

REGULAR RESEARCH ARTICLE

Absolute Measurements of Macrophage Migration Inhibitory Factor and Interleukin-1 β mRNA Levels Accurately Predict Treatment Response in Depressed Patients

Annamaria Cattaneo, PhD; Clarissa Ferrari, PhD; Rudolf Uher, MD;
Luisella Bocchio-Chiavetto, PhD; Marco Andrea Riva, PhD; the MRC
ImmunoPsychiatry Consortium, and Carmine M. Pariante, MD, FRCPsych, PhD

Stress, Psychiatry and Immunology Laboratory, Department of Psychological Medicine, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, United Kingdom (Drs Cattaneo and Pariante); Biological Psychiatry Unit, IRCCS Fatebenefratelli Institute, Brescia, Italy (Dr Cattaneo); Statistical Service, IRCCS Fatebenefratelli Institute, Brescia, Italy (Dr Ferrari); Genetic Unit, IRCCS Fatebenefratelli Institute Brescia, Italy and Faculty of Psychology, eCampus University, Novedrate, Como, Italy (Dr Bocchio-Chiavetto); Department of Psychiatry, Dalhousie University, Halifax, Nova Scotia, Canada (Dr Uher); The Social, Genetic, and Developmental Psychiatry Centre, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, United Kingdom (Dr Uher); Department of Pharmacological and Biomolecular Sciences, University of Milan, Milan, Italy (Dr Riva).

Correspondence: Dr. Annamaria Cattaneo, MSC, PhD, Stress, Psychiatry and Immunology Laboratory (SPI-Lab), Department of Psychological Medicine, Institute of Psychiatry, Psychology and Neuroscience, King's College London, 125 Coldharbour Lane, London, SE5 9NU, UK (annamaria.cattaneo@kcl.ac.uk).

Abstract

Background: Increased levels of inflammation have been associated with a poorer response to antidepressants in several clinical samples, but these findings have had been limited by low reproducibility of biomarker assays across laboratories, difficulty in predicting response probability on an individual basis, and unclear molecular mechanisms.

Methods: Here we measured absolute mRNA values (a reliable quantitation of number of molecules) of Macrophage Migration Inhibitory Factor and interleukin-1 β in a previously published sample from a randomized controlled trial comparing escitalopram vs nortriptyline (GENDEP) as well as in an independent, naturalistic replication sample. We then used linear discriminant analysis to calculate mRNA values cutoffs that best discriminated between responders and nonresponders after 12 weeks of antidepressants. As Macrophage Migration Inhibitory Factor and interleukin-1 β might be involved in different pathways, we constructed a protein-protein interaction network by the Search Tool for the Retrieval of Interacting Genes/Proteins.

Results: We identified cutoff values for the absolute mRNA measures that accurately predicted response probability on an individual basis, with positive predictive values and specificity for nonresponders of 100% in both samples (negative predictive value=82% to 85%, sensitivity=52% to 61%). Using network analysis, we identified different clusters of targets for these 2 cytokines, with Macrophage Migration Inhibitory Factor interacting predominantly with pathways involved in neurogenesis, neuroplasticity, and cell proliferation, and interleukin-1 β interacting predominantly with pathways involved in the inflammasome complex, oxidative stress, and neurodegeneration.

Received: January 5, 2016; Revised: April 26, 2016; Accepted: April 28, 2016

© The Author 2016. Published by Oxford University Press on behalf of CINP.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Conclusion: We believe that these data provide a clinically suitable approach to the personalization of antidepressant therapy: patients who have absolute mRNA values above the suggested cutoffs could be directed toward earlier access to more assertive antidepressant strategies, including the addition of other antidepressants or antiinflammatory drugs.

Keywords: cytokine absolute blood levels, treatment response, predictors, personalized medicine

Introduction

While there is evidence that patients with high peripheral inflammation tend to respond less to conventional antidepressants, we lack biomarkers that are reproducible across laboratories, predict response probability on an individual basis, and have clear molecular mechanisms underlying their predictive effect. The identification of biomarkers that predict treatment response is crucial in reducing the social and economic burden of depression and improving quality of life of patients. Indeed, more than one-half of patients fail to show an adequate response to first-line antidepressants (Trivedi, 2006), and one-third of patients are resistant to all available pharmacological treatments (Rush et al., 2003); therefore, there is a need to establish personalized treatment protocols that can accelerate the escalation toward adjuvant pharmacological strategies in those deemed less likely to respond. The notion of using peripheral inflammation to personalize treatment in depression is supported by a recent randomized controlled trial with the tumor necrosis factor (TNF)- α antagonist, infliximab, providing some evidence that patients with high levels of inflammation are more likely to respond to an adjuvant treatment with antiinflammatory drugs (Raison et al., 2013).

Baseline concentrations of inflammatory markers, like C Reactive Protein (CRP) and circulating cytokines, have been proposed as useful biomarkers for the identification of patients that will fail to respond to antidepressants. While depressed patients in general tend to show higher blood levels of inflammatory biomarkers compared with controls (Howren et al., 2009; Martinez et al., 2012), depressed patients who are resistant to conventional antidepressants tend to have even higher concentrations of these biomarkers, both as plasma/serum proteins (Sluzewska et al., 1997; Lanquillon et al., 2000; Fitzgerald et al., 2006; Rethorst et al., 2013) and as blood mRNA levels (Carvalho et al., 2013; Cattaneo et al., 2013; Lisi et al., 2013; Powell et al., 2013). However, despite this large amount of evidence, none of these findings have been translated yet into clinical practice, partly because different studies use different biomarkers, often measured with assays that are laboratory specific. Moreover, these assays have relative rather than absolute validity: that is, they can separate 2 groups defined as responders/nonresponders but do not necessarily predict the response probability on an individual basis. The recent study using CRP to predict differential response to nortriptyline vs escitalopram is an important step forward, because CRP is a commonly available and standardized test, but it lacks the molecular insight, since CRP is the final outcome of a number of inflammatory pathways (Uher et al., 2014).

Here we propose that the absolute values of Macrophage Migration Inhibitory Factor (MIF) and interleukin (IL)-1 β blood mRNA molecules can be used to accurately predict antidepressant treatment response across different laboratories, because absolute mRNA values are more likely to be comparable independently from the laboratory setting because of the use of standard quantitation. In this study, we build on our previous work in the Genome-Based Therapeutic Drugs for Depression (GENDEP) sample, a part-randomized study with 2 active pharmacological

treatment arms with nortriptyline vs escitalopram, which has been extensively described before (Uher et al., 2009, 2010; Keers et al., 2010). In our previous report (Cattaneo et al., 2013), we measured the blood mRNA relative expression levels of cytokines, that is, we normalized the levels of each cytokine vs the levels of internal controls (housekeeping genes). Of the many cytokines assessed (IL1 α , IL-1 β , IL-4, IL-6, IL-7, IL-8, IL-10, MIF, and TNF- α), only the 3 proinflammatory cytokines, IL-1 β , MIF, and TNF- α , were higher in patients who later did not respond to antidepressants compared with those who did. In the present paper, we aim to: (1) select the strongest predictors (among the 3 cytokines) using multivariate logistic regression model; (2) identify the absolute mRNA values (number of molecules) cutoffs that best allocate individuals to the responders and nonresponders classes; (3) validate the same absolute mRNA values cutoffs in an independent sample recruited in a naturalistic setting; and (4) conduct a network analyses to assess the main targets of these proinflammatory cytokines, thus contributing to mechanistic understanding.

Materials and Methods

Study Design and Sample

GENDEP Study

The GENDEP project is an open-label, part-randomized, multicenter pharmacogenetic study with 2 active pharmacological treatment arms that has been extensively described before (Uher et al., 2009, 2014; Keers et al., 2010; Powell et al., 2012; Cattaneo et al., 2013). For the main study, 811 adults with unipolar major depression of at least moderate severity according to both the ICD-10 (World Health Organization, 1992) and the DSM-IV (American Psychiatric Association, 1994) were recruited and randomly allocated to receive flexible dosage of nortriptyline (50–150 mg daily) or escitalopram (10–30 mg daily) for 12 weeks. Other psychotropic medications were not allowed, with the exception of occasional use of hypnotics. Response to antidepressant medication was quantified as percentage reduction in the Montgomery–Åsberg Depression Rating Scale (MADRS) score from baseline to week 12, and responders were identified as patients with a reduction in MADRS > 50%; according to this definition, approximately 55% of patients in this sample were classified as responders (Uher et al., 2009). Written informed consent was obtained from all participants, and the study was approved by the local ethics committee.

