# NANOPARTICLES BASED ON FRUCTOSE AND ALKALY\_EARTH HALOGENIDES WITH SECOND HARMONIC GENERATION PROPERTIES FOR APPLICATIONS AS **BIO-SENSORS AND FOR RADIOTHERAPY** UNIVERSITÀ UNIVERSITÀ degli Studi DEGLI STUDI di Milano DI TORINO

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In recent years, some Metal Organic Frameworks (MOFs) with Second Harmonic Generation (SHG) properties, based on fructose and alkali-earth alogenides, were investigated to understand the effect of cation size and anion polarizability on crucial quantities correlated to the non-linear optical (NLO) response, such as hyperpolarizability and optical susceptibility [1,2]. The compounds studied are interesting for biomedicine applications, as they combine high biocompatibility, due to their non-toxic components, and significant SH emission, that can permit exploitation for *in vitro* bio-imaging.

Results from our previous work [2] (figure 1) suggested that the SH efficiency is much influenced by the anion, the first static hyperpolarizability and second order susceptibility being higher for bromide compounds with respect to those with chloride.

In order to enhance the SH efficiency of our systems, we decided to synthetized three new MOFs with iodide as anion, instead of bromide and chloride. In particular we obtained three MOFs based on fructose and  $Srl_2$  salts, of formula  $[Sr(fructose)_2]l_2$ ,  $[Sr_2(fructose)_3]l_4 \cdot H_2O$  and [Sr(fructose)(H<sub>2</sub>O)<sub>3</sub>I]I (figure 2), that contain quite the same building blocks but show different structural arrangements. The compounds were characterized by single-crystal and powder XRD, and the first static hyperpolarizability and second-order susceptibility were estimated from theoretical calculations, both in vacuo and in the solid state (table 1) [3], and measured with the Kurtz and Perry method.



Figure 1: Isomorphous structure of the four compounds of formula  $[M(fructose)_2(H_2O)_2]X_2 \cdot H_2O, M=Ca,Sr and X=Cl, Br [2].$ 



**Table 1.** Crystal data and preliminary theoretical results on the fructose-Srl<sub>2</sub> based MOFs.

	[Sr(fructose) <sub>2</sub> ]I <sub>2</sub>	[Sr <sub>2</sub> (fructose) <sub>3</sub> ]I <sub>4</sub> ·H <sub>2</sub> C	D [Sr(fructose)(H <sub>2</sub> O) <sub>3</sub> I]I	Theoretical c	alculation h	has beer	n obtaine	d at the PB	BEO level wi	ith 3-21G sp	lit valence	e basis set. [4]				
Lattice	Monoclinic	Ortorhombic	Ortorhombic			Evolo	niting sym	nmetry in s	olid-state s	simulations	(a ot)	$(2) \cdot (2 \cdot 3) (2 \cdot 3)$	$(2) - x^{(2)}$			
Space group	P2 <sub>1</sub>	P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>			слрк	Jillig Syll	interry in 5	onu-state s	Sinuations	$(t \cdot \mathbf{S}^{t})$	·X··:(a·)(a·k	$() = \chi_{ijk}$			
Cell dimensions	7.8592	12.371685	9.119233	Orthorhombic 222 $xyz, xzy, xzy, xzy, xzy, yzz, yzz, yzz,$												
	12.9355	17.435172	13.090843			$z_{22y}, z_{xy}, z_{yx}, y_{2x}, y_{2x}, y_{2y}, z_{yx}$										
	9.9504	17.660701	13.550428		[Cu/function	- \ 11								$[C_n/f_{n_1}, \dots, f_{n_n}, \infty)/l$		
	90	90	90	COMPONENT	EST(Tructos BETA	e) <sub>2</sub> JI <sub>2</sub> CHI(2)	d(MKS)	d(cgs)	COMPONEN	ISr <sub>2</sub> (fructo	$(2)_{3} JI_{4} \cdot H_{2} O$ CHI(2)	d(MKS) d(cgs)	COMPONENT	EST(fructose)(F BETA	$(1_2O)_3(J)$ CHI(2) d(N	1KS) d(cgs)
	92.803	90	90	XXY	569.3482	0.5247	0.5102	0.0122	XYZ	280.402	13 0.0685	0.0666 0.0016	XYZ	-178.0043	-0.1025 -0.0	996 -0.002
	90	90	90	XYZ	-422.3184	-0.3892	-0.3784	-0.009								
Cell volume	1010.37	3809.46	1617.63	YZZ	-167.1856	-0.7009	-0.6815	-0.018								
Density calc (g cm <sup>-3</sup> )	2.307	2.225	2.364													
μ (mm <sup>-1</sup> )	28.24	29.80	7.179													
Asymmetric unit	Sr <sup>2+</sup>	2 Sr <sup>2+</sup>	Sr <sup>2+</sup>		No.		13									
content	2 FRU	3 FRU	FRU										Substruct	ure decompos	ition of Srfru	ui1
	2 I <sup>-</sup>	3 H <sub>2</sub> O	3 H <sub>2</sub> O				Į Į Į				•					
		4I <sup>-</sup>	2 I⁻		• <del>•</del> •	=	b tak		• •	+	b A		<ul> <li>beta ter</li> <li>if estim</li> </ul>	ms are not pa	irwise addit	IVE are high
		1 H <sub>2</sub> O <sub>crist</sub>			· <del>∳</del> ·ŧ·ŧ·						c 👞		for a g	given specie,	it is reas	onable to
$\beta_{tot}$ <sup>(1)</sup> (10 <sup>-30</sup> cm <sup>5</sup> esu <sup>-1</sup> )	27.0	18.0	14.7										assume	that the cor	ntribution to	o BETA of
χ <sup>(2)</sup> pmV <sup>-1</sup>	5.6	3.2	ND		r.e.		. • • •		<b>•</b> -				that spe	ecie will be re tructure	elevant as v	vell in the
I/I <sub>sucr</sub> (measured) <sup>(2)</sup>	0.7	0.05			COM	PONENT	BETA	CHI(2) d(M	IKS) d(cgs)	COMPONENT	BETA	CHI(2) d(MKS) d(cgs	• Results	the sugar	provides th	ne largest
(1) Coloulated at the D2UVD/C21C(d) lough of the comparison OC						XXY XYZ	140.061 -16.3986 -	0.1291 0.12 -0.0151 -0.0	255 0.003 147 -0.0004	XXY XYZ	-0.4875 - 0.5208 (	0.0004 -0.0004 0	contribu	ution		
<sup>(2)</sup> Measured values obtained from powders, with the Kurtz and Perry method.						YYY	370.3652	0.3413 0.33	319 0.0079	YYY	4.6095	0.0042 0.0041 0.000	01			
						YZZ	-321.518 -	-0.2963 -0.23	881 -0.0069	YZZ	-0.1603 -	0.0001 -0.0001 0				

