

# Causal relationship of hepatic fat with liver damage and insulin resistance in nonalcoholic fatty liver

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**Abstract.** Dongiovanni P, Stender S, Pietrelli A, Mancina RM, Cespiati A, Petta S, Pelusi S, Pingitore P, Badiali S, Maggioni M, Mannisto V, Grimaudo S, Pipitone RM, Pihlajamaki J, Craxi A, Taube M, Carlsson LMS, Fargion S, Romeo S, Kozlitina J, Valenti L (Fondazione IRCCS Ca' Granda Ospedale Policlinico Milano, Milan, Italy; University of Texas Southwestern Medical Center, Dallas, TX, USA; Istituto Nazionale Genetica Molecolare, Milan, Italy; University of Gothenburg, Gothenburg, Sweden; Università di Palermo, Palermo, Italy; Kuopio University Hospital; University of Eastern Finland, Kuopio, Finland; Università degli Studi di Milano, Milan; University Magna Graecia, Catanzaro, Italy; Sahlgrenska University Hospital, Gothenburg, Sweden). Causal relationship of hepatic fat with liver damage and insulin resistance in nonalcoholic fatty liver. *J Intern Med* 2018; **283**: 356–370.

**Background and Aims.** Nonalcoholic fatty liver disease is epidemiologically associated with hepatic and metabolic disorders. The aim of this study was to examine whether hepatic fat accumulation has a causal role in determining liver damage and insulin resistance.

**Methods.** We performed a Mendelian randomization analysis using risk alleles in *PNPLA3*, *TM6SF2*, *GCKR* and *MBOAT7*, and a polygenic risk score for hepatic fat, as instruments. We evaluated complementary cohorts of at-risk individuals and individuals from the general population: 1515 from the

liver biopsy cohort (LBC), 3329 from the Swedish Obese Subjects Study (SOS) and 4570 from the population-based Dallas Heart Study (DHS).

**Results.** Hepatic fat was epidemiologically associated with liver damage, insulin resistance, dyslipidemia and hypertension. The impact of genetic variants on liver damage was proportional to their effect on hepatic fat accumulation. Genetically determined hepatic fat was associated with aminotransferases, and with inflammation, ballooning and fibrosis in the LBC. Furthermore, in the LBC, the causal association between hepatic fat and fibrosis was independent of disease activity, suggesting that a causal effect of long-term liver fat accumulation on liver disease is independent of inflammation. Genetically determined hepatic steatosis was associated with insulin resistance in the LBC and SOS. However, this association was dependent on liver damage severity. Genetically determined hepatic steatosis was associated with liver fibrosis/cirrhosis and with a small increase in risk of type 2 diabetes in publicly available databases.

**Conclusion.** These data suggest that long-term hepatic fat accumulation plays a causal role in the development of chronic liver disease.

**Keywords:** fibrosis, genetics, insulin resistance, mendelian randomization, nonalcoholic fatty liver disease, type 2 diabetes.

\*These authors contributed equally to the study.

## Introduction

Nonalcoholic fatty liver disease (NAFLD) is defined by the accumulation of fat in the liver, in the absence of excessive alcohol consumption. NAFLD encompasses a spectrum of conditions ranging from simple accumulation of excess fat (steatosis) to hepatic inflammation (nonalcoholic steatohepatitis or NASH) and fibrosis. Obesity and insulin resistance are major risk factors for NAFLD [1–3]. Concurrent with the increased prevalence of obesity, NAFLD has emerged as the most frequent liver disorder worldwide, affecting as many as 30% of adults in industrialized countries [4, 5].

Although simple steatosis is generally considered benign, NAFLD is associated with an increased risk of end-stage liver disease, as well as a range of extrahepatic metabolic disorders, including insulin resistance, type 2 diabetes (T2D), dyslipidemia and hypertension. However, the extent to which these associations reflect a causal effect of hepatic fat accumulation remains unclear. Associations observed in epidemiological studies may be influenced by confounding factors (e.g. diet, physical activity and microbiota) and/or reverse causation. Furthermore, hepatic fat accumulation tends to decrease with the progression of fibrosis ('burnt-out nonalcoholic steatohepatitis') [6], potentially limiting the ability of cross-sectional association studies to reliably evaluate the correlates of hepatic fat in patients with severe disease [7]. Weight-loss interventions aimed at reducing liver fat lead to improved metabolic outcomes [8]. However, it is unclear whether these changes are a direct result of the reduction in liver fat or a consequence of the improvement in extrahepatic insulin resistance or improved lifestyle [3].

Mendelian randomization is an epidemiological method that avoids confounding and reverse causation, by using genetic variation as an instrument to establish the causal role of modifiable risk factors in disease [9, 10]. The method relies on the assumption that as an individual's genotype is determined randomly at conception, is not related to lifestyle and other potential confounding factors, and thus can serve as an unconfounded and lifelong proxy for an exposure of interest (e.g. hepatic fat content). As our knowledge of common genetic variation governing variability in quantitative traits has improved over the past decade as a result of genomewide association studies (GWAS), Mendelian randomization has gained widespread

use as a method of testing causal effects of modifiable risk factors in situations where randomized controlled trials are not feasible or ethical [11]. For example, the approach has been used successfully to assess the causal relationship of different lipoprotein fractions with the risk of cardiovascular disease [11]. Previous genomewide association studies have identified several genetic risk factors for NAFLD, including variants in *PNPLA3* [12, 13], *TM6SF2* [14, 15], *GCKR* [13], *MBOAT7* [16, 17] and possibly *LYPLAL1* [13], which can be used as instruments to assess the causal effect of hepatic fat on progressive liver disease. Previous studies have shown most of these variants to be associated with NAFLD development and severity [12–14, 17, 18]. However, whether the magnitude of association of these genetic variants with progressive liver disease is concordant with an increase in risk predicted from their effect on hepatic fat accumulation has not been tested so far. Furthermore, whether genetically determined hepatic fat influences insulin resistance remains a controversial question [3, 19, 20].

Here, we leveraged a Mendelian randomization approach to examine whether hepatic fat causally determines liver damage and metabolic comorbidities. To this end, we evaluated complementary cohorts of individuals at risk of progressive NAFLD due to suspected NASH in the Liver Biopsy Cohort (LBC), or to severe obesity in the Swedish Obese Subjects Study (SOS), and individuals from the population-based Dallas Heart Study (DHS). We sequentially tested whether the steatosis-associated variants, individually or combined into a genetic risk score (GRS), were associated with liver damage (histological features of NAFLD, aminotransferases and noninvasive fibrosis scores) and clinical parameters epidemiologically associated with insulin resistance and NAFLD (presence of arterial hypertension, T2D, homeostasis metabolic assessment-insulin resistance index – HOMA-IR, and circulating HDL cholesterol).

## Materials and methods

### Study cohorts

Part of the LBC has been previously described [17, 21]. Briefly, a total of 1515 adult European individuals who underwent liver biopsy for suspected NASH or severe obesity were consecutively enrolled. Individuals with increased alcohol intake (men, >30 g day<sup>-1</sup>; women, >20 g day<sup>-1</sup>), use of steatogenic medications or other liver diseases

were excluded. Demographic and clinical features of patients included in the LBC were evaluated at the tertiary referral centres at the time of liver biopsy. The SOS is a prospective study comparing the effects of bariatric surgery and usual care in severely obese individuals in Sweden [22]. A total of 3329 subjects, who had complete baseline metabolic characterization and were successfully genotyped, were included. The DHS is a multiethnic population-based probability sample of Dallas County residents [23]. The present investigation included a total of 4570 individuals, who had provided blood samples for genetic and laboratory analysis. Of these, 2736 individuals had available measures of liver fat with proton magnetic resonance spectroscopy ( $^1\text{H-MRS}$ ) [1].

All studies were approved by the competent ethical committees, and all subjects gave written or oral informed consent. The study cohort characteristics and outcomes are summarized in Fig. 1, clinical features of individuals evaluated in the study are presented in Table 1, and

additional details concerning the study design are provided in Supporting information (Data S1) and in Figure S1.

