol and their evaluation in malignant melanoma and pancreatic cell lines, *Bioorganie*, *Med. Chem. Lett.* 20, 2010, 1198–1201, https://doi.org/10.1016/j.bmcl.2009.12.006.
- [265] T. Szekeres, M. Fritzer-Szekeres, P. Saiko and W. Jäger, Resveratrol and resveratrol analogues-structure-activity relationship, Pharm. Res. 27, 2010, 1042-1048, https://doi.org/10.1007/s11095-010-0090-1.
- [266] B. Saha, G.B. Pai, M. Subramanian, P. Gupta, M. Tyagi, B.S. Patro and S. Chattopadhyay, Resveratrol analogue, trans-4,4'-dihydroxystilbene (DHS), inhibits melanoma tumor growth and suppresses its metastatic colonization in lungs, *Biomed. Pharmacother.* 107, 2018, 1104–1114, https://doi.org/10.1016/j.biopha.2018.08.085.
- [267] B.F. Ruan, X. Lu, T.T. Li, J.F. Tang, Y. Wei, X.L. Wang, S.L. Zheng, R.S. Yao and H.L. Zhu, Synthesis, biological evaluation and molecular docking studies of resveratrol derivatives possessing curcumin moiety as potent antitubulin agents, *Bioorganie*, *Med. Chem.* 20, 2012, 1113-1121, https://doi.org/10.1016/j.bmc.2011.11.017.
- [268] V.L. Morris, T. Toseef, F.B. Nazumudeen, C. Rivoira, C. Spatafora, C. Tringali and S.A. Rotenberg, Anti-tumor properties of cis-resveratrol methylated analogs in metastatic mouse melanoma cells, *Mol. Cell. Biochem.* 402 2015, 83-91, https://doi.org/10.1007/s11010-014-2316-8.
- [269] V.P. Androutsopoulos, I. Fragiadaki and A. Tosca, Activation of ERK1/2 is required for the antimitotic activity of the resveratrol analogue 3,4,5,4'-tetramethoxystilbene (DMU-212) in human melanoma cells, Exp. Dermatol. 24, 2015, 632-634, https://doi.org/10.1111/exd.12721.
- [270] V.P. Androutsopoulos, I. Fragiadaki, D.A. Spandidos and A. Tosca, The resveratrol analogue, 3,4,5,4'trans-tetramethoxystilbene, inhibits the growth of A375 melanoma cells through multiple anticancer modes of action, Int. J. Oncol. 49, 2016, 1305–1314, https://doi.org/10.3892/ijo.2016.3635.
- [271] Q. Liu, C.T. Kim, Y.H. Jo, S.B. Kim, B.Y. Hwang and M.K. Lee, Synthesis and biological evaluation of resveratrol derivatives as melanogenesis inhibitors, *Molecules* 20, 2015, 16933-16945, https://doi.org/10.3390/molecules200916933.
- [272] S. Park, J.K. Seok, J.Y. Kwak, Y.-H. Choi, S.S. Hong, H.-J. Suh, W. Park and Y.C. Boo, Anti-melanogenic effects of resveratryl triglycolate, a novel hybrid compound derived by esterification of resveratrol with glycolic acid, Arch. Dermatol. Res. 308, 2016, 325–334, https://doi.org/10.1007/s00403-016-1644-9.
- [273] L. Nivelle, J. Hubert, E. Courot, N. Borie, J.H. Renault, J.M. Nuzillard, D. Harakat, C. Clément, L. Martiny, D. Delmas, P. Jeandet and M. Tarpin, Cytotoxicity of Labruscol, a New Resveratrol Dimer Produced by Grapevine Cell Suspensions, on Human Skin Melanoma Cancer Cell Lnew resveratrol dimer produced by grapevine cell suspensions, on human skin melanoma Cancer cell line HT-144, Molecules 22, 2017, https://doi.org/10.3390/molecules22111940.
- [274] L. Nivelle, V. Aires, D. Rioult, L. Martiny, M. Tarpin and D. Delmas, Molecular analysis of differential antiproliferative activity of resveratrol, epsilon viniferin and labruscol on melanoma cells and normal dermal cells *Food Chem. Toxicol.* 116, 2018, 323–334, https://doi.org/10.1016/j.fct.2018.04.043.
- [275] A. Briones-Herrera, D. Eugenio-Pérez, J.G. Reyes-Ocampo, S. Rivera-Mancía and J. Pedraza-Chaverri, New highlights on the health-improving effects of sulforaphane, *Food Funct.* 9, 2018, 2589-2606, https://doi.org/10.1039/C8FO00018B.
- [276] I. Misiewicz, K. Skupinska and T. Kasprzycka-Guttman, Sulforaphane and 2-oxohexyl isothiocyanate induce cell growth arrest and apoptosis in L-1210 leukemia and ME-18 melanoma cells, *Oncol. Rep.* 10, 2003, 2045–2050 http://www.ncbi.nlm.nih.gov/pubmed/14534741.

