

1 **From optimized monovalent ligands to size-controlled dendrimers: an**
2 **efficient strategy towards high-activity DC-SIGN antagonists**

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9

10 **Abstract**

11 This short review describes our work on the development of dendrimeric antagonists of DC-
12 SIGN, a Dendritic Cells (DCs) receptor recognizing highly mannosylated structures and
13 primarily involved in the recognition of viruses, such as HIV. The structure of pseudo-di-
14 mannoside and pseudo-tri-mannoside compounds was first finely modified to obtain DC-
15 SIGN ligands more stable and selective than mannose. Their DC-SIGN affinity differences
16 were amplified once presented on multivalent dendrimer-like scaffolds, including poly-alkyne
17 terminated and phenylene-ethynylene rod-like ones. Libraries of mannosylated dendrimers
18 were synthesized, improving their stability and maximizing their monodispersity. The effect
19 of the dendrimers valency, structure and size on DC-SIGN affinity and antiviral potency was
20 investigated. Both the valency and the topology of the architectures were revealed as key
21 parameters for activity optimization, together with the intrinsic affinity of the monovalent
22 ligand. The stability, rigidity and length of the scaffolds were also tuned. The design of
23 geometrically adapted scaffolds afforded one of the most potent inhibitors of DC-SIGN-
24 mediated HIV infections to date. This monodispersed, not cytotoxic and highly active
25 compound was also tested with DCs; its internalization into endolysosomal compartments and
26 its ability to induce the overexpression of signaling molecules makes it a good precursor to
27 produce pathogen-entry inhibitors with immunomodulant properties.

28

29 **Key words (3-5):** DC-SIGN, glycomimetics, glycodendrimers, HIV, multivalency.

30

31 **Introduction**

32 Human immunodeficiency virus (HIV) is still a huge health problem of the 21st century,
33 causing the death of over 1 million people per year from AIDS-related illnesses¹. Waiting for
34 an effective HIV vaccine², the search for HIV entry-inhibitors represents a promising strategy
35 to prevent HIV infection. HIV mainly targets Dendritic Cells (DCs), monocyte/macrophage
36 lineage cells, and CD4⁺ T-lymphocytes³. Both receptor-mediated and non-specific
37 electrostatic interactions drive the contact between the virus and the host cells. The discovery
38 of artificial molecules able to efficiently antagonize one or a combination of these interactions
39 goes in the direction of producing powerful microbicides. Nano-sized compounds have shown
40 promising potential as effective inhibitors of viral infections⁴. Several classes of dendrimers
41 have been designed and used to block different HIV-host cells interactions, targeting either
42 cellular receptors or the viral envelope; their application as HIV entry-inhibiting microbicides
43 has been recently reviewed⁵.

44 In this short review, we will describe the research that has been carried out in our group for
45 the past decade, focusing on the discovery of microbicides that operate by antagonizing the
46 dendritic cell receptor DC-SIGN. DC-SIGN (dendritic cell-specific ICAM-3 grabbing non-
47 integrin) is a trans-membrane lectin expressed at the surface of immature dendritic cells,
48 particularly of dermal and mucosal tissue, and involved in the early stages of HIV infection⁶.
49 DC-SIGN recognizes also other pathogens, including Ebola or Dengue viruses, promoting
50 viral transmission and dissemination⁷. This lectin is a tetramer presenting four carbohydrate
51 recognition domains (CRDs) that bind to highly mannosylated and fucosylated structures in a
52 calcium-dependent manner. As for many other lectins, the affinity of DC-SIGN for its glycan
53 ligands is only weak⁸ and strong binding is obtained, both in Nature and by synthetic
54 antagonists, using multivalency. DC-SIGN antagonists have been produced functionalizing
55 various multivalent platforms, including dendrimers, with mannose and oligomannoses^{9,10}. In

56 parallel, our strategy consisted first of the design and the synthesis of unnatural monovalent
57 glycoconjugate mimetics, more complex and possibly more active and selective than mannose
58 itself. Secondly, the most promising candidates were used for the synthesis of polyvalent
59 constructs, mostly based on dendrimers or dendron scaffolds. The effect of the dendrimers
60 valency, structure and size on DC-SIGN affinity and antiviral potency has been investigated.
61 For this endeavour we have used both real dendrimers/dendrons and dendrimer-inspired
62 structures, i.e. molecules that lack a proper nucleus and repeated layers, but still are
63 symmetrical and branched.

64

65 **Monovalent ligand optimization**

66 The high mannose glycan $(\text{Man})_9(\text{GlcNAc})_2$ (Figure 1) is one of the main carbohydrate
67 ligands recognized by DC-SIGN; it is found in multiple copies on the HIV envelope
68 glycoprotein gp120. Its terminal branches $\text{Man}\alpha 1-2\text{Man}$ and $\text{Man}\alpha 1-2 \text{Man}\alpha 1-6 \text{Man}\alpha$
69 both involved in binding with the protein¹¹, although recent studies indicate that the main
70 interaction may occur through the oligosaccharide branched trimannose core¹².