#### Biochemical and clinical end-points

Hypertension was defined as systolic blood pressure  $\geq 140$  mm Hg or diastolic blood pressure  $\geq 90$  mm Hg, or self-reported antihypertensive treatment. T2D was defined as fasting blood glucose level  $\geq 126$  mg dL $^{-1}$ , nonfasting glucose  $\geq 200$  mg dL $^{-1}$ , HbA1c  $\geq 6.5\%$  or use of glucose-lowering medication. HOMA-IR was calculated from fasting levels of glucose and insulin [24]. Plasma levels of lipids and lipoproteins were measured by standard enzymatic assays. As most of the risk variants included in the genetic risk score have been shown to have pleiotropic effects on circulating cholesterol and triglycerides [18], they were not regarded as valid instruments for testing the causal effect of hepatic fat on these outcomes. Therefore, only HDL levels were considered as an outcome in this study.

Cohorts	Liver Biopsy Cohort (LBC); <i>n</i> = 1515	Swedish Obese Subjects Study (SOS); <i>n</i> = 3329	Dallas Heart Study (DHS); <i>n</i> = 4570
Enrolment criteria	Suspected NASH ( <i>n</i> = 884 liver clinic and <i>n</i> = 631 bariatric surgery)	Severe obesity	General population
Hepatic fat evaluation	Liver biopsy; <i>n</i> = 1515	Not available	Nuclear magnetic spectroscopy; <i>n</i> = 2736
Genetic risk score	Derived in the whole LBC	Derived in bariatric patients of LBC	Derived in <i>n</i> = 2736 of DHS
Outcomes	Histological activity and fibrosis Aminotransferases Metabolic features (Hypertension, insulin resistance Type 2 diabetes, HDL cholesterol)	Aminotransferases Metabolic features (Hypertension, insulin resistance Type 2 diabetes, HDL cholesterol)	Fibrosis score (APRI) Aminotransferases Metabolic features (Hypertension, insulin resistance Type 2 diabetes, HDL cholesterol)

**Fig. 1** Study cohorts and outcomes. NASH, nonalcoholic steatohepatitis.

**Table 1** Clinical features of the cohorts of individuals included in the study

	Liver biopsy cohort ( <i>N</i> = 1515)	Swedish Obese Subjects Study ( <i>N</i> = 3329)	Dallas Heart Study ( <i>N</i> = 4570)
Age, years	43 ± 16	48 ± 6	45 ± 11
Sex, female	722 (48)	2338 (70)	2608 (57)
Ethnicity			
European/European descent	1515 (100)	NA <sup>a</sup>	1351 (30)
African American	–	–	2355 (51)
Hispanic	–	–	743 (16)
Recruitment liver disease/morbid obesity <sup>b</sup>	884 (58)/631 (42)	0/3329 (100)	–
ALT, IU L <sup>-1</sup>	42 (23–69)	30 (22–42)	19 (14–27)
AST, IU L <sup>-1</sup>	28 (20–42)	22 (17–28)	21 (17–26)
Statin use, yes	127 (8)	–	390 (9)
Hepatic triglyceride content, %	–	–	3.5 (1.9–7.0) { <i>N</i> = 2736}
Fatty liver, yes <sup>c</sup>	1346 (90)	–	880 (32) { <i>N</i> = 2736}
Fibrosis F3–F4, yes	346 (23)	–	–
BMI, kg m <sup>-2</sup>	32.1 (27–40)	41.0 (38–44)	29.4 (25–34)
Hypertension, yes	459 (36)	2288 (69)	1496 (36)
T2D, yes	400 (26)	501 (15)	561 (12)
Fasting glucose, mg dL <sup>-1d</sup>	95 (86–110)	81 (72–94)	93 (85–102) { <i>N</i> = 4474}
Fasting insulin, IU mL <sup>-1</sup>	15 (9–21) { <i>N</i> = 1082}	17 (12–24)	13 (8–20) { <i>N</i> = 4018}
HOMA-IR, U	3.5 (2.2–5.3) { <i>N</i> = 1082}	3.5 (2.3–5.5)	3.0 (1.7–5.0) { <i>N</i> = 4018}
Total cholesterol, mg dL <sup>-1e</sup>	193 ± 46	220 ± 43	183 ± 40
HDL cholesterol, mg dL <sup>-1e</sup>	46 ± 8	50 ± 11	51 ± 15 { <i>N</i> = 3203}
Triglycerides, mg dL <sup>-1f</sup>	124 (88–177)	159 (115–221)	97 (68–145)
<i>N.</i> of risk alleles	3 (2–4)	2 (1–3)	2 (1–3)
GRS	0.53 (0.47–0.53)	0.43 (0.37–0.48)	0.13 (0.06–0.34)

Values are reported as mean ± SD, number (%), or median (IQR), as appropriate. *N*, number; NA, not assessed; BMI, body mass index; T2D, type 2 diabetes; HOMA-IR, homoeostasis model assessment-insulin resistance index; HDL, high-density lipoprotein; ALT, alanine aminotransferases; AST, aspartate aminotransferases; GRS, genetic risk score. Numbers in curly brackets indicate numbers of individuals with available data for each phenotype. <sup>a</sup>In the SOS, information about ethnicity was not available, but most participants are presumed of European descent; <sup>b</sup>Defined in the presence of BMI ≥ 40 kg m<sup>-2</sup> or ≥35 kg m<sup>-2</sup> in patients with metabolic comorbidities (type 2 diabetes, dyslipidemia or arterial hypertension). <sup>c</sup>In DHS, NAFLD (steatosis) was defined as hepatic triglyceride content (HTGC) >5.5%. <sup>d</sup>Multiply by \*0.055 <sup>e</sup>0.026 <sup>f</sup>0.011 to obtain mmol L<sup>-1</sup>.

In the LBC, steatosis, disease activity and fibrosis were assessed according to the NAFLD clinical research network criteria [25]. Briefly, steatosis

was scored (on a scale from 0 to 3) according to the percentage of affected hepatocytes (0: <5%, 1: 5–33%, 2: 34–66%, 3: 67–100%), and hepatocellular

ballooning (0–2) and lobular necroinflammation (0–3) were also recorded. Fibrosis stage was staged (from 0, no fibrosis to 4, cirrhosis) to evaluate disease progression [25]. The scoring of liver biopsies was performed by independent pathologists unaware of the clinical history and genotype of the patients [21]. The concordance between pathologists within this cohort was very good for fibrosis and good for steatosis with a coefficient of interobserver agreement for fibrosis stage and steatosis grade of 0.89 and 0.76, respectively [26].

In the DHS, we used the APRI score (AST/platelets  $\times$  100) as a noninvasive marker of fibrosis [27], because it showed the strongest association with hepatic fat compared to other indices (Table S1).

#### Genotyping

To test the causal role of hepatic fat, we selected genetic variants with validated effects on hepatic fat accumulation and/or NAFLD severity (in LBC, DHS and previous GWAS [12–14, 16, 28]) and with experimental data supporting the causality of the association, as instruments.

Therefore, the LBC and SOS cohorts were genotyped for rs738409 C>G (*PNPLA3* I148M), rs58542926 C>T (*TM6SF2* E167K), rs1260326 C>T (*GCKR* P446L) and rs641738 C>T in the *MBOAT7* locus, as previously described [17, 21]. The genotyping was performed in duplicate by TaqMan 5'-nuclease assays in the LBC and SOS (Life Technologies, Carlsbad, CA, USA). The duplicate genotype concordance rate was 100%. In the DHS, genotyping was performed using Illumina Human Exome BeadChip, as described [12]. A subset of DHS participants ( $n = 3300$ ) was previously genotyped for *LYPLAL1* rs12137855 using TaqMan assays. As the genotypes for this variant were not available in the LBC and SOS, we did not include this variant in our primary analysis but considered it as part of sensitivity analysis (see Supporting information, Data S2).

