

3

Original Article

1

Critical issues in setting micronutrient recommendations for pregnant women: an insight

4

2

3

4

**Cristiana Berti^{*1}, Tamás Decsi[†], Fiona Dykes[‡], Victoria Hall Moran[‡], Maria Hermoso[§],
Berthold Koletzko[§], Maddalena Massari^{*}, Luis A. Moreno[¶], Luis Serra-Majem^{**} and
Irene Cetin^{*}**

5

6

7

8

9

10

11

**Unit of Obstetrics and Gynecology, Department of Clinical Sciences Hospital 'L. Sacco' and Center for Fetal Research Giorgio Pardi, University of Milan, Milano, Italy, †University of Pécs, Hungary, ‡Maternal and Infant Nutrition and Nurture Unit (MAINN), University of Central Lancashire, UK, §Division of Metabolic Diseases and Nutritional Medicine, Dr von Hauner Children's Hospital, Ludwig-Maximilians-University of Munich, Germany, ¶'Growth, Exercise, Nutrition and Development' (GENUD) Research Group, Escuela Universitaria de Ciencias de la Salud, Universidad de Zaragoza, Zaragoza, Spain, and **Departamento de Ciencias Clínicas, Universidad de Las Palmas de Gran Canaria, Spain*

5

12

13

Abstract

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

The European Micronutrient Recommendations Aligned (EURRECA) project aims to provide standardized approaches to reveal and beneficially influence variability within the European Union in micronutrient recommendations for vulnerable population groups. Characterization of the 'vulnerability' together with the 'variability' of micronutrient needs represents the first step to creating guidelines for setting micronutrient recommendations within target populations. This paper describes some of the key factors and characteristics relevant to assess micronutrient requirements and formulate recommendations of micronutrients in pregnancy. Nutritional requirements during pregnancy increase to support fetal growth and development as well as maternal metabolism and tissue accretion. Micronutrients are involved in both embryonal and fetal organ development and overall pregnancy outcomes. Several factors may affect directly or indirectly fetal nourishment and the overall pregnancy outcomes, such as the quality of diet including intakes and bioavailability of micronutrients, maternal age, and the overall environment. The bioavailability of micronutrients during pregnancy varies depending on specific metabolic mechanisms because pregnancy is an anabolic and dynamic state orchestrated via hormones acting for both redirection of nutrients to highly specialized maternal tissues and transfer of nutrients to the developing fetus. The timing of prenatal intakes or supplementations of specific micronutrients is also crucial as pregnancy is characterized by different stages that represent a continuum, up to lactation and beyond. Consequently, nutrition during pregnancy might have long-lasting effects on the well-being of the mother and the fetus, and may further influence the health of the baby at a later age.

6

32

33

34

Keywords: EURRECA, pregnancy, vulnerability, recommendation, requirement, bioavailability, dietary factors, micronutrient intakes.

35

36

2

Correspondence: Irene Cetin, Unit of Obstetrics and Gynecology, Department of Clinical Sciences, Hospital 'L. Sacco' and Center for Fetal Research Giorgio Pardi, University of Milan, Via G. B. Grassi, 74, 20157 Milano, Italy. E-mail: irene.cetin@unimi.it

37

¹'Invernizzi Foundation' Fellowship.

38

39

40

41

42

43

44

45

Background

Nutrient recommendations are part of the basis for food policy and food-based dietary guidelines, and are used in nutrition labelling. The historical development of the concept of dietary recommendations for populations or groups has been reported by Aggett

et al. (1997). This evolution happened as a consequence of the understanding of the role of nutrients not only in the avoidance of clinical deficiencies but also in the reduction of the risk of chronic degenerative diseases.

A large heterogeneity of micronutrient recommendations exists across Europe, both quantitatively and

46

47

48

49

50

51

52

Table 1. Recommendations of some micronutrients for pregnant women and their related footnotes within some European countries (adapted from the original tables in references)

	Vit A	Vit D	Vit B ₁₂	Folate	Iodine	Zinc	Iron
	(μg day ⁻¹)					(mg day ⁻¹)	
United Kingdom (COMA 1991)	700	10	1.5 [§]	300	140 [§]	7 [§]	14.8 ^{§¶}
Italy (LARN 1996)	700	10 ^{††}	2.2	400 ^{††}	175	7	30 ^{††}
Nordic countries (NNR 2004)	800	10	2.0	500	175	9 ^{§§}	— ^{‡¶¶}
Spain (Moreiras <i>et al.</i> 2007) [†]	800	10	2.2	600 [‡]	135	20	18
D-A-CH (2000)	1.1 mg RE	5	3.5	600	230 (CH: 200)	10	30

