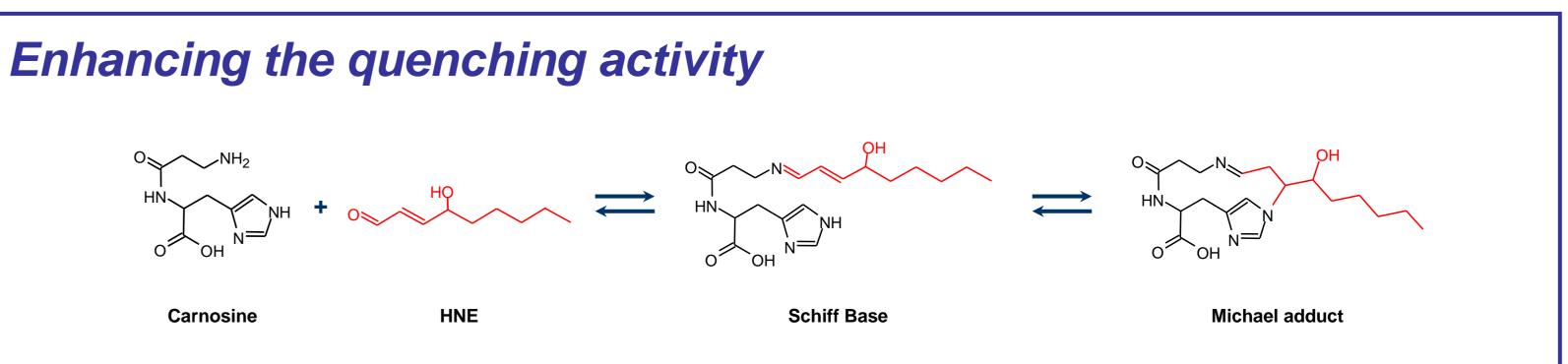
COMBINED IN SILICO APPROACHES FOR DRUG DESIGN AND PHARMACOKINETIC OPTIMIZATION OF A SET OF CARNOSINE ANALOGUES AS POTENT AND SELECTIVE CARBONYL QUENCHERS

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Background

- Reactive carbonyl species (RCS) are cytotoxic mediators generated by lipidoxidation of PUFAs, leading to alteration of cellular functions and inducing irreversible structural modifications to biomolecules.
- > RCS and the corresponding adducts with proteins (that is, carbonylated proteins) are widely used as biomarkers of lipidperoxidation and, in general, of oxidative stress.
- Moreover, there are several convincing evidences supporting a pathogenic role for RCS, such as in the case of diabetic-related diseases, age-dependent tissue dysfunction, and metabolic distress syndrome.
- Consequently, <u>RCS, in addition to being a predictive biomarker, also represents a biological target for drug</u> discovery.
- We recently found that the endogenous dipeptide carnosine (β -alanyl-L-histidine) is a specific quencher of α,β -unsaturated aldehydes [1]. Although carnosine is actively absorbed by intestinal transporter hPepT1, its therapeutic use is limited since it is unstable in human plasma due to the serum carnosinase activity (5). Moreover, the reactivity of carnosine towards RCS is markedly lower compared to other known quenchers. Hence, the rational design of new carnosine analogues should (1) increase the quenching activity of carnosine, maintaining its selectivity (2) confer plasma stability against human serum carnosinase, and (3) conserve an optimal recognition by hPepT1.



The modifications aimed to enhance the reactivity of the amino group are not largely exploitable since they would mine the specificity quenching also physiological aldehydes. Conversely, the specific Michael adduction can be optimized, by (1) modifying the imidazole ring to increase its nucleophilicity and (2) modulating the conformational profile of the Schiff base intermediate in order to favor a close conformation in which the imidazole ring approaches the reactive C3 to form the corresponding Michael adduct.