#### Evaluation of associations of NAFLD-associated variants with T2D and liver-related end-points in GWAS data

We looked up the associations with T2D of the NAFLD-associated genetic variants in publicly available databases from T2D GWAS [29, 30]. Data were downloaded from the Diagram Consortium at <http://diagram-consortium.org/downloads.html>,

and in the case of *TM6SF2* rs58542926 (E167K), extracted directly from Fuchsberger *et al.* [30]. As *GCKR* rs1260326 (P446L) has likely pleiotropic effects on glucose metabolism, we reported the effect but did not consider it as a proxy for hepatic fat when evaluating T2D risk in a Mendelian randomization context.

Two-sample Mendelian randomization analysis was performed using the 'MendelianRandomization' package in R. The per-allele associations of the *PNPLA3*, *TM6SF2* and *MBOAT7* variants with hepatic fat content in the DHS were entered as exposures, and the GWAS associations with T2D were used as outcomes. A summary estimate was calculated using the inverse variance weighted method. As described above, *GCKR* rs1260326 (P446L) was not included in this analysis due to the strong pleiotropic effects of this SNP on plasma glucose levels.

Associations with ICD-code defined liver end-points were extracted from <http://genatlas.roslin.ed.ac.uk/>, a publically available database of genetic associations in the UK Biobank ( $n = 408\,455$ ). Beta coefficients and *P*-values were extracted for associations with ICD-10 K76 ('other diseases of liver') and K74 ('fibrosis and cirrhosis of liver').

#### Statistical analysis

We used four established risk variants for hepatic fat accumulation [17, 31] as instruments in Mendelian randomization analysis. To increase power, we also created a genetic risk score (GRS) for hepatic fat accumulation composed of these variants. The association of each genetic variant with hepatic steatosis was estimated using linear regression models in all individuals with available measures of hepatic steatosis (1515 in LBC and 2736 in DHS). GRS was then calculated across all three cohorts by summing the number of steatosis predisposing alleles, weighted by their effect size (beta coefficient) on steatosis. Weights derived from severely obese individuals in the LBC were used to calculate the GRS in the SOS cohort, due to similar ethnic and clinical make-up of the participants. The GRS was strongly correlated with steatosis, explaining 7.0% and 3.5% of its variability in the LBC and DHS, respectively (Table S2). As steatosis was graded on an ordinal categorical scale in the LBC, we also performed a sensitivity analysis, using ordinal regression models to assess the relationship of genetic variants with steatosis

**Table 2** Observational association of study outcomes with histological steatosis severity in the liver biopsy cohort (LBC; n = 1515 at risk of NASH) and hepatic TG content in the Dallas Heart Study (DHS; n = 2736 from the general population)

	Liver biopsy cohort (N = 1515)			Dallas Heart Study (N = 2736)		
	$\beta$	95% c.i.	P value	$\beta$	95% c.i.	P value
ALT, IU L <sup>-1</sup>	+0.34	(0.30–0.38)	$9.0 \times 10^{-54}$	+0.25	(0.22–0.29)	$2.0 \times 10^{-40}$
AST, IU L <sup>-1</sup>	+0.29	(0.25–0.34)	$4.0 \times 10^{-35}$	+0.17	(0.13–0.21)	$6.6 \times 10^{-17}$
Necroinflammation	+0.53	(0.49–0.58)	$5.0 \times 10^{-115}$	–	–	–
Ballooning	+0.33	(0.29–0.38)	$7.1 \times 10^{-49}$	–	–	–
Fibrosis stage	+0.35	(0.30–0.39)	$2.1 \times 10^{-51}$	–	–	–
APRI score <sup>a</sup>	–	–	–	+0.09	(0.04–0.13)	$9.8 \times 10^{-5}$
Hypertension, yes	+0.13	(0.07–0.18)	$1.0 \times 10^{-5}$	+0.17	(0.06–0.27)	0.0018
T2D, yes	+0.12	(0.08–0.17)	$5.8 \times 10^{-7}$	+0.45	(0.30–0.59)	$2.6 \times 10^{-9}$
HOMA-IR	+0.25	(0.20–0.31)	$1.5 \times 10^{-22}$	+0.29	(0.26–0.33)	$1.3 \times 10^{-60}$
HDL	–0.06	(–0.11 to –0.01)	0.011	–0.15	(–0.19 to –0.12)	$5.5 \times 10^{-15}$

Adjusted standardized beta coefficients and (95% c.i.) are reported. Coefficients were adjusted for age, sex, BMI, use of statins, recruitment centre in the LBC or ethnicity in the DHS. The regression coefficients are expressed in unit of SD of feature per unit increment in histological steatosis grade (which was approximated to a continuous trait and normalized) in the LBC or a 1 SD change in hepatic triglyceride content in the DHS. For binary outcomes, the relationship was assessed using logistic regression, with beta coefficients representing the log of the odds of the outcome per a one-unit increment in steatosis grade or a 1 SD change in hepatic fat content. BMI, body mass index; T2D, type 2 diabetes; HOMA-IR, homoeostasis model assessment-insulin resistance index; HDL, high-density lipoprotein cholesterol; ALT, alanine aminotransferases; AST, aspartate aminotransferases. After correction for multiple comparisons,  $P < 0.0065$  and  $P < 0.0071$  are considered statistically significant in the LBC and DHS, respectively. <sup>a</sup>Available for  $N = 1933$  in DHS.

grade. GRS constructed based on ordinal regression models had very similar associations with liver damage and metabolic outcomes. Therefore, we decided to use the GRS estimated from linear regression models (in both cohorts) for our primary analyses.

Observational associations of hepatic steatosis with NAFLD and NAFLD-associated metabolic traits were assessed using linear and logistic regression models, with steatosis grade (LBC) or hepatic fat content (DHS) as an explanatory variable, and each trait as the outcome. The causal effect of hepatic steatosis on NAFLD and metabolic traits was estimated using the Wald (ratio) method, with standard errors approximated by the delta method [32]. All models were adjusted for age, sex, recruitment centre (in the LBC), ethnicity (in DHS), body mass index (BMI) and statin use. All estimates are standardized. The reported observational estimates are for a 1-SD unit increase in hepatic fat. The causal estimates are for a 1-SD unit increase in genetically determined hepatic fat content. We conducted sensitivity analyses, using alternative GRS, calculated by excluding one genetic variant at a time, and using an unweighted risk score.

Statistical analyses were carried out using R statistical analysis software version 3.3.2 (<http://www.R-project.org/>).  $P$ -values  $< 0.05$  were considered statistically significant.