Nordic countries, Denmark, Finland, Iceland, Norway, Sweden; D-A-CH, Germany, Austria, Switzerland; RE, retinol equivalent; Vit D, 10 μg day⁻¹ corresponds to 400 IU day⁻¹, 5 μg day⁻¹ corresponds to 200 IU day⁻¹; [†]from the second half of pregnancy; [‡]first and second half of gestation; [§]no increment. [¶]Insufficient for women with high menstrual losses where the most practical way of meeting iron requirements is to take iron supplements. ^{††}Dietary supplements or fortified foods may be required. ^{‡‡}The composition of the meal influences the utilization of dietary iron. The availability increases if the diet contains abundant amounts of vitamin C and meat or fish daily, while it is decreased at simultaneous intake of e.g. polyphenols or phytic acid. ^{§§}The utilization of zinc is negatively influenced by phytic acid and positively by animal protein. The recommended intakes are valid for a mixed animal/vegetable diet. For vegetarian cereal-based diets, a 25–30% higher intake is recommended. ^{¶¶}Iron balance during pregnancy requires iron stores of approximately 500 mg at the start of pregnancy. The physiological need of some women for iron cannot be satisfied during the last two-thirds of pregnancy with food only, and supplemental iron is therefore needed.

qualitatively; therefore, a common agreement should be sought on the different uses and applications of nutrient recommendations, as critically discussed by Pijls *et al.* (2009). Table 1 collates differences in the recommendations of some micronutrients for pregnant women from several European countries. The European Micronutrient Recommendations Aligned (EURRECA) project aims at providing standardized approaches to reveal variability within the European Union in micronutrient recommendations for population groups (Doets *et al.* 2008), with particular interest in ‘vulnerable populations’. ‘Vulnerable groups’ are defined as ‘population groups in a healthy population having a higher requirement’.

Pregnant women are considered a ‘vulnerable group’, as their nutritional requirements increase to support fetal and infant growth and development as well as maternal metabolism and tissue accretion. The estimated average requirement (EAR) is the daily intake value that is estimated to meet this

requirement, as defined by the specific indicator of adequacy, in half of the individuals in a life-stage or gender group (WHO/FAO 2004). The estimated total nutrient requirements during pregnancy can be derived from nutrients and energy accumulated in maternal tissues plus those necessary for products of pregnancy and lactation in addition to the baseline requirements for non-pregnant, non-lactating women. Determination of nutrient needs during pregnancy is a complex task because of the alteration of nutrient levels in tissues and fluids as a result of the hormone-induced changes in metabolism, shifts in plasma volume and changes in renal function as well as in patterns of urinary excretion.

When assessing micronutrient recommendations for pregnant women, besides physiological variation, environmental factors must be defined and explored. Macro-level factors such as socio-economic and political contexts, and food availability along with micro-level factors such as local cultural practices,

7

Key messages

- •
- •
- •

1 norms, lifestyles, attitudes and beliefs influence food
2 consumption (Pelto 1987; Hall Moran & Dykes 2009).
3 Moreover, application of any future nutritional guide-
4 lines should also consider new evidence for biological
5 role of micronutrients.

6 The aim of this paper is to discuss the nutritional
7 specificities of pregnant women and the approaches
8 underlying the definition of micronutrient dietary refer-
9 ence values and recommendation, i.e. the physi-
10 ological and environmental factors influencing the
11 bioavailability of micronutrients in pregnancy.

12 This narrative review develops from several
13 integrating meetings and activities within the
14 EURRECA project through evidence-based opinion
15 and explorative work (<http://www.eurreca.org>). Pub-
16 lications were searched using electronic databases
17 and websites, hand searching relevant journals,
18 contact with experts. The databases searched were
19 Embase, Medline and PubMed databases, and
20 Google-indexed scientific literature and periodics
21 from on-line University of Milan Library Service. We
22 used combinations of the following keywords: micro-
23 nutrient, requirement, intake, supplement, status, mal-
24 nutrition, deficiency, excess, overload, food, dietary
25 patterns, pregnancy, pregnancy need, pregnancy
26 health, pregnancy disease, pregnancy outcome, fetus,
27 placenta, newborn and mother. Only human studies
28 were considered, both original studies and reviews.
29 Moreover, official and national documents were used.

