

1 **Physicochemical and rheological properties of gluten-free blends containing**
2 **differently treated chickpea flours**

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24 **Abstract**

25 This study focused on the evaluation of the physicochemical and rheological properties of
26 chickpea flours and blends obtained by partially substituting rice flour (25%) with raw,
27 roasted and dehulled chickpea flour. The characteristics of the resultant doughs were
28 evaluated. In comparison with rice flour, blends containing chickpea flours exhibited
29 high protein and fat content, a reduced retrogradation tendency (setback values of 404-
30 415 vs. 479 BU) and a higher foaming capacity and stability, which can be beneficial for
31 their use in baked food formulations. However, roasting decreased foaming capacity and
32 stability. Even if the rheofermentographic test evidenced a slight reduction in dough
33 development, high CO₂ retention capacity (> 98%) and similar-to-lower leavening times
34 were observed for doughs containing chickpea flours. Incorporating chickpea flours also
35 caused an increase in the viscous and elastic moduli of rice-based doughs, resulting in a
36 good structuring of the dough. The results of this study indicated that chickpea flours
37 could be used as a healthy ingredient in gluten-free rice-based formulations.

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39 **Keywords:** Gluten-free dough; chickpea flour; physicochemical properties; dough
40 viscoelasticity

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42 **1. Introduction**

43 Gluten, a viscoelastic protein complex formed following the kneading process of wheat,
44 rye or barley, may cause health problems such as celiac disease (CD), wheat allergy and
45 non-celiac gluten sensitivity in a broad spectrum of populations. Among these disorders,
46 CD is an autoimmune metabolic disease occurring in 1% of population worldwide (Reilly

47 & Green, 2012). In CD patients, the consumption of gluten-containing foods leads to
48 damage of the small intestine with a consequent reduction in the absorption of nutrients.
49 The remedy for celiac and other gluten-related diseases is to exclude gluten from the diet.
50 The gluten-free (GF) diet is a real challenge, especially for celiac patients, since very low
51 amounts of gluten can trigger the symptoms.

52 In the last few decades, despite some improvements in the technological and nutritional
53 quality of GF bread (Alvarez-Jubete, Auty, Arendt, & Gallagher, 2010; Cappa, Barbosa-
54 Cánovas, Lucisano, & Mariotti et al., 2016; Cappa, Lucisano, & Mariotti, 2016; Mariotti,
55 Lucisano, Pagani, & Ng, 2009; Mariotti et al., 2017), the protein enrichment of GF bread
56 is still a research target for GF producing companies. Thus, the use of legumes is
57 nowadays considered a promising strategy for GF bread production with enhanced
58 nutritional properties.

59 Legumes are plants of the Leguminosae family, which are mostly planted for their seeds
60 called pulses such as peas, beans, chickpeas, lentils and cowpeas (FAO, 2016b).

61 Recently, pulses are of increasing interest due to their nutritional benefits (e.g., high
62 levels of proteins, complex carbohydrates, micronutrients and vitamins), and thus their
63 consumption is highly recommended. Indeed, 2016 was declared the International Year
64 of Pulses by the United Nations. Apart from their consumption as a whole seed, pulses
65 are used, after milling, in many food formulations (e.g., bakery products, pasta, baby
66 foods, etc.).

67 Chickpeas, one of the most important pulses, are mostly produced in Turkey, India,
68 Australia and Pakistan (FAO, 2016a). Chickpeas can be consumed as grains, meal or
69 snack. A special type of roasted chickpea snack, which is widely consumed in Turkey

70 and countries nearby, is called leblebi. During leblebi-processing several steps such as
71 tempering, moistening, resting and roasting are applied (Coşkuner & Karababa, 2004)
72 and the hulls of the chickpea seeds are almost completely removed. Moreover, some
73 chickpeas are split in half during processing and separated from the whole seeds as by-
74 products. After milling, these broken parts could be introduced in different food
75 formulations (e.g., bakery products, desserts, soups, etc.) as a cheap, sustainable and
76 nutritious ingredient.

77 To date, few studies related to the evaluation of doughs containing chickpea flour alone
78 or blended with other ingredients have been published (Aguilar, Albanell, Minarro, &
79 Capellas, 2015; Burešová, Kráčmar, Dvořáková, & Středa, 2014; Ouazib, Garzon, Zaidi,
80 & Rosell, 2016). Furthermore, to the best of our knowledge, no comprehensive study has
81 been published related to the evaluation of the properties of doughs containing chickpea
82 flours in combination with rice flour.

83 The aim of this study was to evaluate the physicochemical properties of raw, dehulled
84 and roasted chickpea flours and to study the effects of their addition to a rice flour-based
85 dough. During the production of roasted chickpeas, heat was applied and hulls were
86 almost completely removed. For this reason, raw and dehulled chickpea flours were also
87 included in the experimental plan in order to assess the effect of the roasting. Dough
88 containing 100% rice flour was considered as reference.

89 **2. Materials and methods**

90 **2.1. Materials**

91 The flours used were rice flour (RF; Beneo-Remy NV, Leuven-Wijgmaal, Belgium),
92 roasted (RCF), dehulled (DCF; Homecraft Pulse 4101, Ingredion, Germany) and raw

93 (CF) chickpea flours. RCF and CF were obtained from a local market and milled by a
94 laboratory mill to obtain flours having particle size ≤ 1 mm. The other ingredients used in
95 the dough were hydroxypropyl methyl cellulose (HPMC, Benecel F4M, Ashland, USA),
96 instant yeast (Pakmaya, Istanbul, Turkey), sugar, salt and sunflower oil.