71 We have shown that pseudo-di-mannoside **1**^{13,14} and pseudo-tri-mannoside **2**^{15,16} (Figure 2),
72 where one of the mannose residues has been replaced by an appropriate conformationally
73 constrained cyclohexane ring, perform as structural and functional mimics of the natural
74 sugars, while being stabilized against enzymatic hydrolysis¹³.

75 Both ligands **1** and **2** bind to DC-SIGN in the carbohydrate recognition domain, as shown by
76 the X-ray structure of their protein complexes^{17,18}. The DC-SIGN affinity of these monovalent
77 ligands was estimated by Surface Plasmon Resonance (SPR) competition experiments,
78 whereby their ability to inhibit DC-SIGN binding to a highly mannosylated surface was
79 measured. In these experiments, where both the lectin and the antagonists are free in solution,
80 the pseudo-trisaccharide **2** (IC_{50} 125 μM) was found to be ca. one order of magnitude more

81 potent than the corresponding pseudo-disaccharide **1**¹⁶, which in turn has the same activity of
82 the natural sugar Man α 1-2Man (IC₅₀ ca. 1 mM)¹⁷, but is significantly less cytotoxic in cellular
83 experiments¹⁴. Mimic **3** (Figure 2c), which is further functionalized on the aglycon, belongs to
84 a group of modified ligands of intermediate DC-SIGN affinity (IC₅₀ 300 μ M) that showed
85 selectivity for DC-SIGN against similar C-type lectin receptors involved in protection
86 mechanisms against HIV¹⁹.

87

88 **Dendrimer optimization**

89 Protein-carbohydrate interactions are relatively weak, with dissociation constants often lying
90 in the low millimolar range. Nonetheless, high affinity can be achieved through multiple
91 interactions. Therefore, in Nature lectin receptors present either multiple carbohydrate
92 recognition domains or an oligomeric structure allowing multiple binding towards the target
93 glycans, usually exposed in several copies as glycoconjugates forming polyglycosylated
94 surfaces. Multivalent interactions affect lectin affinity and also modulate lectin selectivity.
95 For these reasons, to compete with this kind of interactions and efficiently bind to natural
96 proteins, artificial glycoconjugates should be multivalent²⁰. The multivalent binding modes
97 that can be exploited include chelation, i.e. the simultaneous binding of more than one protein
98 binding site by a single multivalent system (Figure 3a), as well as other non-chelating effects,
99 such as the statistical rebinding effect (Figure 3b), caused by enhancement of the local
100 concentration of the active ligands, and the ability of the multivalent structures to
101 cluster/aggregate receptors (Figure 3c)²¹.

102 Notably, the density of sugar residues, the topology of presentation and the orientation have
103 emerged as important parameters able to shape protein/carbohydrate interactions affinity and
104 selectivity^{22,23}. In the design of polyglycosylated DC-SIGN antagonists we made use of
105 dendrimeric core structures as valuable scaffolds for the multivalent presentation of ligands.

106 These architectures enabled efficient generation of monodisperse constructs with control over
107 valency and spatial arrangement. In particular, dendrimers were decorated with the optimized
108 monovalent glycomimetic ligands **1-3**, which have shown a significant affinity for DC-SIGN.
109 In a first attempt, we investigated how valency and the nature of the monovalent ligands
110 affect the affinity towards the receptor. We then tuned the stability, rigidity and length of the
111 scaffolds, in order to produce ligands able to match the distance between two contiguous DC-
112 SIGN binding sites, therefore being able to undergo a plurality of multivalent mechanisms
113 and enhance their bioactivity, while maintaining a moderate valency.

114

115 **Polyester (Boltorn-type) scaffolds**

116 The first class of multivalent glycocompounds was prepared in collaboration with the group
117 of Javier Rojo (CSIC-Seville) using 2,2-bis(hydroxymethyl)propionic acid (bis MPA) derived
118 polyester dendrons and dendrimers (Boltorn-type). Rather than using polydisperse
119 commercial scaffolds, the polyesters were prepared in a monodisperse manner. Both the
120 tetravalent dendron **4** and the 32-valent dendrimer **5** (Figure 4) were synthesized using bis
121 MPA as building block, and introducing carboxylic acid residues at the dendrimer periphery
122 by reaction with conveniently protected succinic acid moieties. Compounds **1** and **2**, both
123 tethered to an amino-terminated linker, were coupled to the dendrimers using HATU as the
124 activating agent, in the presence of *N,N*-diisopropylethylamine. Final compounds **4.1**, **4.2**, **5.1**
125 and **5.2** (Figure 4) were obtained in high yields; the tetravalent compounds were completely
126 monodisperse, whilst both **5.1** and **5.2** were loaded with 30-32 copies of the ligands, on
127 average²⁴.

