

# Pooled Analysis of Clinical Outcome of Patients with Chemorefractory Metastatic Colorectal Cancer Treated within Phase I/II Clinical Studies Based on Individual Biomarkers of Susceptibility: A Single-Institution Experience

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## Abstract

**Background** Patients with metastatic colorectal cancer (mCRC) refractory to standard therapies have a poor prognosis. In this setting, recruitment into clinical trials is warranted, and studies driven by selection according to individual tumor molecular characteristics are expected to provide added value. **Objective** We retrospectively analyzed data from patients with mCRC refractory to or following failure of standard therapies who were enrolled into phase I/II clinical studies at the Niguarda Cancer Center based on the presence of a specific molecular profile expected to represent the target of susceptibility to the experimental drug(s).

**Patients and Methods** From June 2011 to May 2016, 2044 patients with mCRC underwent molecular screening. Eighty patients (3.9%) were enrolled in ad hoc studies; the median age was 60 years (range 36–86) and the median number of previous treatment lines was five (range 2–8). Molecular char-

acteristics exploited within these studies were *MGMT* promoter hypermethylation (48.7%), *HER2* amplification (28.8%), *BRAF*<sup>V600E</sup> mutation (20%), and novel gene fusions involving *ALK* or *NTRK* (2.5%).

**Results** One patient (1%) had RECIST (Response Evaluation Criteria In Solid Tumors) complete response (CR), 13 patients (16.5%) experienced a partial response (PR), and 28 (35%) stable disease (SD). Median progression-free survival (PFS) was 2.8 months (range 2.63–3.83), with 24% of patients displaying PFS >5 months. Median growth modulation index (GMI) was 0.85 (range 0–15.61) and 32.5% of patients had GMI >1.33. *KRAS* exon 2 mutations were found in 38.5% of patients, and among the 78 patients with known *KRAS* status, those with wild-type tumors had longer PFS than those with mutated tumors (3.80 [95% CI 2.80–5.03] vs. 2.13 months [95% CI 1.77–2.87], respectively,  $p = 0.001$ ). Median overall survival (OS) was 7.83 months (range 7.17–9.33) for all patients, and patients with *KRAS* wild-type tumors had longer OS than those with mutated tumors (7.83 [95% CI 7.33–10.80] vs. 7.18 months [95% CI 5.63–9.33], respectively,  $p = 0.06$ ).

**Conclusions** This single-institution retrospective study indicates that in a heavily pretreated population approximately 4% of mCRC tumors display a potential actionable molecular context suitable for therapeutic intervention. Application of molecular selection is challenging but improves clinical outcome even in later lines of treatment.

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## Key Points

Treatment of metastatic colorectal cancer (mCRC) patients driven by selection according to individual tumor molecular characteristics is expected to provide enhanced clinical benefit.

In this single-institution retrospective analysis, 3.9% of 2044 patients were found to harbor biomarkers for *ad hoc* phase I-II clinical studies, including *MGMT* promoter hypermethylation, *HER2* amplification, *BRAF*<sup>V600E</sup> mutation, and gene fusions involving *ALK* or *NTRK*.

14 patients had an objective response, and 28 stable disease. Median progression-free survival (PFS) was 2.8 months, with 24% of patients displaying PFS >5 months.

## 1 Introduction

Colorectal cancer (CRC) remains a significant cause of morbidity and mortality worldwide and many patients present with metastatic disease despite large-scale screening programs in various countries [1]. With the only exception of patients presenting with oligometastatic lesions confined to the liver or lung amenable to resection [2, 3], metastatic disease is considered incurable. In cases where treatment with curative intent is excluded, patients are typically given a combination of cytotoxic chemotherapy, often in conjunction with a targeted therapy.

Although advances in systemic therapy have been made, the 5-year survival rate in this setting is still 12.5% [4], with acquired resistance to therapy being the main reason for treatment failure [5]. Indeed, resistance to targeted therapy, as displayed by disease progression, is observed after a median of 4 months on epidermal growth factor receptor (EGFR) antagonists [45]. The discovery of *RAS* mutations in CRC as a mechanism of innate resistance to these therapies has been an important advance and has ameliorated their clinical use. However, there is an unmet need for effective therapeutic strategies after secondary resistance.

