

# Mapping of 79 loci for 83 plasma protein biomarkers in cardiovascular disease

## Short title

Novel loci for the plasma proteome

## Authors:

Lasse Folkersen<sup>1,3</sup>, Eric Fauman<sup>2</sup>, Maria Sabater-Lleal<sup>3</sup>, Rona J Strawbridge<sup>3</sup>, Mattias Frånberg<sup>3</sup>, Bengt Sennblad<sup>3</sup>, Damiano Baldassarre<sup>4,5</sup>, Fabrizio Veglia<sup>5</sup>, Steve E. Humphries<sup>6</sup>, Rainer Rauramaa<sup>7</sup>, Ulf de Faire<sup>8</sup>, Andries J. Smit<sup>9</sup>, Philippe Giral<sup>10</sup>, Sudhir Kurl<sup>11</sup>, Elmo Mannarino<sup>12</sup>, Stefan Enroth<sup>13</sup>, Åsa Johansson<sup>13</sup>, Sofia Bosdotter Enroth<sup>14</sup>, Stefan Gustafsson<sup>15</sup>, Lars Lind<sup>15</sup>, Cecilia Lindgren<sup>16</sup>, Andrew P Morris<sup>17</sup>, Vilmantas Giedraitis<sup>16</sup>, Angela Silveira<sup>3</sup>, Anders Franco-Cereceda<sup>18</sup>, Elena Tremoli<sup>4,5</sup>, the IMPROVE study group, Ulf Gyllenstein<sup>13</sup>, Erik Ingelsson<sup>15,19</sup>, Søren Brunak<sup>1</sup>, Per Eriksson<sup>3</sup>, Daniel Ziemek<sup>2</sup>, Anders Hamsten<sup>3</sup>, Anders Mälarstig<sup>3,20</sup>

## Affiliations:

1 Department of Systems Biology, Technical University of Denmark, Copenhagen, 2800, Denmark  
2 Pfizer Worldwide Research & Development, Cambridge, MA, 02139, USA  
3 Cardiovascular Medicine Unit, Department of Medicine Solna, Karolinska Institutet, Stockholm, 17176, Sweden  
4 Dipartimento di Scienze Farmacologiche e Biomolecolari, Università di Milano, Milan, Italy.  
5 Centro Cardiologico Monzino, IRCCS, Milan Italy.  
6 British Heart Foundation Laboratories, University College of London, Department of Medicine, Rayne Building, London, WC1E 6BT, United Kingdom  
7 Foundation for Research in Health Exercise and Nutrition, Kuopio Research Institute of Exercise Medicine, Kuopio, 70100, Finland.  
8 Division of Cardiovascular Epidemiology, Institute of Environmental Medicine, Karolinska Institutet, and Department of Cardiology, Karolinska University Hospital, Solna, Karolinska Institutet, Stockholm, 17176, Sweden.  
9 Department of Medicine, University Medical Center Groningen, Groningen, 30001, the Netherlands.  
10 Assistance Publique - Hopitaux de Paris; Service Endocrinologie-Metabolisme, Groupe Hôpitalier Pitie-Salpetriere, Unités de Prévention Cardiovasculaire, Paris, 75013, France.  
11 Institute of Public Health and Clinical Nutrition, University of Eastern Finland, Kuopio Campus, Kuopio, 80101, Finland  
12 Internal Medicine, Angiology and Arteriosclerosis Diseases, Department of Clinical and Experimental Medicine, University of Perugia, Perugia, Italy.  
13 Department of Immunology, Genetics and Pathology, Science for Life Laboratory Uppsala, Uppsala University, Uppsala, 75108, Sweden  
14 Department of Internal Medicine, Uppsala University Hospital, 75185 Uppsala, Sweden  
15 Department of Medical Sciences, Molecular Epidemiology and Science for Life Laboratory, Uppsala University, Uppsala, 75185, Sweden  
16 Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, OX3 7BN, United Kingdom  
17 Department of Biostatistics, University of Liverpool, Liverpool, L69 3BX, United Kingdom  
18 Cardiothoracic Surgery Unit, Department of Molecular Medicine and Surgery, Karolinska Institutet, Stockholm, 17176, Sweden

46 19 Department of Medicine, Division of Cardiovascular Medicine, Stanford University School of  
47 Medicine, Stanford, CA 94305, USA.  
48 20 Pfizer Worldwide Research and Development, Stockholm, 17176, Sweden  
49

50 **Name and complete address of corresponding author**

51 Anders Mälarstig, Centre for Molecular Medicine, L8:02, 171 76 Stockholm, Sweden,

52 Fax number: + 46 8-31 31 47

53 Telephone: +46 8 55052514

54 Email: [anders.malarstig@ki.se](mailto:anders.malarstig@ki.se) , [anders.malarstig@pfizer.com](mailto:anders.malarstig@pfizer.com),

55 **Abstract (max 300 words)**

56 Recent advances in highly multiplexed immunoassays have allowed systematic large-scale  
57 measurement of hundreds of plasma proteins in large cohort studies. In combination with genotyping,  
58 such studies offer the prospect to 1) identify mechanisms involved with regulation of protein  
59 expression in plasma, and 2) determine whether the plasma proteins are likely to be causally  
60 implicated in disease. We report here the results of genome-wide association (GWA) studies of 83  
61 proteins considered relevant to cardiovascular disease (CVD), measured in 3,394 individuals with  
62 multiple CVD risk factors. We identified 79 genome-wide significant ( $p < 5e-8$ ) association signals, 55  
63 of which replicated at  $P < 0.0007$  in separate validation studies ( $n = 2,639$  individuals). Using automated  
64 text mining, manual curation, and network-based methods incorporating information on expression  
65 quantitative trait loci (eQTL), we propose plausible causal mechanisms for 25 trans-acting loci,  
66 including a potential post-translational regulation of stem cell factor by matrix metalloproteinase 9  
67 and receptor-ligand pairs such as RANK-RANK ligand. Using public GWA study data, we further  
68 evaluate all 79 loci for their causal effect on coronary artery disease, and highlight several potentially  
69 causal associations. Overall, a majority of the plasma proteins studied showed evidence of regulation  
70 at the genetic level. Our results enable future studies of the causal architecture of human disease,  
71 which in turn should aid discovery of new drug targets.

72

73 **Author Summary (150-200 words)**

74 Several proteins that circulate in blood have been linked to cardiovascular disease through the use of  
75 classic epidemiology and correlation studies. If individuals with higher risk of disease have higher  
76 levels of a protein, the protein may be associated with disease. However, this does not necessarily  
77 mean that the protein causes disease; it may merely be an innocent bystander or a consequence of the  
78 disease process. To establish whether a protein causes disease, a genetic approach, insensitive to  
79 reverse causation, can be used. Instead of correlating the levels of the protein itself, gene variants that  
80 regulate the protein levels are used in the analysis. This approach requires prior knowledge of which  
81 genetic variants are linked to individual proteins. Therefore we completed a map of how common  
82 genetic variants affect the blood concentration levels of 83 proteins that have been implicated in  
83 cardiovascular disease. By using this map of cause-to-effect findings, we gained insights into the  
84 regulation of a majority of the proteins under study and how they relate to risk of coronary artery  
85 disease. This study provides a map of genetic regulation of important cardiovascular plasma proteins,  
86 insights into their upstream regulatory environment, as well as novel leads for cardiovascular drug  
87 development.

88

## 89 **Introduction**

90 Cardiovascular disease (CVD), especially coronary artery disease (CAD) is a leading cause of human  
91 morbidity and mortality. Data from the world health organization (WHO) showed that CVD caused  
92 approximately 17.5 million deaths in 2012, corresponding to 31% of all deaths globally. Of these 7.4  
93 million were estimated to be due to coronary heart disease and 6.7 million to stroke [1].

94 Specific and mechanistically relevant biomarkers are important tools in risk prediction, disease  
95 diagnosis and successful development of new therapies [2]. Proteins in the circulation have been  
96 extensively explored as biomarkers across numerous disease conditions, not least because of the  
97 relative ease with which blood plasma and serum can be accessed, stored and analysed in  
98 observational studies and randomized controlled trials.

99 The usefulness of a plasma biomarker in disease prediction, or as surrogate endpoint in a clinical trial,  
100 depends on its specificity and sensitivity. These metrics reflect the relationship of the biomarker with  
101 a pre-specified disease endpoint, but are inherently influenced by biological factors such as the tissue  
102 expression, stability, regulation and variability of the biomarker. The genetic contribution to the  
103 variability of plasma biomarkers can be explored in genome-wide association (GWA) studies using  
104 single nucleotide polymorphisms (SNPs), and this approach has been applied to uncover numerous  
105 such relationships [3–5]. For distinct plasma biomarkers such as circulating proteins, the associations  
106 are also known as protein quantitative trait loci (pQTLs) [6–9].

107 Genetic loci for biomarkers and pQTLs have wide applicability in research. Firstly, pQTLs in trans  
108 can identify previously unknown regulatory pathways. Using trans-pQTLs to discover regulatory  
109 pathways is beneficial because it is based on in-vivo human observations that have well-established  
110 direction of causality, flowing from SNP to protein [7]. This approach has been extensively used in-  
111 vitro, for example in yeast studies [8], and the overall goal of such analysis is a deeper understanding  
112 of the regulatory check-points giving rise to a particularly biomarker concentration. For a biomarker  
113 that is causally involved in disease, e.g. low-density lipoprotein cholesterol (LDL-C), this is crucial  
114 knowledge as it allows targeting of upstream factors, e.g. HMG-CoA reductase.

115 Secondly, GWA study loci associated with circulating levels of plasma biomarkers that are predictive  
116 of disease risk enable evaluation of whether the biomarker association with disease is likely to be a  
117 causal relationship, using Mendelian randomization (MR). For example, although both c-reactive  
118 protein (CRP) and LDL-C predict risk of CVD and are lowered by treatment with statins, MR studies  
119 have concluded that plasma LDL-C is an aetiologically important factor, while plasma CRP is a  
120 biomarker that is not causally related to CVD [10,11]. Similarly, all efforts towards HDL-cholesterol  
121 lowering drugs have failed, consistent with MR results showing that SNPs affecting HDL-levels are  
122 unrelated to risk of CVD [12]. Based on these experiences of pharmacological treatment lowering the  
123 LDL-C concentration, one may suggest that a biomarker which is both predictive and causal provides  
124 a more attractive target for novel therapeutics. Numerous associations between biomarkers and  
125 disease have been described in the literature, but the potential causal involvement of these biomarkers  
126 has only been addressed for a limited number, partly due to a lack of robust genetic predictors for  
127 many plasma proteins.

128 In the present study, we analyzed 83 plasma proteins using the Olink ProSeek CVD array in 3,394  
129 European subjects with at least 3 established CVD risk factors. The majority of these proteins are  
130 strong candidates for involvement in atherosclerosis, plaque rupture or thrombosis and many are  
131 upregulated in CVD patients compared to controls or predict future risk of CVD events, such as CAD.

132 The aims of the study were to i) identify genetic loci for circulating plasma proteins that have  
133 previously been connected with CVD, ii) explore the mechanisms underpinning novel loci by  
134 integrating genetics with other biological information and iii) apply the tools to test causality in CAD.

## 135 **Results**

136 Of 83 proteins selected for known involvement in vascular disease and inflammation [13]  
137 (supplementary table S3), we observed 79 SNP-trait associations, consisting of 78 SNPs and their  
138 associations with 56 proteins (figure 1 and table 1). Of the 79 associations, 41 were cis effects, where  
139 the index-SNP is within 500 kb of the gene encoding the measured plasma protein. The functional

140 effect at each of these 41 loci is likely to be a direct effect either on the sequence of the plasma protein  
141 or on regulatory variants proximal to the encoding gene. Additionally, we identified 38 trans effects,  
142 all acting over distances more than 100 MB or at different chromosomes from the gene encoding the  
143 associated protein. Both cis and trans findings represent new understanding of the direct regulation of  
144 candidate CVD proteins, with trans findings additionally providing an opportunity for new insight  
145 into regulatory pathways.

146 We could replicate all but 6 of the pQTLs at nominal significance ( $P < 0.05$ ) measured in three  
147 independent cohorts ( $n = 2,639$ ). All but 16 of the measureable pQTLs were found to be reproducible at  
148  $P < 0.0007$  (Bonferroni corrected value). An additional 8 pQTLs were not measured in the replication  
149 cohorts. All 79 SNP-trait associations are reported in table 1 along with indication of replication  
150 status. Detailed replication statistics is available in supplementary table S1.

#### 151 **Protein quantitative trait loci acting in trans**

152 For each of the reported trans associations, we evaluated the most likely cis-gene intermediary, and  
153 investigated pathways in the direction of the plasma protein (table 2). Analysis of coding proxies  
154 revealed that 10 trans loci had missense mutations in linkage disequilibrium (LD) with the index-  
155 SNPs, providing an obvious explanatory model for the mechanism of action.

156 The analysis of cis-eQTLs in 11 large eQTL data sets provided evidence for an additional 13 mediator  
157 cis-genes. Some of the findings were remarkably independent of tissue and cell-type, and showed  
158 concordant results in several of the 11 eQTL datasets under analysis, as indicated in table 2. At each  
159 locus with significant cis-eQTL association, we additionally investigated neighbouring eQTL and  
160 pQTL effects as LocusZoom plots (supplementary figure S2). In some cases, like rs4810479/KITLG,  
161 the index-SNP shows both the strongest association with KITLG and the strongest cis-gene  
162 association (PLTP in liver). However, cases also exist, like rs200373/CTSL1, where stronger eQTL  
163 effects for the candidate cis-gene intermediary exists from other SNPs, with low LD between the  
164 SNPs precluding straightforward interpretation. Further studies would be required to address this  
165 issue.

166 In pathway analysis using the String-database of protein interactions, an additional 6 trans-genes were  
167 highlighted as possible mediator genes through functional protein connections. The criterion in this  
168 analysis was that less than 5% of randomly re-wired networks had shorter distance, dictating simply  
169 that connections of length 1 from a cis-gene to the trait gene should be selected. Additionally, a more  
170 sophisticated weighted network analysis was performed where each path through the network was  
171 weighted by the strength of the (trans) eQTL of the index-SNP. The eQTL values were calculated  
172 using a large collection of eQTL databases with tissue and cells relevant to cardiovascular disease.  
173 Like in the unweighted network analysis permutation was used to determine significance threshold.  
174 Through this weighted network analysis approach we discovered 11 additional mediator candidates,  
175 examples being the rs61598054 -> *FOXO3* -> *AKT1* -> and the rs693918 -> *XDH* -> *TLR4* -> *IL18*  
176 that are illustrated in figure 2A and 2B.

177 Systematic literature mining suggested an additional 5 possible mediators. Co-occurrence in scientific  
178 abstracts can indicate real biological relationships that may be missing from the String network.  
179 Interestingly, across all trans-pQTL loci, the largest number of abstract co-occurrences was 626 for  
180 the receptor-ligand pair encoded by *TNFSF11* and *TNFRSF11B*, a protein-protein interaction also  
181 reported in String-db.

182 The results of these five mediator-gene assigning approaches are summarised in table 2. While  
183 examples given above provide relatively clear indications of mechanism, more challenging cases do  
184 exist: the IL6-SNP rs10947260 for example gives evidence pointed at three candidate cis-mediator  
185 genes: *NOTCH4*, *AGER*, and *ATF6B*. *NOTCH4* and *ATF6B* were identified as containing FDR5%  
186 significant pathways to the destination IL6 (figure 2D). *AGER* was mentioned together with IL6 in 64  
187 separate publications. *AGER* and *ATF6B* exhibit weak cis-eQTL effects with the index-SNP. All  
188 were found at some distance from the index-SNP (>180 kb). Further experimentation is required to  
189 establish the main mechanism in this case.



190 **Pleiotropy of loci affecting protein levels**

191 Inspection of potential pleiotropic effects of index SNPs on measured protein traits as described in  
192 Methods revealed 6 distinct candidate loci (supplemental figure S1). The *ABO* locus affecting *THBD*,  
193 *TEK*, *F3*, *PECAM1*, and *SELE* in our dataset and the *FUT2* locus affecting *MMP10*, *F3*, and *LGALS3*  
194 are well known for their pleiotropic effects [14]. Furthermore, all SNPs affecting BNP levels seem to  
195 impact NPPB levels as well indicating their effect on steps before cleavage of the precursor protein.  
196 NTproBNP is a prohormone with an inactive N-terminal part that is cleaved to produce the active  
197 BNP. However, because of its half-life NTproBNP is typically used as a prognostic biomarker. A  
198 locus within the *ZFPM2* gene seems to have a strong effect on *PDGFB*, *DDK1*, and, to a lesser extent,  
199 on *VEGFA*. Finally, the cluster of cis-acting variants in the *MMP1*, *MMP3*, and *MMP12* loci are not  
200 specific to only one of the proteins but seem to impact all three of the metalloproteinases in this  
201 genomic region.

202 Additionally, we investigated the known associations of the index-SNPs with a broad range of other  
203 phenotypes, as previously reported in literature (supplemental table S2)..

204 **Associations between plasma proteins and cardiovascular risk**

205 To assess a potential causal involvement of each protein in CAD, we calculated genetic risk scores  
206 from the publically available CARDIoGRAMplusC4D GWAS data with the aim to construct a more  
207 powerful genetic instrument for those markers for which there were multiple SNPs. First, a systematic  
208 look-up of all reported pQTL-SNPs was performed to test for association with CAD (table 3). Then,  
209 we further explored proteins with multiple independent loci by calculating pooled SNP scores per  
210 protein, thus creating more powerful instruments to analyze the causality for proteins with multiple  
211 SNPs. Results show that of the SNPs contributing to the concentrations of proteins (Table 1), eight  
212 were also significantly associated with risk of CAD (Table 3). These findings suggest a causal role for  
213 these proteins, and whilst the cis *IL6R* finding confirms previous observations [15], the other  
214 observations extend our knowledge of important factors in CVD. Results from pooled-scores include  
215 highlights such as the multi-SNP support of *LGALS3* and the contradiction of *CHI3L1* having a  
216 CAD-associated trans-effect but no CAD-association in the cis-loci.

## 217 **Discussion**

218 In this study, we identified 79 pQTLs by measuring 83 plasma proteins of cardiovascular interest in a  
219 cohort of 3,394 subjects with multiple risk CVD risk factors, which may increase the power to detect  
220 genetic variants associated with CAD-associated proteins. The study provided novel insights into 57  
221 of the plasma proteins under investigation, including cis- and trans genetic regulation and effects of  
222 long-distance regulation networks and tentative evidence for causal involvement in CVD. To the best  
223 of our knowledge only a few of the findings were previously known; however reassuringly these  
224 replicated as expected: IL18/rs75649625 and rs4129267/IL6R [16], as well as AGER/sRAGE, CD40  
225 and LGALS3 cis associations [14,17,18] and the rs8176741/TEK trans association [19], and the  
226 rs635634/SELE [20]

### 227 **Insights into specific trans-effects**

228 A proteomics GWA study provides an interesting opportunity for the study of trans-regulatory effects,  
229 because the trait is a well-defined biological entity. In some cases, the trans-pQTL investigating  
230 methods in table 2 converged on a very plausible candidate gene. For example, at the CCL4-  
231 rs62625034 locus the effector transcript is probably the *CCR5* gene, while at the TNFSF11-rs7813952  
232 locus the effector transcript is likely the *TNFRSF11B* gene, two examples of known ligand-receptor  
233 pairs. Another example is the IL27-rs4905 variant, which sits within the *EBI3* gene. The *IL27* and  
234 *EBI3* genes encode the two subunits of the IL27 cytokine complex.

235 The effector transcript at the KITLG-rs4810479 locus may be *MMP9*, which encodes a  
236 metalloproteinase that cleaves the KITLG gene product, a membrane-bound stem cell factor [21].  
237 Thus this trans pQTL may represent an example of genetic regulation via post-translational  
238 modification.

239 At a few loci, we found multiple lines of evidence suggesting different mediator genes at the same  
240 locus. This is not biologically impossible, nor is it uncommon in literature [22], but it does require  
241 more careful analysis. The challenge is illustrated by the IL6-SNP rs10947260, for which separate  
242 lines of evidence pointed to three candidate cis-mediator genes. As shown in figure 2D, a criticism

243 against concluding on the importance of a pathway to IL6 through the *CCND1* gene is that *NOTCH4*  
244 has many neighbours in the String-network, thereby increasing the risk of a spurious discovery.

245 While these examples seem specific, they illustrate challenges that have major consequences for the  
246 general interpretation of any genetic association results. Analyses such as these have driven the  
247 development of popular risk-gene assignment tools (e.g. [23]). Our findings illustrate the increased  
248 power of knowing a certain pathway destination through the use of pQTL.

249

### 250 **Insights into potential causal involvement of the plasma proteins in CVD**

251 The study provided an important opportunity to systematically test each of the plasma proteins for a  
252 potential causal role in CVD by investigating whether identified pQTLs also were associated with  
253 CAD risk. If an instrumental variable, e.g. a SNP or a set of SNPs, exclusively affects one factor, and  
254 also affects an overall phenotype, such as disease risk – then it may be deduced that the protein is  
255 causally involved in the development of this disease. According to this principle, eight proteins  
256 (PECAM1, SELE, F3, IL6R, CHI3L1, LGALS3, MMP12, and PDGFB) showed evidence of  
257 potentially causal involvement in CAD. The connection between IL6R and CAD has already been  
258 described [15], and several drug trials are underway to test whether an ILR6-inhibitor (tocilizumab) is  
259 effective in treatment of CAD (clinicaltrials.org). In light of this, the remaining proteins could be of  
260 interest as therapeutic targets.

261 There are some important limitations to the approach, as compared to a formal MR. A formal MR  
262 study requires that the genetic instrument is specific, is not in LD with other functional variants, and  
263 that there are no hidden population strata [24]. There is no reason to suspect that the second and third  
264 requirements were violated; the study was based on high-resolution imputation of cohorts that were  
265 ethnically homogeneous. Importantly, the specificity requirement was not always satisfied, weakening  
266 the findings for some proteins. This includes all the trans associations, as well as proteins for which  
267 pleiotropy was detected (supplemental figure S1 and table S2). In addition, association between

268 plasma protein concentrations per se and future CVD risk has not been carefully investigated for the  
269 majority of proteins included in the present study.

270 These limitations leave LGALS3, MMP12 and PDGFB as candidates for having a causal effect on  
271 CAD. Of the three SNPs affecting levels of LGALS3, rs1169306, rs7928577 and rs33988101 in trans,  
272 only the first two also contribute to CAD risk, resulting in a pooled CAD association P-value of  
273  $P=1.46e-4$ . For MMP12 and PDGFB, the results are based on single SNPs showing associations with  
274 protein levels. Of the three, only MMP12 is a cis effect thereby strengthening the case for it being a  
275 specific MR instrument. These limitations notwithstanding, the map of pQTLs presented here, and in  
276 particular those acting in cis, should provide the means to systematically assess potential causal roles  
277 of these biomarkers in other common complex diseases. Additionally we highlight the online resource  
278 found at [www.olink-improve.com](http://www.olink-improve.com) where the data pQTL can be browsed in greater detail. This may in  
279 turn help to prioritise drug targets for development of disease-modifying therapies.

## 280 **Conclusion**

281 In conclusion, the main contributions of this paper are: i) identification of 79 pQTLs regulating  
282 important circulating cardiovascular plasma proteins, ii) novel evidence of the regulatory mechanisms  
283 underpinning at least half of these novel loci and iii) evidence of potential causal roles in CAD  
284 development for several plasma proteins. We believe that these three principal findings provide a  
285 strong contribution to the field of cardiovascular biomarkers and beyond.

286

## 287 **Materials and Methods**

### 288 **The IMPROVE study**

289 The IMPROVE study is a multicentre, observational study, which recruited 3,711 men and women  
290 aged between 55 to 79 years with at least three cardiovascular risk factors but without symptoms of  
291 CVD (previously described [25]). Serum and plasma from the study participants were collected at  
292 baseline, dispensed in polypropylene tubes and frozen at  $-80^{\circ}\text{C}$  prior to shipment for centralized

293 biochemical analyses and biobanking at the Karolinska Institutet in Stockholm, Sweden. The study  
294 was approved by the ethics committees of the 7 centers of the discovery cohort [25] and by the  
295 Uppsala University ethics committee for the replication cohorts [26,27]. All participants gave written  
296 informed consent, and the study was conducted in accordance with the declaration of Helsinki.

### 297 **Genotyping, quality control and imputation**

298 DNA genotyping in the IMPROVE study was performed using the Illumina CardioMetabochip and  
299 Immunochip arrays. The combined SNP genotyping data from both platforms were merged and  
300 subjected to the following quality control (QC) using PLINK 1.7: SNPs were excluded for probe to  
301 genome mismatch, incorrect assignment of allelic variants in the array design, failed Hardy-Weinberg  
302 Equilibrium test at  $1 \times 10^{-6}$ , call rate  $< 95\%$  or failed Illumina genotype calling QC. Samples were  
303 excluded if they showed evidence of gender mismatch, abnormal inbreeding coefficient, failed cryptic  
304 relatedness test or had an overall sample call rate  $< 95\%$ . After quality control, a total number of  
305 3,394 subjects remained for analysis. Imputation was performed with IMPUTE 2.0 using the 1000  
306 genomes version 5 as reference panel. The pre-imputation data set contained 244,814 SNPs and the  
307 post-imputation data set contained 5,270,624 SNPs

### 308 **Plasma protein determinations and quality control**

309 In total, there were 3,394 IMPROVE participants for whom quality controlled genotype information  
310 and plasma samples were available. Plasma concentrations were measured in baseline EDTA plasma  
311 samples using the ProSeek CVD array I (Olink Biosciences, Uppsala, Sweden), according to the  
312 standard protocol. The ProSeek method is based on the highly sensitive and specific proximity  
313 extension assay (PEA), which involves the binding of distinct polyclonal oligonucleotide-labelled  
314 antibodies to the target protein followed by quantification by real-time quantitative PCR [13]. In  
315 addition to the controls provided by Olink Biosciences, a pooled plasma control was included in all  
316 plates to enable further quality control (QC) such as calculation of variation coefficients. Prior to  
317 statistical analyses, we excluded individual assays with more than 20 % of samples below the lower  
318 detection limit and those with final inter-plate coefficients of variation above 25 %. After QC, a total  
319 number of 83 proteins out of the 92 remained for analysis. The native scale of Olink protein

320 measurements is  $\log(2)$  but additional  $\log(10)$  transformations were performed to ensure normally  
321 distributed variables. Validation of the OLINK method has been conducted [13], and the method has  
322 been used to validate previous findings obtained with established protein quantification methods  
323 [26,28].

