ADOPTED: 26 January 2016 doi:10.2903/j.efsa.2016.4390



PUBLISHED: 11 February 2016

Safety and efficacy of pyridine and pyrrole derivatives belonging to chemical group 28 when used as flavourings for all animal species

EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP)

Abstract

Following a request from the European Commission, the EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP) was asked to deliver a scientific opinion on the safety and efficacy of nine compounds belonging to chemical group 28 (pyridine, pyrrole and quinoline derivatives). They are currently authorised as flavours in food. The FEEDAP Panel concludes that piperine, 3-methylindole, indole, 2-acetylpyridine and 2-acetylpyrrole are safe at the proposed maximum use level of 0.5 mg/kg complete feed for all animal species; trimethyloxazole, 3-ethylpyridine, pyrrolidine and 2,6-dimethylpyridine are safe at the proposed use level of 0.5 mg/kg complete feed for cattle, salmonids and non-food-producing animals, and at the use level of 0.3 mg/kg complete feed for pigs and poultry. No safety concern would arise for the consumer from the use of these compounds up to the highest safe level in feeds. Hazards for skin and eye contact, and respiratory exposure are recognised for the majority of the compounds under application. Most are classified as irritating to the respiratory system. The concentrations considered safe for the target species are unlikely to have detrimental effects on the terrestrial and fresh water environments. As all the compounds under assessment are used in food as flavourings, and their function in feed is essentially the same as that in food, no further demonstration of efficacy is necessary. In the absence of data on the stability in water for drinking, the FEEDAP Panel is unable to conclude on the safety or efficacy of the substances under this mode of delivery.

© European Food Safety Authority, 2016

Keywords: sensory additives, pyridine, pyrrole, quinoline derivatives, safety, chemical group 28

Requestor: European Commission Question number: EFSA-Q-2010-01171 Correspondence: feedap@efsa.europa.eu



Panel members: Gabriele Aquilina, Giovanna Azimonti, Vasileios Bampidis, Maria de Lourdes Bastos, Georges Bories, Andrew Chesson, Pier Sandro Cocconcelli, Gerhard Flachowsky, Jürgen Gropp, Boris Kolar, Maryline Kouba, Secundino López Puente, Marta López-Alonso, Alberto Mantovani, Baltasar Mayo, Fernando Ramos, Guido Rychen, Maria Saarela, Roberto Edoardo Villa, Robert John Wallace and Pieter Wester.

Acknowledgements: The Panel wishes to thank the members of the Working Group on Feed Flavourings, including Paul Brantom, Birgit Dusemund, Christer Hogstrand, Robert Luttik, Patrick Van Beelen and Johannes Westendorf, for the support provided to this scientific output.

Suggested citation: EFSA FEEDAP Panel (EFSA Panel on Additives and Products or Substances used in Animal Feed), 2016. Scientific opinion on the safety and efficacy of pyridine, pyrrole, and quinoline derivatives belonging to chemical group 28 when used as flavourings for all animal species. EFSA Journal 2016;14(2):4390, 19 pp. doi:10.2903/j.efsa.2016.4390

ISSN: 1831-4732

© European Food Safety Authority, 2016

Reproduction is authorised provided the source is acknowledged.



The EFSA Journal is a publication of the European Food Safety Authority, an agency of the European Union.





Table of contents

Abstrac	t	1
1.	Introduction	
1.1.	Background and Terms of Reference	4
1.2.	Additional information	
2.	Data and Methodologies	5
2.1.	Data	5
2.2.	Methodologies	5
3.	Assessment	5
3.1.	Characterisation	5
3.1.1.	Characterisation of the flavouring additives	5
3.1.2.	Stability and homogeneity	7
3.1.3.	Conditions of use	
3.2.	Safety	
3.2.1.	Absorption, distribution, metabolism and excretion (ADME) and residue studies	7
3.2.2.	Toxicological studies	
3.2.3.	Safety for the target species	
3.2.4.	Safety for the consumer	
3.2.5.	Safety for the user	
3.2.6.	Safety for the environment	13
3.3.	Efficacy	14
4.	Conclusions	
Docum	entation provided to EFSA	15
Referer	nces	15
	iations	
Annex ·	- Executive Summary of the Evaluation Report of the European Union Reference Laboratory	

for Feed Additives on the Method(s) of Analysis for Pyridine, pyrrole, and quinoline derivative19

1. Introduction

1.1. Background and Terms of Reference

Regulation (EC) No 1831/2003¹ establishes the rules governing the Community authorisation of additives for use in animal nutrition. In particular, Article 4(1) of that Regulation lays down that any person seeking authorisation for a feed additive or for a new use of a feed additive shall submit an application in accordance with Article 7, in addition, Article 10(2) of that Regulation also specifies that for existing products within the meaning of Article 10(1), an application shall be submitted in accordance with Article 7, within a maximum of 7 years after the entry into force of this Regulation.

The European Commission (EC) received a request from Feed Flavourings Authorisation Consortium European Economic Interest Grouping (FFAC EEIG)² for authorisation of nine substances belonging to chemical group (CG) 28, when used as feed additives for all animal species (category: sensory additives; functional group: flavouring compounds). CG 28 for flavouring substances is defined in Commission Regulation (EC) No 1565/2000³ as 'pyridine, pyrrole and quinoline derivatives'.

According to Article 7(1) of Regulation (EC) No 1831/2003, the Commission forwarded the application to the European Food Safety Authority (EFSA) as an application under Article 4(1) (authorisation of a feed additive or new use of a feed additive) and under Article 10(2) (re-evaluation of an authorised feed additive). EFSA received directly from the applicant the technical dossier in support of this application. The particulars and documents in support of the application were considered valid by EFSA as of 9 June 2010.

According to Article 8 of Regulation (EC) No 1831/2003, EFSA, after verifying the particulars and documents submitted by the applicant, shall undertake an assessment in order to determine whether the feed additive complies with the conditions laid down in Article 5.

EFSA shall deliver an opinion on the safety for the target animals, consumer, user and the environment and on the efficacy of trimethyloxazole, piperine, 3-methylindole, indole, 2-acetylpyridine, 2-acetylpyridine, 3-ethylpyridine, pyrrolidine and 2,6-dimethylpyridine, when used under the proposed conditions of use (see Section 0).

1.2. Additional information

All nine substances have been assessed by the Joint Food and Agriculture Organization of the United Nations (FAO)/World Health Organization (WHO) Expert Committee on Food Additives (JECFA; WHO 2005, 2006 and 2008) and were considered safe for use in food. No acceptable daily intake values were specified. Subsequently the EFSA Panel on Food Additive, Flavourings, Processing Aids and Materials in Contact with Food (CEF) considered the same compounds for use as food flavourings (EFSA, 2008a,b, 2009; EFSA CEF Panel, 2015a,b) reaching the same conclusions.

The compounds are all currently listed in the European Union (EU) database of flavouring substances⁴ and in the EU Register of Feed Additives, respectively, and thus authorised for use in food and feed in the EU. They have not been previously assessed by EFSA as feed additives.

Regulation (EC) No 429/2008⁵ allows substances already approved for use in human food to be assessed with a more limited procedure than for other feed additives. However, the use of this

¹ Regulation (EC) No 1831/2003 of the European Parliament and of the Council of 22 September 2003 on additives for use in animal nutrition. OJ L 268, 18.10.2003, p. 29.

² On 13/03/2013, EFSA was informed by the applicant that FFAC EEIG was liquidated on 19/12/2012 and their rights as applicant were transferred to FEFANA asbl (EU Association of Specialty Feed Ingredients and their Mixtures). Avenue Louise 130A, Box 1, 1050 Brussels, Belgium.

