

1 **Subaerial biofilms on outdoor stone monuments: changing the perspective towards an**
2 **ecological framework**

3

4 Federica Villa, Philip S. Stewart, Isaac Klapper, Judith M. Jacob, Francesca Cappitelli

5

6 Dr Federica Villa* (corresponding author), double affiliation. Dipartimento di Scienze per gli
7 Alimenti, la Nutrizione e l'Ambiente, Università degli Studi di Milano, via Celoria 2, 20133 Milano,
8 Italy. Center for Biofilm Engineering, Montana State University, 366 EPS Building P.O. Box 173980,
9 Bozeman, MT 59717, USA. E-mail: federica.villa@unimi.it; Phone +39 02 50319121.

10

11 Professor Philip S. Stewart, Center for Biofilm Engineering, Montana State University, 366 EPS
12 Building P.O. Box 173980, Bozeman, MT 59717, USA. E-mail: phil_s@biofilm.montana.edu; Phone
13 +14 406 994-4770.

14

15 Professor Isaac Klapper, Department of Mathematics, Temple University (038-16) 1805 N Broad
16 Street, Philadelphia, PA 19122-6094. E-mail: tue87237@temple.edu; Phone +14 215 215-204-
17 8419.

18

19 Ms. Judith M. Jacob, National Park Service, Northeast Region, Historic Architecture, Conservation,
20 and Engineering Program; New York, NY, USA. E-mail: judy_jacob@nps.gov; Phone +14 212 825-
21 6877.

22

23 Professor Francesca Cappitelli, Dipartimento di Scienze per gli Alimenti, la Nutrizione e l'Ambiente,

24 Università degli Studi di Milano, via Celoria 2, 20133 Milano, Italy. E-mail:

25 francesca.cappitelli@unimi.it; Phone +39 02 5031912.

26

27

28 **Abstract**

29 Despite the appreciation of the role played by outdoor stone heritage in societal well-being and
30 sustainable urban development, research efforts have not been completely successful in tackling
31 the complex issues related to its conservation. One of the main problems is that we are
32 continuously underestimating the role and behavior of microorganisms in form of biofilm
33 (subaerial biofilms, SABs) to the management of stone artifacts. To this end, we discuss the
34 necessity to approach the topic from an ecological perspective, through an overview of the
35 characteristics of SABs that mediate different ecological interactions. Furthermore, we explore the
36 application of functional traits ecology to unravel the mechanisms by which SABs might respond to
37 a changing environment. Finally we guide and prioritize further research, in order to inform policy-
38 makers, and to develop management strategies for protection prior to, or following after, active
39 conservation treatment.

40 **Introduction**

41 Preserving the environment for future generations is one of the key concepts of sustainability,
42 which is grounded in the need for intergenerational equity. The ongoing political and scientific
43 debate on sustainability tends to focus on issues related to carbon emission, energy consumption,
44 natural resource use and waste management, or the economic aspects of urban regeneration and
45 growth (Tweed and Sutherland 2007). Increasingly, however, national governments and
46 international institutions recognize cultural heritage as a non-renewable resource that is unique,
47 non-replaceable or non-interchangeable, highlighting the intrinsic value of cultural heritage in
48 contributing to the societal and economic well-being of communities (*inter alia* MEA 2005, EU
49 Communication 2010, UNESCO 2013).

50 Thus, conservation and management of cultural heritage constitutes a strategic choice for the 21st
51 century.

52 This fundamental principle has been recently recognized in the outcome document of the United
53 Nations Conference on Sustainable Development - or Rio+20 - The Future We Want- (Brazil on 20-
54 22 June 2012), by highlighting how *'many people, especially the poor, depend directly on*
55 *ecosystems for their livelihoods, their economic, social and physical well-being, and their cultural*
56 *heritage'* (emphasis added), or by calling for the *'conservation as appropriate of the natural and*
57 *cultural heritage of human settlements, the revitalization of historic districts, and the rehabilitation*
58 *of city centres.'*

59 The fundamental roles played by cultural heritage are threatened today by a number of factors,
60 including climate change and microbial attack, leading to new challenges for heritage objects,
61 especially those exposed to the outdoor environment. Preserving the fragile character of our
62 cultural heritage, and managing it for the benefit of current and future generations is a major task
63 for researchers and decision-makers worldwide.

64 Many of the world's most precious artworks are made of stone (e.g. marble, limestone and
65 sandstone) with a finite life, and they are slowly but irreversibly disappearing (Schreerer *et al.*
66 2009).

67 Despite the appreciation of the role played by stone heritage in many societies, research efforts
68 have not been completely successful in tackling the complex issues related to its conservation, and
69 the need to develop comprehensive approaches and methodologies for its management. One of
70 the main gaps is that we are still understanding the contribution of microorganisms to the
71 deterioration of stone, as for many decades chemical and physical deterioration was believed the
72 main cause of material decay (Sterflinger and Piñar 2013). Stone monuments, apart from being
73 ancient records that illuminate the cultural history of our planet, are dynamic repositories that
74 support microbial life. The presence of green, yellow-brown or black patinas is all too familiar to
75 anyone who has looked closely at a historic stone building or sculpture. These patinas are
76 composed by densely packed microorganisms that operate within self-organized structures of
77 micron to- millimeter scales (**Figure 1**). These microbial communities at the stone/air interface are
78 called subaerial biofilms (SABs). SABs are made up of many microbial cells, generally of different
79 types, which employ coordinated survival strategies to increase biocide resistance and microbial
80 fitness, and to avoid loss of energy and nutrients (Stewart and Franklin 2008, Stone 2015). SABs
81 can be viewed as multi-component open ecosystems sensitively tuned to the atmosphere and the
82 stone substratum (Gorbushina 2007). As with any other ecosystem, understanding of the
83 ecological and evolutionary mechanisms by which SABs organize themselves and respond to
84 environmental changes will help to predict, and possibly ameliorate, system performance and
85 their response to perturbations, improving the development of comprehensive approaches for the
86 sustainable management of outdoor stone heritage in a changing environment.

87 Thus, in order to obtain a holistic view of the phenomena occurring at the stone surface, we
88 should consider relationships between the biotope (stone), the biocenosis (SABs) and the
89 surrounding environment (macro- and micro-climate).

90 The main goal of this paper is to argue for new lines of research in which SABs inhabiting stone
91 monuments are viewed from an ecological perspective, and moving toward a system-level
92 understanding of biofilm community organization and function. Conversely, SABs on stone
93 monuments could act as interesting models for ecological study, offering exciting new
94 opportunities for the development and testing of ecological principles, broadening understanding
95 of microbial ecosystems and generating new insights in basic ecology.