128 These compounds were tested as DC-SIGN ligands in SPR competition assays. Results
129 showed a dependency on multivalent presentation, since the full dendrimers **5.1** and **5.2** (IC₅₀
130 = 1-2 μM) were two orders of magnitude more active than the corresponding tetravalent

131 dendrons **4** ($IC_{50} = 120-220 \mu M$). Surprisingly, though, the nature of the monovalent ligand
132 did not affect the affinity significantly, since constructs of similar valency showed similar
133 IC_{50} s, despite the one order of magnitude difference observed at the monovalent level
134 between **1** and **2**. The same phenomenon was observed in an Ebola infection model²⁴ and by
135 testing **4.1**, **4.2**, **5.1** and **5.2** in a cellular model of HIV infection¹⁸. These data prompted us to
136 reexamine in more detail the interaction of **2** with DC-SIGN by a number of biophysical
137 techniques, which allowed to conclude that the affinity measured for this molecule in our
138 initial experiments was largely determined by a clustering mechanism in solution (Figure 3c).
139 In other words, the two mannose units at both ends of **2** are able to interact simultaneously
140 with two DC-SIGN tetramers by simultaneously binding to one CRD per tetramer¹⁸. This
141 behavior leads to a high potency measured in all the experimental set ups where both the
142 protein tetramer and **2** are in the same solution (e.g. in SPR competition experiments and in
143 Isothermal Titration Calorimetry (ITC)). However, the clustering mechanism is lost when **2** is
144 conjugated to a polyvalent scaffold, which prevents the reducing end mannose from reaching
145 its target, and is also unlikely to be relevant in physiological conditions, where DC-SIGN is
146 bound to the cellular membrane. As a result, further studies on pseudo-glycodendrimers
147 targeted against DC-SIGN were mostly focused on the modified pseudo-dimannoside **3**.
148 From a practical point of view, the tetravalent dendrons **4.1** and **4.2** represent a good
149 compromise between affinity and synthetic effort. However, both the polyester dendron **4.1**
150 and **4.2** suffer from chemical instability, since the unhindered succinyl ester moieties used to
151 introduce carboxylic groups on the outer layer, are labile to nucleophiles, both under basic
152 and mild acidic catalysis. For instance, 30 % of **4.2** was hydrolyzed after 6 h in water solution
153 at physiological pH (7.4, PBS buffer, NMR investigation), highlighting its limit for *in vivo*
154 applications. MS analysis of the product confirmed that the hydrolysis occurred at the

155 succinyl ester bond. Moreover, this instability prevents purification by reverse phase
156 chromatography and hinders the scale up of the synthetic process.

157 In order to achieve stabilized versions of the hydrolytically unstable constructs **4** (Figure 4),
158 the synthesis of the analogous tetravalent dendron **4N** was undertaken (Scheme 1)²⁵. In **4N** the
159 labile succinyl ester bonds are replaced with more robust amide functionalities, that can be
160 installed starting from the tetramine **6** (Scheme 1). A diglycolate linker **7**, slightly longer and
161 more hydrophilic than succinate, was selected in this design, thus preserving the water
162 solubility of the molecule. Additionally, the length of the linker helped to minimize side
163 reactions due to attack of the scaffold nitrogen atom over the activated carboxylic acids
164 during the final functionalization of the dendron.

165 Condensation of the tetracid **4N** with the amino-tethered **1a** and **2a** led to dendrons **4N.1** and
166 **4N.2** (Figure 5), which were found to be fully stable both to chromatographic conditions and
167 in water solution at physiological pH. These dendrons have also been tested as ligands for
168 mouse Mannose Binding Lectin (MBL-C), a C-type lectin involved in the pathophysiology of
169 brain ischemia/reperfusion injury²⁶, and have shown activities similar to the lead **4.2**.

170 Additionally, functionalization of the dendron focal point with an azido-terminated tether was
171 adopted to allow the convergent synthesis of higher valency constructs. In particular,
172 exploiting Cu(I) catalyzed azide-alkyne cycloaddition (CuAAC), **4N.1** was further connected
173 to a pentaerythritol derived scaffold **8** (Scheme 2). The resulting 16-valent compound **4.4N.1**
174 was almost 2-orders of magnitude more active than its precursor in an MBL-C binding
175 assay²⁵.

176

177 **Polyalkyne terminated scaffolds**

178 In a different approach, to further improve the synthetic accessibility and chemical stability of
179 the dendrimers, new polyalkyne terminated scaffolds, with a valency ranging from 2 to 6,

180 were synthesized²⁷. They were then decorated with azide-bearing ligands **1-3**, or with
181 mannose as a control, through CuAAC. Nondegradable glycodendrons and glycodendrimers
182 with a valency of 3, 4 or 6 (**9-11**, Figure 6) were obtained, characterized by stable ether bonds
183 and triazole connectors. In a convergent approach, the trivalent dendrons **9**, bearing an azido
184 group at the focal point, were used to build higher valency constructs (6-, 9-, 12- and 18-
185 valent **12.9**, **13.9**, **10.9** and **11.9**) by combination with either the divalent, tetravalent or
186 hexavalent scaffolds **12**, **13**, **10** and **11** (Figure 6). CuAAC allowed a full control over the
187 targeted structure: all products were characterized by NMR spectroscopy and MALDI-MS
188 analysis, and were stable for months in water solution.