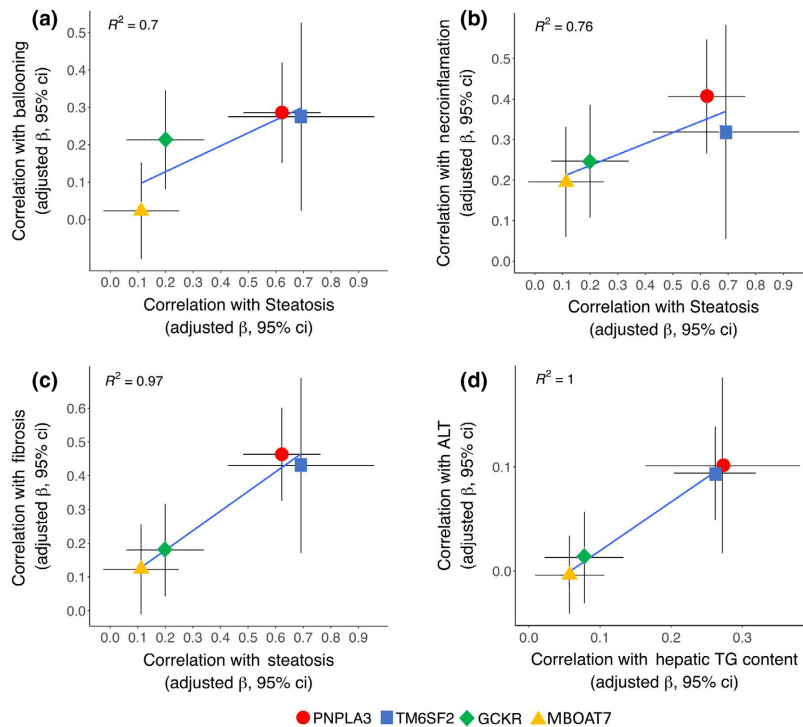
## Results

### *Observational association of hepatic fat with liver damage and metabolic traits*

In the observational analysis, hepatic fat content was strongly associated with aminotransferases ( $P < 10^{-16}$ , Table 2), in two of the study cohorts with available measures of hepatic fat (LBC and DHS). Further, it was associated with histological necroinflammation ( $P < 10^{-115}$ ), ballooning ( $P < 10^{-48}$ ) and fibrosis stage ( $P < 10^{-50}$ ) in the LBC, and with the APRI fibrosis score in the DHS ( $P = 9.8 \times 10^{-5}$ ). In both cohorts, hepatic fat was also associated with the prevalence of hypertension, T2D, increased insulin resistance (HOMA-IR) and with lower HDL (all  $P < 0.05$ ).

### *Association of genetic variants with hepatic fat, liver damage and metabolic traits*

The frequency distribution of the number of risk alleles carried by each individual in the three



**Fig. 2** Comparison of the impact of risk variants: PNPLA3 I148M (rs738409), TM6SF2 E167K (rs58542926), GCKR P446L (rs1260326) and MBOAT7 rs641738 on hepatic fat vs. liver damage. Panel a: histological steatosis vs. ballooning in the LBC ( $R^2 = 0.70$ ). b: histological steatosis vs. necroinflammation in the LBC ( $R^2 = 0.76$ ). c: histological steatosis vs. fibrosis in the LBC ( $R^2 = 0.97$ ). d: hepatic fat content vs. serum ALT levels in the DHS ( $R^2 = 1$ ). Beta coefficients and 95% confidence intervals are shown for each variant. [Correction added on 18 January 2018 after first online publication: In Figure 2B and D, the color labels for PNPLA3 and TM6SF2 have been interchanged for clarity]

cohorts is shown in Figure S2. The impact of the individual risk variants on hepatic fat, liver damage and metabolic traits is presented in Table S3. As previously reported, in the LBC and DHS, PNPLA3 rs738409 (I148M) was the strongest genetic determinant of hepatic fat, followed by TM6SF2 rs58542926 (E167K), while GCKR rs1260326 (P446L) and MBOAT7 rs641738 had smaller but significant effects. Of the four individual variants, PNPLA3 rs738409 (I148M) was also associated with higher liver enzymes in all three cohorts. All four gene variants were associated with the full spectrum of liver damage, related to NAFLD, in the LBC.

The impact of genetic risk variants on liver damage was proportional to their effect on hepatic fat accumulation (Fig. 2), consistent with the notion that their impact on liver damage is mainly mediated by increased liver fat and not by other pleiotropic effects. In a sensitivity analysis using ordinal regression models, we confirmed a close correlation between the impact of genetic variants on steatosis and that on fibrosis in the LBC (Figure S3).

The PNPLA3 rs738409 (I148M) variant was associated with increased HOMA-IR in the LBC and SOS cohorts but not in the DHS (Table S3).

#### Causal effect of hepatic fat on liver damage and metabolic traits

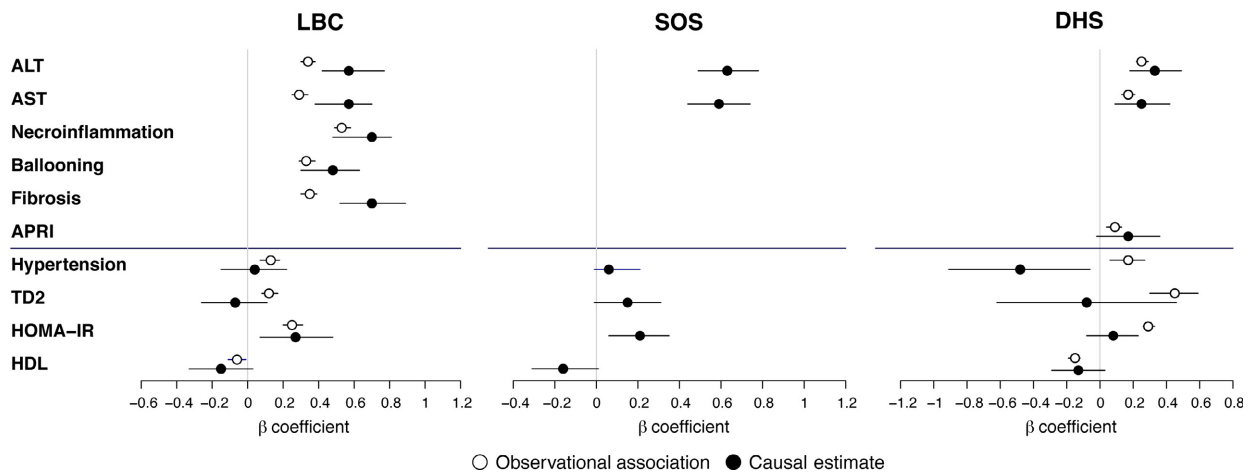
The estimated causal effects of hepatic fat on histological and metabolic traits associated with NAFLD are shown in Table 3 and in Fig. 3. The GRS was not associated with age, sex and anthropometric traits (not shown). A genetically determined increase in hepatic fat was associated with elevated aminotransferases in all three cohorts ( $P < 10^{-9}$  in the LBC,  $P < 10^{-14}$  in the SOS and  $P < 10^{-3}$  in the DHS) and with histological features of NAFLD-related liver damage in the LBC, including necroinflammation and ballooning ( $P < 10^{-16}$  and  $P < 10^{-9}$ , respectively), as well as fibrosis stage ( $P < 10^{-12}$ ). Of note, in the LBC, genetically predicted steatosis had a larger impact on fibrosis than the observed histological steatosis. Finally, the causal association between hepatic fat and fibrosis was independent of necroinflammation and hepatocellular ballooning ( $\beta = +0.37$ , 95% c.i. 0.21–0.52;  $P = 5.2 \times 10^{-6}$ ). In a sensitivity analysis that used two-stage least

**Table 3** Causal association of genetically determined hepatic fat (GRS, genetic risk score) with liver damage and clinical parameters epidemiologically associated with NAFLD in the three study cohorts

Outcome	Liver biopsy cohort (N = 1515)			Swedish Obese Subjects Study (N = 3329)			Dallas Heart Study (N = 4455) <sup>a</sup>		
	$\beta$	95% c.i.	P value	$\beta$	95% c.i.	P value	$\beta$	95% c.i.	P value
ALT	+0.57	(0.42–0.77)	$2.8 \times 10^{-11}$	+0.63	(0.490–0.78)	$1.4 \times 10^{-17}$	+0.33	(0.18–0.49)	$2.8 \times 10^{-5}$
AST	+0.57	(0.38–0.70)	$8.0 \times 10^{-10}$	+0.59	(0.44–0.74)	$1.9 \times 10^{-14}$	+0.25	(0.09–0.41)	0.0019
Necroinflammation	+0.70	(0.48–0.81)	$6.6 \times 10^{-12}$	–	–	–	–	–	–
Ballooning	+0.48	(0.30–0.63)	$2.2 \times 10^{-7}$	–	–	–	–	–	–
Fibrosis	+0.70	(0.52–0.89)	$1.3 \times 10^{-14}$	–	–	–	–	–	–
APRI score <sup>b</sup>	–	–	–	–	–	–	+0.17	(–0.01 to 0.35)	0.061
Hypertension	+0.04	(–0.15 to 0.22)	0.78	+0.06	(–0.01 to 0.21)	0.47	–0.48	(–0.91 to –0.05)	0.028
T2D	–0.07	(–0.26 to 0.11)	0.42	+0.15	(–0.01 to 0.31)	0.057	–0.08	(–0.62 to 0.46)	0.77
HOMA-IR	+0.27	(0.07–0.48)	0.0089	+0.21	(0.06–0.35)	0.006	+0.08	(–0.07 to 0.22)	0.30
HDL	–0.15	(–0.33 to 0.03)	0.10	–0.16	(–0.31 to 0.01)	0.039	–0.13	(–0.28 to 0.02)	0.083