### 324 **Genome-wide quantitative trait locus discovery**

325 Plasma protein readings were  $\log_{10}$  transformed prior to analyses. Standardized residuals for each of  
326 the 83 plasma proteins were calculated using a linear model adjusting for age, sex, recruitment centre,  
327 protein analysis batch, smoking, diabetes and hypertension at baseline. To merge loci in table 1 and  
328 table S1, signals with  $R^2$  higher than 0.1 and distance within 250 KB were omitted, retaining only the  
329 strongest signal in each block, referred to as the index SNP. The standardized residuals were used in  
330 a Wald-test in PLINK 1.9 to test association between genetic data and each plasma protein, using a  
331 significance threshold of  $P < 5e-8$ . All summary statistics can be downloaded at [www.olink-](http://www.olink-improve.com)  
332 [improve.com](http://www.olink-improve.com) [review pass-mail: [rev3@ki.se](mailto:rev3@ki.se)].

333 Narrow-sense heritability for all proteins was calculated using Genome-Wide Complex Trait Analysis  
334 [29]. A genetic relationship matrix was calculated using all imputed autosomal SNPs with imputation  
335 scores above  $R^2=0.9$ , less than 1 % missingness and allele frequency above 5 %, in total 2,617,215  
336 SNPs. The variants were hard called before estimating the phenotypic variance explained by all  
337 autosomal SNPs using the restricted maximum likelihood analysis (REML).

### 338 **Replication of pQTL effects**

339 Replication studies of all pQTLs were performed in three community-based cohorts in which Olink  
340 array protein data and genotypes were available. These cohorts were the NSPHS [27], the Prospective  
341 Investigation of the Vasculature in Uppsala Seniors (PIVUS) and the Uppsala Longitudinal Study of  
342 Adult Men (ULSAM) [26], consisting of samples from 976, 933 and 730 participants, respectively.  
343 Statistics were calculated according to additive association models, and findings were matched either  
344 directly on imputed SNP-id (96% of cases) or using a proxy with  $R^2 > 0.8$  linkage disequilibrium.  
345 Replication P-values were calculated using the METAL meta-analysis software (version 2011-03-25).

346 **Expression quantitative trait analysis**

347 For each index-SNP, cis- and trans-eQTL data were calculated from the following sources: aorta  
348 intima-media, aorta adventitia, liver, mammary artery, and heart from the ASAP study [30],  
349 monocytes and B-cells from the Fairfax et al study [31], and monocytes stimulated with LPS-2h, LPS-  
350 24h and interferon-2h from another Fairfax et al study [32]. Each of these 11 data sets had information  
351 from gene expression microarrays and genotyping microarrays as described in the respective  
352 references. The mean sample size was 223 with a range of 89-367. Data from genotyping microarrays  
353 were imputed using the MACH 1.0 algorithm with 1000 genomes CEU data as reference (mean rsq  
354 quality score 0.89) [33]. The strength of eQTL association was calculated using a linear additive  
355 model between log2-transformed expression value and numerically encoded genotype data. For cis-  
356 eQTL associations, un-corrected p-values from cis-eQTL were reported if the association was  
357 stronger than  $P < 0.0005$  (corresponding to a false discovery rate (FDR)  $< 5\%$ ). For all significant cis-  
358 eQTL associations, locusZoom plots were generated showing regional effect differences between  
359 eQTL and pQTL studies [34].

360 **Network analysis**

361 The network analysis was performed based on the String database network (version 10) [35], using all  
362 edges with a confidence score above 400. For all genes within 0.5 MB of an effect-SNP (“cis-genes”),  
363 the shortest path length was calculated between the cis-gene and the gene encoding the measured  
364 protein biomarker (“trait-gene”) using the *igraph* package in R (version 1.0.1). This was done both  
365 with an unweighted version of the Stringdb-network as well as with a weighted version, wherein each  
366 gene along the path was weighted by the trans-eQTL strength calculated from the effect-SNP (scored  
367 as 1, except if  $P_{eQTL} < 0.05$  which gave score 0.8, and if  $P_{eQTL} < 0.005$ , which gave score 0.6).

368 For both weighted and unweighted networks, significance of a path was calculated as the fraction of  
369 1000 randomly permuted networks that obtained a shorter path length than the one tested. Random  
370 networks were generated using permutation of the original scores and random rewiring of the network  
371 using the *igraph* *rewire* function, as detailed in code repository [http://github.com/lassefolkersen/olink-](http://github.com/lassefolkersen/olink-improve)  
372 *improve*. Given our data, only paths of length 1, i.e. direct links in String-db, were significant at a

373 0.05 level in the unweighted case. For the weighted case, only paths of length 2 with an intermediate  
374 trans-eQTL gene reached significance. Paths were subsequently checked for biological plausibility.

### 375 **Literature analysis**

376 To support the assignment of potential causal genes in pQTLs, we mined the literature for topical co-  
377 occurrences of each gene in a pQTL (defined by a 500kb window) with its associated protein. The  
378 Pfizer-internal LitMS tool can provide such matches based on all PubMed abstracts, a large synonym  
379 dictionary and manually curated rules that limit findings to more relevant articles, e.g. those in which  
380 gene and protein occur in the abstract's title [36]. The system outputs the number of co-occurrences  
381 and underlying article references for each gene-protein input pair. We then reviewed the literature  
382 findings to assign the most plausible causal genes where possible.

### 383 **Pleiotropy**

384 To understand the specificity of all reported index-SNPs we inspected all index SNPs that had at least  
385 2 associations with distinct proteins at  $P < 0.05 / (83 * 79) = 7.7e - 6$ . This cutoff reflects a conservative  
386 approach to the multiple testing burden for all identified index SNPs (79) with all tested protein traits  
387 (83). The resulting association matrix was then clustered and visualized based on the negative log<sub>10</sub>  
388 of the p-values of association. For the clustering, we used a complete-linkage hierarchical clustering  
389 approach based on correlation coefficients as a metric. In addition, index-SNPs were investigated for  
390 other associations in publically available GWAS databases.

### 391 **Calculation of genetic risk scores**

392 To assess the effect on disease, the publicly available CARDIoGRAMplusC4D 1000G imputed data  
393 was interrogated [37]. The goal was to perform *in silico* analysis for every SNP that showed  
394 significant associations with any of the measured traits. For traits that had multiple associated SNPs,  
395 pooled scores per affected protein were calculated using the R-package *gtx* version 0.0.8. Specifically  
396 for the pooled risk scores, the alleles of each protein were encoded so that the coded allele was  
397 increasing CAD risk regardless of its protein concentration effect. This ensured that pooled effect  
398 sizes reflected uniform directionality on CAD risk.



399

400

## 401 **Acknowledgments**

402 LF holds a grant from Innovation Fund Denmark (145-2014-5). APM is a Wellcome Trust Senior  
403 Fellow in Basic Biomedical Science (grant number WT098017). SEH holds a Chair funded by the  
404 British Heart Foundation (PG08/008) and by the National Institute for Health Research University  
405 College London Hospitals Biomedical Research Centre.

406

## 407 **Authorship Contributions**

408 LF, EF, MSL, RJS, DZ and AM analysed the main data and wrote the manuscript. SE, ÅJ, SBE, SG,  
409 LL, CL, AM, VG, UG, and EI provided and analysed the replication data. All other authors  
410 contributed to the collection of samples and provided critical revision for intellectual content.

411

## 412 **Competing interests**

413 Ulf Gyllensten and Stefan Enroth are authors on a patent application entitled “‘Determination and  
414 analysis of Biomarkers in clinical samples’”; United Kingdom Patent Application Nos. 1414913.2 and  
415 1410956.5 (2014, Pending).

## 416 **References**

417 Reference List

418

- 419 1. Shanthi Mendis, Pekka Puska, Bo Norrving (2011) Global Atlas on cardiovascular disease prevention and control.  
420 World Health Organization ISBN 978-92-4-156437-3.: 3-18.
- 421 2. Gashaw I, Ellinghaus P, Sommer A, Asadullah K (2012) What makes a good drug target? Drug Discov Today 17  
422 Suppl: S24-S30. S1359-6446(11)00435-1 [pii];10.1016/j.drudis.2011.12.008 [doi].
- 423 3. Welter D, MacArthur J, Morales J, Burdett T, Hall P, Junkins H, Klemm A, Flicek P, Manolio T, Hindorff L, Parkinson  
424 H (2014) The NHGRI GWAS Catalog, a curated resource of SNP-trait associations. Nucleic Acids Res 42:  
425 D1001-D1006. gkt1229 [pii];10.1093/nar/gkt1229 [doi].

- 426 4. Enroth S, Johansson A, Enroth SB, Gyllensten U (2014) Strong effects of genetic and lifestyle factors on biomarker  
427 variation and use of personalized cutoffs. *Nat Commun* 5: 4684. ncomms5684  
428 [pii];10.1038/ncomms5684 [doi].
- 429 5. Enroth S, Bosdotter ES, Johansson A, Gyllensten U (2015) Effect of genetic and environmental factors on protein  
430 biomarkers for common non-communicable disease and use of personally normalized plasma protein  
431 profiles (PNPPP). *Biomarkers* 20: 355-364. 10.3109/1354750X.2015.1093546 [doi].
- 432 6. Kettunen J, Tukiainen T, Sarin AP, Ortega-Alonso A, Tikkanen E, Lyytikäinen LP, Kangas AJ, Soininen P, Wurtz P,  
433 Silander K, Dick DM, Rose RJ, Savolainen MJ, Viikari J, Kahonen M, Lehtimäki T, Pietiläinen KH, Inouye  
434 M, McCarthy MI, Jula A, Eriksson J, Raitakari OT, Salomaa V, Kaprio J, Jarvelin MR, Peltonen L, Perola M,  
435 Freimer NB, Ala-Korpela M, Palotie A, Ripatti S (2012) Genome-wide association study identifies  
436 multiple loci influencing human serum metabolite levels. *Nat Genet* 44: 269-276. ng.1073  
437 [pii];10.1038/ng.1073 [doi].
- 438 7. Hause RJ, Stark AL, Antao NN, Gorsic LK, Chung SH, Brown CD, Wong SS, Gill DF, Myers JL, To LA, White KP, Dolan  
439 ME, Jones RB (2014) Identification and validation of genetic variants that influence transcription factor  
440 and cell signaling protein levels. *Am J Hum Genet* 95: 194-208. S0002-9297(14)00314-0  
441 [pii];10.1016/j.ajhg.2014.07.005 [doi].
- 442 8. Foss EJ, Radulovic D, Shaffer SA, Goodlett DR, Kruglyak L, Bedalov A (2011) Genetic variation shapes protein  
443 networks mainly through non-transcriptional mechanisms. *PLoS Biol* 9: e1001144.  
444 10.1371/journal.pbio.1001144 [doi];PBIOLGY-D-11-00024 [pii].
- 445 9. Sun W, Kechris K, Jacobson S, Drummond MB, Hawkins GA, Yang J, Chen TH, Quibrera PM, Anderson W, Barr RG,  
446 Basta PV, Bleecker ER, Beaty T, Casaburi R, Castaldi P, Cho MH, Comellas A, Crapo JD, Criner G, Demeo  
447 D, Christenson SA, Couper DJ, Curtis JL, Doerschuk CM, Freeman CM, Gouskova NA, Han MK, Hanania  
448 NA, Hansel NN, Hersh CP, Hoffman EA, Kaner RJ, Kanner RE, Kleerup EC, Lutz S, Martinez FJ, Meyers DA,  
449 Peters SP, Regan EA, Rennard SI, Scholand MB, Silverman EK, Woodruff PG, O'Neal WK, Bowler RP  
450 (2016) Common Genetic Polymorphisms Influence Blood Biomarker Measurements in COPD. *PLoS*  
451 *Genet* 12: e1006011. 10.1371/journal.pgen.1006011 [doi];PGENETICS-D-15-02045 [pii].
- 452 10. Casas JP, Shah T, Cooper J, Hawe E, McMahon AD, Gaffney D, Packard CJ, O'Reilly DS, Juhan-Vague I, Yudkin JS,  
453 Tremoli E, Margaglione M, Di MG, Hamsten A, Kooistra T, Stephens JW, Hurel SJ, Livingstone S, Colhoun  
454 HM, Miller GJ, Bautista LE, Meade T, Sattar N, Humphries SE, Hingorani AD (2006) Insight into the  
455 nature of the CRP-coronary event association using Mendelian randomization. *Int J Epidemiol* 35: 922-  
456 931. dy1041 [pii];10.1093/ije/dy1041 [doi].
- 457 11. Wensley F, Gao P, Burgess S, Kaptoge S, Di AE, Shah T, Engert JC, Clarke R, Davey-Smith G, Nordestgaard BG,  
458 Saleheen D, Samani NJ, Sandhu M, Anand S, Pepys MB, Smeeth L, Whittaker J, Casas JP, Thompson SG,  
459 Hingorani AD, Danesh J (2011) Association between C reactive protein and coronary heart disease:  
460 mendelian randomisation analysis based on individual participant data. *BMJ* 342: d548.
- 461 12. Voight BF, Peloso GM, Orho-Melander M, Frikke-Schmidt R, Barbalic M, Jensen MK, Hindy G, Holm H, Ding EL,  
462 Johnson T, Schunkert H, Samani NJ, Clarke R, Hopewell JC, Thompson JF, Li M, Thorleifsson G, Newton-  
463 Cheh C, Musunuru K, Pirruccello JP, Saleheen D, Chen L, Stewart A, Schillert A, Thorsteinsdottir U,  
464 Thorgeirsson G, Anand S, Engert JC, Morgan T, Spertus J, Stoll M, Berger K, Martinelli N, Girelli D,  
465 McKeown PP, Patterson CC, Epstein SE, Devaney J, Burnett MS, Mooser V, Ripatti S, Surakka I,  
466 Nieminen MS, Sinisalo J, Lokki ML, Perola M, Havulinna A, de FU, Gigante B, Ingelsson E, Zeller T, Wild  
467 P, de Bakker PI, Klungel OH, Maitland-van der Zee AH, Peters BJ, de BA, Grobbee DE, Kamphuisen PW,  
468 Deneer VH, Elbers CC, Onland-Moret NC, Hofker MH, Wijmenga C, Verschuren WM, Boer JM, van der  
469 Schouw YT, Rasheed A, Frossard P, Demissie S, Willer C, Do R, Ordovas JM, Abecasis GR, Boehnke M,  
470 Mohlke KL, Daly MJ, Guiducci C, Burt NP, Surti A, Gonzalez E, Purcell S, Gabriel S, Marrugat J, Peden J,  
471 Erdmann J, Diemert P, Willenborg C, König IR, Fischer M, Hengstenberg C, Ziegler A, Buyschaert I,  
472 Lambrechts D, Van de Werf F, Fox KA, El Mokhtari NE, Rubin D, Schrezenmeier J, Schreiber S, Schafer A,  
473 Danesh J, Blankenberg S, Roberts R, McPherson R, Watkins H, Hall AS, Overvad K, Rimm E, Boerwinkle  
474 E, Tybjaerg-Hansen A, Cupples LA, Reilly MP, Melander O, Mannucci PM, Ardissino D, Siscovick D,  
475 Elosua R, Stefansson K, O'Donnell CJ, Salomaa V, Rader DJ, Peltonen L, Schwartz SM, Altshuler D,  
476 Kathiresan S (2012) Plasma HDL cholesterol and risk of myocardial infarction: a mendelian  
477 randomisation study. *Lancet* 380: 572-580. S0140-6736(12)60312-2 [pii];10.1016/S0140-  
478 6736(12)60312-2 [doi].

- 479 13. Assarsson E, Lundberg M, Holmquist G, Bjorkestén J, Thorsen SB, Ekman D, Eriksson A, Rennel DE, Ohlsson S,  
480 Edfeldt G, Andersson AC, Lindstedt P, Stenvang J, Gullberg M, Fredriksson S (2014) Homogenous 96-  
481 plex PEA immunoassay exhibiting high sensitivity, specificity, and excellent scalability. *PLoS One* 9:  
482 e95192. 10.1371/journal.pone.0095192 [doi];PONE-D-13-52390 [pii].
- 483 14. de Boer RA, Verweij N, van Veldhuisen DJ, Westra HJ, Bakker SJ, Gansevoort RT, Muller Kobold AC, van Gilst WH,  
484 Franke L, Mateo L, I, van der Harst P (2012) A genome-wide association study of circulating galectin-3.  
485 *PLoS One* 7: e47385. 10.1371/journal.pone.0047385 [doi];PONE-D-12-19830 [pii].
- 486 15. Interleukin-6 Receptor Mendelian Randomisation Analysis (IL6R MR) Consortium (2012) The interleukin-6  
487 receptor as a target for prevention of coronary heart disease: a mendelian randomisation analysis.  
488 *Lancet* 379: 1214-1224. S0140-6736(12)60110-X [pii];10.1016/S0140-6736(12)60110-X [doi].
- 489 16. Melzer D, Perry JR, Hernandez D, Corsi AM, Stevens K, Rafferty I, Lauretani F, Murray A, Gibbs JR, Paolisso G,  
490 Rafiq S, Simon-Sanchez J, Lango H, Scholz S, Weedon MN, Arepalli S, Rice N, Washecka N, Hurst A,  
491 Britton A, Henley W, van de Leemput J, Li R, Newman AB, Tranah G, Harris T, Panicker V, Dayan C,  
492 Bennett A, McCarthy MI, Ruukonen A, Jarvelin MR, Guralnik J, Bandinelli S, Frayling TM, Singleton A,  
493 Ferrucci L (2008) A genome-wide association study identifies protein quantitative trait loci (pQTLs).  
494 *PLoS Genet* 4: e1000072. 10.1371/journal.pgen.1000072 [doi].
- 495 17. Jiang DK, Ma XP, Yu H, Cao G, Ding DL, Chen H, Huang HX, Gao YZ, Wu XP, Long XD, Zhang H, Zhang Y, Gao Y,  
496 Chen TY, Ren WH, Zhang P, Shi Z, Jiang W, Wan B, Saiyin H, Yin J, Zhou YF, Zhai Y, Lu PX, Zhang H, Gu X,  
497 Tan A, Wang JB, Zuo XB, Sun LD, Liu JO, Yi Q, Mo Z, Zhou G, Liu Y, Sun J, Shugart YY, Zheng SL, Zhang XJ,  
498 Xu J, Yu L (2015) Genetic variants in five novel loci including CFB and CD40 predispose to chronic  
499 hepatitis B. *Hepatology* 62: 118-128. 10.1002/hep.27794 [doi].
- 500 18. Cheng DT, Kim DK, Cockayne DA, Belousov A, Bitter H, Cho MH, Duvoix A, Edwards LD, Lomas DA, Miller BE,  
501 Reynaert N, Tal-Singer R, Wouters EF, Agusti A, Fabbri LM, Rames A, Visvanathan S, Rennard SI, Jones P,  
502 Parmar H, MacNee W, Wolff G, Silverman EK, Mayer RJ, Pillai SG (2013) Systemic soluble receptor for  
503 advanced glycation endproducts is a biomarker of emphysema and associated with AGER genetic  
504 variants in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 188: 948-  
505 957. 10.1164/rccm.201302-0247OC [doi].
- 506 19. Lieb W, Chen MH, Larson MG, Safa R, Teumer A, Baumeister SE, Lin H, Smith HM, Koch M, Lorbeer R, Volker U,  
507 Nauck M, Volzke H, Wallaschofski H, Sawyer DB, Vasan RS (2015) Genome-wide association study for  
508 endothelial growth factors. *Circ Cardiovasc Genet* 8: 389-397. CIRCGENETICS.114.000597  
509 [pii];10.1161/CIRCGENETICS.114.000597 [doi].
- 510 20. Qi L, Cornelis MC, Kraft P, Jensen M, van Dam RM, Sun Q, Girman CJ, Laurie CC, Mirel DB, Hunter DJ, Rimm E, Hu  
511 FB (2010) Genetic variants in ABO blood group region, plasma soluble E-selectin levels and risk of type 2  
512 diabetes. *Hum Mol Genet* 19: 1856-1862. ddq057 [pii];10.1093/hmg/ddq057 [doi].
- 513 21. Oriss TB, Krishnamoorthy N, Raundhal M, Morse C, Chakraborty K, Khare A, Huff R, Ray P, Ray A (2014) Cutting  
514 Edge: MMP-9 inhibits IL-23p19 expression in dendritic cells by targeting membrane stem cell factor  
515 affecting lung IL-17 response. *J Immunol* 192: 5471-5475. jimmunol.1303183  
516 [pii];10.4049/jimmunol.1303183 [doi].
- 517 22. Musunuru K, Strong A, Frank-Kamenetsky M, Lee NE, Ahfeldt T, Sachs KV, Li X, Li H, Kuperwasser N, Ruda VM,  
518 Pirruccello JP, Muchmore B, Prokunina-Olsson L, Hall JL, Schadt EE, Morales CR, Lund-Katz S, Phillips  
519 MC, Wong J, Cantley W, Racie T, Ejebe KG, Orho-Melander M, Melander O, Kotliansky V, Fitzgerald K,  
520 Krauss RM, Cowan CA, Kathiresan S, Rader DJ (2010) From noncoding variant to phenotype via SORT1  
521 at the 1p13 cholesterol locus. *Nature* 466: 714-719. nature09266 [pii];10.1038/nature09266 [doi].
- 522 23. Pers TH, Karjalainen JM, Chan Y, Westra HJ, Wood AR, Yang J, Lui JC, Vedantam S, Gustafsson S, Esko T, Frayling T,  
523 Speliotes EK, Boehnke M, Raychaudhuri S, Fehrmann RS, Hirschhorn JN, Franke L (2015) Biological  
524 interpretation of genome-wide association studies using predicted gene functions. *Nat Commun* 6:  
525 5890. ncomms6890 [pii];10.1038/ncomms6890 [doi].
- 526 24. Burgess S, Malarstig A (2013) Using Mendelian randomization to assess and develop clinical interventions:  
527 limitations and benefits. *J Comp Eff Res* 2: 209-212. 10.2217/ce.13.14 [doi].

- 528  
529  
530  
531  
532  
533
25. Strawbridge RJ, Deleskog A, McLeod O, Folkersen L, Kavousi M, Gertow K, Baldassarre D, Veglia F, Leander K, Gigante B, Kauhanen J, Rauramaa R, Smit AJ, Mannarino E, Giral P, Dehghan A, Hofman A, Franco OH, Humphries SE, Tremoli E, de FU, Gustafsson S, Ostensson CG, Eriksson P, Ohrvik J, Hamsten A (2014) A serum 25-hydroxyvitamin D concentration-associated genetic variant in DHCR7 interacts with type 2 diabetes status to influence subclinical atherosclerosis (measured by carotid intima-media thickness). *Diabetologia* 57: 1159-1172. 10.1007/s00125-014-3215-y [doi].
- 534  
535  
536
26. Nowak C, Sundstrom J, Gustafsson S, Giedraitis V, Lind L, Ingelsson E, Fall T (2016) Protein Biomarkers for Insulin Resistance and Type 2 Diabetes Risk in Two Large Community Cohorts. *Diabetes* 65: 276-284. db15-0881 [pii];10.2337/db15-0881 [doi].
- 537  
538  
539
27. Igl W, Johansson A, Gyllensten U (2010) The Northern Swedish Population Health Study (NSPHS)--a paradigmatic study in a rural population combining community health and basic research. *Rural Remote Health* 10: 1363. 1363 [pii].
- 540  
541  
542  
543
28. Isgren A, Jakobsson J, Palsson E, Ekman CJ, Johansson AG, Sellgren C, Blennow K, Zetterberg H, Landen M (2015) Increased cerebrospinal fluid interleukin-8 in bipolar disorder patients associated with lithium and antipsychotic treatment. *Brain Behav Immun* 43: 198-204. S0889-1591(14)00474-7 [pii];10.1016/j.bbi.2014.10.001 [doi].
- 544  
545
29. Yang J, Lee SH, Goddard ME, Visscher PM (2011) GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet* 88: 76-82. S0002-9297(10)00598-7 [pii];10.1016/j.ajhg.2010.11.011 [doi].
- 546  
547  
548  
549
30. Folkersen L, Wagsater D, Paloschi V, Jackson V, Petrini J, Kurtovic S, Maleki S, Eriksson MJ, Caidahl K, Hamsten A, Michel JB, Liska J, Gabrielsen A, Franco-Cereceda A, Eriksson P (2011) Unraveling divergent gene expression profiles in bicuspid and tricuspid aortic valve patients with thoracic aortic dilatation: the ASAP study. *Mol Med* 17: 1365-1373. molmed.2011.00286 [pii];10.2119/molmed.2011.00286 [doi].
- 550  
551  
552
31. Fairfax BP, Makino S, Radhakrishnan J, Plant K, Leslie S, Dilthey A, Ellis P, Langford C, Vannberg FO, Knight JC (2012) Genetics of gene expression in primary immune cells identifies cell type-specific master regulators and roles of HLA alleles. *Nat Genet* 44: 502-510. ng.2205 [pii];10.1038/ng.2205 [doi].
- 553  
554  
555
32. Fairfax BP, Humburg P, Makino S, Naranbhai V, Wong D, Lau E, Jostins L, Plant K, Andrews R, McGee C, Knight JC (2014) Innate immune activity conditions the effect of regulatory variants upon monocyte gene expression. *Science* 343: 1246949. 343/6175/1246949 [pii];10.1126/science.1246949 [doi].
- 556  
557
33. Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR (2010) MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genet Epidemiol* 34: 816-834. 10.1002/gepi.20533 [doi].
- 558  
559  
560
34. Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, Boehnke M, Abecasis GR, Willer CJ (2010) LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics* 26: 2336-2337. btq419 [pii];10.1093/bioinformatics/btq419 [doi].
- 561  
562  
563  
564
35. Szklarczyk D, Franceschini A, Wyder S, Forslund K, Heller D, Huerta-Cepas J, Simonovic M, Roth A, Santos A, Tsafou KP, Kuhn M, Bork P, Jensen LJ, von MC (2015) STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Res* 43: D447-D452. gku1003 [pii];10.1093/nar/gku1003 [doi].
- 565
36. Roberts P, Bichko D, Crawford M, Klatte M, Xiang X (2016) LitMS v3.0 (Literature Mining System) (unpublished).
- 566  
567  
568  
569
37. 2015) A comprehensive 1000 Genomes-based genome-wide association meta-analysis of coronary artery disease. *Nat Genet* 47: 1121-1130. ng.3396 [pii];10.1038/ng.3396 [doi].
- 570

