



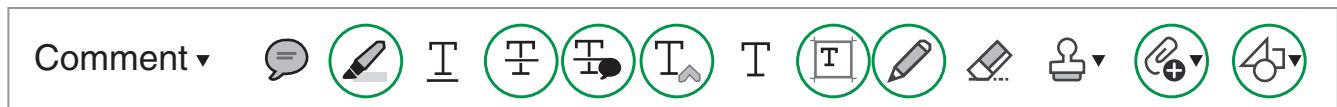
Page Proof Instructions and Queries

Journal Title: European Journal of Preventive Cardiology (CPR)

Article Number: 831500

Thank you for choosing to publish with us. This is your final opportunity to ensure your article will be accurate at publication. Please review your proof carefully and respond to the queries using the circled tools in the image below, which are available by clicking “Comment” from the right-side menu in Adobe Reader DC.*

Please use *only* the tools circled in the image, as edits via other tools/methods can be lost during file conversion. For comments, questions, or formatting requests, please use  Please do *not* use comment bubbles/sticky notes .



*If you do not see these tools, please ensure you have opened this file with Adobe Reader DC, available for free at get.adobe.com/reader or by going to Help > Check for Updates within other versions of Reader. For more detailed instructions, please see us.sagepub.com/ReaderXProofs.

No.	Query
	Please confirm that all author information, including names, affiliations, sequence, and contact details, is correct.
	Please review the entire document for typographical errors, mathematical errors, and any other necessary corrections; check headings, tables, and figures.
	Please confirm that the Funding and Conflict of Interest statements are accurate.
	Please ensure that you have obtained and enclosed all necessary permissions for the reproduction of artistic works, (e.g. illustrations, photographs, charts, maps, other visual material, etc.) not owned by yourself. Please refer to your publishing agreement for further information.
	Please note that this proof represents your final opportunity to review your article prior to publication, so please do send all of your changes now.
AQ: 1	Please check the author affiliations which are amended in the journal format of department, organization and country.
AQ: 2	Please check that the full postal address is provided for the corresponding author.
AQ: 3	Please confirm the keywords.
AQ: 4	Please define the abbreviation ATHEROREMO-IVUS in the text on first use.
AQ: 5	Please define VFAQ on first use in the text and in the Figure 1 caption.
AQ: 6	Please define INSIG in the text.
AQ: 7	Please define COPII in the text.
AQ: 8	Please define MBD2 in the text.
AQ: 9	Please check the reference for Kera et al. in the text as the author name does not correspond?
AQ: 10	Please define Rac1-GTP in the text.
AQ: 11	Please define HepG2 and Caco-2.
AQ: 12	Please review the last sentence of the section on Ezetimibe as it is not clearly understood what is meant by ‘if it is indisputable’ - do you mean ‘although it is undisputable’ or something else?
AQ: 13	Please define APOE in the text.
AQ: 14	Please define DPA in the text.
AQ: 15	Please check the referencing for the sentence beginning ‘Initially, this hypothesis’ in the section on red yeast rice as two studies are mentioned but only one is cited?

AQ: 16	Please define ESC/EAS in the text.
AQ: 17	Please define RR on first use in the text.
AQ: 18	Please define IVUS in the text.
AQ: 19	Please clarify the meaning of 'reduced twice' - do you mean 'halved'?
AQ: 20	Please define ACS.
AQ: 21	Please define OCT.
AQ: 22	Please define ORION.
AQ: 23	Please define GlycA and NMR.
AQ: 24	Please check that the author/organization name is correct in reference 2.
AQ: 25	Please check the journal volume number and page range in reference 3.
AQ: 26	Please check the epublication date in reference 4.
AQ: 27	Please check that the author/organization name is correct in reference 8.
AQ: 28	Please check the journal volume number and page range in reference 14.
AQ: 29	Please check the year, journal volume number and page range in reference 23.
AQ: 30	Please check the page range in reference 94.
AQ: 31	Please check the page range in reference 115.
AQ: 32	Please check the article title for reference 117 as it is identical to that for reference 116?
AQ: 33	Please check the epublication date in reference 124.
AQ: 34	Please provide the date of epublication in reference 126.
AQ: 35	Please check the epublication date in reference 131.
AQ: 36	Please check the journal volume number and page range in reference 132.
AQ: 37	Please check the epublication date in reference 133.
AQ: 38	Please check the epublication date in reference 135.
AQ: 39	Please check the journal volume number and page range in reference 136.
AQ: 40	Please check the journal volume number and page range in reference 143.
AQ: 41	Please check the journal volume number and page range in reference 144.
AQ: 42	Please check the epub before print date in reference 151.
AQ: 43	Please check the epub before print date in reference 158.
AQ: 44	Please check the journal volume number and page range in reference 159.
AQ: 45	Please provide the journal volume number in reference 162.
AQ: 46	Please check the journal volume number and page range in reference 164.
AQ: 47	Please check the journal volume number and page range in reference 165.
AQ: 48	Please check the journal volume number and page range in reference 173.
AQ: 49	Please check the journal volume number and page range in reference 174.
AQ: 50	Please check that all abbreviations in Figure 1 are correctly defined in the caption.
AQ: 51	Please check throughout the text as some changes have been made for consistency of use of abbreviations and layout. Please check the definition of ODYSSEY in the abstract and on first use in the text.

Changes in circulating pro-protein convertase subtilisin/kexin type 9 levels – experimental and clinical approaches with lipid-lowering agents

C Macchi¹, M Banach^{2,3,4}, A Corsini^{1,5}, CR Sirtori⁶, N Ferri⁷ and M Ruscica¹

Abstract

Regulation of pro-protein convertase subtilisin/kexin type 9 (PCSK9) by drugs has led to the development of a still small number of agents with powerful activity on low-density lipoprotein cholesterol levels, associated with a significant reduction of cardiovascular events in patients in secondary prevention. The Further Cardiovascular Outcomes Research with PCSK9 Inhibition in Subjects with Elevated Risk (FOURIER) and Evaluation of Cardiovascular Outcomes After an Acute Coronary Syndrome During Treatment With Alirocumab (ODYSSEY [AQ51] OUTCOMES) studies, with the two available PCSK9 antagonists, i.e. evolocumab and alirocumab, both reported a 15% reduction in major adverse cardiovascular events. Regulation of PCSK9 expression is dependent upon a number of factors, partly genetic and partly associated to a complex transcriptional system, mainly controlled by sterol regulatory element binding proteins. PCSK9 is further regulated by concomitant drug treatments, particularly by statins, enhancing PCSK9 secretion but decreasing its stimulatory phosphorylated form (S688). These complex transcriptional mechanisms lead to variable circulating levels making clinical measurements of plasma PCSK9 for cardiovascular risk assessment a debated matter. Determination of total PCSK9 levels may provide a diagnostic tool for explaining an apparent resistance to PCSK9 inhibitors, thus indicating the need for other approaches. Newer agents targeting PCSK9 are in clinical development with a major interest in those with a longer duration of action, e.g. RNA silencing, allowing optimal patient compliance. Interest has been expanded to areas not only limited to low-density lipoprotein cholesterol reduction but also investigating other non-lipid pathways raising cardiovascular risk, in particular inflammation associated to raised high-sensitivity C-reactive protein levels, not significantly affected by the present PCSK9 antagonists.

Keywords

Alirocumab, evolocumab, inclisiran, pro-protein convertase subtilisin/kexin type 9 circulating levels, pro-protein convertase subtilisin/kexin type 9 [AQ3]

Received 13 November 2018; accepted 25 January 2019

Introduction

The reduction of low-density lipoprotein cholesterol (LDL-C) has been an extensively evaluated pharmacological target. LDL-C is the modifiable risk factor associated with cardiovascular (CV) risk that has been thoroughly investigated in the last decades, with an outstanding number of therapeutic options.¹ Analysis of large primary and secondary prevention patients has generally indicated that a 38.6 mg/dl LDL-C reduction is associated with, roughly, a 22% lowering of CV risk.² This benefit is consistent also in

¹Dipartimento di Scienze Farmacologiche e Biomolecolari, Università degli Studi di Milano, Italy

²Department of Hypertension, Medical University of Lodz, Poland

³Polish Mother's Memorial Hospital Research Institute (PMMHRI), Poland

⁴Cardiovascular Research Centre, University of Zielona Gora, Poland

⁵Multimedica IRCCS, Italy

⁶Dyslipidemia Center, A.S.S.T. Grande Ospedale Metropolitano Niguarda, Italy

⁷Dipartimento di Scienze del Farmaco, Università degli Studi di Padova, Italy [AQ1]

Corresponding author:

Nicola Ferri, Dipartimento di Scienze del Farmaco, Università degli Studi di Padova Padua, Italy. [AQ2]

Email: nicola.ferri@unipd.it

patients starting with LDL-C levels as low as a median of 63 mg/dl and achieving levels as low as a median of 21 mg/dl.³ The relevance of LDL-C in the initiation and progression of atherosclerosis is also based on ultrasound studies, e.g. those evaluating carotid intima media thickness as a non-invasive tool to assess CV risk; indeed, the rate of atherosclerotic plaque progression is directly linked to the absolute LDL-C level.⁴ Very recently, a person's total atherosclerotic plaque burden has been associated with his/her cumulative LDL exposure evaluated in mg-years by means of the following formula: age \times LDL-C, measured in mg/dl.⁵

On this background of epidemiological and pharmacological findings, new approaches have been investigated in order to further reduce LDL-cholesterolaemia and CV risk.⁶ In 2006, the Atherosclerosis Risk in Communities (ARIC) epidemiological study demonstrated that 'loss of function' mutations of pro-protein convertase subtilisin/kexin type 9 (PCSK9) were associated with lower levels of LDL-C (–28%, –37 mg/dl) and an astonishing 88% risk reduction in myocardial infarction (MI), fatal coronary heart diseases (CHDs) or coronary revascularization, despite a significant prevalence of other non-lipid-related CV-risk factors.⁷

More importantly, a genome-wide association study led to the identification of nine CHD/MI susceptibility loci including PCSK9.⁸ This finding, together with the discovery in 2003 of a missense 'gain of function' mutation in PCSK9, directly linked to LDL-C,⁹ led to the identification of PCSK9 as a new pharmacological target and paved the way for the development of a number of PCSK9 antagonists,¹⁰ e.g. monoclonal antibodies (mAbs) and small interfering RNA (siRNA).

Beyond cholesterol metabolism, a direct role of PCSK9 in atherosclerotic plaque formation has been also identified. The ATHEROREMO-IVUS study showed that higher serum PCSK9 levels were linearly associated with a higher necrotic core fraction in coronary atherosclerosis, regardless of serum LDL-C,¹¹ confirming data from our group in PCSK9 knock-out (KO) mice partially protected from neointimal formation.¹² [AQ4] A significant association between PCSK9 levels and carotid intima-media thickening¹³ as well as with arterial stiffness¹⁴ and liver fat accumulation¹⁵ was also shown. PCSK9 expression has been described in different cell types involved in atherosclerosis, e.g. endothelial and smooth muscle cells¹⁶ and macrophages.¹⁷ Circulating platelets were added to the list of PCSK9 targets,¹⁸ together with the description of pro-inflammatory effects on macrophages¹⁹ and the involvement in aortic valve calcification.²⁰

The clinical relevance of measuring circulating PCSK9 plasma levels may be hindered by the complexity of PCSK9 biology at the transcriptional and translational levels,²¹ leading to concentrations varying over

approximately a 100-fold range.²² Indeed, PCSK9 expression and plasma levels are tightly regulated by nutritional and hormonal status, e.g. hepatic glucagon receptor signaling²³ as well as by a diurnal rhythm.²⁴ Thus, in order to assess whether PCSK9 levels may contribute to risk prediction, even in patients already on preventive pharmacological therapy, i.e. on statins, it is of major interest to better understand whether drugs affecting lipid metabolism modulate PCSK9 gene expression and protein levels.

The present review article is aimed at covering major biological and clinical aspects of PCSK9 activity and antagonism. Discussion of studies investigating the mechanistic aspects of PCSK9 activity will be followed by detailed evaluation of the impact of lipid-lowering drugs and nutraceuticals on PCSK9 regulation. These different aspects of the present role of PCSK9 in system biology and in atherosclerosis induction/prevention will thus allow a potential forecast of events leading to more effective drug utilization in CV disease prevention, as well indications of possible new approaches to this therapeutic area.

The biology

PCSK9 is the ninth member and last-discovered protein belonging to the family of mammalian proprotein convertases (PCs), i.e. endoprotease enzymes. The first seven proteins (PCSK1–PCSK7) are structurally and biochemically similar to each other and to bacterial and yeast proteins-subtilisin and kexin, respectively - from which they probably derive.²⁵ A further classification is based on the predilection for the substrate cleavage motif which leads to precursor proteolysis;²⁶ the first seven PCSKs cleave precursors at basic residues, whereas subtilisin kexin isozyme 1/site 1 protease (SKI-1/S1P), the eighth member, does not require basic residues for cleavage and finally PCSK9 cleaves itself at its internal VFAQ₁₅₂ sequence.²⁷ [AQ5]

PCSK9 acts as a protein binding to specific cell surface receptors.²⁸ PCSK9 possesses the unique feature of being unable to get rid of its prodomain, thus leading to a secreted inactive protease. Produced in the endoplasmic reticulum (ER), PCs navigate through the secretory pathway to their final destination, which is one of the Golgi, the trans-Golgi network, secretory granules, endosomes, the plasma membrane, the extracellular matrix and the cell surface.