- [277] T.P. Hamsa, P. Thejass and G. Kuttan, Induction of apoptosis by sulforaphane in highly metastatic B16F-10 melanoma cells, Drug Chem. Toxicol. 34, 2011, 332-340, https://doi.org/10.3109/01480545.2010.538694.
- [278] K. Rudolf, M. Cervinka and E. Rudolf, Sulforaphane-induced apoptosis involves p53 and p38 in melanoma cells, Apoptosis 19, 2014, 734-747, https://doi.org/10.1007/s10495-013-0959-7.
- [279] P. Arcidiacono, F. Ragonese, A. Stabile, A. Pistilli, E. Kuligina, M. Rende, U. Bottoni, S. Calvieri, A. Crisanti and R. Spaccapelo, Antitumor activity and expression profiles of genes induced by sulforaphane in human melanoma cells, Eur. J. Nutr. 57, 2018, 2547–2569, https://doi.org/10.1007/s00394-017-1527-7.
- [280] P. Arcidiacono, A.M. Stabile, F. Ragonese, A. Pistilli, S. Calvieri, U. Bottoni, A. Crisanti, R. Spaccapelo and M. Rende, Anticarcinogenic activities of sulforaphane are influenced by Nerve Growth Fnerve growth factor in human melanoma A375 cells, Food Chem. Toxicol. 113, 2018, 154-161, https://doi.org/10.1016/j.fct.2018.01.051.
- [281] P. Thejass and G. Kuttan, Antimetastatic activity of Sulforaphane, Life Sci. 78, 2006, 3043-3050, https://doi.org/10.1016/j.lfs.2005.12.038.
- [282] P. Thejass and G. Kuttan, Modulation of cell-mediated immune response in B16F-10 melanoma-induced metastatic tumor-bearing C57BL/6 mice by sulforaphane, *Immunopharmacol. Immunotoxicol.* 29, 2007, 173-186, https://doi.org/10.1080/08923970701511728.
- [283] M.L. Fisher, G. Adhikary, D. Grun, D.M. Kaetzel and R.L. Eckert, The Ezh2 polycomb group protein drives an aggressive phenotype in melanoma cancer stem cells and is a target of diet derived sulforaphane, *Mol. Carcinog.* 55, 2016, 2024–2036, https://doi.org/10.1002/mc.22448.
- [284] D.P. Do, S.B. Pai, S.A.A. Rizvi and M.J. D'Souza, Development of sulforaphane-encapsulated microspheres for cancer epigenetic therapy, *Int. J. Pharm.* 386, 2010, 114-121, https://doi.org/10.1016/j.ijpharm.2009.11.009.
- [285] G.G. Enriquez, S.A.A. Rizvi, M.J. D'Souza and D.P. Do, Formulation and evaluation of drug-loaded targeted magnetic microspheres for cancer therapy, *Int. J. Nanomedicine.* 8, 2013, 1393–1402, https://doi.org/10.2147/IJN.S43479.
- [286] S. Tahata, S.V. Singh, Y. Lin, E.-R. Hahm, J.H. Beumer, S.M. Christner, U.N. Rao, C. Sander, A.A. Tarhini, H. Tawbi, L.K. Ferris, M. Wilson, A. Rose, C.M. Dietz, E. Hughes, J.W. Fahey, S.A. Leachman, P.B. Cassidy, L.H. Butterfield H.M. Zarour and J.M. Kirkwood, Evaluation of Biodistribution of Sulforaphane after Administration of Oral Broccoli Sprout Extract in Melanoma Patients with Multiple Atypical Nevi, Cancer Prev. Res. 11, 2018, 429-438, https://doi.org/10.1158/1940-6207.CAPR-17-0268.
- [287] Ł. Woźniak, S. Skąpska and K. Marszałek, Ursolic Acid—A Pentacyclic Triterpenoid with a Wide Spectrum of Pharmacological Activities Molecules. acid—a pentacyclic triterpenoid with a wide Spectrum of pharmacological activities, Molecules 20, 2015, 20614-20641, https://doi.org/10.3390/molecules201119721.
- [288] P.O. Harmand, R. Duval, C. Delage and A. Simon, Ursolic acid induces apoptosis through mitochondrial intrinsic pathway and caspase-3 activation in M4Beu melanoma cells, *Int. J. Cancer* 114, 2005, 1-11, https://doi.org/10.1002/ijc.20588.
- [289] R.E. Duval, P.-O. Harmand, C. Jayat-Vignoles, J. Cook-Moreau, A. Pinon, C. Delage and A. Simon, Differential involvement of mitochondria during ursolic acid-induced apoptotic process in HaCaT and M4Beu cells, *Onco. Rep.* 19, 2008, 145–149, https://doi.org/10.3892/or.19.1.145.
- [290] K.A. Manu and G. Kuttan, Ursolic acid induces apoptosis by activating p53 and caspase-3 gene expressions and suppressing NF-kB mediated activation of bcl-2 in B16F-10 melanoma cells, *Int. Immunopharmacol.* 8, 2008, 974-981, https://doi.org/10.1016/j.intimp.2008.02.013.
- [291] M. Mahmoudi, S.Z.T. Rabe, M. Balali-Mood, G. Karimi, N. Tabasi and B. Riahi-Zanjani, Ursolic acid induced apoptotic cell death following activation of caspases in isolated human melanoma cells, *Cell Biol. Int.* 39, 2015 230-236, https://doi.org/10.1002/cbin.10376.
- [292] C. Oprean, A. Ivan, F. Bojin, M. Cristea, C. Soica, L. Drăghia, A. Caunii, V. Paunescu and C. Tatu, Selective in vitro anti-melanoma activity of ursolic and oleanolic acids, *Toxicol. Mech. Methods* 28, 2018, 148–156, https://doi.org/10.1080/15376516.2017.1373881.
- [293] A. Pinon, Y. Limami, L. Micallef, J. Cook-Moreau, B. Liagre, C. Delage, R.E. Duval and A. Simon, A novel form of melanoma apoptosis resistance: Mmelanogenesis up-regulation in apoptotic B16-F0 cells delays ursolic acid-triggered cell death, Exp. Cell Res. 317, 2011, 1669–1676, https://doi.org/10.1016/j.yexcr.2011.04.014.