97 **2.2. Design of experiments**

98 In order to evidence the effects of raw, roasted and dehulled chickpea flours on a rice
99 flour-based dough, the physicochemical properties of RF, CF, RCF, DCF were first
100 assessed. According to preliminary trails and the results of a previous study (Kahraman,
101 2016), three flour blends composed of 75.15% RF and 24.85% CF, RCF or DCF
102 and their respective bread doughs (RF+CF, RF+RCF and RF+DCF) were prepared and
103 characterized.

104 **2.3. Flour and blends properties**

105 **2.3.1. Proximate composition and particle size distribution**

106 The moisture content of the flour samples was determined via oven drying at 105 °C until
107 a constant weight was reached. The total nitrogen content of samples was determined
108 according to the Official Standard Method AOAC 920.87 (AOAC, 1999) by using a
109 block digestion system (Kjeldatherm, C. Gerhardt GmbH & Co. KG, Germany) and a
110 distillation system (Vapodest 50s, C. Gerhardt GmbH & Co. KG, Germany). Protein
111 content was calculated using 5.95 and 6.25 as conversion factors for rice and chickpea
112 flours, respectively. For fat content determination, flour samples (4 g) were extracted
113 with n-hexane (Sigma-Aldrich, Germany) by using an automatic extraction system
114 (Soxtherm, Gerhardt, Germany). Ash content was analyzed according to AACC (1999)

115 by using a muffle furnace (Protherm, Turkey). The evaluations were done at least in
116 duplicate and the results were expressed as percentages on a dry basis (db).
117 For the flour particle size distributions, 50 g of sample was placed in an analytical sieve
118 shaker (Octagon Digital, Endecotts Ltd., England) equipped with 6 sieves with 40, 90,
119 125, 250, 500 and 1000 μm openings. Plastic balls having diameters of 3 cm were placed
120 on sieves in order to facilitate sample distribution. Each fraction was collected after
121 sieving at amplitude 8 for 10 min. Results are the average of two determinations and are
122 given as percentages of each fraction per 100 g flour.

123 **2.3.2. Scanning electron microscopy (SEM) analysis**

124 In order to investigate the effects of the different treatment on the flour microstructure the
125 images of flours were captured by scanning electron microscope (XL 30S FEG, Philips)
126 under a voltage of 2.0 kV. Double-sided carbon tape having flour samples on one side
127 was attached to an aluminum stub and coated with gold under vacuum (0.09 mbar).

128 **2.3.3. Water binding capacities and foaming properties**

129 To assess water binding capacity (WBC) of blends and RF, each sample (2 g) was mixed
130 with deionized water (24 mL), shaken for 60 min (KS 130 Basic, IKA, Germany) and
131 centrifuged at 3460 x g at 25 °C for 10 min (Universal 320R, Hettich, Germany). The
132 supernatant was carefully discarded and the weights of the tubes were recorded. The
133 results (average of at least two measurements) were given as percentages of water held by
134 the dry sample.

135 Foam capacity (FC) and stability (FS) of blends and RF were determined according to
136 Shevkani, Singh, Kaur, & Rana (2015) with some modifications. The sample (2 g) was
137 mixed with deionized water (50 mL) and homogenized for 2 min (Ultra-Turrax T 25,

138 18G Dispenser, IKA, Germany). FC (%) was calculated as the volume ratio of foam and
139 initial volume, and FS (%) was given as the ratio of foam volume measured after 60 min
140 with respect to initial foam volume. At least three replications were performed.

141 **2.3.4. Pasting properties**

142 Blends and RF were analyzed for their pasting properties by using Brabender[®] Micro-
143 Visco-Amylograph (MVA) (Brabender OHG, Duisburg, Germany) according to Cappa et
144 al. (2013). Sample slurry was prepared by dispersing sample (12 g) in distilled water (100
145 mL), scaling sample and water weight on 14% sample moisture basis. The measured
146 indices were: gelatinization temperature (GT, °C; temperature at which an initial increase
147 in viscosity occurs); peak viscosity (PV, Brabender units, BU; maximum paste viscosity
148 achieved during heating), breakdown (BD, BU; viscosity decrease index while kept at 95
149 °C); final viscosity (FV, BU; paste viscosity at the end of the cooling), and setback (SB,
150 BU; index of the viscosity increase during cooling). The analysis was performed in
151 triplicate.

152 **2.4. Dough preparation**

153 The doughs (RF, RF+CF, RF+RCF and RF+DCF) were prepared by using the Brabender
154 Farinograph (Brabender OHG, Germany). In addition to RF or flour blends, the bread
155 dough formulation included HPMC (1.72%), sugar (2%), salt (1.5%), instant yeasts
156 (2.5%), sunflower oil (5.27%) and water. All these percentages were based on flour
157 weight. The amount of water added to each formulation was determined in order to
158 achieve a dough consistency of 125±5 BU (Kahraman, 2016). Dry components (rice flour
159 or blends, HPMC, instant yeast, sugar and salt) were added to the farinograph bowl (300
160 g capacity) and mixed for 1 min. Then, within 2 min, part of the water, vegetable oil and

161 the remaining water were added. The dough was eventually mixed for 8 min at 25 °C and
162 consistency was recorded.