Similar to other members of the proprotein convertase family, PCSK9 shares a N-terminal prodomain, a subtilisin-like catalytic domain and a cysteine- and histidine-rich domain. Once in the ER, the signal peptide is cleaved and the zymogen proPCSK9 (aa 31–692, about 74 kDa) undergoes an autocatalytic cleavage of the prodomain (aa 31–152 at VFAQ₁₅₂: S1P),^{29,30}

this last remains firmly attached to its catalytic pocket by strong noncovalent interactions, thereby preventing any trans-catalytic activity of the enzyme. This phenomenon is a prerequisite for the release of the mature PCSK9 (aa 153–692, about 62 kDa) from the ER to the Golgi, likely because the prodomain serves as a chaperone (Figure 1).^{31,32} Loss-of-function mutations in the prosegment can result in lower PCSK9 circulating levels due to impaired autocatalytic processing and secretion.³³ A similar phenotype has been found when misfolded precursors in the ER are present; indeed, they act in a dominant negative manner by strongly decreasing secretion of PCSK9.³⁴

Common to other subtilisins, trademarks of the catalytic domain (aa 153–454), are the triad Asp₁₈₆, His₂₂₆ and Ser₃₈₆ as well as the amino acid sequence 367–380.^{31,32} The catalytic domain is responsible for the binding of PCSK9 to the epidermal growth factor-like repeat homology domain A (EGF-A) of human LDL receptor (LDLR). Finally, the PCSK9 C-terminal domain (aa 455–692) although not required for LDLR binding, plays a pivotal role in targeting

LDLRs for subsequent degradation.³⁵ In circulation, mature PCSK9 undergoes a furin enzymatic inactivation at position 218 of the catalytic domain (RFHR218:QA) resulting in the formation of a 55-kDa-truncated form.³⁶ This last, representing 15–40% of the total circulating PCSK9, is believed to be inactive. PCSK9 exists in a phosphorylated and non-phosphorylated form and after statin treatment the phosphorylated state is reduced by 25%. Considering that phosphorylation at site S688 could be stimulatory, statin therapy may reduce PCSK9 activity by decreasing the phosphorylated state.³⁷

Another unclear aspect of PCSK9 biology remains its ability to circulate bound to lipoproteins (20–40%: one PCSK9 molecule for every 500–1000 LDL particles). Upon lipoprotein-apheresis half of plasma PCSK9 is removed.³⁸ This has led to the hypothesis that the cellular regulation of cholesterol concentrations is under the tight control of a stochastic extracellular system in which LDLR fate is regulated by the interaction with PCSK9-carrying LDL; this could explain, at least in part, why LDLR recycles hundreds

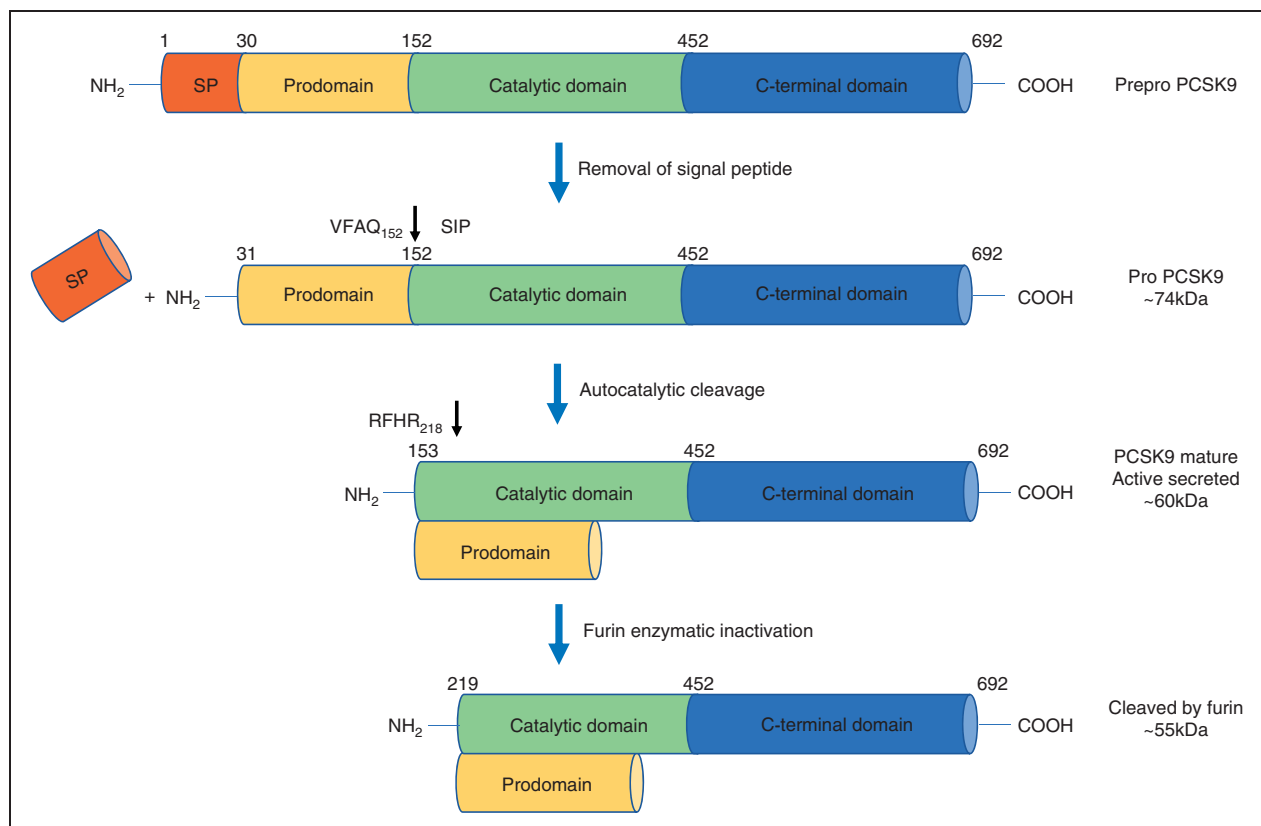


Figure 1. [AQ50] The mature form of pro-protein convertase subtilisin/kexin type 9 (PCSK9) consists of a signal peptide (SP) (aa: 1–30), a prodomain (aa: 31–152), a catalytic domain (aa: 153–452) and a C-terminal domain (aa: 453–692). Once in the endoplasmic reticulum, the SP is cleaved and autocatalysis of the PCSK9 zymogen occurs between Gln₁₅₂ and Ser₁₅₃ (VFAQ₁₅₂: SIP). Mature PCSK9 (aa: 153–692, about 60 kDa) undergoes a furin enzymatic inactivation at RFHR₂₁₈:QA leading to a 55-kDa truncated form. Adapted from Abifadel et al.¹⁷⁵ SIP: site 1 protease.

of times.³⁹ PCSK9 was also found in association with lipoprotein(a) (Lp(a)) particles in humans with high Lp(a) levels and in mice carrying human Lp(a), whereas *in vitro* data do not support this conclusion.⁴⁰ Interestingly, after treatment with an anti-Lp(a) antibody, total PCSK9 levels were not affected whereas there was a steady decline in those bound to Lp(a), i.e. PCSK9-Lp(a).⁴¹ The binding with high-density lipoprotein (HDL) remains controversial.^{42,43}

Transcriptional regulation of PCSK9

Discovered in 2003,⁴⁴ PCSK9 is one of the 33 genes regulated by the sterol regulatory element binding protein (SREBP) family of transcription factors.⁴⁵ In the same year, Abifadel et al. discovered the association between mutations in the neural apoptosis-regulated convertase-1 (NARCI or PCSK9) human orthologue and the autosomal dominant form of hypercholesterolaemia.⁹

The regulation of PCSK9 by SREBPs is not a matter of surprise, since these transcription factors are finely regulated by intracellular sterols and lipid levels. The SREBP family consists of three different proteins, SREBP1a, SREBP1c and SREBP2. SREBP1a is a potent activator of all SREBP-responsive genes, including those mediating cholesterol, fatty acid and triglyceride (TG) biosynthesis, whereas the roles of SREBP1c and SREBP2 are more selective.⁴⁶ It is generally assumed that genes involved in fatty acid metabolism, e.g. fatty-acid-synthase and stearoyl-CoA desaturase-1, are preferentially regulated by SREBP1c, whereas the expression of enzymes involved in cholesterol metabolism (3-hydroxy-3-methyl glutaryl coenzyme A (HMG-CoA) synthase), LDL uptake and intestinal absorption (Niemann-Pick C1-like 1 (NPC1L1)) are equally modulated by the activity of both SREBP1c and SREBP2.⁴⁷

The SREBP cleavage-activating protein (SCAP) contains a sterol-sensing domain (SSD) and, in response to a reduced availability of cholesterol, permits the dissociation of SREBP from the ER-resident membrane protein, INSIG. **[AQ6]** The conformational change of SCAP exposes a portion of the protein, that signals inclusion as a cargo in the COPII vesicles moving from the ER to the Golgi apparatus. **[AQ7]** In these vesicles, SREBP – linked to SCAP – is transported to the Golgi.⁴⁸ Within the Golgi, SREBP undergoes two sequential cleavage processes, mediated by the S1P₁ and the site-2 protease (S2P), that release the soluble amino-terminal portion of SREBP, then translocated into the nucleus driving the transcription of target genes.⁴⁹

The SREBP-dependent regulation of PCSK9 was explained by the identification of the sterol-regulatory element (SRE) in its promoter region. Thus, in response to cell cholesterol depletion or to inhibition of

intracellular synthesis, i.e. statins, PCSK9 promoter activity is induced determining increased transcription and higher circulating levels⁵⁰ (Figure 2). Interestingly, the expression of both NPC1L1 and PCSK9 is regulated by a second transcription factor, the hepatocyte nuclear factor 1 (HNF1) α .^{51–53} The highly conserved HNF1-binding site has been identified in close proximity to the SRE in the promoter region of PCSK9.⁵⁴

A further characterization of the transcriptional control of the PCSK9 gene expression brought the discovery of the histone nuclear factor P (HINFP), also binding PCSK9 promoter sequence. HINFP interacts with a highly conserved HINFP-recognition sequence located between HNF1-binding site and the SRE of PCSK9 promoter.⁵⁵ HINFP can exert either a positive or negative impact on target genes through an interaction with the MBD2 protein.⁵⁵ **[AQ8]** In the case of PCSK9, HINFP behaves as a transactivator, by mediating the histone H4 acetylation on the PCSK9 promoter region.⁵⁵ Along this line, deficiency of the histone deacetylase sirtuin 6 in the liver leads to elevated PCSK9 expression.⁵⁶ Sirtuin 6 is recruited to the PCSK9 promoter by the transcription factor forkhead box protein O3 (FoxO3), leading to deacetylation of histone H3 and repression of PCSK9 transcription by suppression of HNF1 activity.⁵⁶ With a similar mechanism, the flavonoid pinostrobin can inhibit PCSK9 expression by up-regulating FoxO3a in hepatic cells.⁵⁶ Potent inhibitory effects are also exerted by the epigenetic drug 5-azacytidine, reducing expression of PCSK9 and of additional key genes, i.e. HMG-CoA reductase and fatty acid synthase.⁵⁷ Azacytidine activation of the related deacetylase Sirt1 further leads to a marked reduction in PCSK9 secretion by an unknown posttranslational mechanism.⁵⁷ These findings confirm a major role of the epigenetic regulation of PCSK9 expression through DNA methylation and histone acetylation.

By using bioinformatics and *in vitro* functional analysis, microRNAs (miRNA, miR)-191, miR-222 and miR-224 were identified as natural regulators of PCSK9.⁵⁸ These small endogenous non-coding RNAs regulate a wide range of molecular pathways by target genes, such as PCSK9, controlling expression at the post-transcriptional level. In particular, miR-122 and miR-33 appear to play a major role in fatty acid and cholesterol metabolism.⁵⁸

Pharmacological modulation of PCSK9 levels by drugs affecting lipid metabolism

Statins

Statins are the most commonly prescribed class of lipid-lowering drugs. They act as competitive inhibitors of HMG-CoA reductase, the rate-limiting enzyme in the

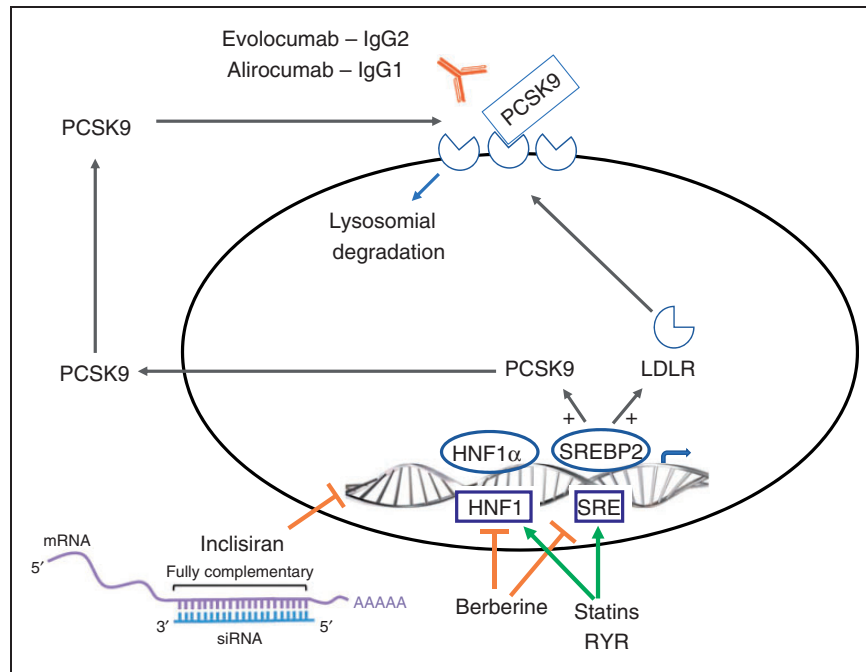


Figure 2. Statin increases the activity/nuclear translocation of sterol regulatory element-binding protein-2 (SREBP2), a transcription factor activating the low-density lipoprotein receptor (LDLR) and pro-protein convertase subtilisin/kexin type 9 (PCSK9) genes. Statin treatment also stimulates gene and protein expression of hepatocyte nuclear factor I (HNF1) α , a further transcriptional activator of PCSK9. This results in increased expression and secretion of PCSK9 protein, which binds the LDLR and targets it for lysosomal degradation. Berberine reduces PCSK9 mRNA transcription activity due to a negative interaction with the SREBP2 and HNF1 transcription factors on the PCSK9 promoter. Aloricomab and evolocumab are fully humanised monoclonal antibodies against PCSK9. Inclisiran, a synthetic silencing RNA, targets the 3' UTR of the PCSK9 mRNA. Adapted from Konrad et al.¹⁷⁶ IgG: immunoglobulin G; RYR: red yeast rice; SRE: sterol regulatory element.

cellular cholesterol biosynthetic pathway. The end process is increased expression of LDLRs and uptake of LDL-C from the circulation. Statins have provided outstanding contributions to the primary and secondary prevention of CHD.^{59,60} Analysis of large populations in primary and secondary prevention have generally indicated that a 38.6 mg/dl LDL-C reduction is associated with an approximate 22% lowering of CV risk.²

A paradoxical effect of statins on PCSK9 expression has been detected. By inhibiting cholesterol biosynthesis, statins favour the nuclear translocation of SREBP2 and, through a direct interaction with the SRE-1 element, raise both *LDLR* and *PCSK9* cellular gene expression,^{50,61} resulting in an elevation of circulating PCSK9 levels.