We have previously demonstrated that different molecular alterations that drive resistance can occur simultaneously in the same patient [7]. Identifying relevant molecular subtypes within this heterogeneous disease and matching patients with appropriate single agents or combinations of targeted therapies at resistance is crucial to therapeutic progress [8]. Therefore, recruitment into precision oncology clinical trials

based on selection according to individual tumor molecular characteristics is expected to provide added value.

We retrospectively collected data from patients with metastatic CRC (mCRC) resistant to standard therapies treated at the Niguarda Cancer Center (NCC) (Milan, Italy) in phase I/II clinical studies based on the presence of specific tumor molecular profiles conferring susceptibility to experimental drugs, and performed a pooled analysis for measuring results according to main clinical and other molecular variables.

## 2 Methods

### 2.1 Patients

We retrospectively collected data from patients with mCRC resistant to standard therapies treated at NCC between June 2011 and May 2016 in phase I/II clinical studies, including one phase I first-in-human study, based on the presence of specific biomarkers that confer susceptibility to experimental drugs (Table 1). These included tumor genetic alterations (i.e., gene mutations, amplifications, or fusions) or a certain genetic context (i.e., methylation of specific genes). Consecutive eligible patients were offered participation in clinical trials. All patients gave written informed consent and the study and all treatments were conducted in accordance with the guidelines of the Institutional Review Board at Ospedale Niguarda.

The presence of the particular biomarker was investigated according to specific study protocol criteria or retrieved by medical history, where applicable. Further molecular characterization of Kirsten rat sarcoma viral oncogene homolog (*KRAS*) mutations, a well-known biomarker associated with poor prognosis, was performed in all available samples of patients included in the studies by the Pathology Department of NCC as per standard of care. The registration of patients in the database, pathology assessment, and additional mutation analyses if needed were performed at NCC.

### 2.2 Treatment and Evaluation

Treatment was carried out according to the specific requisites of treatment protocols and was continued until disease progression, unacceptable toxicity, or withdrawal of consent. Assessments, including history, physical examination, and laboratory evaluations, were performed as specified in each protocol, typically before the initiation of therapy, and longitudinally during the treatment cycles. Efficacy was assessed from computed tomography scans and/or magnetic resonance imaging at baseline before treatment initiation and then every 6–8 weeks depending on the study protocol. All radiographs were evaluated by the Department of Radiology at NCC and responses were categorized per RECIST (Response Evaluation Criteria In Solid Tumors) 1.1 criteria and were

**Table 1** Distribution of patients in clinical trials with actionable molecular alterations treated with matched targeted agents included in the pooled analysis

Biomarker	Study drug(s)	EudraCT N°	Patients (n)
<i>MGMT</i> promoter hypermethylation	Temozolomide [9]	2012–003338-17	27
<i>HER2</i> amplification	Trastuzumab + lapatinib [10]	2012–002128-33	23
<i>MGMT</i> promoter hypermethylation	Dacarbazine [11]	2011–002080-21	12
<i>BRAF</i> mutation	MEK162 + LGX818 [NCT01543698]	2011–005875-17	9
<i>BRAF</i> mutation	MEK162 + panitumumab [NCT01927341]	2013–001986-18	7
<i>NTRK</i> or <i>ALK</i> gene fusions	Entrectinib [12]	2012–000148-88	2

reported as best response [13]. Progression-free survival of the treatment (PFS<sub>n</sub>) was recorded for each individual patient as in the case report form of the respective trial, whereas progression-free survival achieved with the previous treatment line received by the patient (PFS<sub>n-1</sub>) was calculated based on available clinical documentation.

### 2.3 Statistical Analysis

Fisher's exact test was used to assess the association among categorical variables and the presence of the biomarker used for treatment selection. Progression-free survival (PFS) was defined as the time interval from the start of therapy to the first observation of disease progression or death, whichever occurred first. All tests were 2-sided, and  $p < 0.05$  was considered statistically significant. Multivariate analysis was performed by Cox logistic regression; the model as a whole was evaluated by likelihood ratio test, whereas the significance of the single independent variables was evaluated by means of the Wald test. Results of the multivariate analysis should be interpreted with caution due to the small sample size and sampling bias of our study.

PFS and overall survival (OS) for all patients were computed using the Kaplan-Meier method and differences in PFS and OS were calculated with log-rank test where applicable. The growth modulation index (GMI) has been applied as previously described [14]. We consider a GMI >1.33, i.e., an increase in the PFS<sub>n</sub>/PFS<sub>n-1</sub> ratio of 30%, as clinically meaningful. All statistical analyses were carried out using R software [15] and graphics were generated with ggplot2 [16].