31 **Factors influencing micronutrient** 32 **recommendations in pregnancy**

33 The physiological requirement for a nutrient should be the
34 basis for calculating a reference intake. The ideal definition
35 of a physiological requirement is the amount and chemical
36 form of a nutrient that is needed systematically to maintain
37 normal health and development without disturbance of the
38 metabolism of any other nutrient. The corresponding dietary
39 requirement would be the intake sufficient to meet the
40 physiological requirement.

(Aggett *et al.* 1997)

43 When assessing recommendations, quality of diet,
44 genetics, physiological stress, pre-pregnancy body
45 mass index, body composition, gestational weight
46 gain, time of gestation, maternal age, lifestyle, socio-

economic status, culture, ethnicity, etc. must be taken
into account. This means that the bioavailability of
nutrients, depending not only on the composition of
diet or the chemical form of the nutrient but also on
the nutritional status or physiological stage, is a
crucial issue.

54 **Pregnancy physiological–metabolic factors**

55 Pregnancy is an anabolic state in which the body
56 undergoes significant physiological and anatomical
57 changes (Munro & Eckerman 1998). Hormones act
58 towards a redirection of nutrients to highly special-
59 ized maternal tissues (placenta and mammary gland)
60 and for the transfer of nutrients to the developing
61 fetus. Biochemical, metabolic and physiological
62 adjustments of the maternal organism meet the extra
63 demands of the developing fetus and placenta
64 (Kalhan 2000; Lain & Catalano 2007; Carlin &
65 Alfirevic 2008) and support the homeostasis of micro-
66 nutrients such as iodine (Zimmermann 2009), iron
67 (Milman 2006) and calcium (Kovacs 2008). Body
68 composition changes dramatically with maternal fat
69 accretion. Maternal fat storage increases in early to
70 mid-gestation and, during late gestation, these mater-
71 nal energy reserves are mobilized, following changes
72 in maternal insulin production, to provide an
73 increased supply of energy to the fetus. Improved
74 availability of substrates and precursors for fetal–
75 placental metabolism and hormone production is
76 mediated through increments in dietary intake and
77 endocrine changes that increase the availability of
78 nutritional substrates (Weissgerber & Wolfe 2006).

79 Pregnancy is characterized by different stages that
80 represent a continuum (Fig. 1) both in a life cycle
81 context and from a nutritional point of view. In
82 details, the first trimester is the time for the fetus
83 when organogenesis (embryogenesis) takes place, and
84 tissue patterns and organ systems are established; in
85 the second trimester, the fetus undergoes major cel-
86 lular adaptation and an increase in body size; during
87 the third trimester, organ systems mature, and there is
88 a significant increase in fetal body weight (Mullis &
89 Tonella 2008). Nutritional deficiencies occurring
90 during pregnancy might have long-lasting effects on
91 both maternal and infantile and adult health. In par-

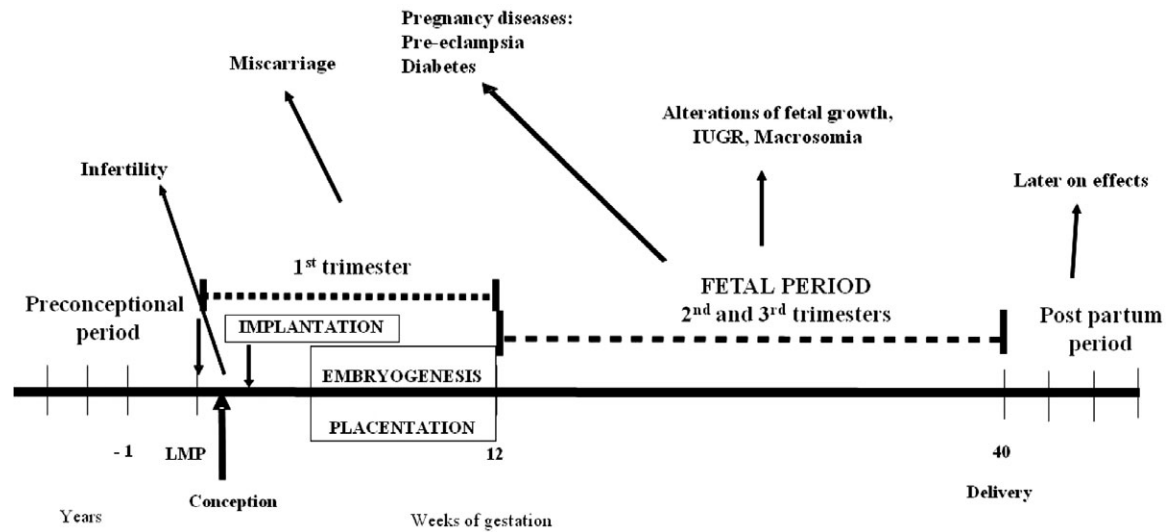


Fig. 1. Different pregnancy stages from the preconceptional to the post-partum period. Several specific malformations and pregnancy-related disorders may originate during each phase. LMP, last menstrual period. Adapted from Cetin *et al.* (2009).