- [294] L. Hassan, A. Pinon, Y. Limami, J. Seeman, C. Fidanzi-Dugas, F. Martin, B. Badran, A. Simon and B. Liagre, Resistance to ursolic acid-induced apoptosis through involvement of melanogenesis and COX-2/PGE2 pathways in human M4Beu melanoma cancer cells, *Exp. Cell Res.* 345, 2016, 60-69, https://doi.org/10.1016/j.yexcr.2016.05.023.
- [295] M. Kanjoormana and G. Kuttan, Antiangiogenic activity of ursolic acid, Integr. Cancer Ther. 9, 2010, 224-235, https://doi.org/10.1177/1534735410367647.
- [296] L. Xiang, T. Chi, Q. Tang, X. Yang, M. Ou, X. Chen, X. Yu, J. Chen, R.J.Y. Ho, J. Shao and L. Jia, A pentacyclic triterpene natural product, ursolic acid and its prodrug US597 inhibit targets within cell adhesion pathway and prevent cancer metastasis, Oncotarget 6, 2015, https://doi.org/10.18632/oncotarget.3261.
- [297] Y.-H. Lee, E. Wang, N. Kumar and R.D. Glickman, Ursolic acid differentially modulates apoptosis in skin melanoma and retinal pigment epithelial cells exposed to UVVIS broadband radiation. Apoptosis: vis broadband radiation. Apoptosis 19, 2014, 816-828, https://doi.org/10.1007/s10495-013-0962-z.
- [298] S.J. Koh, J.K. Tak, S.T. Kim, W.S. Nam, S.Y. Kim, K.M. Park and J.W. Park, Sensitization of ionizing radiation-induced apoptosis by ursolic acid, *Free Radic. Res.* 46, 2012, 339-345, https://doi.org/10.3109/10715762.2012.656101.
- [299] X. Yang, Y. Li, W. Jiang, M. Ou, Y. Chen, Y. Xu, Q. Wu, Q. Zheng, F. Wu, L. Wang, W. Zou, Y.J. Zhang and J. Shao, Synthesis and biological evaluation of novel ursolic acid derivatives as potential anticancer prodrugs, *Chen. Biol. Drug Des.* 86, 2015, 1397-1404, https://doi.org/10.1111/cbdd.12608.
- [300] J. Wiemann, L. Heller and R. Csuk, Targeting cancer cells with oleanolic and ursolic acid derived hydroxamates, Bioorganie, Med. Chem. Lett. 26, 2016, 907-909, https://doi.org/10.1016/j.bmcl.2015.12.064.
- [301] R. Baishya, D.K. Nayak, D. Kumar, S. Sinha, A. Gupta, S. Ganguly and M.C. Debnath, Ursolic Acid Loaded PLGA Nanoparticles: in vitro and in vivo Evaluation to Explore Tumor Targeting Ability on B16F10 Melanoma Cell Lacid loaded PLGA nanoparticles: in vitro and in vivo evaluation to explore tumor targeting ability on B16F10 melanoma cell lines, Pharm. Res. 33, 2016, 2691–2703, https://doi.org/10.1007/s11095-016-1994-1.
- [302] H.L. Alvarado, A.C. Calpena, M.L. Garduño-Ramírez, R. Ortiz, C. Melguizo, J.C. Prados and B. Clares, Nanoemulsion Anticancer strategy for ursolic and oleanic acids isolates from Plumeria obtusa improves antioxidant and cytotoxic activity in melanoma cells, Anticancer Agents Med. Chem. 18, 2018, 847–853, https://doi.org/10.2174/1871520618666180111151846.
- [303] W. Cheng, F.Z. Dahmani, J. Zhang, H. Xiong, Y. Wu, L. Yin, J. Zhou and J. Yao, Anti-angiogenic activity and antitumor efficacy of amphiphilic twin drug from ursolic acid and low molecular weight heparin, Nanotechnology 28, 2017, 075102https://doi.org/10.1088/1361-6528/aa53c6.
- [304] C. Soica, C. Oprean, F. Borcan, C. Danciu, C. Trandafirescu, D. Coricovac, Z. Crăiniceanu, C. Dehelean and M. Munteanu, The Synergistic Biologic Activity of Oleanolic and Ursolic Acids in Complex with Hydroxypropyl-y-Cyclodextrin, Molecules 19, 2014, 4924-4940, https://doi.org/10.3390/molecules19044924.
- [305] C. Oprean, M. Mioc, E. Csányi, R. Ambrus, F. Bojin, C. Tatu, M. Cristea, A. Ivan, C. Danciu, C. Dehelean, V. Paunescu and C. Soica, Improvement of ursolic and oleanolic acids' antitumor activity by complexation with hydrophilic cyclodextrins, *Biomed. Pharmacother.* 83, 2016, 1095–1104, https://doi.org/10.1016/j.biopha.2016.08.030.
- [306] P. Mohanan, S. Subramaniyam, R. Mathiyalagan and D.-C. Yang, Molecular signaling of ginsenosides Rb1, Rg1, and Rg3 and their mode of actions, *J. Ginseng Res.* 42, 2018, 123-132, https://doi.org/10.1016/j.jgr.2017.01.008.
- [307] T. Ota, M. Maeda, S. Odashima, J. Ninomiya-Tsuji and M. Tatsuka, G1 phase-specific suppression of the Cdk2 activity by ginsenoside Rh2 in cultured murine cells, *Life Sci.* 60, 1996, PL39-PL44, https://doi.org/10.1016/S0024-3205(96)00608-X.
- [308] X.-F. Fei, B.-X. Wang, S. Tashiro, T.-J. Li, J.-S. Ma and T. Ikejima, Apoptotic effects of ginsenoside Rh2 on human malignant melanoma A375-S2 cells, *Acta Pharmacol. Sin.* 23, 2002, 315-322 http://www.ncbi.nlm.nih.gov/pubmed/11931705.
- [309] J. Chen, H. Peng, X. Ou-Yang and X. He, Research on the antitumor effect of ginsenoside Rg3 in B16 melanoma cells, Melanoma Res. 18, 2008, 322-329, https://doi.org/10.1097/CMR.0b013e32830b3536.
- [310] X. Shan, Y.S. Fu, F. Aziz, X.Q. Wang, Q. Yan and J.W. Liu, Ginsenoside Rg3 inhibits melanoma cell proliferation through down-regulation of histone deacetylase 3 (HDAC3) and increase of p53 acetylation, *PLoS One* 2014, e115401https://doi.org/10.1371/journal.pone.0115401.

- [311] X. Shan, F. Aziz, L.L. Tian, X.Q. Wang, Q. Yan and J.W. Liu, Ginsenoside Rg3-induced EGFR/MAPK pathway deactivation inhibits melanoma cell proliferation by decreasing FUT4/LeY expression, *Int. J. Oncol.* 46, 2015, 1667–1676, https://doi.org/10.3892/ijo.2015.2886.
- [312] X. Shan, L.L. Tian, Y.M. Zhang, X.Q. Wang, Q. Yan and J.W. Liu, Ginsenoside Rg3 suppresses FUT4 expression through inhibiting NF-xB/p65 signaling pathway to promote melanoma cell death, *Int. J. Oncol.* 47, 2015, 701–709, https://doi.org/10.3892/ijo.2015.3057.
- [313] J.S. Kim, E.J. Joo, J. Chun, Y.W. Ha, J.H. Lee, Y. Han and Y.S. Kim, Induction of apoptosis by ginsenoside Rk1 in SK-MEL-2-human melanoma, Arch. Pharm. Res. 35, 2012, 717-722, https://doi.org/10.1007/s12272-012-0416-0.
- [314] S. Kang, J.E. Kim, N.R. Song, S.K. Jung, M.H. Lee, J.S. Park, M.H. Yeom, A.M. Bode, Z. Dong and K.W. Lee, The ginsenoside 20-O-β-D-glucopyranosyl-20(S)-protopanaxadiol induces autophagy and apoptosis in human melanoma via AMPK/JNK phosphorylation, *PLoS One* 9, 2014, e104305https://doi.org/10.1371/journal.pone.0104305.
- [315] Y.-M. Jeong, W.K. Oh, T.L. Tran, W.-K. Kim, S.H. Sung, K. Bae, S. Lee and J.-H. Sung, Aglycone of Rh 4 Inhibits Melanin Synthesis in B16 Melanoma Cells: Possible Involvement of the Protein Kinase A Pinhibits melanin synthesis in B16 melanoma cells: possible involvement of the protein kinase a pathway, Biosci. Biotechnol. Biochem. 77, 2013, 119-125, https://doi.org/10.1271/bbb.120602.
- [316] L. Wang, A.P. Lu, Z.L. Yu, R.N. Wong, Z.X. Bian, H.H. Kwok, P.Y. Yue, L.M. Zhou, H. Chen, M. Xu and Z. Yang, The melanogenesis-inhibitory effect and the percutaneous formulation of ginsenoside Rb1, AAPS