Adjusted standardized beta coefficients and (95% c.i.) are reported. Coefficients were adjusted for age, sex, BMI, use of statins, recruitment centre in the LBC or ethnicity in the DHS. The regression coefficients are expressed in unit of SD of feature per unit increment in GRS. For binary outcomes, the relationship was assessed using logistic regression, with beta coefficients representing the log of the odds of the outcome per a one-unit increment in GRS. BMI, body mass index; T2D, type 2 diabetes; HOMA-IR, homeostasis model assessment–insulin resistance index; HDL, high-density lipoprotein; LDL, low-density lipoprotein; ALT, alanine aminotransferases; AST, aspartate aminotransferases; HF, hepatic fat, that is normalized histological steatosis of hepatic triglycerides content. After correction for multiple comparisons,  $P < 0.0065$ ,  $P < 0.008$  and  $P < 0.0071$  are considered statistically significant in the LBC, SOS and DHS, respectively. <sup>a</sup>All individuals with available genotype, phenotype and covariate data (up to  $N = 4455$ ) were included. <sup>b</sup>Available for  $N = 3133$  in DHS.





**Fig. 3** Comparison of the epidemiological association of observed hepatic fat with NAFLD features (open circles) with the causal association of hepatic fat with NAFLD-related features (filled circles) in the LBC ( $n = 1515$ , panel a), the SOS ( $n = 3329$ , panel b) and the DHS ( $n = 2736$ , panel c). T2D, type 2 diabetes; HOMA-IR, homoeostasis model assessment-insulin resistance index; HDL, high-density lipoprotein; ALT, alanine aminotransferases; AST, aspartate aminotransferases. Results were adjusted for age, sex, BMI, modality of recruitment in the LBC and ethnicity in the DHS, use of statins and (for the metabolic parameters) the severity of liver fibrosis. Estimates are beta coefficients, and error bars are 95% confidence intervals.

squares regression (rather than the ratio method) to calculate causal estimates, the results did not change substantially (Data S1 and Table S4), except that genetically determined hepatic fat was additionally associated with fibrosis (as estimated by the APRI score) in the DHS ( $P < 0.05$ ). These results suggest that hepatic fat accumulation causally promotes the full spectrum of liver damage associated with NAFLD.

In the LBC, genetically determined hepatic fat was associated with HOMA-IR, to an extent similar to that of observed steatosis, suggesting that hepatic fat causes insulin resistance, consistent with the reported epidemiological association (Table 3 and Fig. 3). The effect of steatosis on HOMA-IR became more marked after excluding patients with T2D ( $\beta = +0.41$ , 95% c.i. 0.15–0.64;  $P = 0.00078$ ), but it was attenuated after correction for fibrosis ( $\beta = +0.10$ , 95% c.i.  $-0.10$ – $0.33$ ;  $P = 0.26$ ). Similarly, in the SOS, genetically determined hepatic fat was causally associated with HOMA-IR ( $\beta = +0.21$ ; 95% c.i. 0.06–0.35; Table 3 and Fig. 3), but the association was lost after correction for serum ALT, used as a measure of liver damage ( $\beta = +0.10$ ; 95% c.i.  $-0.05$ – $0.24$ ;  $P = 0.20$ ). We did not detect a significant effect of genetically determined hepatic fat on insulin resistance in the population-based DHS (Table 3).

To validate the causal association between hepatic fat and insulin resistance, we extracted results from the largest publicly available GWAS on T2D (Table 4). The risk alleles of the two strongest steatogenic variants, *PNPLA3* rs738409 (I148M) and *TM6SF2* rs58542926 (E167K), were associated with a small increase in the risk of T2D (allelic ORs 1.04, 95% c.i. 1.01–1.07 and 1.14, 95% c.i. 1.10–1.19, respectively). The association of the *MBOAT7* variant was directionally concordant but not significant (OR 1.02, 95% c.i. 0.99–1.05;  $P = 0.19$ ). The major (P) allele of *GCKR* rs1260326 P446L (which reduces the risk of NAFLD) was associated with a modestly increased risk of T2D (OR 1.06, 95% c.i. 1.03–1.09). However, *GCKR* rs1260326 P446L is known to associate strongly with plasma glucose levels, a pleiotropic association that makes it problematic to use this variant as a proxy for hepatic fat in a Mendelian randomization context with T2D as an outcome.

In two-sample Mendelian randomization analysis using the *PNPLA3*, *TM6SF2* and *MBOAT7* variants as instruments for hepatic fat, the causal OR for risk of T2D for a 1 standardized unit increase in genetically determined hepatic fat was 1.31 (95% c.i., 1.20–1.43;  $P = 1.2 \times 10^{-9}$ ).

**Table 4** Association of NAFLD-variants with type 2 diabetes in published GWAS

Gene	SNP	Effect allele	other		95% c.i.	P value	N	Data source
			allele	T2D OR				
<i>PNPLA3</i>	rs738409	G (minor)	C	1.04	1.01–1.07	0.0045	100 323	diagram.mega-meta
<i>TM6SF2</i>	rs58542926	T (minor)	C	1.14	1.10–1.19	$3.2 \times 10^{-10}$	92 794	Fuchsberger 2016 [30]
<i>MBOAT7</i>	rs641738	T (minor)	C	1.02	0.99–1.05	0.19	76 306	diagram.mega-meta
<i>GCKR</i>	rs1260326	C (major) <sup>a</sup>	T	1.06	1.03–1.09	$3.4 \times 10^{-5}$	100 584	diagram.mega-meta

Data were downloaded from <http://diagram-consortium.org/downloads.html> or extracted from Fuchsberger *et al.* [30].

<sup>a</sup>The effect allele in *GCKR* is the major allele, encoding *GCKR* 446P. The 446P-allele causes a relative gain-of-function compared to the L-allele, leading to less hepatic phosphorylation of glucose, increased blood glucose and thus an increased risk of type 2 diabetes (T2D). Due to these strong pleiotropic effects of *GCKR* on glucose metabolism, it is problematic to use this variant as a proxy for NAFLD in Mendelian randomization analyses with T2D as an outcome.

### Sensitivity analyses

To rule out the possibility that the association of the GRS with NAFLD-associated traits was biased by the specific mechanism of action of single genetic risk variants, we conducted sensitivity analyses using alternative GRS, calculated by excluding a single risk variant at a time (Table S2). The results evaluating the association of alternative GRS with NAFLD features are shown in Table S5.

All GRS were associated with hepatic fat, although, as expected, those excluding *PNPLA3* rs738409 (I148M) were less robustly associated than the other scores, across all cohorts. The associations between alternative GRS and NAFLD-associated traits were generally consistent with those for the full score, based on four variants. Importantly, most alternative models confirmed a causal association of hepatic fat with liver enzymes and histological liver damage. The only exception was that the associations with aminotransferases in the DHS were no longer statistically significant when excluding the *PNPLA3* variant, likely due to the lack of power (Table S5). In the LBC, we also confirmed a trend for association of the alternative GRS with increased insulin resistance.