96 This review is organized to provide the reader with: (1) an overview of what is known of ecology of
97 SABs inhabiting stone monuments and the gaps in the literature, suggesting an objective
98 framework for the factors that influence the structure and function of the microbial communities
99 inhabiting stone surfaces; (2) the application of functional traits ecology to unravel the
100 mechanisms by which SABs might respond to a changing environment; and (3) a summary of the
101 salient points of the presented review and identification of the highest priority research areas for
102 targeted research.

103

104 **Ecology of SABs inhabiting outdoor stone materials**

105 SABs and their inhabitants are shaped by the complex dipartite interactions between the
106 atmosphere and the stone. The stone substratum acts as a putative source of minerals, while the
107 air chemistry might offer inorganic and organic compounds (Villa *et al.* 2015). Furthermore,
108 surface irregularities such as fissures, cracks and pores, provide microorganisms safe places
109 against harsh environmental conditions. There, microorganisms take advantage of the
110 accumulated moisture, as well as of the shelter from intense solar radiation, temperature

111 fluctuations, wind and desiccation, and therefore they successfully colonize the lithic material
112 (Gorbushina *et al.* 2002). Not surprising is the occurrence of endolithic communities, including
113 photosynthetic communities, inside micro-cracks and pores of stone monuments (Crispim and
114 Gaylarde 2005).

115 Microbial growth on stone surfaces follows the complex topography of the substrate and
116 generates a patchy biofilm that spreads between the mineral grains filling depressions, fissures
117 and intergranular spaces (Gorbushina 2007). However, SABs do not simply cover the lithic surface,
118 but rather they interact with the stone in myriad ways, revealing a tight and clearly defined
119 coupling between geochemical and biological processes that affect the lithic substrate in different
120 ways (**Table 1**). These properties translate into a characteristic set of ecological impacts, making
121 SABs effective ecosystem engineers by their substantial effects on the physical and chemical
122 properties of the habitat in which they live.

123 Taxonomic and phylogenetic studies of SABs have revealed lower diversity in SABs on stone
124 surfaces compared to most natural systems (Gorbushina and Broughton 2009). The relatively low
125 diversity is attributed to the extreme and fluctuating environmental conditions that
126 microorganisms must endure. In fact, outdoor stone monuments are often stressful environments
127 characterized by desiccation, low nutrient concentrations, large temperature variations and high
128 exposure to wind, UV radiation and physical damage (Viles and Cutler 2012). Only microorganisms
129 with a very broad range of tolerance to multiple and fluctuating stresses can establish themselves
130 under these conditions (Zakharova *et al.* 2013).

131 However, despite the relatively low genetic diversity, SABs contain metabolically interactive, self-
132 sustaining microbial communities, which promote cooperative interactions within the biofilm
133 (Villa *et al.* 2015). An over-riding characteristic of SAB communities is that together, constituent
134 microorganisms overcome environmental stresses better than any could individually.

135 This joint protection is rooted in the presence of the biofilm matrix, in the close contacts between
136 different biofilm partners (e.g. mutually beneficial associations with cooperating microorganisms
137 with different nutritional requirement) and in interactions with the mineral substrate and the
138 atmosphere (Gorbushina 2007, **Figure 1**). Furthermore, the biofilm microenvironment provides
139 the community as a whole with an enormous capability to become resistant to biocide exposure.
140 Bacteria embedded in the biofilm matrix are remarkably more tolerant to biocides, up to 1,000-
141 fold relative to planktonic cultures of the same bacterial strains, depending on the species-drug
142 combination (Davies 2003). Conservation treatments with traditional doses of biocides are
143 sometimes insufficient to destroy all members of the biofilm community, and this is a cause of
144 concern for conservators (Cappitelli *et al.* 2011). Consequently, in the last few years, the efforts
145 have been directed towards implementing and developing preventive strategies (Cappitelli *et al.*
146 2011).

147 The documented presence of specialized microorganisms (*Inter alia* Golubic *et al.* 1981, Friedmann
148 and Ocampo-Friedmann 1984, Eppard *et al.* 1996, Laiz *et al.* 2009, Bastian *et al.* 2010, Cappitelli *et*
149 *al.* 2012, Polo *et al.* 2012, Ettenauer *et al.* 2014), highlights the existence of multiple trophic levels
150 (McNamara and Mitchell 2005) with a simultaneous bottom up (resource supply-driven) and top-
151 down (food web structure-driven) control of ecosystem structure and function, emergent patterns
152 of organization (Gorbushina 2007), ecological succession (Hoppert and König 2006) and ecosystem
153 stability founded on diversity (Miller *et al.* 2009, **Figure 2**).

154 The above-mentioned characteristics are the hallmarks of a 'complete' and 'complex'
155 environmental biological system.

156 Moreover, despite conditions perceived by us as 'extreme', the primary production rate of
157 epilithic communities can be high, comparable on a $\text{gC m}^{-2} \text{y}^{-1}$ basis to rates for many terrestrial
158 and ocean ecosystems (Büdel 1999). Interestingly, this suggests that carbon fixation rates in less

159 then 100 μm thick biofilms are broadly equivalent to those achieved across the ocean photic zone
160 (Deneff *et al.* 2010).

161 The relatively low species complexity, the defined ecological succession patterns and trophic level,
162 the tight biological-geochemical coupling, and the high biological productivity are important
163 features that make SABs inhabiting stone surfaces a good model system to generate simple and
164 clearly defined hypotheses to be tested across a range of environments.

165 As these biological systems are involved in the processing of weathered rock material, they might
166 be considered perfect model systems for studying biogeochemical processes and pedogenesis and
167 promising indicators of climate changes, being coupling agents between the atmosphere and the
168 lithosphere (Warscheid and Braams 2000, Gorbushina 2007, Villa *et al.* 2015). In addition, SABs
169 demonstrate mutually neutral or even beneficial associations as, in such hostile environments, the
170 metabolic costs of survival are so high that antibiosis is often an unaffordable luxury, making them
171 potential system to study symbiosis (Gorbushina *et al.* 2005, Gorbushina and Broughton 2009).

172 The ability of SABs to impact the lithic substrate and to buffer and adapt to both natural and
173 anthropogenic changes provides a number of significant ecosystem services essential to human
174 communities and societies (**Table 2**).

175

176 **Towards a traits-based approach to SAB ecology**

177 We advocate that an improved appreciation of the ecology of SABs inhabiting outdoor stone
178 materials will strengthen our ability to predict the impact of environmental change and to develop
179 management strategies for protection prior to, or following after, an active conservation
180 treatment.

181 Until now, the scientific community traditionally viewed SABs through a taxonomic lens, often
182 resulting in the loss of ecological generality. Although genomes and metagenomes give a detailed

183 cross section of the functional potential of a community, the functional traits (morphological,
184 biochemical, physiological, structural, phenological, or behavioral characteristics) are those
185 properties that interact directly with the environment, providing more relevant information in a
186 community analysis with special emphasis on feedback responses to environmental change and
187 biodecay phenomena of cultural heritage.