## 3 Results

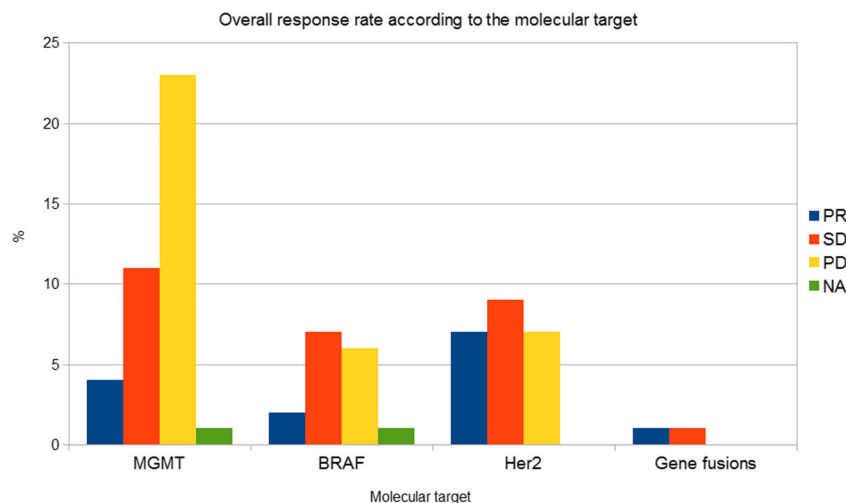
From June 2011 to May 2016, 2044 patients with mCRC were referred to NCC for molecular screening for the inclusion into phase I/II trials encompassing the targeting

of actionable molecular alterations or molecular contexts of susceptibility. In total, 80 patients (3.9%) were enrolled into six ad hoc studies: one phase I/II and five phase II trials.

The median age was 60 years (range 36–86) and the median number of previous treatment lines was five (range 2–8). The six studies included therapies based on *MGMT* promoter hypermethylation (48.7%), *HER2* amplification (28.8%), *BRAF*<sup>V600E</sup> mutation (20%), and gene fusions involving *ALK* or *TRKA* (2.5%) (Table 1 and Fig. 1). Among the 78 of 80 patients evaluable for *KRAS* mutations, any *KRAS* (exon 2) mutation was found in 30 (38.5%) of patients.

According to RECIST 1.1 criteria, one patient (1%) had complete response (CR), 13 patients (16.5%) had partial response (PR), and 28 (35%) had stable disease (SD), accounting for a 52.5% disease control rate (DCR = CR + PR + SD). The DCR was higher in patients with *KRAS* wild-type tumors (66 vs. 38%,  $p = 0.02$ ). The median PFS was 2.8 months (range 2.63–3.83) (Fig. 2a), and 24% of patients displayed a PFS >5 months. A multivariate analysis of clinicopathological factors including age, gender, performance status, carcinoembryonic antigen (CEA) levels, and *KRAS* status showed that *KRAS* status and age were significantly associated with PFS ( $p = 0.011$  and 0.047, respectively), whereas CEA and age were significantly associated with OS ( $p = 0.0063$  and 0.0279, respectively). Patients with *KRAS* wild-type tumors had longer PFS than those with mutated tumors (3.80 [95% CI 2.80–5.03] vs. 2.13 months [95% CI 1.77–2.87], respectively) ( $p = 0.001$ ) (Fig. 2b). Median OS was 7.83 months (range 7.17–9.33) (Fig. 3a), and patients with *KRAS* wild-type tumors had longer OS than those with mutated tumors (7.83 [95% CI 7.33–10.80] vs. 7.18 months [95% CI 5.63–9.33], respectively,  $p = 0.06$ ) (Fig. 3b). The median GMI was 0.85 (range 0–15.61) and 32.5% of patients had a GMI >1.33 (Fig. 4). The Spearman Rho between PFS<sub>n</sub> and PFS<sub>n-1</sub> was 0.10 ( $p = 0.372$ ).