³ Commission Regulation (EC) No 1565/2000 of 18 July 2000 laying down the measures necessary for the adoption of an evaluation programme in application of Regulation (EC) No 2232/96 of the European Parliament and of the Council. OJ L 180, 19.7.2000, p. 8.

⁴ Commission Implementing Regulation (EU) No 872/2012 of 1 October 2012 adopting the list of flavouring substances provided for by Regulation (EC) No 2232/96 of the European Parliament and of the Council, introducing it in Annex I to Regulation (EC) No 1334/2008 of the European Parliament and of the Council and repealing Commission Regulation (EC) No 1565/2000 and Commission Decision 1999/217/EC. OJ L 267, 2.10.2012, p. 1.

⁵ Commission Regulation (EC) No 429/2008 of 25 April 2008 on detailed rules for the implementation of Regulation (EC) No 1831/2003 of the European Parliament and of the Council as regards the preparation and the presentation of applications and the assessment and the authorisation of feed additives. OJ L 133, 22.5.2008, p. 1.



procedure is always subject to the condition that food safety assessment is relevant to the use in feed.

2. Data and Methodologies

2.1. Data

The present assessment is based on data submitted by the applicant in the form of a technical dossier⁶ in support of the authorisation request for the use of the pyridine, pyrrole and quinoline derivatives as a feed additive. The technical dossier was prepared following the provisions of Article 7 of Regulation (EC) No 1831/2003, Regulation (EC) No 429/2008 and the applicable EFSA guidance documents.

The FEEDAP Panel has sought to use the data provided by the applicant together with data from other sources, such as previous risk assessments by EFSA or other expert bodies, peer-reviewed scientific papers and experts' knowledge, to deliver the present output.

EFSA has verified the European Union Reference Laboratory (EURL) report as it relates to the methods used for the control of flavourings from CG 28 – pyridine, pyrrole and quinoline derivatives – in animal feed. The Executive Summary of the EURL report can be found in Annex A.⁷

2.2. Methodologies

The approach followed by the FEEDAP Panel to assess the safety and the efficacy of the pyridine, pyrrole and quinoline derivatives is in line with the principles laid down in Regulation (EC) No 429/2008 and the relevant guidance documents: Guidance for the preparation of dossiers for sensory additives (EFSA FEEDAP Panel, 2012a), Technical Guidance for assessing the safety of feed additives for the environment (EFSA, 2008c), Guidance for the preparation of dossiers for additives already authorised for use in food (EFSA FEEDAP Panel, 2012b), Guidance for establishing the safety of additives for the consumer (EFSA FEEDAP Panel, 2012c) and Guidance on studies concerning the safety of use of the additive for users/workers (EFSA FEEDAP Panel, 2012d).

3. Assessment

3.1. Characterisation

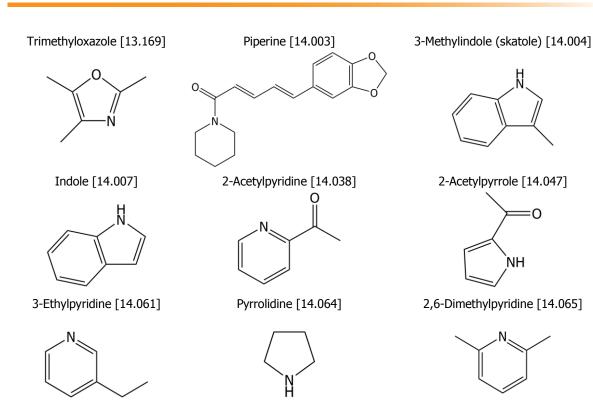
3.1.1. Characterisation of the flavouring additives

The molecular structures of the nine additives under application are shown in Figure 1 and their physico-chemical characteristics in Table 1.

⁶ FEED dossier reference: FAD-2010-0117.

⁷ The full report is available on the EURL website https://ec.europa.eu/jrc/sites/default/files/FinRep-FAD-2010-0117.pdf





- Figure 1: Molecular structures and [EU Flavour Information System (FLAVIS) numbers] of the nine flavouring compounds under assessment
- **Table 1:** Chemical Abstracts Service (CAS) and FLAVIS numbers and some characteristics of the chemically defined flavourings under assessment

EU Register name	CAS No	Flavis No	Molecular formula	Molecular weight	Physical state	Log Kow ^(a)
Trimethyloxazole	20662-84-4	13.169	C ₆ H ₉ ON	111.14	Liquid	1.09
Piperine	94-62-2	14.003	C ₁₇ H ₁₈ O ₃ N	285.34	Solid	3.69
3-Methylindole	83-34-1	14.004	C ₉ H ₉ N	131.18	Solid	2.6
Indole	120-72-9	14.007	C ₈ H ₇ N	115.15	Solid	2.14
2-Acetylpyridine	1122-62-9	14.038	C ₇ H ₇ ON	121.14	Liquid	0.85
2-Acetylpyrrole	1072-83-9	14.047	C ₆ H ₇ ON	109.13	Solid	0.93
3-Ethylpyridine	536-78-7	14.061	C ₇ H ₉ N	107.06	Liquid	1.66
Pyrrolidine	123-75-1	14.064	C ₄ H ₉ N	71.12	Liquid	0.46
2,6-Dimethylpyridine	108-48-5	14.065	C ₇ H ₉ N	107.16	Liquid	1.68

EU: European Union; CAS No: Chemical Abstract Service No.; Flavis number: EU Flavour Information System numbers; (a): Logarithm of octanol–water partition coefficient.

These substances are produced by chemical synthesis. Typically several routes of synthesis are available and described in the dossier. 8

Batch-to-batch variation data were provided for five batches of each additive. The mean content of the active substance for all compounds except piperine exceeded the JECFA specifications (Table 2).

⁸ Technical dossier/Section II.



EU Register name	Flavis No	JECFA specification	Assay %		
		minimum % ^(a)	Average	Range	
Trimethyloxazole	13.169	> 95	99.7	99.2–100	
Piperine	14.003	> 97	95.3	95.0–95.9	
3-Methylindole	14.004	> 97	99.7	98.9–100	
Indole	14.007	> 97	99.7	99.3–100	
2-Acetylpyridine	14.038	> 97	97.9	95.7–100	
2-Acetylpyrrole	14.047	> 97	99.9	99.4–100	
3-Ethylpyridine	14.061	> 98	99.8	99.6–100	
Pyrrolidine	14.064	> 95	99.7	99.6–99.7	
2,6-Dimethylpyridine	14.065	> 99	99.6	99.4–100	

Table 2: Identity of the substances and data on purity

EU: European Union; Flavis number: EU Flavour Information System numbers; JECFA: The Joint FAO/WHO Expert Committee on Food Additives

(a): FAO, 2006.

Potential contaminants are considered as part of the product specification and are monitored as part of the Hazard Analysis and Critical Control Point procedure applied by all consortium members. The parameters considered include residual solvents, heavy metals and other undesirable substances. However, no evidence of compliance was provided for these parameters.

3.1.2. Stability and homogeneity

A minimum shelf-life for the compounds under assessment range from 6 to 24 months when stored in closed containers under recommended conditions. This assessment is made on the basis of compliance with the original specification over this storage period.

Although no data are required for the stability of flavours in premixtures and feed, their use in water for drinking introduces other issues relating to product stability, such as degradation due to microbial activity. The FEEDAP Panel notes that three out of the nine compounds in CG 28 have low water solubility (log $K_{OW} > 2$). Considering this, and in the absence of data on the short-term stability and homogeneity of the all additives in water for drinking, the FEEDAP Panel is not in the position to conclude on the use of the additives in water for drinking.

3.1.3. Conditions of use

The applicant proposes the use of the nine additives in feed or water for drinking for all animal species without withdrawal period. The normal use level is 0.1 mg/kg feed and a high use level of 0.5 mg/kg feed. No specific proposals are made for doses to be used in water for drinking.