189 Compounds affinity for DC-SIGN was evaluated through SPR inhibition assays and the
190 relative inhibitory potency (R.I.P.) values were compared (Figure 7, Relative inhibitory
191 potencies (R.I.P.) are valency-corrected and calculated according as: $\frac{IC_{50,monovalent}}{IC_{50,multivalent} \times valency}$).

192 Comparing constructs of the same valency, glycodendrimers bearing the monovalent ligand **3**
193 had lower IC₅₀ values than the ones carrying **1**, and all of them were more active than the
194 simply mannosylated analogs (Figure 7) . Once again, dendrimers based on **2** behaved
195 similarly to the corresponding derivatives of **1**; moreover, the tetravalent and the hexavalent
196 compounds **10.2** and **11.2** showed a R.I.P. < 1. This confirmed that the high activity of the
197 monovalent ligand **2** is due to an outperforming binding mode not allowed for its multivalent
198 derivatives. On the contrary, the activity of the multivalent presentations of compounds **1** and
199 **3** gradually increased by incrementing the valency. Remarkably, R.I.P. values obtained for
200 dendrimers based on **3** grew faster with valency than those shown by the corresponding
201 dendrimers based on **1** or on mannose (Figure 7). These results highlight that affinity
202 differences at the monovalent level are amplified by multivalent presentation and support the
203 importance of optimizing the structure of monovalent ligands. Tetravalent **10.1** and **10.2** had
204 an analogous affinity for DC-SIGN with respect to **4.1** and **4.2**, suggesting that, being the

205 valency equal, the shape of this type of dendrimers does not influence their activity to a
206 significant extent. .

207 Compounds **10.1**, **10.3** and **11.3** were tested also in cellular infection models, by evaluating
208 their ability to inhibit the DC-SIGN-mediated HIV infection of CD4⁺ T-cells. Also in this
209 case, the activity of the monovalent ligand had a positive role, since **10.3** was more active
210 than **10.1**. The importance of the valency was also confirmed: the hexavalent compound **11.3**
211 was around 10 times more active than the tetravalent **10.3**. Remarkably, despite the good
212 results shown by glycodendrimers bearing mimic **3**, a further valency increase, as in **13.9.3**
213 (Figure 6), was not practical, due to low water solubility of the constructs.

214 Molecular dynamic simulations were used to estimate the maximum distance spanned by
215 dendrimers **10.1** and **11.1** (as simpler models of **10.3** and **11.3**, respectively), which in turn
216 allowed to dissect the possible mechanisms of interaction between the glycodendrimers and
217 DC-SIGN. Since the calculated maximum extension of **11.1** was 35.4 Å and the distance
218 between two contiguous DC-SIGN binding sites in one tetramer is at least 38 Å²⁸, the
219 chelation mechanism was excluded. Remarkably, the hexavalent compound **11.1** had an IC₅₀
220 value comparable to that of **12.9.1**, which is also hexavalent, but is built on a longer PEG
221 core, suggesting that a flexible linker is not suitable to make the active ligands assuming the
222 proper conformation for chelation.

223

224 **Phenylene-ethynylene rod-like scaffolds**

225 With the goal of further improving dendrimers activity by exploiting the chelation
226 mechanism, the length and the rigidity of the scaffolds were optimized. In a collaboration
227 with the group of Roland J. Pieters (Utrecht University), three phenylene-ethynylene rod-like
228 spacers of different lengths (i.e. 8, 12.5 and 22 Å) were synthesized through multiple
229 Sonogashira reactions and deprotection steps²⁹. Their terminal triple bonds were exploited to

230 connect the glycomimetic moieties by CuAAC reactions. To this aim, trivalent dendrons **9.1**
231 and **9.3** were employed, as well as the monovalent ligands **1** and **3** and their elongated
232 versions, characterized by a PEG linker having the same length of the dendron chain³⁰. The
233 structures of the obtained hexavalent dendrimers (**14.9.1**, **15.9.1**, **16.9.1**, **14.9.3**, **15.9.3**,
234 **16.9.3**) and divalent controls (**16.1**, **16.3** and **14.1L**, **15.1L**, **16.1L**, **14.3L**, **15.3L**, **16.3L**) are
235 depicted in Figure 8.