163 **2.5. Dough evaluation**

164 **2.5.1. Leavening properties**

165 The leavening behavior of the dough samples was evaluated with a Chopin
166 Rheofermentometer F3 (Chopin, Villeneuve-La-Garenne, Cedex, France) according to a
167 method developed for gluten-free dough samples (Cappa et al., 2013). The proofing was
168 carried out at 30 °C for 60 min. Maximum and final heights (Hm and Hf, mm) of the
169 doughs, time necessary to reach maximum height (T1, min), time for dough porosity to
170 appear (Tx, min), total CO₂ production (CO_{2-TOT}, ml), CO₂ retention (CO_{2-RET}, ml),
171 released CO₂ (CO_{2-REL}, ml) and coefficient of retention (Rc, %) were measured.

172 As a parallel test to the rheofermentometric test, the leavening properties of dough
173 samples were measured by using image analysis with the method developed by Cappa et
174 al. (2013). The dough area increase (%) during proofing time was calculated. Six petri
175 dishes per each sampling time (every 10 min) were analyzed.

176 **2.5.3. Rheological properties**

177 The fundamental rheological behavior of the dough was studied by dynamic oscillatory
178 measurements performed on a Physica MCR300 Rheometer (Anton Paar, Graz, Austria).
179 The measurements were carried out with a corrugated parallel plate system (PP25/P2,
180 diameter: 25 mm) having a gap of 2 mm. The dough samples were prepared and rested
181 for 60 min at 25 °C before each measurement. The instant yeast was not included in the
182 formulations to avoid perturbation of the system. The dough was loaded between the
183 plates and the excess amount was trimmed off. In order to avoid moisture loss during

184 analysis, a humidity cover (H-PTD 150) having a water trap and wet pads was used, and
185 mineral oil was carefully applied to dough borders. After five minutes of resting to relax
186 stresses, the measurements were carried out and the data were recorded by using
187 Universal Software US200 (version 2.5) (Anton Paar, Ostfildern, Germany). The strain
188 sweep test was performed at a constant frequency of 1 Hz and in the range of 0.01-100%
189 strain to determine the maximum strain amplitude at which the viscoelastic properties
190 such as G' and G'' were independent of strain (linear viscoelastic region). According to
191 strain sweep tests, 0.04% strain was selected as the strain amplitude for all the doughs
192 analyzed. Frequency sweep tests were carried out in the range of 10 to 0.1 Hz at a
193 constant strain of 0.04%. For both tests, storage modulus (G' , Pa), loss modulus (G'' , Pa)
194 and damping factor ($\tan \delta$, the ratio of G'' to G') were calculated. For each formulation,
195 the analysis was performed on two separate doughs, each having at least two replications.

196 **2.6. Statistical evaluation**

197 Statistical evaluation of the data was performed by using MINITAB 16 (Minitab Inc.,
198 U.S.). The results were given as “mean \pm SD”. The significance of the data was tested by
199 analysis of variance (ANOVA) at $p < 0.05$ and, in the significant models, means were
200 compared by Tukey’s test at 95% confidence interval.

201 **3. Results and discussion**

202 **3.1. Flour and blend physicochemical properties**

203 **3.1.1. Proximate composition and particle size distribution**

204 The proximate composition of rice and chickpea flours is listed in Table 1. The chickpea
205 flours resulted richer in protein, fat and ash in comparison to RF. In particular, the protein
206 content of chickpea flours was approximately three times the RF amount thus making the

207 addition of chickpea flours into GF formulations an interesting strategy to increase the
208 protein content of GF products. The fat content was comparable to the values reported by
209 Alajaji & El-Adawy (2006) and Kaur, Singh, & Sodhi (2005). It must be emphasized that
210 chickpea contains a higher level of linoleic and oleic acid and polyunsaturated fatty acids
211 (PUFA) (Jukanti, Gaur, Gowda, & Chibbar, 2012) in comparison with other pulses.
212 Comparing the different chickpea treatments, since heat was applied during the roasting
213 process, RCF showed almost three times lower moisture content than CF. Also the
214 dehulling process resulted in a reduction of moisture, but to a lower extent than roasting.
215 Even if some modifications in carbohydrates and proteins were reported by Coşkuner &
216 Karababa (2004), roasting caused no statistical change in the amount of protein. No
217 significant changes in protein content, as well as in ash and fiber, during roasting
218 processing were previously observed by Sağlam (2006). the removal of hulls, rich in
219 minerals, caused a 25% reduction in ash as evidenced by Ghavidel & Prakash (2007).
220 Incorporating unconventional flours in baked products has to consider the particle size
221 distribution of the new ingredients as it may affect some properties such as the hydration
222 rate and the pasting behavior. The particle size distribution of the flours was quite
223 different and covered a wide range ($90 \geq x > 500 \mu\text{m}$). In general, the chickpea flours had a
224 larger particle size than RF. In fact, if the sample particle size is divided into two classes
225 (i.e., below or higher $250 \mu\text{m}$), the flours could be ranked as CF (24% fine particles) >
226 RCF (48% fine particles) > DCF (55% fine particles) > RF (85% fine particles). The
227 particle size distribution may be affected by the structure of the seed and the milling
228 process (Schober, 2009). Comparing the two samples milled on laboratory scale (CF and
229 RCF), the effect of the roasting process on the chickpea structure appears clear: due to the

230 expansion of air, gas cells are formed inside the seed. The presence of air gaps made the
231 seed very brittle and after milling, the resulting flour consisted of smaller particles.