  PharmSciTech 15, 2014, 1252-1262, https://doi.org/10.1208/s12249-014-0138-3.
- [317] Q. Le Zhou and X.W. Yang, Four new ginsenosides from red ginseng with inhibitory activity on melanogenesis in melanoma cells, *Bioorganie*, *Med. Chem. Lett.* 25, 2015, 3112-3116, https://doi.org/10.1016/j.bmcl.2015.06.017.
- [318] D.Y. Lee, H.G. Kim, Y.G. Lee, J.H. Kim, J.W. Lee, B.R. Choi, I.B. Jang, G.S. Kim and N.I. Baek, Isolation and quantification of ginsenoside rh23, a new anti-melanogenic compound from the leaves of panax ginseng, *Molecules* 23, 2018, E267, https://doi.org/10.3390/molecules23020267.
- [319] J.H. Kim, E.J. Baek, E.J. Lee, M.H. Yeom, J.S. Park, K.W. Lee and N.J. Kang, Ginsenoside F1 attenuates hyperpigmentation in B16F10 melanoma cells by inducing dendrite retraction and activating Rho signalling, *Exp. Dermatol.* 24, 2015, 154-162, https://doi.org/10.1111/exd.12586.
- [320] Y.-S. Jiang, Z.-X. Jin, H. Umehara and T. Ota, Cholesterol-dependent induction of dendrite formation by ginsenoside Rh2 in cultured melanoma cells, *Int. J. Mol. Med.* 26, 2010, 787-793 http://www.ncbi.nlm.nih.gov/pubmed/21042771.
- [321] K. Sato, M. Mochizuki, I. Saiki, Y. Yoo, K. Samukawa and I. Azuma, Inhibition of Tumor Angiogenesis and Metastasis by a Stumor angiogenesis and metastasis by a saponin of Panax ginseng, Ginsenoside-Rb2, Biol. Pharm.

  Bull. 17, 1994, 635-639, https://doi.org/10.1248/bpb.17.635.
- [322] M. Mochizuki, Y.C. Yoo, K. Matsuzawa, K. Sato, I. Saiki, S. Tono-oka, K. Samukawa and I. Azuma, Inhibitory effect of tumor metastasis in mice by saponins, ginsenoside-Rb2, 20(R)- and 20(S)-ginsenoside-Rg3, of red ginseng, *Biol. Pharm. Bull.* 18, 1995, 1197-1202.
- [323] K. Shinkai, H. Akedo, M. Mukai, F. Imamura, A. Isoai, M. Kobayashi and I. Kitagawa, Inhibition of in vitro tumor cell invasion by ginsenoside Rg3, Japanese pn. J. Cancer Res. 87, 1996, 357–362, https://doi.org/10.1111/j.1349-7006.1996.tb00230.x.
- [324] T.Y. Park, M.H. Park, W.C. Shin, M.H. Rhee, D.W. Seo, J.Y. Cho and H.M. Kim, Anti-metastatic Potential of Ginsenoside Rp1, a Novel Ginsenoside Dpotential of ginsenoside Rp1, a novel ginsenoside derivative, Biol. Pharm. Bull. 31, 2008, 1802-1805, https://doi.org/10.1248/bpb.31.1802.
- [325] S.G. Lee, Y.J. Kang and J.O. Nam, Anti-metastasis effects of ginsenoside Rg3 in B16F10 cells, J. Microbiol. Biotechnol. 25, 2015, 1997-2006, https://doi.org/10.4014/jmb.1506.06002.
- [326] M. Wang, S.-J. Yan, H.-T. Zhang, N. Li, T. Liu, Y.-L. Zhang, X.-X. Li, Q. Ma, X.-C. Qiu, Q.-Y. Fan and B.-A. Ma, Ginsenoside Rh2 enhances the antitumor immunological response of a melanoma mice model, *Oncol. Lett.* 13, 2017, 681–685, https://doi.org/10.3892/ol.2016.5490.
- [327] K. Son, K. ryung Choi, S.J. Lee and H. Lee, Immunogenic Cell Death Induced by Ginsenoside Rg3: Significance in Dendritic Cell Deadh induced by ginsenoside Rg3: significance in dendritic cell-based anti