For the present study, we selected all patients who had been drug free for at least 2 weeks before entering into the trial and provided a baseline blood PaxGene tube for mRNA gene expression analysis ($n=74$). On average, they were in their second episode of moderately severe depression and scored, at baseline, 28.7 (± 4.2) on the MADRS; according to the percent change in the MADRS score, 69% (51 of 74) were defined as responders (the selective inclusion of drug free patients may have led to a slightly more antidepressant-responsive group compared with

the total sample). The main demographic and clinical features are summarized in **Table 1**. There were no significant differences between patients treated with escitalopram ($n=38$) or nortriptyline ($n=36$) in age (38 ± 12.4 vs 36 ± 9.4 , $P=.25$), gender (F/M was 20/18 vs 23/13, $P=.2$), or response rate (responders/nonresponders were 26/12 vs 25/11, $P=.6$).

Validation Sample

A second independent sample of $n=68$ depressed patients was recruited within a European multicenter collaboration on depression and analysed at the IRCCS Fatebenefratelli Brescia. The study was approved by the local Ethic Committees of the Institutes. Diagnosis of depression was confirmed by clinical interviews using the Structured Clinical Interview for DSM-IV, and severity (and treatment response) was assessed using the MADRS at baseline and at week 12. The mean (SD) MADRS score at baseline was $29.5 (\pm 3.9)$; using the same criteria used in the GENDEP sample to classify responders and nonresponders, 66% (45 of 68) were defined as responders. At baseline, most patients were drug free; they then started pharmacological treatment with clinician-determined antidepressant drugs: 20 took an selective serotonin reuptake inhibitors (SSRI) (mostly escitalopram and paroxetine), 20 a serotonin and noradrenaline reuptake inhibitor (mostly duloxetine and venlafaxine), and 14 a tricyclic (mostly amitriptyline and desipramine). Participants were excluded if they were taking antipsychotics or mood stabilizers, or if they had a history of neurological or comorbid psychiatric disorders (Axis I or Axis II), substance abuse, or severe medical illness or head injury. Written informed consent was obtained from all participants, and the study was approved by the local ethics committee. Demographic and clinical features are summarized in **Table 1**.

Blood Sample Collection and RNA Isolation

In both clinical groups, blood sample collection for gene expression analyses in leukocytes was performed at baseline (before starting antidepressant treatment) using PaxGene tubes. After blood collection, PaxGene tubes were kept for 2 hours at room temperature and then stored at -20°C for 24 hours and then at -80°C until their processing. RNA isolation was performed using the PaxGene Blood RNA Kit (Qiagen) according to the manufacturer's protocols. The RNA quantity was assessed by evaluation of the A260/280 and A260/230 ratios using a Nanodrop spectrometer (NanoDrop Technologies, Wilmington, DE), and RNA quality was determined using an Agilent Bioanalyzer (Agilent Technologies). RNA samples were then stored at -80°C until their processing for gene expression analyses.

Generation of Absolute mRNA Molecule Number through External Calibration Curves

After the initial multivariate logistic regression analysis (see Results below), which identified MIF and IL-1 β as the strongest predictors of treatment response in the GENDEP sample, subsequent quantification analyses concentrated on only these 2 cytokines. We performed an absolute gene expression analyses to get absolute levels of cytokines mRNA expression, a novel

approach that does not require normalization with "housekeeping" genes and is more likely to be comparable across different laboratories because of the use of standard quantitation. cDNA clones for human MIF and IL-1 β were available from Origene (MIF NM_002415 and NM_000576, 10 μg). Purified plasmid clones were then quantified using the PicoGreen method. Knowing the copy numbers and the concentration of plasmid cDNA, the precise number of molecules added to real time-PCR runs can be calculated providing a standard for specific cDNA quantification. Oligonucleotides specific for MIF and IL-1 β , as well as 3 housekeeping genes (β -actin, β 2-microglobulin [B2M], and GAPDH) were designed with Primer 3 software (<http://www.bioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi>) and used in subsequent real time reactions. Primer and probes sequences were: for β -actin, FW primer: CACACGCAGCTATTGTAGA, RW primer: GGCATGGGTCAGAAGGATT, probe: GAAATCTGGCACACACCT; for B2M, FW primer ATGCCCTGCCGTGTGAACCATGT, RW primer: TCAACCCTCCATGATGCTGCT, probe: CACTGAATTCA CCCACTGA; for GAPDH, FW primer TTCACACCCATGACGAACAT, RW primer: CGAGATCCCTCCAAAATCAA, probe: GCCAAAGGGTC ATCATCTC; for IL-1 β , FW primer GGAGAATGACCTGACCACCT, RW primer: GGAGGTGGAGAGCTTCAGT, probe: ACGATGCACC TGTACGATCA; for MIF, FW primer CGCAGAACCGCTCTACAG, RW primer: AGGCGAAGGTGGAGTTGTT, probe: CGGGACAGGGTCTACATCAA. For each primer and probe set, we ran a standard curve whose slope was used to calculate the PCR efficiencies, which were $E_{\text{MIF}} = 2.0$ and $E_{\text{IL-1}\beta} = 1.9$. Real time PCR runs were performed in triplicate, with 1 μL of template (10 ng of RNA) unknown cDNA from patients, or 1 μL of clone cDNA (8 dilution points ranging from 1e03 to 1e06), in a final volume of 2.5 μL .

The standard curve was built up with external clones containing MIF or IL-1 β mRNA. Specifically, a known amount of plasmid was used to construct a calibration curve, and then unknown samples (mRNA from patients) were quantified as number of mRNA molecules (Li and Wang, 2000). After having checked for the efficiency of the assays, standard curves for MIF and IL-1 β were run together (in the same reaction plate) with the mRNA samples of patients, and the threshold cycle (Ct) values of MIF or IL-1 β for samples from patients were compared with the Ct values of the dilution points of the standard curve to get an absolute value of expression of each cytokine (total number of MIF or IL-1 β mRNA molecules per nanogram total RNA) in each patient. In particular, a standard curve was obtained by plotting the natural log of the Ct against the natural log of the number of molecules.

To ensure that any differences in absolute levels of cytokines mRNA expression were not due to generic differences in the levels of mRNA, we measured the mRNA levels of 3 housekeeping genes in both independent samples, and we then expressed the values in term of relative expression Ratio, with "responders" as reference group.

MIF and IL-1 β Network Analysis

As MIF and IL-1 β might be involved in different pathways, we constructed a protein-protein interaction network by using

Table 1. Demographic and Clinical Information for the Two Samples of Depressed Patients

	Age	Gender (% Female)	Baseline MADRS	Responders (%)
GENDEP sample ($n=74$)	38.3 ± 10.9	58.1% (31M/43F)	28.7 ± 4.2	69% (51/74)
Validation sample ($n=68$)	39 ± 9.5	52.9% (32M/36F)	29.5 ± 3.9	66% (45/68)

the Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) Network Analyses software (<http://string-db.org/>) to identify the closest interacting genes and the main networks that are regulated.