3.2. Safety

The assessment of safety is based on the highest use level proposed by the applicant (0.5 mg/kg complete feed).

3.2.1. Absorption, distribution, metabolism and excretion (ADME) and residue studies

Absorption, distribution and excretion

In its evaluation of pyridine and pyrrole derivatives, JECFA reported that flavourings from this group are expected to be rapidly absorbed from the gastrointestinal tract (WHO, 2005). JECFA also reported that oxazoles, such as trimethyloxazole [13.169] are rapidly absorbed. Pyrroles and indoles are weak bases with pKa values of > 3 and are readily absorbed from the intestine by passive diffusion because the base will not be ionised and pass through intestinal membranes with ease. Pyridines are weak bases and undergo rapid absorption in the gastrointestinal tract (WHO, 2006).

The excretion of indole in rats is fairly rapid with 75% of the dosage excreted after 24 h, and more than 80% of the radioactivity recovered from the pooled 48-h urine of female albino Wistar rats given

a single oral dose of 64 to 80 mg/kg $[2^{-14}C]$ -indole. The radioactivity measured in urine, faeces and expired air after 48 h was 80.6, 11.1 and 2.4% of the dose (King et al., 1966).

Twenty-four male Holtzman rats were fed diets supplemented with 0, 0.25, 0.50 or 0.75% (corresponding to 0, 150, 300 and 450 mg/kg body weight (bw) per day) of indole for 3 weeks. During week 3, all groups (including controls) received a single dose of [2-14C]-indole via stomach tube. Urine collected for 72 h after the single dose administration revealed that 32, 47, 49 and 51% of the radioactive dose were recovered from the 0 (control), 0.25, 0.50 and 0.75% indole groups (Martinez and Roe, 1972).

Mice and rats given single intraperitoneal injections of 400 mg/kg of [¹⁴C]-3-methylindole (skatole) excreted 69.4% and 66.2% of the total radioactivity, respectively, after 48 h (Skiles and Yost, 1992).

When piperine was administered to male albino rats by gavage (170 mg/kg bw per day), about 97% was absorbed, the remaining 3% of the administered dose was excreted as piperine in the faeces. Piperine was not detectable in urine (the limit of detection was not known), indicating a complete metabolic conversion of this compound. Examination of the passage of piperine through the gut indicated that the highest concentration in the stomach and small intestine was attained at about 6 h. Only traces (less than 0.15%) of piperine were detected in serum, kidney and spleen from 30 minutes to 24 h. For intraperitoneally administered piperine (85 mg/kg bw per day), about 1–2.5% of the dose was detected in the liver during 0.5–6 h after administration as contrasted with 0.1–0.25% of the orally administered dose (Bhat and Chandrasekhara, 1986).

When everted sacs of rat intestines were incubated with $200-1,000 \mu g$ piperine, about 47-64% of the added piperine disappeared from the mucosal side. Only piperine was present in the serosal fluid and also in the intestinal tissue, indicating that piperine did not undergo any metabolic change during absorption (Bhat and Chandrasekhara, 1986).

Metabolism

Once absorbed, pyridine and pyrrole derivatives are oxidised to polar metabolites, and eliminated primarily in the urine and, to a minor extent, in the faeces. Alkyl-substituted pyrroles and indoles may undergo cytochrome P450-mediated side-chain oxidation to yield the corresponding alcohol. Unsubstituted indole is metabolised primarily by ring hydroxylation at the C-3 position. The resulting hydroxyl derivatives are conjugated with glucuronic acid or sulfate and excreted in the urine. To a lesser extent, the double bond of the pyrrole ring may undergo epoxidation and subsequent glutathione conjugation. Alkyl-substituted pyridines are principally subject to side-chain oxidation, primarily at the C-1 position. N-Oxide formation has also been reported (WHO, 2005, 2006). Alicyclic secondary amines such as pyrrolidine undergo C-oxidation at the a-carbon, but oxidation can also occur at other carbons on the ring (WHO, 2006). The metabolism of oxazoles, such as trimethyloxazole, involves oxazole ring cleavage, and depending on the degree of ring substitution, ring hydroxylation. The presence of a substituent at the 2-position tends to stabilise the oxazole ring.

Metabolism studies in laboratory animals are available for a number of compounds belonging to CG 28 and have been extensively reviewed by the CEF Panel (EFSA CEF Panel, 2013, 2014).

Hydroxylation of both rings of indole derivatives has been well documented. 3-Hydroxyindole (indoxyl), resulting from electrophilic attack at C-3, is the main metabolite of indole in various species. Minor metabolites are oxindole and 5-hydroxyoxindole, resulting from hydroxylation at C-2 and subsequent C-5 hydroxylation (Damani and Crooks, 1982). The metabolism of $[2^{-14}C]$ -indole was investigated by King et al. (1966). When single oral doses of 64–74 mg/kg bw of $[2^{-14}C]$ -indole were administered to three albino Wistar rats, 63% of the dose was detected in 48-h urine pools as 3-hydroxyindole-sulfate (49.6%) and 3-hydroxyindole-glucuronide (13.2%). Minor metabolites identified in the urine were 5-hydroxyoxindole (3.5%), indole-2-one (1.4%) and indole-2,3-dione (or isatin, 5.8%). In faeces, indole, 3-hydroxyindole-sulfate and total 3-hydroxyindole metabolites represented 0.14, 0.40 and 0.64% of the dose, respectively; in the bile, 5-hydroxyoxindole, 3-hydroxyindole-sulfate and total 3-hydroxyindole metabolites, accounted for 0.56, 0.80 and 0.82% of the dose, respectively. Similar findings were reported by Martinez and Roe (1972). Twenty-four male Holtzman rats were administered diets supplemented with 0, 0.25, 0.50 or 0.75% (corresponding to 0, 150, 300 and 450 mg/kg bw per day) indole for 3 weeks. After a single dose administration of $[2^{-14}C]$ -indole by gavage (week 3), sulfate and glucuronide conjugates of hydroxylated indole were the two main



radioactive metabolites detected in the urine collected for 72 h. The excretion of glucuronide was increased with increasing doses of administered indole.

The major pathways of metabolism of 3-methylindole in mice and goat (Skiles et al., 1991; Smith et al., 1993) are oxygenation of the double bond in position 2 and 3 (epoxidation product), and stepwise oxidation of the methyl group and hydroxylation of the ring structure, leading to metabolites mainly conjugated with glucuronic acid, sulfate and glutathione. Mice and rats given single intraperitoneal injections of 400 mg/kg of [methyl-14C]-3-methylindole excreted 2.6 and 7.3%, respectively, as the mercapturic acid conjugate of 3-methylindole, 3-[(*N*-acetylcystein-*S*-yl)methyl]indole (Skiles et al., 1991). Three primary pathways were characterised and at least six metabolites isolated from the urine after 36 h of male Swiss-Webster mice receiving 400 mg/kg of ring labelled [ring-UL-14C]-3-methylindole by intraperitoneal injection. In the first pathway, side-chain oxidation yields indole-3-carbinol, which is further oxidised to the corresponding carboxylic acid. Alternatively, 3-methylindole can be converted to the reactive substance 3-methyleneindolenine, which subsequently is conjugated with glutathione to yield 3-[(N-acetylcystein-S-yl)methyl]indole. In the third pathway, the 2,3-alkene is epoxidised to yield 3-methyloxindole or 3-hydroxy-3methylindolenine intermediates. These metabolites are conjugated with glucuronic acid or sulfate, followed by excretion in the urine or are further oxidised to yield a series of dihydroxy-3methyloxindole metabolites that are also conjugated and excreted. The second pathway through epoxidation of 3-methylindole predominates (Smith et al., 1993).