236 Molecular dynamics simulations showed that all these constructs have the potential to chelate
237 two contiguous DC-SIGN binding sites: at maximum extension both the shortest dendrimer
238 **14.9.1** and the longest one **16.9.1** are longer than 4 nm. As before, SPR competition assays
239 were performed to measure their IC₅₀ and R.I.P. values (Figure 9). The hexavalent derivatives
240 of **3** (**14.9.3**, **15.9.3** and **16.9.3**) could not be evaluated appropriately in this format, because
241 their affinity reached the lower limit of the assay (Figure 9). Nonetheless, valuable
242 information could be gathered concerning the effect of the spacers on the dendrimers'
243 activity.

244 As expected, compounds based on the more active ligand **3** outperformed those based on **1**
245 (see, for instance, divalent **16.1** and **16.3**). The bioactivity was also clearly dependent on the
246 valency, since hexavalent compounds were more active than the corresponding divalent ones
247 (compare **16.9.1** and **16.1**), demonstrating the positive effect of increasing the local ligand
248 concentration. Entropic penalties were paid by the structures characterized by the flexible
249 PEG linker (**1.L** and **3.L** derivatives); indeed, **16.1** and **16.3** were about two times more active
250 than **16.1.L** and **16.3.L**, respectively. Finally, IC₅₀s decreased regularly with the length of the
251 rod, suggesting that the chelation mechanism may become increasingly accessible.

252 For the hexavalent derivatives of **3** a more sensitive cellular HIV infection study was
253 performed, revealing again a clear dependency on the length of the rod core. In particular, the
254 bioactivity increased by about one order of magnitude from **11.3** to **14.9.3**, by simply

255 inserting the shortest rod as the dendrimer core, and it was highest for the longest **16.9.3**. The
256 valency effect was also confirmed, since hexavalent compounds were more active than the
257 divalent ones. IC₅₀ values were measured for **14.9.3**, **16.9.3**, **16.3** as 67 nM, 24 nM and 161
258 nM, respectively. To the best of our knowledge, **16.9.3** is one of the most potent inhibitors of
259 the DC-SIGN-mediated *trans* HIV infection described up to date, despite its relatively
260 moderate valency and strengthened by its controlled and well characterized structure. This
261 result clearly highlights how the combination of optimized monovalent ligands with fine-
262 tuned multivalent constructs of controllable size can maximize the effect of valency when
263 targeting a multivalent receptor.

264 Remarkably, even if these dendrimer-like compounds, bearing a lipophilic core and
265 hydrophilic carbohydrate groups, are endowed with amphiphilic properties, they do not
266 significantly aggregate in aqueous solution, as shown using Dynamic Light Scattering,
267 Analytical Ultracentrifugation and a number of other biophysical techniques³¹. Hence, their
268 performances in infection studies can be correctly interpreted based on their individual
269 structure.

270

271 **Biological characterization**

272 Three of the dendrimers described above, **4.2**³², **11.3**³³ and **16.9.3**³⁴, underwent further
273 biological assays to achieve a fuller characterization of their effect. Importantly, none of the
274 compounds was found to be toxic against the tested cell lines, and both **4.2** and **11.3** did not
275 alter the viability of human cervical explants up to a concentration of 1 mM.

276 In particular, dendron **4.2** and dendrimer **11.3** were tested as inhibitors of HIV infection on
277 explants obtained from human uterine cervix, to achieve a better approximation of *in vivo*
278 conditions. To this aim, tissue explants were exposed to different types of R5 tropic HIV-1,
279 including the laboratory adapted BaL and clinical primary isolates, in the presence of

280 glycoderivatives. Infection was reduced by more than 85 % with 1 mM **4.2** and 0.5 mM **11.3**,
281 demonstrating that these inhibitors are good precursors to be further optimized and formulated
282 as microbicide drugs.

283 It is well-known that the DC-SIGN binding to a ligand can activate and regulate the immune
284 response³⁵, therefore DC-SIGN binders are also studied for the development of vaccines and
285 immunomodulants. The activation of DC-SIGN signaling by the dendrimers synthesized in
286 our group was investigated to assess if they stimulate the production of factors able to
287 interfere with HIV infection and if they are suitable as immunomodulants and vaccine
288 adjuvants. In this study, the expression of cytokines, chemokines and genes involved in
289 immune responses was evaluated after immature Monocyte Derived Dendritic Cells
290 (iMDDCs) interaction with DC-SIGN antagonists. Preliminarily, the behavior of **4.2** was
291 explored³², showing that it induced an increase of the production of β chemokines known to
292 suppress HIV-1 R5 tropic strain replication³⁶. A similar activity was observed for **11.3**³³ and
293 **16.9.3**³⁴, that also stimulated an impressive production of antiviral and pro-inflammatory
294 cytokines, as well as of co-stimulatory molecules having a role in the activation of naïve T
295 cells. Additional cytokines and proteins that promote Dendritic Cells differentiation,
296 development, survival and activation were overexpressed by treating iMDDCs with **16.9.3**.
297 Although a direct comparison between the effects of the tested compounds could not be
298 drawn, since experiments were conducted on different healthy donors, using different
299 techniques and experimental settings, it was noticed that the inflammatory response
300 stimulated by **16.9.3** was less pronounced than that caused by **8.3**. This is an important
301 element to acquire, since an excessive immune response can result in tissue damage (and even
302 death). It was therefore concluded that these compounds and, in particular, the most active
303 DC-SIGN antagonist **16.9.3**, have a potential use as adjuvants and immunostimulants in the

304 formulation of vaccines for HIV and, generally, mucosal pathogens. Further studies in both *ex*
305 *vivo* and *in vivo* models will be required.