Beyond the SREBP2-induced activation of PCSK9, statin treatment also stimulates gene and protein expression of HNF1 α , a further transcriptional activator of PCSK9. In this respect, Dong et al., in dyslipidaemic hamsters treated with rosuvastatin, reported an unexpected increment of LDL-C levels, caused by a higher induction of PCSK9 compared with that of LDLRs. In an attempt to clarify the mechanism, the authors found that statins increased both

SREBP2 and HNF1 α , resulting in a stronger activation of PCSK9 compared with that of LDLRs.⁶¹ More recently Dong et al. demonstrated that, besides HNF1 α , HNF1 β is also a positive regulator of liver PCSK9. A transient liver-specific knockdown of either HNF1 α or HNF1 β blunted the rosuvastatin-driven elevation of serum PCSK9 with a concomitant reduction of cholesterolaemia.⁶²

Overall, a consequence of raised PCSK9 levels upon statin treatment may be the increment of LDLR lysosomal degradation, with a reduced efficacy of statins. This mechanism could explain why increasing doses of statins fail to achieve proportional LDL-C lowering, leading to the 'rule of 6%'. According to this, each doubling of statin dose improves LDL-C reduction by only approximately 6%.⁶³

Atorvastatin. Definition of the effects of atorvastatin on PCSK9 levels came from a meta-analysis on the effects of statin therapy. A significant PCSK9 rise was observed in all subgroups (80 mg, 40 mg or 20 mg/day) with atorvastatin.⁶⁴ The effect appears to be dose-dependent, as described by Kera et al.⁶⁴ reporting that atorvastatin 10 mg/day increases PCSK9 levels by 19%,

with a stronger effect (+27%) of atorvastatin 80 mg. In other cohorts, atorvastatin 10 mg/day led to non significant PCSK9 increases, whereas atorvastatin 20 mg raised circulating PCSK9 levels in a range between 30% and 35%,⁶⁵ atorvastatin 40 mg by 34–38%^{66,67} and atorvastatin 80 mg by 46%.⁶⁸ [AQ9] In addition to dose-dependency, the impact on PCSK9 is relatively rapid; 24-hour treatment with atorvastatin 10 and 80 mg raises PCSK9 by 13% and 27%, respectively.⁶⁵

Rosuvastatin. In a clinical trial with 60 patients with stable angina, 14-day therapy with rosuvastatin (10 mg) led to a 19% rise in circulating PCSK9 levels (together with a 60% reduction of LDL-C).⁶⁹ In a sub-analysis of the Justification for the Use of Statins in Prevention: An Intervention Trial Evaluating Rosuvastatin (JUPITER) study, one-year treatment with rosuvastatin (20 mg) increased PCSK9 by 35% in women and 28% in men vs a similar fall of LDL-C in the two genders (–55% for women and –53% for men). The largest LDL-C reduction was associated with the largest rise in plasma PCSK9 concentrations, an effect consistent across genders.⁶⁹ In a series of 27 hypercholesterolaemic subjects, administration of rosuvastatin for three months resulted in a positive correlation between PCSK9 plasma levels and LDL-C reduction, with a 27% rise of PCSK9 with a dose increase from five to 40 mg.⁷⁰ In homozygous familial hypercholesterolaemic (HoFH) patients, LDL-C levels were reduced by 16.5% and in heterozygous familial hypercholesterolaemic (HeFH) patients levels were reduced by 48.5%. Conversely, PCSK9 levels increased by 17% in HoFH patients and by 27% in HeFH patients.⁷⁰ Overall, PCSK9 levels are elevated in untreated familial hypercholesterolaemia (FH) patients, particularly HoFH and high-dose statin therapy further increases PCSK9 levels. Thus, the positive correlation between PCSK9 and LDL-C, found in untreated HoFH and HeFH patients, was lost.⁷¹ Finally, the statin-driven stimulatory effect on PCSK9 levels remained consistent when rosuvastatin was administered in combination with ezetimibe.⁶⁹

Simvastatin. In a single centre, randomised clinical trial, subjects with fasting LDL-C < 190 mg/dl, treated for 14 days with simvastatin (40 mg/day) showed a reduction of LDL-C levels by 41%, and a marked rise of PCSK9 concentrations (+67.8%). The association with ezetimibe lowered LDL-C by 60%, with a comparable effect on PCSK9 to simvastatin monotherapy, i.e. +67.3%.⁷²

A similar trend was found in a cross-over study, enrolling men with mixed dyslipidaemia, given simvastatin (40 mg/day) for two weeks. Along with a LDL-C lowering effect of –24%, simvastatin raised PCSK9

levels significantly in both fed (+29%) and fasted (+16%) states.⁷³

A different conclusion was instead reached in a double-blind, placebo-controlled, crossover trial involving 215 African- and European-American men. Six-week simvastatin (10 mg/day) treatment did not affect PCSK9 concentrations, despite a 25% drop in LDL-C levels. No significant correlations were found between simvastatin-mediated LDL-C lowering and baseline or on-treatment PCSK9 levels.⁷⁴

Finally, looking at the pleiotropic effects of statins beyond cholesterol-lowering, an interesting effect on PCSK9 expression of the isoprenoid cholesterol precursor geranylgeraniol was shown. Studies in Caco-2 cells indicated that induction of PCSK9 mRNA and protein driven by simvastatin was significantly prevented by the addition of geranylgeraniol. The mechanism involved the suppression of Rac1-GTP levels driven by simvastatin (–35.7%).⁷⁵ [AQ10]

Pravastatin and pitavastatin. The effect of pravastatin (20 mg/day) and pitavastatin (4 mg/day) treatment on PCSK9 has been evaluated in the Treatment With Statin on Atheroma Regression Evaluated by Intravascular Ultrasound With Virtual Histology (TRUTH) study, i.e. an eight-month prospective, open-labeled, randomised, multicentre trial. Among 164 patients with coronary artery disease (CAD), not on lipid-lowering therapy, assignment to pravastatin or pitavastatin significantly reduced LDL-C by 28% and 41%, respectively, with concomitant rises of total PCSK9, by 39% and 78%, respectively.⁷⁶

Ezetimibe

Among other commonly prescribed LDL-C lowering medications, ezetimibe is a cholesterol-absorption inhibitor acting on the absorption of biliary and dietary cholesterol across the brush border membrane of the intestinal enterocyte, by blocking the transport protein NPC1L1. By reducing cholesterol absorption ezetimibe increases liver LDLR expression.⁷⁷

Studies of the effects on PCSK9 levels have provided contrasting findings. The previously quoted trial in 215 African- and European-American with LDL-C between 130–175 mg/dl, showed that six-week treatment with ezetimibe (10 mg/q.d.), simvastatin (10 mg/q.d.) and both in combination did not significantly raise plasma PCSK9 levels.⁷⁴ Similar conclusions were reached in healthy⁷² and type 2 diabetes (T2D) subjects,⁷⁸ for whom ezetimibe alone or in addition to a statin did not affect PCSK9 levels, despite an incremental reduction of LDL-C.

Contrasting findings were provided by Davignon and Dubuc who showed that hypercholesterolaemic

patients on ezetimibe in combination with a statin had higher PCSK9 levels compared with controls and statin-treated subjects.⁷⁹ These data were confirmed in patients on statins or on combined statin-ezetimibe treatment who presented with roughly 45% and 77% higher PCSK9 levels, respectively, vs controls. The combination regimen was associated with 22% higher plasma PCSK9 levels⁷⁰ compared with statin alone.

A meta-analysis showed a non-significant impact of combining statin with ezetimibe vs statin monotherapy in terms of altering PCSK9.⁶⁴ Removal from the analysis of the study by Lakoski et al.⁷⁴ led to a significant elevation of plasma PCSK9 after ezetimibe. The authors justified this discrepancy, speculating that the lowest intensity of statin therapy (simvastatin 10 mg/day) was used and that African-Americans have a reduced response to statins.⁶⁴ More importantly, African-Americans have a higher rate of non-response to ezetimibe, due to frequent NPC1L1 mutations.⁸⁰ Finally, if a molecular mechanism is searched, HepG2 and Caco-2 cells treated with ezetimibe 25 μ M or 50 μ M showed a ~1.5 to two-fold increase of PCSK9 mRNA with no changes in protein secretion.⁷⁹ [AQ11]

Overall, if it is indisputable that PCSK9 levels are raised by cholesterol synthesis and absorption inhibitors, treatment with PCSK9-antibodies did not affect the balance between cholesterol synthesis and absorption.⁸¹ [AQ12]

Fibrates

Fibrates are activators of the transcription factor peroxisome proliferator-activated receptor (PPAR) system, mainly PPAR- α . They have shown significant benefit in clinical trials of CV prevention, i.e. reducing occurrence of nonfatal MI in patients with concomitant TG elevation and HDL-C reduction. PPAR- α agonists, i.e. fibrates, are powerful TG-lowering agents. They mainly affect TG catabolism and consequently raise, particularly fibrates, HDL-C levels.⁸² Of minor significance, their effect on cholesterol metabolism is characterised by a modest reducing activity of HMG-CoA reductase and an increase of cholesterol 7 α -hydroxylase (CYP7A1), raising bile acid excretion, in some cases associated with an increased risk of gallstone formation.⁸³

Although studies in rodents have clearly showed a decrement of PCSK9 protein content upon fenofibrate treatment,⁸⁴ discordant findings have been reported in humans.^{85–87} The effects of bezafibrate and fenofibrate on PCSK9 plasma levels have been assessed in an eight-week open randomised crossover study enrolling 14 dyslipidaemic subjects with impaired glucose tolerance or T2D. Fibrates raised PCSK9 levels by 40% (bezafibrate) and 67% (fenofibrate).⁸⁸ In line with these

studies, two other studies reported that 12- or 24-week treatments with fibrates raised serum PCSK9 levels by 25%⁸⁶ and 17%,⁸⁵ respectively. A different conclusion was reached instead in a post-hoc analysis of the Fenofibrate Intervention and Event Lowering in Diabetes (FIELD) study showing that, in diabetics, six-week treatment with fenofibrate (200 mg/day) modestly, but significantly, reduced plasma PCSK9 concentrations by 8.5%, paralleling a 13% reduction in LDL-C.⁸⁷

Again, also in case of concomitant administration of a statin with fenofibrate, inconclusive evidence has been reported. Six-week treatment with atorvastatin (10 mg) or fenofibrate (160 mg) raised PCSK9 by 14% and 26%, respectively; combination therapy enhanced PCSK9 by 19%, thus showing no additive effect.⁸⁹ Conversely, in statin-treated T2D patients, fenofibrate (145 mg) as monotherapy led to a 13% drop in PCSK9 levels.⁹⁰

Fibrates and statins share a common pathway regulating PCSK9, i.e. SREBP2 activation. Indeed, like statins, fibrates decrease LDL-C by modest activity on HMG-CoA reductase,⁹¹ thus upregulating SREBP2. This mechanism may be valid when fibrates are administered as monotherapy, whereas they are less likely to further influence cholesterol synthesis and SREBP2 expression when given with background statins. In this condition, PPAR- α activation may counteract PCSK9 induction by statins, repressing PCSK9 promoter activity and furthermore raising furin expression, leading to protein degradation.⁹²

Omega-3 fatty acids (omega-3s)

Fatty acids of the n-3 series, i.e. those with multiple double bonds – the first one being in the n-3 position from the terminal methyl group – have provided an important addition to dietary treatment in syndromes characterised by elevated TGs. Omega-3s act as ‘fraudulent fatty acids’,⁹³ i.e. somewhat similar to drugs with a fatty acid-like structure, particularly fibrates. They do not enter the liver metabolic handling by the classical fatty acetyl coenzyme A (acetylCoA) oxidative mechanism with carnitine-mediated transport to mitochondria. Omega-3s rather stimulate the metabolism of fatty acids coming from diet or end products of TG metabolism by the peroxisomal PPAR- α mediated pathway (see review by Botta et al.).⁹⁴

The potential activity of omega-3s on PCSK9 has been evaluated in numerous experimental studies. The transcription factor SREBP1c⁹⁵ has been shown to be inhibited by docosahexaenoic acid (DHA), thus reducing expression of PCSK9. In rats, 12-week supplementation with eicosapentaenoic acid (EPA)+DHA reduced liver cholesterol synthesis by downregulating

HMG-CoA reductase and SREBP2.⁹⁶ The SREBP involvement was confirmed in a study testing the efficacy of a novel conjugate of EPA with niacin, i.e. CAT-2003, on the prevention of nonalcoholic steatohepatitis. This formulation, designed to be hydrolysed by a fatty acid amide hydrolase-releasing EPA, inhibited processing of SREBP1 and SREBP2 to their mature nuclear forms, thus, reducing expression of many SREBP target genes, e.g. *PCSK9*, *HMGCR*, and *LDLR*. CAT-2003, when administered to APOE*3-Leiden mice fed a cholesterol-containing Western diet, lowered PCSK9 along with total cholesterol and TGs.⁹⁷ [AQ13]