**Fig. 1** RECIST (Response Evaluation Criteria In Solid Tumors) objective response rates according to molecular targets in the pooled patient population. *PR* partial response, *SD* stable disease, *PD* progressive disease, *NA* not assessed



## 4 Discussion

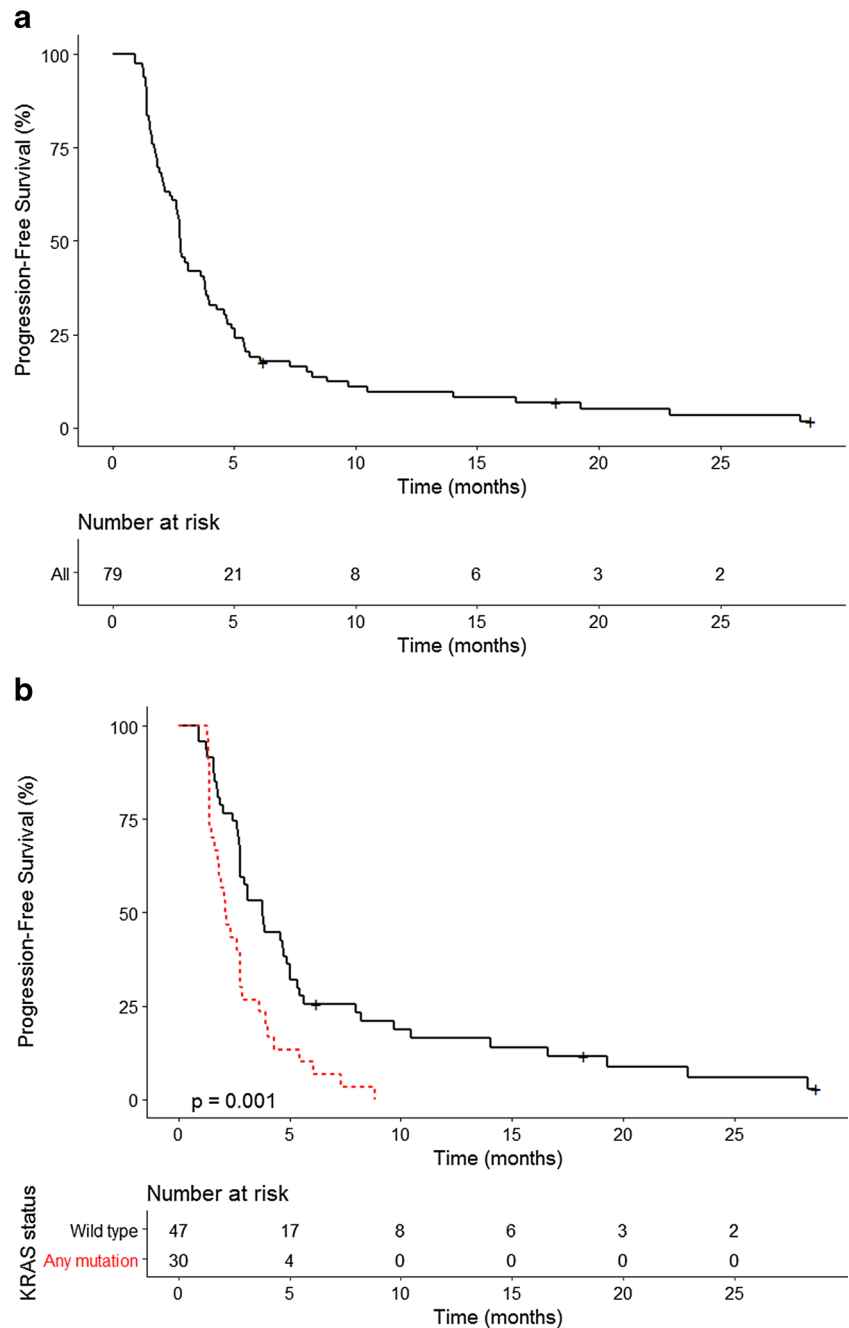
In this single-institution retrospective analysis we found that 4.0% of mCRC patients refractory to standard therapies were eligible for enrollment into phase I/II clinical trials based on molecular target selection. These patients, who had experienced a median number of five lines of previous treatments, achieved a median PFS of 2.8 months (with approximately one-quarter of patients displaying a PFS >5 months) and a median OS of 7.8 months. These data should be evaluated in the context of advanced mCRC, where the median PFS with newer agents has been less than 2 months [20], and rechallenge with standard chemotherapy [18, 19] or further chemotherapy [17] showed only poor efficacy and was burdened by adverse effects [21]. With regards to objective response rates (ORRs), 14 patients (17.5%) had PR and 28 (35%) SD, accounting for a 52.5% DCR. These data compare favorably with those achieved with licensed third-line therapies such as the multikinase inhibitor regorafenib (ORR = 1–4%) and the trifluridine–tipiracil combination TAS-102 (ORR = 2%), both of which are administered without the selection of a molecular target, or salvage chemotherapy with mitomycin-C or oxaliplatin rechallenge [17, 22, 23].