571 **Tables**572 **Table 1**

SNP id	Trait	-log(P)	SNP id	Trait	-log(P)
Cis-acting loci			Trans-acting loci		
rs1580006‡	ADM	14.69	rs184243355*	CCL3	7.65
rs2070600‡	AGER (RAGE)	9.52	rs73062378	CCL4	12.35
rs549596*	BNP	13.76	rs62625034	CCL4	40.51
rs2188974	CCL3	17.31	rs28601761	CHI3L1	8.3
rs6607368	CCL4	30.2	rs200373	CTSL1	8.37
rs1569723	CD40	48.52	rs6993770	DKK1	8.79
rs2153101	CHI3L1	107.13	rs495828	F3	9.34
rs17610659	CSF1	9.19	rs200433550*	F3	9.25
rs35285321	CSTB	42.93	rs1260326	FST (Follistatin)	8.69
rs111693235	CTSD	25.69	rs4672375	GAL	10.15
rs670211	CX3CL1	11.13	rs76519098†	GDF15	9.95
rs74544699	CXCL1	11.88	rs693918	IL18	10.62
rs35186877	CXCL16	8.76	rs7599125‡	IL18	7.95
rs72650832	CXCL6	41.21	rs35166255	IL1RL1	8.93
rs982764	FAS	11.7	rs11599750	IL27	9.85
rs3195944	GDF15	7.65	rs10947260†	IL6	9.74
rs6555820	HAVCR1	86.89	rs4810479	KITLG	10.35
rs13236526	HSPB1	16.96	rs7928577	LGALS3	8.67
rs139879640*	IL16	61.53	rs1169306‡	LGALS3	8.19
rs75649625	IL18	20.84	rs33988101‡	LGALS3	8.45
rs1420101	IL1RL1	131.69	rs12570111†	MMP1	7.33
rs4905	IL27	79.93	rs492602	MMP10	8.11
rs4129267	IL6R	264.67	rs12469459	MUC16	44.15
rs62115757	KLK11	61.91	rs61598054*	NGF	7.42
rs11667946	KLK6	14.47	rs75416436†	NGF	7.38
rs9323280	LGALS3	61.25	rs6557662*	NPPB	7.83
rs471994	MMP1	34.63	rs140000161	PAPPA	9.84
rs17368659	MMP12	96.26	rs16873402‡	PDGFB	7.62
rs7946057	MMP3	107.92	rs635634	PECAM1	44.72
rs56378716	MPO	8.73	rs117538444†	PGF	8.18
rs35207557*	NPPB	24.59	rs635634	SELE (E-selectin)	219.02
rs880949‡	PGF	7.8	rs8176741	TEK	49.06
rs116661163	REN (Renin)	7.99	rs8176693	THBD	9.95
rs1969539	SPON1	21.82	rs241771‡	TNFRSF11B	9.22
rs79250370	TEK (TIE2)	12.71	rs142552223	TNFSF11 (TRANCE)	16.47
rs3176123	THBD	23.64	rs7813952	TNFSF11 (TRANCE)	15.67

<b>rs6469811</b>	TNFRSF11B (Osteoprotegerin)	10.54	<b>rs35538083</b> †	XPNPEP2	7.51
<b>rs76769120</b> ‡	TNFRSF1B (TRAIL)	10.87	<b>rs11150189</b> ‡	XPNPEP2	13.16
<b>rs344560</b>	TNFSF14	17.53			
<b>rs2050011</b> *	XPNPEP2	67.62			
<b>rs2271025</b>	AGRP	8.63			

573

#### 574 **Overview of pQTL associations**

575 More commonly used non-systematic names indicated in parenthesis for some proteins. \* pQTL that  
 576 was not measured in replication cohorts, † pQTL that was measured in replication cohorts, but did not  
 577 replicate at  $P < 0.05$ , ‡ pQTL that did not replicate at Bonferroni corrected value of  $P < 0.0007$ . A more  
 578 detailed version of this table is found as supplemental table S1.

579

580

581

582

583

584

585

586

587 **Table 2**

trait-gene	SNP	cis-gene	Distance (kb)	Dist-rank	Coding-proxy	Cis-eQTL	Un-weighted-eQTL-pathway	Weighted-eQTL-pathway	Literature-score
CCL4	rs62625034	CCR5	0	1	rs62625034 (R <sup>2</sup> =1)				59
CTSL1	rs200373	IFI30	0	1		Monocytes+LPS (P=2.6e-05), Monocytes+IFN (P=1e-04)			
		MAST3	24	5	rs8108738 (R <sup>2</sup> =0.64)				
F3	rs495828	SURF6	43	2		Monocytes (P=2.9e-05), B-cells (P=3.4e-05)			
		MED22	53	3			Via PPARD (P=0.00321)		
FST	rs1260326	GCKR	0	1	rs1260326 (R <sup>2</sup> =1)				
		KRTCAP3	62	4		B-cells (P=3.4e-08)			
GDF15	rs76519098	MAPK8	283	4			Yes	Yes, short	
IL18	rs693918	XDH	-231	3				Via TLR4 (P=0.00085)	
IL18	rs7599125	LTBP1	-311	3				Via TGFB2 (P=0.00321)	
		NLRC4	-371	5			Yes	Yes, short	
IL1RL1	rs35166255	TIRAP	137	4			Yes	Yes, short	
		RPUSD4	-220	8		Monocytes+IFN (P=0.00034)			
IL27**	rs11599750	CWF19L1	187	6		4 eQTL-sets show cis-eQTL effect**			
IL6*	rs10947260	BTNL2	0	1	rs60263670 (R <sup>2</sup> =1)				
		NOTCH4	-181	6				Via CCND1 (P=0.00427)	
		AGER	-221	9					64
		ATF6B	-277	18				Via ATF3 (P=0.00349)	
KITLG	rs4810479	PLTP	-4	1		Liver (P=4.2e-09), B-cells (P=4.3e-07)			
		PCIF1	-18	3		Monocytes+IFN (P=5.4e-05)			
		ACOT8	-59	9		Monocytes+IFN (P=0.00021)			
		MMP9	-92	12			Yes	Yes, short	
LGALS3	rs7928577	TIRAP	63	3				Via IL6 (P=0.000463)	
		CDON	-295	9				Via CTNNA1 (P=0.00494)	
LGALS3	rs1169306	HNF1A	0	1	rs2464196 (R <sup>2</sup> =0.71)				
		C12orf43	3	2		5 eQTL-sets show cis-eQTL effect			
LGALS3	rs33988101	RASIP1	6	2	rs2287922 (R <sup>2</sup> =0.88)				
		FUT2	9	3	rs602662 (R <sup>2</sup> =0.68)				
		FGF21	-41	6				Via EGFR (P=0.000853)	
		BCAT2	80	10				Via GAPDH (P=0.000584)	



MMP10	rs492602	FUT2	0	1	rs601338 (R <sup>2</sup> =0.99)			
		RASIP1	17	3	rs2287922 (R <sup>2</sup> =0.68)			
		PPP1R15A -169		18				Via GADD45A (P=0.0045)
		BAX	-252	26				Via TNF (P=0.00461)
MUC16	rs12469459	GAL3ST2	0	1	rs12469459 (R <sup>2</sup> =1)			
		D2HGDH	8	2				Monocytes (P=9.6e-06)
NGF	rs61598054	FOXO3	-70	2				Via AKT1 (P=0.00376)
PAPPA	rs140000161	PRG2	0	1			Monocytes+IFN (P=5.4e-06)	Yes Yes, short
PECAM1	rs635634	SURF6	43	2			B-cells (P=1.7e-05), Monocytes (P=3.3e-05)	
SELE	rs635634	SURF6	43	2			B-cells (P=1.7e-05), Monocytes (P=3.3e-05)	
		MED22	53	3				Via PPARD (P=0.00277)
TEK	rs8176741	ABO	0	1	rs8176747 (R <sup>2</sup> =0.98)			
		MED22	76	5				Via ALB (P=0.00266)
		RPL7A	-84	6				Via UBC (P=0.000421)
		GBGT1	-92	9				Via ALB (P=0.00266)
THBD	rs8176693	ABO	0	1	rs8176746 (R <sup>2</sup> =1)			
TNFSF11	rs7813952	TNFRSF11-159 B		3			Yes	Yes, short 626

588

589 **Systematic analysis of potential mechanisms behind trans-pQTL associations.**

590 For each of 41 SNPs that had an effect in trans, cis-genes within 500 kb were analysed using 5  
591 different methods for evaluation of mediator cis-gene: 1) presence of non-synonymous coding SNP in  
592 LD with index SNP at R<sup>2</sup>>0.65, 2) presence of FDR5% cis-eQTL effect, 3) presence of significant  
593 pathway to trait-gene shorter than 95% of randomly permuted pathways, 4) presence of eQTL-  
594 weighted pathway to trait-gene shorter than 95% of randomly permuted pathways and/or 5) literature  
595 matching score above 50. A total of 1618 SNP-cis-gene pairs were considered, but only pairs that  
596 satisfied at least one of the tests are shown. \* Shown in figure 1B. \*\* Shown in figure 1A.

597

598

599

600

601 **Table 3**

<b>SNP</b>	<b>Trait- protein</b>	<b>Cis / trans</b>	<b>P<sub>protein</sub></b>	<b>β<sub>CAD</sub></b>	<b>P<sub>CAD</sub></b>	<b>β<sub>CAD-pool</sub></b>	<b>P<sub>CAD-pool</sub></b>
<b>rs635634</b>	PECAM1	trans	1.9E-45	0.08	<i>4.47E-11</i>		
<b>rs635634</b>	SELE	trans	9.6E-220	0.08	<i>4.47E-11</i>		
<b>rs495828</b>	F3	trans	4.5E-10	0.07	<i>1.29E-10</i>		
<b>rs4129267</b>	IL6R	cis	2.1E-265	0.05	<i>2.21E-07</i>		
<b>rs28601761</b>	CH13L1	trans	5.1E-09	0.05	<i>1.00E-06</i>	0.03	<i>2.3E-05</i>
<b>rs1169306</b>	LGALS3	trans	6.5E-09	0.03	<i>5.69E-04</i>	0.02	<i>5.9E-05</i>
<b>rs7928577</b>	LGALS3	trans	2.2E-09	0.06	<i>1.28E-03</i>	0.02	<i>5.9E-05</i>
<b>rs17368659</b>	MMP12	cis	5.5E-97	0.05	<i>1.39E-03</i>		
<b>rs16873402</b>	PDGFB	trans	2.4E-08	0.03	<i>1.47E-03</i>		
<b>rs6993770</b>	DKK1	trans	1.6E-09	0.03	<i>6.90E-03</i>		
<b>rs880949</b>	PGF	cis	1.6E-08	0.02	<i>2.00E-02</i>	0.02	<i>2.0E-02</i>
<b>rs17610659</b>	CSF1	cis	6.5E-10	0.02	<i>2.25E-02</i>		
<b>rs112579976</b>	CCL4	trans	2.5E-13	0.05	<i>3.03E-02</i>		
<b>rs9323280</b>	LGALS3	cis	5.6e-62	0.02	<i>3.20E-01</i>	0.02	<i>5.9E-05</i>
<b>rs2153101</b>	CH13L1	cis	7.5E-108	0.01	<i>4.68E-01</i>	0.03	<i>2.3E-05</i>
<b>rs33988101</b>	LGALS3	trans	3.6E-09	0.01	<i>5.16E-01</i>	0.02	<i>5.9E-05</i>

<b>rs117538444</b>	PGF	trans	6.5E-09	0.01	7.64E-01	0.02	2.0E-02
--------------------	-----	-------	---------	------	----------	------	---------

602 **Association between pQTLs and coronary artery disease (CAD) risk.**

603 Each SNP from supplemental table S1 was investigated in the CARDIoGRAMplusC4D data, and the  
604 P-values for the pQTL and CAD risk were extracted. An additional pooled analysis was performed in  
605 cases where one plasma protein had multiple pQTLs,. The table shows all pQTLs for which either a  
606 single-SNP or pooled CAD association had a  $P < 0.05$ . P-values highlighted in italics indicate that the  
607 association was also significant after correction for multiple testing.

608

609 Figure Legends

610 **Figure 1**

611 Genome-wide association strength of all measured plasma proteins.

612 The extent of each stack indicates the negative log P of association between the plasma protein and  
613 SNPs. Stacks with black dots and black text labels indicate cis-associations. Stacks with hollow  
614 circles and grey text labels indicate trans-associations; their targets are indicated with central colour  
615 coded lines. Consequently, plasma proteins having both cis- and trans-effects can be identified as  
616 those with a black dot stack as well as connecting lines from hollow dots, e.g. XPNPEP2 or CCL4.  
617 Fully drawn circle shows  $P=5e-8$ . Dashed circle shows  $1e-15$ . A detailed table of the genome-wide  
618 significant associations in this figure is available as supplemental table S1.

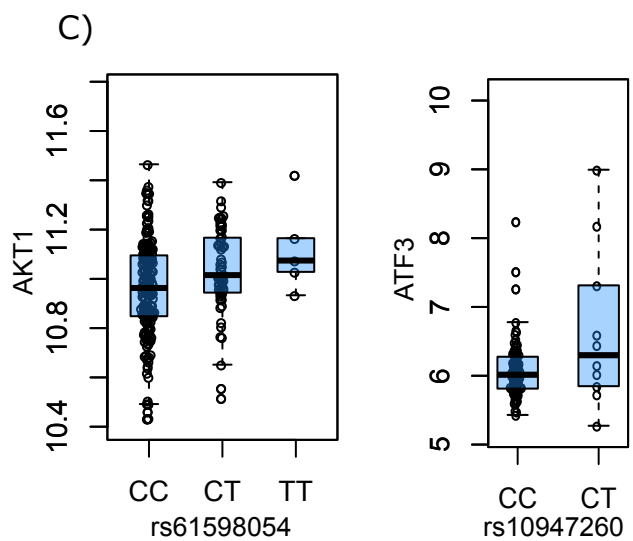
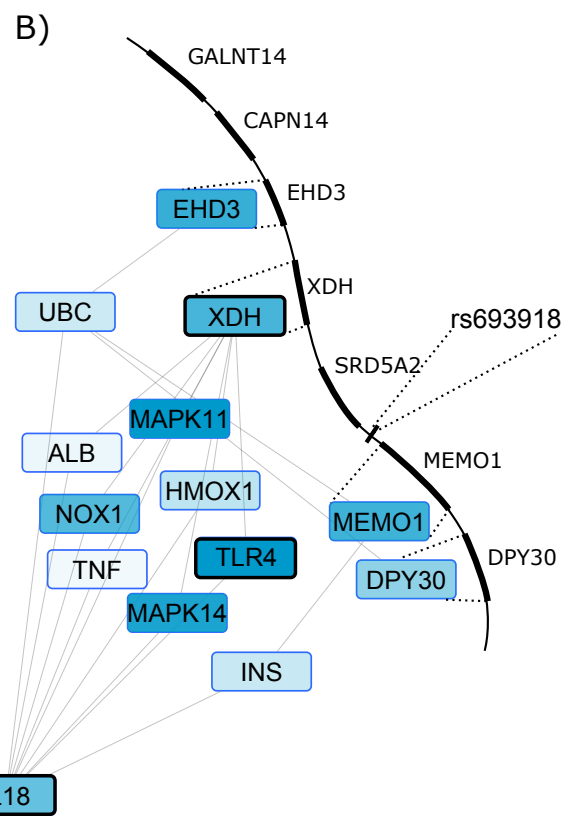
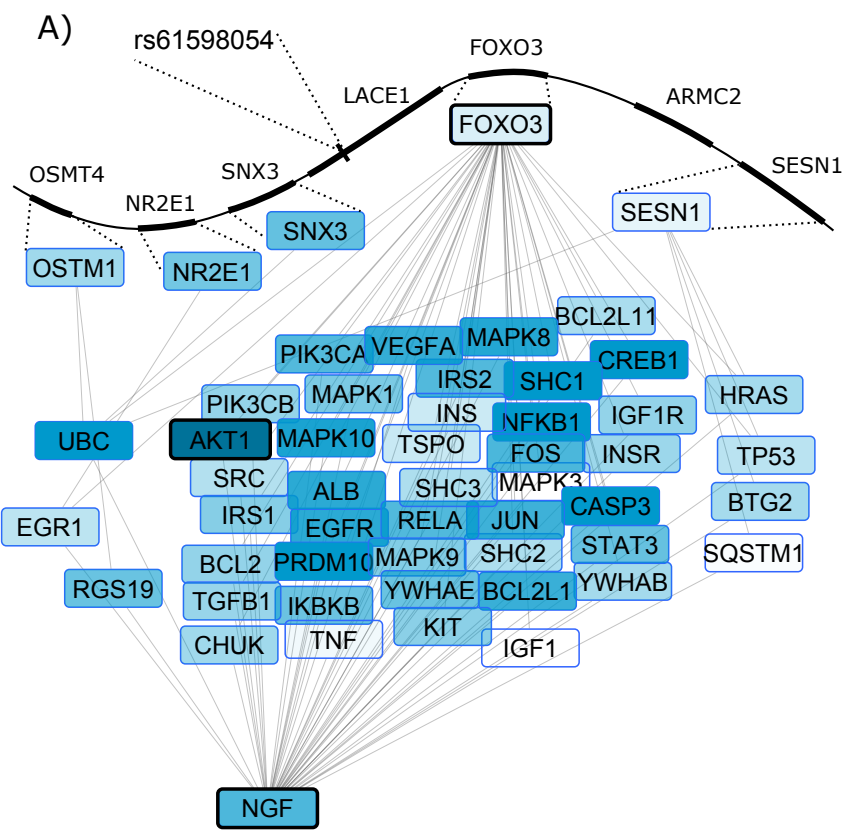
619

620 **Figure 2**

621 String-database network connections between proximal cis-gene and target plasma protein.

622 All short String paths that connect proximal cis-genes with the target plasma protein are shown. The  
623 colour intensity of each gene shows the eQTL association-strength with the index-SNP. The nodes  
624 highlighted with bold border show paths that satisfy  $P<0.05$  in network permutation analysis. **A)** the  
625 rs61598054-SNP is harboured in an intron of the *LACE1* gene, but have no paths to the target gene  
626 *NGF* and a more likely mechanism is therefore *FOXO3* -> *AKT1* -> *NGF*, which involves a  
627 rs61598054-trans-eQTL effect on *AKT1*. In permutation analysis of re-wired networks this is stronger  
628 than 95% of random networks. **B)** Similarly for rs693918, while located between *SRD5A2* and  
629 *MEMO1*, the path *XDH* -> *TLR4* -> *IL18* is a more likely mechanistic path, supported by eQTL  
630 effects on both *XDH* and *TLR4*. **C)** The rs61598054-*AKT1* trans-eQTL from panel A in 235 IFN-  
631 stimulated monocytes and the rs10947260-*ATF3* trans-eQTL from panel D in 89 mammary artery  
632 samples. **D)** Although rs10947260 is found in an intron of *BTNL2*, no obvious path exists between  
633 *BTNL2* and *IL6*. However, both *ATF6B* -> *ATF3* -> *IL6* and *NOTCH4* -> *CCND1* -> *IL6* are  
634 significant. DNA-strands are not to scale.

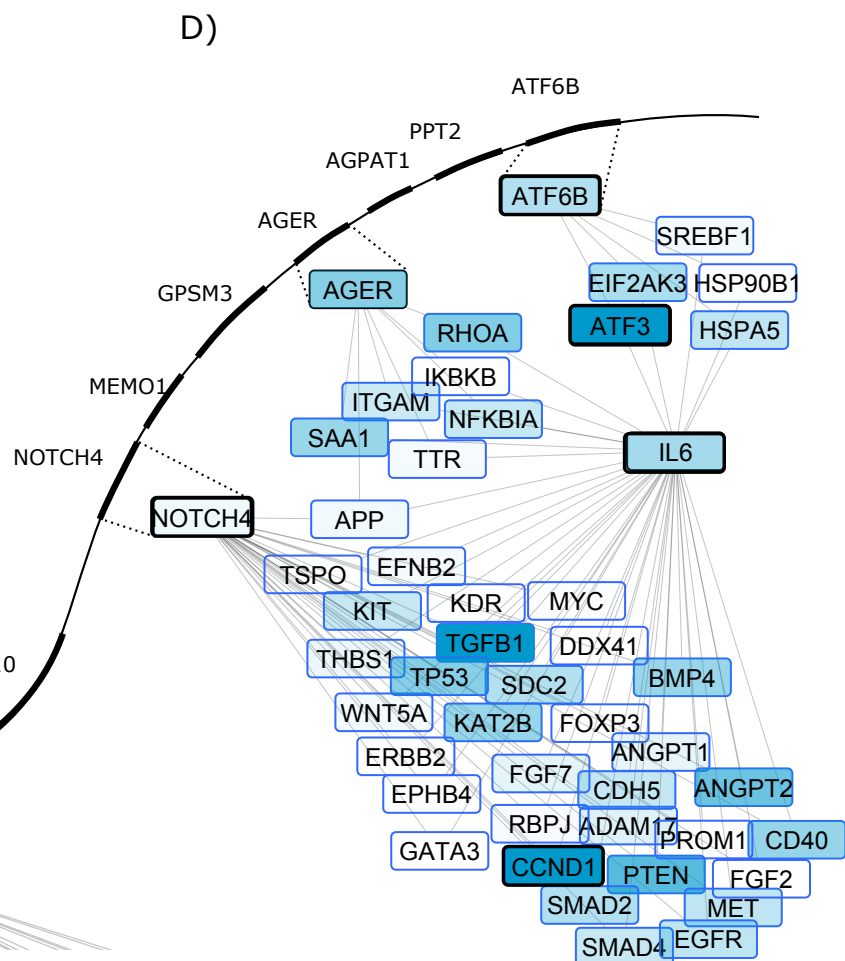




P = 0.05

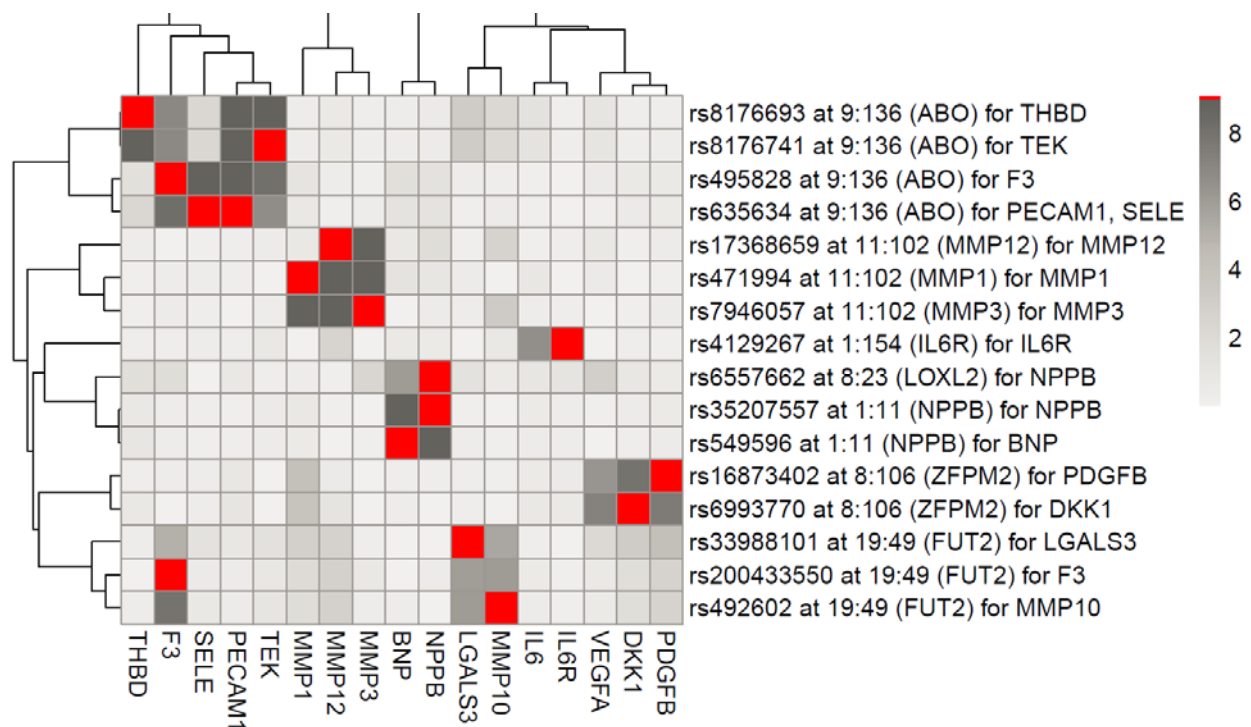
Trans-eQTL:

String-DB edge:



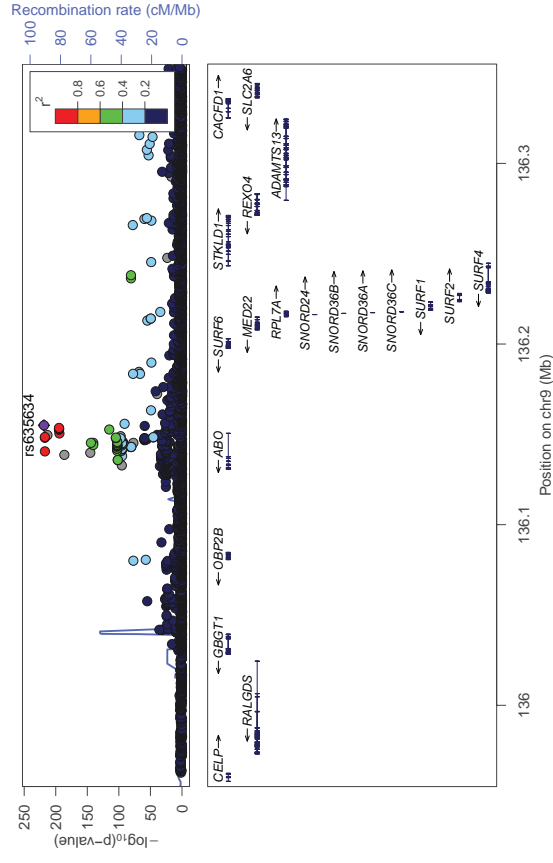
## SUPPLEMENTAL MATERIAL

**Figure S1**

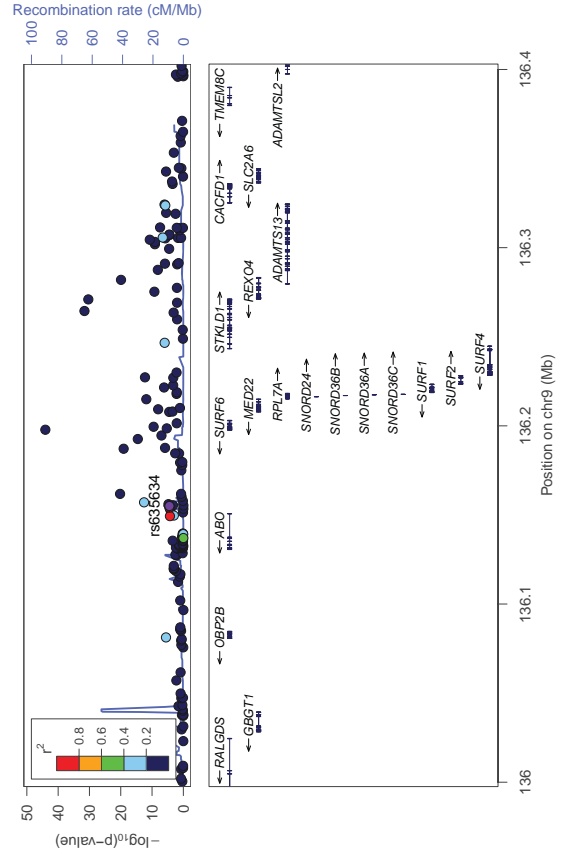


Potential pleiotropy between genome-wide significant SNPs and measured trait proteins. This figure shows all lead SNPs that have at least 2 associations with distinct proteins at  $P < 0.05 / (83 * 79) = 7.7e-6$ . This cutoff reflects a conservative approach to the multiple testing burden for all identified lead SNPs (79) with all tested protein traits (83). Protein traits are not displayed if they have no associations with the selected SNPs at the defined threshold. Red colour indicates the main effect as reported in table S1. Grey-scale colours indicate the effect strength on a  $-\log_{10}(P)$  scale as indicated.