Statistical Analyses

Preliminary descriptive analysis, based on Mann-Whitney nonparametric test (for non-Gaussian distributions), was carried out to assess any significant differences in mRNA relative cytokines between the responder and nonresponder categories. Initially, a multiple logistic regression model was carried out in the GENDEP sample through the backward Wald method for the selection of the strongest predictors of treatment response using cytokines relative mRNA values of IL-1 β , MIF, and TNF- α . Subsequently, univariate logistic models were performed to evaluate the associations between selected biomarkers with treatment response. The absolute values of the selected cytokines (numbers of molecules per nanogram total RNA) were then calculated, as described above. Subsequently, we used a linear discriminant analysis (LDA) to analyze the linear combination of the absolute values of the cytokines in relationship with treatment response to define a rule able to discriminate responders or nonresponders (Mc Lachlan, 2004). The resulting combination, named linear discriminant function (LDF), was then used as a linear classifier able to separate patients according to the scores of this function, indicating the probability of each individual of belonging to the responder or nonresponder classes based on cutoff values of MIF and IL-1 β mRNA molecules. Finally, to validate these findings, we assessed the performance of the same cutoff values (numbers of molecules) on generating probability scores in the second validation sample. Positive and negative predictive values (PPV, NPV), specificity, and sensitivity were calculated in both samples for the probability of being a nonresponder. Statistical analyses were carried out with SPSS 21.0 and R language and environmental v.3.0.3. (R Development Core Team, 2013). Statistical significance was set at $P < .05$.

Results

Baseline mRNA relative Expression of MIF and IL-1 β , but Not TNF- α , Predicts Treatment Response in the GENDEP Sample

As mentioned above, we had previously found that nonresponders in the GENDEP sample had higher baseline mRNA relative expression levels of 3 proinflammatory cytokines, IL-1 β , MIF, and TNF- α , compared with responders; these findings were present for both escitalopram and nortriptyline nonresponders (Cattaneo et al., 2013). In those analyses, we did not apply any variable selection procedure; however, we found that TNF- α had the lowest predicting power (adjusted $R^2 = 0.19$) compared with both IL-1 β (adjusted $R^2 = 0.31$) and MIF (adjusted $R^2 = 0.37$) (Cattaneo et al., 2013).

In the present study, we detected the biomarkers that were more strongly associated with treatment response by using a multiple logistic regression model applying the backward Wald method for the variable selection, which automatically removes variables that do not add more significance to the model. We found that MIF and IL-1 β were strongly associated with treatment response: the odd ratios (ORs) of being a nonresponder vs being a responder (obtained by univariate logistic models) were OR = 1.32 (95% CI 1.04–1.70, $P = .034$) for MIF, and OR = 1.24 (95% CI 1.02–1.54, $P = .049$) for IL-1 β , with higher levels predicting lack of response. Moreover, the power of the model increased (with an R^2 of Nagelkerke index increasing from 0.86 to 0.95) when IL-1 β was evaluated together with MIF in the same logistic regression

model (OR = 1.44, $P = .030$). TNF- α , in contrast, did not significantly increase the predictive power ($P > .05$) and was removed from the model by backward selection method. Of note, the ORs were similar when we analyzed separately the response prediction to escitalopram (OR = 1.34, $P = .029$ for MIF, and OR = 1.29, $P = .035$ for IL-1 β) and to nortriptyline (OR = 1.35, $P = .038$ for MIF, and OR = 1.20, $P = .045$ for IL-1 β), indicating that these cytokines predict treatment response to both antidepressants.

Baseline mRNA Absolute Expression of MIF and IL-1 β Accurately Discriminate between Responders and Nonresponders at the Individual Level

We measured the absolute blood mRNA expression of MIF and IL-1 β in the GENDEP sample in order to get absolute (and not relative) mRNA measurements. Overall, means \pm SEM in the whole sample were $66.8 \pm 3.5 \times 10^6$ for IL-1 β and $130.2 \pm 5.2 \times 10^6$ for MIF. Consistently with our previous findings using relative gene expression approach (Cattaneo et al., 2013), we found that the mean absolute number of molecules for MIF and IL-1 β were significantly higher in nonresponders compared with responders (mean \pm SEM; IL-1 β : $83.1 \pm 4.8 \times 10^6$ in nonresponders vs $50.4 \pm 2.1 \times 10^6$ in responders, $t = 11.7$, DF = 72, $P < .001$; MIF: $102.5 \times 10^6 \pm 4.2$ in nonresponders vs $55.4 \times 10^6 \pm 1.9$ in responders, $t = 7.2$, DF = 72, $P < .001$).

We then used an LDA to analyze the linear combination of the absolute values of MIF and IL-1 β in relationship with treatment response to define a rule able to discriminate between responders and nonresponders (Mc Lachlan, 2004). Specifically, we first used LDA to calculate the coefficients for each biomarker (resulting in 0.056 for MIF and 0.036 for IL-1 β , respectively) and then used the LDF to calculate the probability of each individual of belonging to the responder or nonresponder classes based on cutoff values of MIF and IL-1 β mRNA molecules (Table 2). The table shows the relative probability of being a responder (in green) or nonresponder (in red) based on the LDF and according to the quantitative chosen cutoff values. The top-left cells represent patients with the lowest levels of inflammation and the highest probability of being a responder, virtually equal to 100%; with increasing levels of either or both cytokines, the probability of being a responder decreases dramatically, with bottom-right cells being characterized by a probability of being a responder virtually equal to zero. Specifically, green (responder) indicates >73% probability of being a responder and <27% probability of being a nonresponder; red (nonresponder) indicates <32% probability of being a responder and >72% probability of being a nonresponder; and orange indicates intermediate values. Based on these criteria, this sample had $n = 35$ (47%) of subjects classified as responders ($MIF \leq 60 \times 10^6$ and $IL-1\beta \leq 85 \times 10^6$), $n = 23$ (31%) of subjects classified as nonresponders ($MIF > 95 \times 10^6$ and $IL-1\beta > 50 \times 10^6$), and $n = 16$ (22%) of subjects classified as intermediates ($MIF = 60 \times 10^6$ – 95×10^6 and $IL-1\beta = 50 \times 10^6$ – 85×10^6).

We then calculated PPV, NPV, sensitivity, and specificity for being a nonresponder using the specific cutoff values ($MIF > 95 \times 10^6$ and $IL-1\beta > 50 \times 10^6$; red cells in the table). 100% of those classified as nonresponders ($n = 14$) were true nonresponders (PPV = 100%), and all true responders ($n = 51$) were identified as responders (specificity = 100%). This is ideal in terms of clinical practice: using this approach, only true nonresponder patients would be exposed to a more rapid or assertive antidepressant strategies (see also Discussion). Nine (39%) of nonresponders were not identified as such (NPV = 85%, sensitivity = 61%): in terms of clinical practice, they would continue to receive standard treatment.

We found no difference in the expression levels of the 3 housekeeping genes in this sample; mean \pm SEM in responders

vs nonresponders were: β -actin: 1.08 ± 0.7 vs 1.09 ± 0.6 ; B2M: 1.11 ± 0.8 vs 1.11 ± 0.4 ; GAPDH: 1.03 ± 0.4 vs 1.09 ± 0.6 (all $P > .05$).

MIF and IL-1 β mRNA Molecules as Predictors of Treatment Response Are Validated in an Independent Sample

We also measured the number of MIF and IL-1 β mRNA molecules in the second independent sample of 68 depressed patients. Overall, the absolute RNA levels observed in this second validation sample were similar to those observed in the first group (mean and SEM were $66.5 \pm 3.4 \times 10^6$ for IL-1 β and $74.8 \pm 2.9 \times 10^6$ for MIF). As a first validation, we found that, similarly to the GENDEP sample, nonresponders had higher numbers of MIF and IL-1 β molecules compared with responders (mean \pm SEM; IL-1 β : $78.5 \pm 3.5 \times 10^6$ in nonresponders vs $54.4 \pm 3.2 \times 10^6$ in responders, $t = 9.8$, $DF = 50$, $P < .01$; MIF: $94.3 \pm 3.2 \times 10^6$ in nonresponders vs $55.3 \pm 2.7 \times 10^6$ in responders, $t = 8.2$, $DF = 50$, $P < .001$). The results remain the same also when analyzing separately responders and nonresponders to SSRI ($P = .008$ for IL-1 β and $P = .009$ for MIF) or SNRI ($P = .003$ for IL-1 β and $P = .009$ for MIF), replicating the evidence from the GENDEP sample that these cytokines predict treatment response to both antidepressants.