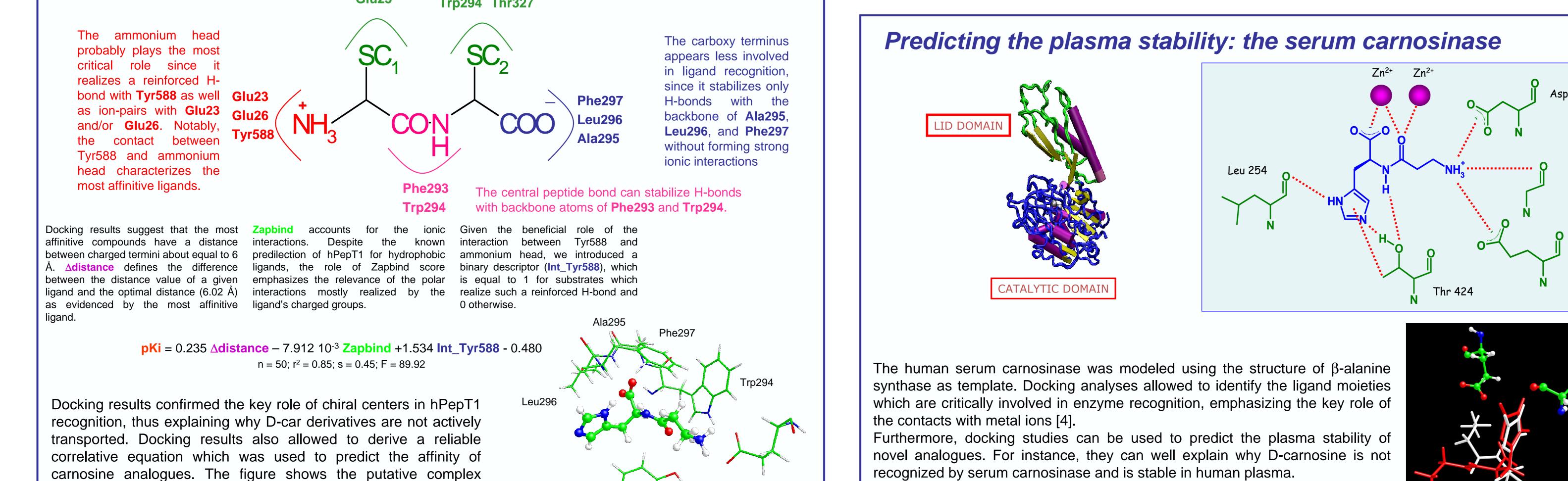
The key role of conformational profile: the case of **Schiff base carnosine-HNE**

- Accordingly, in silico approaches can support the design of carnosine derivatives by (1) parameterizing the factors which govern the quenching activity, (2) predicting the effects of serum carnosinase and (3) modeling the recognition by hPepT1.
- When the modifications were so significant to prevent the active transport, the marked hydrophilicity of carnosine analogues was modulated by designing prodrugs whose hydrolysis was predicted in silico by docking simulations with the major human carboxylesterases.