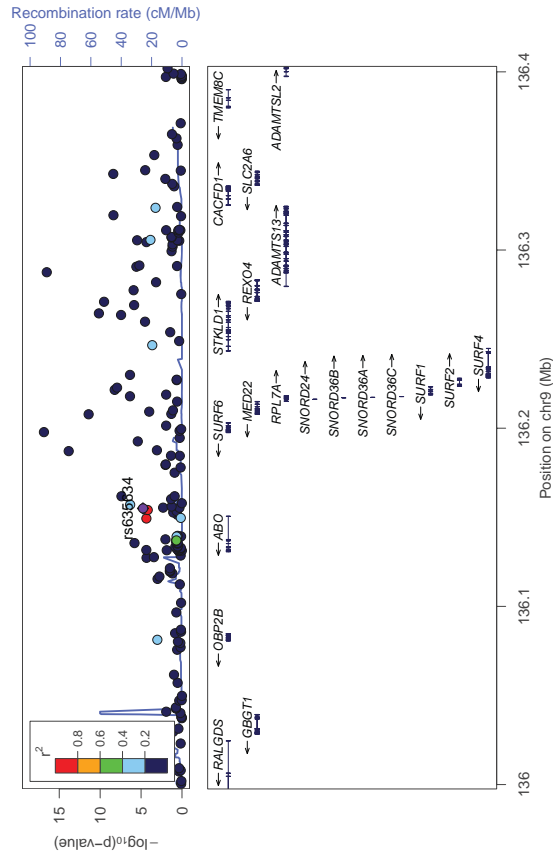
GWAS: rs635634/SURF6



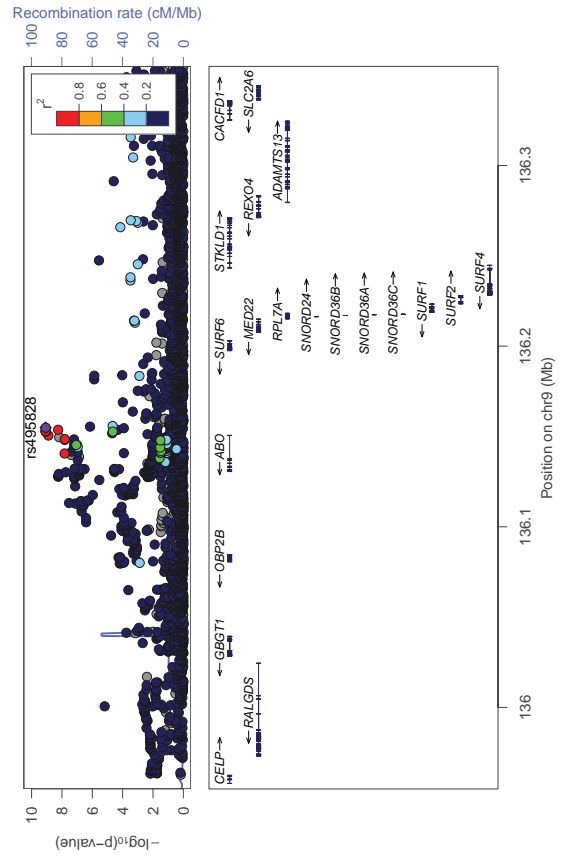
eQTL: rs635634/SURF6 in Monocytes



eQTL: rs635634/SURF6 in B-cells



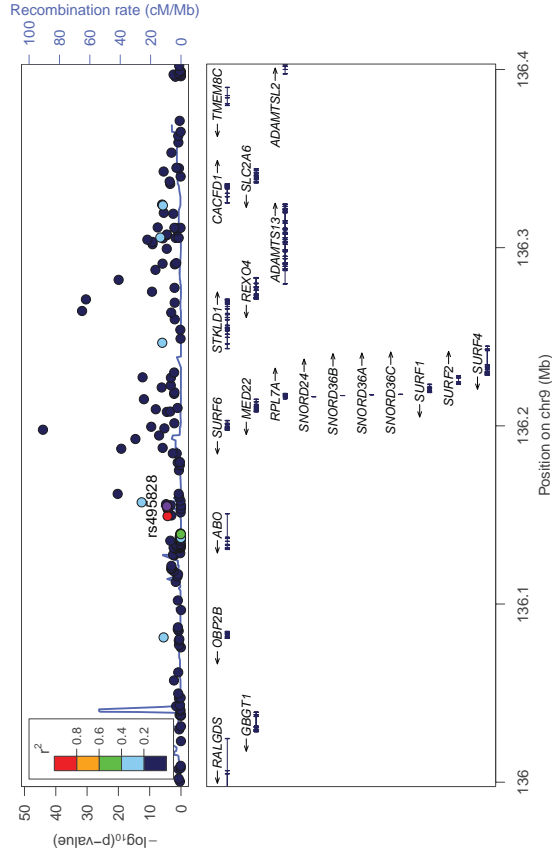
GWAS: rs495828/SURF6



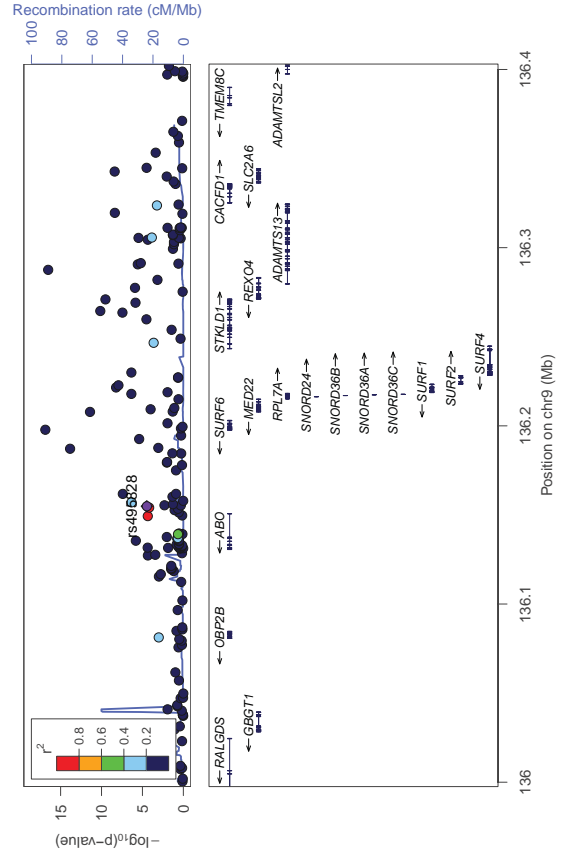


**Figure S2**

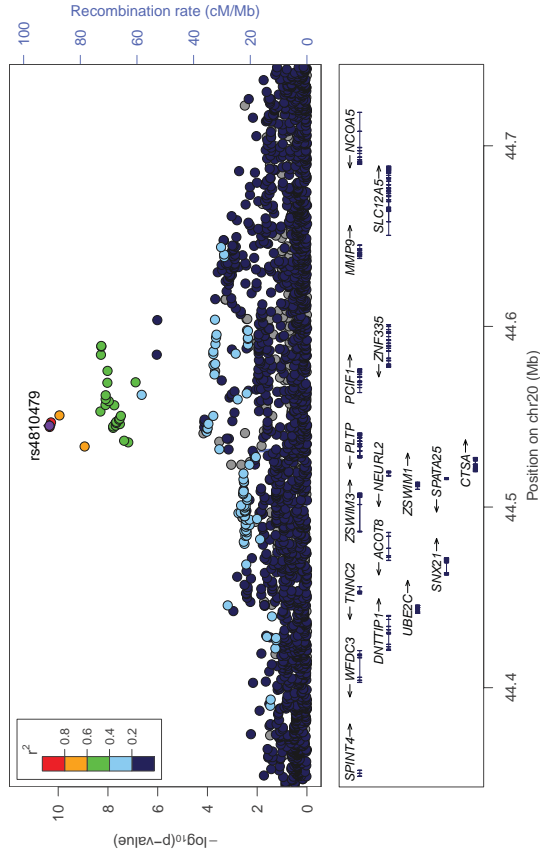
**eQTL: rs495828/SURF6 in Monocytes**



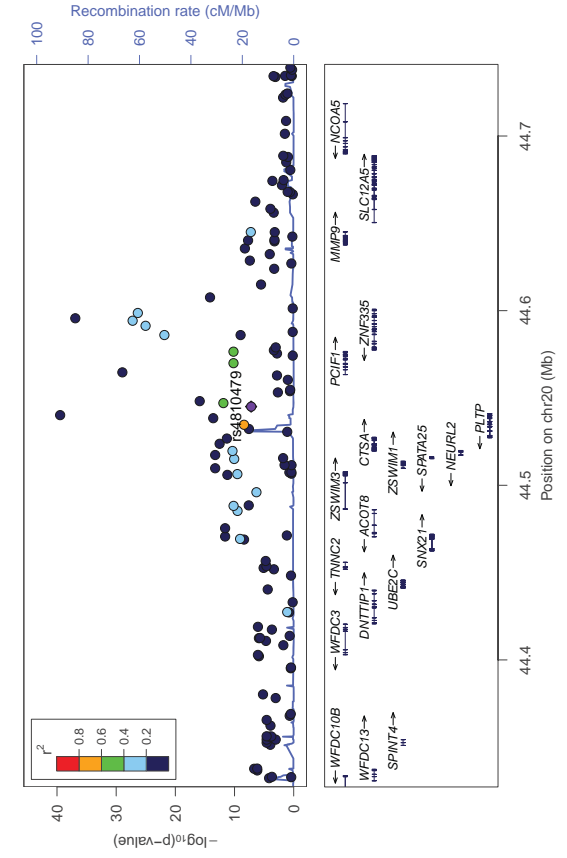
**eQTL: rs495828/SURF6 in B-cells**



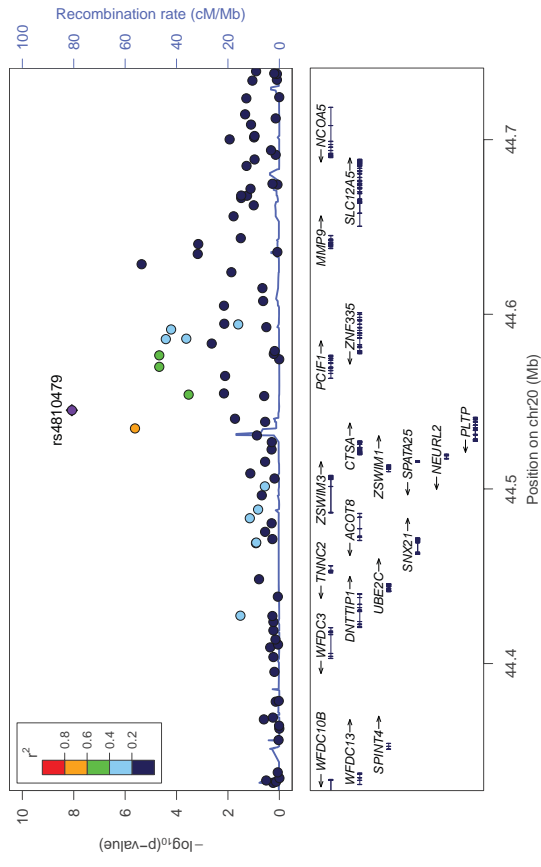
**GWAS: rs4810479/PLTP**



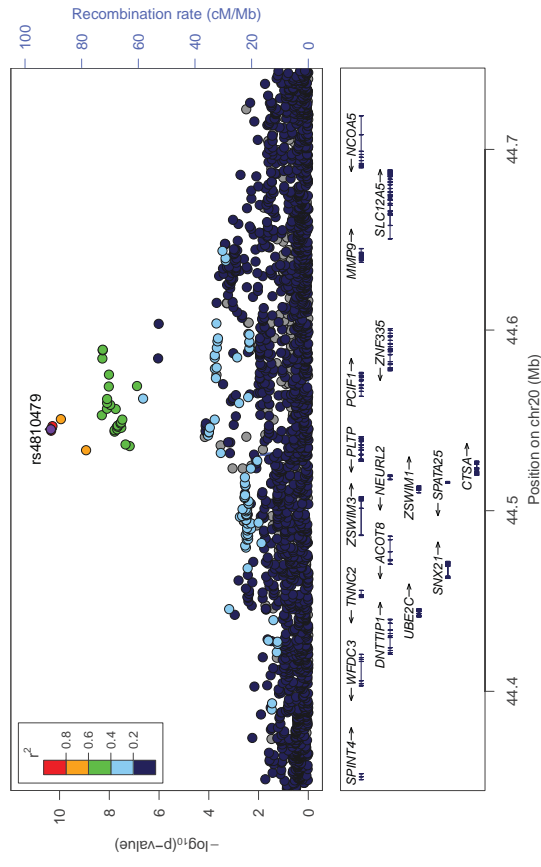
**eQTL: rs4810479/PLTP in B-cells**



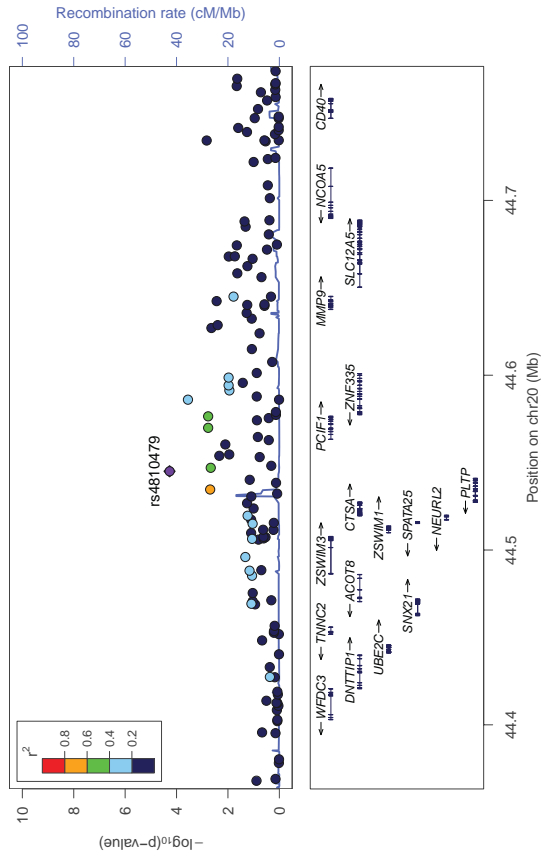
eQTL: rs4810479/PLTP in Liver



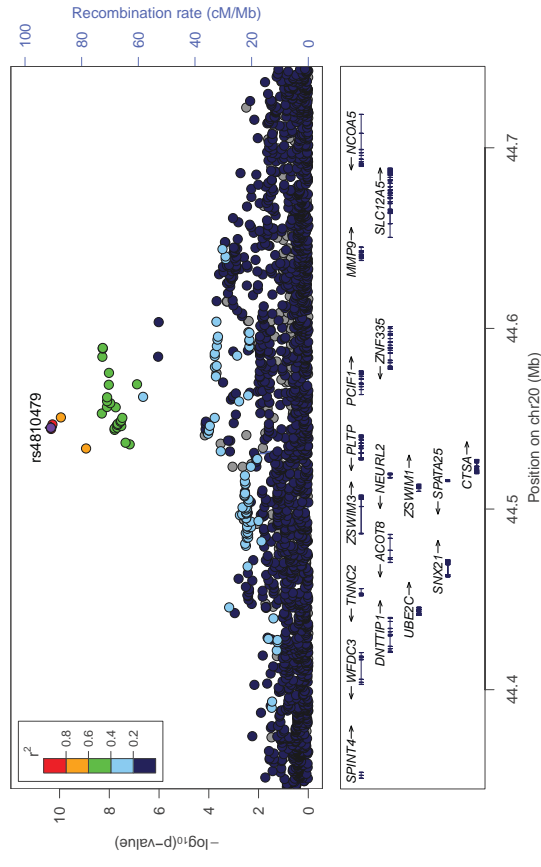
GWAS: rs4810479/PCIF1



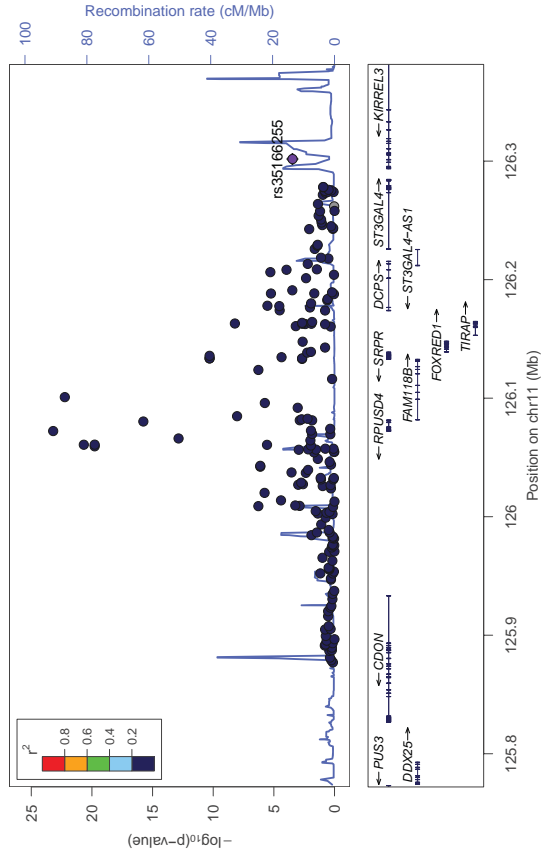
eQTL: rs4810479/PCIF1 in Monocytes+IFN



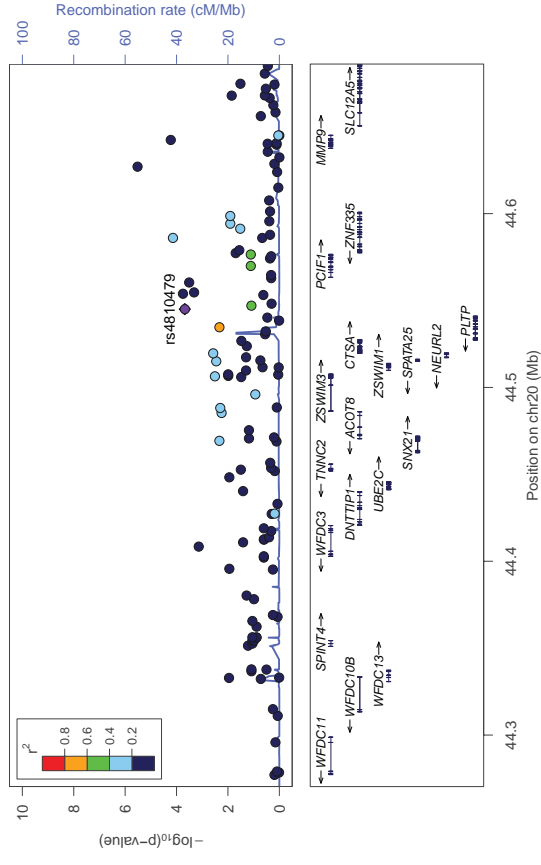
GWAS: rs4810479/ACOT8



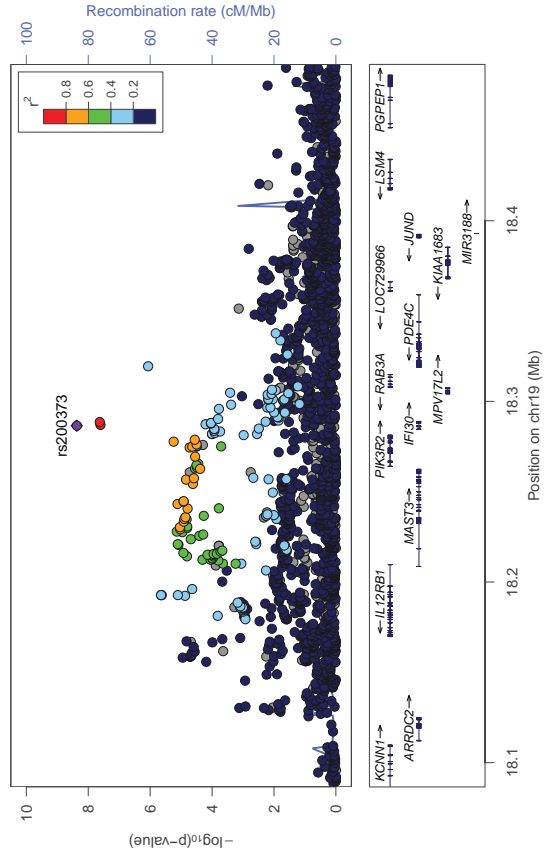
eQTL: rs35166255/RPUSD4 in Monocytes+IFN