Clinical studies have examined different aspects of omega-3 activity. In a double-blinded, parallel-group, placebo-controlled intervention study, involving 90 pre- or post-menopausal women, 12-day supplementation with 2.2 g omega-3 polyunsaturated fatty acids (PUFAs) (38.5% EPA, 25.9% DHA and 6.0% DPA) vs control thistle oil led to a PCSK9 drop of 16.1% in premenopausal and 13.1% in postmenopausal women. Neither group showed LDL-C changes.⁹⁸ [AQ14]

Cholesterol ester transfer protein (CETP) inhibitors

CETP is a hydrophobic glycoprotein mainly secreted by the liver. CETP promotes a net mass transfer of cholesteryl esters (CEs) from the HDL fraction to the potentially proatherogenic non-HDL fractions in exchange for TGs. The discovery of the CETP system has led to the development of agents interfering with CETP, thus elevating HDL-C, decreasing LDL-C and potentially preventing CV disease. However, this pharmacological approach has provided convincing evidence of HDL-C raising activity, but disappointing results in trials of CV prevention.⁹⁹

In mildly hypercholesterolaemic patients, anacetrapib as monotherapy or in combination with a statin reduced LDL by increasing fractional catabolic rate (FCR), most likely reflecting a rise in the number of LDLRs, rather than an involvement of PCSK9. A significant 19% reduction of PCSK9 levels was found only in subjects treated with anacetrapib monotherapy, whereas – given in combination with atorvastatin – anacetrapib had no effect on either the production rate or the FCR of PCSK9.¹⁰⁰ This was confirmed in another study in similar patients.¹⁰¹

In an effort to shed light on the mechanism of the side effects observed in the Investigation of Lipid Level Management to Understand its Impact in Atherosclerotic Events (ILLUMINATE) trial, PCSK9 levels were evaluated in a subset of the study.¹⁰² Atorvastatin treatment was positively associated with PCSK9 levels in a dose-dependent fashion at the end of the run-in period, during which patients were titrated to doses of 10, 20, 40 or 80 mg/day, in order to achieve an

LDL-cholesterolaemia ≤ 100 mg/dl. PCSK9 concentrations were 37 ng/ml higher in patients receiving 80 mg atorvastatin vs those on the lowest 10 mg dose. Conversely, after three months of treatment with either atorvastatin or torcetrapib+atorvastatin PCSK9 levels rose modestly in the torcetrapib group (+3.7%) vs no change with placebo (+0.7%),¹⁰² an effect evident only in patients with T2D.

Overall, in humans, it appears that the impact of CETP inhibition on PCSK9 modulation is mainly explained by the background lipid-lowering therapy.

Nicotinic acid

Nicotinic acid (niacin or vitamin B3) is a natural compound exerting a variety of activities on lipids, both cholesterol and TGs, at pharmacological doses, i.e. about 1–2 g/day.

Limited information is available on the activity of niacin on PCSK9 levels. In a study of drug combinations in 70 patients with carotid atherosclerosis, a comparison between simvastatin 20 mg/day alone and simvastatin 20 mg+extended-release (ER) niacin 2 g/day was carried out. PCSK9 levels were raised by simvastatin alone (+13%) whereas they fell with the combination (–13%). This trend was also found in 19 dyslipidaemic patients randomised to atorvastatin 10 mg with the serial addition of fenofibrate 135 mg followed by ER niacin titrated to 2 g/day. Fenofibrate raised PCSK9 by 23% whereas the addition of niacin reduced it by 17%.¹⁰³ These findings may provide a demonstration of a possible mechanism whereby niacin lowers LDL-C levels.

Nutraceuticals

The role of dietary interventions to lower LDL-C levels is well established with a major impact of low fat and omega-6 and omega-3 fatty acid diets. Recent reports have, however, been focused on the potential of functional foods or food additives with a significant activity on LDL-C levels, promoted for patients with LDL-C elevation.¹⁰⁴ Consumers have free access to these products and some of them have definitely provided an indication of significant LDL-C lowering activity, in some cases with clear changes in PCSK9 levels.¹⁰⁵

Berberine

Berberine, a natural isoquinoline alkaloid present in several plants such as *Berberis vulgaris*, *Coptis chinensis* and *Berberis aristata*, has a traditional use as an antimicrobial, immunomodulatory and anti-diarrhoeal agent. Berberine administration can reduce total cholesterol (–23.5 mg/dl), LDL-C (–25.1 mg/dl) and TGs

(−44.2 mg/dl) as well as raise HDL-C (+1.93 mg/dl).¹⁰⁶ The main mechanisms of the lipid-lowering effects are dependent on increased liver LDLR expression and reduction of PCSK9. The raised LDLR expression does not appear to be dependent on SREBP1 and 2, but consequent to the activation of the extra-signal regulated kinase (ERK).¹⁰⁷ The berberine-associated PCSK9 lowering, from in vitro and in vivo studies, is instead linked to a reduction of hepatic PCSK9 mRNA and protein levels. A study of a combination of berberine and mevastatin showed an increase of LDLR mRNA levels with a suppressed upregulation of PCSK9 mRNA, induced by mevastatin alone.¹⁰⁸ The berberine-mediated drop of PCSK9 thus appears to be directly consequent to a reduction in mRNA transcription activity¹⁰⁸ due to a negative interaction with the SREBP2 and HNF1 transcription factors on the PCSK9 promoter. Berberine can also enhance HNF1 α protein ubiquitination and proteasomal degradation.¹⁰⁹ Alternatively, berberine could induce AMP-activated protein kinase (AMPK), in turn activating the PPAR α downregulation of PCSK9 mRNA levels.¹⁰⁸ Somewhat contrasting findings were reported in rats in which berberine raised plasma PCSK9 levels with a concomitant increase of SREBP2 and reduction of HNF1.¹⁰⁸

In clinical trials, berberine is often administered together with other nutraceuticals or with drugs, with contrasting effects on PCSK9 regulation. In patients with HeFH, treated with statins or a combination of statins and ezetimibe, supplementation with a nutraceutical tablet containing berberine, policosanol and red yeast rice (RYR), further reduced plasma LDL-C levels by 10.6%, possibly mediated by berberine through a modulation of PCSK9 expression.¹¹⁰ ~~In 23 dyslipidaemic subjects, a combination of berberine (531.2 mg), monacolin K (3.3 mg), and *Morus alba* extract (200 mg) given for four weeks did not change PCSK9 plasma levels, a neutral effect explained by a rise of PCSK9 levels by monacolin K, with negative modulation by berberine and *Morus alba*.~~¹¹¹

RYR

RYR is a food product made by fermenting the yeast *Monascus purpureus* on white rice. RYR contains several monacolins inhibiting HMG-CoA reductase. RYR is an effective natural cholesterol-lowering agent.¹¹² In statin-intolerant patients, it has been indicated as an alternative to statin therapy for patients with mild to moderate hypercholesterolaemia.¹¹³

Since monacolin K has the same chemical structure as lovastatin, it can be inferred that administration of RYR may lead to a rise in PCSK9 levels. Initially, this hypothesis was confirmed in two different studies

showing that three-day administration of a purified extract of RYR, Xuezhikang (1200 mg/kg/day), led to an increment in PCSK9 levels in the range of 56–70%.¹¹⁴ **AQ15**

The effect of RYR on the regulation of PCSK9 appears to be counteracted when given in combination with other nutraceuticals, e.g. berberine. In particular, in 23 dyslipidaemic subjects, a nutraceutical combination of RYR (monacolin K 3.3 mg), berberine 531.25 mg and leaf extract of *Morus alba* 200 mg did not modify PCSK9 levels. A possible explanation could be the opposite impact on PCSK9 modulation, i.e. up-regulated by RYR and down-regulated by berberine. Similar results were obtained with berberine 200 mg+monacolin K 3 mg, in individuals with non-HDL-cholesterol levels \geq 160 mg/dl.¹¹⁵ When given in combination with ezetimibe (10 mg/kg/day), Xuezhikang led to an increment of PCSK9 levels by 63%, an effect similar to that obtained by the administration of RYR as monotherapy.¹¹⁴

PCSK9 clinical evaluation – results from the Further Cardiovascular Outcomes Research with PCSK9 Inhibition in Subjects with Elevated Risk (FOURIER) and Evaluation of Cardiovascular Outcomes After an Acute Coronary Syndrome During Treatment With Alirocumab (ODYSSEY OUTCOMES) trials

PCSK9 is at present the most extensively studied drug target for treating dyslipidaemia and several strategies are being explored to reduce plasma levels or to inhibit either PCSK9 protein synthesis or binding to LDLR, by way of mAbs, vaccines, small protein inhibitors (peptides/adnectins), antisense oligonucleotides or siRNA. Among these to date, two fully human mAbs, alirocumab and evolocumab, have been approved in the USA and in the European Union, whereas bococizumab, a humanised mAbs, has been discontinued in November 2016^{116,117} and will not be given further space in this review article. Alirocumab and evolocumab contain two human heavy chains each covalently linked to a fully human kappa light chain through a disulfide bond. Molecular weights are approximately 146 and 144 kDa, respectively.^{118,119}

Alirocumab is indicated as an adjunct to diet and maximally tolerated statin therapy for the treatment of adults with HeFH or clinical atherosclerotic cardiovascular disease (ASCVD), who require additional lowering of LDL-C; evolocumab is also authorised in patients with HoFH requiring additional lowering of LDL-C, in order to reduce the risk of MI, stroke,

and coronary revascularization in adults with CVD. Overall, in the context of CVD prevention in FH patients, PCSK9 inhibitors are to be considered in patients with severe HeFH without ASCVD with LDL-C ≥ 200 mg/dl or LDL-C ≥ 175 mg/dl in the presence of comorbidities, i.e. diabetes mellitus or hypertension. HoFH, excluding those with null LDLR mutations, should receive maximal lipid-lowering therapy including lipoprotein apheresis plus a PCSK9 inhibitor, namely evolocumab.¹²⁰ A recent approach, based on the estimation of the number needed to treat (NNT) determined by patients' absolute risk and estimated relative reduction in risk, has identified the addition of ezetimibe to PCSK9 mAbs to maximally tolerated statin therapy to be cost effective in very high and high-risk patients.¹²¹

The practical clinical guidance of the European ESC/EAS Task Force advises use of PCSK9 inhibitors in (a) patients with ASCVD with elevated LDL-C levels, despite maximally tolerated statin with or without ezetimibe therapy; (b) patients at very high risk who do not tolerate appropriate doses of at least three statins; (c) in FH subjects without clinically diagnosed ASCVD, at high or very high CV risk despite maximally tolerated statin plus ezetimibe therapy.¹²²

AQ16

Absolute bioavailability is 72% for evolocumab and 85% for alirocumab, with distribution volumes of 3.31 and 3.0–3.81, respectively, confirming their limited tissue distribution. Effective plasma half-lives are 11–17 days for evolocumab and 17–20 days for alirocumab, reduced by statin co-administration. Indeed, evolocumab clearance is increased by about 20%, partially due to the statin-driven upregulation of PCSK9.¹¹⁸ Similarly, alirocumab exposure was reduced by about 40%, 15% and 35% when administered with statins, ezetimibe and fenofibrate, respectively. These drug-drug interactions are not to be rated as clinically meaningful and do not generally require dose adjustment.

Evolocumab

The efficacy of evolocumab as a lipid-lowering drug was tested in the clinical development programme, Program to Reduce LDL-C and Cardiovascular Outcomes Following Inhibition of PCSK9 In Different Populations (PROFICIO).

The FOURIER study evaluated the efficacy of evolocumab in patients with ASCVD on major adverse cardiovascular events (MACEs), i.e. the composite of CV death, MI, stroke, hospitalization for unstable angina, or coronary revascularization. Evolocumab reduced the primary end-point by 15%, an effect primarily ascribable to MI (–27%), stroke (–21%) and

coronary revascularization (–22%) reductions. Over the course of the trial, LDL-C had a median value of 30 mg/dl with 42% of patients reaching <25 mg/dl. Non-HDL was reduced by 52%, similar to apolipoprotein B (apoB) (–49%); Lp(a) levels were reduced by 27%.¹²³ A post-hoc analysis reported that when baseline Lp(a) levels were >37 nmol/l, evolocumab reduced MACEs by 2.5% (NNT=40). Conversely, if baseline Lp(a) levels were \leq the median, the absolute benefit was reduced to 0.95% (NNT=105).¹²⁴ Similar conclusions were reached in the presence of diabetes, with a 17% RR in diabetics vs 13% in non-diabetics.¹²⁵ **AQ17**

Overall, the FOURIER trial highlighted how severity and extent of CAD are leading features in order to identify people who benefit the most from LDL-C lowering. Recent MI, multiple prior MIs and residual multivessel coronary disease were independent predictors of CV outcomes, leading to an absolute risk reduction by treatment of over 3% in high risk vs approximately 1% in low-risk groups, respectively. In patients with at least one high-risk feature there was a relative risk reduction in CV death, MI or stroke of 19% during the first year and of 27% beyond the first year.¹²⁶ A further sub-analysis showed that in patients with peripheral artery disease (PAD) absolute RR for the primary CV endpoint was 3.5% (NNT=29) vs 1.6% (NNT=63) for patients without PAD. Major adverse limb events were further reduced by 42% with evolocumab. This benefit followed a linear correlation with LDL-C reduction.¹²⁷ Evolocumab did not raise either the risk of new-onset diabetes¹²⁵ or of cognition impairment.¹²⁸

Evolocumab was also the object of assessment of plaque regression, evaluated by IVUS in the GLOBAL Assessment of Plaque regression With a PCSK9 antibody as Measured by intraVascular Ultrasound (GLAGOV) study. **AQ18** Drug treatment was associated with a small, yet significant, reduction in percentage and total atheroma volume,¹²⁹ but with inconsistent changes in coronary fibrous, fibrofatty and necrotic volumes as well as in coronary calcium by virtual histology.¹³⁰

Alirocumab

The efficacy of alirocumab has been tested in the ODYSSEY programme including 16 clinical trials, designed to evaluate the antibody together with other lipid-lowering agents, e.g. statins, or as monotherapy across a wide range of patients, including those with high CV risk, intolerant to statins and HeFH.