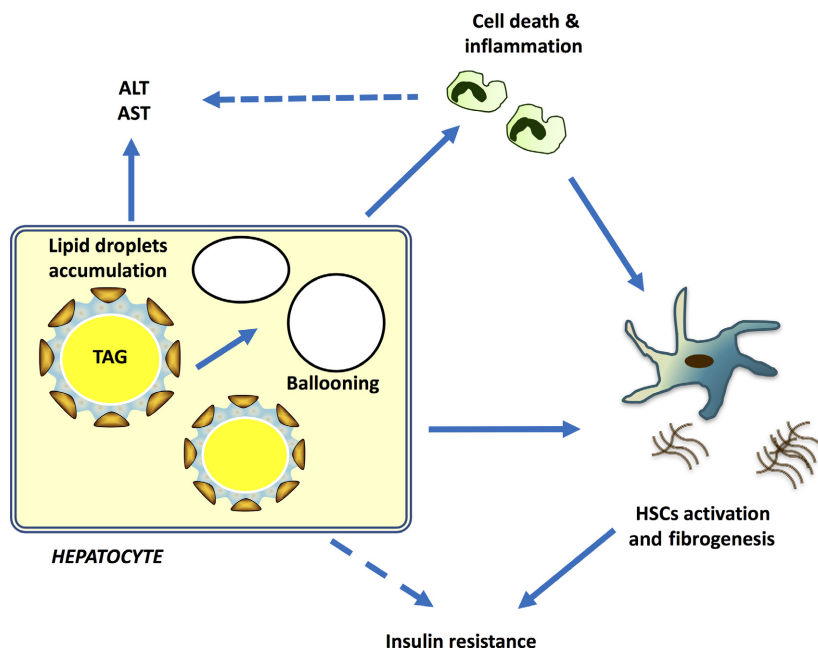
To confirm the association of the steatogenic variants with liver disease in a separate cohort, we performed a lookup in a publically available database of genetic associations in 408 455 participants from UK Biobank (as reported in the Methods and in Supporting information, Data S1). The steatogenic alleles in *PNPLA3*, *TM6SF2*, *GCKR* and *MBOAT7* all associated with increased risk of ICD-defined 'other diseases of liver' (K76), which

includes NAFLD ( $n = 408\ 455$ ,  $P = 0.0075$ – $2 \times 10^{-12}$ , Table S6 and Figure S4). Variants in *PNPLA3*, *TM6SF2* and *MBOAT7* associated with increased risk of 'fibrosis and cirrhosis of liver' (K74;  $P = 0.002$ – $3 \times 10^{-6}$ , Table S6). In two-sample Mendelian randomization analysis, a standardized 1-unit increase in genetically determined hepatic fat content was associated with an increased risk of 'other diseases of liver' ( $P = 5 \times 10^{-19}$ ), and with 'fibrosis and cirrhosis of liver' ( $P = 9 \times 10^{-11}$ ).

Additional sensitivity analyses, evaluating an alternative GRS including the *LYPLAL1* rs12137855 variant, using beta coefficients derived from external cohorts, and showing the results in patients stratified according to modality of recruitment and ethnicity are reported in Supporting information.

### Discussion

The current study represents the first attempt to apply a formal Mendelian randomization framework to test whether hepatic fat is causally related to liver damage and fibrosis. The main findings of this work are as follows: (i) the magnitude of association of genetic risk variants with fibrosis severity, the major prognostic determinant in NAFLD, is proportional to their effect on steatosis, suggesting that the association between these genetic variants and liver disease is explained by hepatic fat accumulation; (ii) the association between hepatic fat accumulation and fibrosis was independent of inflammation, suggesting that it is not influenced by pleiotropic effects of genetic variants on inflammation; (iii) the effect size of genetically predicted steatosis on fibrosis was



**Fig. 4** Schematic representation of main study findings. Hepatic fat accumulation is causally associated with increased liver enzymes, hepatocellular damage, necroinflammation and fibrosis. TAG, triglycerides, AST, aspartate aminotransferases, ALT, alanine aminotransferases, HSCs, hepatic stellate cells.

consistent with that of observational steatosis, suggesting that long-term exposure to steatosis causes fibrosis independently of confounders; (iv) the association of genetically determined hepatic fat with insulin resistance was restricted to high-risk individuals, suggesting that insulin resistance is not a direct consequence of hepatic fat accumulation but a phenomenon possibly mediated by liver damage. A schematic representation of these study findings is presented in Fig. 4.

Although the associations of the four gene variants included in our risk score with NAFLD progression have been previously reported [17, 18], whether these effects are consistent with a causal role of hepatic fat or are explained by other pleiotropic effects of the genetic variants has not been examined so far. Here, we observed a clear dose-response relationship between the genetic effects on hepatic fat content and liver damage, in particular the severity of fibrosis, the major prognostic determinant in patients with NAFLD [33]. Furthermore, genetically determined liver fat was strongly associated with liver fibrosis and cirrhosis in the UK Biobank database. These data suggest that the impact of these variants on liver damage predisposition is mainly determined by hepatic fat accumulation.

To increase statistical power, we constructed a GRS from the four risk variants. The GRS was

strongly associated with hepatic fat content and with aminotransferase levels, as well as with the entire spectrum of histological liver damage. Interestingly, the estimated causal effect of hepatic steatosis was larger than the observational association of histological steatosis with fibrosis. This likely reflects the lifelong exposure to steatosis in carriers of steatogenic variants [11] and suggests that the histological grade of steatosis at the time of biopsy may underestimate the full impact of hepatic fat accumulation on fibrosis. In addition, the causal effect of hepatic fat on fibrosis was partly independent of disease activity at the time of biopsy, supporting the active role of hepatic fat accumulation in inducing progressive liver disease over the entire lifetime. These data are consistent with experimental evidence linking hepatic fat accumulation and insulin resistance with fibrosis in NAFLD, independent of inflammatory pathways [34].

Hepatic steatosis has been hypothesized to represent an ‘innocent bystander’ in the pathogenesis of NASH, or even a protective response to increased free fatty acids availability [35]. In contrast to these hypotheses, rare mutations predisposing to severe hepatic fat accumulation frequently lead to cirrhosis and hepatocellular carcinoma even in the absence of other hepatotoxic insults [36]. Furthermore, a recent population-based twin study found that liver steatosis and fibrosis tend to be

co-inherited [37]. The present results suggest that hepatic steatosis promotes the full spectrum of liver disease. Potential mechanisms include reduced ability to incorporate fatty acids in lipid droplets under stress conditions, spillover or peroxidation of fatty acids and activation of lipid-droplet dependent signalling pathways [38]. However, these results do not necessarily imply that triglyceride accumulation are the driver of liver disease progression, because NAFLD risk variant are associated with hepatic accumulation of different lipid species [39]. Therefore, experimental studies are still required to investigate the mechanism underlying this association.

Finally, we found that hepatic steatosis is causally associated with insulin resistance in individuals at risk of NASH because of severe liver disease or obesity, in a liver damage dependent fashion. The association between genetically determined hepatic fat and insulin resistance was not observed in the DHS, possibly due to the low prevalence of individuals with advanced liver fibrosis in this general population. Alternatively, the lack of association may be a result of reduced power, owing to the fact that the impact of the genetic risk variants for NAFLD on liver damage was more marked in individuals with acquired risk factors such as obesity [40]. The steatogenic variants were associated with an increased T2D risk in the largest T2D GWAS [30] and in a recently published study including >300 000 individuals with metabolic characterization [41]. In our study, the association between a genetically increased liver fat content and insulin resistance appeared to depend on the presence of liver damage. This might reflect decreased hepatic insulin signalling induced by accumulation of specific lipid species or a fibrosis-mediated decrease in hepatic insulin clearance [42, 43]. Taken together, these data support the hypothesis that inhibition of hepatic fat accumulation might lead to improvement of insulin resistance and a reduction in risk of T2D, secondary to the improvement of liver disease.