306 Exploiting the intrinsic fluorescence of its rod core, the cellular uptake and internalization
307 route of **16.9.3** within human iMDDCs was investigated by means of fluorescence
308 microscopy at varying time and temperature intervals³⁴. It was observed that **16.9.3** is
309 internalized by DCs already after 10 min of incubation at room temperature, but not at 4 °C,
310 an indication that uptake occurs through an energy-dependent mechanism, such as the
311 receptor-mediated one.

312 Finally, confocal images with fluorescent tracers used to mark specific sub-cellular organelles
313 revealed that **16.9.3** is internalized mainly to lysosomes and that it also transits through and
314 resides in early endosomes for a quite prolonged period of time. These pulse-chase
315 experiments showed that the dendrimer uptake is rapid and confirmed that the molecule has
316 the potential to elicit an immune response that may be exploited for the development of
317 vaccines.

318

319 **Summary and outlook**

320 We have here presented the results of our search for efficient antagonists of DC-SIGN, a C-
321 type lectin recognizing highly mannosylated structures, and involved in the recognition
322 process of pathogens such as HIV, Ebola and Dengue viruses. Rather than directly using
323 mannose- or oligomannose derivatives, our strategy has first been devoted to the optimization
324 of glycomimetic monovalent DC-SIGN ligands. We have then combined the most promising
325 monovalent ligands with several types of dendrimer-like scaffolds, to produce multivalent
326 materials targeted against C-type lectins and in particular DC-SIGN. The activity trend of the
327 monovalent ligands was reflected in the bioactivity of the dendrimers, thus highlighting the
328 importance of ligand optimization. Also the valency influenced dendrimers activity; indeed,

329 among each dendrimer sub-class, the activity generally increased by increasing the valency.
330 The size of the dendrimers was finally tuned using rigid rod-like cores of controlled length, in
331 order to control the relative orientation and the distance spanned by the active ligands. In this
332 way, we synthesized a class of dendrimer-like molecules that can reach simultaneously two
333 adjacent DC-SIGN binding sites within the tetramer carrying to each site a small cluster of
334 glycomimetic ligands. These systems, thus, can exploit more than one multivalency
335 mechanism (e.g. chelation, statistical rebinding), which results in a major potency
336 enhancement compared to shorter or more flexible constructs. With an appropriate
337 combination of monovalent ligands, rod-core and valency, the hexavalent compound **16.9.3**,
338 one of the most potent inhibitors of the DC-SIGN-mediated HIV infection up to date, was
339 obtained. This result showed how the rational design of artificial ligands helps in obtaining
340 powerful compounds while retaining a moderate valency, which allows to deal with perfectly
341 monodisperse and well-characterized species. DC-SIGN antagonists such as **16.9.3** are
342 effective precursors for further derivatizations and appropriate formulations, with the aim of
343 producing topical microbicides as pathogen-entry inhibitors. Moreover, we have observed that
344 **16.9.3** is internalized by dendritic cells and routed to endolysosomal compartments. These
345 features, together with its ability to induce the overexpression of signaling molecules involved
346 in immune responses, make **16.9.3** interesting as an immunomodulator and vaccine adjuvant.