54 55 58

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29

ticular, the periconceptional period, which encompasses preconception, conception, implantation, placentation and embryo- or organogenesis, is a stage of pregnancy representing a critical step in determining fetal development (reviewed in Cetin *et al.* 2009). Later on, placental function regulates fetal growth and development (Desoye & Hauguel-De Mouzon 2007). Several fetal diseases originate in the placenta and develop only later on in the fetus (Pardi & Cetin 2006). The ‘fetal’ or ‘early’ origins of adult disease hypothesis suggests that environmental factors, particularly nutrition, act through the processes of developmental plasticity (i.e. the ability of the fetus to respond to environmental cues by choosing a trajectory of development that often offers an adaptive advantage) to alter the development of the organism to such an extent that affects its capacity to cope with the environment in adult life, and therefore influences disease risk in adult life (Gluckman *et al.* 2005; McMillen & Robinson 2005).

The main characteristics of pregnancy that need to be highlighted from the nutritional perspective are as follows:

1. Pregnancy is characterized by a three-compartment model, i.e. mother/placenta/fetus. Each

of them has different metabolism; placenta transport function determines the composition of the umbilical cord blood providing nutrients and oxygen to the fetus to assure appropriate fetal growth (Cetin *et al.* 2005). Fetal growth is regulated by the balance between fetal nutrient demand and maternal–placental nutrient supply. Maternal nutrition and metabolism, utero-placental blood flow, size and transfer capabilities of the placenta all determine the maternal–placental supply of nutrients (Pardi & Cetin 2006).

2. Pregnancy is a dynamic state, during which adjustments in nutrient metabolism evolve continuously as the mother switches from an anabolic condition during early pregnancy to a catabolic state during late pregnancy (Catalano *et al.* 2002; Hauguel-de Mouzon *et al.* 2006). This switch is illustrated, e.g. in lipid metabolism going from fat deposition as a result of both hyperphagia and enhanced lipogenesis during the first and second trimesters to fat breakdown during the third trimester (Herrera 2000). Consequently, qualitative differences in dietary requirements exist during early and late pregnancy.

3. Maternal stores that have been developed before and throughout pregnancy will influence the composition of breast milk during lactation (Picciano 2003).

30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55

1 **Period of gestation**

2 Adequate maternal micronutrient status and intake
3 prior to conception and throughout the entire preg-
4 nancy is critical to ensure satisfactory birth outcomes
5 (reviewed in Picciano 2003 and Allen 2005). Timing of
6 maternal nutritional intake and status impacting spe-
7 cifically and differently on the embryonal/fetal organ
8 development, time of initiation and dose of prenatal
9 supplementation influencing maternal micronutrient
10 status, as well as the role of the interaction between
11 the pre- and post-natal environment in determining
12 final health outcomes, are important issues (Gardiner
13 2007). Preconceptional nutritional status appears to
14 be crucial for an optimal onset and development of
15 pregnancy (reviewed by Cetin *et al.* 2009), suggesting
16 the importance of adequate micronutrient intake of
17 all women of childbearing age. This may be, for
18 instance, of particular concern for calcium; if
19 adequate bone mass has not been accrued before
20 pregnancy and the intake of calcium from maternal
21 diet is low, calcium is taken from the maternal skel-
22 eton (Thomas & Weisman 2006).

23 Specifying the micronutrient recommendations for
24 specific periods of gestation may improve the overall
25 outcome of pregnancy. In this regard, mineral recom-
26 mendations by WHO/FAO (2004) are separated for
27 the first, second and third trimesters of pregnancy.
28 Spain recommends increased intakes of calcium,
29 iodine, magnesium, niacin and vitamin E from the
30 second half of pregnancy and an increased folate
31 intake in the first half of gestation (Moreiras *et al.*
32 2007). In the UK, increased thiamin intake is recom-
33 mended for the last trimester of pregnancy only
34 (COMA 1991). However, micronutrient recommen-
35 dations in most European countries do not differen-
36 tiate between specific periods of gestation.