**Periodic LCGTF calculations (CRYSTAL14)** 



Orthorhombic	
222	xyz, xzy, yzx, yxz, zxy,

#### Synthesys of nanoparticles.

Nanoparticles of  $[Sr(fructose)_2(H_2O)_2]Cl_2 H_2O$  were obtained by fast precipitation of crystals from isobutanol solution, dryed in a stove at 70°C and grinded in a bill mill. Then the nanoparticles were proofed through reaction of the –OH groups on surface with dodecanoyl chloride and encapsulated in a phospholipidic m-PEG shell. They were characterized by Dynamic light scattering technique (DLS), showing a narrow band centered at ca. 245 nm (polidispersity 0.005).

# Applications

### Cell uptake and viability tests.

Preliminary activity studies on target cells are in progress. Nanoparticles were tested on two cell lines, hTERT-HME1 (breast epithelial cells immortalized with hTERT) and HT-29 (colorectal adenocarcinoma) to evaluate cellular uptake and viability. To study cellular uptake, nanoparticles were loaded with

1100.00 hTERT-HME1 HT-29 **2** 700,00 -500,00 -300,00 -100,00 1:1000 1:100 1:1000 1:100 1:10



Next step provide for the functionalization of the nanoparticles surfice in order to direct them towards specific cancer cells.

#### References

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[4] J. Stephen Binkley, John A. Pople, Warren J. Hehre, J. Am. Chem. Soc, 102, **1980**, 939; K. D. Dobbs, W. J. Hehre, J. Comput. Chem. 8, **1987**, 880

Table 2. Mean diameter (nm) and multimodal size distribution of nanoparticles determined by DLS.



fluorescein (1 mg/20 mg of the compound) before treatment with dodecanoyl chloride and their internalization was expressed as median fluorescence intensity. Preliminary data suggested that cellular uptake was dose- and time-dependent.

Cell viability was evaluated after 1, 2 and 5 days of treatment and was measured as ATP production.

Nanoparticles inhibited the growth of cells by about 50% at the maximum concentration after 5 days of treatment.



## **Radiotherapy applications.**

Applications in radiotherapy of MOFs based on the co-crystallization of fructose and salts containing various radionuclides are presently under investigation. For example, <sup>89</sup>Sr, is a  $\beta$  emitter with a half-life of 50.57 d obtained by nuclear fission and used in the form of chloride for metastasized bone cancer. The average energy of the  $\beta$  emitted by <sup>89</sup>Sr is 0.58 MeV. Based on a linear energy transfer in water of 1.75 MeV cm<sup>-1</sup>, we calculate an average range of 3.3 mm and we assume a tumor size at least one order of magnitude larger. Under this hypothesis, the estimated dose delivered by functionalized particles of  $[Sr(fructose)_2(H_2O)_2]Cl_2 H_2O$  of radius 0.5  $\mu$ m prepared with commercial strontium chloride with specific activity 4.57 MBq/mg would be 23.7 Gy. However, the selectivity towards tumor cells would be greatly enhanced with respect to strontium chloride itself.