Following the seminal work by von Hoff et al. [14], several reports investigating the use of molecularly targeted agents on the basis of the identification of molecular alterations provided conflicting results [43, 44]. Provocative findings from Janku et al. highlighted that in phase I studies the lack of matching between molecular alterations and targeted treatment provides only limited clinical benefit in CRC and other solid tumors [24]. Recently, the same authors reported findings of a prospective, single-center ‘navigation’ study conducted in 500 patients with several different refractory cancers who underwent comprehensive genomic profiling by sequencing of 236 genes, offering proof-of-concept for the utility of this approach in assigning therapy to patients with refractory malignancies, especially in those patients with multiple genomic

aberrations for whom combination therapies could be implemented [25]. However, patients with CRC were only a small proportion of this cohort and therefore definitive conclusions could not be drawn for this histology. In mCRC, Dienstmann et al. reported a series of 68 patients treated with targeted agents in phase I trials based on molecular profiling, concluding that this approach did not confer a significant clinical benefit on the basis of a comparison between the median time to treatment failure  $n$  ( $TTF_n$ ) of 7.9 weeks and  $TTF_{n-1}$  of 16.3 weeks [26]. Similarly, in our present analysis of 80 patients, we observed that  $PFS_n$  was inferior to  $PFS_{n-1}$  (12.0 vs. 15.6 weeks). However, there are substantial differences between these two cohorts. Our series included only 2.5% patients treated within a phase I study versus 100% of the patients included in the study by Dienstmann et al. [26], and there are differences in the molecular alterations on which the treatments are based (*KRAS/BRAF/PIK3CA* mutations, *PTEN* and *MET* expression vs. *HER2*, *BRAF*, *MGMT*, and gene fusions, respectively). Assessment of clinical benefit by means of the ratio between PFS with the actual targeted therapy and that achieved with the previous line of treatment, referred to as the GMI, has also been proposed [14]. When we applied this index to our cohort, we found that at least one-third of patients benefited from the enrollment in clinical trials with targeted agents matched to selected actionable molecular alterations, making our results meaningful.

Among the molecular abnormalities selected in our cohort, we included gene fusions involving the *ALK* and *NTRK* genes. These alterations have only recently been recognized as targets occurring at low prevalence in CRC [27]. In particular, in 2014 we reported the characterization of the *TPM3-NTRK1* gene rearrangement as a recurring, although rare, event in CRC. The concomitant discovery of entrectinib (NMS-P626; RXDX-101) as a novel highly potent and selective pan-Trk inhibitor by the group of Ardini et al. [28] enhanced the interest in this target, with further development across different