37 *Effect of prenatal micronutrients on pregnancy outcomes*

38 The biological role and reliable functional markers or
39 indicators of status of the micronutrients that are of
40 considerable public health significance during preg-
41 nancy are shown in Table 2.

42 Increasing the intake of folic acid before and during
43 the first weeks of pregnancy can reduce birth defects
44

(MRC, 1991; Czeizel & Dudás 1992; Czeizel *et al.*
1999); hence, periconceptional folic acid supplements
in doses of 4000 and 400 µg daily are recommended
in addition to adequate dietary folate to prevent, respec-
tively, recurrence and occurrence of neural tube
defects (NTDs) (de Bree *et al.* 1997). Folate and/or
vitamin B₆/B₁₂ deficiencies as a result of deregulation
of their normal metabolism and/or low dietary intake
(reviewed in Steen *et al.* 1998, and Tamura & Picciano
2006) may induce elevation in plasma total homocys-
teine or hyperhomocysteinemia as a consequence of
decrease in the methylation cycle. Some 'placenta
events' are postulated to arise from deficiencies
of either folate and/or vitamin B₁₂ or defect within
the methionine-homocysteine metabolic pathways
(Goddijn-Wessel *et al.* 1996; Ray & Laskin 1999; ¹¹
Braekke *et al.* 2007; Dodds *et al.* 2008). Moreover,
altered homocysteine metabolism leading to hyper-
homocysteinemia has been proposed as the mecha-
nism involved in NTDs (Locksmith & Duff 1998).

Prenatal vitamin A or beta-carotene supplementa-
tion or fortification may reduce maternal mortality in
vitamin A-deficient mothers (West *et al.* 1999).
Although an excessive intake has been shown to be
teratogenic (McCaffery *et al.* 2003; Williamson 2006),
adequate maternal vitamin A status is crucial for fetal
lung development and maturation (reviewed in
Strobel *et al.* 2007). Interestingly, liver stores of retinol ¹²
in human fetuses were found to increase with the
progress of gestation and to vary with maternal
retinol levels, with the influence of maternal status
being greater in later pregnancy than in the earlier
stages (Shah & Rajalakshmi 1987). Consequently, ¹³
supplementation after mid-pregnancy at physiologi-
cal levels can improve fetal stores without the risk of
teratogenic effects. Insufficient vitamin A intake
seems to be associated to low birthweight (LBW)
(Strobel *et al.* 2007).

Dietary antioxidants (i.e. vitamin C, vitamin E, sele-
nium, zinc, beta-carotene) enhance many aspects of
the immune response and limit pathological aspects
of the cytokine-mediated response (Bendich 2001;
Arrigoni & De Tullio 2002). Recent reports associate
poor maternal selenium status as a nutritional factor
predisposing mothers to pre-eclampsia, as women
who develop pre-eclampsia have a lower selenium

Table 2. The biological role and the reliable biomarkers or indicators of status of micronutrients of considerable public health significance

Micronutrient	Function	Indicators of status	References
Folate	Involvement in the DNA cycle (cell replication); methylation cycle (amino acids cysteine and methionine cycle)	Erythrocyte folate [†] ; serum/plasma folate [†] ; serum/plasma total homocysteine [†]	WHO/FAO 2004; McNulty & Scott 2008
Vitamin B ₁₂	Conversion of homocysteine to methionine as cofactor of the methionine synthase	Serum/plasma vitamin B ₁₂ [†] ; serum/plasma methylmalonic acid (MMA) [†]	Ryan-Harshman & Aldoori 2008; Hoey <i>et al.</i> 2009
Vitamin A	Growth and differentiation of a number of cells and tissues	Serum retinol [†]	Ross 2006
Vitamin D	Bone resorption, intestinal calcium transport (calcium and bone homeostasis), modulation of transcription of cell cycle proteins, and cell-differentiating, anti-inflammatory and immunomodulatory properties	Plasma 25-hydroxyvitamin-D [25(OH)D]	WHO/FAO 2004
Iodine	Synthesis of thyroid hormones	Urinary Iodine excretion in 24 h [†] ; serum thyroid-stimulating hormone (TSH) [†]	WHO/FAO 2004; Zimmerman 2008; Ristic-Medic <i>et al.</i> 2009
Iron	Haematopoiesis; nucleic acid metabolism; carrier of oxygen to the tissues by red blood cell haemoglobin; transport medium for electrons within cells; integrated part of important enzyme systems	Haemoglobin [†] ; serum ferritin [†] ; serum transferrin receptor [†]	WHO/FAO 2004; Wood & Ronnenberg, 2006; Zimmerman 2008
Zinc	Structural, regulatory and catalytic functions as cofactor for numerous metalloenzymes	Plasma/serum zinc [†] ; prevalence of inadequate intakes of dietary zinc [†]	McCall <i>et al.</i> 2000; Lowe <i>et al.</i> 2009
Selenium	Protection of body tissues against oxidative stress, maintenance of defences against infection, and modulation of growth and development	Plasma/serum selenium [†] ; platelet or erythrocyte selenium [†] ; selenium-related proteins [†]	WHO/FAO 2004; Sunde <i>et al.</i> 2008