188 In a recent work, Villa and colleagues (2015) used a traits-based approach to reveal the metabolic
189 capabilities of SABs inhabiting historic limestone tombstones in response to atmospheric sulfur
190 pollution. They elucidated functional interaction networks and syntrophic interplays that enable
191 cooperative growth in SAB communities (**Figure 3**). This study showed also the ability of SABs to
192 perceive the external environment and to buffer environmental perturbations.

193 Thus, the long-standing question “what is there?” should switch to the questions “why is it
194 there?”, “how does it interact with the external environment?”, and “how does it respond to a
195 disturbance event?”. In addition, recent developments in community ecology have begun to
196 recognize that microbial assemblages cannot be defined without reference to their environments
197 (Konopka 2009). An appreciation for the tight interrelationship between microbes and their
198 physical and chemical environments is particularly important for delineation of microbial
199 communities and their ability to respond to a changing environment (O’Donnell *et al.* 2007).

200 Because functional traits mediate the interactions among microorganisms as well as between
201 microorganisms and the environment, it has been argued that trait-based approaches provide
202 more relevant information in a community analysis and ecosystem service than taxonomic or
203 phylogenetic attributes (Violle *et al.* 2007, Boon *et al.* 2014, Krause *et al.* 2014). As Cohan & Perry
204 (2007) state, ‘...the recognized “species” of bacterial systematics frequently contain a diversity of
205 populations that are distinct in their biochemistry, physiology, genome content and ecology;
206 classifying an unknown organism to its species thus tells us only vaguely about the organism’s way

207 *of life*'. Functional differences, even in only few critical pathways, could reflect dramatically
208 altered ecosystem properties and could impact the services or disservices that human societies
209 derive from them (Luck *et al.* 2009). This functional approach is instrumental for unraveling the
210 role of SABs in biodeterioration or bioprotection of stone monuments, as detecting
211 microorganisms does not automatically imply an involvement in the biodecay process of the lithic
212 substrate. The axiomatic correlation among microorganisms and stone decay is matter of
213 controversy, as it is far from clear why some communities are deteriorative and others are
214 protective or, indeed, why can be deteriorative under some environmental conditions and
215 bioprotective under others (Viles and Cutler 2012, Bartoli *et al.* 2014, Pinna 2014). Only a traits-
216 based approach may reveal the dual role of SABs and their inhabitants, and how this dual role
217 affects conservation strategies.

218

219 **Trait-based approach to predicting feedback responses of SABs to a changing environment**

220 Frameworks that group microorganisms into functional groups along a few trait axes have helped
221 to summarize biological variation and has led to the development of hypotheses to explain the
222 origins of functional diversity, the distribution and abundance of species, and the consequences of
223 functional traits for ecosystem functioning (Chagnon *et al.* 2013).

224 For example, a simple model sees the characterization of microorganisms according to their life-
225 history strategy: r-strategists (termed copiotrophs in microbial ecology) have high growth rates
226 and low resource use efficiency, and K-strategists (termed oligotrophs in microbial ecology) have
227 low growth rates and high resource use efficiency (Fierer *et al.* 2007). This assumed fundamental
228 trade-off between growth rate and resource use efficiency might underlie the capacity of
229 microbial communities to respond to disturbance, as community structure will change if the taxa
230 present differences in this trade-off (Wallenstein and Hall 2012). There is evidence from both plant

231 and soil communities that K-strategists are more resistant (the ability of a community property or
232 process to remain unchanged in the face of a specific disturbance), but less resilient (the ability of
233 a community property or process to recover after a specific disturbance, often reported as a rate
234 of return), to climate change-related disturbances than r-strategists (Bapiri *et al.* 2010, Lennon *et*
235 *al.* 2012), and a trade-off between resistance and resilience is widely documented (De Vries *et al.*
236 2012). De Vries and Shade (2013) proposed that simple measures that characterize microbial
237 communities along the r-K spectrum could inform their ability to resist and recover from climate
238 change related disturbances.

239 Nevertheless, the r-K framework has been criticized for its oversimplification of life history
240 strategies along a single axis that combines both disturbance and resource availability. Other
241 models that integrate additional axes have thus been proposed to more completely characterize
242 diversity while at the same time remain simple and tractable (Chagnon *et al.* 2013). The
243 Competitor-Stress tolerant-Ruderal (CSR) framework developed for plants, overcomes some
244 limitations of other models by classifying plant life history strategies according to the functional
245 traits associated with responses to two major environmental filters, namely stress and disturbance
246 (Grime 1977). Stress refers to persistent adverse environmental conditions (e.g. increasing
247 temperature and UV levels, decreasing moisture levels), whereas disturbance refers to episodic
248 events leading to significant loss of functional biomass (e.g. fire, drought, storms or erosion). The
249 C-S-R framework identifies three main life history strategies: 1) 'competitors' are adapted for rapid
250 resource utilization and long-term site occupation, 2) 'stress tolerators' are adapted to persist in
251 low-resource environments owing to resource conservation strategies, and 3) 'ruderals' cope with
252 frequent disturbance by relying on high colonization ability, rapid production of low cost biomass
253 and short reproductive cycles (Prosser *et al.* 2007).

254 Recently, Viles and Cutler (2012) employed the C-S-R framework as an example to show how trait-
255 based classification approach can predict the responses of heritage biota in terms of
256 biodeterioration, bioprotection and biological soiling to environmental changes. According to
257 Hoppert and König (2006), opportunistic and ruderal taxa within SABs colonizing stone
258 monuments are more likely to be deteriorative, as they colonize rapidly after disturbance and use
259 a range of strategies to derive nutrients from the substrate (e.g. rapid, destabilizing, endolithic
260 growth). Such strategies may cause further disturbance to the surface through weathering,
261 favoring ongoing ruderal colonization. By contrast, stress-tolerant species are likely to be less
262 deteriorative as, according to Hoppert and König (2006), they do not cause disruption of the
263 surface. Indeed, some of the strategies they use to cope with stress (e.g. pigmentation) may even
264 have bioprotective role by protecting the artistic surface from weathering (Viles and Cutler 2012).
265 Following this path, Viles and Cutler (2012) predicted that areas likely to experience increased
266 frequency of climatic disturbances are likely to experience a shift from bioprotective to
267 biodeteriorative conditions. Furthermore, areas that are likely to face increased stresses (e.g.
268 decreased precipitation) will show a reduction in soiling rates, a switch to stress-tolerators and
269 knock-on decline in biodeterioration. They envisioned situations where conditions change from
270 stressed to disturbed (or *vice versa*), producing no net change in soiling rate, but a switch between
271 biodeterioration and bioprotection.