As reported in Table 3, we then used the second sample to validate our predictive model based on the absolute mRNA cut-off values identified in the first sample. Indeed, we obtained probability scores for being responders/non-responders that were very similar to those obtained in the first sample ($P > .05$ for all comparisons between probability scores), again, with top-left cells representing patients with the lowest levels of inflammation and the highest probability of being a responder, and bottom-right cells representing those with the highest levels of inflammation and the lowest probability of being a responder. Specifically, green (responder) indicates $>82\%$ probability of being a responder and $<18\%$ probability of being a nonresponder; red (nonresponder) indicates $<28\%$ probability of being a responder and $>72\%$ probability of being a nonresponder; and orange indicate intermediate values. Based on these criteria, this sample had $n = 29$ (43%) of subjects classified as responders (MIF $\leq 60 \times 10^4$ and IL-1 $\beta \leq 85 \times 10^4$), $n = 13$ (19%) of subjects classified as non-responders (that is, with MIF $> 95 \times 10^4$ and IL-1 $\beta > 50 \times 10^4$), and $n = 26$ (38%) of subjects classified as intermediates (that is, MIF = 60×10^4 – 95×10^4 and IL-1 $\beta = 50 \times 10^4$ – 85×10^4). Similar to the GENDEP sample, 100% of those classified as non-responders ($n = 13$) were true nonresponders (PPV = 100%), and

all true responders ($n = 45$) were identified as responders (specificity = 100%). Ten (43%) of nonresponders were not identified as such (NPV = 82%, sensitivity = 56%).

Again, we found no difference in the expression levels of the three housekeeping genes in this sample; mean \pm SEM in responders vs nonresponders were: β -actin: 1.09 ± 0.8 vs 1.10 ± 0.7 ; B2M: 1.08 ± 0.5 vs 1.10 ± 0.7 ; GAPDH: 1.06 ± 0.5 vs 1.08 ± 0.7 (all $P > .05$).

MIF and IL-1 β Network Analysis Identifies Different Molecular Targets

The network analysis showed that MIF and IL-1 β were directly linked as reciprocal targets (data not shown), indicating that increased activation of one cytokine has downstream effects also on the other. However, the 2 cytokines connected to different and specific clusters of targets. As shown in Figure 1a, MIF did not show any interactions with other proinflammatory cytokines, and it interacted mainly with ubiquitin C via matrix metalloproteinase 9, which, in turn, is involved in the regulation of genes known to have an effect on neurogenesis, neuroplasticity, and cell proliferation, like Endothelial Growth Factor, Notch, and SMAD proteins (Anacker et al., 2013; Marschallinger et al., 2014). On the contrary, the neighbor targets of IL-1 β (Figure 1b) were mainly represented by proteins with inflammatory properties, including IL-6 and CRP, and by other molecules involved in the upstream or downstream regulation of the inflammatory signal, like Toll Like Receptors, Caspases (CASP), and nuclear factor kappa-light-chain-enhancer of activated B cells; interestingly all of these partners are related to the inflammasome complex or are mediators of oxidative stress, and they are well known to cause neurodegenerative effects (Leemans et al., 2011; Radi et al., 2014).

Discussion

In this study we report that the absolute numbers of MIF and IL-1 β mRNA molecules are both accurate and reliable predictors of antidepressant response, identifying, for the first time, an mRNA-based biomarker approach that is independent from local experimental settings and does not require “relative” quantification using housekeeping genes. Of note, the predictive power of these proinflammatory cytokines, and in particular the absolute values of mRNA molecules identified as the best cutoffs for prediction of nonresponse on an individual basis,

Table 2. Cutoff Values of MIF and IL-1 β Molecules and Probability Scores of Being a Responder or a Nonresponder in the First Sample of Depressed Patients (GENDEP Sample)

Number of molecules (IL-1 β or MIF) per nanogram total RNA	IL-1 $\beta \leq 50 \times 10^4$	$50 \times 10^4 < IL-1\beta \leq 85 \times 10^4$	$IL-1\beta > 85 \times 10^4$
$MIF \leq 60 \times 10^4$	Responder probability >0.995 Nonresponder probability <0.005	Responder probability >0.73 Nonresponder probability <0.27	Responder probability >0.37 Nonresponder probability <0.63
$60 \times 10^4 < MIF \leq 95 \times 10^4$	Responder probability >0.32 Nonresponder probability <0.68	Responder probability 0.008 – 0.995 Nonresponder probability 0.005 – 0.99	Responder probability <0.73 Nonresponder probability >0.27
$MIF > 95 \times 10^4$	Responder probability 0.03 – 0.93 Nonresponder probability 0.07 – 0.97	Responder probability <0.32 Nonresponder probability >0.72	Responder probability <0.008 Nonresponder probability >0.99

White columns on the right represent MIF cutoffs (in term of number of molecules) and white rows on the top represent IL-1 β cutoffs (in term of number of molecules); each cells or in green, orange, or red indicates the probability of being a responder or a nonresponder: green (responder) indicates $>73\%$ probability of being a responder and $<27\%$ probability of being a nonresponder; red (nonresponder) indicates $<32\%$ probability of being a responder and $>72\%$ probability of being a nonresponder; and orange indicate intermediate values.

Table 3. Cutoff Values of MIF and IL-1 β Molecules and Probability Scores of Being a Responder or a Nonresponder in the Second Independent Sample of Depressed Patients

Number of Molecules (IL-1 β or MIF)/ng Total RNA	IL-1 β $\leq 50 \times 10^4$	$50 \times 10^4 < IL-1\beta \leq 85 \times 10^4$	IL-1 β $> 85 \times 10^4$
MIF $\leq 60 \times 10^4$	Responder probability >0.99 Nonresponder probability <0.001	Responder probability >0.82 Nonresponder probability <0.18	Responder probability >0.39 Nonresponder probability <0.61
$60 \times 10^4 < MIF \leq 95 \times 10^4$	Responder probability >0.28 Nonresponder probability <0.72	Responder probability $0.001-0.99$ Nonresponder probability $0.001-0.99$	Responder probability <0.82 Nonresponder probability >0.18
MIF $> 95 \times 10^4$	Responder probability $0.007-0.97$ Nonresponder probability $0.03-0.99$	Responder probability <0.28 Nonresponder probability >0.72	Responder probability <0.001 Nonresponder probability >0.99

White columns on the right represent MIF cutoffs (in term of number of molecules) and white rows on the top represent IL-1 β cutoffs (in term of number of molecules); each cells or in green, orange, or red indicates the probability of being a responder or a nonresponder: green (responder) indicates $>82\%$ probability of being a responder and $<18\%$ probability of being a nonresponder; red (nonresponder) indicates $<28\%$; probability of being a responder and $>72\%$ probability of being a nonresponder; and orange indicate intermediate values.

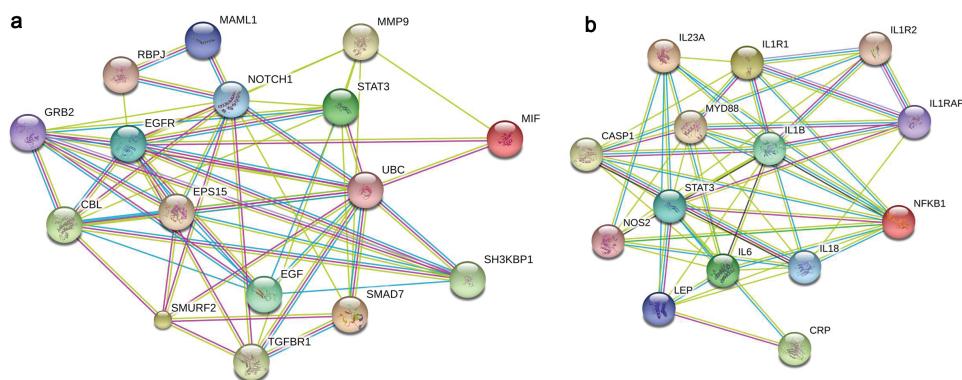


Figure 1. Representative interaction between Macrophage Migration Inhibitory Factor (MIF) (Figure 1a) or interleukin (IL)-1 β (Figure 1b) and their neighbors' targets, where nodes (genes) can be either colored (if they are directly linked to the input, in that case MIF) or white (nodes of a higher interaction/depth-this is not the case). Lines represent predicted functional edges of interaction between nodes, and they are represented with eight different colors according to the type of evidence and the predictive method used (neighborhood, gene fusion, co-occurrence, coexpression, experiments, databases, and textmining): green line, activation; red line, inhibition; blue line, binding; light blue line, phenotype; violet, catalyzes; pink, posttranslational mechanism; black, reaction; yellow, coexpression (<http://string-db.org>).