Indicators of status were taken from a table compiled by the Biomarkers of Status Working Party, which comprised a group of international micronutrient experts and EURRECA partners (Fairweather-Tait 2008), and successive updates[†]. Biomarkers reported here are those rated Excellent or Good according to a star rating used to classify the range of biomarkers available for each mineral/vitamin in relation to the limitations of the method.

[†]Also reviewed in 'Biomarkers of status/exposure. Minerals and vitamins'. RA1.2 Status Methods/IA3 Individuality, Vulnerability and Variability. July, 2008; 'BIOMARKERS OF STATUS/EXPOSURE. Iron, Zinc, Vitamin A, Vitamin B12, Folate, Iodine & Selenium'. RA1.2 Status Methods/IA3 Individuality, Vulnerability and Variability. February, 2010 (<http://www.eurreca.org>).

status (Rayman *et al.* 2003). Through the selenoproteins, selenium plays a critical role in regulating the antioxidant status. The various demands of pregnancy impose oxidative, metabolic and inflammatory stresses on the mother (Redman *et al.* 1999). When occurring during embryogenesis and in the placenta, oxidative stress causes adverse pregnancy outcomes such as birth defects, early pregnancy failure, miscarriage and pre-eclampsia (Agarwal *et al.* 2005; Jauniaux *et al.* 2006; Forges *et al.* 2007). Oxidative stress and inflammatory mediators seem to be involved in the abnormal implantation associated with pre-eclampsia (Roberts *et al.* 2003; Vanderlelie *et al.* 2005). Dietary antioxidants seem to play a crucial role in regulating the antioxidant status, thereby aiding in maintaining health. As an example,

when comparing women with a higher level of prenatal vitamin C ($\geq 11.734 \mu\text{g mL}^{-1}$) to women with a lower level of prenatal vitamin C ($< 8.997 \mu\text{g mL}^{-1}$), a significant lower trophoblast expression for the endothelial scavenger receptor low-density lipoprotein receptor-1 (LOX-1) and the apoptotic index in normal full-term pregnancy was detected in women with a higher level of prenatal vitamin C (Ahn *et al.* 2007). This seems to indicate that placental oxidative stress and apoptotic activity were associated with the gestational vitamin C status.

Increasing calcium intake can reduce the risk of pregnancy-induced hypertensive disorders (Thomas & Weisman 2006). A significant association was also observed between low 25-hydroxyvitamin D [25(OH)D] concentrations in early pregnancy and

1 subsequent pre-eclampsia (Bodnar *et al.* 2007). More-
2 over, a significant association was found between
3 maternal plasma 25(OH)D concentrations in mid-
4 gestation and the risk of developing gestational dia-
5 betes mellitus (Clifton-Bligh *et al.* 2008; Maghbooli
6 *et al.* 2008). Adequate maternal calcium intake can
7 affect positively both maternal and fetal bone health
8 because the fetus is dependent on maternal sources
9 for the total calcium load. On the contrary, maternal
10 bone loss during pregnancy might lead to osteoporosis
11 and fracture either contemporaneously or by
12 reducing peak bone mass in later life (Prentice 1994).
13 It was shown that whole body bone mineral content
14 of fetus increases between 32 and 33 and 40–41 weeks
15 of gestation. Several findings suggest that the greatest
16 fetal calcium accumulation occurs during the third
17 trimester. Hence, calcium consumption should be
18 encouraged during pregnancy to replace maternal
19 skeletal calcium stores that are depleted during this
20 period (Thomas & Weisman 2006). Similarly,
21 25 mg day⁻¹ of vitamin D should be given during the
22 last 3 months of pregnancy or 2500 mg day⁻¹ in one
23 dose at the beginning of the last trimester in countries
24 where sunshine exposure is negligible (i.e. in northern
25 countries) or to women avoiding dairy products for
26 cultural or dietary reasons (Salle *et al.* 2000).