272 We summarized current knowledge of functional traits of the main microbial groups of a mature
273 SAB (**Table 3**), and incorporate them into the C-S-R framework to conceptualize SAB life strategies
274 in order to better predict of their responses to environmental changes (**Figure 4**).

275 The trait-based approach proposed provides a simplified representation of SAB life strategies.
276 Associations in nature will likely be much more complex because SAB communities will rarely be at
277 any of the three extremes of the C-S-R triangle, but most of the time will rather have a mixed life

278 history. Moreover, microorganisms can display competitive, ruderal or stress-tolerant
279 morphotypes at different stages of SAB development and under different environmental
280 conditions. However, we argue that integrating such a trait-based approach into an established life
281 history classification scheme, such as the C-S-R framework, can provide more mechanistic insights
282 about the relationship among SABs, stone and the environment. The idea would be to assign
283 taxonomic and functional information of a specific biofilm community retrieved on the artistic
284 surface within the three dimensions of C-S-R classification framework, providing the basis to
285 predict and assess SAB distribution, prevalence and response to stresses and disturbances. The
286 same approach was recently used by Ho *et al.* (2013) to classify the observational ecological
287 characteristics of methane-oxidizing bacteria and exploiting their life strategies to optimize the
288 performance of this community in respect to a desirable set of outputs.

289

290 **Moving ahead: future research directions**

291 Understanding the ecology of SABs is arguably one of the most compelling intellectual challenges
292 facing contemporary ecology. Although worthy for its intellectual merits alone, developing such an
293 understanding is essential to the management of outdoor cultural heritage for their benefits in
294 culture-related economic activities, socio-political development, urban sustainability, education
295 and environmental protection.

296 Predicting how under a changing environment SABs will influence the ecosystem processes they
297 mediate requires an approach that links change in fitness of individuals to population dynamics,
298 community composition and function. In particular, looking at the structure of functional traits on
299 a community-wide scale could provide us insight about the processes carried out by SABs and, in
300 turn, about what traits are associated with a particular environmental condition. A deeper

301 understanding of ecosystem function might represent a way to manipulate the growth of SABs on
302 surfaces.

303 We think that several topics of research should be prioritized in order to predict the feedback
304 response of SABs to anthropogenic changes, and to develop microorganism-mediated approaches
305 to protect artistic surfaces and mitigate the effect of stresses and disturbances.

306 First, we need to understand and quantify the functional traits of SABs that may impact their
307 fitness in a given habitat and their responses to a changing environment. Second, we need to map
308 taxonomic information into a functional space, in order to assign ecological niches to different
309 microbial taxa and elucidate SAB/stone and SAB/atmosphere interactions. Third, we need to
310 quantify the biodeterioration of artistic surfaces and the effects of environmental changes on
311 stone geochemistry. Fourth, we need to improve our understanding of microbial responses to
312 simultaneous environmental stressors (Staudt *et al.* 2013). Finally, we need to create a framework
313 to incorporate biological (omics), environmental, chemical and geological data into mathematical
314 models, in order to offer a system-level understanding of the phenomenon, reducing uncertainty
315 and improving quantitative estimation and prediction.

316 Recent advances in 'omic' technologies, computational science and the ease with which data can
317 be shared and forwarded will provide the opportunity to integrate knowledge across disciplines to
318 generate an increasingly comprehensive understanding of the SABs responses to a changing
319 environment, and how they will influence the ecosystem where they are growing in. Progress will
320 require collaborative research among different disciplines. We envision that contributions by five
321 different groups will be particularly useful:

322 1) Partnerships between conservators, heritage managers and ecosystem scientists to sample
323 SABs on outdoor stone surfaces across a global-scale gradient of biomes. The comparison
324 of the taxonomic and functional dimension of SABs over a wide range of both spatial- and

325 timescales, will lead to hypotheses about the relationships between environmental
326 changes and potential microbiological damage. This effort should be coordinated with
327 ongoing long-term research networks and utilize existing data sets to the fullest possible
328 extent.

329 2) Collaborations with molecular biologists and bioinformaticians to apply next generation
330 sequencing technologies to look for functional patterns in the samples collected from
331 around the world. This information would allow testing hypotheses about the time and
332 mode of SABs response to a changing environment.

333 3) Collaborations with biochemists to identify biomarkers in the form of metabolites, proteins
334 or transcript pools that signify ecosystem state at the onset of a transition. This knowledge
335 will inform hypotheses about the potential role for SABs in C, N, P and S dynamics in a
336 changing environment.

337 4) Work together with mathematicians to incorporate space- and time-resolved omics and
338 environmental data into new models to test hypotheses about the role of SABs for
339 biogeochemical cycles, biodeterioration vs. bioprotection of stone, ecosystem productivity
340 and climate.

341 5) Most importantly, research findings should be used to build relationship and open lines of
342 communication between researchers and stakeholders, to facilitate the translation of
343 research findings into actions. Researchers can profile their achievements and stakeholders
344 can be informed of research outcomes and influence research challenges.

345 The complexity of the phenomenon under investigation requires interdisciplinary research if we
346 are to attain the predictive capability that could inform policy makers. The potential for
347 interdisciplinary research ultimately hinges on the extent to which individuals want to engage in it,
348 and equally importantly if they have the opportunity to do so. Granting agencies are encouraging

349 multidisciplinary approaches by increasingly providing support for crosscutting research efforts.
350 There is no better time for seizing the opportunity to establish and fine-tune the collaboration
351 with co-workers in other fields.

352

353 **Acknowledgements**

354 This research has received funding from the European Union Seventh Framework Programme
355 (FP7-PEOPLE-2012-IOF) under grant agreement n° 328215.

356 The authors gratefully acknowledge the National Park Service for providing access to the roofs of
357 the Lincoln Memorial, Thomas Jefferson Memorial, and Federal Hall National Memorial.

359 **Table 1:** Mechanisms by which SABs can alter and engineer their habitat.