CV outcomes have been studied in the ODYSSEY OUTCOMES trial enrolling a total of 18,924 patients, with an acute coronary syndrome 1–12 months earlier, from 57 countries with a 2.8 year follow-up. In the

on-treatment analysis, the mean LDL-C reduction at 4, 12 and 48 months was 38 mg/dl, 42 mg/dl and 53 mg/dl, respectively. If compared with placebo, decrements were 63% (four months), 61% (12 months) and 55% (48 months). The primary efficacy endpoint, i.e. a composite of CHD death, non-fatal MI, fatal or non-fatal ischaemic stroke, or unstable angina requiring hospitalization was reduced by 15%. The absolute risk reduction in these patients already on intensive or maximum-tolerated statin therapy was -1.6% (-9.5 vs -11.1%). This effect was mainly driven by non-fatal MI and ischaemic stroke lowering, i.e. -14% and -27% , respectively. To prevent the occurrence of one primary endpoint event, 49 patients would need to be treated for four years (NNT = 49). Without extrapolation the 2.8-year NNT was 63.¹³¹ Deaths from coronary causes were non-significantly reduced, whereas there was a 15% reduction of all-cause mortality.¹³² Overall, when total (first and subsequent) nonfatal CV events and all-cause deaths are jointly estimated, alirocumab reduced twice the number of first events prevented.¹³³ [AQ19]

When the analysis was stratified by baseline LDL-C, MACE reduction went down to 24% in patients with baseline LDL-C ≥ 100 mg/dl with NNT = 16 for four years; a non-significant effect was found in those with LDL-C < 100 mg/dl.¹³² The percentage of LDL-C reduction was highest in the group with LDL-C < 80 mg/dl (i.e. -57%) vs -55% for the 80–100 mg/dl baseline group and -53% for the patients with baseline LDL-C > 100 mg/dl.¹³² Paradoxically, in the FOURIER study, patients who benefited the most were those with LDL-C < 80 mg/dl and between 80–92 mg/dl. The significant effect was lost for LDL-C > 92 mg/dl.¹²³ No changes were noted in high-sensitivity C-reactive protein (hsCRP) levels. Local injection-site reactions were 3.8% in the alirocumab group vs 2.1% in the placebo group.¹³²

Comparing the ODYSSEY OUTCOMES with the FOURIER study, it can be noted that (a) the ODYSSEY OUTCOMES trial recruited ACS patients vs those with CAD in the FOURIER study, (b) 89.5% of ODYSSEY OUTCOMES patients were on high-intensity statin (atorvastatin 40–80 mg/q.d. or rosuvastatin 20–40 mg/q.d.) vs 69.2% in the FOURIER study, and (c) the ODYSSEY OUTCOMES endpoints were specific for CAD and ischaemic stroke vs CVD and stroke in the FOURIER study.¹³⁴ [AQ20] A study with a similar objective as GLAGOV with evolocumab is being carried out in Japan, i.e. the ALirocumab for Thin-cap fibroatheroma in patients with coronary Artery disease estimated by optical coherence tomography (ALTAIR) study using OCT imaging combined with virtual histology, in order to characterise the drug efficacy on rupture-prone plaques.¹³⁵

[AQ21]

Overall, recent data from pooled analysis of the ODYSSEY programme indicate that alirocumab is effective with a similar safety profile in high-risk patients with or without prior coronary revascularization,¹³⁶ in individuals with T2D¹³⁷ and high CV risk with or without mixed dyslipidaemia.¹³⁸

Inclisiran

Among therapeutic strategies aimed at reducing circulating PCSK9 levels, the use of siRNA represents a different strategy for reducing PCSK9 secretion. siRNAs target RNA directly destroying it before the protein is synthesised. Inclisiran, targeting the 3' UTR of the PCSK9 mRNA, is a long-acting, subcutaneously delivered, synthetic siRNA, conjugated to triantennary N-acetylgalactosamine carbohydrates. It binds to asialoglycoprotein receptors on hepatocytes, leading to the uptake of inclisiran and suppression of hepatic PCSK9 production.¹³⁹ The drug is rapidly distributed in plasma with peak concentrations occurring at the end of the infusion, with a roughly dose-proportional increment.¹⁴⁰

The first study evaluating intracellular PCSK9 inhibition vs LDL-C reduction was a Phase 1 dose-escalation randomised controlled trial (RCT) enrolling 32 healthy volunteers with LDL-C > 116 mg/dl. PCSK9 and LDL-C were reduced in a dose-dependent manner with maximal 70% and 40% falls when the highest dose of inclisiran (0.40 mg/kg) was administered.¹⁴⁰ Effects were maintained for at least six months if inclisiran was administered at the dose of 300 mg or more, either in single- or multiple-dose regimens. No serious adverse events were observed.¹⁴¹

These data were corroborated in the ORION-1 study, a Phase 2 dose-ranging trial evaluating the efficacy of inclisiran vs placebo in lowering LDL-C at six months. [AQ22] Six different doses (three single doses and three two-dose starting regimens administered 90 days apart) were tested. In high CV-risk patients, already at maximally tolerated statin therapy, inclisiran, either as single- or two-dose regimens, dramatically reduced LDL-C, by 42% and 52.6%, respectively. These percentage reductions were in a range similar to those achieved with mAbs with an overall optimal dosage of 300 mg given twice as the starting regimen.¹⁴² Whether or not there was the presence of diabetes at baseline, inclisiran was equally effective in the management of dyslipidaemias; LDL-lowering was between 22.8% and 25.2% in patients without diabetes, and between 22.8% and 25.5% in diabetics.¹⁴³ Although Lp(a) was reduced, this did not reach statistical significance.¹⁴⁴

Phase 3 trials evaluating the efficacy of inclisiran on HeFH (NCT03397121) or ASCVD (NCT03705234)

patients are ongoing. A CV outcomes trial involving approximately 15,000 subjects with high ASCVD risk is being planned.

Summary and conclusions

The pharmacological inhibition of PCSK9 has led to indisputable benefits in terms of LDL-C and CV-risk lowering, but the validity of clinical measurements of plasma PCSK9 for CV-risk prediction is not fully supported by the analysis of RCTs with PCSK9 inhibitors and thus remains an open question. While a number of studies did not report an association between PCSK9 levels and events,^{145–148} different conclusions were reached by others. Specifically, looking at meta-analyses, an association between CV events and PCSK9 levels was found,¹⁴⁹ a conclusion supported by a genome-wide association study of circulating PCSK9 levels showing that genetic reduction of PCSK9 levels by 50% is associated with a similar percentage reduction of CAD risk.¹⁵⁰ Such discrepancies may be based on the following observations: (a) plasma PCSK9 levels

differ across study design and clinical outcomes, (b) assay methods for PCSK9 do not discriminate between active and inactive truncated forms and (c) PCSK9 is tightly regulated at the transcriptional level.³⁹ Besides a diurnal rhythm, nutritional status, environmental pollutants (PM₁₀ particles)¹⁵¹ and gender differences regulate plasma PCSK9 levels, significantly higher in women than in men, with lower levels in postmenopausal vs premenopausal women.¹⁵² Most recently, evaluation of two cohort studies in diabetics provided discordant results, one study (Non-Insulin Dependent Diabetes, Hypertension, Microalbuminuria or Proteinuria, Cardiovascular Events, and Ramipril (DIABHYCAR)) showing a clear association between PCSK9 levels and incidence of MI, all CV events and stroke, the other (Survie, Diabète de type 2 et Génétique (SURDIAGENE)) reporting no such association, confirming the still inconsistent link between PCSK9 level and CV risk.¹⁵³ Finally, in the context of plasma PCSK9 as a possible independent factor beyond the traditional CV-risk biomarkers, in the obese subject PCSK9 appeared to be associated with

Table 1. Impact of hypolipidaemic agents on pro-protein convertase subtilisin/kexin type 9 (PCSK9) levels.

Drug class		Effect on PCSK9
Statins	Atorvastatin	10 mg, + 19%
		20 mg, + 30–35%
		40 mg, + 34–38%
		80 mg, + 46%
	Rosuvastatin	Women, + 35%
		Men, + 28%
	HeFH, + 38%	
	HoFH, + 21%	
	Simvastatin	+16–68%
	Pravastatin	+39%
	Pitavastatin	+78%
Ezetimibe	Ezetimibe	Neutral effect
Fibrates	Fenofibrate	+67%
	Bezafibrate	+40%
	Subgroup of FIELD study	–8.5%
Fatty acids	Omega 3	–16.1%
CETP inhibitors	Anacetrapib	–19%
	Subgroup of ILLUMINATE (<i>n</i> = 1745): after three months of treatment:	
	Atorvastatin+torcetrapib	+3.7% vs run-in with atorvastatin (significant)
	Atorvastatin+placebo	+0.7% vs run-in with atorvastatin (not significant)
Nicotinic acid		–13%
Nutraceuticals	Berberine	↔
	Red yeast rice	↔

CETP: cholesterol ester transfer protein; FIELD: Fenofibrate Intervention and Event Lowering in Diabetes; HeFH: heterozygous familial hypercholesterolaemic; HoFH: homozygous familial hypercholesterolaemic; ILLUMINATE: Investigation of Lipid Level Management to Understand its Impact in Atherosclerotic Events.

a significant rise of the Framingham Risk Score, a well-established predictor of CV risk.¹⁵¹

The pharmacological regulation of PCSK9 provides a pointer to estimate possible alterations in the overall response to lipid-lowering medications in patients at high CV risk. Prominent in this area has been the discovery that statins, via SREBP2 involvement, up-regulate PCSK9 gene expression (Table 1). Although clinical trials confirm that statin therapy raises PCSK9, irrespective of statin type, the magnitude of this effect remains unclear. Notably, the discovery that PCSK9 exists in an active S688-phosphorylated form, reduced by 25% after statin treatment, has led to the speculation that HMG-CoA reductase inhibition may reduce PCSK9 activity by decreasing the active phosphorylated state.³⁷

The pharmacological inhibition of PCSK9 in the clinic, with fully human mAbs against PCSK9, i.e. evolocumab and alirocumab, has provided convincing evidence of a dramatic LDL-C reducing activity, as well as of significant lowering of CV events.¹⁵⁴ Interestingly, also in this context, measuring PCSK9 circulating levels has become important. Upon administration of mAbs, total plasma PCSK9 concentrations increase up to 10-fold. This may be consequent either to reduced clearance of the immune complex vs free PCSK9, or to entrapment in the circulation of newly secreted PCSK9 with the antibody or, finally, to the up-regulation of PCSK9 secretion by the liver as a counter-regulatory mechanism.¹⁰ These findings have led some authors to hypothesise the use of PCSK9 total concentrations as a diagnostic tool assessing apparent cases of resistance to PCSK9 inhibitors;¹⁵⁵ indeed, if PCSK9 elevations are not detected after mAbs administration, this would suggest an altered tissue distribution of the drug. However, when free PCSK9 levels are considered, these are suppressed by 90–100% within one week of administration followed by a 65% fall of LDL-C.¹⁵⁶

Before the introduction of PCSK9 inhibitors, there was little to be done to achieve target LDL-C values, except for adding ezetimibe to statin treatment. This left out most of the very high-risk patients, e.g. those with FH, who could not reach LDL-C target values and could not be adequately treated.¹⁵⁷ The newly published American College of Cardiology/American Heart Association (AHA/ACC) 2018 cholesterol guidelines recommend – for ASCVD patients who are at very high risk and LDL-C not adequately controlled by statin therapy – a stepped approach of ezetimibe, in addition to the statin. If ~~that~~ combination does not work adequately, i.e. LDL-C levels ≥ 70 mg/dl, PCSK9 inhibitors could be added.¹⁵⁸ It should be noted that a very recent analysis of applicability and cost implications for PCSK9 inhibitors, based on the results of the ODYSSEY OUTCOMES study and on

these new guidelines, concludes that only a selective use of PCSK9 antagonists is justified.¹⁵⁹

~~In the context of~~ the interplay between hypercholesterolaemia and the inflammatory burden,¹⁶⁰ the clinical CV preventive activity of PCSK9 antagonists may possibly be affected by the poor efficacy of these agents on circulating inflammatory markers, e.g. hsCRP.^{161,162} In carriers of the PCSK9 R46L loss-of-function mutation, no changes in GlycA,¹⁶³ a composite NMR biomarker of systemic inflammation correlated with CRP,¹⁶⁴ was noted. **AQ23**

This lack of anti-inflammatory efficacy is also detected in FH patients when they are switched from lipoprotein apheresis to PCSK9 mAbs.¹⁶⁵ In the large FOURIER trial, patients with elevated hsCRP levels (>3 mg/dl) showed the best preventive response to treatment: those with a lower baseline hsCRP had an absolute risk reduction of 1.8%, vs a reduction of 2.6% for those with the highest baseline levels.¹⁶⁶ Open questions still remain on the association of PCSK9 not only with inflammation but also with infections as well; indeed, PCSK9 may play a critical role in the innate immune defense against bacterial and/or viral infections.¹⁶⁷

The very recent introduction of a siRNA PCSK9 antagonist, inclisiran, has given the possibility of widely spaced treatments, allowing for optimal compliance. This type of approach, still available to a very limited extent, may provide a very active cholesterol-lowering treatment with minimal effort by the patients.¹⁶⁸

Relative to safety concerns on the use of PCSK9 inhibitors, uncertainties persist ~~relative to~~ the potential risk for new-onset diabetes mellitus. While the use of PCSK9 inhibitors led to significant CV-risk reduction, generally similar between individuals with and without diabetes mellitus, with no further worsening of diabetes mellitus,¹⁶⁹ data from Mendelian randomization analyses reported that genetic inactivation of *PCSK9* is associated with an increased risk of new-onset diabetes mellitus.^{170–172} We recently showed that mice lacking *PCSK9* have an impaired insulin secretion vs wild-type animals, an effect lost in double KO mice for *PCSK9* and *LDLR*.¹⁷³ The hypothesis that the target responsible for the phenotype of defective glucose homeostasis was the LDLR is supported by the lack of changes of LDLR expression in β cells from mice selectively deficient in liver *PCSK9*. Overall, pharmacological approaches aimed at targeting liver *PCSK9* may not be associated with an increased risk of developing diabetes mellitus.¹⁷⁴

Author contribution

CM wrote the manuscript. CRS and MR conceived the topic and wrote the manuscript. AC conceived the topic and critically revised the manuscript. MB critically revised the manuscript. NF wrote and critically revised the manuscript.