were independently replicated in two clinical samples: a randomized trial that we had previously used to measure relative mRNA expression and to identify the top mRNA transcripts of interest (Cattaneo et al., 2013); and a naturalistic cohort that has been independently recruited for this study. Both samples demonstrate identical PPV and specificity of 100% for nonresponders. This is the ideal scenario in clinical practice: patients with mRNA numbers above the suggested cutoffs ("red" in Tables 2 and 3) could be directed toward early access to more assertive antidepressant strategies, including augmentation with other antidepressants or antiinflammatory drugs, while those with mRNA numbers below the suggested cutoffs could be directed toward standard care. Most importantly, this would allow a therapeutically conservative approach, where no true responders would be exposed unnecessarily to more assertive pharmacological strategies.

Depressed patients who are resistant to conventional antidepressants have higher concentrations of inflammatory biomarkers in plasma or serum (Sluzewska et al., 1997; Lanquillon et al., 2000; Fitzgerald et al., 2006), and successful antidepressant treatment has been associated with a reduction in the levels of proinflammatory cytokines (Abdel-Salam et al., 2003; Roumestan et al., 2007; Cattaneo et al., 2013). As mentioned above, we

conducted the first study showing that (relative) mRNA expression levels of IL-1 β , MIF, and TNF- α are higher in nonresponders. Our findings have been corroborated by Belzeaux and colleagues (2012), who identified a combination of mRNA signatures of TNF α and IL-1 β as best predictors of antidepressant response in a case study, and by Powell and colleagues (2013) in a separate study of a different GENDEP sample, who investigated the mRNA expression of 84 genes related to inflammation and found that nonresponders have significantly higher baseline mRNA levels of TNF and other TNF-target genes. Cytokine mRNA levels may also be involved in the genetic regulation of treatment response: for example, Uher et al. (2010) have found, in yet another GENDEP sample, a single nucleotide polymorphism (rs1126757) in the IL-11 gene with suggestive genome-wide significance, and Powell et al. (2013) have found that only carriers of the IL-11 A allele (who have a better response) show a significant reduction in IL-11 mRNA expression following citalopram. Together with the present findings showing accurate and reliable prediction of antidepressant response by absolute mRNA levels in 2 independent samples, all these studies show that peripheral blood mRNA analyses are one of the most promising approaches for biomarker discovery in mental health (Hepgul et al., 2013). This notion is supported by the fact that about 80% of genes

are coexpressed, and similarly modulated as mRNA levels, in peripheral blood cells and brain tissues (Sullivan et al., 2006). Moreover, molecular mechanisms activated by early life stress, a well-known risk factor for antidepressant nonresponse (Nanni et al., 2012), produce similar epigenetic changes in human blood cells and human neuronal cells, leading to long-term dysregulation of the stress hormone system and global changes in the function of immune cells as well as of brain areas associated with stress regulation (Klengel et al., 2013; Uher et al., 2014).

In our study, we find that MIF and IL-1 β mRNA levels predict treatment response across antidepressants classes, that is, for both escitalopram and nortriptyline in the GENDEP samples as well as for both SSRIs and SNRIs in the naturalistic cohort. These findings may be perceived as in contrast with the recent paper by Uher et al. (2014) showing (in a larger, different GENDEP sample) that high levels of serum CRP predicted lack of treatment response to the SSRI, escitalopram, but not to the tricyclic (and noradrenergic uptake inhibitor) nortriptyline. The authors explain this by speculating that antidepressants with a noradrenergic action may have antiinflammatory properties (Uher et al., 2014), and, in fact, we and others have found that noradrenergic or SNRI antidepressants have antiinflammatory properties *in vitro* (Horowitz et al., 2014). However, the antiinflammatory properties of SSRI drugs are also well known (Tynan et al., 2012; Al-Amin et al., 2013; Cattaneo et al., 2013). Our network analysis may shed some light on this apparent discrepancy, as CRP is only loosely connected with IL-1 β (and it is not connected with MIF), indicating that subjects with high levels of CRP may be different from those with high levels of MIF or IL-1 β ; indeed CRP levels in the aforementioned paper predicted only around 11% of MADRS-based response, as opposed to 40% to 50% of the variance explained by cytokines mRNA levels in our study (Cattaneo et al., 2013). Moreover, because MIF and IL-1 β are “neighbor targets,” it is possible that having high levels of either of these cytokines (*i.e.*, above the top cutoff) is enough to activate downstream targets of both, thus affecting both the neurogenesis/neuroplasticity targets of MIF and the inflammasome/neurodegeneration targets of IL-1 β . These widespread molecular abnormalities would likely inhibit the response to a wide range of antidepressants. Of note, taken together with the other GENDEP studies mentioned above by Uher et al. (2010) and Powell et al. (2013), our study and the study by Uher et al. (2014) all point to a prominent role of increased inflammation in the lack of antidepressant response in the GENDEP cohort. However, as these studies have been conducted in different and only partially overlapping samples, it is difficult to determine if and how these genetic, serum, and mRNA biomarker cluster together.

High levels of inflammation can prevent response to antidepressants, because it can interfere with the same biological processes that are crucial for antidepressant therapeutic action. For example, high levels of inflammation increase the expression and activation of monoamine transporters, reduce tryptophan availability, inhibit neuropeptide and growth factors involved in neuroplasticity, and interfere with the kynurenine pathways, leading in turn to a reduction in neurogenesis and glutamate dysfunction (Zunszain et al., 2012; Felger and Lotrich, 2013). Of note, inflammation can be triggered by childhood trauma (Danese et al., 2007, 2008; Hepgul et al., 2013; Baumeister et al., 2015), which is in itself associated with poor antidepressant response (Nanni et al., 2012); thus, it is possible to speculate that a cluster of individuals have both a history of childhood trauma and increased inflammation, eventually leading to lack of antidepressant response (Nemeroff et al., 2003; Nanni et al., 2012; Coelho et al., 2014). However, it is also known that genetic

variability in inflammatory genes, such as IL-6, IL-11, and TNF- α , as well as in enzymes involved in the prostaglandins and kynurenine pathways, contributes to both the risk of depression and to antidepressant response (Bull et al., 2009; Dowlati et al., 2010; Uher et al., 2010; Bufalino et al., 2013), thus suggesting that a different cluster of individuals expresses high levels of inflammation and lack of treatment response through a predominantly genetic path. Indeed, it is possible that the better accuracy and validity of mRNA gene expression as biomarkers of treatment response are due to the ability of these molecules to integrate genomic variability and environmental effects. Importantly, inflammatory genes could represent not only biomarkers predicting lack of antidepressant response but also novel targets for antidepressant therapies, and indeed clinical trial have been conducted to test the efficacy of antiinflammatory drugs. Müller et al. (2006) was the first to conduct a randomized controlled clinical trial to evaluate the efficacy of celecoxib added to antidepressant drugs and reported improved antidepressant effects. Subsequent studies confirmed these findings (Akhondzadeh et al., 2009; Hashemian et al., 2011; Abbasi et al., 2012). Moreover, Raison and colleagues (2013) have recently reported that a specific antagonist of TNF α improves depressive symptoms in patients with high inflammation at baseline. No clinical trials have been conducted with drugs able to inhibit MIF or IL-1 β ; however, there are promising data in animals that IL-1 β receptor antagonism prevents long-term cognitive impairment and memory deficits, which are clinical features observed in depressed patients (Barichello et al., 2015).