232 **3.1.2. Microstructure**

233 The scanning electron micrographs of RF and chickpea flours can be seen in Figure 1.
234 According to micrograph, the granule sizes of starches ranged from 5 to 12 μm (average
235 size: 10 μm) for RF, from 13 to 30 μm (average size: 22 μm) for CF, from 14 to 26 μm
236 (average size: 19 μm) for RCF, and from 11 to 27 μm (average size: 19 μm) for DCF.
237 Previously, average granule sizes of 19-35 μm and 19-26 μm for raw and roasted pulse
238 flours were reported, respectively (Ma et al., 2011). As seen in Figure 1 rice starch
239 granules had a polyhedral shape and, although large granules ($\leq 12 \mu\text{m}$) were observed,
240 they were smaller and in aggregated form in comparison with chickpea granules.
241 Chickpea flours (CF, RCF, DCF), on the other hand, exhibited starch granules of
242 spherical shapes covered with protein fragments. For all samples, intact starch granules
243 were detected, although partial gelatinization of starch occurred in RCF, suggesting the
244 roasting and dehulling treatments do not affect starch granula organization so much.
245 Similarly, Köksel, Sivri, Scanlon, & Bushuk (1998) stated that starch was not completely
246 gelatinized during roasting due to limited kernel hydration.

247 **3.1.3. Water binding capacity and foaming properties**

248 The addition of CF, RCF, and DCF to RF caused a slight decrease in WBC (Figure 2),
249 which was significant ($p < 0.05$) only for DCF. In the literature it is reported that flour
250 having small particle size has high water binding capacity (Kim & Shin, 2014) and this
251 behavior has been related to the greater specific surface area of the flour. However, DCF,
252 the chickpea flour with the smallest particle size, had the lowest water binding capacity.

253 This contradictory finding can be attributed to husk removal during the dehulling process
254 and thus to the different sample composition (Table 1). Since husks are sources of non-
255 starch polysaccharides and proteins, their removal might have modified flour water
256 binding capacity and dough rheology (Witczak, Ziobro, Juszczak, & Korus, 2015).
257 As in GF dough production, the absence of gluten penalizes dough structuring,
258 hydrocolloids and proteins are often used for their property of binding water, and
259 foaming capacity. The foaming capacity and stability of RF and chickpea blends are
260 reported in Figure 2. Increased foam formation was observed in chickpea-containing
261 blends ($p < 0.05$). However, RF+RCF showed significantly lower FC compared to CF and
262 DCF containing blends. The reduced foaming capacity of roasted chickpeas in
263 comparison to the raw chickpea flour was previously reported by Ma et al. (2011) and
264 related to the lower solubility of proteins as a result of the heat treatment.
265 As regards foam stability, rice flour exhibited no detectable foam after 60 min at room
266 temperature; whereas the replacement of 24.85% of RF with chickpea flours highly
267 improved foam stability. These findings are promising as the chickpea flours here are
268 suggested as ingredients for GF bread dough in which a high capacity to retain leavening
269 gas during baking is desirable. Of all the chickpea-containing samples, RF+RCF
270 exhibited the lowest foam stability. Conversely, Ma et al. (2011) did not find any
271 difference between raw and roasted chickpea flours in terms of foam stability. This can
272 be related to the procedure of the roasting process; in this study, roasting was directly
273 applied to whole chickpea seeds and a heating and a roasting process was applied in
274 sequence instead of one stage of roasting, as carried out on flour by Ma et al. (2011).

275 **3.1.4. Pasting properties**

276 The pasting curves of RF and blends (*data not shown*) were characterized by an increase
277 in viscosity upon heating, since starch granules started to uptake water and swelled, and
278 then a paste was obtained. During the holding period at 95 °C the starch gel viscosity
279 decreased due to the shearing force applied. With cooling, the reordering of starch
280 molecules caused a new increase in viscosity. This trend was displayed by all samples but
281 to different extents. The pasting curve indices are reported in Table 2. It is well known
282 that the pasting properties are mainly affected by the quantity and quality of starch and by
283 flour particle size (Kim & Shin, 2014). All blends, in comparison to RF, exhibited lower
284 peak, final viscosity, breakdown and setback values. This behavior is mainly due to the
285 composition of chickpea flours and it indicates that the addition of chickpea flour, besides
286 producing a lower strength gel, may slow down paste retrogradation (i.e., low SB) and
287 thus limit the staling of baked food.

288 Of the three chickpea-containing blends, the maximum viscosity reached during the
289 heating period (PV) and the final viscosity were lower for RF+RCF, which indicates that
290 roasting had a slight effect on starch gelatinization in accordance with the scanning
291 images. It is to note that roasting was performed with the addition of a small amount of
292 water and thus the heat treatment did not exhibit a relevant effect on pasting behavior.