347

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353

354 **References**

- 355 (1) www.unaids.org Web. Accessed on 10 Jan 2017.
- 356 (2) Wang, H. B.; Mo, Q. H.; Yang, Z. *J. Immunol. Res.* **2015**, *2015*, 503978.
- 357 (3) Pan, X.; Baldauf, H. M.; Keppler, O. T.; Fackler, O. T. *Cell Res.* **2013**, *23*, 876.
- 358 (4) Szunerits, S.; Barras, A.; Khanal, M.; Pagneux, Q.; Boukherroub, R. *Molecules* **2015**,
359 *20*, 14051.
- 360 (5) Sepulveda-Crespo, D.; Cena-Diez, R.; Jimenez, J. L.; Angeles Munoz-Fernandez, M.
361 *Med. Res. Rev.* **2017**, *37*, 149.
- 362 (6) Gringhuis, S. I.; den Dunnen, J.; Litjens, M.; van Het Hof, B.; van Kooyk, Y.;
363 Geijtenbeek, T. B. *Immunity* **2007**, *26*, 605.
- 364 (7) van Kooyk, Y.; Geijtenbeek, T. B. *Nat. Rev. Immunol.* **2003**, *3*, 697.
- 365 (8) Tabarani, G.; Reina, J. J.; Ebel, C.; Vives, C.; Lortat-Jacob, H.; Rojo, J.; Fieschi, F.
366 *FEBS Lett.* **2006**, *580*, 2402.
- 367 (9) Reina, J. J.; Rojo, J. *Braz. J. Pharm. Sci.* **2013**, *49*, 109.
- 368 (10) Sattin, S.; Bernardi, A. *Trends Biotechnol.* **2016**, *34*, 483.
- 369 (11) Adams, E. W.; Ratner, D. M.; Bokesch, H. R.; McMahon, J. B.; O'Keefe, B. R.;
370 Seeberger, P. H. *Chem. Biol.* **2004**, *11*, 875.
- 371 (12) Shahzad-Ul-Hussan, S.; Sastry, M.; Lemmin, T.; Soto, C.; Loesgen, S.; Scott, D. A.;
372 Davison, J. R.; O'Connor, R.; Kwong, P. D.; Bewley, C. A. *ChemBioChem* **2017**.
- 373 (13) Mari, S.; Posterl, H.; Marcou, G.; Potenza, D.; Micheli, F.; Cañada, F. J.; Jimenez-
374 Barbero, J.; Bernardi, A. *Eur. J. Org. Chem.* **2004**, *2004*, 5119.
- 375 (14) Reina, J. J.; Sattin, S.; Invernizzi, D.; Mari, S.; Martinez-Prats, L.; Tabarani, G.;
376 Fieschi, F.; Delgado, R.; Nieto, P. M.; Rojo, J.; Bernardi, A. *ChemMedChem* **2007**, *2*, 1030.
- 377 (15) Mari, S.; Sanchez-Medina, I.; Mereghetti, P.; Belvisi, L.; Jimenez-Barbero, J.;
378 Bernardi, A. *Carbohydr. Res.* **2007**, *342*, 1859.

379 (16) Sattin, S.; Daggetti, A.; Thepaut, M.; Berzi, A.; Sanchez-Navarro, M.; Tabarani, G.;
380 Rojo, J.; Fieschi, F.; Clerici, M.; Bernardi, A. *ACS Chem. Biol.* **2010**, *5*, 301.

381 (17) Thepaut, M.; Guzzi, C.; Sutkeviciute, I.; Sattin, S.; Ribeiro-Viana, R.; Varga, N.;
382 Chabrol, E.; Rojo, J.; Bernardi, A.; Angulo, J.; Nieto, P. M.; Fieschi, F. *J. Am. Chem. Soc.*
383 **2013**, *135*, 2518.

384 (18) Sutkeviciute, I.; Thepaut, M.; Sattin, S.; Berzi, A.; McGeagh, J.; Grudinin, S.; Weiser,
385 J.; Le Roy, A.; Reina, J. J.; Rojo, J.; Clerici, M.; Bernardi, A.; Ebel, C.; Fieschi, F. *ACS*
386 *Chem. Biol.* **2014**, *9*, 1377.

387 (19) Varga, N.; Sutkeviciute, I.; Guzzi, C.; McGeagh, J.; Petit-Haertlein, I.; Gugliotta, S.;
388 Weiser, J.; Angulo, J.; Fieschi, F.; Bernardi, A. *Chem. - Eur. J.* **2013**, *19*, 4786.

389 (20) Lundquist, J. J.; Toone, E. J. *Chem. Rev.* **2002**, *102*, 555.

390 (21) Pieters, R. J. *Org. Biomol. Chem.* **2009**, *7*, 2013.

391 (22) Renaudet, O.; Roy, R. *Chem. Soc. Rev.* **2013**, *42*, 4515.

392 (23) Chabre, Y. M.; Roy, R. *Adv. Carbohydr. Chem. Biochem.* **2010**, *63*, 165.

393 (24) Luczkowiak, J.; Sattin, S.; Sutkeviciute, I.; Reina, J. J.; Sanchez-Navarro, M.;
394 Thepaut, M.; Martinez-Prats, L.; Daggetti, A.; Fieschi, F.; Delgado, R.; Bernardi, A.; Rojo, J.
395 *Bioconjug. Chem.* **2011**, *22*, 1354.

396 (25) Goti, G.; Palmioli, A.; Stravalaci, M.; Sattin, S.; De Simoni, M. G.; Gobbi, M.;
397 Bernardi, A. *Chem. - Eur. J.* **2016**, *22*, 3686.

398 (26) Gesuete, R.; Storini, C.; Fantin, A.; Stravalaci, M.; Zanier, E. R.; Orsini, F.; Vietsch,
399 H.; Mannesse, M. L.; Ziere, B.; Gobbi, M.; De Simoni, M. G. *Ann. Neurol.* **2009**, *66*, 332.