The targets identified by the network analyses of MIF and IL-1 β shed some interesting light on the putative molecular mechanisms underlying their predictive effects. MIF promotes the expression and function of multiple cytokines and chemokines, including TNF-alpha, IL-6, CXCL1, and CCL2 (Gregory et al., 2006; Toh et al., 2006; Fan et al., 2011; Santos et al., 2011). However, MIF also plays a role as a physiological counter-regulator of glucocorticoid antiinflammatory action (Leech et al., 2000; Aeberli et al., 2006) by regulating glucocorticoid-induced leucine zipper, which in turn not only has antiinflammatory functions but also is involved in the effects of stress on neurogenesis (Anacker et al., 2013). Finally, MIF also interacts with genes involved in neurogenesis, neuroplasticity, and cell proliferation, like Endothelial Growth Factor, Notch, and SMAD proteins (Anacker et al., 2013; Marschallinger et al., 2014). Indeed, MIF has been shown to promote neuroplasticity and neuroprotective processes under physiological conditions, but it can also increase the production of proinflammatory cytokines under conditions of stress. Moreover, MIF is modulated by glucocorticoids, and high, antiinflammatory doses of glucocorticoids inhibit MIF secretion; however, during pathological conditions characterized by glucocorticoid resistance, such as depression, it is conceivable that levels of MIF are increased (Bloom et al., 2014). In contrast, IL-1 β contributes to the activation of other cytokines (IL-18, IL-6) and mediators of oxidative stress, like CASP1, CASP9, and Nitric oxide synthase 2, related to the inflammasome complex, also comprising the Nod-like Receptor, the precursor pro-caspase-1, and the adaptor ASC. Interestingly an overactivation of the inflammasome in peripheral blood cells of depressed patients has been recently demonstrated, providing support to its putative role in the pathogenesis of depression and its treatment (Alcocer-Gomez and Cordero, 2014). We would advocate that any of these pathways might represent a target for novel antidepressant therapies.

The finding that markers of peripheral inflammation can predict the response to an antidepressant can be interpreted as an

example of a peripheral correlate of central (immune) alterations. Indeed, increased neuro-inflammation has been reported in the brain of patients with depression; for example, postmortem studies have found an increased proportion of primed microglia and higher mRNA levels of several chemokines involved in the recruitment of monocytes (Torres-Platas et al., 2014); and a recent neuroimaging study has found increased microglia activation (translocator protein density) using positron emission tomography (Setiawan, et al., 2015). Indeed, it has recently been proposed that microglial activation in psychiatric disorders could not only contribute to the brain pathology but also influence treatment response (Reus et al., 2015). It is also, however, important to highlight that readily available, blood-based peripheral biomarkers are relevant in psychiatry even if they do not directly reflect brain-based mechanisms when they reliably predict important clinical features such as risk, vulnerability, outcome, and treatment response.

The findings of the present study need to be interpreted in the light of important limitations. First, although we have validated our results in 2 independent samples, the sample size is relatively small, and therefore these findings should be replicated in larger datasets. Indeed, we have assessed "response" rather than "remission," as the latter would have led to even smaller groups available for the analyses. Second, the identified biomarkers predict the response to antidepressants in general and are not specific to one or the other of the antidepressant classes; therefore, they cannot be used to guide the choice of antidepressants but rather to identify patients that may benefit from an early access to adjuvant therapies. Third, these findings may be relevant only to patients that, as in our samples, show no historical evidence of chronicity or of treatment resistance to multiple antidepressants, although we would argue that this is indeed the kind of patients that would benefit most from our proposed personalized approach of early access to assertive antidepressant strategies based on the biomarkers profile.

In summary, here we provide evidence that the measurements of the number of MIF and IL-1 β mRNA molecules, which is an absolute and thus reliable quantitation that could be adopted across laboratories, may help the clinicians in the prediction of antidepressant response, so that patients with mRNA numbers above the suggested cutoffs ("red") could be directed toward early access to more rapid escalation of assertive antidepressant strategies. However, randomized controlled trials testing the use of biomarkers vs treatment as usual need to be conducted in order to deliver clear guidelines, especially considering the increased risk of adverse effects when combining conventional antidepressants with antiinflammatories (Andrade et al., 2010).

Acknowledgments

The MRC ImmunoPsychiatry Consortium include: 1) University of Cambridge (Edward Bullmore, Petra E. Vértes, Rudolf Cardinal); 2) MRC Biostatistics Unit, Cambridge (Sylvia Richardson, Gwenael Leday); 3) University of Edinburgh (Tom Freeman, David Hume, Tim Regan, Zhaozong Wu); 4) King's College London (Carmine Pariante, Annamaria Cattaneo, Patricia Zunszain, Alessandra Borsini, Robert Stewart, David Chandran); 5) University College London (Livia Carvalho, Joshua Bell, Luis Henrique Souza-Teodoro); 6) University of Southampton (Hugh Perry); 7) University of Sussex (Neil Harrison); 8) Johnson&Johnson (Wayne Drevets, Gayle M Witteberg, Yu Sun, Declan Jones); 9) GlaxoSmithKline (Edward Bullmore, Shahid Khan, Annie Stylianou, Robert B Henderson).

This work was supported by the grants "Immunopsychiatry: a consortium to test the opportunity for immunotherapeutics

in psychiatry" (MR/L014815/1) and 'Persistent Fatigue Induced by Interferon-alpha: A New Immunological Model for Chronic Fatigue Syndrome' (MR/J002739/1), from the Medical Research Council (UK). Additional support has been offered by the National Institute for Health Research Mental Health Biomedical Research Centre in Mental Health at South London and Maudsley NHS Foundation Trust and King's College London. Dr. Cattaneo is also funded by the Eranet Neuron 'Inflame-D'. The GENDEP project was supported by a European Commission Framework 6 grant (contract reference: LSHB-CT-2003-503428).

Statement of Interest

Professor Pariante has received research funding from Johnson & Johnson as part of a programme of research on depression and inflammation, and speaker's fee from Lundbeck. In addition, Professor Pariante has received research funding from the Medical Research Council (UK) and the Wellcome Trust for research on depression and inflammation as part of two large consortia that also include Johnson & Johnson, GSK, Pfizer and Lundbeck.

References

- Abdel-Salam OM, Nofal SM, El-Shenawy SM (2003) Evaluation of the anti-inflammatory and anti-nociceptive effects of different antidepressants in the rat. *Pharmacol Res* 48:157–165.
- Aeberli D, Yang Y, Mansell A, Santos L, Leech M, Morand EF (2006) Endogenous macrophage migration inhibitory factor modulates glucocorticoid sensitivity in macrophages via effects on MAP kinase phosphatase-1 and p38 MAP kinase. *FEBS Lett* 580:974–981.
- Al-Amin MM, Uddin MM, Rahman MM, Reza HM, Rana MS (2013) Effect of diclofenac and antidepressants on the inflammatory response in astrocyte cell culture. *Inflammopharmacol* 21:421–5. doi: 10.1007/s10787-013-0181-9.
- Alcocer-Gomez E, Cordero MD (2014) NLRP3 inflammasome: a new target in major depressive disorder. *CNS Neurosci Ther* 20:294–295. doi: 10.1111/cns.12230.
- Anacker C, Cattaneo A, Luoni A, Musaelyan K, Zunszain PA, Milanesi E, Rybka J, Berry A, Cirulli F, Thuret S, Price J, Riva MA, Gennarelli M, Pariante CM (2013) Glucocorticoid-related molecular signaling pathways regulating hippocampal neurogenesis. *Neuropsychopharmacology* 38:872–883. doi: 10.1038/npp.2012.253.
- Andrade C, Sandarsh S, Chethan KB, Nagesh KS (2010) Serotonin reuptake inhibitor antidepressants and abnormal bleeding: a review for clinicians and a reconsideration of mechanisms. *J Clin Psychiatry* 71:1565–1575. doi: 10.4088/JCP.09r05786blu. Review.
- Baumeister D, Akhtar R, Ciufolini S, Pariante CM, Mondelli V (2015) Childhood trauma and adulthood inflammation: a meta-analysis of peripheral C-reactive protein, interleukin-6 and tumour necrosis factor- α . *Mol Psychiatry*:2015 Jun 2.
- Belzeaux R, Bergon A, Jeanjean V, Loriod B, Formisano-Treziny C, Verrier L, Loundou A, Baumstarck-Barrau K, Boyer L, Gall V, Gabert J, Nguyen C, Azorin JM, Naudin J, Ibrahim EC (2012) Responder and nonresponder patients exhibit different peripheral transcriptional signatures during major depressive episode. *Transl Psychiatry* 2:e185. doi: 10.1038/tp.2012.112.
- Bloom J, Al-Abed Y (2014) MIF: mood improving/inhibiting factor? *J Neuroinflammation* 21:11. doi: 10.1186/1742-2094-11-11. Review.
- Bufalino C, Hepgul N, Aguglia E, Pariante CM (2013) The role of immune genes in the association between depression and