293 **3.2. Dough properties**

294 In order to reach the desired (125 ± 5 BU; Kahraman, 2016) dough consistency, and in
295 accordance with the WBC data, RF+ DCF required less water in comparison to the other
296 dough (90.07% vs. 101.14%, 99.47%, 104.70%, for RF+DCF, RF, RF+CF, RF+RCF,
297 respectively). These differences are related to flour composition and to their ability to
298 adsorb water, as previously discussed.

299 **3.2.1. Dough proofing properties**

300 The best leaving performance in terms of maximum and final dough height was
301 evidenced by RF dough (Table 3). This is in accordance with the general findings that
302 flour ingredients containing fiber – such as chickpea flours – resulted in weak dough
303 structure. However, the CO₂ retention capacity of RF+RCF (99.0%) was higher than
304 RF+CF (97.9%) and RF+DCF (98.1%), and slightly lower than RF (99.4%). This
305 suggests a slight weakening of the dough containing chickpea flours, which were in any
306 case able to retain the majority of the CO₂ produced by the yeast. The advantages of
307 using chickpea flours can be seen in terms of leavening time before the dough porosity
308 appearance (Tx) and total CO₂ produced. In fact, for the three formulations containing
309 chickpea flours, the production of CO₂ was 5-12% higher than in RF. This can be due to
310 the faster action of the yeast in these samples, presumably due to a higher amount of
311 sugars (10.85 g sugar/100 g chickpea flour; 0.12 g sugar/100 g rice flour) (USDA, 2016).
312 Accordingly, RF+CF and RF+DCF were also characterized by an earlier appearance of
313 Tx, thus a shorter leavening process is recommended for their baking.

314 Dough leavening behavior was also monitored by means of image analysis. This
315 technique has been proposed as an alternative tool to the Rheofermentometer by
316 evaluating dough development as an increase in dough area (Cappa et al., 2013). All
317 doughs were able to increase their area (up to a maximum value of 114% after 60 min of
318 proofing) and for each formulation, the dough area increase during leavening was highly
319 correlated to the values of dough height obtained from the Rheofermentometer test
320 ($R^2 \geq 0.989$), suggesting that both tests can be used to study dough development.

321 **3.2.2. Dough rheology**

322 Even if the deformation applied during the dynamic test are often very different from
323 those experienced by the dough during real processing (i.e., mixing, leavening, baking),
324 these measurements provide unique information about the viscoelastic characteristic of
325 the dough by preserving the dough structure during the test; thus, the viscoelastic
326 properties measured can be used to compare different dough formulations. The strain
327 sweep test was firstly performed to delineate the region of linear viscosity in order to
328 define when dough characteristics do not depend on the magnitude of the deforming
329 strain. For all the samples, G' and G'' remained almost constant at least up to 0.04% strain
330 (*data not shown*). Beyond this limit, the storage and loss moduli decrease, indicating a
331 progressive destruction of the dough structure. Similar limits of linear viscoelasticity
332 were found in the literature for GF doughs (Mariotti et al., 2009).

333 According to the region of linear viscosity, the frequency sweep test was performed at a
334 constant strain of 0.04%, in the range of 10 to 0.1 Hz. Frequency sweep curves are
335 reported in Figure 3. Although all the dough samples had the same final farinographic
336 consistency (125 ± 5 BU), they differed in terms of fundamental rheological properties.
337 According to Mariotti et al., 2009, no relationships were found between the different
338 water levels in the GF doughs and the values of their respective dynamic moduli. For all
339 dough formulations, G' values were higher than G'' to indicate a solid-like behavior. This
340 behavior is in agreement with the literature regarding the rheology of GF batters (Hüttner,
341 Bello, & Arendt, 2010; Mariotti et al., 2009; Sciarini, Ribotta, León, & Pérez, 2012) and
342 gels (Cappa et al., 2016). The addition of chickpea flours to rice dough formulations
343 caused an increase in both G' and G'' . In particular, CF and DCF showed the highest
344 values for all frequencies investigated and RCF had an intermediate behavior in

345 comparison to RF. A similar increase in G' and G'' was previously observed after the
346 addition of chickpea flour (Aguilar et al., 2015) in starch-based gluten-free dough. The
347 damping factor was lower than 1 at all frequencies indicating the prevalence of a solid-
348 like behavior and was slightly affected by the addition of chickpea flour as it remained
349 constant for all formulations; in particular values of 0.41 ± 0.02 , 0.41 ± 0.02 , 0.42 ± 0.02 ,
350 0.37 ± 0.01 for RF, RCF+RF, CF+RCF and RF+DCF respectively, were obtained at 1 Hz.

351 **4. Conclusions**

352 The nutritional quality of food products is drawing considerable interest due to the
353 increasing awareness of healthy diet. The use of pulse flours in food formulations is
354 becoming an interesting strategy due to the important nutritional benefits of pulses. In
355 this study, the effects of roasted, dehulled and raw chickpea flour when added (25%) to
356 rice dough formulations were evaluated. Besides increasing protein and fat content, the
357 addition of all types of chickpea flours created positive effects on the technological
358 performance of the doughs. Although the dough development of the formulations
359 containing chickpea flours was slightly lower than the reference samples, high CO_2
360 retention was evidenced and shorter leavening times were necessary to obtain maximum
361 dough development. Also the viscoelastic properties of the dough were positively
362 affected; higher storage moduli were obtained for the samples containing raw and
363 dehulled chickpea flours. Furthermore, the viscoamylographic test indicated a slower
364 retrogradation tendency of the slurry containing chickpea flours, which is a promising
365 result for baking food applications. This study showed the potential of using raw,
366 dehulled and roasted chickpea flour in combination with rice flour in gluten-free bread
367 formulations.