400 (27) Varga, N.; Sutkeviciute, I.; Ribeiro-Viana, R.; Berzi, A.; Ramdasi, R.; Daggetti, A.;
401 Vettoretti, G.; Amara, A.; Clerici, M.; Rojo, J.; Fieschi, F.; Bernardi, A. *Biomaterials* **2014**,
402 *35*, 4175.

- 403 (28) Tabarani, G.; Thepaut, M.; Stroebel, D.; Ebel, C.; Vives, C.; Vachette, P.; Durand, D.;
404 Fieschi, F. *J. Biol. Chem.* **2009**, *284*, 21229.
- 405 (29) Pertici, F.; Varga, N.; van Duijn, A.; Rey-Carrizo, M.; Bernardi, A.; Pieters, R. J.
406 *Beilstein J. Org. Chem.* **2013**, *9*, 215.
- 407 (30) Ordanini, S.; Varga, N.; Porkolab, V.; Thepaut, M.; Belvisi, L.; Bertaglia, A.;
408 Palmioli, A.; Berzi, A.; Trabattoni, D.; Clerici, M.; Fieschi, F.; Bernardi, A. *Chem. Commun.*
409 **2015**, *51*, 3816.
- 410 (31) Ordanini, S.; Zanchetta, G.; Porkolab, V.; Ebel, C.; Fieschi, F.; Guzzetti, I.; Potenza,
411 D.; Palmioli, A.; Podlipnik, C.; Meroni, D.; Bernardi, A. *Macromol. Biosci.* **2016**, *16*, 896.
- 412 (32) Berzi, A.; Reina, J. J.; Ottria, R.; Sutkeviciute, I.; Antonazzo, P.; Sanchez-Navarro,
413 M.; Chabrol, E.; Biasin, M.; Trabattoni, D.; Cetin, I.; Rojo, J.; Fieschi, F.; Bernardi, A.;
414 Clerici, M. *AIDS* **2012**, *26*, 127.
- 415 (33) Berzi, A.; Varga, N.; Sattin, S.; Antonazzo, P.; Biasin, M.; Cetin, I.; Trabattoni, D.;
416 Bernardi, A.; Clerici, M. *Viruses* **2014**, *6*, 391.
- 417 (34) Berzi, A.; Ordanini, S.; Joosten, B.; Trabattoni, D.; Cambi, A.; Bernardi, A.; Clerici,
418 M. *Sci. Rep.* **2016**, *6*, 35373.
- 419 (35) Svajger, U.; Anderluh, M.; Jeras, M.; Obermajer, N. *Cell. Signalling* **2010**, *22*, 1397.
- 420 (36) Cocchi, F.; DeVico, A. L.; Garzino-Demo, A.; Arya, S. K.; Gallo, R. C.; Lusso, P.
421 *Science* **1995**, *270*, 1811.

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425 **Figure captions**

426 Figure 1: Chemical structure of (Man)₉(GlcNAc)₂ glycan; the disaccharide Man α 1-2Man and
427 the trisaccharide Man α 1-2 Man α 1-6 Man α are encircled.

428 Figure 2: Structure of the synthesized pseudo-mannosylated compounds: a) pseudo-
429 disaccharide **1** mimics the Man α 1-2Man disaccharide; b) pseudo-trisaccharide **2** mimics the
430 Man α 1-2Man α 1-6Man trisaccharide; c) bis-amide-substituted pseudo-disaccharide **3**.

431 Figure 3: Possible affinity-enhancement mechanisms occurring between a multivalent ligand
432 (black) and a multivalent protein (gray): a) chelation; b) statistical rebinding; c) protein
433 aggregation.

434 Figure 4: Structure of scaffolds **4** and **5** and the corresponding tetravalent **4.1** and **4.2**
435 glycosylated dendrons and 32-valent glycosylated **5.1** and **5.2** dendrimers.

436 Figure 5: Structure of tetravalent dendrons **4N.1** and **4N.2**, stabilized by replacing the succinyl
437 ester bond with an amido bond.

438 Figure 6: Structure of the glycodendrimers based on monovalent mannose, **1**, **2** or **3** and
439 multivalent polyalkyne terminated scaffolds.

440 Figure 7: IC₅₀ (empty white bars) and relative inhibitory potency (R.I.P.) (black bars) values
441 of **1**, **2**, **3** and **mannose** multivalent derivatives, tested as DC-SIGN binders through SPR
442 competition assays.

443 Figure 8: Structure of the glycodendrimers based on monovalent ligands **1** or **3** and rigid
444 phenylene-ethynylene scaffolds.

445 Figure 9: IC₅₀ (empty white bars) and relative inhibitory potency (R.I.P.) (black bars) values
446 of rod-based derivatives of **1** and **3**, tested as DC-SIGN binders by SPR competition assays.

447 Scheme 1: Synthesis of tetramine **6** and of the tetravalent dendron **4N**

448 Scheme 2: Synthesis of the 16-valent compound **4.4N.1** from dendron **4N.1**.

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