Mechanisms	References
Respiration of bacteria and fungi increases local CO ₂ concentrations: <ul style="list-style-type: none"> • Formation of H₂CO₃, which decreases the pH of the stone surface and leaches out carbonates, phosphates and silicates. 	Warscheid and Braams 2000, Dakal and Cameotra 2012, Sterflinger and Piñar 2014
Production of ligand-based agents (e.g. organic anions, siderophores): <ul style="list-style-type: none"> • Chelation of Ca, Mg, and Fe, which promote the dissolution of cationic constituents. 	Warscheid and Braams 2000, Hoffland <i>et al.</i> 2004, Dakal and Cameotra 2012
Production of acids: <ul style="list-style-type: none"> • Promotion of the dissolution and/or chelation of cations. • Weakening of the mineral lattice by dissolution of metal cations. • Precipitation of calcium oxalate. 	Warscheid and Braams 2000, McNamara and Mitchell 2005, Gorbushina 2007, Sterflinger and Piñar 2014
Production of extracellular polymeric substances (EPS): <ul style="list-style-type: none"> • Dessiccation/hydration cycles of the EPS cause separation of particles. • Regulation of the humidity, thermal transmission and water vapor diffusion, reducing thermo-hydric stresses to the stone. • Wrapping the grains with a biogenic matrix temporarily stabilizes the surface and reduces weathering. 	Warscheid and Braams 2000, Crispim and Gaylarde 2004, Gorbushina 2007, Pinna 2014, Sterflinger and Piñar 2014
Uptake and accumulation of sulfur and calcium into the cells: <ul style="list-style-type: none"> • Weakening of the stone matrix. • Growth of cells forces separation of mineral grains. 	Crispim and Gaylarde 2004, Scheerer <i>et al.</i> 2009, Dakal and Cameotra 2012
Endolithic growth: <ul style="list-style-type: none"> • Contribution to the breakdown of rock crystalline structures. 	Golubic <i>et al.</i> 1981, Crispim and Gaylarde 2004, Scheerer <i>et al.</i> 2009
Hyphae and filamentous growth: <ul style="list-style-type: none"> • Contribution to the breakdown of rock crystalline structures. 	Sterflinger and Krumbein 1997, Warscheid and Braams 2000, Hoffland <i>et al.</i> 2004
Create a multitude of varnish-like coatings: <ul style="list-style-type: none"> • Discoloration. • Discolored areas may absorb more sunlight, which increases physical stress by expansion and contraction caused by temperature changes. 	Warscheid and Braams 2000, Gorbushina <i>et al.</i> 2002, Crispim and Gaylarde 2004, Noack-Schönmann <i>et al.</i> 2014

361 **Table 2:** Ecosystem services provided by SABs inhabiting stone surfaces
 362

Service	Mechanism
Biogeochemical cycles	Nutrient cycling, specific elemental transformation (e.g. nitrification and sulfur oxidation).
Atmospheric change indicators	By intercepting compounds carried by the air, SABs and their activity are under the direct influence of the atmospheric input.
Climate regulators	Carbon sequestration, nutrient cycling, specific elemental transformation (e.g. nitrification, sulfur oxidation).
Culture and conservation of stone monuments with impacts on recreation, tourism and economy	Cultural heritage is often associated with the identity of an individual, a community or a society. Cultural heritage provides experiences shared across generations, as well as settings for communal interactions important to cultural ties. Conservation of stone monuments has indirect impacts on tourism and recreation activities. Tourism and recreation activities are estimated to contribute € 415 billion to the EU GDP and 3.4 million tourism enterprises account for 15.5 million jobs (EU Communication 2014). In addition, visitor's expenditure generates income for the local communities and infrastructure development.

363

364

365 **Table 3:** Ecological characteristics of the main microbial groups of SABs (Fungi, Bacteria and Algae)
 366 inhabiting stone surfaces. C: Competitor; R: Ruderal; S: Stress tolerator

FUNGI			
Group	Ecological characteristics	References	Class
Hyphomycetes (Hyp)	<ul style="list-style-type: none"> • Fast growing in comparison to MCF. • Different abilities to access limiting resources (e.g. production of siderophores). • Ability to scavenge nutrients from the air and rain. • Pigment production. • Hyphal growth and reproductive structures. • Production of asexual spores. • High dispersal rates in comparison to MCF. • Epilithic and endolithic growth. 	Sterflinger and Krumbein 1997, Cutler and Viles 2010, Nai <i>et al.</i> 2013, Sterflinger and Piñar 2013	C C/R
Micro colonial fungi (MCF)	<ul style="list-style-type: none"> • Slow growing in comparison to Hyphomycetes. • Accumulation of storage compounds. • High resistance to desiccation, UV radiation and osmotic stress. • Swollen, isodiametric cells with thick, melanin containing cell walls. • Compact microcolonies on and inside the stone. • No aerial mycelium. • Capacity to survive long period of suspended metabolism. • Create a multitude of varnish-like coatings. • Production of survival propagules. 	Sterflinger and Krumbein 1997, Nai <i>et al.</i> 2013, Cutler and Viles 2010, Sterflinger and Piñar 2013	C C/S

367

BACTERIA			
Group	Ecological characteristics	References	Class
Cyanobacteria (Cya)	<ul style="list-style-type: none"> • Simple nutritional requirement. • Slow growing. • Ability to store essential nutrients and metabolites. • Production of photosynthetic and/or protective pigments. • Production of exopolymers. • Harbor a number of repair and tolerance mechanisms to counter the effects of UV and oxidative stress. • Efficient response to moisture status. • Epilithic and endolithic growth. • Limited mobility. 	Crispim and Gaylarde 2005, Scheerer <i>et al.</i> 2009, Sterflinger and Piñar 2013	S S/C
Actinobacteria (Act)	<ul style="list-style-type: none"> • High growth rate. • High cellular turnover rates and short life cycle. • Small cell size. • Metabolic plasticity and rapid response to different substrates. • Production of soluble pigments. • Hyphal growth. • Endolithic growth. • Early production of asexual spores. • More efficient dispersal mechanisms. 	Eppard <i>et al.</i> 1996, Gorbushina 2007, Scheerer <i>et al.</i> 2009, Sterflinger and Piñar 2013	R R/C R/S
Lithotrophs (Lit)	<ul style="list-style-type: none"> • Simple nutritional requirement. • Slow growing. • Release of inorganic and organic acids. • Accumulation of storage compounds. • Small cell size. 	Golubic <i>et al.</i> 1981, Warscheid and Braams 2000	S/C

368
369

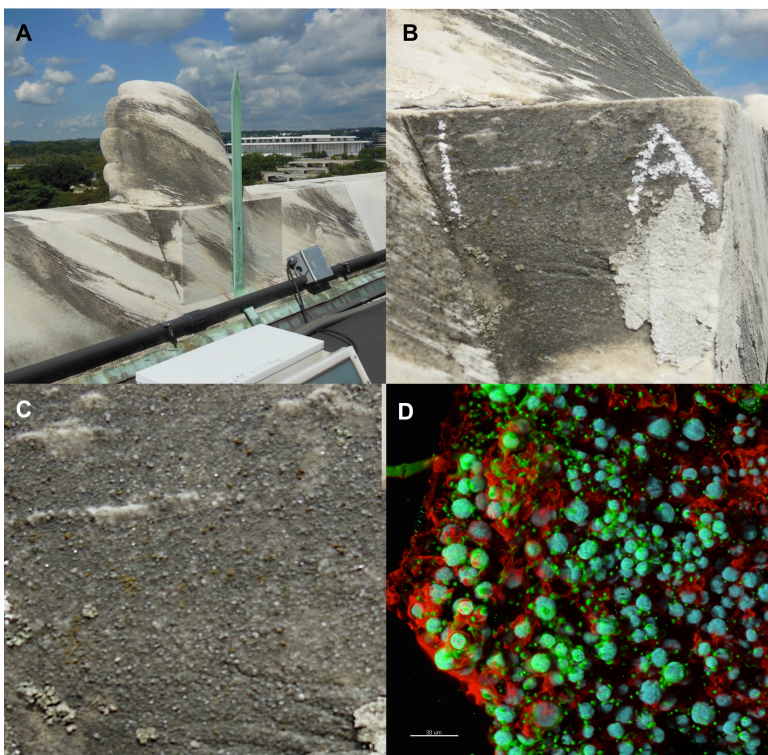
ALGAE			
Group	Ecological characteristics	References	Class
Green algae (GA)	<ul style="list-style-type: none"> • Simple nutritional requirement. • Slow growing. • Accumulate organic osmolytes to face osmotic stress. • Protection against oxidative stress via non-photochemical quenching. • Cope with high light condition by producing protective carotenes and xanthophyles. • Able to use water vapor. • Mixotrophy. • Algal propagules can remain viable in the atmosphere for extended period. 	Gorbushina 2007, Scheerer <i>et al.</i> 2009, Cutler and Viles 2010	S S/C