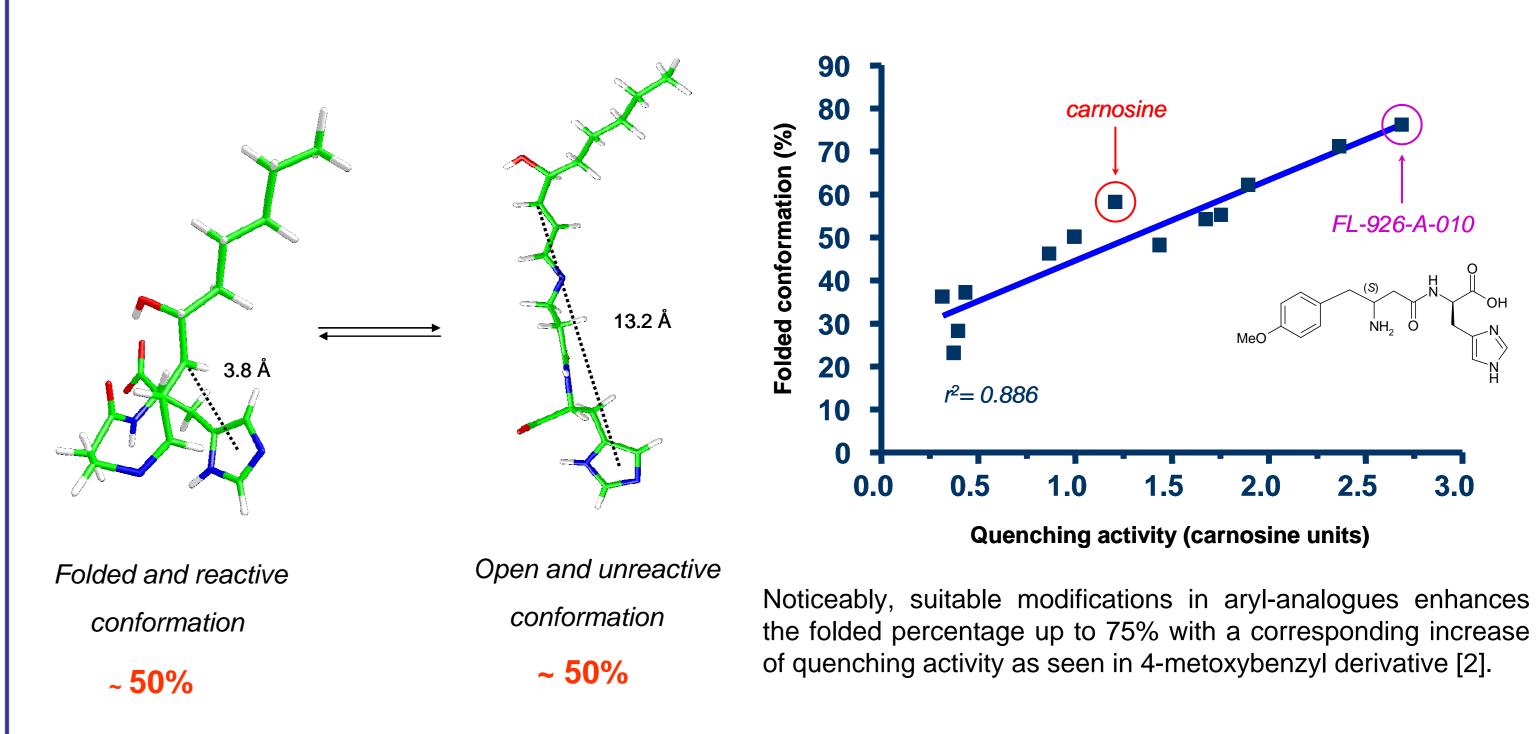
Predicting the active absorption: the homology model for hPepT1

The hPepT1 structure was modeled by fragments based on the resolved structure of lactose permease, LacY. Docking analyses involved a set of 50 known substrates and allowed the identification of a common pattern of interactions [3].