16

27 The role of iron supplementation during pregnancy
28 is more controversial. An adequate iron intake is
29 mandatory for normal fetal growth and development,
30 although evidence for either a beneficial or harmful
31 effect of iron prophylaxis on pregnancy outcomes is
32 inconclusive and routine supplementation in preg-
33 nancy is a matter of debate (Breyman 2002). Iron-
34 deficiency anaemia (IDA), early in pregnancy, has
35 been found to be inversely related to placental size
36 and associated with reduced infant growth and
37 increased risk of adverse pregnancy outcomes (Scholl
38 & Hediger 1994; Hindmarsh *et al.* 2000; Ronnenberg
39 *et al.* 2004; Buckley *et al.* 2005). Moreover, maternal
40 anaemia during the second trimester has been associ-
41 ated with an increased risk of preterm delivery
42 (Scholl 2005). New insights are emerging into the role
43 of iron on neurocognitive and neurobehavioural
44 development of the fetus during the last two-thirds of
45 gestation and into the long-term consequences of
46 their perinatal deficiency (Beard 2003; Beard 2008).

17

18

47 Human brain growth spurt begins in the latter part of
48 the second trimester (Lukas & Campbell 2000), but
49 its peak velocity is during the last trimester of gesta-
50 tion and the first post-natal months (reviewed in Innis
51 2003). As the physiological needs of some women for
52 iron are not achieved during the last two-thirds of
53 pregnancy with food only, supplemental iron is there-
54 fore needed (Beaton 2000). Pregnant women using
55 iron supplements have a better iron status and a lower
56 frequency of IDA compared with women receiving no
57 supplement (Makrides *et al.* 2003; Milman *et al.* 2005).
58 Hence, pre-partum IDA seems to be prevented by
59 oral iron supplements (30–40 mg day⁻¹) taken from
60 the 20th week of gestation until delivery (Milman
61 *et al.* 2005). Women supplemented with iron pre-
62 sented a higher mean birth weight and a lower
63 preterm delivery incidence compared with the control
64 group (Cogswell *et al.* 2003; Siega-Riz *et al.* 2006). On
65 the contrary, high dose of iron supplement (more than
66 100 mg day⁻¹) was observed to be significantly associ-
67 ated to gestational diabetes (Bo *et al.* 2009).

19

68 Iodine intake is required to prevent the onset of
69 subclinical hypothyroidism of mother and fetus
70 during pregnancy, thus to prevent the possible risk of
71 brain damage of the fetus (WHO/FAO 2004). Mater-
72 nal iodine deficiency leads to fetal hypothyroidism
73 results in cretinism as thyroid hormones are critical
74 for normal brain development and maturation
75 (WHO/FAO 2004). However, if hypothyroidism
76 develops late in pregnancy, the neurological damage
77 is not as severe as when it is already present in early
78 pregnancy (WHO/FAO 2004). Third trimester preg-
79 nant women with urinary iodine concentrations
80 below 50 mcg L⁻¹ are significantly more likely to have
81 a small-for-gestational-age (SGA) infant. Higher
82 levels of thyroid-stimulating hormone were also asso-
83 ciated with a higher risk of having an SGA or LBW
84 newborn (Alvarez-Pedrerol *et al.* 2009). A random-
85 ized trial showed that a daily dose of 200 µg iodine
86 starting from 16–20th week of gestation in marginal
87 iodine deficiency appeared to be effective in prevent-
88 ing gestational goiter without enhancing the fre-
89 quency of post-partum thyroiditis (Antonangeli *et al.*
90 2002).