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376 respectively.

377 **Declaration of conflict of interest**

378 The Authors declare that there is no conflict of interest.

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Table 1

Table 1. Proximate composition of rice and chickpea flours

Sample	Moisture (%)	Protein (% db)	Fat (% db)	Ash (% db)
RF	11.85 ± 0.09 ^a	8.29 ± 0.18 ^c	1.28 ± 0.19 ^c	0.68 ± 0.00 ^b
CF	9.15 ± 0.06 ^b	23.52 ± 0.30 ^a	5.71 ± 0.24 ^b	3.09 ± 0.01 ^a
RCF	3.25 ± 0.06 ^d	23.10 ± 0.79 ^a	7.57 ± 0.08 ^a	2.68 ± 0.03 ^a
DCF	7.12 ± 0.00 ^c	21.15 ± 0.41 ^b	7.55 ± 0.05 ^a	2.32 ± 0.77 ^a

RF, rice flour; CF, chickpea flour; RCF, roasted chickpea flour; DCF, dehulled chickpea flour. Values are mean ± SD.

Means having different letters in the same column are significantly different (p<0.05).

Table 2. Pasting properties

Sample	GT (°C)	PV (BU)	BD (BU)	SB (BU)	FV (BU)
RF	77.15 ± 0.07 ^{bc}	716.50 ± 7.78 ^a	326.50 ± 12.02 ^a	478.50 ± 9.19 ^a	868.50 ± 13.44 ^a
RF+CF	76.80 ± 0.14 ^{ab}	503.50 ± 2.12 ^b	196.50 ± 6.36 ^b	410.00 ± 4.24 ^b	717.00 ± 0.01 ^b
RF+RCF	77.60 ± 0.01 ^a	435.50 ± 12.02 ^c	161.50 ± 9.19 ^c	404.00 ± 7.07 ^b	678.00 ± 9.90 ^c
RF+DCF	77.40 ± 0.14 ^c	498.00 ± 5.66 ^b	164.00 ± 4.24 ^{bc}	415.00 ± 5.66 ^b	749.00 ± 4.24 ^b

RF, rice flour; CF, chickpea flour; RCF, roasted chickpea flour; DCF, dehulled chickpea flour; GT, gelatinization temperature; PV, peak viscosity; BD, breakdown; SB, setback; FV, final viscosity. Values are mean ± SD. Means having different letters in the same column are significantly different (p<0.05).

Table 3

Table 3. Dough proofing properties

Sample	Hm (mm)	Hf (mm)	Tx (min)	CO ₂ -TOT (mL)	CO ₂ -REL (mL)	CO ₂ -RET (mL)	Rc (%)
RF	49.2	49.2	>60	783	5	779	99.4
RF+CF	41.3	36.9	40.5	875	18	857	97.9
RF+RCF	41.7	35.8	>60	821	8	812	99.0
RF+DCF	43.2	41.0	45.0	879	17	862	98.1

RF, rice flour; CF, chickpea flour; RCF, roasted chickpea flour; DCF, dehulled chickpea flour; Hm, dough maximum height; Hf, dough final height; Tx, time of dough porosity appearance; CO₂-TOT, total gas production; CO₂-REL, CO₂ released by the dough; CO₂-RET, CO₂ retained by the dough; Rc, gas retention coefficient.