**Fig. 2** Progression-free survival **a** for all patients; and **b** according to *KRAS* status

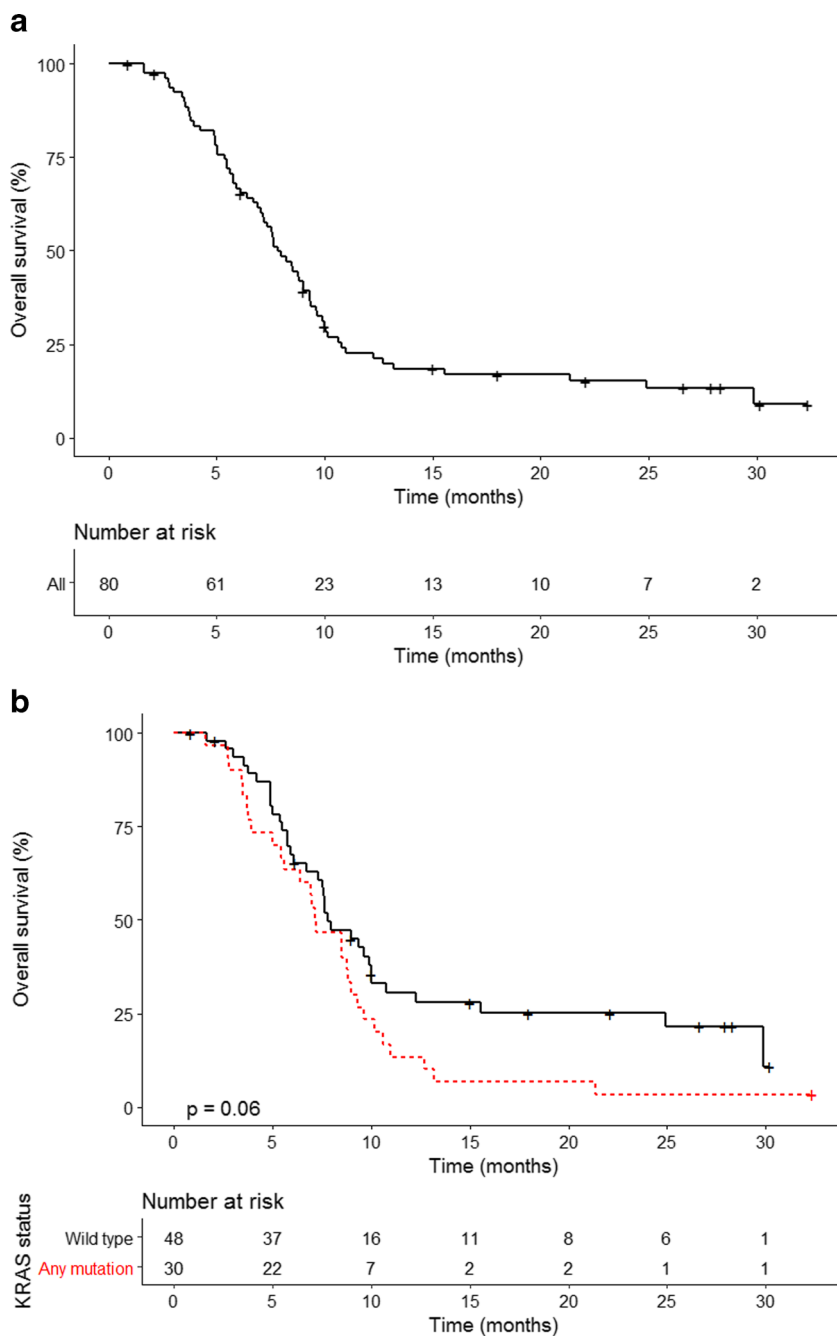


histologies and currently ongoing phase II studies, including in CRC [12, 27]. Interestingly enough, the two gene fusions involving *NTRK* and *ALK* described in the present cohort were previously unreported [29, 30], highlighting the challenge of a comprehensive molecular pathology that should not merely include gene fusions together with mutations and amplifications but also elucidate the exact fusion partner and oncogenic potential for treatment with specific inhibitors.

During the last decade, the assessment of *KRAS* and neuroblastoma RAS viral oncogene homolog (*NRAS*) gained increasing importance for an optimal management of mCRC

[31]. Mutations in *KRAS* codons 12 or 13 (in exon 2) affect approximately 40% of patients with metastatic CRC [32]. About an additional 5% of CRC patients display mutations in *KRAS* exons 3 or 4, usually at codon 61 or 146, and another 5% of CRC patients have mutations in *NRAS* in exons 2, 3, or 4. Extended *RAS* status (*KRAS* and *NRAS* exons 2, 3, and 4 mutations) is the validated biomarker of response to anti-EGFR antibodies [31]. *RAS* mutant mCRC exhibit a significantly higher cumulative incidence of lung, bone, and brain metastasis [33–35]. The prognostic role of *RAS* mutations is controversial, but increasing evidence shows a negative effect