370

371 **Figure captions**

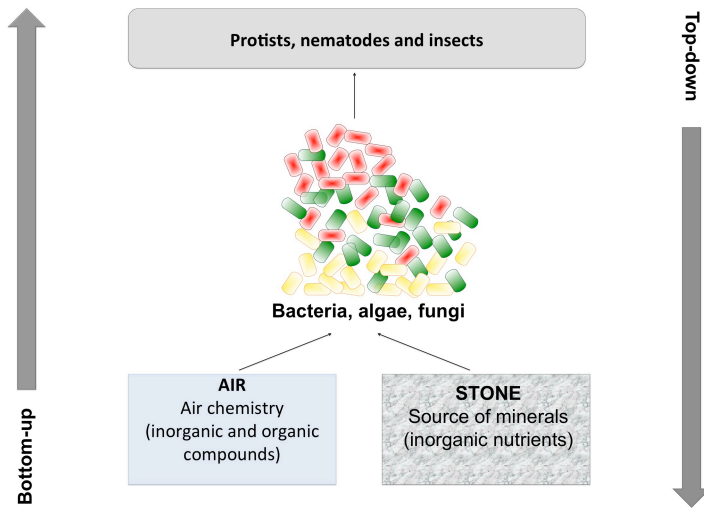
372

373 **Figure 1:** A SAB growing on the white marble of the Lincoln Memorial, Washington, DC. (A) Antefix
374 on the roof of the Lincoln Memorial. (B-C) Close-up shots of a vertical SAB on the Jefferson
375 Memorial. (D) Confocal laser scanning imaging of a biofilm taken from this location. Blue are
376 microcolonies of photoautotrophic microbes, green are chemoheterotrophic microbes, and red
377 are extracellular polymeric substances.



378

379 **Figure 2:** Multiple trophic levels in SABs inhabiting stone surfaces. The microbial food web in SABs
380 is influenced by both bottom-up (resource-supply driven) and top-down (predation-driven) forces.



381

382

383

384

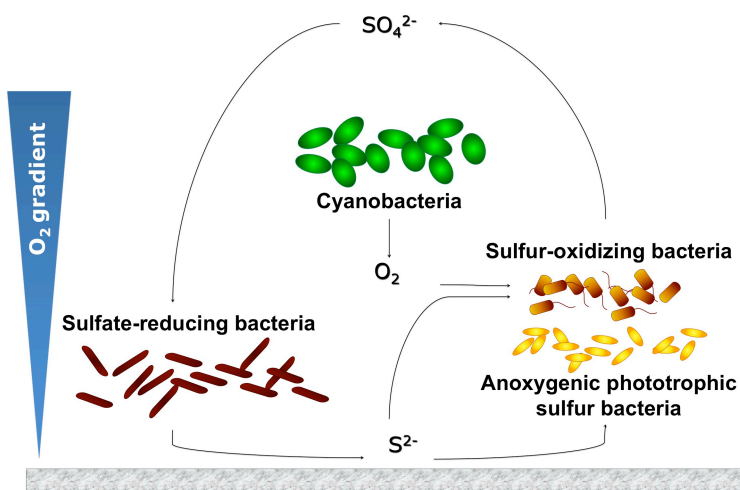
385

386

387

388

Figure 3: Interdependent cycling of nutrients that occurs among the main functional groups retrieved on a tombstone located in a polluted environment. The organic carbon produced by cyanobacteria during photosynthesis supports the growth of microorganisms that require organic matter as energy source such as sulfate reducing bacteria (SRB), and sulfur oxidizing bacteria (SOB). SOB consumes the oxygen produced by cyanobacteria, creating the anaerobic environment for SRB and anoxygenic phototrophic sulfur bacteria. The SOB quickly remove the metabolic products of SRB, S^{2-} , that could inhibit cyanobacteria and at higher concentrations also SRB.



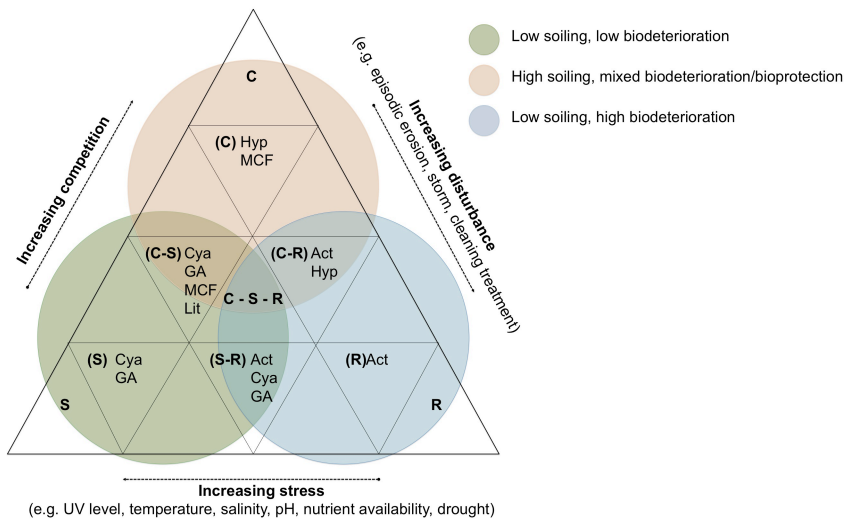
389

390

391

392

Figure 4: Reflection of SAB microbial traits on the Competitor (C)-Ruderal (R)-Stress tolerator (S) life strategy framework as was proposed for plants by Grime (1977). Hyp: Hyphomycetes; MCF: Micro Colonial Fungi; Cya: Cyanobacteria; Act: Actinobacteria; GA: Green Algae; Lit: Lithotrophs.