Figure 1
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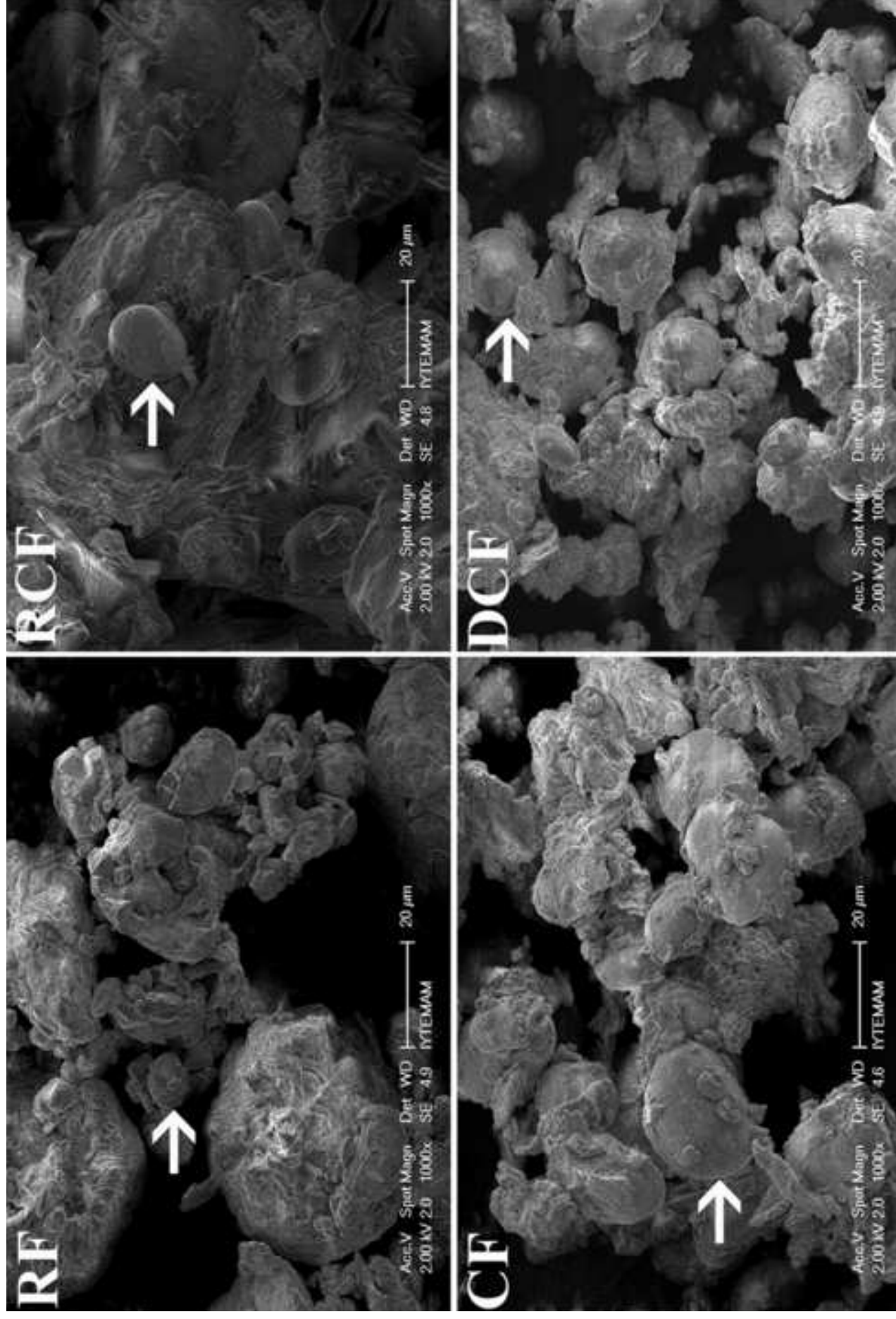


Figure 2
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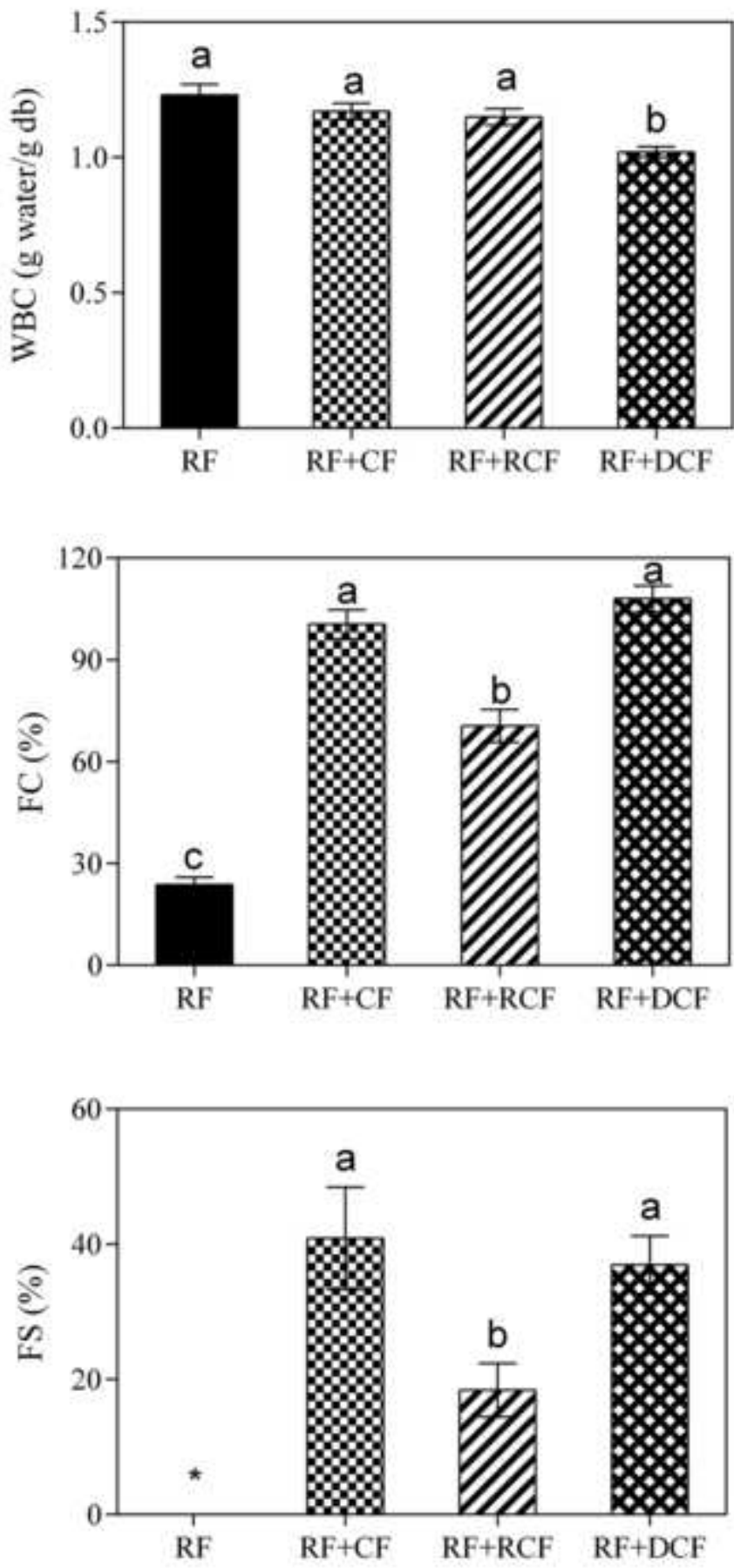