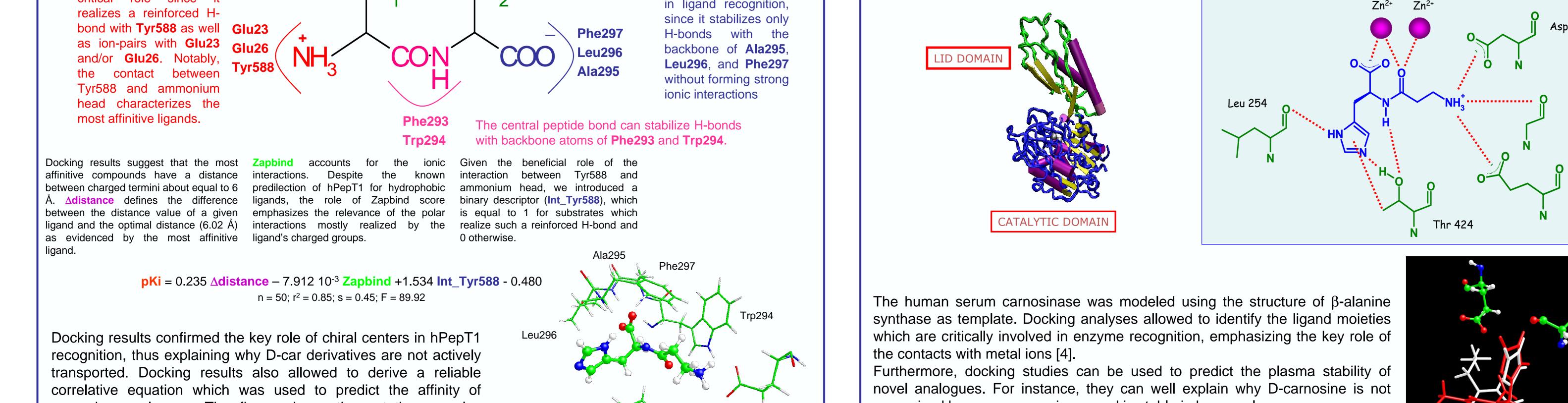
> The residues which interact with the side chains are heterogeneous, justifying the ability of hPepT1 to interact with structurally diverse substrates. It is possible to recognize a set of residues involved in the interaction with the N-terminal side chain (SC1) such as Asn22, Glu23, and Phe293, while the Cterminal side chain (SC2) contacts Trp294, Ile331, Glu291 and Thr327.