393

394 **References**

395

396 Bapiri A, Bååth E, Rousk J. 2010. Drying-rewetting cycles affect fungal and bacterial growth
397 differently in an arable soil. *Microbial Ecology* 60: 419-28.

398 Bartoli F, Casanova Municchia A, Futagami Y, Kashiwadanic H, Moond KH, Caneva G. 2014.

399 Biological colonization patterns on the ruins of Angkor temples (Cambodia) in the

400 biodeterioration vs bioprotection debate. *International Biodeterioration Biodegradation* 96:

401 157-65.

402 Bastian F, Jurado V, Nováková A, Alabouvette C, Saiz-Jimenez C. 2010. The microbiology of Lascaux

403 Cave. *Microbiology* 156: 644-52.

404 Boon E, Meehan CJ, Whidden C, Wong DH, Langille MG, Beiko RG. 2014. Interactions in the

405 microbiome: communities of organisms and communities of genes. *FEMS Microbiology Reviews*

406 38: 90-118.

407 Büdel B. 1999. Ecology and diversity of rock-inhabiting cyanobacteria in tropical regions. *European*

408 *Journal of Phycology* 34: 361-70.

409 Cappitelli F, Salvadori O, Albanese D, Villa F, Sorlini C. 2012. Cyanobacteria cause black staining of

410 the National Museum of the American Indian Building, Washington, DC, USA. *Biofouling* 28:

411 257-66.

412 Cappitelli F, Villa F, Sorlini C. 2011. New environmentally friendly approaches against

413 biodeterioration of outdoor cultural heritage. In *Biocolonization of Stone: Control and*

414 *Preventive Methods, Proceedings from the MCI Workshop Series*, eds. Charola A. E., McNamara

415 C., Koestler R. J. Washington: Smithsonian Institution Scholarly Press (SISP), 51-58.

416 Chagnon PL, Bradley RL, Maherali H, Klironomos JN. 2013. A trait-based framework to understand

417 life history of mycorrhizal fungi. *Trends in Plant Science* 18: 484-91.

418 Cohan FM, Perry EB. 2007. A systematics for discovering the fundamental units of bacterial
419 diversity. *Current Biology* 17: R373-86.

420 Crispim CA, Gaylarde CC. 2005. Cyanobacteria and biodeterioration of cultural heritage: a review.
421 *Microbial Ecology* 49: 1-9.

422 Dakal TC, Cameotra SS. 2012. Microbially induced deterioration of architectural heritages: routes
423 and mechanisms involved. *Environmental Sciences Europe* 24: 36.

424 Davies D. 2003. Understanding biofilm resistance to antibacterial agents. *Nature Reviews Drug*
425 *Discovery* 2: 114-22.

426 Denev VJ, Mueller RS, Banfield JF. 2010. AMD biofilms: using model communities to study
427 microbial evolution and ecological complexity in nature. *ISME J.* 4: 599-610.

428 de Vries FT, Liiri ME, Strandmark LB, Bowker MA, Christensen S, Setälä HM, Bardgett RD. 2012.
429 Land use alters the resistance and resilience of soil food webs to drought. *Nature Climate*
430 *Change* 2: 276-80.

431 de Vries FT, Shade A. 2013. Controls on soil microbial community stability under climate change.
432 *Frontiers in Microbiology* 4: 265.

433 Eppard M, Krumbein WE, Koch C, Rhiel E, Staley JT, Stackebrandt E. 1996. Morphological,
434 physiological, and molecular characterization of actinomycetes isolated from dry soil, rocks,
435 and monument surfaces. *Archives of Microbiology* 166: 12-22.

436 Ettenauer JD, Jurado V, Piñar G, Miller AZ, Santner M, Saiz-Jimenez C, Sterflinger K. 2014.
437 Halophilic microorganisms are responsible for the rosy discolouration of saline environments in
438 three historical buildings with mural paintings. *PLoS ONE* 9: e103844.

439 EU Communication. 2010. Europe 2020. A strategy for smart, sustainable and inclusive growth.
440 Available at [http://eur-](http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=COM:2010:2020:FIN:EN:PDF)
441 [lex.europa.eu/LexUriServ/LexUriServ.do?uri=COM:2010:2020:FIN:EN:PDF](http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=COM:2010:2020:FIN:EN:PDF)

442 EU Communication. 2014. Towards an integrated approach to cultural heritage for Europe.
443 Available at [http://ec.europa.eu/culture/library/publications/2014-heritage-](http://ec.europa.eu/culture/library/publications/2014-heritage-communication_en.pdf)
444 [communication_en.pdf](http://ec.europa.eu/culture/library/publications/2014-heritage-communication_en.pdf)

445 Fierer N, Bradford MA, Jackson RB. 2007. Toward an ecological classification of soil bacteria.
446 *Ecology* 88: 1354-64.

447 Friedmann EI, Ocampo-Friedmann R. 1984. Endolithic microorganisms in extremely dry
448 environments: analysis of a lithobiontic microbial habitat. *Current Perspectives in Microbial*
449 *Ecology*. M. J. Klug and C. A. Reddy. Washington, D.C., American Society for Microbiology: 177-
450 85.

451 Golubic S, Friedmann I, Schneider J. 1981. The lithobiontic ecological niche with special reference
452 to microorganisms. *Journal of Sedimentary Petrology* 51: 475-78.

453 Gorbushina AA, Beck A, Schulte A. 2005. Microcolonial rock inhabiting fungi and lichen
454 photobionts: evidence for mutualistic interactions. *Mycological Research* 109: 1288-96.

455 Gorbushina AA, Broughton WJ. 2009. Microbiology of the atmosphere-rock interface: how
456 biological interactions and physical stresses modulate a sophisticated microbial ecosystem.
457 *Annual Review of Microbiology* 63: 431-50.

458 Gorbushina AA. 2007. Life on the rocks. *Environmenta Microbiology* 9: 1613-31.

459 Gorbushina AA, Krumbein WE, Volkmann M. 2002. Rock surfaces as life indicators: new ways to
460 demonstrate life and traces of former life. *Astrobiology* 2: 203-13.

461 Grime J. 1977. Evidence for the existence of three primary strategies in plants and its relevance to
462 ecological and evolutionary theory. *American Naturalist* 111: 1169-97.

463 Ho A, Kerckhof FM, Luke C, Reim A, Krause S, Boon N, Bodelier PLE. 2013. Conceptualizing
464 functional traits and ecological characteristics of methane-oxidizing bacteria as life strategies.
465 *Environmental Microbiology Reports* 5: 335-45.

466 Hoffland E, Kuyper TW, Wallander H, Plassard C, Gorbushina AA, Haselwandter K. 2004. The role of
467 fungi in weathering. *Frontiers in Ecology and the Environment* 2: 258-64.