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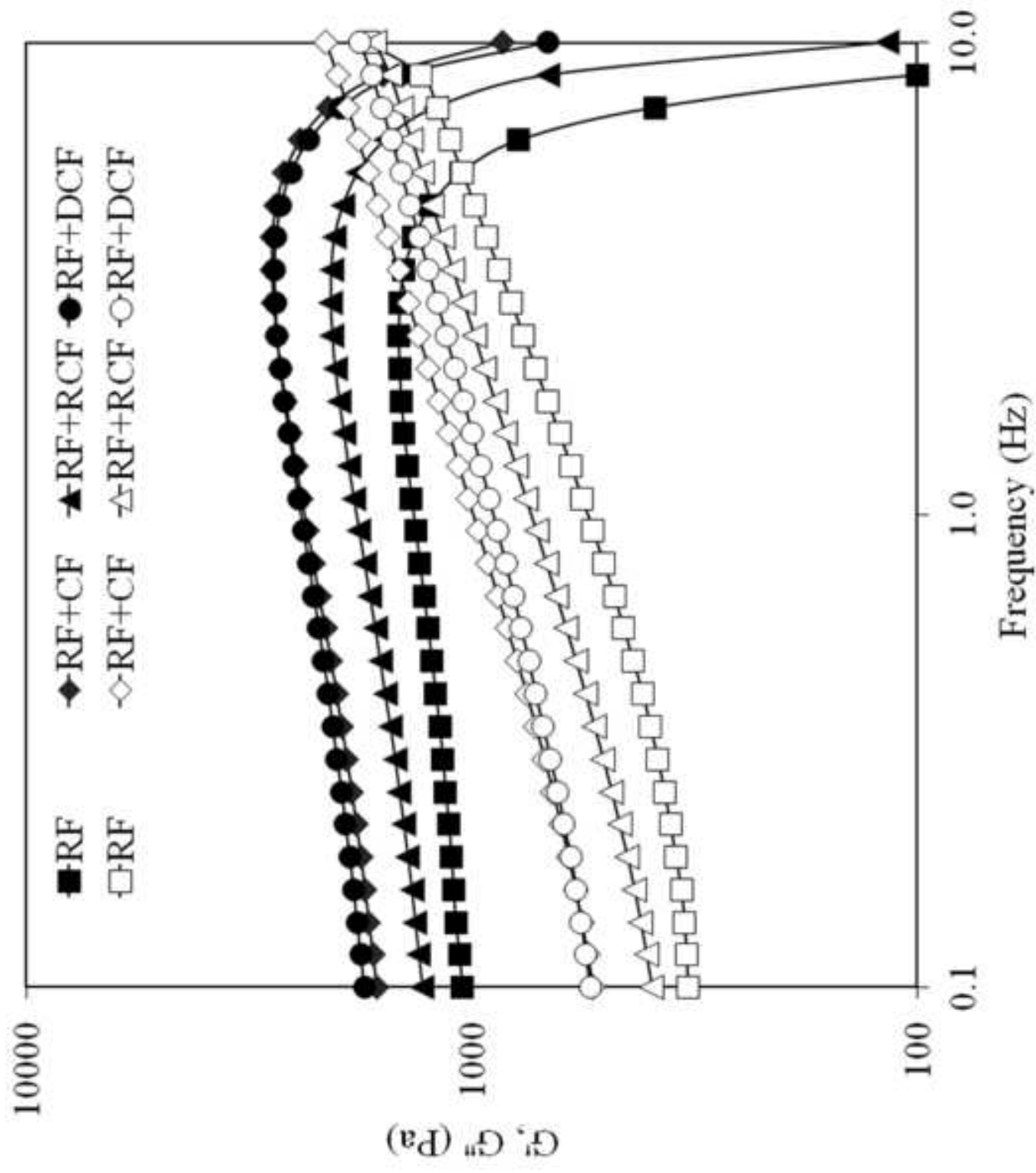


FIGURE CAPTIONS

Figure 1. SEM images of flour samples (x1000). The starch granules were shown with the arrow. RF, rice flour; CF, chickpea flour; RCF, roasted chickpea flour; DCF, dehulled chickpea flour.

Figure 2. Water binding capacities (WBC), foaming capacity (FC) and foam stability (FS) of RF and blends. RF, rice flour; CF, chickpea flour; RCF, roasted chickpea flour; DCF, dehulled chickpea flour; * no detectable foam after 60 min. Means having different letters are significantly different ($p < 0.05$).

Figure 3. Dough viscoelastic properties: frequency sweep tests. RF, rice flour; CF, chickpea flour; RCF, roasted chickpea flour; DCF, dehulled chickpea flour. Storage modulus (G' , dark), loss modulus (G'' , white), RF (■), RF+CF (◆), RF+RCF (▲) and RF+DCF (●).