There are limitations to our study. We evaluated a small number of variants, but given the high heritability of NAFLD [31], the GRS explained a larger fraction of phenotypic variability than most scores based on a large number of variants in similar studies conducted for other complex traits [32]. An underlying assumption of the Mendelian randomization method is that genetic variants used as proxies for exposure influence the outcome

solely via their effect on the exposure and not via pleiotropic effects. *TM6SF2* rs58542926 (E167K) impairs hepatic secretion of lipoproteins and reduces plasma levels of triglycerides and cholesterol. However, reduced levels of lipoproteins are not likely to directly cause liver damage. *GCKR* rs1260326 (P446L) increases the trapping of glucose in the liver and has an impact on plasma levels of glucose and insulin. This may pose a problem when using the variant as a proxy for hepatic fat with T2D as an outcome. *MBOAT7* rs641738 is associated with changes in circulating levels of phosphatidylinositols [17], which are unlikely to influence liver damage. Even though we cannot rule out a possibility that *PNPLA3* rs738409 (I148M) also promotes liver damage by alternate mechanisms, for example by altering retinol metabolism [44], our results held consistent when excluding the *PNPLA3* rs738409 (I148M) from the GRS. In summary, pleiotropic effects are unlikely to explain the main findings of our study, even if pleiotropy remains an unavoidable limitation of the Mendelian randomization approach [11, 45].

To reduce bias, we performed several sensitivity analyses, including the evaluation of alternative GRS, and validated the results in three independent cohorts. Steatosis grade (a categorical phenotype) was approximated to a continuous trait in the LBC. However, using steatosis grade as an ordinal categorical trait did not change the results. Finally, direct measurement of hepatic fat content was not available in the SOS, and causes of fatty liver other than NAFLD could not be ruled out in the DHS and SOS.

## Conclusion

Genetic variants that increase hepatic fat content are associated with an increase in biochemical markers of liver damage and the risk of hepatic fibrosis, to the degree predicted by their steatogenic effects. This supports the hypothesis that hepatic steatosis per se is likely to be a causal risk factor for the development of liver fibrosis, independent of inflammation. In addition, genetic variants that increase hepatic fat content were associated with modest increases in insulin resistance and risk of T2D. However, these associations were only observed in individuals with liver disease, suggesting that liver disease, rather than hepatic steatosis, could be the underlying causal factor. Taken together, these data suggest that

interventions aimed at reducing hepatic steatosis are likely to have long-term beneficial effects on liver disease and potentially on insulin resistance in patients with NAFLD.

#### Conflict of interest statement

Authors declare that they do not have any conflict of interest relevant to this manuscript. SR has been consulting for Chiesi Farmaceutici Group, Amgen, Sanofi, Novonordisk, Akcea therapeutics, Genzyme, AstraZeneca and Aegerion in the last 5 years.

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#### References

- Browning JD, Szczepaniak LS, Dobbins R, *et al.* Prevalence of hepatic steatosis in an urban population in the United States: impact of ethnicity. *Hepatology* 2004; **40**: 1387–95.
- Non-alcoholic Fatty Liver Disease Study G, Lonardo A, Bellentani S, *et al.* Epidemiological modifiers of non-alcoholic fatty liver disease: Focus on high-risk groups. *Dig Liver Dis* 2015; **47**: 997–1006.
- Valenti L, Bugianesi E, Pajvani U, Targher G. Nonalcoholic fatty liver disease: cause or consequence of type 2 diabetes? *Liver Int* 2016; **36**: 1563–79.
- Dyson J, Jaques B, Chattopadhyay D, *et al.* Hepatocellular cancer: The impact of obesity, type 2 diabetes and a multi-disciplinary team. *J Hepatol* 2014; **60**: 110–7.
- Charlton MR, Burns JM, Pedersen RA, Watt KD, Heimbach JK, Dierkhising RA. Frequency and outcomes of liver transplantation for nonalcoholic steatohepatitis in the United States. *Gastroenterology* 2011; **141**: 1249–53.
- van der Poorten D, Samer CF, Ramezani-Moghadam M, *et al.* Hepatic fat loss in advanced nonalcoholic steatohepatitis: are alterations in serum adiponectin the cause? *Hepatology* 2013; **57**: 2180–8.
- Bril F, Barb D, Portillo-Sanchez P, *et al.* Metabolic and histological implications of intrahepatic triglyceride content in nonalcoholic fatty liver disease. *Hepatology* 2017; **65**: 1132–44.
- Romero-Gomez M, Zelber-Sagi S, Trenell M. Treatment of NAFLD with diet, physical activity and exercise. *J Hepatol* 2017; **67**: 829–46.
- Lawlor DA, Harbord RM, Sterne JA, Timpson N, Davey Smith G. Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat Med* 2008; **27**: 1133–63.
- Davey Smith G, Hemani G. Mendelian randomization: genetic anchors for causal inference in epidemiological studies. *Hum Mol Genet* 2014; **23**: R89–98.
- Holmes MV, Ala-Korpela M, Smith GD. Mendelian randomization in cardiometabolic disease: challenges in evaluating causality. *Nat Rev Cardiol* 2017; **14**: 577–90.
- Romeo S, Kozlitina J, Xing C, *et al.* Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease. *Nat Genet* 2008; **40**: 1461–5.
- Speliotes EK, Yerges-Armstrong LM, Wu J, *et al.* Genome-wide association analysis identifies variants associated with nonalcoholic fatty liver disease that have distinct effects on metabolic traits. *PLoS Genet* 2011; **7**: e1001324.
- Kozlitina J, Smagris E, Stender S, *et al.* Exome-wide association study identifies a TM6SF2 variant that confers susceptibility to nonalcoholic fatty liver disease. *Nat Genet* 2014; **46**: 352–6.
- Holmen OL, Zhang H, Fan Y, *et al.* Systematic evaluation of coding variation identifies a candidate causal variant in TM6SF2 influencing total cholesterol and myocardial infarction risk. *Nat Genet* 2014; **46**: 345–51.
- Buch S, Stickel F, Trepo E, *et al.* A genome-wide association study confirms PNPLA3 and identifies TM6SF2 and MBOAT7 as risk loci for alcohol-related cirrhosis. *Nat Genet* 2015; **47**: 1443–8.
- Mancina RM, Dongiovanni P, Petta S, *et al.* The MBOAT7-TMC4 Variant rs641738 Increases Risk of Nonalcoholic Fatty Liver Disease in Individuals of European Descent. *Gastroenterology* 2016; **150**: 1219–30. e6
- Dongiovanni P, Romeo S, Valenti L. Genetic factors in the pathogenesis of nonalcoholic fatty liver and steatohepatitis. *Biomed Res Int* 2015; **2015**: 460190.
- Yki-Jarvinen H. Non-alcoholic fatty liver disease as a cause and a consequence of metabolic syndrome. *Lancet Diabetes Endocrinol* 2014; **2**: 901–10.
- Kim DS, Jackson AU, Li YK, *et al.* Novel association of TM6SF2 rs58542926 genotype with increased serum tyrosine