- inflammation: a review of recent clinical studies. *Brain Behav Immun* 31:31–47. doi: 10.1016/j.bbi.2012.04.009.
- Bull SJ, Huezo-Diaz P, Binder EB, Cubells JF, Ranjith G, Maddock C, Miyazaki C, Alexander N, Hotopf M, Cleare AJ, Norris S, Cassidy E, Aitchison KJ, Miller AH, Pariante CM (2009) Functional polymorphisms in the interleukin-6 and serotonin transporter genes, and depression and fatigue induced by interferon-alpha and ribavirin treatment. *Mol Psychiatry* 14:1095–1104. doi: 10.1038/mp.2008.48.
- Carvalho LA, Torre JP, Papadopoulos AS, Poon L, Juruena MF, Markopoulou K, Cleare AJ, Pariante CM (2013) Lack of clinical therapeutic benefit of antidepressants is associated overall activation of the inflammatory system. *J Affect Disord* 148:136–140. doi: 10.1016/j.jad.2012.10.036.
- Cattaneo A, Gennarelli M, Uher R, Breen G, Farmer A, Aitchison KJ, Craig IW, Anacker C, Zunsztain PA, McGuffin P, Pariante CM (2013) Candidate genes expression profile associated with antidepressants response in the GENDEP study: differentiating between baseline ‘predictors’ and longitudinal ‘targets’. *Neuropsychopharmacology* 38:377–385. doi: 10.1038/npp.2012.191.
- Coelho R, Viola TW, Walss-Bass C, Brietzke E, Grassi-Oliveira R (2014) Childhood maltreatment and inflammatory markers: a systematic review. *Acta Psychiatr Scand* 129:180–192. doi: 10.1111/acps.12217.
- Danese A, Moffitt TE, Pariante CM, Ambler A, Poulton R, Caspi A (2008) Elevated inflammation levels in depressed adults with a history of childhood maltreatment. *Arch Gen Psychiatry* 65:409–415. doi: 10.1001/archpsyc.65.4.409.
- Danese A, Pariante CM, Caspi A, Taylor A, Poulton R (2007) Childhood maltreatment predicts adult inflammation in a life-course study. *Proc Natl Acad Sci U S A* 104:1319–1324. doi: 10.1073/pnas.0610362104.
- Dowlati Y, Herrmann N, Swardfager W, Liu H, Sham L, Reim EK, Lanctot KL (2010) A meta-analysis of cytokines in major depression. *Biol Psychiatry* 67:446–457. doi: 10.1016/j.biopsych.2009.09.033.
- Fan H, Hall P, Santos LL, Gregory JL, Fingerle-Rowson G, Bucala R, Morand EF, Hickey MJ (2011) Macrophage migration inhibitory factor and CD74 regulate macrophage chemotactic responses via MAPK and Rho GTPase. *J Immunol* 186:4915–4924. doi: 10.4049/jimmunol.1003713.
- Felger JC, Lotrich FE (2013) Inflammatory cytokines in depression: neurobiological mechanisms and therapeutic implications. *Neuroscience* 246:199–229. doi: 10.1016/j.neuroscience.2013.04.060.
- Fitzgerald P, O’Brien SM, Scully P, Rijkers K, Scott LV, Dinan TG (2006) Cutaneous glucocorticoid receptor sensitivity and pro-inflammatory cytokine levels in antidepressant-resistant depression. *Psychol Med* 36:37–43. doi: 10.1017/S003329170500632X.
- Gregory JL, Morand EF, McKeown SJ, Ralph JA, Hall P, Yang YH, McColl SR, Hickey MJ (2006) Macrophage migration inhibitory factor induces macrophage recruitment via CC chemokine ligand 2. *J Immunol* 177:8072–8079.
- Hegpul N, Cattaneo A, Zunsztain PA, Pariante CM (2013) Depression pathogenesis and treatment: what can we learn from blood mRNA expression? *BMC Med* 11:28. doi: 10.1186/1741-7015-11-28.
- Horowitz MA, Wertz J, Zhu D, Cattaneo A, Musaelyan K, Nikkheisl N, Thuret S, Pariante CM, Zunsztain PA (2014) Antidepressant compounds can be both pro- and anti-inflammatory in human hippocampal cells. *Int J Neuropsychopharmacol* 18: pii: pyu076. doi: 10.1093/ijnp/pyu076.
- Howren MB, Lamkin DM, Suls J (2009) Associations of depression with C-reactive protein, IL-1, and IL-6: a meta-analysis. *Psychosom Med* 71:171–186. doi: 10.1097/PSY.0b013e3181907c1b.
- Keers R, Uher R, Gupta B, Rietschel M, Schulze TG, Hauser J, Skibinska M, Henigsberg N, Klember P, Maier W, Zobel A, Mors O, Kristensen AS, Kozel D, Giovannini C, Mendlewicz J, Kumar S, McGuffin P, Farmer AE, Aitchison KJ (2010) Stressful life events, cognitive symptoms of depression and response to antidepressants in GENDEP. *J Affect Disord* 127:337–342. doi: 10.1016/j.jad.2010.06.011.
- Klengel T, Mehta D, Anacker C, Rex-Haffner M, Pruessner JC, Pariante CM, Pace TW, Mercer KB, Mayberg HS, Bradley B, Nemeroff CB, Holsboer F, Heim CM, Ressler KJ, Rein T, Binder EB (2013) Allele-specific FKBP5 DNA demethylation mediates gene-childhood trauma interactions. *Nat Neurosci* 16:33–41. doi: 10.1038/nn.3275.
- Lanquillon S, Krieg JC, Bening-Abu-Shach U, Vedder H (2000) Cytokine production and treatment response in major depressive disorder. *Neuropsychopharmacology* 22:370–379. doi: 10.1016/S0893-133X(99)00134-7.
- Leech M, Metz C, Bucala R, Morand EF (2000) Regulation of macrophage migration inhibitory factor by endogenous glucocorticoids in rat adjuvant-induced arthritis. *Arthritis Rheum* 43:827–833. doi: 10.1002/1529-0131(200004)43:4<827::AID-ANR13>3.0.CO;2-K.
- Leemans JC, Cassel SL, Sutterwala FS (2011) Sensing damage by the NLRP3 inflammasome. *Immunol Rev* 243:152–162. doi: 10.1111/j.1600-065X.2011.01043.x.
- Li X, Wang X (2000) Application of real-time polymerase chain reaction for the quantitation of interleukin-1beta mRNA upregulation in brain ischemic tolerance. *Brain Res Brain Res Protoc* 5:211–217.
- Lisi L, Camardese G, Treglia M, Tringali G, Carrozza C, Janiri L, Dello Russo C, Navarra P (2013) Monocytes from depressed patients display an altered pattern of response to endotoxin challenge. *PLoS One* 8:e52585. doi: 10.1371/journal.pone.0052585.
- Marschallinger J, Krampert M, Couillard-Despres S, Heuchel R, Bogdahn U, Aigner L (2014) Age-dependent and differential effects of Smad7DeltaEx1 on neural progenitor cell proliferation and on neurogenesis. *Exp Gerontol* 57C:149–154. doi: 10.1016/j.exger.2014.05.011.
- Martinez JM, Garakani A, Yehuda R, Gorman JM (2012) Proinflammatory and “resiliency” proteins in the CSF of patients with major depression. *Depress Anxiety* 29:32–38. doi: 10.1002/da.20876.
- Mc Lachlan 2004 Discriminant analysis and statistical pattern recognition, Book
- Müller N, Schwarz MJ, Dehning S, Douhe A, Cerovecki A, Goldstein-Müller B, Spellmann I, Hetzel G, Maino K, Kleindienst N, Möller HJ, Arolt V, Riedel M (2006) The cyclooxygenase-2 inhibitor celecoxib has therapeutic effects in major depression: results of a double-blind, randomized, placebo controlled, add-on pilot study to reboxetine. *Mol Psychiatry* 11(7):680–684.
- Nanni V, Uher R, Danese A (2012) Childhood maltreatment predicts unfavorable course of illness and treatment outcome in depression: a meta-analysis. *Am J Psychiatry* 169: 141–151.
- Nemeroff CB, Heim CM, Thase ME, Klein DN, Rush AJ, Schatzberg AF, Ninan PT, McCullough JP Jr, Weiss PM, Dunner DL, Rothbaum BO, Kornstein S, Keitner G, Keller MB (2003) Differential responses to psychotherapy versus pharmacotherapy in patients with chronic forms of major depression and