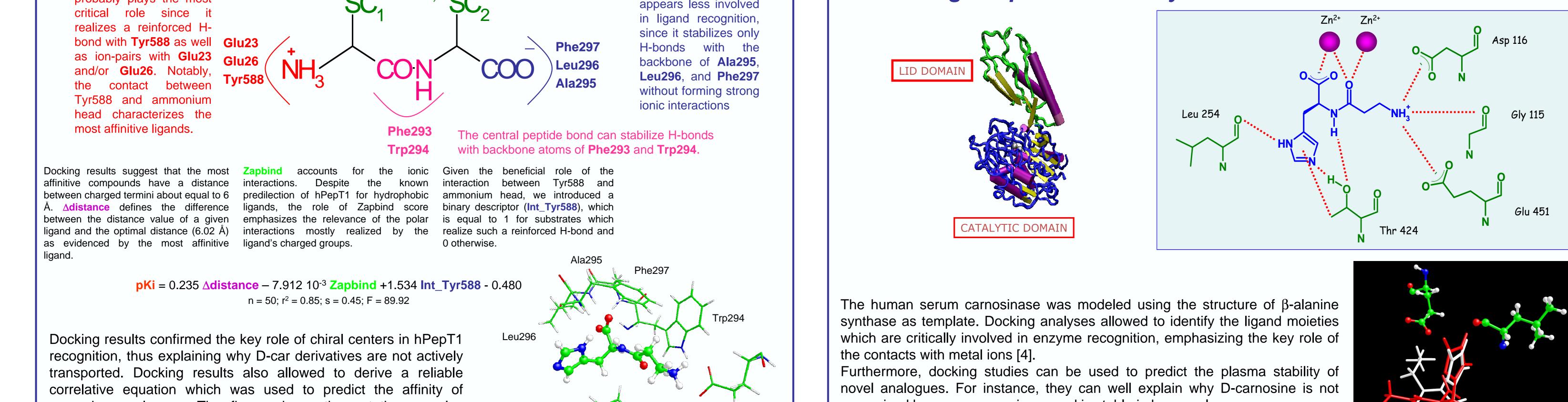


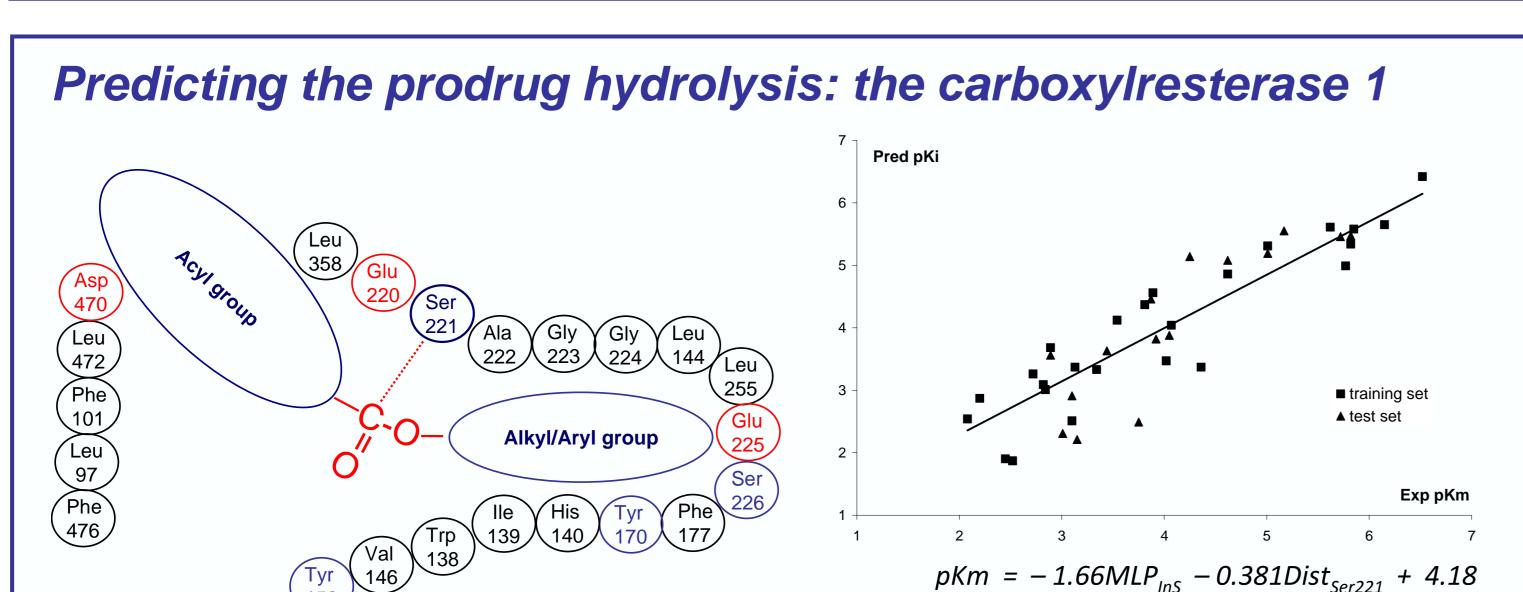


The Michael adduction can be parameterized calculating the nucleophilicity of new analogues using suitable indices mainly based on *ab initio* simulations. Preliminary analyses confirmed the significant reactivity of imidazole ring and unveiled the promising activity of furan ring.



Glu23





Conclusions: summarizing the obtained SARs

Irrelevant for quenching but crucial for PepT1 It can be largely modified. Required by CN2 but not by PepT1

Mandatory for quenching It can be replaced by other moieties able to yield Michael adducts Important for CN2 but not for PepT1

Based on obtained SARs and the performed simulations, design rational of new carnosine analogues followed such a pathway:

-carnosin

L-carnosine

Active, absorbed but unstable

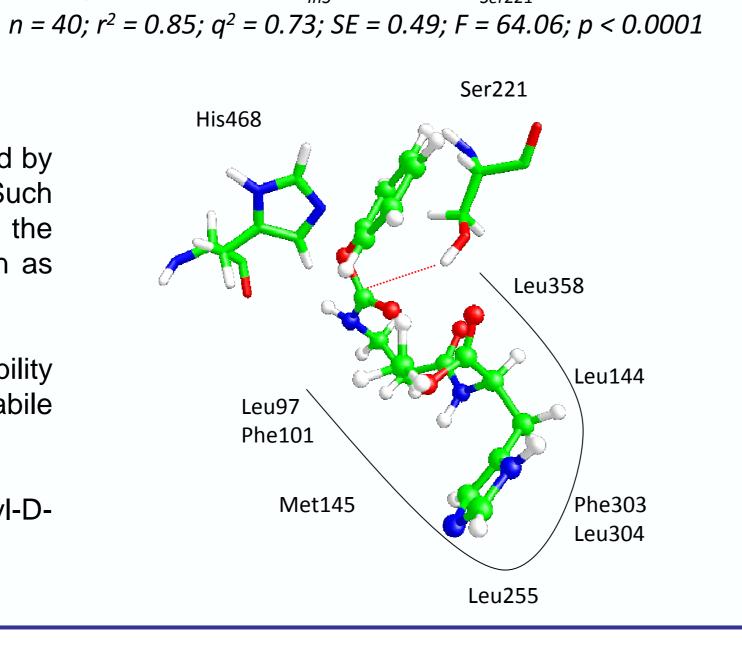
The design of suitable prodrugs for D-carnosine was supported by docking analyses involving the human carboxylesterase-1. Such studies firstly involved a set ok known substrates and allowed the identification of key residues involved in complex stabilization as well as the development of a robust predictive equations [5].

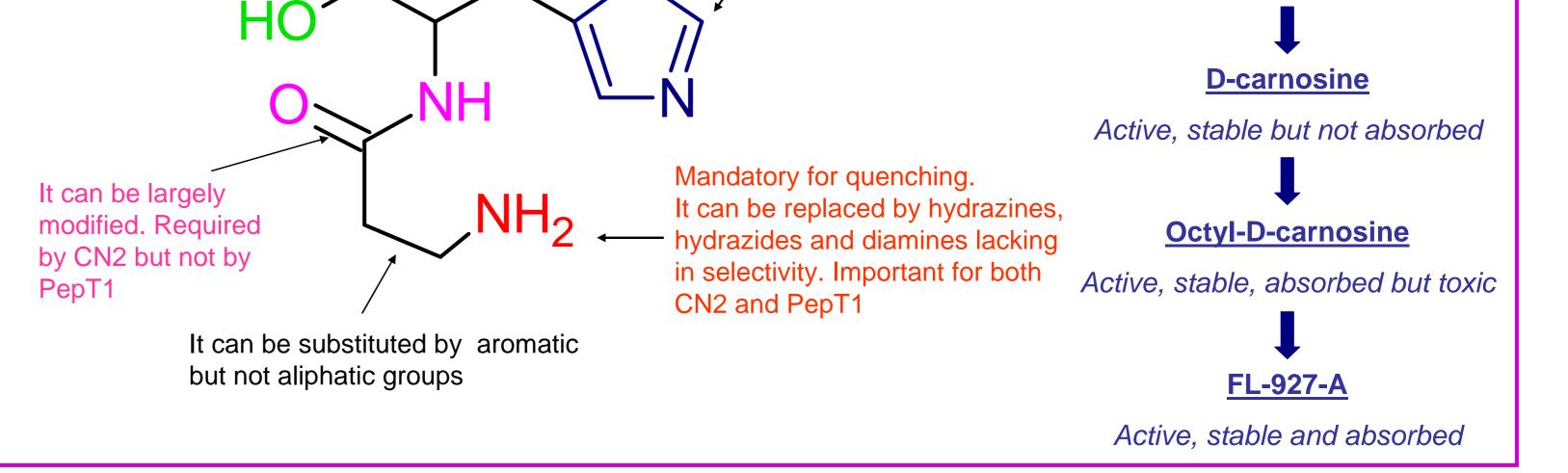
carnosine-hPepT1. The predicted affinity is in line with the

experimental value: 1.41 mM (pred) vs. 2.48 mM (exp).

The obtained results were then exploited to rationalize the stability of a set of D-carnosine prodrugs also predicting the more labile function for the bi-protected derivatives.

Globally, the performed simulations lead to the design of octyl-Dcarnosine as most promising prodrug for clinical studies.





References

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