- levels and decreased apolipoprotein B-100 particles in Finns. *J Lipid Res* 2017; **58**: 1471–81.
- 21 Dongiovanni P, Petta S, Maglio C, *et al.* Transmembrane 6 superfamily member 2 gene variant disentangles nonalcoholic steatohepatitis from cardiovascular disease. *Hepatology* 2015; **61**: 506–14.
  - 22 Carlsson LM, Peltonen M, Ahlin S, *et al.* Bariatric surgery and prevention of type 2 diabetes in Swedish obese subjects. *N Engl J Med* 2012; **367**: 695–704.
  - 23 Victor RG, Haley RW, Willett DL, *et al.* The Dallas Heart Study: a population-based probability sample for the multi-disciplinary study of ethnic differences in cardiovascular health. *Am J Cardiol* 2004; **93**: 1473–80.
  - 24 Bonora E, Targher G, Alberiche M, *et al.* Homeostasis model assessment closely mirrors the glucose clamp technique in the assessment of insulin sensitivity: studies in subjects with various degrees of glucose tolerance and insulin sensitivity. *Diabetes Care* 2000; **23**: 57–63.
  - 25 Kleiner DE, Brunt EM, Van Natta M, *et al.* Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005; **41**: 1313–21.
  - 26 Petta S, Valenti L, Marra F, *et al.* MERTK rs4374383 polymorphism affects the severity of fibrosis in non-alcoholic fatty liver disease. *J Hepatol* 2016; **64**: 682–90.
  - 27 Maida M, Macaluso FS, Salomone F, Petta S. Non-invasive assessment of liver injury in non-alcoholic fatty liver disease: a review of literature. *Curr Mol Med* 2016; **16**: 721–37.
  - 28 Kitamoto T, Kitamoto A, Yoneda M, *et al.* Genome-wide scan revealed that polymorphisms in the PNPLA3, SAMM50, and PARVB genes are associated with development and progression of nonalcoholic fatty liver disease in Japan. *Hum Genet* 2013; **132**: 783–92.
  - 29 Manning AK, Hivert MF, Scott RA, *et al.* A genome-wide approach accounting for body mass index identifies genetic variants influencing fasting glycemic traits and insulin resistance. *Nat Genet* 2012; **44**: 659–69.
  - 30 Fuchsberger C, Flannick J, Teslovich TM, *et al.* The genetic architecture of type 2 diabetes. *Nature* 2016; **536**: 41–7.
  - 31 Dongiovanni P, Valenti L. Genetics of nonalcoholic fatty liver disease. *Metabolism* 2016; **65**: 1026–37.
  - 32 Burgess S, Thompson SG. Use of allele scores as instrumental variables for Mendelian randomization. *Int J Epidemiol* 2013; **42**: 1134–44.
  - 33 Angulo P, Kleiner DE, Dam-Larsen S, *et al.* Liver fibrosis, but no other histologic features, is associated with long-term outcomes of patients with nonalcoholic fatty liver disease. *Gastroenterology* 2015; **149**: 389–97. e10.
  - 34 Dongiovanni P, Meroni M, Baselli GA, *et al.* Insulin resistance promotes Lysyl Oxidase Like 2 induction and fibrosis accumulation in non-alcoholic fatty liver disease. *Clin Sci* 2017; **131**: 1301–15.
  - 35 Yamaguchi K, Yang L, McCall S, *et al.* Inhibiting triglyceride synthesis improves hepatic steatosis but exacerbates liver damage and fibrosis in obese mice with nonalcoholic steatohepatitis. *Hepatology* 2007; **45**: 1366–74.
  - 36 Di Filippo M, Moulin P, Roy P, *et al.* Homozygous MTTP and APOB mutations may lead to hepatic steatosis and fibrosis despite metabolic differences in congenital hypocholesterolemia. *J Hepatol* 2014; **61**: 891–902.
  - 37 Cui J, Chen CH, Lo MT, *et al.* Shared genetic effects between hepatic steatosis and fibrosis: A prospective twin study. *Hepatology* 2016; **64**: 1547–58.
  - 38 Mashek DG, Khan SA, Sathyanarayan A, Ploeger JM, Franklin MP. Hepatic lipid droplet biology: Getting to the root of fatty liver. *Hepatology* 2015; **62**: 964–7.
  - 39 Luukkonen PK, Zhou Y, Nidhina Haridas PA, *et al.* Impaired hepatic lipid synthesis from polyunsaturated fatty acids in TM6SF2 E167K variant carriers with NAFLD. *J Hepatol* 2017; **67**: 128–36.
  - 40 Stender S, Kozlitina J, Nordestgaard BG, Tybjaerg-Hansen A, Hobbs HH, Cohen JC. Adiposity amplifies the genetic risk of fatty liver disease conferred by multiple loci. *Nat Genet* 2017; **49**: 842–7.
  - 41 Liu DJ, Peloso GM, Yu H, *et al.* Exome-wide association study of plasma lipids in >300,000 individuals. *Nat Genet* 2017; **49**: 1758–66.
  - 42 Seppala-Lindroos A, Vehkavaara S, Hakkinen AM, *et al.* Fat accumulation in the liver is associated with defects in insulin suppression of glucose production and serum free fatty acids independent of obesity in normal men. *J Clin Endocrinol Metab* 2002; **87**: 3023–8.
  - 43 Kumashiro N, Erion DM, Zhang D, *et al.* Cellular mechanism of insulin resistance in nonalcoholic fatty liver disease. *Proc Natl Acad Sci USA* 2011; **108**: 16381–5.
  - 44 Pirazzi C, Valenti L, Motta BM, *et al.* PNPLA3 has retinyl-palmitate lipase activity in human hepatic stellate cells. *Hum Mol Genet* 2014; **23**: 4077–85.
  - 45 Boyle EA, Li YI, Pritchard JK. An expanded view of complex traits: from polygenic to omnigenic. *Cell* 2017; **169**: 1177–86.
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### Supporting Information

Additional Supporting Information may be found in the online version of this article:

**Data S1.** Supplementary methods.

**Data S2.** Supplementary results.

**Table S1.** Noninvasive fibrosis scores tested in the DHS for association with hepatic fat content.

**Table S2.** Coefficient used to develop genetic risk scores (GRS) applied in the study cohorts (full models and sensitivity analysis conducted by removing one genetic variant at time).

**Table S3.** Association of genetic variants included in the GRS (*PNPLA3* rs738409 I148M, *TM6SF2* rs58542926 E167K, *GCKR* rs1260326 P446L, and *MBOAT7* rs641738) with liver damage and clinical parameters epidemiologically associated with NAFLD, in the LBC ( $N = 1515$ ), SOS ( $N = 3329$ ), and DHS ( $N = 4570$ ). Estimates of  $\beta_{\text{variant-feature}}$  are reported for adjusted models.

**Table S4.** Causal effect estimates of hepatic fat content, as evaluated by instrumental regression analysis by the 2SLS method, on metabolic and hepatic correlates of NAFLD in the LBC ( $n = 1515$  at risk of NASH) and the DHS ( $n = 2736$  from the general population).

**Table S5.** Sensitivity analyses of IV regression analysis evaluating the causal role of hepatic fat on the hepatic and metabolic NAFLD features in the study cohorts with alternative GRS excluding one genetic risk variant per time. Estimate of causality -  $\beta$  coefficient - (95% confidence intervals) and  $P$  values are reported for adjusted models.

**Table S6.** Associations of steatogenic variants with ICD-code defined liver endpoints in 408 455 participants from UKBiobank.

**Table S7.** Weights used in GRS construction in DHS.

**Table S8.** Impact of the *LYPLAL1* rs12137855 variant on hepatic and metabolic features of NAFLD in the DHS. Estimate of  $\beta_{\text{variant-feature}}$  are reported for adjusted models.

**Table S9.** Association of genetically determined hepatic fat with liver damage and metabolic features, using alternative GRS-S1 and GRS-S2.

**Table S10.** Sensitivity analysis: causal effect estimates of hepatic fat content on hepatic and metabolic correlates of NAFLD in the LBC, stratified by modality of recruitment (Hepatology clinics vs. Bariatric surgery centers) in the LBC.

**Table S11.** Causal effect estimates of hepatic fat content on hepatic and metabolic correlates of NAFLD in the DHS, stratified by ethnicity.

**Figure S1.** Schematic representation of the principles of Mendelian randomization.

**Figure S2.** Frequency distribution of the number of risk variants for hepatic fat accumulation in *PNPLA3*, *TM6SF2*, *GCKR*, and *MBOAT7* carried by each individual in the LBC, SOS, and in the DHS.

**Figure S3.** Comparison of the impact of the evaluated common risk variants in *PNPLA3*, *TM6SF2*, *GCKR*, and *MBOAT7* on steatosis grade vs. fibrosis stage in the LBC by ordinal regression analysis.

**Figure S4.** Comparison of the impact of the evaluated common risk variants in *PNPLA3*, *TM6SF2*, *GCKR*, and *MBOAT7* on hepatic fat content vs. other liver diseases (ICD9-K76, including NAFLD) in the UKBiobank ( $n = 408\ 455$ ). Beta coefficients and 95% confidence intervals are shown. ■