- childhood trauma. *Proc Natl Acad Sci U S A* 100:14293–14296. doi: 10.1073/pnas.2336126100.
- Powell TR, Schalkwyk LC, Heffernan AL, Breen G, Lawrence T, Price T, Farmer AE, Aitchison KJ, Craig IW, Danese A, Lewis C, McGuffin P, Uher R, Tansey KE, D'souza UM (2013) Tumor necrosis factor and its targets in the inflammatory cytokine pathway are identified as putative transcriptomic biomarkers for escitalopram response. *Eur Neuropsychopharmacol* 23:1105–1114. doi: 10.1016/j.euroneuro.2012.09.009.
- Radi E, Formichi P, Battisti C, Federico A (2014) Apoptosis and Oxidative Stress in Neurodegenerative Diseases. *J Alzheimers Dis.* doi: 10.3233/JAD-132738.
- Raison CL, Rutherford RE, Woolwine BJ, Shuo C, Schettler P, Drake DF, Haroon E, Miller AH (2013) A randomized controlled trial of the tumor necrosis factor antagonist infliximab for treatment-resistant depression: the role of baseline inflammatory biomarkers. *JAMA Psychiatry* 70:31–41. doi: 10.1001/2013.jamapsychiatry.4.
- Rethorst CD, Toups MS, Greer TL, Nakonezny PA, Carmody TJ, Grannemann BD, Huebinger RM, Barber RC, Trivedi MH (2013) Pro-inflammatory cytokines as predictors of antidepressant effects of exercise in major depressive disorder. *Mol Psychiatry* 18:1119–1124. doi: 10.1038/mp.2012.125.
- Réus GZ, Fries GR, Stertz L, Badawy M, Passos IC, Barichello T, Kapczinski F, Quevedo J (2015) The role of inflammation and microglial activation in the pathophysiology of psychiatric disorders. *Neuroscience*. 300, 141–154. doi: 10.1016/j.neuroscience.2015.05.018.
- Roumestan C, Michel A, Bichon F, Portet K, Detoc M, Henriet C, Jaffuel D, Mathieu M (2007) Anti-inflammatory properties of desipramine and fluoxetine. *Respir Res* 8:35. doi: 10.1186/1465-9921-8-35.
- Rush AJ, Thase ME, Dube S (2003) Research issues in the study of difficult-to-treat depression. *Biol Psychiatry* 53:743–753.
- Santos LL, Fan H, Hall P, Ngo D, Mackay CR, Fingerle-Rowson G, Bucala R, Hickey MJ, Morand EF (2011) Macrophage migration inhibitory factor regulates neutrophil chemotactic responses in inflammatory arthritis in mice. *Arthritis Rheum* 63:960–970. doi: 10.1002/art.30203.
- Setiawan E, Wilson AA, Mizrahi R, Rusjan PM, Miler L, Rajkowska G, Suridjan I, Kennedy JL, Rekkas PV, Houle S, Meyer JH (2015) Role of translocator protein density, a marker of neuroinflammation, in the brain during major depressive episodes. *JAMA Psychiatry* 72:268–275.
- Sluzewska A, Sobieska M, Rybakowski JK (1997) Changes in acute-phase proteins during lithium potentiation of antidepressants in refractory depression. *Neuropsychobiology* 35:123–127.
- Sullivan PF, Fan C, Perou CM (2006) Evaluating the comparability of gene expression in blood and brain. *Am J Med Genet B Neuropsychiatr Genet* 141B:261–268. doi: 10.1002/ajmg.b.30272.
- Toh ML, Aeberli D, Lacey D, Yang Y, Santos LL, Clarkson M, Sharma L, Clyne C, Morand EF (2006) Regulation of IL-1 and TNF receptor expression and function by endogenous macrophage migration inhibitory factor. *J Immunol* 177:4818–4825.
- Torres-Platas SG, Cruceanu C, Chen GG, Turecki G, Mechawar N (2014) Evidence for increased microglial priming and macrophage recruitment in the dorsal anterior cingulate white matter of depressed suicides. *Brain Behav Immun* 42:50–59.
- Trivedi MH (2006) Major depressive disorder: remission of associated symptoms. *J Clin Psychiatry* 67 Suppl 6:27–32.
- Tynan RJ, Weidenhofer J, Hinwood M, Cairns MJ, Day TA, Walker FR (2012) A comparative examination of the anti-inflammatory effects of SSRI and SNRI antidepressants on LPS stimulated microglia. *Brain Behav Immun*. 26:469–79. doi: 10.1016/j.bbi.2011.12.011.
- Uher R, Huezo-Diaz P, Perroud N, Smith R, Rietschel M, Mors O, Hauser J, Maier W, Kozel D, Henigsberg N, Barreto M, Placentino A, Dernovsek MZ, Schulze TG, Kalember P, Zobel A, Czerski PM, Larsen ER, Souery D, Giovannini C, Gray JM, Lewis CM, Farmer A, Aitchison KJ, McGuffin P, Craig I (2009) Genetic predictors of response to antidepressants in the GENDEP project. *Pharmacogenomics J* 9:225–233. doi: 10.1038/tpj.2009.12.
- Uher R, Perroud N, Ng MY, Hauser J, Henigsberg N, Maier W, Mors O, Placentino A, Rietschel M, Souery D, Zagar T, Czerski PM, Jerman B, Larsen ER, Schulze TG, Zobel A, Cohen-Woods S, Pirlo K, Butler AW, Muglia P, Barnes MR, Lathrop M, Farmer A, Breen, G, Aitchison KJ, Craig I, Lewis CM, McGuffin P (2010) Genome-wide pharmacogenetics of antidepressant response in the GENDEP project. *Am J Psychiatry* 167:555–564. doi: 10.1176/appi.ajp.2009.09070932.
- Uher R, Tansey KE, Dew T, Maier W, Mors O Hauser, J Dernovsek MZ, Henigsberg N, Souery D, Farmer A, McGuffin P (2014) An Inflammatory Biomarker as a Differential Predictor of Outcome of Depression Treatment With Escitalopram and Nortriptyline. *Am J Psychiatry*. doi: 10.1176/appi.ajp.2014.14010094.
- Zunszain PA, Anacker C, Cattaneo A, Choudhury S, Musaelyan K, Myint AM, Thuret S, Price J, Pariante CM (2012) Interleukin-1beta: a new regulator of the kynurene pathway affecting human hippocampal neurogenesis. *Neuropsychopharmacology* 37:939–949. doi: 10.1038/npp.2011.277.