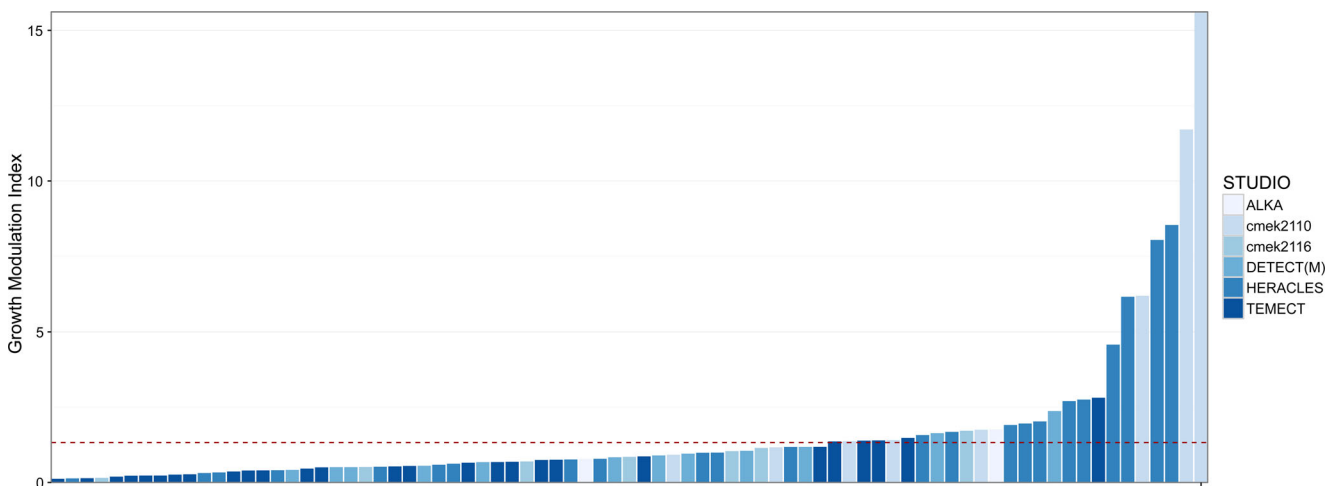
**Fig. 3** Overall survival **a** for all patients; and **b** according to *KRAS* status



in the adjuvant and metastatic setting [36–38]. In our cohort, *KRAS* mutations showed an overall negative prognostic value, since patients harboring *KRAS*-mutated tumors had shorter PFS and OS. Also, DCR was more frequent in patients with *KRAS* wild-type tumors. However, it should be taken into account that half of the selected phase I/II trials were designed for patients with *KRAS* wild-type tumors (Cmek2110 and Cmek2116 for BRAF-mutated tumors, HER2 Amplification for Colo-rectal Cancer Enhanced Stratification-HERACLES

for HER2 amplified tumors) and most of the *KRAS*-mutated patients were treated in the two studies (DETECT and TEMECT with dacarbazine and temozolomide, respectively) that were not based on directly targeting an oncogenic driver but rather exploiting a molecular context of susceptibility.

Limitations of our study include the small sample size and potential sampling bias, the notion that the use of GMI as an index of clinical benefit in CRC has been criticized as there is a lack of correlation between PFS<sub>n</sub> and PFS<sub>n-1</sub> that is the



**Fig. 4** Growth modulation index for each patient included in the pooled analysis and corresponding phase I/II clinical trials. *M\** subset of patients within the study with hypermethylation of *MGMT*

statistical assumption for applying this method [39, 40], and that *KRAS*-limited instead of extended *RAS* analysis was performed in our cohort.

In conclusion, our single-institution study indicates that in a heavily pretreated mCRC population about 4% of tumors display a potential actionable molecular context suitable for phase I/II trials with matched therapeutics. Comprehensive molecular selection is challenging, encompassing inclusion of actionable alterations occurring at low frequency in CRC such as *HER2* amplification [10] and rare gene fusions involving the *ALK* and *NTRK* genes [41], but enhances therapeutic options to be exploited in the advanced setting. This knowledge, coupled with recent advancements in the understanding of acquired resistance mechanisms to targeted agents through liquid biopsy [8, 42] is expanding opportunities of precision oncology therapies for CRC.

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**Author Contributions** AS-B provided patients' care, collected patients' samples, supervised data collection and analysis, and was the major contributor in writing the manuscript. AA provided patients' care, participated in data collection, statistical analysis, and writing the manuscript. EB provided patients' care and participated in data collection and writing the manuscript. SSt performed data collection and participated in collection of patients' samples. LG, GC, IS, KB, CF, RR, TC, SDB, MS, VG, LP, GC-S, FR, GB and ET provided patients' care. FP, SG, AC and AG participated in data and patients' samples collection. GM supervised data and patients' samples collection. SV, EV, CL and MT performed diagnostic pathology and molecular analysis on tumor tissue specimens as per clinical studies. AV supervised local radiological assessment of patients as per clinical studies. SSi supervised patients' care and participated in data analysis and writing of the manuscript. All authors read and approved the final manuscript.

### Compliance with Ethical Standards

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**Conflicts of Interest** The authors declare no conflicts of interest.

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