- tumor immunotherapy, Immune Netw. 16, 2016, 75, https://doi.org/10.4110/in.2016.16.1.75.
- [328] Z. Wang, Q. Zheng, K. Liu, G. Li and R. Zheng, Ginsenoside Rh2 Enhances Antitumour Activity and Decreases Genotoxic Effect of Cenhances antitumour activity and decreases genotoxic effect of cyclophosphamide, Basic Clin Pharmacol. Toxicol. 98, 2006, 411-415, https://doi.org/10.1111/j.1742-7843.2006.pto 348.x.
- [329] J. Lei, X. Li, X.J. Gong and Y.N. Zheng, Isolation, synthesis and structures of cytotoxic ginsenoside derivatives, Molecules 12, 2007, 2140-2150, https://doi.org/10.3390/12092140.
- [330] C.-H. Cui, D.J. Kim, S.-C. Jung, S.-C. Kim and W.-T. Im, Enhanced Production of Gypenoside LXXV Using a Novel Ginsenoside Transforming Glucosidase from Ginseng Cultivating Soil Bacteria and Its Anti-Cancer Property Molecules. Production of gypenoside LXXV using a novel ginsenoside-transforming β-Glucosidase from ginseng-cultivating soil Bacteria and its anti-cancer property, Molecules 22, 2017, 844, https://doi.org/10.3390/molecules22050844.
- [331] H. Zare-Zardini, A. Taheri-Kafrani, A. Amiri and A.-K. Bordbar, New generation of drug delivery systems based on ginsenoside Rh2-, Lysine- and Arginine-treated highly porous graphene for improving anticancer activity, *Sci. Rep.* 8, 2018, 586, https://doi.org/10.1038/s41598-017-18938-y.
- [332] H. Ahsan, A. Ahad, J. Igbal and W.A. Siddiqui, Pharmacological potential of tocotrienols: a review, Nutr. Metab. (Lond). 11, 2014, 52, https://doi.org/10.1186/1743-7075-11-52.
- [333] M. Montagnani Marelli, M. Marzagalli, F. Fontana, M. Raimondi, R.M. Moretti and P. Limonta, Anticancer properties of tocotrienols: Az review of cellular mechanisms and molecular targets, *J. Cell. Physiol.* 234, 2019, 1147–1164, https://doi.org/10.1002/jcp.27075.
- [334] F. Fontana, M. Raimondi, M. Marzagalli, R.M. Moretti, M. Montagnani Marelli and P. Limonta, Tocotrienols and Cancer: From the State of the Art to Promising Novel Pfrom the state of the art to promising novel patents, Recent Pat. Anticancer. Drug Discov. 2019, https://doi.org/10.2174/1574892814666190116111827.
- [335] F. Fontana, R.M. Moretti, M. Raimondi, M. Marzagalli, G. Beretta, P. Procacci, P. Sartori, M. Montagnani Marelli and P. Limonta, & Tocotrienol induces apoptosis, involving endoplasmic reticulum stress and autophagy, and paraptosis in prostate cancer cells, Cell Prolif. 2019, e12576, https://doi.org/10.1111/cpr.12576.
- [336] K.D. Tang, J. Liu, P.J. Russell, J.A. Clements and M.T. Ling, Gamma-Tocotrienol Induces Apoptosis in Prostate Cancer Cells by Tocotrienol induces apoptosis in prostate Cancer cells by targeting the Ang-1/Tier-2 Signalling pathway, Int. J. Mol. Sci. 20, 2019, E1164, https://doi.org/10.3390/ijms20051164.
- [337] L.D. Rajasinghe, M. Hutchings and S.V. Gupta, Delta Tocotrienol Modulates Glutamine Dependence by Itocotrienol modulates glutamine dependence by Inhibiting ASCT2 and LAT1 Transporters in Non-Small Cell Lung Cancer (NSCLC) Cells: a metabolomic approach, Metabolites 9, 2019, E50, https://doi.org/10.3390/metabo9030050.
- [338] M. Montagnani Marelli, M. Marzagalli, R.M. Moretti, G. Beretta, L. Casati, R. Comitato, G.L. Gravina, C. Festuccia and P. Limonta, Vitamin E 6\_tocotrienol triggers endoplasmic reticulum stress-mediated apoptosis in human melanoma cells, Sci. Rep. 6, 2016, 30502, https://doi.org/10.1038/srep30502.
- [339] G. Beretta, F. Gelmini, F. Fontana, R.M. Moretti, M. Montagnani Marelli and P. Limonta, Semi-preparative HPLC purification of 6-tecetrienel (-tocotrienel (6-T3)) from Elaeis guineensis Jacq. and Bixa orellana L. and evaluation of its in vitro anticancer activity in human A375 melanoma cells, Nat. Prod. Res. 32, 2018, 1130-1135, https://doi.org/10.1080/14786419.2017.1320793.
- [340] M. Marzagalli, R.M. Moretti, E. Messi, M.M. Marelli, F. Fontana, A. Anastasia, M.R. Bani, G. Beretta and P. Limonta, Targeting melanoma stem cells with the Vitamin E derivative & tocotrienol, Sci. Rep. 8, 2018, 587, https://doi.org/10.1038/s41598-017-19057-4.
- [341] P.N. Chang, W.N. Yap, D.T.W. Lee, M.T. Ling, Y.C. Wong and Y.L. Yap, Evidence of gamma-tocotrienol as an apoptosis-inducing, invasion-suppressing, and chemotherapy drug-sensitizing agent in human melanoma cells, *Nutr. Cancer* 61, 2009, 357-366, https://doi.org/10.1080/01635580802567166.
- [342] J. a McAnally, J. Gupta, S. Sodhani, L. Bravo and H. Mo, Tocotrienols potentiate lovastatin-mediated growth suppression in vitro and in vivo, *Exp. Biol. Med. (Maywood)* 232, 2007, 523-531 http://www.ncbi.nlm.nih.gov/pubmed/17392488.
- [343] S. Yamashita, K. Baba, A. Makio, M. Kumazoe, Y. Huang, I.-C. Lin, J. Bae, M. Murata, S. Yamada and H. Tachibana, y-Tocotrienol upregulates aryl hydrocarbon receptor expression and enhances the anticancer effect of