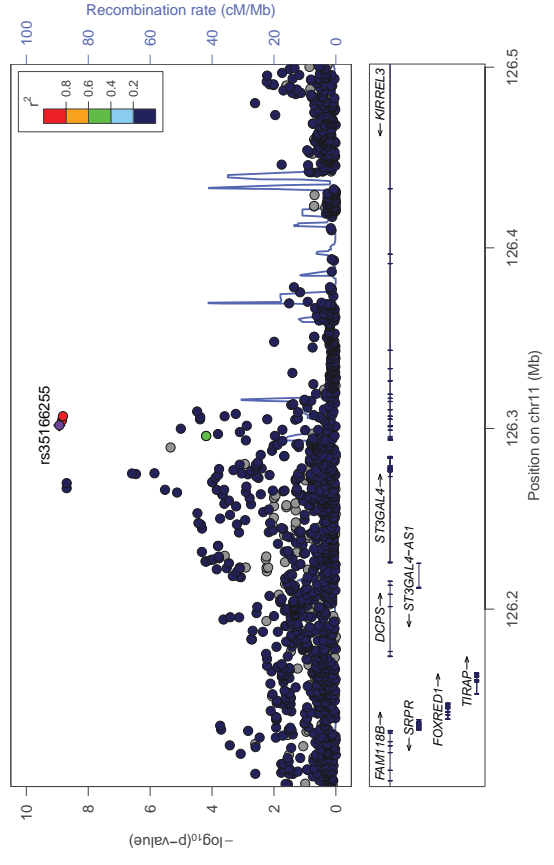
eQTL: rs4810479/ACOT8 in Monocytes+IFN



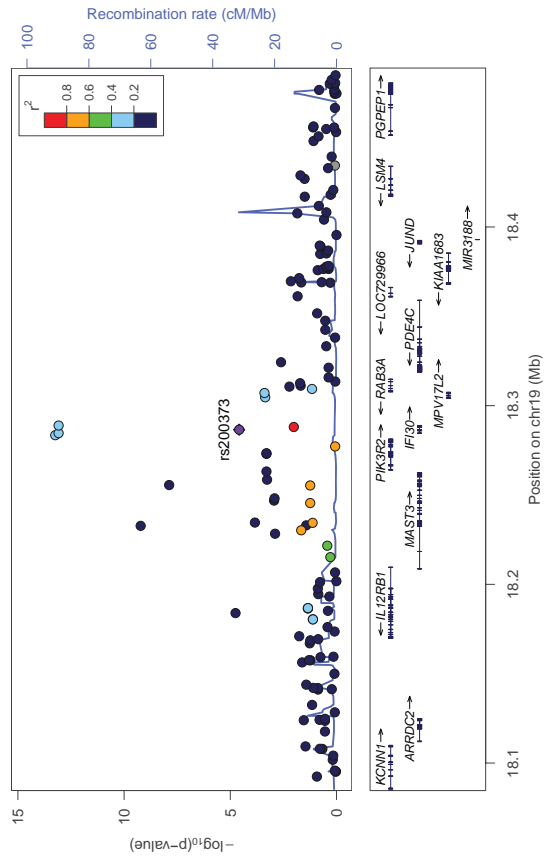
GWAS: rs200373/IFI30



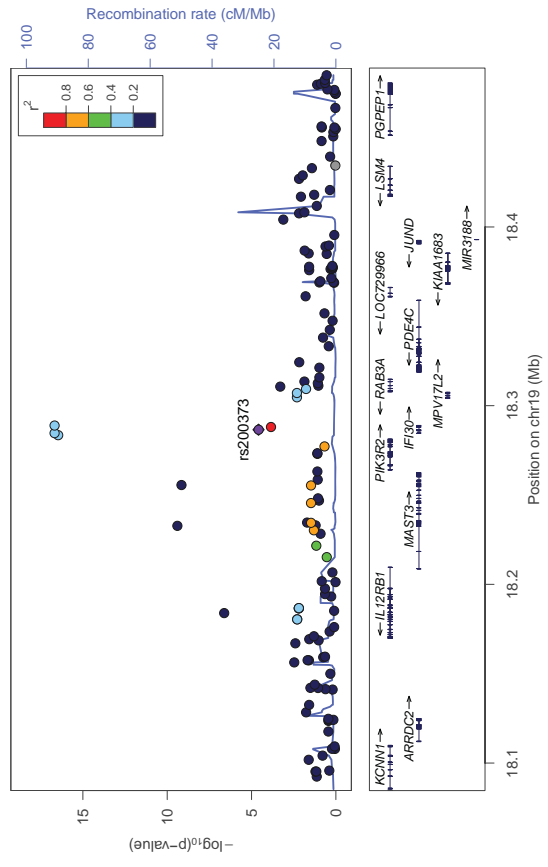
GWAS: rs35166255/RPUSD4



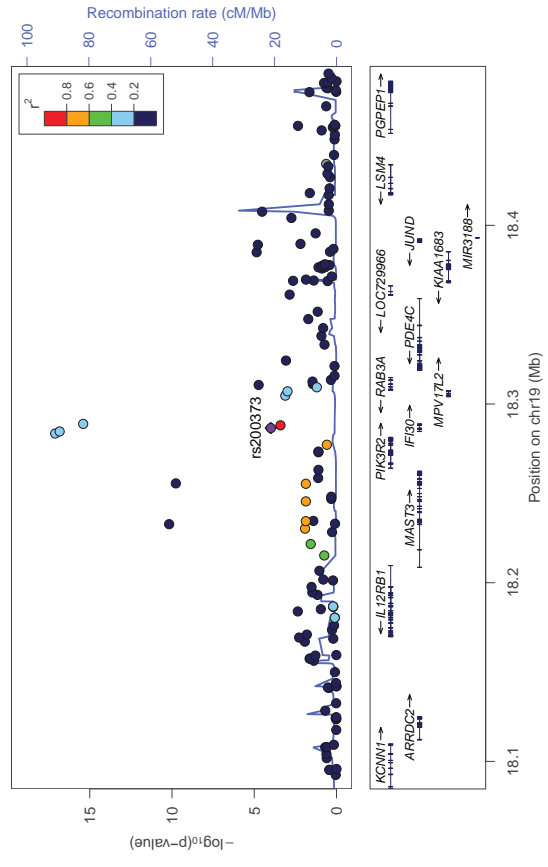
eQTL: rs200373/IFI30 in Monocytes+LPS



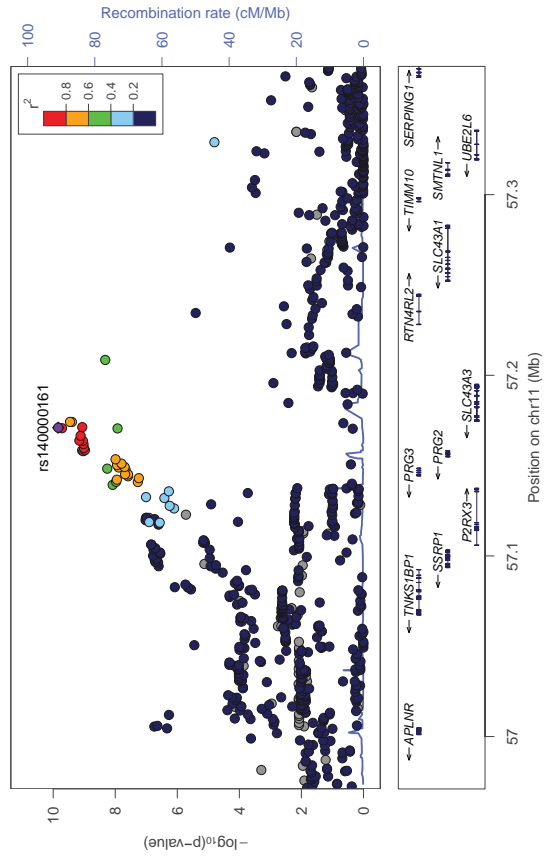
eQTL: rs200373/IFI30 in Monocytes+LPS



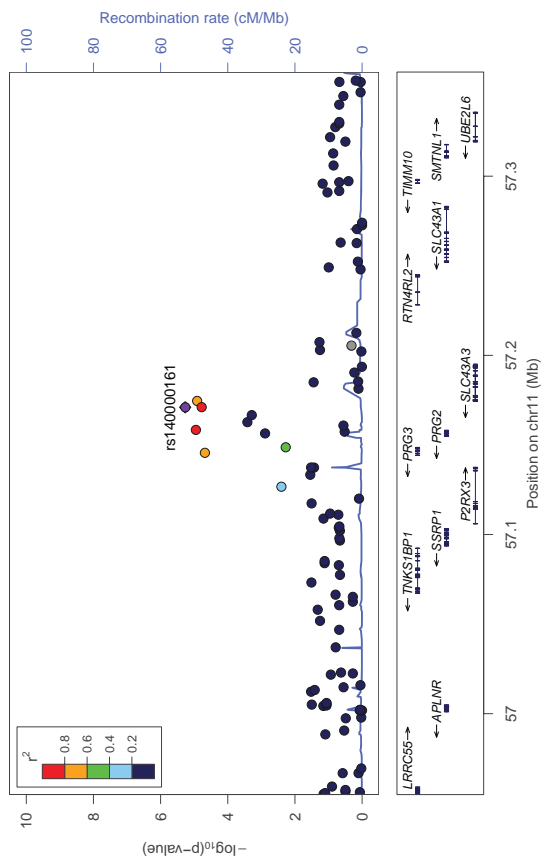
eQTL: rs200373/IFI30 in Monocytes+IFN



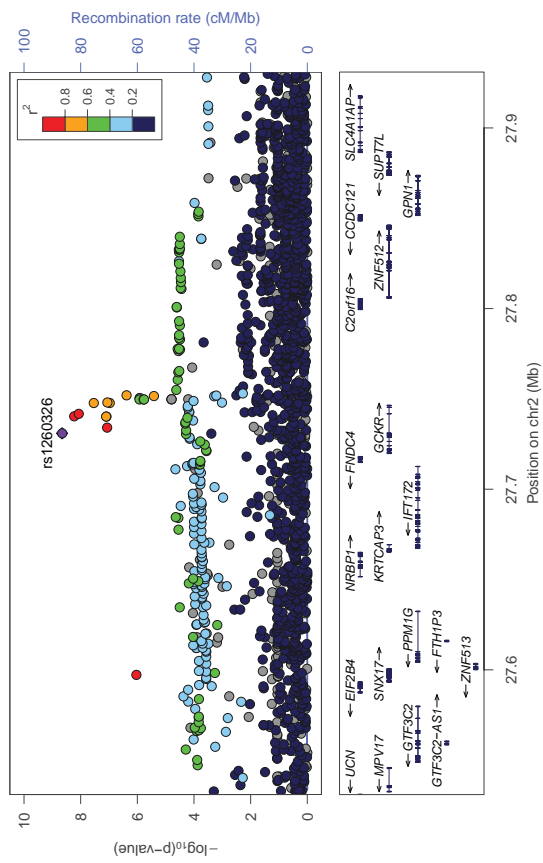
GWAS: rs140000161/PRG2



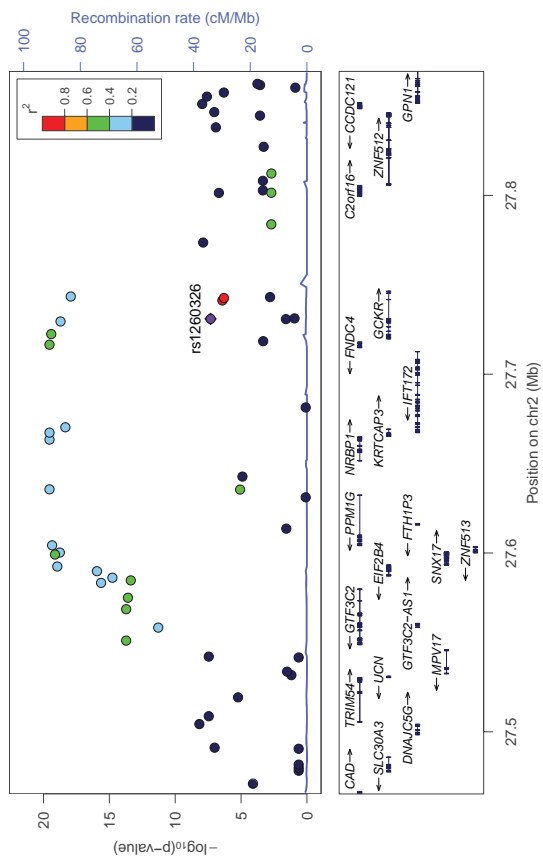
eQTL: rs140000161/PRG2 in Monocytes+IFN



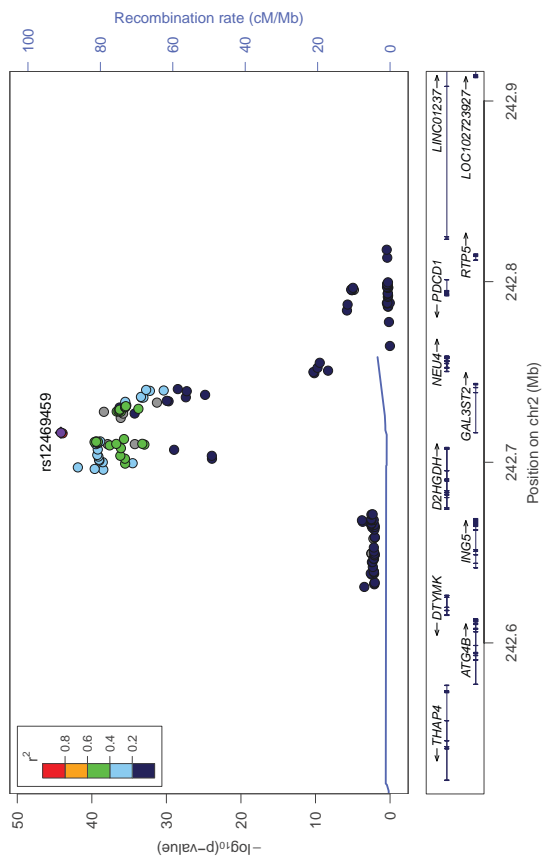
GWAS: rs1260326/KRTCAP3



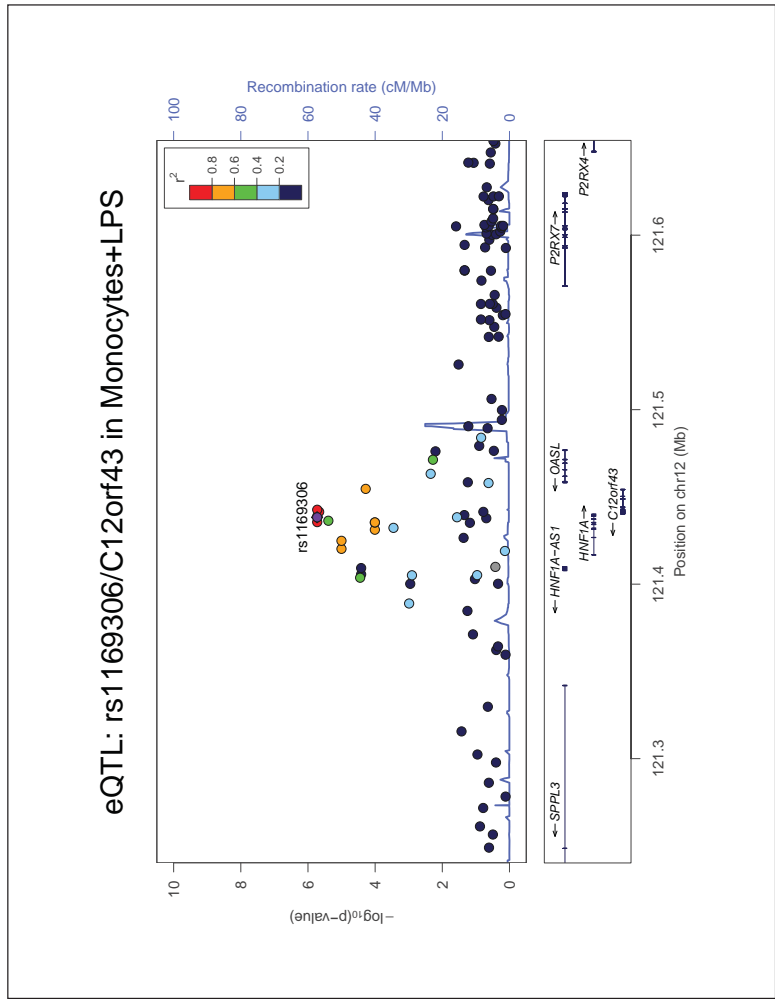
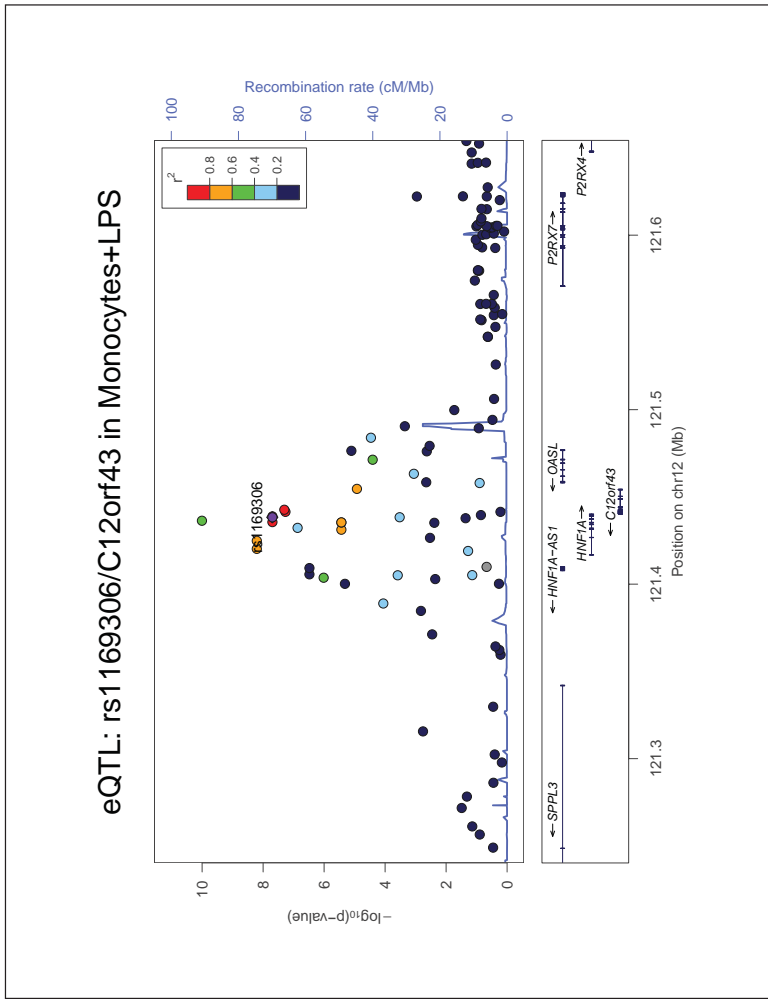
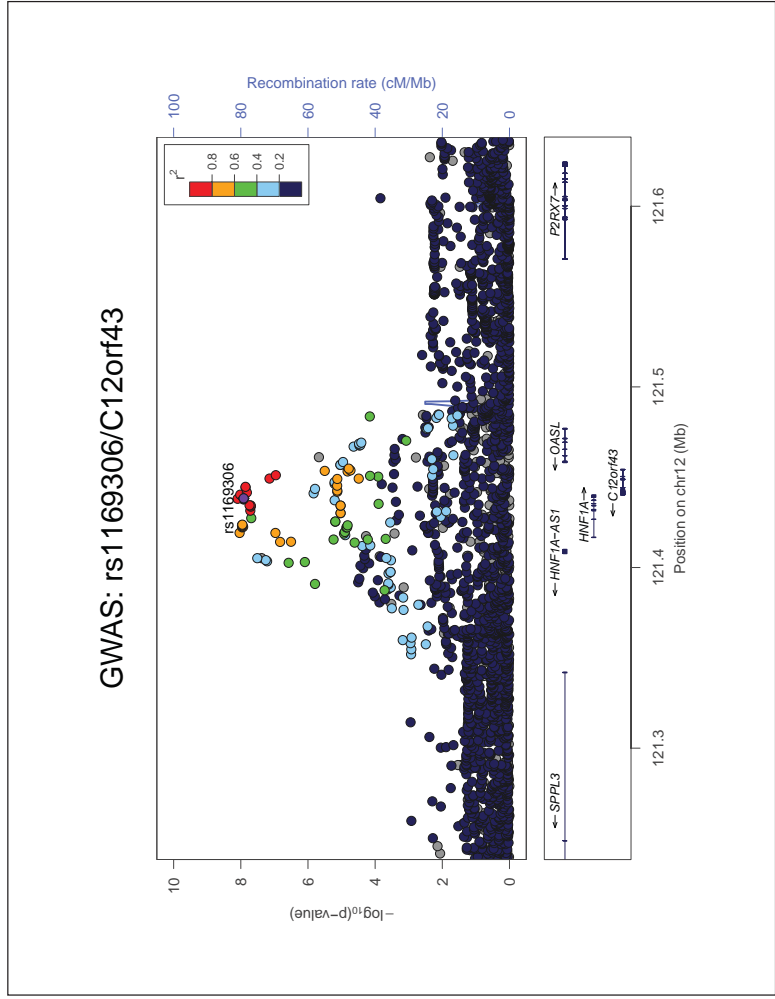
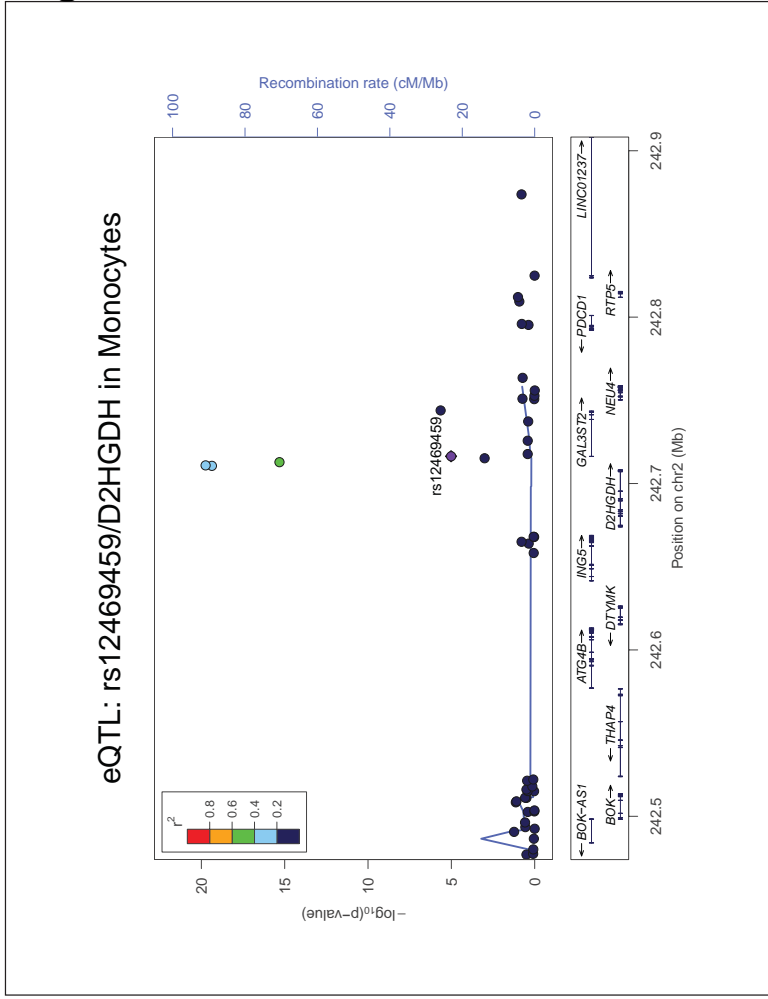
eQTL: rs1260326/KRTCAP3 in B-cells



GWAS: rs12469459/D2HGDH

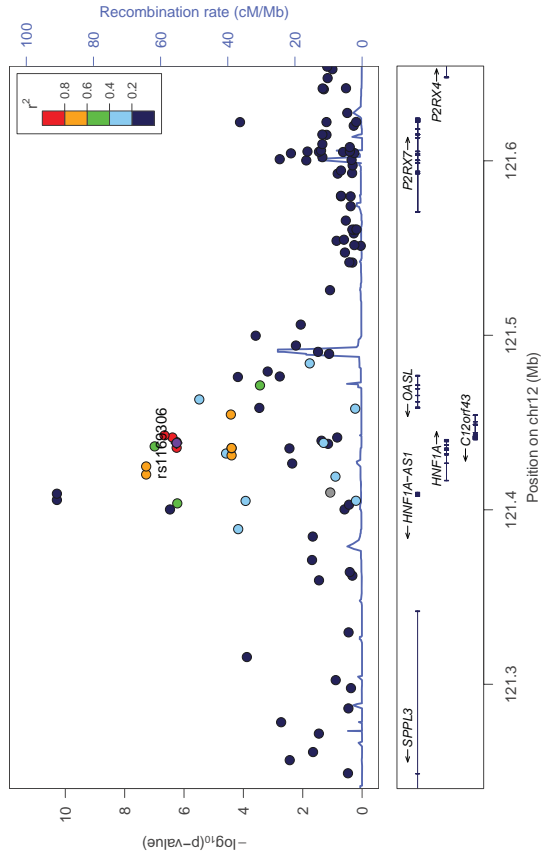


**Figure S2**

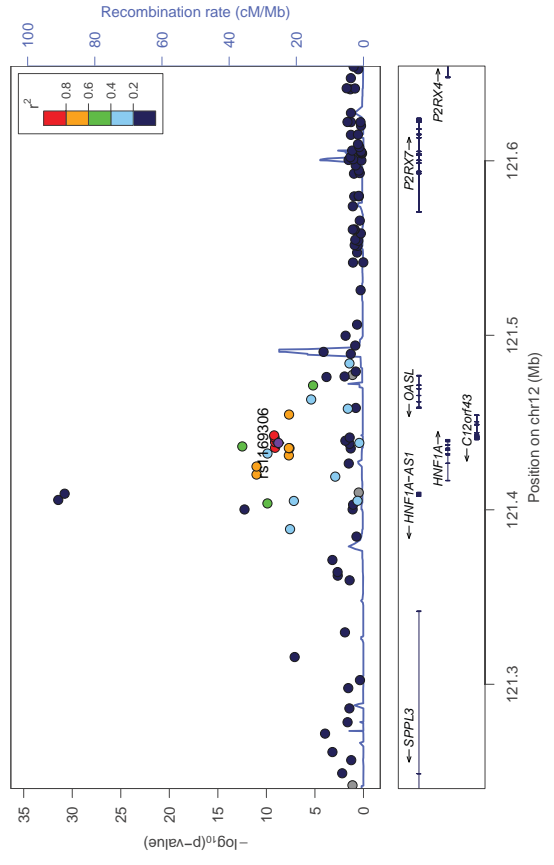


**Figure S2**

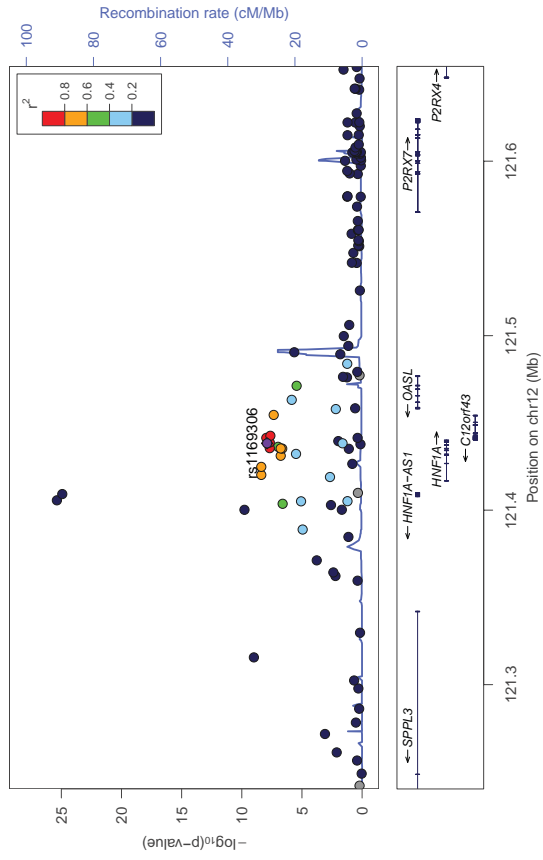
**eQTL: rs1169306/C12orf43 in Monocytes+IFN**



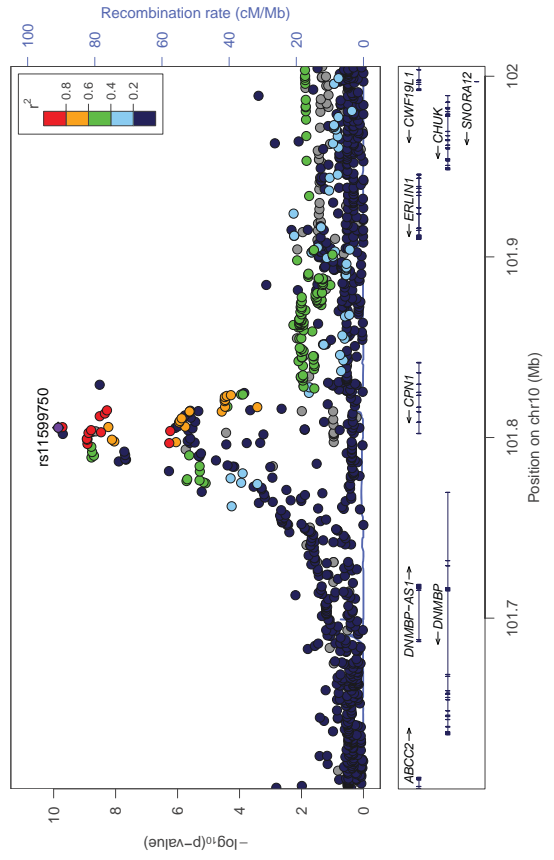
**eQTL: rs1169306/C12orf43 in Monocytes**