No unchanged piperine was found in the urine of male albino rats administered piperine either by gavage (170 mg/kg bw per day) or intraperitoneally (85 mg/kg bw per day). Cleavage of the methylenedioxy group of piperine followed by glucuronidation and sulfation appear to be the major steps in the metabolism of piperine in the rat (Bhat and Chandrasekhara, 1986). After oral administration of piperine (170 mg/kg bw per day) to rats, bile and urine were examined for metabolites. Four metabolites, piperonylic acid, piperonyl alcohol, piperonal and vanillic acid were identified in the free form in 0–96 h urine whereas only piperic acid was detected in 0–6 hours bile (Bhat and Chandrasekhara, 1987). This metabolic profile could not be confirmed in a more recent study (Bajad et al., 2003), where a new major metabolite was identified in the urine of rats and characterised as $5-(3,4-methylenedioxy-phenyl)-2E_AE$ -pentadienoic acid-*N*-(3-yl-propionic acid)-amide. This metabolite is proposed to be formed by the oxidation of the piperidine ring at position 4 followed by loss of the ethylene group by a-carbon cleavage and further oxidation.

Mendes (2012) detected three major compounds in the urine of male and female rats dosed with 2acetylpyrrole [14.047] by oral gavage. Unchanged 2-acetylpyrrole and pyrrol-2,5-dione were identified by gas chromatography–mass spectrometry (GC–MS); the third compound was thought to be 1,5dihydropyrrol-2-one, but its identity was not confirmed (EFSA CEF Panel, 2015a).

3-Ethylpyridine was converted to the corresponding N-oxide by fortified hepatic microsomal preparations from hamster, guinea-pig, rabbit, rat and mouse, and by pulmonary microsomal preparations from guinea-pig and rabbit (Cowan et al., 1978 as quoted by EFSA CEF Panel 2013). *In vivo*, no urinary N-oxidation products were found for dimethylpyridines, the major metabolism being the result of the oxidation of one of the methyl groups. When 2,6-dimethylpyridine (100 mg/kg bw) was administered via gavage to male Wistar rats, 90% was excreted as the glycine conjugate of the 6-methyl-2-carboxylic acid (Hawksworth and Scheline, 1975).

Studies of metabolism of compounds belonging to CG 28 in animals, other than rats, are lacking in the scientific literature. Oxidation is ubiquitous and phase II conjugation via glucuronidation or sulfation occurs in mammals, although the predominance of one pathway over another varies among animal species (Gupta, 2007). Data collected in a review by Ioannides (2006) show that the cytochrome P450 enzymes responsible for the majority of oxidation reactions are expressed in the liver of the main food-producing animals (cattle, pig, sheep, goat) as well as in the rabbit and chicken (Nebbia et al., 2003). Biotransformation through oxidation followed by conjugation with glucuronic acid and sulfate has also been reported for birds (Pan and Fouts, 1978). A recent study showed that the principal cytochrome P450 enzymes responsible for oxidation of xenobiotics, as well as glutathione transferases, are present in the liver of chickens (Blevins et al., 2012). Fish have analogous mechanisms for handling xenobiotic compounds, including both phase I and phase II biotransformation reactions, and many of the same microsomal and cytosolic enzymes as mammals (Wolf and Wolfe, 2005). Thus, fish can transform endobiotic and xenobiotic compounds through



oxidation or hydroxylation, conjugate the metabolites to polar substrates through sulfate, and glucuronide conjugation (James and Pritchard, 1987) with further elimination via bile or urine (Di Giulio and Hinton, 2008). Therefore, food-producing animals, including fish and birds, can also be assumed to have the ability to metabolise and excrete the flavouring substances from CG 28 and there is no evidence that they or their metabolites would accumulate in tissues.

3.2.2. Toxicological studies

Toxicological data (subchronic, repeated-dose studies, with multiple doses tested) could be found for piperine [14.003], 2-acetylpyridine [14.038] and 2-acetylpyrrole [14.047].

A no observed adverse effect level (NOAEL) of 37 mg 2-acetylpyridine/kg bw per day was identified in a 90-day study in rats (doses: 0, 37, 110, 330 and 1,000 mg/kg bw per day, 6 days a week; M/F, 20 animals each group, administration route: gavage). Slight anaemia was observed in females from 110 mg/kg bw per day and in males from 330 mg/kg bw per day. Liver enlargement was observed at 330 and 1,000 mg/kg bw per day (Til and van der Meulen, 1971, unpublished).

A 90-day study was performed with piperine [14.003] (Bauter, 2013). The study was performed according to The Organisation for Economic Co-operation and Development (OECD) Guideline 408. Four groups of adult Crl: Sprague–Dawley[®] CD[®] IGS rats (10/sex and group) were maintained on diets, calculated to provide piperine intake levels of 4.8, 14.5 and 47.8 mg/kg bw per day in males, and 4.8, 14.6 and 48.4 mg/kg bw per day in females, giving an average daily intake of 0 (vehicle), 5, 15 or 50 mg/kg bw per day for males and females.

There were no mortalities, clinical or ophthalmological changes, attributable to piperine administration. Decreased male body weight gain (20% reduction) and reduced male (15% reduction) and female (12% reduction) food consumption in the top dose group were attributed to the possible decrease in palatability caused by the test substance at high dietary levels. No effect was observed on the final body weights.

There were no gross and microscopic changes or clinical pathology or organ weight changes attributed to the administration of piperine. A statistical significant and dose-dependent increase in plasma cholesterol in males was observed of approximately 30% in the 15 mg/kg bw per day and 55% in the 50 mg/kg bw/day groups. No effect on cholesterol was observed in females. Based on the dose-dependent increase in plasma cholesterol levels in males at the mid and high dose, the CEF Panel decided that the lowest dose level of 5 mg/kg bw per day should be considered as the NOAEL. This conclusion is supported by the FEEDAP Panel.

In a 90-day study compliant with OECD guideline 408, groups of rats (10/sex and dietary intake level) of male and female Sprague–Dawley CD rats were fed a diet designed to provide 0 (dietary control), 1050, 2100 and 4200 mg 2-acetylpyrrole [14.047]/kg feed daily (Bauter, 2012). These dietary levels correspond to the calculated average daily intakes of 0, 68, 133 and 263 mg/kg bw per day for males and 0, 79, 155 and 298 mg/kg bw per day or females, respectively.

The test material was not stable in the diet, and in the report (Bauter, 2012) it is suggested that part of it was probably not detected by the extraction method employed due to formation of complexes with metal ions in the feed. It is calculated that over the course of the study the animals received concentrations of 35–40% of the target intake level on average. Therefore, values for exposure levels based on the measured intake are proportionally lower. Based on this analysis of the test diets, the mean daily intakes were calculated to 367, 754 and 1705 mg/kg feed. Assuming that possible toxicity is only related to the free 2-acetylpyrrole, these dietary concentrations correspond to average daily intakes of 24, 48 and 107 mg/kg bw per day for males, and 28, 56 and 121 mg/kg bw per day for females, respectively, over 90 days.

Statistically significant dose-dependent reductions in body weight, body weight gain, feed consumption (males and females) and feed to gain ratio (females) at the highest dietary level (1705 mg/kg feed measured concentration) during the study were attributed to the possible decrease in palatability caused by the test substance at high dietary levels.

Female rats of the high intake groups displayed minimal to moderate dark bilateral thyroid glands. Microscopic changes were slight thyroid hypertrophy/hyperplasia among 4/10 and 10/10 high intake group males and females, respectively. This was characterised by enlarged subgross tall columnar



appearance of the follicular epithelial cells which appeared with fine cytoplasmic vacuolation with intermittent focally piled papillary projections into the follicular lumen.