- baicalein, Biochem. Biophys. Res. Commun. 473, 2016, 801-807, https://doi.org/10.1016/j.bbrc.2016.03.111.
- [344] R. Karim, S. Somani, M. Al Robaian, M. Mullin, R. Amor, G. McConnell and C. Dufès, Tumor regression after intravenous administration of targeted vesicles entrapping the vitamin E α-tocotrienol, *J. Control. Release*-246, 2017, 79-87, https://doi.org/10.1016/j.jconrel.2016.12.014.
- [345] M. Imenshahidi and H. Hosseinzadeh, Berberis Vulgaris and Berberine: An Update Ryulgaris and berberine: an update review, Phytother. Res. 30, 2016, 1745-1764, https://doi.org/10.1002/ptr.5693.
- [346] T.L. Serafim, P.J. Oliveira, V.A. Sardao, E. Perkins, D. Parke and J. Holy, Different concentrations of berberine result in distinct cellular localization patterns and cell cycle effects in a melanoma cell line, *Cancer Chemother: Pharmacol.* 61, 2008, 1007–1018, https://doi.org/10.1007/s00280-007-0558-9.
- [347] A. Burgeiro, C. Gajate, E.H. Dakir, J.A. Villa-Pulgarín, P.J. Oliveira and F. Mollinedo, Involvement of mitochondrial and B-RAF/ERK signaling pathways in berberine-induced apoptosis in human melanoma cells, Anticancer, Drugs. Drugs 22, 2011, 507-518, https://doi.org/10.1097/CAD.0b013e32834438f6.
- [348] S. Letasiová, S. Jantová, L. Cipák and M. Múcková, Berberine-antiproliferative activity in vitro and induction of apoptosis/necrosis of the U937 and B16 cells, *Cancer Lett.* 239, 2006, 254-262, https://doi.org/10.1016/j.canlet.2005.08.024.
- [349] Y.C. Song, Y. Lee, H.M. Kim, M.Y. Hyun, Y.Y. Lim, K.Y. Song and B.J. Kim, Berberine regulates melanin synthesis by activating PI3K/AKT, ERK and GSK3β in B16F10 melanoma cells, *Int. J. Mol. Med.* 35, 2015, 1011-1016, https://doi.org/10.3892/ijmm.2015.2113.
- [350] Y. Kou, L. Li, H. Li, Y. Tan, B. Li, K. Wang and B. Du, Berberine suppressed epithelial mesenchymal transition through cross-talk regulation of PI3K/AKT and RARα/RARβ in melanoma cells, *Biochem. Biophys. Res. Commun.* 479, 2016, 290-296, https://doi.org/10.1016/j.bbrc.2016.09.061.
- [351] J.F. Liu, K.C. Lai, S.F. Peng, P. Maraming, Y.P. Huang, A.C. Huang, F.S. Chueh, W.W. Huang and J.G. Chung, Berberine inhibits human melanoma A375.S2 cell migration and invasion via affecting the FAK, uPA, and NF-κE signaling pathways and inhibits PLX4032 resistant A375.S2 cell migration in vitro, *Molecules* 23, 2018, E2019, https://doi.org/10.3390/molecules23082019.
- [352] T.P. Hamsa and G. Kuttan, Berberine inhibits pulmonary metastasis through down-regulation of MMP in metastatic B16F-10 melanoma cells, Phyther. Res. 26, 2012, 568-578, https://doi.org/10.1002/ptr.3586.
- [353] H.S. Kim, M.J. Kim, E.J. Kim, Y. Yang, M.S. Lee and J.S. Lim, Berberine-induced AMPK activation inhibits the metastatic potential of melanoma cells via reduction of ERK activity and COX-2 protein expression, Biochem. Pharmacol. 83, 2012, 385-394, https://doi.org/10.1016/j.bcp.2011.11.008.
- [354] T.P. Hamsa and G. Kuttan, Antiangiogenic activity of berberine is mediated through the downregulation of hypoxia-inducible factor-1, VEGF, and proinflammatory mediators, *Drug Chem. Toxicol.* 35, 2012, 57-70, https://doi.org/10.3109/01480545.2011.589437.
- [355] A. Mittal, S. Tabasum and R.P. Singh, Berberine in combination with doxorubicin suppresses growth of murine melanoma B16F10 cells in culture and xenograft, *Phytomedicine* 21, 2014, 340-347, https://doi.org/10.1016/j.phymed.2013.09.002.
- [356] P.W. Mannal, J. Schneider, A. Tangada, D. McDonald and D.W. McFadden, Honokiol produces anti-neoplastic effects on melanoma cells in vitro, J. Surg. Oncol. 104, 2011, 260-264, https://doi.org/10.1002/jso.21936.
- [357] K.-L. Lan, K.-H. Lan, M.-L. Sheu, M.-Y. Chen, Y.-S. Shih, F.-C. Hsu, H.-M. Wang, R.-S. Liu and S.-H. Yen, Honokiol inhibits hypoxia-inducible factor-1 pathway, *Int. J. Radiat. Biol.* 87, 2011, 579–590, https://doi.org/10.3109/09553002.2011.568572.
- [358] G. Kaushik, S. Ramalingam, D. Subramaniam, P. Rangarajan, P. Protti, P. Rammamoorthy, S. Anant and J.M.V. Mammen, Honokiol induces cytotoxic and cytostatic effects in malignant melanoma cancer cells, *Am. J. Surg.* 204, 2012, 868–873, https://doi.org/10.1016/j.amjsurg.2012.09.001.
- [359] R. Guillermo-Lagae, S. Santha, M. Thomas, E. Zoelle, J. Stevens, R.S. Kaushik and C. Dwivedi, Antineoplastic effects of honokiol on melanoma, *Biomed Res. Int.* 2017, 2017, 5496398https://doi.org/10.1155/2017/5496398.
- [360] A.P. Trotta, J.D. Gelles, M.N. Serasinghe, P. Loi, J.L. Arbiser and J.E. Chipuk, Disruption of mitochondrial electron transport chain function potentiates the pro-apoptotic effects of MAPK inhibition, J. Biol. Chem. 292,