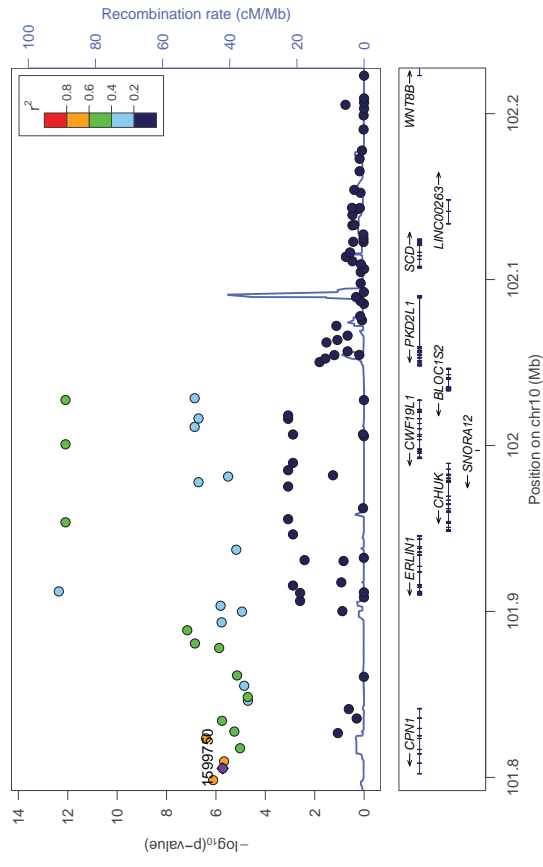
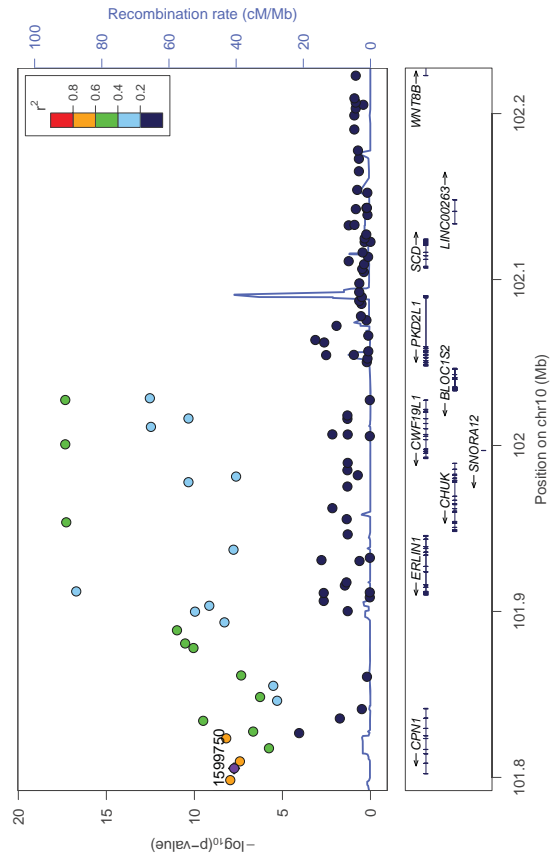
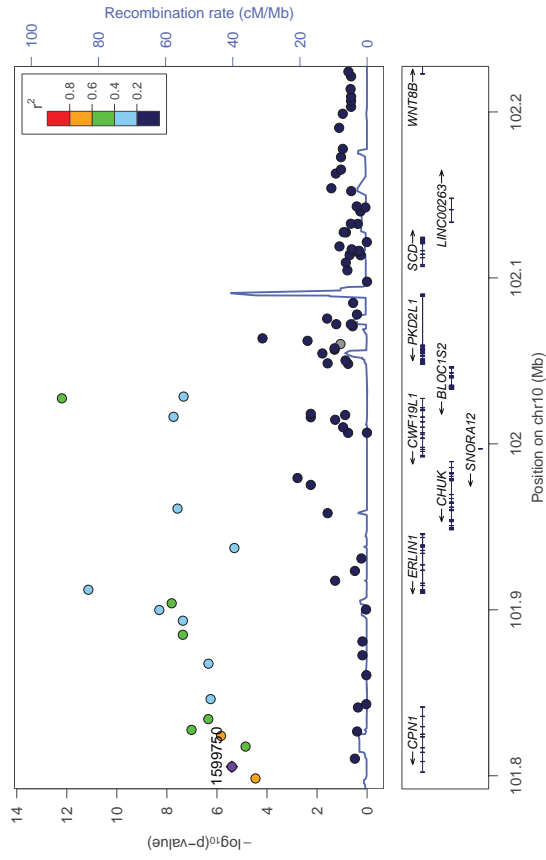
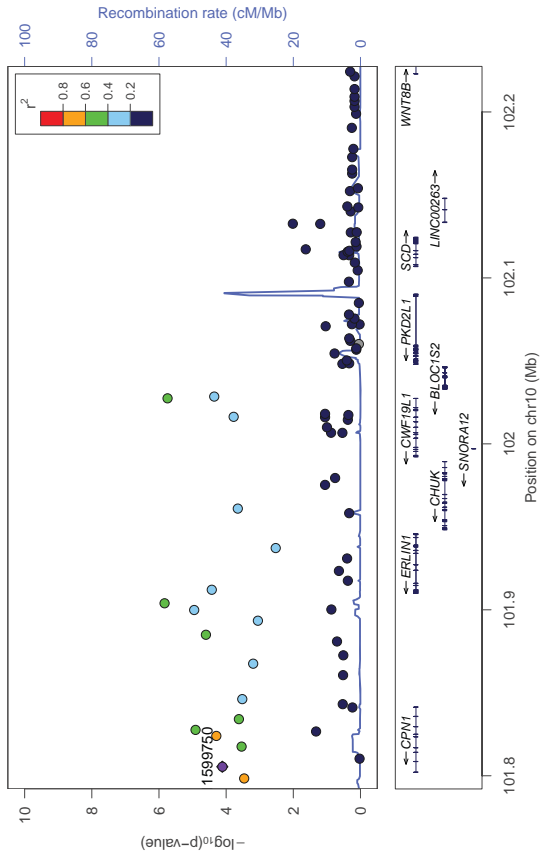


**eQTL: rs1169306/C12orf43 in B-cells**



**GWAS: rs11599750/CWF19L1**



**Figure S2****eQTL: rs11599750/CWF19L1 in Monocytes+LPS****eQTL: rs11599750/CWF19L1 in Monocytes****eQTL: rs11599750/CWF19L1 in Liver****eQTL: rs11599750/CWF19L1 in Heart**



**Table S1**

SNP	Position (chr:MB)	Trait	Dist (kb)	Likely mediator gene	Discovery P	Discovery Beta	A1/A2	A1 freq	INFO	Protein Name	Replication P	Combined P	Directions
rs2153101	chr1:203.2	CHI3L1	-12	CHI3L1	7E-108	-0.62	A/T	0.21	1.05	Chitinase-3-like protein 1	7.01e-136	1.8e-236	---
rs635634	chr9:136.2	SELE	trans	SURF6	1E-219	-0.86	T/C	0.22	0.99	E-selectin	8.1e-133	<1e-256	---
rs1420101	chr2:103	IL1RL1 (ST2)	0	IL1RL1	2E-132	0.58	C/T	0.63	1.03	Interleukin-1 receptor-like 1	5.06e-125	5.86e-254	+++
rs72650832	chr4:74.7	CXCL6	-29	CXCL6	6.2E-42	-0.53	T/C	0.65	0.41	C-X-C motif chemokine 6	8.79e-104	3.12e-130	---
rs635634	chr9:136.2	PECAM1	trans		1.9E-45	-0.41	T/C	0.22	0.99	Platelet endothelial cell adhesion molecule	3.27e-90	6.91e-125	---
rs1569723	chr20:44.7	CD40	-5	CD40	3E-49	-0.39	C/A	0.28	1.00	Tumor necrosis factor receptor superfamily member 5	1.156e-79	1.81e-121	---
rs17368659	chr11:102.7	MMP12	0	MMP12	5.5E-97	0.82	G/T	0.85	0.70	Macrophage metalloelastase	2.15e-76	2.84e-171	+++
rs6555820	chr5:156.5	HAVCR1 (TIM)	0	HAVCR1	1.3E-87	0.46	C/A	0.52	1.03	Hepatitis A virus cellular receptor1	8.443e-60	1.76e-145	+++
rs4129267	chr1:154.4	IL6R (IL6RA)	0	IL6R	2E-265	0.81	T/C	0.36	0.99	Interleukin-6 receptor subunit alpha	1.617e-56	1.53e-297	++
rs7946057	chr11:102.7	MMP3	7	MMP3	1E-108	0.53	A/T	0.53	0.97	Stromelysin-1	1.41e-53	5.21e-160	+?+
rs471994	chr11:102.7	MMP1	-29	MMP1	2.3E-35	0.32	G/A	0.65	0.97	Interstitial collagenase	9.129e-52	2.182e-82	+++
rs62625034	chr3:46.4	CCL4	trans	CCR5	3.1E-41	-0.55	G/T	0.9	0.99	C-C motif chemokine 4	2.178e-44	7.618e-83	---
rs4905	chr19:4.2	IL27 (IL27A)	0	EBI3	1.2E-80	-0.49	G/A	0.3	1.02	Interleukin-27 subunit alpha	1.59e-40	4.52e-119	-?-
rs9323280	chr14:56	LGALS3 (GAL3)	189	LGALS3	5.6E-62	0.77	A/C	0.87	0.60	Galectin-3	4.052e-34	6.6e-94	+++
rs12469459	chr2:242.7	MUC16 (CA125)	trans	GAL3ST2	7.1E-45	0.65	A/T	0.68	0.31	Mucin-16	1.05e-33	3.202e-75	+?+
rs8176741	chr9:136.1	TEK (TIE2)	trans	RALGDS	8.7E-50	-0.61	G/A	0.91	1.02	Angiopoietin-1 receptor	1.769e-30	2.264e-78	---
rs74544699	chr4:74.7	CXCL1	0	CXCL1	1.3E-12	-0.45	A/G	0.95	0.78	Growth-regulated alpha protein	5.48e-25	1.042e-30	-?-

<b>rs3176123</b>	chr20:23	THBD (TM)	0	THBD	2.3E-24	-0.3	T/G	0.81	1.04	Thrombomodulin	1.441e-23	5.828e-46	---
<b>rs982764</b>	chr10:90.8	FAS	0	FAS	2E-12	-0.18	C/T	0.32	1.04	Tumor necrosis factor receptor superfamily member 6	1.06e-21	1.792e-28	-?-
<b>rs79250370</b>	chr9:27.2	TEK (TIE2)	0	TEK	1.9E-13	-0.58	G/A	0.94	0.44	Angiopoietin-1 receptor	5.353e-20	1.141e-30	---
<b>rs2188974</b>	chr17:34.4	CCL3	1	CCL3	4.9E-18	-0.31	A/G	0.81	0.75	C-C motif chemokine 3	9.466e-19	7.919e-35	---
<b>rs6607368</b>	chr17:34.8	CCL4	386	CCL4	6.3E-31	0.51	A/C	0.8	0.46	C-C motif chemokine 4	1.78e-18	2.081e-47	+?+
<b>rs111693235</b>	chr11:1.8	CTSD	4	CTSD	2E-26	0.35	C/G	0.71	0.65	Cathepsin D	2.1e-17	1.071e-41	+?+
<b>rs4810479</b>	chr20:44.5	KITLG (SCF)	trans	MMP9	4.5E-11	-0.18	T/C	0.74	1.01	Kit ligand	8.921e-17	2.802e-25	---
<b>rs62115757</b>	chr19:51.5	KLK11 (HK11)	4	KLK11	1.2E-62	0.77	T/G	0.79	0.39	Kallikrein-11	6.13e-16	6.693e-76	+?+
<b>rs670211</b>	chr16:57.4	CX3CL1	0	CX3CL1	7.4E-12	0.18	G/A	0.41	0.84	Fractalkine	3.654e-15	7.084e-25	+++
<b>rs344560</b>	chr19:6.7	TNFSF14	0	TNFSF14	3E-18	0.75	C/T	0.96	0.46	Tumor necrosis factor ligand superfamily member 14	5.478e-15	1.313e-31	++
<b>rs35285321</b>	chr21:45.2	CSTB	-6	CSTB	1.2E-43	-0.43	A/G	0.38	0.64	Cystatin-B	7.56e-15	7.873e-57	--?
<b>rs75649625</b>	chr11:112.1	IL18	-17	IL18	1.4E-21	0.29	G/A	0.76	0.86	Interleukin-18	3.323e-13	4.094e-33	+++
<b>rs1969539</b>	chr11:14	SPON1	0	SPON1	1.5E-22	-0.25	G/A	0.5	0.89	Spondin-1	5.531e-13	8.121e-34	---
<b>rs693918</b>	chr2:31.9	IL18	trans		2.4E-11	0.19	G/A	0.55	0.72	Interleukin-18	2.905e-12	7.815e-22	+++
<b>rs1260326</b>	chr2:27.7	FST (FS)	trans	GCKR	2E-09	0.14	T/C	0.44	1.07	Follistatin	8.217e-11	1.863e-18	+++
<b>rs4672375</b>	chr2:60.5	GAL	trans		7.1E-11	0.17	G/A	0.58	0.90	Galanin peptides	3.79e-10	8.019e-19	+?+
<b>rs116661163</b>	chr1:204.6	REN	-475	REN	1E-08	-0.72	C/G	0.98	0.38	Renin	4.168e-10	4.455e-17	---
<b>rs142552223</b>	chr3:172.2	TNFSF11 (TRANCE)	trans		3.4E-17	0.81	G/A	0.97	0.58	Tumor necrosis factor ligand superfamily member 11	6.042e-10	1.636e-25	+++
<b>rs13236526</b>	chr7:75.9	HSPB1 (HSP27)	0	HSPB1	1.1E-17	0.36	A/G	0.7	0.44	Heat shock protein beta-1	1.59e-09	2.35e-25	++?
<b>rs3195944</b>	chr19:18.5	GDF15	-20	GDF15	2.2E-08	-0.33	A/G	0.88	0.39	Growth differentiation factor 15	8.05e-09	5.618e-15	-?-
<b>rs56378716</b>	chr17:56.4	MPO	0	MPO	1.9E-09	-0.5	G/A	0.02	0.99	Myeloperoxidase	4.798e-08	5.002e-16	---
<b>rs73062378</b>	chr3:45.8	CCL4	trans		4.5E-13	-0.29	T/C	0.81	0.61	C-C motif chemokine 4	7.16e-08	2.257e-19	-?-
<b>rs140000161</b>	chr11:57.2	PAPPA	trans		1.4E-10	-0.37	A/G	0.91	0.55	Pappalysin-1	9.1e-08	1.27e-16	-?-
<b>rs28601761</b>	chr8:126.5	CHI3L1	trans		5E-09	0.14	C/G	0.61	1.03	Chitinase-3-like protein 1	3.746e-07	9.569e-15	+++

<b>rs8176693</b>	chr9:136.1	THBD (TM)	trans	MED22	1.1E-10	-0.27	C/T	0.91	1.03	Thrombomodulin	4.021e-07	2.427e-16	---
<b>rs7813952</b>	chr8:120.1	TNFSF11 (TRANCE)	trans	TNFRSF11B	2.1E-16	0.22	C/T	0.58	0.81	Tumor necrosis factor ligand superfamily member 11	4.092e-07	1.284e-21	+++
<b>rs200373</b>	chr19:18.3	CTSL1	trans		4.3E-09	-0.14	T/A	0.48	1.03	Cathepsin L1	5.46e-07	2.116e-14	-?-
<b>rs6993770</b>	chr8:106.6	DKK1	trans		1.6E-09	-0.17	T/A	0.27	0.99	Dickkopf-related protein 1	8.545e-06	7.278e-14	---
<b>rs2271025</b>	chr16:67	AGRP	530+		2.3E-09	-0.36	G/A	0.92	0.55	Agouti-related protein	1.866e-05	2.346e-13	---
<b>rs35186877</b>	chr17:4.6	CXCL16	11	CXCL16	1.7E-09	0.21	G/A	0.81	0.76	C-X-C motif chemokine 16	3.07e-05	2.54e-13	+?+
<b>rs35166255</b>	chr11:126.3	IL1RL1 (ST2)	trans		1.2E-09	-0.46	G/A	0.96	0.68	Interleukin-1 receptor-like 1	3.69e-05	2.02e-13	-?-
<b>rs492602</b>	chr19:49.2	MMP10	trans	FUT2, RASIP1	7.8E-09	0.14	G/A	0.45	1.03	Stromelysin-2	5.71e-05	2.364e-12	+++
<b>rs11667946</b>	chr19:51.5	KLK6	-1	KLK6	3.4E-15	0.3	C/T	0.53	0.41	Kallikrein-6	0.000116	2.909e-18	+?+
<b>rs7928577</b>	chr11:126.2	LGALS3 (GAL3)	trans	TIRAP	2.1E-09	0.26	T/G	0.08	0.98	Galectin-3	0.0001312	1.834e-12	+++
<b>rs495828</b>	chr9:136.2	F3 (TF)	trans	SURF6	4.6E-10	-0.17	T/G	0.25	0.99	Tissue factor	0.0001693	6.249e-13	---
<b>rs17610659</b>	chr1:110.5	CSF1	30	CSF1	6.5E-10	0.15	T/C	0.48	0.97	Macrophage colony-stimulating factor 1	0.000228	1.202e-12	+++
<b>rs11599750</b>	chr10:101.8	IL27 (IL27A)	trans	CWF19L1	1.4E-10	0.16	C/T	0.62	1.00	Interleukin-27 subunit alpha	0.000429	2.816e-13	+?+
<b>rs6469811</b>	chr8:120.1	TNFRSF11B (OPG)	-137	TNFRSF11B	2.9E-11	-0.18	G/A	0.55	0.81	Osteoprotegerin	0.0005594	2.52e-13	---
<b>rs76769120</b>	chr1:12.2	TNFRSF1B (TRAIL/TNFR2)	0	TNFRSF1B	1.3E-11	0.42	G/T	0.91	0.46	Tumor necrosis factor receptor superfamily member 1B	0.000869	6.649e-14	+?+
<b>rs11150189</b>	chr16:79.7	XPNPEP2 (MAMP)	trans		6.9E-14	0.2	A/G	0.69	0.97	Xaa-Pro aminopeptidase 2	0.00104	3.739e-16	++?
<b>rs2070600</b>	chr6:32.2	AGER (RAGE)	0	AGER	3E-10	-0.37	T/C	0.04		Advanced glycosylation end product-specific receptor	0.001181	4.749e-12	+-
<b>rs241771</b>	chr17:26.6	TNFRSF11B (OPG)	trans		6E-10	-0.15	T/C	0.45	0.98	Osteoprotegerin	0.003457	3.469e-11	---
<b>rs1169306</b>	chr12:121.4	LGALS3 (GAL3)	trans	HNF1A, C12orf43	6.5E-09	0.14	T/C	0.38	1.02	Galectin-3	0.003868	2.847e-10	++
<b>rs16873402</b>	chr8:106.6	PDGFB	trans		2.4E-08	0.15	C/T	0.71	0.94	Platelet-derived growth factor subunit B	0.005719	1.407e-09	+++
<b>rs7599125</b>	chr2:32.9	IL18	trans	NLRC4	1.1E-08	-0.23	G/A	0.44	0.35	Interleukin-18	0.006416	8.746e-10	---
<b>rs33988101</b>	chr19:49.2	LGALS3 (GAL3)	trans	FUT2, RASIP1	3.5E-09	0.14	T/G	0.48	1.04	Galectin-3	0.009141	5.557e-10	+++

<b>rs880949</b>	chr14:75.4	PGF (PLGF)	6	PGF	1.6E-08	-0.17	G/A	0.58	0.68	Placenta growth factor	0.01379	3.281e-09	---
<b>rs1580006</b>	chr11:10.4	ADM (AM)	42	ADM	2E-15	-0.19	A/T	0.54	1.00	Pro-adrenomedullin	0.02987	7.124e-14	---
<b>rs10947260</b>	chr6:32.4	IL6	trans	NOTCH4, AGER, ATF6B	1.8E-10	0.25	C/T	0.1		Interleukin-6	0.1524	6.21e-09	+++
<b>rs35538083</b>	chr1:27.1	XPNPEP2 (MAMP)	trans	PIGV	3.1E-08	-0.27	T/C	0.93	0.95	Xaa-Pro aminopeptidase 2	0.181	3.717e-08	--?
<b>rs75416436</b>	chr13:42.6	NGF (BETANGF)	trans		4.2E-08	-0.5	G/A	0.96	0.44	Beta-nerve growth factor	0.286	9.912e-08	--?
<b>rs12570111</b>	chr10:12.3	MMP1	trans		4.7E-08	0.14	T/C	0.56	0.94	Interstitial collagenase	0.616	1.4e-05	+?-
<b>rs76519098</b>	chr10:49.9	GDF15	trans	MAPK8	1.1E-10	-0.83	C/T	0.98	0.39	Growth differentiation factor 15	0.6989	2.571e-06	--+
<b>rs117538444</b>	chr15:89.9	PGF (PLGF)	trans		6.6E-09	0.78	C/T	0.98	0.37	Placenta growth factor	0.9649	9.685e-06	++
<b>rs549596</b>	chr1:11.9	BNP	1	NPPB	1.7E-14	-0.19	T/C	0.59	0.97	Binatriuretic peptides		1.738e-14	-??
<b>rs35207557</b>	chr1:11.9	NPPB (NTPROBNP)	0	NPPB	2.6E-25	-0.26	T/TA	0.6	0.98	Natriuretic peptides B		2.57e-25	-??
<b>rs184243355</b>	chr5:153.2	CCL3	trans		2.2E-08	-0.41	T/C	0.94	0.47	C-C motif chemokine 3		2.239e-08	-??
<b>rs61598054</b>	chr6:108.8	NGF (BETANGF)	trans	FOXO3	3.8E-08	-0.31	C/T	0.87	0.42	Beta-nerve growth factor		3.802e-08	-??
<b>rs6557662</b>	chr8:23.2	NPPB (NTPROBNP)	trans		1.5E-08	0.23	A/G	0.73	0.44	Natriuretic peptides B		1.479e-08	+??
<b>rs139879640</b>	chr15:81.6	IL16	0	IL16	3E-62	0.86	TCTCA/	0.94	0.96	Pro-interleukin-16 [Cleaved into: Interleukin-16		2.951e-62	+??
<b>rs200433550</b>	chr19:49.2	F3 (TF)	trans		5.6E-10	0.16	TA/T	0.58	0.91	Tissue factor		5.623e-10	+??
<b>rs2050011</b>	chrX:128.9	XPNPEP2 (MAMP)	-1	XPNPEP2	2.4E-68	-0.36	T/G	0.33	1.05	Xaa-Pro aminopeptidase 2		2.399e-68	-??

Overview of all associations between plasma protein and SNPs significant at genome-wide level. Trait – the plasma protein target; Dist (kb) – if cis, the distance between SNP and protein encoding gene; Likely mediator gene – most likely cis-mediator gene, protein-encoding gene in cis-cases, based on table 2 in trans cases; Discovery P – the pQTL association P-value from the Olink-Improve discovery cohort (n=3,394); Discovery Beta – the Olink-Improve effect size; A1/A2 – encoded allele and alternative allele; A1 freq – frequency of encoded allele; INFO – imputation quality score (MACH 1.0). Protein name; Replication P - the pQTL association P-value from the replication cohorts (n=976, n=933, n=730); Combined P – the meta-analysis P-value of both discovery and replication; Directions - for replication meta-analysis are indicated as IMPROVE (discovery), NSPHS (replication), ULSAM-PIVUS (merged replication). †while 530.7 kb is formally outside of the pre-defined cis-limit of 500 kb, the AGRP association was classified as cis-acting; all other pQTL associations were either acting across chromosomes or at distances more than 100 MB. SE: standard error.