All gave final approval and agree to be accountable for all aspects of work ensuring integrity and accuracy.

Declaration of conflicting interests

The authors declared the following potential conflicts of interest with respect to the research, authorship and/or publication of this article: CM, CRS and MR have nothing to declare. MB speakers bureau: Abbott/Mylan, Abbott Vascular, Actavis, Akcea, Amgen, Biofarm, KRKA, MSD, Sanofi-Aventis, Servier and Valeant; consultant to Abbott Vascular, Akcea, Amgen, Daichii Sankyo, Esperion, Lilly, MSD, Resverlogix, Sanofi-Aventis; grants from Sanofi and Valeant. NF has received honoraria from DOC, Mylan Pfizer and AC has received honoraria from AstraZeneca, AMGEN, Sanofi, Recordati, Novartis, MSD, Mediolanum, DOC, Mylan and Pfizer.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported by Fondazione Cariplo to MR (2015-0552) and Intramural Grant by Università degli Studi di Milano to MR (piano di sostegno alla ricerca dotazione 2018 LINEA 2 AZIONE A).

References

1. Ference BA, Ginsberg HN, Graham I, et al. Low-density lipoproteins cause atherosclerotic cardiovascular disease. 1. Evidence from genetic, epidemiologic, and clinical studies. A consensus statement from the European Atherosclerosis Society Consensus Panel. *Eur Heart J* 2017; 38: 2459–2472.
2. Fulcher J, O'Connell R, et al. Cholesterol Treatment Trialists C Efficacy and safety of LDL-lowering therapy among men and women: Meta-analysis of individual data from 174,000 participants in 27 randomised trials. *Lancet* 2015; 385: 1397–1405 [AQ24].
3. Sabatine MS, Wiviott SD, Im K, et al. Efficacy and safety of further lowering of low-density lipoprotein cholesterol in patients starting with very low levels: A meta-analysis. *JAMA Cardiol* 2018; 3: 823–828 [AQ25].
4. Koenig W, Giovas P and Nicholls SJ. Combining cholesterol-lowering strategies with imaging data: A visible benefit? *Eur J Prev Cardiol*. Epub ahead of print 30 August 2018. DOI: 10.1177/2047487318798059 [AQ26].
5. Ference BA, Graham I, Tokgozoglul L, et al. Impact of lipids on cardiovascular health: JACC Health Promotion Series. *J Am Coll Cardiol* 2018; 72: 1141–1156.
6. De Backer GG. Prevention of cardiovascular disease: Much more is needed. *Eur J Prev Cardiol* 2018; 25: 1083–1086.
7. Cohen JC, Boerwinkle E, Mosley TH Jr, et al. Sequence variations in PCSK9, low LDL, and protection against coronary heart disease. *N Engl J Med* 2006; 354: 1264–1272.
8. Myocardial Infarction Genetics C, Kathiresan S, Voight BF, et al. Genome-wide association of early-onset myocardial infarction with single nucleotide polymorphisms and copy number variants. *Nat Genet* 2009; 41: 334–341 [AQ27].
9. Abifadel M, Varret M, Rabes JP, et al. Mutations in PCSK9 cause autosomal dominant hypercholesterolemia. *Nat Genet* 2003; 34: 154–156.
10. Rosenson RS, Hegele RA, Fazio S, et al. The evolving future of PCSK9 inhibitors. *J Am Coll Cardiol* 2018; 72: 314–329.
11. Cheng JM, Oemrawsingh RM, Garcia-Garcia HM, et al. PCSK9 in relation to coronary plaque inflammation: Results of the ATHEROREMO-IVUS study. *Atherosclerosis* 2016; 248: 117–122.
12. Ferri N, Marchiano S, Tibolla G, et al. PCSK9 knockout mice are protected from neointimal formation in response to perivascular carotid collar placement. *Atherosclerosis* 2016; 253: 214–224.
13. Lee CJ, Lee YH, Park SW, et al. Association of serum proprotein convertase subtilisin/kexin type 9 with carotid intima media thickness in hypertensive subjects. *Metabolism* 2013; 62: 845–850.
14. Ruscica M, Ferri N, Fogacci F, et al. Circulating levels of proprotein convertase subtilisin/kexin type 9 and arterial stiffness in a large population sample: Data from the Brisighella Heart Study. *J Am Heart Assoc* 2017; 6: e005764 [AQ28].
15. Ruscica M, Ferri N, Macchi C, et al. Liver fat accumulation is associated with circulating PCSK9. *Ann Med* 2016; 48: 384–391.
16. Ferri N, Tibolla G, Pirillo A, et al. Proprotein convertase subtilisin kexin type 9 (PCSK9) secreted by cultured smooth muscle cells reduces macrophages LDLR levels. *Atherosclerosis* 2012; 220: 381–386.
17. Giunzioni I, Tavori H, Covarrubias R, et al. Local effects of human PCSK9 on the atherosclerotic lesion. *J Pathol* 2016; 238: 52–62.
18. Camera M, Rossetti L, Barbieri SS, et al. PCSK9 as a positive modulator of platelet activation. *J Am Coll Cardiol* 2018; 71: 952–954.
19. Ricci C, Ruscica M, Camera M, et al. PCSK9 induces a pro-inflammatory response in macrophages. *Sci Rep* 2018; 8: 2267.
20. Poggio P, Songia P, Cavallotti L, et al. PCSK9 involvement in aortic valve calcification. *J Am Coll Cardiol* 2018; 72: 3225–3227.
21. Seidah NG, Chretien M and Mbikay M. The ever-expanding saga of the proprotein convertases and their roles in body homeostasis: Emphasis on novel proprotein convertase subtilisin kexin number 9 functions and regulation. *Curr Opin Lipidol* 2018; 29: 144–150.
22. Lakoski SG, Lagace TA, Cohen JC, et al. Genetic and metabolic determinants of plasma PCSK9 levels. *J Clin Endocrinol Metab* 2009; 94: 2537–2543.
23. Spolitu S, Okamoto H, Dai W, et al. Hepatic glucagon signaling regulates PCSK9 and LDL-Cholesterol. *Circ Res* 2019; 124: 38–51 [AQ29].
24. Persson L, Cao G, Stahle L, et al. Circulating proprotein convertase subtilisin kexin type 9 has a diurnal rhythm synchronous with cholesterol synthesis and is reduced by fasting in humans. *Arterioscler Thromb Vasc Biol* 2010; 30: 2666–2672.

25. Artenstein AW and Opal SM. Proprotein convertases in health and disease. *N Engl J Med* 2011; 365: 2507–2518.
26. Seidah NG, Mayer G, Zaid A, et al. The activation and physiological functions of the proprotein convertases. *Int J Biochem Cell Biol* 2008; 40: 1111–1125.
27. Seidah NG and Prat A. The biology and therapeutic targeting of the proprotein convertases. *Nat Rev Drug Discov* 2012; 11: 367–383.
28. Zhang DW, Lagace TA, Garuti R, et al. Binding of proprotein convertase subtilisin/kexin type 9 to epidermal growth factor-like repeat A of low density lipoprotein receptor decreases receptor recycling and increases degradation. *J Biol Chem* 2007; 282: 18602–18612.
29. Mayne J, Dewpura T, Raymond A, et al. Novel loss-of-function PCSK9 variant is associated with low plasma LDL cholesterol in a French-Canadian family and with impaired processing and secretion in cell culture. *Clin Chem* 2011; 57: 1415–1423.
30. Benjannet S, Hamelin J, Chretien M, et al. Loss- and gain-of-function PCSK9 variants: Cleavage specificity, dominant negative effects, and low density lipoprotein receptor (LDLR) degradation. *J Biol Chem* 2012; 287: 33745–33755.
31. Cunningham D, Danley DE, Geoghegan KF, et al. Structural and biophysical studies of PCSK9 and its mutants linked to familial hypercholesterolemia. *Nat Struct Mol Biol* 2007; 14: 413–419.
32. Piper DE, Jackson S, Liu Q, et al. The crystal structure of PCSK9: A regulator of plasma LDL-cholesterol. *Structure* 2007; 15: 545–552.
33. Zhao Z, Tuakli-Wosornu Y, Lagace TA, et al. Molecular characterization of loss-of-function mutations in PCSK9 and identification of a compound heterozygote. *Am J Hum Genet* 2006; 79: 514–523.
34. Cariou B, Ouguerram K, Zair Y, et al. PCSK9 dominant negative mutant results in increased LDL catabolic rate and familial hypobetalipoproteinemia. *Arterioscler Thromb Vasc Biol* 2009; 29: 2191–2197.
35. Yamamoto T, Lu C and Ryan RO. A two-step binding model of PCSK9 interaction with the low density lipoprotein receptor. *J Biol Chem* 2011; 286: 5464–5470.
36. Essalmani R, Susan-Resiga D, Chamberland A, et al. In vivo evidence that furin from hepatocytes inactivates PCSK9. *J Biol Chem* 2011; 286: 4257–4263.
37. Gauthier MS, Awan Z, Bouchard A, et al. Posttranslational modification of proprotein convertase subtilisin/kexin type 9 is differentially regulated in response to distinct cardiometabolic treatments as revealed by targeted proteomics. *J Clin Lipidol* 2018; 12: 1027–1038.
38. Tavori H, Giunzioni I, Linton MF, et al. Loss of plasma proprotein convertase subtilisin/kexin 9 (PCSK9) after lipoprotein apheresis. *Circ Res* 2013; 113: 1290–1295.
39. Shapiro MD, Tavori H and Fazio S. PCSK9: From basic science discoveries to clinical trials. *Circ Res* 2018; 122: 1420–1438.
40. Romagnuolo R, Scipione CA, Boffa MB, et al. Lipoprotein(a) catabolism is regulated by proprotein convertase subtilisin/kexin type 9 through the low density lipoprotein receptor. *J Biol Chem* 2015; 290: 11649–11662.
41. Viney NJ, Yeang C, Yang X, et al. Relationship between ‘LDL-C’, estimated true LDL-C, apolipoprotein B-100, and PCSK9 levels following lipoprotein(a) lowering with an antisense oligonucleotide. *J Clin Lipidol* 2018; 12: 702–710.
42. Ferri N, Corsini A, Macchi C, et al. Proprotein convertase subtilisin kexin type 9 and high-density lipoprotein metabolism: Experimental animal models and clinical evidence. *Transl Res* 2016; 173: 19–29.
43. Ruscica M, Simonelli S, Botta M, et al. Plasma PCSK9 levels and lipoprotein distribution are preserved in carriers of genetic HDL disorders. *Biochim Biophys Acta* 2018; 1863: 991–997.
44. Seidah NG, Benjannet S, Wickham L, et al. The secretory proprotein convertase neural apoptosis-regulated convertase 1 (NARC-1): Liver regeneration and neuronal differentiation. *Proc Natl Acad Sci U S A* 2003; 100: 928–933.
45. Horton JD, Shah NA, Warrington JA, et al. Combined analysis of oligonucleotide microarray data from transgenic and knockout mice identifies direct SREBP target genes. *Proc Natl Acad Sci U S A* 2003; 100: 12027–12032.
46. Horton JD, Goldstein JL and Brown MS. SREBPs: Activators of the complete program of cholesterol and fatty acid synthesis in the liver. *J Clin Invest* 2002; 109: 1125–1131.
47. Bennett MK, Toth JI and Osborne TF. Selective association of sterol regulatory element-binding protein isoforms with target promoters in vivo. *J Biol Chem* 2004; 279: 37360–37367.
48. Sun LP, Li L, Goldstein JL, et al. Insig required for sterol-mediated inhibition of Scap/SREBP binding to COPII proteins in vitro. *J Biol Chem* 2005; 280: 26483–26490.
49. Shao W and Espenshade PJ. Sterol regulatory element-binding protein (SREBP) cleavage regulates Golgi-to-endoplasmic reticulum recycling of SREBP cleavage-activating protein (SCAP). *J Biol Chem* 2014; 289: 7547–7557.
50. Dubuc G, Chamberland A, Wassef H, et al. Statins upregulate PCSK9, the gene encoding the proprotein convertase neural apoptosis-regulated convertase-1 implicated in familial hypercholesterolemia. *Arterioscler Thromb Vasc Biol* 2004; 24: 1454–1459.
51. Pramfalk C, Jiang ZY, Cai Q, et al. HNF1alpha and SREBP2 are important regulators of NPC1L1 in human liver. *J Lipid Res* 2010; 51: 1354–1362.
52. Shende VR, Wu M, Singh AB, et al. Reduction of circulating PCSK9 and LDL-C levels by liver-specific knockdown of HNF1alpha in normolipidemic mice. *J Lipid Res* 2015; 56: 801–809.
53. Ruscica M, Ricci C, Macchi C, et al. Suppressor of cytokine signaling-3 (SOCS-3) induces proprotein convertase subtilisin kexin type 9 (PCSK9) expression in hepatic HepG2 cell line. *J Biol Chem* 2016; 291: 3508–3519.
54. Li H, Dong B, Park SW, et al. Hepatocyte nuclear factor 1alpha plays a critical role in PCSK9 gene transcription and regulation by the natural hypocholesterolemic compound berberine. *J Biol Chem* 2009; 284: 28885–28895.