468 Hoppert M, Konig S. 2006. The succession of biofilms on building stone and its possible impact on
469 biogenic weathering. *Proceedings of the Congress Heritage, Weathering and Conservation;*
470 *London (2006), Taylor & Francis Group, pp. 311-15.*

471 Konopka A. 2009. What is microbial community ecology? *ISME Journal* 3: 1223-30.

472 Krause S, Le Roux X, Niklaus PA, Van Bodegom PM, Lennon JT, Bertilsson S, Grossart HP, Philippot
473 L, Bodelier PL. 2014. Trait-based approaches for understanding microbial biodiversity and
474 ecosystem functioning. *Frontiers in Microbiology* 5: 251.

475 Laiz L, Miller AZ, Jurado V, Akatova E, Sanchez-Moral S, Gonzalez JM, Dionísio A, Macedo MF, Saiz-
476 Jimenez C. 2009. Isolation of five Rubrobacter strains from biodeteriorated monuments.
477 *Naturwissenschaften* 96: 71-79.

478 Lennon JT, Aanderud ZT, Lehmkuhl BK, Schoolmaster DR Jr. 2012. Mapping the niche space of soil
479 microorganisms using taxonomy and traits. *Ecology* 93: 1867-79.

480 Luck GW, et al. 2009. Quantifying the contribution of organisms to the provision of ecosystem
481 services. *BioScience* 59: 223-35.

482 MA 2005. Ecosystems and human well-being: current state and trends: Findings of the Conditions
483 and Trends Working Group. In: Hassan R, Scholes R, Ash N. (Eds.), *Millennium Ecosystem*
484 *Assessment (MA)*. Island Press, Washington.

485 Macedo MF, Miller AZ, Dionísio A, Saiz-Jimenez C. 2009. Biodiversity of cyanobacteria and green
486 algae on monuments in the Mediterranean Basin: an overview. *Microbiology* 155: 3476-90.

487 Macedo MF, Miller AZ, Dionísio A, Saiz-Jimenez C. 2009. Growth of phototrophic biofilms from
488 limestone monuments under laboratory conditions. *International Biodeterioration*
489 *Biodegradation* 63: 860-67.

490 McNamara CJ, Mitchell R. 2005. Microbial deterioration of historic stone. *Frontiers in Ecology and*
491 *the Environment* 3: 445-51.

492 Nai C, Wong HY, Pannenbecker A, Broughton WJ, Benoit I, de Vries RP, Gueidan C, Gorbushina AA.
493 2013. Nutritional physiology of a rock-inhabiting, model microcolonial fungus from an ancestral
494 lineage of the Chaetothyriales (Ascomycetes). *Fungal Genetics and Biology* 56: 54-66.

495 Noack-Schönmann S, Spagin O, Gründer K-P, Breithauptb M, Günterb A, Muschikb B, Gorbushinaa
496 AA. 2014. Sub-aerial biofilms as blockers of solar radiation: spectral properties as tools to
497 characterise material-relevant microbial growth. *International Biodeterioration Biodegradation*
498 86: 286-93.

499 O'Donnell AG, Young IM, Rushton SP, Shirley MD, Crawford JW. 2007. Visualization, modelling and
500 prediction in soil microbiology. *Nature Review Microbiology* 5: 689-99.

501 Pinna D. 2014. Biofilms and lichens on stone monuments: do they damage or protect? *Frontiers in*
502 *Microbiology* 5: 133.

503 Polo A, Gulotta D, Santo N, Di Benedetto C, Fascio U, Toniolo L, Villa F, Cappitelli F. 2012.
504 Importance of subaerial biofilms and airborne microflora in the deterioration of stonework: a
505 molecular study. *Biofouling* 28: 1093-106.

506 Prosser JI, et al. 2007. The role of ecological theory in microbial ecology. *Nature Reviews*
507 *Microbiology* 5: 384-92.

508 Scheerer S, Ortega-Morales O, Gaylarde C. 2009. Microbial deterioration of stone monuments--an
509 updated overview. *Advances in Applied Microbiology* 66: 97-139.

510 Staudt A, Leidner AK, Howard J, Brauman KA, Dukes JS, Hansen LJ, Paukert C, Sabo J, Solórzano LA.
511 2013. The added complications of climate change: understanding and managing biodiversity
512 and ecosystems. *Frontiers in Ecology and the Environment* 11: 494-501.

513 Sterflinger K, Piñar G. 2013. Microbial deterioration of cultural heritage and works of art--tilting at
514 windmills? *Applied Microbiology and Biotechnology* 97: 9637-46.

515 Sterflinger K, Krumbein WE. 1997. Dematiaceous fungi as a major agent for biopitting on
516 Mediterranean marbles and limestones. *Geomicrobiology Journal* 14: 219-22.

517 Stewart PS, Franklin MJ. 2008. Physiological heterogeneity in biofilms. *Nature Reviews*
518 *Microbiology* 6: 199-210.

519 Stone M. 2015. Small Talk: The Evolution of Bacterial Languages. *BioScience* 65: 336.

520 Tweed C, Sutherland M. 2007. Built cultural heritage and sustainable urban development.
521 *Landscape Urban Plan* 83: 62-69.

522 UNESCO–April 2013 1 Sessions 3A and 3A-a Introducing cultural heritage in to the sustainable
523 development agenda. Available at
524 <http://www.unesco.org/new/fileadmin/MULTIMEDIA/HQ/CLT/images/HeritageENG.pdf>

525 United Nations Conference on Sustainable Development. 2012. Rio de Janeiro, Brazil, 20-22 June,
526 2012.

527 Viles HA, Cutler NA. 2012. Global environmental change and the biology of heritage structures.
528 *Global Change Biology* 18: 2406-18.

529 Villa F, Vasanthakumar A, Mitchell R, Cappitelli F. 2015. RNA-based molecular survey of
530 biodiversity of limestone tombstone microbiota in response to atmospheric sulphur pollution.
531 *Letters in Applied Microbiology* 60: 92-102.

532 Violle C, Navas ML, Vile D, Kazakou E, Fortunel C, Hummel I, Garnier E. 2007. Let the concept of
533 trait be functional! *Oikos* 116: 882-92.

534 Wallenstein MD, Hall EK. 2012. A trait-based framework for predicting when and where microbial
535 adaptation to climate change will affect ecosystem functioning. *Biogeochemistry* 109: 35-47.

- 536 Warscheid T, Braams J. 2000. Biodeterioration of stone: a review. *International Biodeterioration &*
537 *Biodegradation* 46: 343-68.
- 538 Zakharova K, Tesei D, Marzban G, Dijksterhuis J, Wyatt T, Sterflinger K. 2013. Microcolonial fungi
539 on rocks: a life in constant drought? *Mycopathologia* 175: 537-47.
- 540
- 541