The thyroid effects at the high exposure level are considered adverse. Therefore, a NOAEL for 2-acetylpyrrole is derived from the dose of 48 mg/kg bw per day in males.

3.2.3. Safety for the target species

The first approach to the safety assessment for target species takes account of the applied use levels in animal feed relative to the maximum reported exposure of humans on the basis of the metabolic body weight. Human exposure in the EU ranges from 0.12 to 50 µg/person per day (EFSA, 2008a,b, 2009; EFSA CEF Panel 2015a,b). This corresponds from 0.006 to 2.31 µg/kg^{0.75} per day. Table 3 summarises the result of the comparison with human exposure for representative target animals. The body weight of target animals is taken from the default values shown in Table 4.

Table 3: Comparison of exposure of humans and target animals (calculated from the proposed maximum feed concentrations, see 3.1.3) to the flavourings under application

Flavouring	Use level in feed	Human exposure (µg/kg bw ⁰⁷⁵	Target animal exposure (µg/kg bw ⁰⁷⁵ per day)			
	(mg/kg)	per day) ^(a)	Salmon	Piglet	Dairy cow	
Trimethyloxazole	0.5	0.17	11.8	52.6	77.7	
Piperine	0.5	0.93	11.8	52.6	77.7	
3-Methylindole	0.5	0.11	11.8	52.6	77.7	
Indole	0.5	1.21	11.8	52.6	77.7	
2-Acetylpyridine	0.5	2.32	11.8	52.6	77.7	
2-Acetylpyrrole	0.5	0.15	11.8	52.6	77.7	
3-Ethylpyridine	0.5	0.43	11.8	52.6	77.7	
Pyrrolidine	0.5	0.006	11.8	52.6	77.7	
2,6-Dimethylpyridine	0.5	0.012	11.8	52.6	77.7	

(a): Metabolic body weight (kg $bw^{0.75}$) for a 60-kg person = 21.6.

Table 3 shows that for all nine compounds the intake by the target animals greatly exceeds that of humans resulting from use in food. As a consequence, safety for the target species at the feed concentration applied cannot be derived from the risk assessment for food use.

As an alternative, the maximum feed concentration considered as safe for the target animals can be derived from the lowest NOAEL available. Toxicological data could only be found for piperine [14.003], 2-acetylpyridine [14.038] and acetylpyrrole [14.047]. The FEEDAP Panel notes that the purity of piperine for feed use is lower than in the JECFA specification (95 vs 97%). However, considering the intended use level and the availability of relevant toxicological studies, this difference is not considered of concern. Applying an uncertainty factor (UF) of 100 to the NOAELs, the maximum safe intake for the target species was derived for the three compounds following the EFSA guidance for sensory additives (EFSA FEEDAP Panel, 2012a), and thus the maximum safe feed concentration was calculated. The results are summarised in Table 4.

Table 4:	Maximum safe concentration in feed for different target animals for piperine (A), 2-
	acetylpyridine (B) and 2-acetylpyrrole (C)

Target animal	Default values		Maximum safe intake/feed concentration					
	Body weight (kg) Feed intake (g/day) ^(a)		Intake (mg/day)			Concentration (mg/kg feed) ^(b)		
			Α	В	С	Α	В	С
Salmonids	2	40	0.1	1	1	2.5	19	24
Veal calves (milk replacer)	100	2,000	5	37	48	2.5	19	24
Cattle for fattening	400	8,000	20	148	192	2.2	16	21
Pigs for fattening	100	3,000	5	37	48	1.7	12	16
Sows	200	6,000	10	74	96	1.7	12	16
Dairy Cows	650	20,000	32.5	241	312	1.4	11	14
Turkeys for fattening	12	400	0.6	4	6	1.5	11	14
Piglets	20	1,000	1	7	10	1.0	7	10



Target animal	Default	Default values		Maximum safe intake/feed concentration					
	Body weight (kg)	Feed intake (g/day) ^(a)	Intake (mg/day)			Concentration (mg/kg feed) ^(b)			
			Α	В	С	Α	В	С	
Chickens for fattening	2	120	0.1	1	1	0.8	6	8	
Laying hens	2	120	0.1	1	1	0.8	6	8	
Dogs	15	250	0.75	6	7	2.6	20	25	
Cats	3	60	0.15	1	1	2.2	16	21	

(a): Complete feed with 88% dry matter (DM), except milk replacer for veal calves (94.5% DM), and for cattle for fattening, dairy cows, dogs and cats for which the values are DM intake.

(b): Complete feed containing 88% DM, milk replacer 94.5% DM.

For the six remaining compounds, adequate subchronic, repeated-dose studies performed with the additive under assessment were not available. Therefore, the threshold of toxicological concern (TTC) approach was followed to derive the maximum safe feed concentration (EFSA FEEDAP Panel, 2012a).

For the Cramer class I compounds, 3-methylindole [14.004] and indole [14.007], the calculated safe use level is 1.5 mg/kg complete feed for cattle, salmonids and non-food-producing animals, and 1.0 mg/kg complete feed for pigs and poultry.

For the Cramer Class II compounds, trimethyloxazole [13.169], 3-ethylpyridine [14.061], pyrrolidine [14.064] and 2,6-dimethylpyridine [14.065], the calculated safe use level is 0.5 mg/kg complete feed for cattle, salmonids and non-food-producing animals, and 0.3 mg/kg complete feed for pigs and poultry.

Conclusions on safety for the target species

The FEEDAP Panel concludes that:

- piperine [14.003], 3-methylindole [14.004], indole [14.007], 2-acetylpyridine [14.038] and 2acetylpyrrole [14.047] are safe at the proposed maximum dose level (0.5 mg/kg complete feed) for all animal species;
- trimethyloxazole [13.169], 3-ethylpyridine [14.061], pyrrolidine [14.064] and 2,6dimethylpyridine [14.065] are safe at the proposed use level of 0.5 mg/kg complete feed for cattle, salmonids and non-food-producing animals, and at the use level of 0.3 mg/kg complete feed for pigs and poultry.

3.2.4. Safety for the consumer

The safety for the consumer of the compounds in CG 28, used as food flavours, has already been assessed by JECFA (WHO, 2005, 2006 and 2008) and EFSA (EFSA, 2008a,b, 2009; EFSA CEF Panel 2015a,b). All these compounds are presently authorised as food flavourings without limitations.

Given the low use levels of CG 28 compounds to be applied in feed, and the expected extensive metabolism and excretion in target animals (see Section 3.2.1), the FEEDAP Panel considers that the possible residues in food derived from animals fed with these flavourings would not appreciably increase the human intake levels of these compounds from natural sources and food use. Therefore, the FEEDAP Panel concludes that the use of the nine additives under assessment in animal nutrition at the proposed use levels is safe for the consumer.

3.2.5. Safety for the user

No specific data on the safety for the user were provided. In the material safety data sheets,⁹ hazards for skin and eye contact, and respiratory exposure are recognised for the majority of the compounds under application. Most are classified as irritating to the respiratory system.

⁹ Technical dossier/Section II/Annex II.3



3.2.6. Safety for the environment

The additions of naturally occurring substances that will not result in a substantial increase of the concentration in the environment are exempt from further assessment (EFSA, 2008c). Examination of the published literature shows that this applies to piperine [14.003], 3-methylindole [14.004], indole [14.007], 2-acetylpyridine [14.038], 2-acetylpyrrole [14.047] and pyrrolidine [14.064], which occur in the environment at levels above the application rate of 0.5 mg/kg feed [data taken from the Netherlands Organisation for Applied Scientific Research (TNO) database Volatile Compounds in Food *ver.* 14.1; Burdock, 2003].¹⁰

The applicant did not demonstrate that the other three compounds trimethyloxazole [13.169], 3-ethylpyridine [14.061] and 2,6-dimethylpyridine [14.065] occur in the environment at levels above the application rate of 0.5 mg/kg feed. These substances are therefore assessed in a predicted environmental concentration (PEC) calculation for soil (PEC_{soil}) arising from the application rate. When the calculations are performed according to the EFSA guidance (EFSA, 2008c) with a fixed concentration in feed, there is a fixed order of PEC_{soil} from each species, with the lamb being the most critical.