**Table S2**

Other trait	Other SNP	r <sup>2</sup> (EUR 1000G)	Other P-value	Pubmed ID	Olink SNP	Olink Trait Protein	Olink P-value
DBP	rs6668659	0.67	5.1E-10	24560520	rs549596	BNP	1.75E-14
Mean Arterial Pressure	rs6668659	0.67	1.2E-09	24560520	rs549596	BNP	1.75E-14
SBP	rs6668659	0.67	4.5E-09	24560520	rs549596	BNP	1.75E-14
HDL cholesterol	rs12748152	0.92	1.0E-15	24097068	rs35538083	XPNPEP2	3.11E-08
LDL cholesterol	rs12748152	0.92	3.0E-12	24097068	rs35538083	XPNPEP2	3.11E-08
Triglycerides	rs12748152	0.92	1.0E-09	24097068	rs35538083	XPNPEP2	3.11E-08
Interleukin-6_receptor CSF (gene=IL6R)	rs4129267	1.00	2.7E-62	25340798	rs4129267	IL6R	2.14E-265
IL6RA plasma (gene=IL6R)	rs4129267	1.00	4.4E-58	25147954	rs4129267	IL6R	2.14E-265
Protein quantitative trait loci (sIL-6R)	rs4129267	1.00	2.0E-57	18464913	rs4129267	IL6R	2.14E-265
C-reactive protein	rs4129267	1.00	2.0E-48	21300955	rs4129267	IL6R	2.14E-265
Fibrinogen plasma	rs61812598	1.00	2.7E-36	26561523	rs4129267	IL6R	2.14E-265
Fibrinogen (EA)	rs4129267	1.00	6.0E-27	23969696	rs4129267	IL6R	2.14E-265
Ankylosing spondylitis	rs4129267	1.00	3.4E-13	23749187	rs4129267	IL6R	2.14E-265
Asthma	rs4129267	1.00	2.0E-08	21907864	rs4129267	IL6R	2.14E-265
Fibrinogen	rs2228145	0.99	2.0E-11	20031577	rs4129267	IL6R	2.14E-265
Rheumatoid arthritis	rs2228145	0.99	4.0E-09	24390342	rs4129267	IL6R	2.14E-265
YKL-40 levels	rs4950928	0.90	1.0E-13	18403759	rs2153101	CHI3L1	7.50E-108
Triglycerides	rs1260326	1.00	2.0E-239	24097068	rs1260326	FST	2.02E-09
Blood metabolite ratios (glucose/mannose)	rs1260326	1.00	3.0E-148	24816252	rs1260326	FST	2.02E-09
Blood metabolite levels (mannose)	rs1260326	1.00	1.0E-77	24816252	rs1260326	FST	2.02E-09
Urate levels	rs1260326	1.00	1.0E-44	23263486	rs1260326	FST	2.02E-09
Tryglycerides transethnic	rs1260326	1.00	1.6E-42	23555291	rs1260326	FST	2.02E-09
Cholesterol, total	rs1260326	1.00	3.0E-42	24097068	rs1260326	FST	2.02E-09
Fasting glucose- BMI-adjusted	rs1260326	1.00	1.0E-40	22885924	rs1260326	FST	2.02E-09
Fasting glucose	rs1260326	1.00	1.0E-40	22885924	rs1260326	FST	2.02E-09
C-reactive protein	rs1260326	1.00	5.0E-40	21300955	rs1260326	FST	2.02E-09
Lipid metabolism phenotypes (TG.assay, whole)	rs1260326	1.00	1.0E-37	19936222	rs1260326	FST	2.02E-09

Lipid metabolism phenotypes (TG.by.NMR, whole)	rs1260326	1.00	3.0E-35	19936222	rs1260326	FST	2.02E-09
Lipid metabolism phenotypes (TG.assay, fasting)	rs1260326	1.00	4.0E-32	19936222	rs1260326	FST	2.02E-09
Lipid metabolism phenotypes (TG.by.NMR, fasting)	rs1260326	1.00	3.0E-29	19936222	rs1260326	FST	2.02E-09
Lipid metabolism phenotypes (VLDL.large, whole)	rs1260326	1.00	3.0E-28	19936222	rs1260326	FST	2.02E-09
Lipid metabolism phenotypes (VLDL.large, fasting)	rs1260326	1.00	4.0E-24	19936222	rs1260326	FST	2.02E-09
Fasting Insulin- BMI adjusted	rs1260326	1.00	2.7E-22	22885924	rs1260326	FST	2.02E-09
Serum albumin level	rs1260326	1.00	4.0E-19	23022100	rs1260326	FST	2.02E-09
Metabolite levels (Ala, Gln)	rs1260326	1.00	3.0E-18	22286219	rs1260326	FST	2.02E-09
2 hr glucose -FG and BMI adjusted	rs1260326	1.00	9.0E-15	22885924	rs1260326	FST	2.02E-09
Chronic kidney disease (eGFRcrea)	rs1260326	1.00	3.0E-14	20383146	rs1260326	FST	2.02E-09
Serum albumin level (EA)	rs1260326	1.00	3.0E-14	23022100	rs1260326	FST	2.02E-09
Fasting Insulin	rs1260326	1.00	3.8E-14	22885924	rs1260326	FST	2.02E-09
Blood metabolite levels (alanine)	rs1260326	1.00	6.0E-14	24816252	rs1260326	FST	2.02E-09
Hypertriglyceridemia	rs1260326	1.00	2.0E-13	23505323	rs1260326	FST	2.02E-09
Liver enzyme levels (gamma-glutamyl transferase)	rs1260326	1.00	4.0E-13	22001757	rs1260326	FST	2.02E-09
Glycemic traits (pregnancy) (FPG)	rs1260326	1.00	6.0E-13	23903356	rs1260326	FST	2.02E-09
Metabolite levels	rs1260326	1.00	1.0E-12	22916037	rs1260326	FST	2.02E-09
Alanine	rs1260326	1.00	7.6E-12	23823483	rs1260326	FST	2.02E-09
PC aa C40:5	rs1260326	1.00	1.3E-11	26068415	rs1260326	FST	2.02E-09
Lactate	rs1260326	1.00	3.3E-11	23823483	rs1260326	FST	2.02E-09
Glycemic traits (pregnancy) (FCP)	rs1260326	1.00	6.0E-11	23903356	rs1260326	FST	2.02E-09
Two-hour glucose challenge	rs1260326	1.00	3.0E-10	20081857	rs1260326	FST	2.02E-09
Metabolic traits (TG)	rs1260326	1.00	4.0E-10	19060910	rs1260326	FST	2.02E-09
Platelet counts	rs1260326	1.00	9.0E-10	22139419	rs1260326	FST	2.02E-09
$\alpha$ -hydroxybutyrate	rs1260326	1.00	1.3E-09	23823483	rs1260326	FST	2.02E-09
Non-albumin protein levels (ALB)	rs1260326	1.00	3.0E-09	22558069	rs1260326	FST	2.02E-09
TAG 50:4	rs1260326	1.00	3.4E-09	23823483	rs1260326	FST	2.02E-09

Hematological and biochemical traits (ALB)	rs1260326	1.00	4.0E-09	20139978	rs1260326	FST	2.02E-09
Cardiovascular disease risk factors (TRIG)	rs1260326	1.00	2.0E-08	21943158	rs1260326	FST	2.02E-09
Serum albumin level (Japanese)	rs1260326	1.00	2.0E-08	23022100	rs1260326	FST	2.02E-09
TAG 48:2	rs1260326	1.00	2.5E-08	23823483	rs1260326	FST	2.02E-09
TAG 50:3	rs1260326	1.00	2.6E-08	23823483	rs1260326	FST	2.02E-09
PC 34:3	rs1260326	1.00	2.9E-08	23823483	rs1260326	FST	2.02E-09
PC 32:2	rs1260326	1.00	3.9E-08	23823483	rs1260326	FST	2.02E-09
Waist circumference and related phenotypes (triglycerides)	rs1260326	1.00	4.0E-08	18454146	rs1260326	FST	2.02E-09
TAG 48:3	rs1260326	1.00	4.9E-08	23823483	rs1260326	FST	2.02E-09
Urate levels (Urate)	rs780093	0.91	4.0E-17	20884846	rs1260326	FST	2.02E-09
Sex hormone-binding globulin levels (Men + Women)	rs780093	0.91	2.0E-16	22829776	rs1260326	FST	2.02E-09
Waist Circumference - Triglycerides (WC-TG)	rs780093	0.91	2.0E-12	21386085	rs1260326	FST	2.02E-09
Crohn's disease	rs780093	0.91	5.0E-11	21102463	rs1260326	FST	2.02E-09
Sex hormone-binding globulin levels (Women)	rs780093	0.91	9.0E-11	22829776	rs1260326	FST	2.02E-09
Triglycerides-Blood Pressure (TG-BP)	rs780093	0.91	3.0E-10	21386085	rs1260326	FST	2.02E-09
Palmitoleic acid (16:1n-7) plasma levels	rs780093	0.91	1.0E-09	23362303	rs1260326	FST	2.02E-09
Metabolic traits (glucose/mannose + 54 other traits)	rs780094	0.90	6.0E-53	21886157	rs1260326	FST	2.02E-09
Fasting glucose-related traits (FPG)	rs780094	0.90	6.0E-38	20081858	rs1260326	FST	2.02E-09
Fasting glucose-related traits (HOMA-IR)	rs780094	0.90	3.0E-24	20081858	rs1260326	FST	2.02E-09
Fasting insulin-related traits (HOMA-IR)	rs780094	0.90	3.0E-24	20081858	rs1260326	FST	2.02E-09
Fasting glucose-related traits (interaction with BMI)	rs780094	0.90	4.0E-24	22581228	rs1260326	FST	2.02E-09
Fasting insulin-related traits (FI)	rs780094	0.90	4.0E-20	20081858	rs1260326	FST	2.02E-09
Fasting glucose-related traits (FI)	rs780094	0.90	4.0E-20	20081858	rs1260326	FST	2.02E-09
Metabolic syndrome (TG)	rs780094	0.90	6.0E-20	22399527	rs1260326	FST	2.02E-09
height	rs780094	0.90	7.5E-12	25282103	rs1260326	FST	2.02E-09
Calcium levels	rs780094	0.90	1.0E-10	24068962	rs1260326	FST	2.02E-09
Fasting insulin-related traits (interaction with BMI)	rs780094	0.90	3.0E-10	22581228	rs1260326	FST	2.02E-09

Uric acid levels	rs780094	0.90	1.0E-09	19503597	rs1260326	FST	2.02E-09
Fasting plasma glucose (East Asian)	rs780094	0.90	3.6E-09	25187374	rs1260326	FST	2.02E-09
Triglycerides (Hispanic)	rs780094	0.90	7.0E-09	23726366	rs1260326	FST	2.02E-09
Phospholipid levels (plasma) (DPA)	rs780094	0.90	9.0E-09	21829377	rs1260326	FST	2.02E-09
Triglycerides (AA)	rs4665972	0.87	1.0E-08	23726366	rs1260326	FST	2.02E-09
Inflammatory biomarkers (IL18)	rs7577696	0.77	3.0E-19	24182552	rs693918	IL18	2.38E-11
Eosinophil counts (EA)	rs1420101	1.00	5.0E-14	19198610	rs1420101	IL1RL1	2.03E-132
Serum protein levels (sST2)	rs950880	0.97	7.0E-94	23999434	rs1420101	IL1RL1	2.03E-132
Pulmonary function (interaction) (FEV1/FVC, Pack-years)	rs2070600	1.00	1.0E-21	23284291	rs2070600	AGER	3.01E-10
Pulmonary function (FEV1/FVC)	rs2070600	1.00	3.0E-14	20010835	rs2070600	AGER	3.01E-10
Crohn's disease	rs10947261	0.97	3.0E-12	23850713	rs10947260	IL6	1.83E-10
Knee osteoarthritis	rs10947262	0.97	5.0E-09	20305777	rs10947260	IL6	1.83E-10
Vascular endothelial growth factor levels	rs6993770	1.00	5.0E-23	21757650	rs6993770	DKK1	1.61E-09
Platelet counts	rs6993770	1.00	4.0E-17	22139419	rs6993770	DKK1	1.61E-09
Vascular endothelial growth factor levels	rs6993770	0.80	5.0E-23	21757650	rs16873402	PDGFB	2.42E-08
Platelet counts	rs6993770	0.80	4.0E-17	22139419	rs16873402	PDGFB	2.42E-08
Triglycerides	rs2954029	0.74	1.0E-107	24097068	rs28601761	CHI3L1	5.06E-09
Cholesterol, total	rs2954029	0.74	2.0E-65	24097068	rs28601761	CHI3L1	5.06E-09
LDL cholesterol	rs2954029	0.74	2.0E-50	24097068	rs28601761	CHI3L1	5.06E-09
HDL cholesterol	rs2954029	0.74	3.0E-29	24097068	rs28601761	CHI3L1	5.06E-09
CAD	rs2954029	0.74	4.8E-09	23202125	rs28601761	CHI3L1	5.06E-09
Lipid metabolism phenotypes (APOB.assay, whole)	rs6982636	0.71	7.0E-12	19936222	rs28601761	CHI3L1	5.06E-09
Lipid metabolism phenotypes (TG.assay, whole)	rs6982636	0.71	1.0E-09	19936222	rs28601761	CHI3L1	5.06E-09
Liver enzyme levels (alkaline phosphatase)	rs2954021	0.64	2.0E-13	22001757	rs28601761	CHI3L1	5.06E-09
Liver enzyme levels (alanine transaminase)	rs2954021	0.64	5.0E-09	22001757	rs28601761	CHI3L1	5.06E-09
Elevated serum carcinoembryonic antigen levels	rs8176741	1.00	2.0E-24	24941225	rs8176741	TEK	8.63E-50
Tumor biomarkers (CEA)	rs8176749	0.98	7.0E-105	23300138	rs8176741	TEK	8.63E-50
sTie-2 plasma (gene=TEK)	rs8176693	0.98	1.8E-33	25552591	rs8176741	TEK	8.63E-50
vWF	rs8176693	0.98	1.6E-17	23381943	rs8176741	TEK	8.63E-50



End-stage coagulation (vWF)	rs8176743	0.98	2.0E-17	23381943	rs8176741	TEK	8.63E-50
FVIII Ag	rs8176693	0.98	1.2E-14	23381943	rs8176741	TEK	8.63E-50
Urinary metabolites (H-NMR features) (5.2625, Unknown)	rs8176749	0.98	4.0E-12	24586186	rs8176741	TEK	8.63E-50
Ang-2 plasma (gene=MCPH1)	rs8176746	0.98	2.1E-08	25552591	rs8176741	TEK	8.63E-50
Mean corpuscular hemoglobin concentration	rs8176746	0.98	4.0E-08	20139978	rs8176741	TEK	8.63E-50
Malaria	rs8176722	0.70	9.0E-10	23717212	rs8176741	TEK	8.63E-50
Tumor biomarkers (CEA)	rs8176749	1.00	7.0E-105	23300138	rs8176693	THBD	1.12E-10
sTie-2 plasma (gene=TEK)	rs8176693	1.00	1.8E-33	25552591	rs8176693	THBD	1.12E-10
vWF	rs8176693	1.00	1.6E-17	23381943	rs8176693	THBD	1.12E-10
End-stage coagulation (vWF)	rs8176743	1.00	2.0E-17	23381943	rs8176693	THBD	1.12E-10
FVIII Ag	rs8176693	1.00	1.2E-14	23381943	rs8176693	THBD	1.12E-10
Urinary metabolites (H-NMR features) (5.2625, Unknown)	rs8176749	1.00	4.0E-12	24586186	rs8176693	THBD	1.12E-10
Ang-2 plasma (gene=MCPH1)	rs8176746	1.00	2.1E-08	25552591	rs8176693	THBD	1.12E-10
Mean corpuscular hemoglobin concentration	rs8176746	1.00	4.0E-08	20139978	rs8176693	THBD	1.12E-10
Elevated serum carcinoembryonic antigen levels	rs8176741	0.98	2.0E-24	24941225	rs8176693	THBD	1.12E-10
Malaria	rs8176722	0.71	9.0E-10	23717212	rs8176693	THBD	1.12E-10
Liver enzyme levels (alkaline phosphatase)	rs579459	1.00	3.0E-123	22001757	rs495828	F3	4.54E-10
E-selectin plasma (gene=SELE)	rs651007	1.00	1.9E-103	23300549	rs495828	F3	4.54E-10
E-selectin levels	rs651007	1.00	2.0E-82	20147318	rs495828	F3	4.54E-10
Hematological and biochemical traits (ALP)	rs495828	1.00	4.0E-59	20139978	rs495828	F3	4.54E-10
Serum alkaline phosphatase levels	rs651007	1.00	1.0E-56	24094242	rs495828	F3	4.54E-10
Soluble levels of adhesion molecules (P-Selectin)	rs579459	1.00	2.0E-41	20167578	rs495828	F3	4.54E-10
Blood metabolite ratios (ADSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs649129	1.00	9.0E-37	24816252	rs495828	F3	4.54E-10
Blood metabolite ratios (DSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs495828	1.00	6.0E-34	24816252	rs495828	F3	4.54E-10
Urinary metabolites (H-NMR features) (5.1825, Unknown)	rs579459	1.00	2.0E-32	24586186	rs495828	F3	4.54E-10

Soluble E-selectin levels	rs579459	1.00	1.0E-29	19729612	rs495828	F3	4.54E-10
Blood metabolite ratios (ADpSGEGDFXAEGGGVR/X-14304-- leucylalanine)	rs579459	1.00	1.0E-28	24816252	rs495828	F3	4.54E-10
Urinary metabolites (H-NMR features) (2.0525, Unknown)	rs579459	1.00	1.0E-28	24586186	rs495828	F3	4.54E-10
End-stage coagulation (FVIII)	rs651007	1.00	2.0E-25	23381943	rs495828	F3	4.54E-10
Blood metabolite levels (ADpSGEGDFXAEGGGVR)	rs651007	1.00	6.0E-20	24816252	rs495828	F3	4.54E-10
Red blood cell traits (EA, RBCC)	rs579459	1.00	9.0E-18	23222517	rs495828	F3	4.54E-10
Venous thromboembolism	rs495828	1.00	3.0E-16	22672568	rs495828	F3	4.54E-10
Soluble levels of adhesion molecules (ICAM)	rs649129	1.00	1.0E-15	20167578	rs495828	F3	4.54E-10
Coronary heart disease	rs579459	1.00	4.0E-14	21378990	rs495828	F3	4.54E-10
vWF	rs495828	1.00	9.4E-13	23381943	rs495828	F3	4.54E-10
Red blood cell count	rs495828	1.00	3.0E-12	20139978	rs495828	F3	4.54E-10
Hematological and biochemical traits (Hb)	rs495828	1.00	1.0E-11	20139978	rs495828	F3	4.54E-10
Hematological and biochemical traits (Ht)	rs495828	1.00	6.0E-10	20139978	rs495828	F3	4.54E-10
Coronary artery disease or ischemic stroke	rs579459	1.00	2.0E-09	24262325	rs495828	F3	4.54E-10
Metabolite levels (LDL)	rs651007	1.00	6.0E-09	21909109	rs495828	F3	4.54E-10
Ferritin (log)   serum   (gene=FTL   FTH1)	rs651007	1.00	1.3E-08	25352340	rs495828	F3	4.54E-10
CAD	rs579459	1.00	2.7E-08	23202125	rs495828	F3	4.54E-10
Coronary artery disease or large artery stroke	rs579459	1.00	3.0E-08	24262325	rs495828	F3	4.54E-10
Angiotensin-converting enzyme activity	rs495828	1.00	3.0E-08	20066004	rs495828	F3	4.54E-10
FVIII Ag	rs495828	1.00	3.7E-08	23381943	rs495828	F3	4.54E-10
LDL cholesterol	rs635634	0.83	2.0E-41	24097068	rs495828	F3	4.54E-10
Cholesterol, total	rs635634	0.83	3.0E-35	24097068	rs495828	F3	4.54E-10
invasive epithelial ovarian cancer	rs635634	0.83	4.4E-09	25581431	rs495828	F3	4.54E-10
Soluble ICAM-1	rs507666	0.83	3.0E-91	21533024	rs495828	F3	4.54E-10
Lipid traits (LDL)	rs507666	0.83	2.0E-11	24386095	rs495828	F3	4.54E-10
Lipid traits (TC)	rs507666	0.83	4.0E-11	24386095	rs495828	F3	4.54E-10
LDL-C transethnic	rs2519093	0.82	2.2E-13	23555291	rs495828	F3	4.54E-10

LDL cholesterol	rs635634	1.00	2.0E-41	24097068	rs635634	PECAM1	1.89E-45
Cholesterol, total	rs635634	1.00	3.0E-35	24097068	rs635634	PECAM1	1.89E-45
E-selectin   plasma   (gene=SELE)	rs635634	1.00	5.0E-16	25147954	rs635634	PECAM1	1.89E-45
invasive epithelial ovarian cancer	rs635634	1.00	4.4E-09	25581431	rs635634	PECAM1	1.89E-45
Soluble ICAM-1	rs507666	0.99	3.0E-91	21533024	rs635634	PECAM1	1.89E-45
Lipid traits (LDL)	rs507666	0.99	2.0E-11	24386095	rs635634	PECAM1	1.89E-45
Lipid traits (TC)	rs507666	0.99	4.0E-11	24386095	rs635634	PECAM1	1.89E-45
Venous thromboembolism	rs2519093	0.98	8.0E-16	22672568	rs635634	PECAM1	1.89E-45
LDL-C transethnic	rs2519093	0.98	2.2E-13	23555291	rs635634	PECAM1	1.89E-45
CAD	rs2519093	0.98	1.2E-11	26343387	rs635634	PECAM1	1.89E-45
Liver enzyme levels (alkaline phosphatase)	rs579459	0.83	3.0E-123	22001757	rs635634	PECAM1	1.89E-45
E-selectin levels	rs651007	0.83	2.0E-82	20147318	rs635634	PECAM1	1.89E-45
Hematological and biochemical traits (ALP)	rs495828	0.83	4.0E-59	20139978	rs635634	PECAM1	1.89E-45
Serum alkaline phosphatase levels	rs651007	0.83	1.0E-56	24094242	rs635634	PECAM1	1.89E-45
Soluble levels of adhesion molecules (P-Selectin)	rs579459	0.83	2.0E-41	20167578	rs635634	PECAM1	1.89E-45
Blood metabolite ratios (ADSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs649129	0.83	9.0E-37	24816252	rs635634	PECAM1	1.89E-45
Blood metabolite ratios (DSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs495828	0.83	6.0E-34	24816252	rs635634	PECAM1	1.89E-45
Urinary metabolites (H-NMR features) (5.1825, Unknown)	rs579459	0.83	2.0E-32	24586186	rs635634	PECAM1	1.89E-45
Soluble E-selectin levels	rs579459	0.83	1.0E-29	19729612	rs635634	PECAM1	1.89E-45
Blood metabolite ratios (ADpSGEGDFXAEGGGVR/X-14304--leucylalanine)	rs579459	0.83	1.0E-28	24816252	rs635634	PECAM1	1.89E-45
Urinary metabolites (H-NMR features) (2.0525, Unknown)	rs579459	0.83	1.0E-28	24586186	rs635634	PECAM1	1.89E-45
End-stage coagulation (FVIII)	rs651007	0.83	2.0E-25	23381943	rs635634	PECAM1	1.89E-45
Blood metabolite levels (ADpSGEGDFXAEGGGVR)	rs651007	0.83	6.0E-20	24816252	rs635634	PECAM1	1.89E-45

Red blood cell traits (EA, RBCC)	rs579459	0.83	9.0E-18	23222517	rs635634	PECAM1	1.89E-45
Soluble levels of adhesion molecules (ICAM)	rs649129	0.83	1.0E-15	20167578	rs635634	PECAM1	1.89E-45
Coronary heart disease	rs579459	0.83	4.0E-14	21378990	rs635634	PECAM1	1.89E-45
vWF	rs579459	0.83	9.9E-13	23381943	rs635634	PECAM1	1.89E-45
Red blood cell count	rs495828	0.83	3.0E-12	20139978	rs635634	PECAM1	1.89E-45
Hematological and biochemical traits (Hb)	rs495828	0.83	1.0E-11	20139978	rs635634	PECAM1	1.89E-45
Hematological and biochemical traits (Ht)	rs495828	0.83	6.0E-10	20139978	rs635634	PECAM1	1.89E-45
Coronary artery disease or ischemic stroke	rs579459	0.83	2.0E-09	24262325	rs635634	PECAM1	1.89E-45
Metabolite levels (LDL)	rs651007	0.83	6.0E-09	21909109	rs635634	PECAM1	1.89E-45
Ferritin (log)   serum   (gene=FTL   FTH1)	rs651007	0.83	1.3E-08	25352340	rs635634	PECAM1	1.89E-45
FVIII Ag	rs579459	0.83	2.3E-08	23381943	rs635634	PECAM1	1.89E-45
Coronary artery disease or large artery stroke	rs579459	0.83	3.0E-08	24262325	rs635634	PECAM1	1.89E-45
Angiotensin-converting enzyme activity	rs495828	0.83	3.0E-08	20066004	rs635634	PECAM1	1.89E-45
LDL cholesterol	rs635634	1.00	2.0E-41	24097068	rs635634	SELE	9.64E-220
Cholesterol, total	rs635634	1.00	3.0E-35	24097068	rs635634	SELE	9.64E-220
E-selectin   plasma   (gene=SELE)	rs635634	1.00	5.0E-16	25147954	rs635634	SELE	9.64E-220
invasive epithelial ovarian cancer	rs635634	1.00	4.4E-09	25581431	rs635634	SELE	9.64E-220
Soluble ICAM-1	rs507666	0.99	3.0E-91	21533024	rs635634	SELE	9.64E-220
Lipid traits (LDL)	rs507666	0.99	2.0E-11	24386095	rs635634	SELE	9.64E-220
Lipid traits (TC)	rs507666	0.99	4.0E-11	24386095	rs635634	SELE	9.64E-220
Venous thromboembolism	rs2519093	0.98	8.0E-16	22672568	rs635634	SELE	9.64E-220
LDL-C transethnic	rs2519093	0.98	2.2E-13	23555291	rs635634	SELE	9.64E-220
CAD	rs2519093	0.98	1.2E-11	26343387	rs635634	SELE	9.64E-220
Liver enzyme levels (alkaline phosphatase)	rs579459	0.83	3.0E-123	22001757	rs635634	SELE	9.64E-220
E-selectin levels	rs651007	0.83	2.0E-82	20147318	rs635634	SELE	9.64E-220
Hematological and biochemical traits (ALP)	rs495828	0.83	4.0E-59	20139978	rs635634	SELE	9.64E-220
Serum alkaline phosphatase levels	rs651007	0.83	1.0E-56	24094242	rs635634	SELE	9.64E-220
Soluble levels of adhesion molecules (P-Selectin)	rs579459	0.83	2.0E-41	20167578	rs635634	SELE	9.64E-220
Blood metabolite ratios (ADSGEGDFAEGGGVR/ADpSGEGDFAEGGGVR)	rs649129	0.83	9.0E-37	24816252	rs635634	SELE	9.64E-220