55. Li H and Liu J. The novel function of HINFP as a co-activator in sterol-regulated transcription of PCSK9 in HepG2 cells. *Biochem J* 2012; 443: 757–768.
56. Tao R, Xiong X, DePinho RA, et al. FoxO3 transcription factor and Sirt6 deacetylase regulate low density lipoprotein (LDL)-cholesterol homeostasis via control of the proprotein convertase subtilisin/kexin type 9 (Pcsk9) gene expression. *J Biol Chem* 2013; 288: 29252–29259.
57. Poirier S, Samami S, Mamarbachi M, et al. The epigenetic drug 5-azacytidine interferes with cholesterol and lipid metabolism. *J Biol Chem* 2014; 289: 18736–18751.
58. Naeli P, Mirzadeh Azad F, Malakootian M, et al. Post-transcriptional regulation of PCSK9 by miR-191, miR-222, and miR-224. *Front Genet* 2017; 8: 189.
59. Reiner Z. Statins in the primary prevention of cardiovascular disease. *Nat Rev Cardiol* 2013; 10: 453–464.
60. Hobbs FD, Banach M, Mikhailidis DP, et al. Is statin-modified reduction in lipids the most important preventive therapy for cardiovascular disease? A pro/con debate. *BMC Med* 2016; 14: 4.
61. Dong B, Wu M, Li H, et al. Strong induction of PCSK9 gene expression through HNF1alpha and SREBP2: mechanism for the resistance to LDL-cholesterol lowering effect of statins in dyslipidemic hamsters. *J Lipid Res* 2010; 51: 1486–1495.
62. Dong B, Singh AB, Shende VR, et al. Hepatic HNF1 transcription factors control the induction of PCSK9 mediated by rosuvastatin in normolipidemic hamsters. *Int J Mol Med* 2017; 39: 749–756.
63. Nozue T. Lipid lowering therapy and circulating PCSK9 concentration. *J Atheroscler Thromb* 2017; 24: 895–907.
64. Sahebkar A, Simental-Mendia LE, Guerrero-Romero F, et al. Effect of statin therapy on plasma proprotein convertase subtilisin kexin 9 (PCSK9) concentrations: A systematic review and meta-analysis of clinical trials. *Diabetes Obes Metab* 2015; 17: 1042–1055.
65. Guo YL, Liu J, Xu RX, et al. Short-term impact of low-dose atorvastatin on serum proprotein convertase subtilisin/kexin type 9. *Clin Drug Investig* 2013; 33: 877–883.
66. Careskey HE, Davis RA, Alborn WE, et al. Atorvastatin increases human serum levels of proprotein convertase subtilisin/kexin type 9. *J Lipid Res* 2008; 49: 394–398.
67. Tremblay AJ, Lamarche B, Lemelin V, et al. Atorvastatin increases intestinal expression of NPC1L1 in hyperlipidemic men. *J Lipid Res* 2011; 52: 558–565.
68. Welder G, Zineh I, Pacanowski MA, et al. High-dose atorvastatin causes a rapid sustained increase in human serum PCSK9 and disrupts its correlation with LDL cholesterol. *J Lipid Res* 2010; 51: 2714–2721.
69. Zhang J, Long M and Yu Y. The effects of additional ezetimibe treatment to baseline rosuvastatin on circulating PCSK9 among patients with stable angina. *J Thorac Dis* 2017; 9: 1226–1233.
70. Dubuc G, Tremblay M, Pare G, et al. A new method for measurement of total plasma PCSK9: Clinical applications. *J Lipid Res* 2010; 51: 140–149.
71. Raal F, Panz V, Immelman A, et al. Elevated PCSK9 levels in untreated patients with heterozygous or homozygous familial hypercholesterolemia and the response to high-dose statin therapy. *J Am Heart Assoc* 2013; 2: e000028.
72. Berthold HK, Seidah NG, Benjannet S, et al. Evidence from a randomized trial that simvastatin, but not ezetimibe, upregulates circulating PCSK9 levels. *PLoS One* 2013; 8: e60095.
73. Chen F, Maridakis V, O'Neill EA, et al. The effects of simvastatin treatment on plasma lipid-related biomarkers in men with dyslipidaemia. *Biomarkers* 2011; 16: 321–333.
74. Lakoski SG, Xu F, Vega GL, et al. Indices of cholesterol metabolism and relative responsiveness to ezetimibe and simvastatin. *J Clin Endocrinol Metab* 2010; 95: 800–809.
75. Ferri N, Marchiano S, Lupo MG, et al. Geranylgeraniol prevents the simvastatin-induced PCSK9 expression: Role of the small G protein Rac1. *Pharmacol Res* 2017; 122: 96–104.
76. Nozue T, Hattori H, Ishihara M, et al. Comparison of effects of pitavastatin versus pravastatin on serum proprotein convertase subtilisin/kexin type 9 levels in statin-naive patients with coronary artery disease. *Am J Cardiol* 2013; 111: 1415–1419.
77. Telford DE, Sutherland BG, Edwards JY, et al. The molecular mechanisms underlying the reduction of LDL apoB-100 by ezetimibe plus simvastatin. *J Lipid Res* 2007; 48: 699–708.
78. Miyoshi T, Nakamura K, Doi M, et al. Impact of ezetimibe alone or in addition to a statin on plasma PCSK9 concentrations in patients with type 2 diabetes and hypercholesterolemia: A pilot study. *Am J Cardiovasc Drugs* 2015; 15: 213–219.
79. Davignon J and Dubuc G. Statins and ezetimibe modulate plasma proprotein convertase subtilisin kexin-9 (PCSK9) levels. *Trans Am Clin Climatol Assoc* 2009; 120: 163–173.
80. Cohen JC, Pertsemlidis A, Fahmi S, et al. Multiple rare variants in NPC1L1 associated with reduced sterol absorption and plasma low-density lipoprotein levels. *Proc Natl Acad Sci U S A* 2006; 103: 1810–1815.
81. Silbernagel G, Steiner LK, Hollstein T, et al. The interrelations between PCSK9 metabolism and cholesterol synthesis and absorption. *J Lipid Res* 2019; 60: 161–167.
82. Ferri N, Corsini A, Sirtori C, et al. PPAR-alpha agonists are still on the rise: An update on clinical and experimental findings. *Expert Opin Investig Drugs* 2017; 26: 593–602.
83. Post SM, Duez H, Gervois PP, et al. Fibrates suppress bile acid synthesis via peroxisome proliferator-activated receptor-alpha-mediated downregulation of cholesterol 7alpha-hydroxylase and sterol 27-hydroxylase expression. *Arterioscler Thromb Vasc Biol* 2001; 21: 1840–1845.
84. Lambert G, Jarnoux AL, Pineau T, et al. Fasting induces hyperlipidemia in mice overexpressing proprotein convertase subtilisin kexin type 9: Lack of modulation of very-low-density lipoprotein hepatic output by the low-density lipoprotein receptor. *Endocrinology* 2006; 147: 4985–4995.
85. Mayne J, Dewapura T, Raymond A, et al. Plasma PCSK9 levels are significantly modified by statins and fibrates in humans. *Lipids Health Dis* 2008; 7: 22.

86. Troutt JS, Alborn WE, Cao G, et al. Fenofibrate treatment increases human serum proprotein convertase subtilisin kexin type 9 levels. *J Lipid Res* 2010; 51: 345–351.
87. Lambert G, Ancellin N, Charlton F, et al. Plasma PCSK9 concentrations correlate with LDL and total cholesterol in diabetic patients and are decreased by fenofibrate treatment. *Clin Chem* 2008; 54: 1038–1045.
88. Noguchi T, Kobayashi J, Yagi K, et al. Comparison of effects of bezafibrate and fenofibrate on circulating proprotein convertase subtilisin/kexin type 9 and adipocytokine levels in dyslipidemic subjects with impaired glucose tolerance or type 2 diabetes mellitus: Results from a crossover study. *Atherosclerosis* 2011; 217: 165–170.
89. Costet P, Hoffmann MM, Cariou B, et al. Plasma PCSK9 is increased by fenofibrate and atorvastatin in a non-additive fashion in diabetic patients. *Atherosclerosis* 2010; 212: 246–251.
90. Chan DC, Hamilton SJ, Rye KA, et al. Fenofibrate concomitantly decreases serum proprotein convertase subtilisin/kexin type 9 and very-low-density lipoprotein particle concentrations in statin-treated type 2 diabetic patients. *Diabetes Obes Metab* 2010; 12: 752–756.
91. Brewer HB. Comparative evaluation of fibric acid derivatives and HMG-CoA reductase inhibitors. *Atherosclerosis* 1992; 97: S11–S19.
92. Kourimate S, Le May C, Langhi C, et al. Dual mechanisms for the fibrate-mediated repression of proprotein convertase subtilisin/kexin type 9. *J Biol Chem* 2008; 283: 9666–9673.
93. Sirtori CR, Galli C and Franceschini G. Fraudulent (and non fraudulent) fatty acids for human health. *Eur J Clin Invest* 1993; 23: 686–689.
94. Botta M, Audano M, Sahebkar A, et al. PPAR agonists and metabolic syndrome: An established role?. *Int J Mol Sci* 2018; 19: 1197 **[AQ30]**.
95. Costet P, Cariou B, Lambert G, et al. Hepatic PCSK9 expression is regulated by nutritional status via insulin and sterol regulatory element-binding protein 1c. *J Biol Chem* 2006; 281: 6211–6218.
96. Jump DB. N-3 polyunsaturated fatty acid regulation of hepatic gene transcription. *Curr Opin Lipidol* 2008; 19: 242–247.
97. Zimmer M, Bista P, Benson EL, et al. CAT-2003: A novel sterol regulatory element-binding protein inhibitor that reduces steatohepatitis, plasma lipids, and atherosclerosis in apolipoprotein E*3-Leiden mice. *Hepatology* 2017; 1: 311–325.
98. Graversen CB, Lundbye-Christensen S, Thomsen B, et al. Marine n-3 polyunsaturated fatty acids lower plasma proprotein convertase subtilisin kexin type 9 levels in pre- and postmenopausal women: A randomized study. *Vascul Pharmacol* 2016; 76: 37–41.
99. Ferri N, Corsini A, Sirtori CR, et al. Present therapeutic role of cholesteryl ester transfer protein inhibitors. *Pharmacol Res* 2018; 128: 29–41.
100. Millar JS, Reyes-Soffer G, Jumes P, et al. Anacetrapib lowers LDL by increasing ApoB clearance in mildly hypercholesterolemic subjects. *J Clin Invest* 2015; 125: 2510–2522.
101. Girona J, Ibarretxe D, Plana N, et al. Circulating PCSK9 levels and CETP plasma activity are independently associated in patients with metabolic diseases. *Cardiovasc Diabetol* 2016; 15: 107.
102. Arsenault BJ, Petrides F, Tabet F, et al. Effect of atorvastatin, cholesterol ester transfer protein inhibition, and diabetes mellitus on circulating proprotein subtilisin kexin type 9 and lipoprotein(a) levels in patients at high cardiovascular risk. *J Clin Lipidol* 2018; 12: 130–136.
103. Khera AV, Qamar A, Reilly MP, et al. Effects of niacin, statin, and fenofibrate on circulating proprotein convertase subtilisin/kexin type 9 levels in patients with dyslipidemia. *Am J Cardiol* 2015; 115: 178–182.
104. Poli A, Barbagallo CM, Cicero AFG, et al. Nutraceuticals and functional foods for the control of plasma cholesterol levels. An intersociety position paper. *Pharmacol Res* 2018; 134: 51–60.
105. Momtazi AA, Banach M, Pirro M, et al. Regulation of PCSK9 by nutraceuticals. *Pharmacol Res* 2017; 120: 157–169.
106. Johnston TP, Korolenko TA, Pirro M, et al. Preventing cardiovascular heart disease: Promising nutraceutical and non-nutraceutical treatments for cholesterol management. *Pharmacol Res* 2017; 120: 219–225.
107. Kong W, Wei J, Abidi P, et al. Berberine is a novel cholesterol-lowering drug working through a unique mechanism distinct from statins. *Nat Med* 2004; 10: 1344–1351.
108. Cameron J, Ranheim T, Kulseth MA, et al. Berberine decreases PCSK9 expression in HepG2 cells. *Atherosclerosis* 2008; 201: 266–273.
109. Dong B, Li H, Singh AB, et al. Inhibition of PCSK9 transcription by berberine involves down-regulation of hepatic HNF1alpha protein expression through the ubiquitin-proteasome degradation pathway. *J Biol Chem* 2015; 290: 4047–4058.
110. Pisciotta L, Bellocchio A and Bertolini S. Nutraceutical pill containing berberine versus ezetimibe on plasma lipid pattern in hypercholesterolemic subjects and its additive effect in patients with familial hypercholesterolemia on stable cholesterol-lowering treatment. *Lipids Health Dis* 2012; 11: 123.
111. Adorni MP, Ferri N, Marchiano S, et al. Effect of a novel nutraceutical combination on serum lipoprotein functional profile and circulating PCSK9. *Ther Clin Risk Manag* 2017; 13: 1555–1562.
112. Ruscica M, Gomasaschi M, Mombelli G, et al. Nutraceutical approach to moderate cardiometabolic risk: Results of a randomized, double-blind and crossover study with Armolipid Plus. *J Clin Lipidol* 2014; 8: 61–68.
113. Banach M, Patti AM, Giglio RV, et al. The role of nutraceuticals in statin intolerant patients. *J Am Coll Cardiol* 2018; 72: 96–118.
114. Zhang Y, Liu J, Li S, et al. Impact of currently prescribed lipid-lowering drugs on plasma PCSK9 concentration: Single or in combination study in rats. *Lipids Health Dis* 2014; 13: 35.
115. Spigoni V, Aldigeri R, Antonini M, et al. Effects of a New nutraceutical formulation (berberine, red yeast rice