EU Register name	CAS No	Dose mg/kg	PEC soil (µg/kg)	PEC pore water (µg/L)
Trimethyloxazole	20662-84-4	0.5	11	1.8
3-Ethylpyridine	536-78-7	0.5	11	2.7
2,6-Dimethylpyridine	108-48-5	0.5	11	3.2

EU: European Union; PEC: predicted environmental concentration; CAS No: Chemical Abstracts Service.

Table 5 shows the PEC_{soil} for lambs. The values are slightly above the threshold of 10 μ g/kg (EFSA, 2008c). The PEC for pore water, however, is dependent on the sorption, which is different for each compound. For these calculations, the substance-dependent constants organic carbon sorption constant (K_{oc}), molecular weight, vapour pressure and solubility are needed. These were estimated from the Simplified Molecular Input Line Entry Specification (SMILES) notation of the chemical structure using EPIWEB 4.1 (Table 6).¹¹ This program was also used to derive the SMILES notation from the CAS numbers. The K_{oc} value derived from the first-order molecular connectivity index was used, as recommended by the EPIWEB program.

Table 6:	Physico-chemical	properties pred	dicted by EPIWEB 4.1
----------	------------------	-----------------	----------------------

EU Register name	CAS No	.1				
		DT ₅₀ ^(a) (days)	Molecular weight (g/mol)	Vapour pressure (Pa)	Solubility (mg/L)	<i>K</i> _{oc} ^(b) (L/kg)
Trimethyloxazole	20662-84-4	16	111.14	762	2,801	337
3-Ethylpyridine	536-78-7	18	107.16	264	84,780	219
2,6-Dimethylpyridine	108-48-5	22	107.16	702	81,500	185

EU: European Union; CAS No: Chemical Abstract Service number; DT₅₀: predicted environmental concentration.

(a): DT₅₀, half-life of the additive (by BioWin3).

(b): K_{oc} , organic carbon sorption constant.

The half-life (DT_{50}) was calculated using BioWin3 (Ultimate Survey Model), which gives a rating number. This rating number r was translated into a half-life using the formula by Arnot et al. (2005):

$$\mathsf{DT}_{50} = 10^{(-r \times 1.07 + 4.12)}$$

This is the general regression used to derive estimates of aerobic environmental biodegradation halflives from BioWin3 model output.

All three substances in Table 5 have $PEC_{pore water}$ above 0.1 µg/L. Therefore, they are subjected to phase II risk assessment.

¹⁰ Technical dossier/Supplementary information June 2011.

¹¹ Available online: http://www.epa.gov/opptintr/exposure/pubs/episuitedl.htm



In the absence of experimental data, the phase II risk assessment was performed using ECOSAR v1.11, which estimates the half-maximal effective concentration (EC_{50}) for earthworms, fish, algae and *Daphnia* from the SMILES notation of the substance.

Table 7: The Predicted Environmental Concentration for surface water for lambs compared with
the EC_{50} values in mg/L predicted by ECOSAR 1.11

EU Register name	LC ₅₀ ^(a) Earthworm (mg/kg)	LC ₅₀ Fish (mg/L)	LC ₅₀ Daphnia (mg/L)	EC ₅₀ ^(b) Algae (mg/L)	PEC surface water (µg/L)
Trimethyloxazole	200	123	69	49	0.57
3-Ethylpyridine	193	122	68	48	0.89
2,6-Dimethylpyridine	191	108	61	44	1.05

EU: European Union; EC₅₀: half-maximal effective concentration; LC₅₀: lethal concentration 50; PEC: predicted environmental concentration.

(a): LC_{50} , the concentration of a test substance which results in a 50% mortality of the test species.

(b): EC₅₀, the concentration of a test substance which results in 50% of the test animals being adversely affected (i.e. both mortality and sublethal effects).

The LC_{50} and EC_{50} values (Table 7) divided by a UF of 1,000 were much higher than the PEC values for soil and surface water, for all compounds indicating that, according to the guidance (EFSA, 2008c), there is no risk to the environment at the doses mentioned in Table 5.

The use of all additives in fish feed in land-based aquaculture systems does not give a predicted environmental concentration of the additive (parent compound) in surface water (PEC_{swaq}) above the trigger value of 0.1 μ g/L when calculated according to the guidance. For sea cages, a safe dose of 0.047 mg/kg feed was calculated according to the EFSA guidance (EFSA, 2008c). This dose would give a sediment concentration of 10 μ g/kg which is the threshold level of no concern.

Conclusions on safety for the environment

The concentrations considered safe for the target species (see Section 3.2.3) are unlikely to have detrimental effects on the terrestrial and fresh water environments. For the marine environment, the safe use level is estimated to be 0.05 mg/kg feed.

3.3. Efficacy

Since all nine compounds are used in food as flavourings, and their function in feed is essentially the same as that in food, no further demonstration of efficacy is necessary.

4. Conclusions

The FEEDAP Panel concludes that piperine [14.003], 3-methylindole [14.004], indole [14.007], 2-acetylpyridine [14.038] and 2-acetylpyrole [14.047] are safe at the proposed maximum use level of 0.5 mg/kg complete feed for all animal species; trimethyloxazole [13.169], 3-ethylpyridine [14.061], pyrrolidine [14.064] and 2,6-dimethylpyridine [14.065] are safe at the proposed use level of 0.5 mg/kg complete feed for cattle, salmonids and non-food-producing animals, and at the use level of 0.3 mg/kg complete feed for pigs and poultry.

No safety concern would arise for the consumer from the use of these compounds up to the highest proposed level in feeds.

Hazards for skin and eye contact, and respiratory exposure are recognised for the majority of the compounds under application. Most are classified as irritating to the respiratory system.

The concentrations considered safe for the target species are unlikely to have detrimental effects on the terrestrial and fresh water environments.

As all of the compounds under assessment are used in food as flavourings and their function in feed is essentially the same as that in food, no further demonstration of efficacy is necessary.

In the absence of data on the stability in water for drinking, the FEEDAP Panel is unable to conclude on the safety or efficacy of the substances under this mode of delivery.



Documentation provided to EFSA

- 1. Chemically defined flavourings from Flavouring Group 28 Pyridine, pyrrole and quinoline derivatives (CDG 28). October 2010. Submitted by Feed Flavourings Authorisation Consortium European Economic Interest Grouping (FFAC EEIG).
- 2. Chemically defined flavourings from Flavouring Group 28 Pyridine, pyrrole and quinoline derivatives (CDG 28).Supplementary information. June 2011. Submitted by Feed Flavourings Authorisation Consortium European Economic Interest Grouping (FFAC EEIG).
- 3. Chemically defined flavourings from Flavouring Group 28 Pyridine, pyrrole and quinoline derivatives (CDG 28). Supplementary information. February 2012. Submitted by Feed Flavourings Authorisation Consortium European Economic Interest Grouping (FFAC EEIG).
- 4. Chemically defined flavourings from Flavouring Group 28 Pyridine, pyrrole and quinoline derivatives (CDG 28). Supplementary information. August 2015. Submitted by Feed Flavourings Authorisation Consortium European Economic Interest Grouping (FFAC EEIG)
- 5. Evaluation report of the European Union Reference Laboratory for Feed Additives on the methods(s) of analysis for CDG 28 Pyridine, pyrrole, and quinoline derivatives.
- 6. Comments from Member States.