- 2017, 11727-11739, https://doi.org/10.1074/jbc.M117.786442.
- [361] S. Martin, H.K. Lamb, C. Brady, B. Lefkove, M.Y. Bonner, P. Thompson, P.E. Lovat, J.L. Arbiser, A.R. Hawkins and C.P.F. Redfern, Inducing apoptosis of cancer cells using small-molecule plant compounds that bind to GRP78, Br. J. Cancer 109, 2013, 433-443, https://doi.org/10.1038/bjc.2013.325.
- [362] C.S. Chiu, C.H. Tsai, M.S. Hsieh, S.C. Tsai, Y.J. Jan, W.Y. Lin, D.W. Lai, S.M. Wu, H.Y. Hsing, J.L. Arbiser and M.L. Sheu, Exploiting Honokiol-induced ER stress CHOP activation inhibits the growth and metastasis of melanoma by suppressing the MITF and β-catenin pathways, *Cancer Lett.* 442, 2019, 113-125, https://doi.org/10.1016/j.canlet.2018.10.026.
- [363] R. Prasad, J.C. Kappes and S.K. Katiyar, Inhibition of NADPH oxidase 1 activity and blocking the binding of cytosolic and membrane-bound proteins by honokiol inhibit migratory potential of melanoma cells, Oncotarget -7, 2016, 7899-7912, https://doi.org/10.18632/oncotarget.6860.
- [364] G. Kaushik, D. Kwatra, D. Subramaniam, R.A. Jensen, S. Anant and J.M.V. Mammen, Honokiol affects melanoma cell growth by targeting the AMP-activated protein kinase signaling pathway, Am. J. Surg. 208, 2014, 995–1002, https://doi.org/10.1016/j.amjsurg.2014.09.014.
- [365] G. Kaushik, A. Venugopal, P. Ramamoorthy, D. Standing, D. Subramaniam, S. Umar, R.A. Jensen, S. Anant and J.M.V. Mammen, Honokiol inhibits melanoma stem cells by targeting notch signaling, *Mol. Carcinog.* 54, 2015, 1710–1721, https://doi.org/10.1002/mc.22242.
- [366] M.Y. Bonner, I. Karlsson, M. Rodolfo, R.S. Arnold, E. Vergani and J.L. Arbiser, Honokiol bis-dichloroacetate (Honokiol DCA) demonstrates activity in vemurafenib-resistant melanoma in vivo, Oncotarget 7, 2016, 12857–12868, https://doi.org/10.18632/oncotarget.7289.
- [367] M. Ramacher, V. Umansky and T. Efferth, Effect of artesunate on immune cells in ret-transgenic mouse melanoma model, Anticancer, Drugs 20, 2009, 910-917, https://doi.org/10.1097/CAD.0b013e328330caba.
- [368] E. Buommino, A. Baroni, N. Canozo, M. Petrazzuolo, R. Nicoletti, A. Vozza and M.A. Tufano, Artemisinin reduces human melanoma cell migration by down-regulating αVβ3 integrin and reducing metalloproteinase 2 production, *Invest. New Drugs* -27, 2009, 412-418, https://doi.org/10.1007/s10637-008-9188-2.
- [369] C.M. Cabello, S.D. Lamore, W.B. Bair, S. Qiao, S. Azimian, J.L. Lesson and G.T. Wondrak, The redox antimalarial dihydroartemisinin targets human metastatic melanoma cells but not primary melanocytes with induction of NOXA-dependent apoptosis, *Invest. New Drugs* 30, 2012, 1289-1301, https://doi.org/10.1007/s10637-011-9676-7.
- [370] A. Dwivedi, A. Mazumder, L. du Plessis, J.L. du Preez, R.K. Haynes and J. du Plessis, In vitro anti-cancer effects of artemisone nano-vesicular formulations on melanoma cells, Nanotechnology Nanomedicine Nanotechnology Nanotechnology Nanomedicine Nanotechnology Nano
- [371] N.H. Zuma, F.J. Smit, C. de Kock, J. Combrinck, P.J. Smith and D.D. N'Da, Synthesis and biological evaluation of a series of non-hemiacetal ester derivatives of artemisinin, Eur. J. Med. Chem. 122, 2016, 635-646, https://doi.org/10.1016/j.ejmech.2016.07.027.
- [372] L. Zheng and J. Pan, The Anti-malarial Drug Artesunate Blocks W The anti-malarial drug artesunate blocks wnt/β-catenin Pat hway and Curr. Cancer Drug Targets. inhibits growth, migration and invasion of uveal melanoma cells Curr. Cancer Drug Targets 18, 2018, 988-998, https://doi.org/10.2174/1568009618666180425142653.
- [373] C.-L. Zhang, L.-J. Wu, H.-J. Zuo, S.-I. Tashiro, S. Onodera and T. Ikejima, Cytochrome c release from oridonin-treated apoptotic A375-S2 cells is dependent on p53 and extracellular signal-regulated kinase activation, J. Pharmacol. Sci. 96, 2004, 155-163, https://doi.org/10.1254/jphs.FPJ04008X.
- [374] C.L. Zhang, L.J. Wu, S.I. Tashiro, S. Onodera and T. Ikejima, Oridonin induced A375-S2 cell apoptosis via bax-regulated caspase pathway activation, dependent on the cytochrome C/caspase-9 apoptosome, *J. Asian Nat. Prod. Res.* 6, 2004, 127-138, https://doi.org/10.1080/1028602031000147375.
- [375] K.K. Ren, H.Z. Wang, L.P. Xie, D.W. Chen, X. Liu, J. Sun, Y.C. Nie and R.Q. Zhang, The effects of oridonin on cell growth, cell cycle, cell migration and differentiation in melanoma cells, *J. Ethnopharmacol.* 103, 2006, 176-180, https://doi.org/10.1016/j.jep.2005.07.020.
- [376] H.J. Wang, D. Li, F.Y. Yang, S.I. Tashiro, S. Onodera and T. Ikejima, Oridonin induces human melanoma A375-S2 cell death partially through inhibiting insulin-like growth factor 1 receptor signaling, J. Asian Nat. Prod.

- Res. 10, 2008, 787-798, https://doi.org/10.1080/10286020802030918.
- [377] Z. Gu, X. Wang, R. Qi, L. Wei, Y. Huo, Y. Ma, L. Shi, Y. Chang, G. Li and L. Zhou, Oridonin induces apoptosis in uveal melanoma cells by upregulation of Bim and downregulation of Fatty Acid Sfatty acid synthase,