)							
Blood metabolite ratios (DSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs495828	0.83	6.0E-34	24816252	rs635634	SELE	9.64E-220
Urinary metabolites (H-NMR features) (5.1825, Unknown)	rs579459	0.83	2.0E-32	24586186	rs635634	SELE	9.64E-220
Soluble E-selectin levels	rs579459	0.83	1.0E-29	19729612	rs635634	SELE	9.64E-220
Blood metabolite ratios (ADpSGEGDFXAEGGGVR/X-14304--leucylalanine)	rs579459	0.83	1.0E-28	24816252	rs635634	SELE	9.64E-220
Urinary metabolites (H-NMR features) (2.0525, Unknown)	rs579459	0.83	1.0E-28	24586186	rs635634	SELE	9.64E-220
End-stage coagulation (FVIII)	rs651007	0.83	2.0E-25	23381943	rs635634	SELE	9.64E-220
Blood metabolite levels (ADpSGEGDFXAEGGGVR)	rs651007	0.83	6.0E-20	24816252	rs635634	SELE	9.64E-220
Red blood cell traits (EA, RBCC)	rs579459	0.83	9.0E-18	23222517	rs635634	SELE	9.64E-220
Soluble levels of adhesion molecules (ICAM)	rs649129	0.83	1.0E-15	20167578	rs635634	SELE	9.64E-220
Coronary heart disease	rs579459	0.83	4.0E-14	21378990	rs635634	SELE	9.64E-220
vWF	rs579459	0.83	9.9E-13	23381943	rs635634	SELE	9.64E-220
Red blood cell count	rs495828	0.83	3.0E-12	20139978	rs635634	SELE	9.64E-220
Hematological and biochemical traits (Hb)	rs495828	0.83	1.0E-11	20139978	rs635634	SELE	9.64E-220
Hematological and biochemical traits (Ht)	rs495828	0.83	6.0E-10	20139978	rs635634	SELE	9.64E-220
Coronary artery disease or ischemic stroke	rs579459	0.83	2.0E-09	24262325	rs635634	SELE	9.64E-220
Metabolite levels (LDL)	rs651007	0.83	6.0E-09	21909109	rs635634	SELE	9.64E-220
Ferritin (log)   serum   (gene=FTL   FTH1)	rs651007	0.83	1.3E-08	25352340	rs635634	SELE	9.64E-220
FVIII Ag	rs579459	0.83	2.3E-08	23381943	rs635634	SELE	9.64E-220
Coronary artery disease or large artery stroke	rs579459	0.83	3.0E-08	24262325	rs635634	SELE	9.64E-220
Angiotensin-converting enzyme activity	rs495828	0.83	3.0E-08	20066004	rs635634	SELE	9.64E-220
Height	rs11599750	1.00	2.0E-13	20881960	rs11599750	IL27	1.40E-10
height	rs11599750	1.00	4.7E-13	25282103	rs11599750	IL27	1.40E-10
Bone mineral density (FNBMD)	rs7084921	0.88	9.0E-10	22504420	rs11599750	IL27	1.40E-10
Age-related hearing impairment (interaction)	rs4462272	0.62	8.0E-09	24939585	rs11599750	IL27	1.40E-10
Circulating vasoactive peptide levels (ADM)	rs2957692	0.63	1.0E-12	23381795	rs1580006	ADM	2.04E-15

Matrix_Metalloproteinase-3 CSF (gene=MMP3)	rs471994	1.00	2.3E-21	25340798	rs471994	MMP1	2.34E-35
Matrix metalloproteinase levels	rs495366	0.72	6.0E-34	20031604	rs471994	MMP1	2.34E-35
Matrix_Metalloproteinase-3 CSF (gene=MMP3)	rs7946057	1.00	2.4E-42	25340798	rs7946057	MMP3	1.22E-108
Matrix metalloproteinase levels	rs11225434	0.94	9.0E-29	20031604	rs7946057	MMP3	1.22E-108
Chronic obstructive pulmonary disease (severe)	rs626750	0.64	3.0E-09	24621683	rs17368659	MMP12	5.52E-97
Interleukin-18 levels	rs1834481	0.79	1.0E-08	20150558	rs75649625	IL18	1.46E-21
C-reactive protein	rs1169310	1.00	2.0E-08	18439552	rs1169306	LGALS3	6.47E-09
N-glycan levels (DG11)	rs735396	1.00	4.0E-08	21203500	rs1169306	LGALS3	6.47E-09
C-reactive protein (HA women)	rs2259816	0.99	3.0E-10	22939635	rs1169306	LGALS3	6.47E-09
Liver enzyme levels (GGT)	rs1169313	0.98	2.0E-10	18940312	rs1169306	LGALS3	6.47E-09
Gamma glutamyl transpeptidase	rs2393791	0.73	7.0E-30	21909109	rs1169306	LGALS3	6.47E-09
C-reactive protein levels	rs2393791	0.73	3.0E-10	24763700	rs1169306	LGALS3	6.47E-09
C-reactive protein and white blood cell count (CRP)	rs2393791	0.73	3.0E-09	22788528	rs1169306	LGALS3	6.47E-09
Liver enzyme levels (gamma-glutamyl transferase)	rs7310409	0.73	7.0E-45	22001757	rs1169306	LGALS3	6.47E-09
Urate levels	rs7188445	0.98	2.0E-09	23263486	rs11150189	XPNPEP2	7.00E-14
Thyroid hormone levels (TSH)	rs3813582	0.92	8.0E-18	23408906	rs11150189	XPNPEP2	7.00E-14
Thyroid hormone levels (TSH - Males)	rs3813582	0.92	6.0E-17	23408906	rs11150189	XPNPEP2	7.00E-14
Thyroid volume (thyroid volume)	rs17767419	0.92	9.0E-15	21565293	rs11150189	XPNPEP2	7.00E-14
Thyroid function	rs3813582	0.92	6.0E-10	22494929	rs11150189	XPNPEP2	7.00E-14
Vitamin B12 levels	rs492602	1.00	5.0E-17	18776911	rs492602	MMP10	7.69E-09
Cholesterol, total	rs492602	1.00	1.0E-16	24097068	rs492602	MMP10	7.69E-09
Blood metabolite ratios (ADSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs601338	0.99	2.0E-20	24816252	rs492602	MMP10	7.69E-09
Crohn's disease	rs516246	0.99	1.0E-15	23128233	rs492602	MMP10	7.69E-09
Blood metabolite levels (ADpSGEGDFXAEGGGVR)	rs601338	0.99	3.0E-11	24816252	rs492602	MMP10	7.69E-09
Liver enzyme levels (gamma-glutamyl	rs516246	0.99	8.0E-10	22001757	rs492602	MMP10	7.69E-09

transferase)							
Folate pathway vitamin levels (vitamin B12)	rs602662	0.89	3.0E-20	19303062	rs492602	MMP10	7.69E-09
Folate pathway vitamin levels (Plasma Vitamin B12)	rs602662	0.89	2.0E-15	19744961	rs492602	MMP10	7.69E-09
Metabolic traits (SM-11 + 2 other traits)	rs503279	0.89	4.0E-20	21886157	rs492602	MMP10	7.69E-09
Blood metabolite ratios (DSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs503279	0.89	1.0E-13	24816252	rs492602	MMP10	7.69E-09
Pubertal anthropometrics (Single Height-males)	rs281379	0.83	5.0E-08	23449627	rs492602	MMP10	7.69E-09
Liver enzyme levels (alkaline phosphatase)	rs281377	0.70	1.0E-15	22001757	rs492602	MMP10	7.69E-09
Urinary metabolites (H-NMR features) (5.2125, Fucose)	rs2287921	0.68	3.0E-36	24586186	rs492602	MMP10	7.69E-09
Retinal vascular caliber (Retinal venular caliber)	rs2287921	0.68	2.0E-25	21060863	rs492602	MMP10	7.69E-09
Urinary metabolites (H-NMR features) (5.2825, Fucose)	rs2287921	0.68	7.0E-19	24586186	rs492602	MMP10	7.69E-09
Urinary metabolites (H-NMR features) (5.2275, Fucose)	rs2287921	0.68	1.0E-12	24586186	rs492602	MMP10	7.69E-09
Dietary macronutrient intake (Fat)	rs838145	0.64	4.0E-10	23636237	rs492602	MMP10	7.69E-09
Urinary metabolites (H-NMR features) (5.2125, Fucose)	rs2287921	0.89	3.0E-36	24586186	rs33988101	LGALS3	3.57E-09
Retinal vascular caliber (Retinal venular caliber)	rs2287921	0.89	2.0E-25	21060863	rs33988101	LGALS3	3.57E-09
Urinary metabolites (H-NMR features) (5.2825, Fucose)	rs2287921	0.89	7.0E-19	24586186	rs33988101	LGALS3	3.57E-09
Urinary metabolites (H-NMR features) (5.2275, Fucose)	rs2287921	0.89	1.0E-12	24586186	rs33988101	LGALS3	3.57E-09
Crohn's disease	rs281379	0.73	7.0E-12	21102463	rs33988101	LGALS3	3.57E-09
Pubertal anthropometrics (Single Height-males)	rs281379	0.73	5.0E-08	23449627	rs33988101	LGALS3	3.57E-09
Dietary macronutrient intake (Fat)	rs838145	0.71	4.0E-10	23636237	rs33988101	LGALS3	3.57E-09
Folate pathway vitamin levels (vitamin B12)	rs602662	0.68	3.0E-20	19303062	rs33988101	LGALS3	3.57E-09
Folate pathway vitamin levels (Plasma Vitamin B12)	rs602662	0.68	2.0E-15	19744961	rs33988101	LGALS3	3.57E-09
Metabolic traits (SM-11 + 2 other traits)	rs503279	0.68	4.0E-20	21886157	rs33988101	LGALS3	3.57E-09
Blood metabolite ratios (DSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs503279	0.68	1.0E-13	24816252	rs33988101	LGALS3	3.57E-09

Blood metabolite ratios (ADSGEGDFXAEGGGVR/ADpSGEGDFXAEGGGVR)	rs601338	0.67	2.0E-20	24816252	rs33988101	LGALS3	3.57E-09
Blood metabolite levels (ADpSGEGDFXAEGGGVR)	rs601338	0.67	3.0E-11	24816252	rs33988101	LGALS3	3.57E-09
Liver enzyme levels (gamma-glutamyl transferase)	rs516246	0.67	8.0E-10	22001757	rs33988101	LGALS3	3.57E-09
Vitamin B12 levels	rs492602	0.66	5.0E-17	18776911	rs33988101	LGALS3	3.57E-09
Cholesterol, total	rs492602	0.66	1.0E-16	24097068	rs33988101	LGALS3	3.57E-09
Metabolite levels	rs4810479	1.00	2.0E-42	22916037	rs4810479	KITLG	4.44E-11
Lipid metabolism phenotypes (HDL.large, fasting)	rs6065904	0.72	4.0E-40	19936222	rs4810479	KITLG	4.44E-11
Lipid metabolism phenotypes (L-HDL-L/M-HDL-L)	rs6065904	0.72	2.0E-31	22286219	rs4810479	KITLG	4.44E-11
Inflammatory bowel disease	rs1569723	1.00	1.0E-13	23128233	rs1569723	CD40	3.01E-49
Kawasaki disease	rs1569723	1.00	6.0E-09	22446961	rs1569723	CD40	3.01E-49
Rheumatoid arthritis	rs4810485	0.99	3.0E-09	20453842	rs1569723	CD40	3.01E-49
Rheumatoid arthritis (EA)	rs4239702	0.85	1.0E-16	24390342	rs1569723	CD40	3.01E-49
Multiple sclerosis	rs2425752	0.64	5.0E-10	21833088	rs1569723	CD40	3.01E-49

Pleiotropy of reported trait protein SNPs with findings from previously published GWAS studies. Publically available studies were investigated and associations were reported for proxy SNPs with  $r^2$  LD above 0.6 and association P-value stronger than  $5e-8$ . Other trait – the trait investigated in the published GWAS; Other SNP – the index SNP in the published GWAS;  $r^2$  (EUR 1000G) – linkage disequilibrium between Olink-improve study index SNP and the other SNP; Other P-value – P-value as reported in published GWAS; Pubmed ID – the pubmed ID of the published GWAS; Olink SNP – the index SNP of the Olink-improve study; Olink Trait Protein – the trait protein associated in the Olink-improve study; Olink P-value – the P-value as also reported in table 1.



**Table S3**

Gene	Protein Name	#samples below LOD	CV%	Incl- uded	Mean (SD)	Median (IQR)	V(G)/Vp	R2
<b>ADM</b>	Pro-adrenomedullin	1	0.2	yes	0.87 (0.029)	0.87 (0.85-0.88)	-4.3% -4.3%	
<b>AGER</b>	Advanced glycosylation end product-specific receptor	0	0.15	yes	0.66 (0.043)	0.67 (0.64-0.69)	-3.9% -3.9%	
<b>AGRP</b>	Agouti-related protein	0	0.17	yes	0.64 (0.049)	0.64 (0.61-0.67)	-2.5% -0.9%	
<b>BNP</b>	Natriuretic peptides B	1434	0.2	yes	0.15 (0.16)	0.14 (0.001-0.22)	0.8% 6%	
<b>CASP8</b>	Caspase-8	518		no				
<b>CCL2</b>	Monocyte chemoattractant protein 1	0	0.2	yes	0.6 (0.069)	0.61 (0.56-0.65)	3.3% 10.9%	
<b>CCL20</b>	C-C motif chemokine 20	0	0.15	yes	0.79 (0.07)	0.78 (0.74-0.83)	34.4% 74.1%	
<b>CCL3</b>	C-C motif chemokine 3	0	0.17	yes	0.37 (0.095)	0.38 (0.31-0.44)	6.3% 17.3%	
<b>CCL4</b>	C-C motif chemokine 4	0	0.13	yes	0.88 (0.036)	0.88 (0.86-0.9)	-2.9% -2.1%	
<b>CD40</b>	Tumor necrosis factor receptor superfamily member 5	0	0.15	yes	0.97 (0.03)	0.96 (0.95-0.98)	-4.1% -4.1%	
<b>CD40LG</b>	CD40 ligand	0	0.2	yes	0.77 (0.1)	0.76 (0.69-0.84)	-3.5% -3.1%	
<b>CHI3L1</b>	Chitinase-3-like protein 1	0	0.16	yes	0.8 (0.066)	0.8 (0.76-0.84)	1.1% 6.6%	
<b>CSF1</b>	Macrophage colony-stimulating factor 1	0	0.15	yes	0.95 (0.015)	0.95 (0.94-0.96)	7.6% 20.1%	
<b>CSTB</b>	Cystatin-B	0	0.16	yes	0.74 (0.04)	0.74 (0.71-0.76)	-3.7% -3.7%	
<b>CTSD</b>	Cathepsin D	1	0.15	yes	0.87 (0.03)	0.87 (0.85-0.89)	3.4% 11.3%	

<b>CTSL1</b>	Cathepsin L1	0	0.16	yes	0.78 (0.034)	0.78 (0.75-0.8)	-4.1% -4.1%
<b>CX3CL1</b>	Fractalkine	0	0.26	yes	0.73 (0.045)	0.73 (0.7-0.76)	-0.6% 2.9%
<b>CXCL1</b>	Growth-regulated alpha protein	0	0.15	yes	0.79 (0.093)	0.8 (0.73-0.86)	1.4% 7.0%
<b>CXCL16</b>	C-X-C motif chemokine 16	0	0.17	yes	0.58 (0.041)	0.58 (0.56-0.61)	10.9% 26.6%
<b>CXCL6</b>	C-X-C motif chemokine 6	0	0.15	yes	0.78 (0.066)	0.78 (0.74-0.83)	-1.8% 0.5%
<b>DKK1</b>	Dickkopf-related protein 1	0	0.13	yes	0.72 (0.063)	0.72 (0.68-0.77)	-0.8% 2.7%
<b>EGF</b>	Pro-epidermal growth factor	0	0.15	yes	0.65 (0.15)	0.66 (0.55-0.76)	-4.1% -4.1%
<b>ESM1</b>	Endothelial cell-specific molecule 1	0	0.17	yes	0.58 (0.053)	0.58 (0.54-0.61)	-0.1% 4.1%
<b>F2R</b>	Proteinase-activated receptor 1	0	0.18	yes	0.84 (0.034)	0.84 (0.82-0.86)	3.4% 11.4%
<b>F3</b>	Tissue factor	0	0.18	yes	0.81 (0.03)	0.81 (0.79-0.83)	-3.8% -3.8%
<b>FABP4</b>	Fatty acid-binding protein, adipocyte	4	0.2	yes	0.48 (0.096)	0.49 (0.43-0.55)	2.5% 9.4%
<b>FAS</b>	Tumor necrosis factor receptor superfamily member 6	0	0.13	yes	0.89 (0.023)	0.9 (0.88-0.91)	8% 21%
<b>FGF23</b>	Fibroblast growth factor 23	0	0.17	yes	0.46 (0.08)	0.47 (0.41-0.51)	-3.9% -3.9%
<b>FIGF</b>	Vascular endothelial growth factor D	0	0.19	yes	0.84 (0.038)	0.84 (0.82-0.86)	6.1% 16.6%
<b>FST</b>	Follistatin	0	0.18	yes	0.74 (0.041)	0.74 (0.71-0.77)	0.7% 5.7%
<b>GAL</b>	Galanin peptides	0	0.15	yes	0.72 (0.06)	0.73 (0.69-0.76)	3.6% 11.6%
<b>GDF15</b>	Growth/differentiation factor	0	0.13	yes	0.99	0.99	-0.7%

	15				(0.027)	(0.97-1.0)	2.8%
<b>GH1</b>	Somatotropin	0	0.15	yes	0.92 (0.11)	0.93 (0.84-1.0)	-3.7% -3.2%
<b>HAVCR1</b>	Hepatitis A virus cellular receptor 1	0	0.16	yes	0.78 (0.06)	0.78 (0.74-0.82)	-2.7% -0.8%
<b>HBEGF</b>	Proheparin-binding EGF-like growth factor	0	0.18	yes	0.67 (0.042)	0.67 (0.65-0.7)	11% 26.6%
<b>HGF</b>	Hepatocyte growth factor	0	0.13	yes	0.83 (0.03)	0.83 (0.82-0.85)	2.8% 10%
<b>HSPB1</b>	Heat shock protein beta-1	303	0.16	yes	0.34 (0.24)	0.32 (0.14-0.53)	-0.7% 2.8%
<b>IKBKG</b>	NF-kappa-B essential modulator	163	0.22	yes	0.43 (0.22)	0.43 (0.29-0.58)	1.3% 7%
<b>IL16</b>	Pro-interleukin-16 [Cleaved into: Interleukin-16]	0	0.2	yes	0.63 (0.053)	0.63 (0.6-0.67)	6.4% 17.3%
<b>IL18</b>	Interleukin-18	0	0.12	yes	1.0 (0.026)	1.0 (0.99-1.01)	15.6% 36.1%
<b>IL1RL1</b>	Interleukin-1 receptor-like 1	0	0.13	yes	0.54 (0.072)	0.55 (0.5-0.59)	6.4% 17.3%
<b>IL1RN</b>	Interleukin-1 receptor antagonist protein	0		no			
<b>IL27</b>	Interleukin-27 subunit alpha	0	0.2	yes	0.42 (0.071)	0.43 (0.38-0.47)	-3.3% -2.3%
<b>IL4</b>	Interleukin-4	2559		no			
<b>IL6</b>	Interleukin-6	0	0.13	yes	0.7 (0.078)	0.69 (0.65-0.74)	-1.3% 1.7%
<b>IL6R</b>	Interleukin-6 receptor subunit alpha	0	0.14	yes	0.85 (0.029)	0.85 (0.83-0.87)	-3.7% -3.7%
<b>IL8</b>	Interleukin-8	1	0.15	yes	0.73 (0.055)	0.73 (0.69-0.76)	3.6% 11.9%
<b>ITGB1BP2</b>	Integrin beta-1-binding protein 2	1788		no			
<b>KITLG</b>	Kit ligand	0	0.12	yes	0.87 (0.031)	0.87 (0.85-0.89)	2.9% 10.3%

<b>KLK11</b>	Kallikrein-11	0	0.19	yes	0.72 (0.039)	0.72 (0.7-0.75)	12.7% 30.3%
<b>KLK6</b>	Kallikrein-6	0	0.15	yes	0.83 (0.028)	0.83 (0.81-0.85)	-1.2% 1.7%
<b>LEP</b>	Leptin	82	0.18	yes	0.47 (0.16)	0.5 (0.39-0.59)	-4.2% -4.2%
<b>LGALS3</b>	Galectin-3	0	0.16	yes	0.78 (0.03)	0.78 (0.76-0.8)	-4% -4%
<b>MB</b>	Myoglobin	3	0.17	yes	0.74 (0.048)	0.74 (0.71-0.77)	7.6% 19.6%
<b>MMP1</b>	Interstitial collagenase	7	0.19	yes	0.49 (0.19)	0.51 (0.4-0.61)	54.8% 114.2%
<b>MMP10</b>	Stromelysin-2	0	0.16	yes	0.82 (0.043)	0.82 (0.79-0.85)	1.6% 7.6%
<b>MMP12</b>	Macrophage metalloelastase	0	0.14	yes	0.85 (0.043)	0.85 (0.82-0.88)	-3.8% -3.8%
<b>MMP3</b>	Stromelysin-1	20	0.13	yes	-0.027 (0.3)	0.031 (-0.15-0.17)	6.3% 17.3%
<b>MMP7</b>	Matrilysin	1	0.2	yes	0.83 (0.11)	0.86 (0.76-0.92)	5.9% 16.5%
<b>MPO</b>	Myeloperoxidase	0	0.15	yes	0.57 (0.04)	0.57 (0.54-0.59)	-4.1% -4.1%
<b>MUC16</b>	Mucin-16	1	0.22	yes	0.71 (0.076)	0.72 (0.67-0.76)	-4.2% -4.2%
<b>NGF</b>	Beta-nerve growth factor	510	0.18	yes	-0.059 (0.13)	-0.064 (-0.16-0.029)	0.5% 5.5%
<b>NPPB</b>	Pro-Natriuretic peptides B	135	0.2	yes	0.56 (0.12)	0.57 (0.48-0.65)	7.2% 19.1%
<b>OLR1</b>	Oxidized low-density lipoprotein receptor 1	0	0.17	yes	0.67 (0.059)	0.66 (0.63-0.7)	7.1% 18.6%
<b>PAPPA</b>	Pappalysin-1	60	0.17	yes	0.32 (0.14)	0.34 (0.25-0.41)	4.6% 13.8%
<b>PDGFB</b>	Platelet-derived growth factor subunit B	0	0.13	yes	0.82 (0.091)	0.83 (0.77-0.88)	-3.9% -3.9%

<b>PECAM1</b>	Platelet endothelial cell adhesion molecule	0	0.2	yes	0.78 (0.032)	0.77 (0.75-0.8)	-3.1% -2.6%
<b>PGF</b>	Placenta growth factor	1	0.15	yes	0.88 (0.024)	0.88 (0.87-0.9)	6.5% 17.7%
<b>PLAT</b>	Tissue-type plasminogen activator	0	0.24	yes	0.89 (0.037)	0.89 (0.87-0.91)	-4% -4%
<b>PLAUR</b>	Urokinase plasminogen activator surface receptor	0	0.14	yes	1 (0.015)	1.0 (0.99-1.01)	4% 12.5%
<b>PRL</b>	Prolactin	3	0.15	yes	0.62 (0.081)	0.62 (0.57-0.67)	8.8% 22.1%
<b>PTX3</b>	Pentraxin-related protein PTX3	914		no			
<b>REN</b>	Renin	0	0.17	yes	0.87 (0.056)	0.87 (0.83-0.91)	0.9% 6%
<b>RETN</b>	Resistin	0	0.16	yes	0.84 (0.036)	0.84 (0.82-0.87)	21% 47%
<b>RNASE3</b>	Eosinophil cationic protein	2	0.16	yes	0.77 (0.053)	0.77 (0.74-0.8)	-2.8% -1.8%
<b>S100A12</b>	Protein S100-A12	669		no			
<b>SELE</b>	E-selectin	0	0.16	yes	0.77 (0.052)	0.77 (0.73-0.8)	5.5% 15.6%
<b>SELPLG</b>	P-selectin glycoprotein ligand 1	1081		no			
<b>SIRT2</b>	NAD-dependent protein deacetylase sirtuin-2	585		no			
<b>SPON1</b>	Spondin-1	0	0.17	yes	0.69 (0.033)	0.69 (0.67-0.71)	-3.6% -3.6%
<b>SRC</b>	Proto-oncogene tyrosine-protein kinase Src	0	0.21	yes	0.74 (0.12)	0.77 (0.65-0.85)	-2.8% -1.5%
<b>TEK</b>	Angiopoietin-1 receptor	0	0.14	yes	0.82 (0.023)	0.82 (0.8-0.83)	-4.2% -4.2%
<b>THBD</b>	Thrombomodulin	0	0.16	yes	1.0 (0.018)	1.0 (0.99-1)	2% 8.4%
<b>TNFRSF10B</b>	Tumor necrosis factor	0	0.24	yes	0.19	0.26	0.9%

	receptor superfamily member 10B				(0.21)	(0.086-0.33)	6.4%
<b>TNFRSF11B</b>	Osteoprotegerin	1	0.13	yes	1 (0.023)	1 (0.99-1)	10.7% 26.2%
<b>TNFRSF1A</b>	Tumor necrosis factor receptor superfamily member 1A	0		no			
<b>TNFRSF1B</b>	Tumor necrosis factor receptor superfamily member 1B	6	0.2	yes	0.63 (0.071)	0.64 (0.59-0.68)	6.6% 17.9%
<b>TNFSF10</b>	Tumor necrosis factor ligand superfamily member 10	0	0.16	yes	0.35 (0.11)	0.36 (0.28-0.43)	-3.9% -3.9%
<b>TNFSF11</b>	RANK ligand	0	0.15	yes	1.1 (0.014)	1.1 (1.1-1.1)	13.6% 32.1%
<b>TNFSF14</b>	Tumor necrosis factor ligand superfamily member 14	0	0.15	yes	0.73 (0.038)	0.73 (0.7-0.75)	3.5% 11.4%
<b>VEGFA</b>	Vascular endothelial growth factor A	0	0.13	yes	1 (0.017)	1 (1-1)	0.5% 5.4%
<b>XPNPEP2</b>	Xaa-Pro aminopeptidase 2	462	0.19	yes	0.47 (0.14)	0.48 (0.33-0.58)	-0.1% 4.1%

Overview of all 92 measured proteins, with quality control parameters, descriptive statistics and heritability estimates. All descriptive statistics are reported on the log10-transformed data that was used for analysis; *#samples below LOD* – the number of samples below limit of detection; *CV%* - coefficient of variation; *Included* – final choice on inclusion in analysis; *Mean (SD)* – mean and standard-deviation; *Median (IQR)* – median and inter-quartile range; *V(G)/Vp*– The GCTA calculated narrow-sense heritability. Missing values correspond to GCTA invertible matrix errors - meaning that too few samples were available for the V(G)/Vp estimation ; **R2 – description still missing**.