- and chitosan) on non-HDL cholesterol levels in individuals with dyslipidemia: Results from a randomized, double blind, placebo-controlled study. *Int J Mol Sci* 2017; 18: 1498 [AQ31].
116. Ferri N, Corsini A, Sirtori CR, et al. Bococizumab for the treatment of hypercholesterolaemia. *Expert Opin Biol Ther* 2017; 17: 237–243.
 117. Ferri N, Corsini A, Sirtori CR, et al. Bococizumab for the treatment of hypercholesterolaemia. *Expert Opin Biol Ther* 2017; 17: 909–910 [AQ32].
 118. Kasichayanula S, Grover A, Emery MG, et al. Clinical pharmacokinetics and pharmacodynamics of evolocumab, a PCSK9 inhibitor. *Clin Pharmacokinet* 2018; 57: 769–779.
 119. Farnier M. An evaluation of alirocumab for the treatment of hypercholesterolemia. *Expert Rev Cardiovasc Ther* 2015; 13: 1307–1323.
 120. Pecin I, Hartgers ML, Hovingh GK, et al. Prevention of cardiovascular disease in patients with familial hypercholesterolaemia: The role of PCSK9 inhibitors. *Eur J Prev Cardiol* 2017; 24: 1383–1401.
 121. Robinson JG, Huijgen R, Ray K, et al. Determining when to add nonstatin therapy: A quantitative approach. *J Am Coll Cardiol* 2016; 68: 2412–2421.
 122. Landmesser U, Chapman MJ, Stock JK, et al. 2017 Update of ESC/EAS Task Force on practical clinical guidance for proprotein convertase subtilisin/kexin type 9 inhibition in patients with atherosclerotic cardiovascular disease or in familial hypercholesterolaemia. *Eur Heart J* 2018; 39: 1131–1143.
 123. Sabatine MS, Giugliano RP, Keech AC, et al. Evolocumab and clinical outcomes in patients with cardiovascular disease. *N Engl J Med* 2017; 376: 1713–1722.
 124. O'Donoghue ML, Fazio S, Giugliano RP, et al. Lipoprotein(a), PCSK9 inhibition and cardiovascular risk: Insights from the FOURIER Trial. *Circulation*. Epub ahead of print 30 November 2018. DOI: 10.1161/CIRCULATIONAHA.118.037184 [AQ33].
 125. Sabatine MS, Leiter LA, Wiviott SD, et al. Cardiovascular safety and efficacy of the PCSK9 inhibitor evolocumab in patients with and without diabetes and the effect of evolocumab on glycaemia and risk of new-onset diabetes: A prespecified analysis of the FOURIER randomised controlled trial. *Lancet Diabetes Endocrinol* 2017; 5: 941–950.
 126. Sabatine MS, De Ferrari GM, Giugliano RP, et al. Clinical benefit of evolocumab by severity and extent of coronary artery disease: An analysis from FOURIER. *Circulation*. DOI: 10.1161/CIRCULATIONAHA.118.034309 [AQ34].
 127. Bonaca MP, Nault P, Giugliano RP, et al. Low-density lipoprotein cholesterol lowering with evolocumab and outcomes in patients with peripheral artery disease: Insights from the FOURIER Trial (Further Cardiovascular Outcomes Research With PCSK9 Inhibition in Subjects With Elevated Risk). *Circulation* 2018; 137: 338–350.
 128. Giugliano RP, Mach F, Zavitz K, et al. Cognitive function in a randomized trial of evolocumab. *N Engl J Med* 2017; 377: 633–643.
 129. Nicholls SJ, Puri R, Anderson T, et al. Effect of evolocumab on progression of coronary disease in statin-treated patients: The GLAGOV randomized clinical trial. *JAMA* 2016; 316: 2373–2384.
 130. Nicholls SJ, Puri R, Anderson T, et al. Effect of evolocumab on coronary plaque composition. *J Am Coll Cardiol* 2018; 72: 2012–2021.
 131. Wilson PWF, Polonsky TS, Miedema MD, et al. Systematic review for the 2018 AHA/ACC/AACVPR/AAPA/ABC/ACPM/ADA/AGS/APhA/ASPC/NLA/PCNA guideline on the management of blood cholesterol: A report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. *J Am Coll Cardiol*. Epub ahead of print 3 November 2018. DOI: 10.1016/j.jacc.2018.11.004 [AQ35].
 132. Schwartz GG, Steg PG, Szarek M, et al. Alirocumab and cardiovascular outcomes after acute coronary syndrome. *N Engl J Med* 2018; 379: 2097–2107 [AQ36].
 133. Szarek M, White HD, Schwartz GG, et al. Alirocumab reduces total nonfatal cardiovascular and fatal events in the ODYSSEY OUTCOMES trial. *J Am Coll Cardiol*. Epub ahead of print 27 October 2018. DOI: 10.1016/j.jacc.2018.10.039 [AQ37].
 134. Hadjiphilippou S and Ray KK. PCSK9 inhibition and atherosclerotic cardiovascular disease prevention: Does reality match the hype? *Heart* 2017; 103: 1670–1679.
 135. Otake H, Sugizaki Y, Toba T, et al. Efficacy of alirocumab for reducing plaque vulnerability: Study protocol for ALTAIR, a randomized controlled trial in Japanese patients with coronary artery disease receiving rosuvastatin. *J Cardiol*. Epub before print 19 December 2018. DOI: 10.1016/j.jcc.2018.11.012 [AQ38].
 136. Kereiakes DJ, Lepor NE, Gerber R, et al. Efficacy and safety of alirocumab in patients with or without prior coronary revascularization: Pooled analysis of eight ODYSSEY phase 3 trials. *Atherosclerosis* 2018; 277: 211–218 [AQ39].
 137. Leiter LA, Tinahones FJ, Karalis DG, et al. Alirocumab safety in people with and without diabetes mellitus: Pooled data from 14 ODYSSEY trials. *Diabet Med* 2018; 35: 1742–1751.
 138. Taskinen MR, Del Prato S, Bujas-Bobanovic M, et al. Efficacy and safety of alirocumab in individuals with type 2 diabetes mellitus with or without mixed dyslipidaemia: Analysis of the ODYSSEY LONG TERM trial. *Atherosclerosis* 2018; 276: 124–130.
 139. Khvorova A. Oligonucleotide therapeutics – a new class of cholesterol-lowering drugs. *N Engl J Med* 2017; 376: 4–7.
 140. Fitzgerald K, Frank-Kamenetsky M, Shulga-Morskaya S, et al. Effect of an RNA interference drug on the synthesis of proprotein convertase subtilisin/kexin type 9 (PCSK9) and the concentration of serum LDL cholesterol in healthy volunteers: A randomised, single-blind, placebo-controlled, phase 1 trial. *Lancet* 2014; 383: 60–68.
 141. Fitzgerald K, White S, Borodovsky A, et al. A highly durable RNAi therapeutic inhibitor of PCSK9. *N Engl J Med* 2017; 376: 41–51.

142. Ray KK, Landmesser U, Leiter LA, et al. Inclisiran in patients at high cardiovascular risk with elevated LDL cholesterol. *N Engl J Med* 2017; 376: 1430–1440.
143. Leiter LA, Teoh H, Kallend D, et al. Inclisiran lowers LDL-C and PCSK9 irrespective of diabetes status: The ORION-1 randomized clinical trial. *Diabetes Care* 2018; 42: 173–176 [AQ40].
144. Ray KK, Stoekenbroek RM, Kallend D, et al. Effect of an siRNA therapeutic targeting PCSK9 on atherogenic lipoproteins: Pre-specified secondary end points in ORION 1. *Circulation* 2018; 138: 1304–1316 [AQ41].
145. Werner C, Hoffmann MM, Winkler K, et al. Risk prediction with proprotein convertase subtilisin/kexin type 9 (PCSK9) in patients with stable coronary disease on statin treatment. *Vascul Pharmacol* 2014; 62: 94–102.
146. Zhu YM, Anderson TJ, Sikdar K, et al. Association of proprotein convertase subtilisin/kexin type 9 (PCSK9) with cardiovascular risk in primary prevention. *Arterioscler Thromb Vasc Biol* 2015; 35: 2254–2259.
147. Ridker PM, Rifai N, Bradwin G, et al. Plasma proprotein convertase subtilisin/kexin type 9 levels and the risk of first cardiovascular events. *Eur Heart J* 2016; 37: 554–560.
148. Gencer B, Montecucco F, Nanchen D, et al. Prognostic value of PCSK9 levels in patients with acute coronary syndromes. *Eur Heart J* 2016; 37: 546–553.
149. Vlachopoulos C, Terentes-Printzios D, Georgiopoulos G, et al. Prediction of cardiovascular events with levels of proprotein convertase subtilisin/kexin type 9: A systematic review and meta-analysis. *Atherosclerosis* 2016; 252: 50–60.
150. Pott J, Schlegel V, Teren A, et al. Genetic regulation of PCSK9 (proprotein convertase subtilisin/kexin type 9) plasma levels and its impact on atherosclerotic vascular disease phenotypes. *Circ Genom Precis Med* 2018; 11: e001992.
151. Macchi C, Ferri N, Favero C, et al. Long-term exposure to air pollution raises circulating levels of proprotein convertase subtilisin/kexin type 9 in obese individuals. *Eur J Prev Cardiol*. Epub before print 26 November 2018. DOI: 10.1177/2047487318815320 [AQ42].
152. Ferri N and Ruscica M. Proprotein convertase subtilisin/kexin type 9 (PCSK9) and metabolic syndrome: Insights on insulin resistance, inflammation, and atherogenic dyslipidemia. *Endocrine* 2016; 54: 588–601.
153. El Khoury P, Roussel R, Fumeron F, et al. Plasma proprotein-convertase-subtilisin/kexin type 9 (PCSK9) and cardiovascular events in type 2 diabetes. *Diabetes Obes Metab* 2018; 20: 943–953.
154. Khan SU, Talluri S, Riaz H, et al. A Bayesian network meta-analysis of PCSK9 inhibitors, statins and ezetimibe with or without statins for cardiovascular outcomes. *Eur J Prev Cardiol* 2018; 25: 844–853.
155. Shapiro MD, Miles J, Tavori H, et al. Diagnosing resistance to a proprotein convertase subtilisin/kexin type 9 inhibitor. *Ann Intern Med* 2018; 168: 376–379.
156. Desai NR, Giugliano RP, Wasserman SM, et al. Association between circulating baseline proprotein convertase subtilisin kexin type 9 levels and efficacy of evolocumab. *JAMA Cardiol* 2017; 2: 556–560.
157. Reiner Z. PCSK9 inhibitors in clinical practice: Expectations and reality. *Atherosclerosis* 2018; 270: 187–188.
158. Grundy S, Stone N, Bailey A, et al. 2018 AHA/ACC/AACVPR/AAPA/ABC/ACPM/ADA/AGS/APhA/ASPC/NLA/PCNA guideline on the management of blood cholesterol: A report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. *Circulation*. Epub before print 10 November 2018. DOI: 10.1161/CIR.0000000000000625 [AQ43].
159. Virani S, Akeroyd J, Nambi V, et al. Applicability and cost implications for PCSK9 inhibitors based on the ODYSSEY OUTCOMES trial: Insights from the Department of Veterans Affairs. *Circulation* 2018; 139: 410–412 [AQ44].
160. Tunon J, Back M, Badimon L, et al. Interplay between hypercholesterolaemia and inflammation in atherosclerosis: Translating experimental targets into clinical practice. *Eur J Prev Cardiol* 2018; 25: 948–955.
161. Ruscica M, Ferri N, Corsini A, et al. PCSK9 antagonists and inflammation. *Atherosclerosis* 2018; 268: 235–236.
162. Ruscica M, Ferri N, Macchi C, et al. Lipid lowering drugs and inflammatory changes: An impact on cardiovascular outcomes? *Ann Med* 2018; 1–24 [AQ45].
163. Sliz E, Kettunen J, Holmes MV, et al. Metabolomic consequences of genetic inhibition of PCSK9 compared with statin treatment. *Circulation* 2018; 138: 2499–2512.
164. Kianoush S, Bittencourt MS, Lotufo PA, et al. Association between smoking and serum GlycA and high-sensitivity C-reactive protein levels: The Multi-Ethnic Study of Atherosclerosis (MESA) and Brazilian Longitudinal Study of Adult Health (ELSA-Brasil). *J Am Heart Assoc* 2017; 3: e001221 [AQ46].
165. Zenti MG, Altomari A, Lupo MG, et al. From lipoprotein apheresis to proprotein convertase subtilisin/kexin type 9 inhibitors: Impact on low-density lipoprotein cholesterol and C-reactive protein levels in cardiovascular disease patients. *Eur J Prev Cardiol* 2018; 25: 1843–1851 [AQ47].
166. Bohula EA, Giugliano RP, Leiter LA, et al. Inflammatory and cholesterol risk in the FOURIER trial. *Circulation* 2018; 138: 131–140.
167. Khademi F, Momtazi-Borojeni AA, Reiner Z, et al. PCSK9 and infection: A potentially useful or dangerous association?. *J Cell Physiol* 2018; 233: 2920–2927.
168. Nishikido T and Ray KK. Inclisiran for the treatment of dyslipidemia. *Expert Opin Investig Drugs* 2018; 27: 287–294.
169. Colhoun HM, Ginsberg HN, Robinson JG, et al. No effect of PCSK9 inhibitor alirocumab on the incidence of diabetes in a pooled analysis from 10 ODYSSEY Phase 3 studies. *Eur Heart J* 2016; 37: 2981–2989.
170. Ference BA, Robinson JG, Brook RD, et al. Variation in PCSK9 and HMGCR and risk of cardiovascular disease and diabetes. *N Engl J Med* 2016; 375: 2144–2153.
171. Lotta LA, Sharp SJ, Burgess S, et al. Association between low-density lipoprotein cholesterol-lowering

- genetic variants and risk of type 2 diabetes: A meta-analysis. *JAMA* 2016; 316: 1383–1391.
172. Schmidt AF, Swerdlow DI, Holmes MV, et al. PCSK9 genetic variants and risk of type 2 diabetes: A Mendelian randomisation study. *Lancet Diabetes Endocrinol* 2017; 5: 97–105.
173. Da Dalt L, Ruscica M, Bonacina F, et al. PCSK9 deficiency reduces insulin secretion and promotes glucose intolerance: The role of the low-density lipoprotein receptor. *Eur Heart J* 2018; 4: 357–368 [AQ48].
174. Paneni F and Costantino S. PCSK9 in diabetes: Sweet, bitter or sour? *Eur Heart J* 2018; 40: 369–371 [AQ49].
175. Abifadel M, Elbitar S, El Khoury P, et al. Living the PCSK9 adventure: From the identification of a new gene in familial hypercholesterolemia towards a potential new class of anticholesterol drugs. *Curr Atheroscler Rep* 2014; 16: 439.
176. Konrad RJ, Troutt JS and Cao G. Effects of currently prescribed LDL-C-lowering drugs on PCSK9 and implications for the next generation of LDL-C-lowering agents. *Lipids Health Dis* 2011; 10: 38.