  Biochem. Biophys. Res. Commun. 457, 2015, 187-193, https://doi.org/10.1016/j.bbrc.2014.12.086.
- [378] C.-Y. Li, Q. Wang, S. Shen, X.-L. Wei and G.-X. Li, Oridonin inhibits migration, invasion, adhesion and TGF-β1-induced epithelial-mesenchymal transition of melanoma cells by inhibiting the activity of PI3K/Akt/GSK-3β signaling pathway, *Oncol. Lett.* 15, 2018, 1362-1372, https://doi.org/10.3892/ol.2017.7421.
- [379] T.P. Hamsa and G. Kuttan, Harmine inhibits tumour specific neo-vessel formation by regulating VEGF, MMP, TIMP and pro-inflammatory mediators both in vivo and in vitro, Eur. J. Pharmacol. 649, 2010, 64-73, https://doi.org/10.1016/j.ejphar.2010.09.010.
- [380] T.P. Hamsa and G. Kuttan, Harmine activates intrinsic and extrinsic pathways of apoptosis in B16F-10 melanoma, Chin. Med. 6, 2011, 11, https://doi.org/10.1186/1749-8546-6-11.
- [381] T.P. Hamsa and G. Kuttan, Studies on Anti-invasive Effects of Harmine Using Highly Metastatic Murine B16F-10 Melanoma Canti-metastatic and anti-invasive effects of harmine using highly metastatic murine B16F-10 melanoma cells, J. Environ. Pathol. Toxicol. Oncol. 30, 2012, 123-137, https://doi.org/10.1615/jenvironpatholtoxicoloncol.v30.i2.40.
- [382] X.F. Zhang, R.Q. Sun, Y.F. Jia, Q. Chen, R.F. Tu, K.K. Li, X.D. Zhang, R.L. Du and R.H. Cao, Synthesis and mechanisms of action of novel harmine derivatives as potential antitumor agents, Sci. Rep. 6, 2016, 33204, https://doi.org/10.1038/srep33204.
- [383] A. Carvalho, J. Chu, C. Meinguet, R. Kiss, G. Vandenbussche, B. Masereel, J. Wouters, A. Kornienko, J. Pelletier and V. Mathieu, Data in support of a harmine-derived beta-carboline in vitro effects in cancer cells through protein synthesis, *Data Br.* 12, 2017, 546-551, https://doi.org/10.1016/j.dib.2017.05.006.
- [384] D.J. Morré, E. Sun, C. Geilen, L.Y. Wu, R. De Cabo, K. Krasagakis, C.E. Orfanos and D.M. Morré, Capsaicin inhibits plasma membrane NADH oxidase and growth of human and mouse melanoma lines, Eur. J. Cancer Part A 32A, 1996, 1995–2003, https://doi.org/10.1016/0959-8049(96)00234-1.
- [385] P.S. Patel, M.L. Varney, B.J. Dave and R.K. Singh, Regulation of Constitutive and Induced NF-B Activation in Malignant Melanoma Cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin Mconstitutive and induced NF- K B activation in malignant melanoma cells by Capsaicin M
- [386] H.S. Jun, T. Park, C.K. Lee, M.K. Kang, M.S. Park, H. Il Kang, Y.J. Surh and O.H. Kim, Capsaicin induced apoptosis of B16-F10 melanoma cells through down-regulation of Bcl-2, Food Chem. Toxicol. 45, 2007, 708-715, https://doi.org/10.1016/j.fct.2006.10.011.
- [387] D.H. Shin, O.H. Kim, H.S. Jun and M.K. Kang, Inhibitory effect of capsaicin on B16-F10 melanoma cell migration via the phosphatidylinositol 3-kinase/Akt/Rac1 signal pathway, Exp. Mol. Med. 40, 2008, 486-494, https://doi.org/10.3858/emm.2008.40.5.486.
- [388] K.C. Pramanik, S.K. Kudugunti, N.M. Fofaria, M.Y. Moridani and S.K. Srivastava, Caffeic acid phenethyl ester suppresses melanoma tumor growth by inhibiting PI3K/AKT/XIAP pathway, *Carcinogenesis*. 34, 2013, 2061–2070, https://doi.org/10.1093/carcin/bgt154.
- [389] S.K. Kudugunti, N.M. Vad, A.J. Whiteside, B.U. Naik, M.A. Yusuf, K.S. Srivenugopal and M.Y. Moridani, Biochemical mechanism of Caffeic Acid Phenylethyl Ester (CAPE) selective toxicity towards melanoma cell lines, *Chem. Biol. Interact.* 188, 2010, 1-14, https://doi.org/10.1016/j.cbi.2010.05.018.
- [390] S.K. Kudugunti, H. Thorsheim, M.S. Yousef, L. Guan and M.Y. Moridani, The metabolic bioactivation of caffeic acid phenethyl ester (CAPE) mediated by tyrosinase selectively inhibits glutathione S-transferase, *Chem. Biol. Interact.* 192, 2011, 243-256, https://doi.org/10.1016/j.cbi.2011.03.015.
- [391] S.K. Kudugunti, N.M. Vad, E. Ekogbo and M.Y. Moridani, Efficacy of Caffeic acid Phenethyl Ester (CAPE) in skin B16-F0 melanoma tumor bearing C57BL/6 mice, *Invest. New Drugs* 29, 2011, 52-62, https://doi.org/10.1007/s10637-009-9334-5.
- [392] L.P. Pelinson, C.E. Assmann, T.V. Palma, I.B.M. da Cruz, M.M. Pillat, A. Mânica, N. Stefanello, G.C.C. Weis, A. de Oliveira Alves, C.M. de Andrade, H. Ulrich, V.M.M. Morsch, M.R.C. Schetinger and M.D. Bagatini, Antiproliferative and apoptotic effects of caffeic acid on SK-Mel-28 human melanoma cancer cells, *Mol. Biol. Rep.* 2019, https://doi.org/10.1007/s11033-019-04658-1.

- [393] T.-W. Chung, S.-J. Kim, H.-J. Choi, C.-H. Kwak, K.-H. Song, S.-J. Suh, K.-J. Kim, K.-T. Ha, Y.-G. Park, Y.-C. Chang, H.W. Chang, Y.-C. Lee and C.-H. Kim, CAPE suppresses VEGFR-2 activation, and tumor neovascularization and growth, J. Mol. Med. 91, 2013, 271-282, https://doi.org/10.1007/s00109-012-0952-6.
- [394] J.-Y. Lee, H.-J. Choi, T.-W. Chung, C.-H. Kim, H.-S. Jeong and K.-T. Ha, Caffeic Acid Phenethyl Ester Inhibits Alpha Melanocyte Stimulating Hormone Induced Melanin Synthesis through Suppressing Transactivation Activity of Microphthalmia-Associated Transactivation activity of microphthalmia-associated transcription factor, J. Nat. Prod. 76, 2013, 1399-1405, https://doi.org/10.1021/np400129z.
- [395] H. Maruyama, F. Kawakami, T.-T. Lwin, M. Imai and F. Shamsa, Biochemical Characterization of Ferulic Acid and Caffeic Acid Which Effectively Inhibit Melanin Synthesis via Different Mechanisms in B16 Melanoma Cells, Biol. Pharm. Bull. 41, 2018, 806-810, https://doi.org/10.1248/bpb.b17-00892.

### **Queries and Answers**

**Query:** Your article is registered as belonging to the Special Issue/Collection entitled "Melanoma biology". If this is NOT correct and your article is a regular item or belongs to a different Special Issue please contact r.nair@elsevier.com immediately prior to returning your corrections.

**Answer:** This is correct

Query: The author names have been tagged as given names and surnames (surnames are highlighted in teal color). Please confirm if they have been identified correctly.

Answer: There is an error. Marina is the first name and Montagnani Marelli the surnames (2 surnames)

Query: Have we correctly interpreted the following funding source(s) and country names you cited in your article: MIUR?

**Answer:** Yes