References

- Arnot J, Gouin T and Mackay D, 2005. Practical Methods for Estimating Environmental Biodegradation Rates, Report to Environment Canada. CEMN Report No 200503. Canadian Environmental Modelling Network, Trent University, Peterborough, ON, Canada.
- Bajad S, Khajuria RK, Suri OP and Bedi KL, 2003. Characterisation of a new minor urinary metabolite of piperine, an omnipresent food component, by LC-MS/MS. Journal of separation science.
- Bauter MR, 2012. Draft Report. 2-Acetylpyrrole: A 90-day dietary study in rats. Product Safety Labs. Study no. 33327. Marts 2, 2012. Unpublished report submitted by EFFA to FLAVIS Secretariat.
- Bauter MR, 2013. Piperine: a 90-day dietary study in rats. Product safety Labs. Study no 35233. May 23, 2013. Unpublished report submitted by EFFA to FLAVIS Secretariat.
- Bhat BG and Chandrasekhara N, 1986. Studies on the metabolism of piperine: absorption, tissue distribution and excretion of urinary conjugates in rats. Toxicology, 40, 83–92.
- Bhat BG and Chandrasekhara N, 1987. Metabolic disposition of piperine in the rat. Toxicology 44, 99-106.
- Blevins S, Siegel PB, Blodget DJ, Ehrich M and Lewis RM, 2012. Liver enzymes in White Leghorns selected for the sheep red blood cell immune response. Poultry Science, 91, 322–326.
- Cowan DA, Damani LA and Gorrod JW, 1978. Metabolic N-oxidation of 3-substituted pyridines: Identification of products by mass spectrometry. Biomedical Mass Spectrometry 5, 551–556.
- Damani LA and Crooks PA, 1982. Oxidative metabolism of heterocyclic ring systems. In: Jakoby, W.B., Bend, J.R., Caldwe, J. (Ed.). Metabolic Basis of Detoxication. 2nd Ed. Academic Press, New York, pp. 69-89.
- Di Giulio RT and Hinton DE, 2008. The toxicology of fishes. CRC Press, Boca Raton, FL, USA, 153–234.
- EFSA (European Food Safety Authority), 2008a. Scientific Opinion of the Panel on Food Additives, Flavourings, Processing Aids and Materials in Contact with Food on Flavouring Group Evaluation Flavouring Group Evaluation 86 (FGE.86). Considerations of aliphatic and aromatic amines and amides evaluated by JECFA (65th meeting). The EFSA Journal, 745, 1–46.
- EFSA (European Food Safety Authority), 2008b. Scientific Opinion of the Panel on Food Additives, Flavourings, Processing Aids and Materials in Contact with Food on Flavouring Group Evaluation 85



(FGE.85). Considerations of miscellaneous nitrogen-containing substances evaluated by JECFA (65th meeting). The EFSA Journal, 804, 1–30.

- EFSA (European Food Safety Authority), 2008c. Technical Guidance of the Scientific Panel on Additives and Products or Substances used in Animal Feed (FEEDAP) for assessing the safety of feed additives for the environment. The EFSA Journal, 842, 1–28.
- EFSA (European Food Safety Authority), 2009. Scientific Opinion of the Panel on Food Additives, Flavourings, Processing Aids and Materials in Contact with Food on Flavouring Group Evaluation 77 (FGE.77). Considerations of Pyridiens, Pyrrole and Quinoline Derivatives evaluated by JECFA (63rd meeting) structurally related to Pyridine, Pyrrole and Quinoline derivatives evaluated by EFSA in FGE.24Rev1 (2008). The EFSA Journal, 936, 1–40.
- EFSA CEF Panel (EFSA Panel on Food Contact Materials, Enzymes, Flavourings and Processing Aids), 2013. Scientific Opinion on Flavouring Group Evaluation 24, Revision 2 (FGE.24Rev2): Pyridine, pyrrole, indole and quinoline derivatives from chemical group 28. EFSA Journal 2013;11(11):3453, 64 pp. doi:10.2903/j.efsa.2013.3453
- EFSA CEF Panel (EFSA Panel on Food Contact Materials, Enzymes, Flavourings and Processing Aids), 2014. Scientific Opinion on Flavouring Group Evaluation 300, Revision 1 (FGE.300Rev1): One cycloaliphatic amide from chemical group 33. EFSA Journal 2014;12(11):3887, 34 pp. doi:10.2903/j.efsa.2014.3887
- EFSA CEF Panel (EFSA Panel on Food Contact Materials, Enzymes, Flavourings and Processing Aids), 2015a. Scientific Opinion on Flavouring Group Evaluation 77, Revision 2 (FGE.77Rev2): Consideration of Pyridine, Pyrrole and Quinoline Derivatives evaluated by JECFA (63rd meeting) structurally related to Pyridine, Pyrrole, Indole and Quinoline Derivatives evaluated by EFSA in FGE.24Rev2 (2013). EFSA Journal 2015;13(1):3997, 58 pp. doi:10.2903/j.efsa.2015.3997
- EFSA CEF Panel (EFSA Panel on Food Contact Materials, Enzymes, Flavourings and Processing Aids), 2015b. Scientific Opinion on Flavouring Group Evaluation 86, Revision 2 (FGE.86Rev2): Consideration of aliphatic and arylalkyl amines and amides evaluated by JECFA (65th meeting). EFSA Journal 2015;13(1):3998, 49 pp. doi:10.2903/j.efsa.2015.3998
- EFSA FEEDAP Panel (EFSA Panel on Additives and Products or Substances used in Animal Feed), 2012a. Guidance for the preparation of dossiers for sensory additives. EFSA Journal 2012;10(1):2534, 26 pp. doi:10.2903/j.efsa.2012.2534
- EFSA FEEDAP Panel (EFSA Panel on Additives and Products or Substances used in Animal Feed), 2012b. Guidance for the preparation of dossiers for additives already authorised for use in food. EFSA Journal 2012;10(1):2538, 4 pp. doi:10.2903/j.efsa.2012.2538
- EFSA FEEDAP Panel (EFSA Panel on Additives and Products or Substances used in Animal Feed), 2012c. Guidance for establishing the safety of additives for the consumer. EFSA Journal 2012;10(1):2537, 12 pp. doi:10.2903/j.efsa.2012.2537
- EFSA FEEDAP Panel (EFSA Panel on Additives and Products or Substances used in Animal Feed), 2012d. Guidance on studies concerning the safety of use of the additive for users/workers. EFSA Journal 2012;10(1):2539, 5 pp. doi:10.2903/j.efsa.2012.2539
- FAO (Food and Agricultural Organization of the United Nations), 2006. FAO JECFA Monographs 1: Combined Compendium of Food Additive Specifications—Joint FAO/WHO Expert Committee on Food Additives—All specifications monographs from the 1st to the 65th meeting (1956–2005). Volume 4. Analytical methods, test procedures and laboratory solutions used by and referenced in the food additive specifications. Food and Agricultural Organization of the United Nations, Rome Italy.
- Gupta RC, 2007. Veterinary toxicology. Basic and clinical principles. Elsevier, Amsterdam, the Netherlands.
- Hawksworth G and Scheline RR. 1975. Metabolism in the rat of some pyrazine derivatives having flavour importance in foods. Xenobiotica, 5, 389–399.
- Ioannides C, 2006. Cytochrome P450 expression in the liver of food-producing animals. Current Drug Metabolism, 7, 335–348.



- James MO and Pritchard JB, 1987. *In vivo* and *in vitro* renal metabolism and excretion of benzoic acid by a marine teleost, the southern flounder. Drug Metabolism and Disposition, 15, 665–670.
- King LJ, Parke DV and Williams RT, 1966. The metabolism of [2-¹⁴C]-indole in the rat. Biochemical Journal, 98, 266–277.
- Martinez BO and Roe DA, 1972. Interrelationships of Sulfate and Glucuronide Conjugation in Indolefed Rats. Journal of Nutrition, 102, 365–374.
- Mendes O, 2012. Draft Report. 2-Acetylpyrrole: A 10-day oral metabolism/toxicity study in rats. Product Safety Labs. Study no. 34496. June 11-21, 2012. Unpublished report submitted by EFFA to FLAVIS Secretariat.
- Nebbia C, Dacasto M, Rossetto Giaccherino A, Giuliano Albo A and Carletti M, 2003. Comparative expression of liver cytochrome P450-dependent monooxygenases in the horse and in other agricultural and laboratory species. Veterinary Journal, 165, 53–64.
- Pan HP and Fouts J, 1978. Drug metabolism in birds: Part 2. Drug Metabolism Reviews, 39, 194–196.
- Skiles GL, Appleton ML, Smith DJ, Carlson JR and Yost GS, 1991. Isolation of a mercapturate adduct produced subsequent to glutathione conjugation of bioactivated 3-methylindole. Toxicology and Applied Pharmacology 108, 531–537.
- Skiles GL and Yost GS, 1992. Stable-isotope mechanistic studies on the oxidation of 3-methylindole. Toxicologist, 121, 289–295.
- Smith DJ, Skiles GL, Appleton ML, Carlson JR, Yost GS, 1993. Identification of goat and mouse urinary metabolites of the pneumotoxin, 3 methylindole. Xenobiotica, 23, 1025–1044.
- Til HP and van der Meulen HC, 1971. Subchronic (90-day) toxicity study with 2-acetylpyridine in albino rats. Centraal Instituut Voor Voedingsonderzioek, Netherlands. Report no. R3373. February, 1971. Unpublished report submitted by EFFA to FLAVIS.
- WHO (World Health Organization), 2005. Evaluation of certain food additives. Sixty-third report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series, no. 928. Geneva, 8–17 June 2004.
- WHO (World Health Organization), 2006. Evaluation of certain food additives. Sixty-fifth report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series, no. 934. Geneva, 7–16 June 2005.
- WHO (World Health Organization), 2008. Joint FAO/WHO Expert Committee on Food Additives. Sixtyninth meeting, Geneva, 17–26 June 2008. Summary and conclusions issued 4 July 2008.
- Wolf JC and Wolfe MJ, 2005. A brief overview of nonneoplastic hepatic toxicity in fish. Toxicologic Pathology, 33, 75–85.



Abbreviations

ADME	absorption, distribution, metabolism and excretion
bw	body weight
CAS	Chemical Abstracts Service
CD	Commission Decision
CEF	EFSA Scientific Panel on Food Contact Materials, Enzymes, Flavourings and Processing
	Aids
CG	chemical group
CDG	chemically defined group
DM	dry matter
DT ₅₀	degradation half-time
EC	European Commission
EC ₅₀	half-maximal effective concentration
ECOSAR	component program of EPI suite [™]
EPI suite	Estimation Programs Interface (EPI) Suite [™]
EURL	European Union Reference Laboratory
FAO	Food and Agriculture Organization
FEEDAP	EFSA Scientific Panel on Additives and Products or Substances used in Animal Feed
FFAC	Feed Flavourings authorisation Consortium of (FEFANA) the EU Association of
	Specialty Feed Ingredients and their Mixtures
FGE	Flavouring Group Evaluation
FID	flame ionisation detector
FLAVIS	the EU Flavour Information System
FL-No	FLAVIS number
GC-MS	gas chromatography–mass spectrometry
JECFA	The Joint FAO/WHO Expert Committee on Food Additives
K _{oc}	organic carbon sorption constant
K _{ow}	octanol-water partition coefficient
LC ₅₀	lethal concentration 50
log K _{ow}	logarithm of octanol-water partition coefficient
NOAEL	no observed adverse effect level
OECD	Organisation for Economic Co-operation and Development
PEC	predicted environmental concentration
PEC _{swaq}	predicted environmental concentration of the additive (parent compound) in surface
	water
RTL	retention time locking
SMILES	Simplified Molecular Input Line Entry Specification
TNO	Netherlands Organisation for Applied Scientific Research
TTC	threshold of toxicological concern
UF	uncertainty factor
WHO	World Health Organization



Annex – Executive Summary of the Evaluation Report of the European Union Reference Laboratory for Feed Additives on the Method(s) of Analysis for Pyridine, pyrrole, and quinoline derivative

The *Chemically Defined Flavourings* – *Group 28* (*Pyridine, pyrrole and quinoline derivatives*), in this application comprises nine substances, for which authorisation as feed additives is sought under the category 'sensory additives', functional group 2(b) 'flavouring compounds', according to the classification system of Annex I of Regulation (EC) No 1831/2003.

In the current application submitted according to Article 4(1) and Article 10(2) of Regulation (EC) No 1831/2003, the authorisation for all species and categories is requested. The flavouring compounds of interest have a purity ranging from 95% to 99%.

Mixtures of flavouring compounds are intended to be incorporated only into *feedingstuffs* or drinking *water*. The Applicant suggested no minimum or maximum levels for the different flavouring compounds in *feedingstuffs* or in *water*.

For the identification of volatile chemically defined flavouring compounds *chemically defined group (CDG) 28* in the *feed additive,* the Applicant submitted a qualitative multianalyte gas chromatographymass spectrometry (GC–MS) method, using retention time locking (RTL), which allows a close match of retention times on GC–MS. By making an adjustment to the inlet pressure, the retention times can be closely matched to those of a reference chromatogram. It is then possible to screen samples for the presence of target compounds using a mass spectral database of RTL spectra. The Applicant maintained two FLAVOR2 databases/libraries (for retention times and for MS spectra) containing data for more than 409 flavouring compounds. These libraries were provided to the European Union Reference Laboratory (EURL). The Applicant provided the typical chromatogram for the *CDG 28* of interest.

In order to demonstrate the transferability of the proposed analytical method (relevant for the method verification), the Applicant prepared a model mixture of flavouring compounds on a solid carrier to be identified by two independent expert laboratories. This mixture contained twenty chemically defined flavourings belonging to twenty different chemical groups to represent the whole spectrum of compounds in use as feed flavourings with respect to their volatility and polarity. Both laboratories properly identified all the flavouring compounds in all the formulations. As the substances of *CDG 28* are within the volatility and polarity range of the model mixture tested, the Applicant concluded that the proposed analytical method is suitable to determine qualitatively the presence of the substances from *CDG 28* in the *mixture of flavouring compounds*.

Based on the satisfactory experimental evidence provided, the EURL recommends for official control for the qualitative identification in the *feed additive* of the individual (or mixture of) *flavouring compounds* of interest listed in Table 1 (*) the GC–MS–RTL (Agilent specific) method submitted by the Applicant.

For the identification of *piperine* in the *feed additive*, the Applicant proposed a single laboratory validated and further verified method based on GC coupled to a flame ionisation detector (GC-FID). Based on the satisfactory performance characteristics presented, the EURL recommends for official control the single laboratory validated and further verified GC-FID method, submitted by the Applicant, for the qualitative identification of *piperine* in the *feed additive*.

As no experimental data were provided by the Applicant for the identification of the *active substance(s)* in *feedingstuffs* and *water*, no methods could be evaluated. Therefore, the EURL is unable to recommend a method for the official control to identify the *active substance(s)* of interest listed in Table 1 (*) in *feedingstuffs* or *water*.

Further testing or validation of the methods to be performed through the consortium of National Reference Laboratories as specified by Article 10 (Commission Regulation (EC) No 378/2005) is not considered necessary.

(*) Full list provided in EURL evaluation report, available from the EURL website.