91 Docosahexaenoic acid (DHA; n-3) and arachidonic
92 acid are essential for fetal and neonatal growth and

development (Eilander *et al.* 2007), as long-chain polyunsaturated fatty acids are involved in modifications of neuronal membrane fluidity, function of neuronal membrane ionic channels and production of neurotransmitters and brain peptides (Innis 2007). If maternal DHA supply is limited, the fetus is particularly vulnerable to developmental deficits in the third trimester. Adequate maternal DHA intake or supplementation from the second trimester seems to be crucial in avoiding the potential perturbation of cellular environments in the offspring (reviewed in Innis 2003). The PeriLip Steering Committee and the Project Coordinating Committee of the early Nutrition Programming project stated that pregnant women should aim to achieve an average dietary intake of at least 200 mg DHA day⁻¹, and women of childbearing age should be recommended to consume one or two portions of sea fish per week, including oily fish such as salmon, herrings, etc. (Koletzko *et al.* 2008). Beneficial effects on subsequent infant visual function and neurodevelopment were also reported (reviewed in Judge *et al.* 2007 and Innis & Friesen 2008). Potential benefit of enhanced supply of n-3 fatty acids in preventing pre-eclampsia has been suggested in a recent prospective cohort study (Oken *et al.* 2007). Moreover, associations between maternal long-chain polyunsaturated fatty acids supplementation and a small reduction of risk of early preterm delivery in women with high-risk pregnancies (Horvath *et al.* 2007) as well as small increment in the duration of pregnancy (Szajewska *et al.* 2006) have been observed in several studies.

Effect of prenatal micronutrients on lactation and post-natal outcomes

The timing of prenatal micronutrient status and intake has been observed to condition breast milk composition. In particular, maternal vitamin A status from the second trimester of gestation seems to influence both retinol (vitamin A) concentration in breast milk (Mulismatun *et al.* 2001) and newborn development, and inadequacies during pregnancy are not compensated by post-natal supplementation (Strobel *et al.* 2007). Similarly, levels of vitamin E in transitional milk seem to be dependent on vitamin E and

polyunsaturated fatty acids intakes during the third trimester (Ortega *et al.* 1999).

The timing of prenatal nutrition seems to impact differently on the nature of adult diseases by programming post-natal pathophysiology. Accumulating data suggest that the early environment may modify the effects of the genome (Newnham *et al.* 2002; Fleming *et al.* 2004; Buckley *et al.* 2005; De Bo & Harding 2006; Gluckman *et al.* 2008). Molecular, cellular, metabolic, neuroendocrine and physiological adaptations in the early nutritional environment may cause a permanent alteration of the developmental pattern of cellular proliferation and differentiation in tissue and organ systems that may result in pathological consequences in adult life (Koletzko *et al.* 1998; McMillen & Robinson 2005). Studies in the offspring of women exposed to the Dutch Winter Famine showed that the nutrient challenge in the first trimester of pregnancy was linked to increased prevalence of coronary heart disease and obesity, and to raised blood lipids (Ravelli *et al.* 1999; Roseboom *et al.* 2000; Roseboom *et al.* 2001), whereas famine occurring during late gestation led to decreased glucose tolerance in adult life (Ravelli *et al.* 1998).

Poor maternal vitamin D status early in pregnancy may result in impaired maternal–fetal transfer of 25(OH)D and consequently reduced bone mineral content during infancy and childhood (Javaid *et al.* 2006). There are also arguments that low maternal vitamin D intake from the second trimester of pregnancy may be associated with the risk of recurrent wheeze at 3 or 5 years, suggesting that childhood asthma may be influenced by maternal diet during pregnancy (Camargo *et al.* 2007; Devereux *et al.* 2007). Maternal IDA during the last two-thirds of gestation is suggested to result in irreversible effects on neurochemistry and neurobiology (Beard 2003) such as schizophrenia in later life (Brown & Susser 2008; Insel *et al.* 2008). Data collected from a population-based cohort born from 1959 to 1967, and followed up for development of schizophrenia spectrum disorders from 1981 through to 1997, suggested that second and third trimester exposure to maternal haemoglobin concentrations ≤ 10.0 g dL⁻¹ was associated with a fourfold significantly increased rate of schizophrenia disorders in adult offspring (Insel *et al.*

2008). Similarly, in a cohort of births from 1978 to 1998, and followed from their 10th birthday, cohort members whose mothers were diagnosed with anaemia during pregnancy had a 1.60-fold increased risk of schizophrenia (Sørensen *et al.* 2010). It may be proposed that low haemoglobin concentrations compromise oxygen delivery to the developing fetus. In addition, insufficient iron *in utero* exposure may crucially disrupt neurodevelopment given that iron is essential for several metabolic processes involved in the development of brain structures and functions (i.e. dopaminergic neurotransmission, myelination and energy metabolism).

Birth spacing

It has been suggested that short birth intervals, by giving the mother insufficient time to recover from the nutritional burden of pregnancy, could adversely affect the nutritional status of both mother and child (King 2003). This nutritional burden may increase significantly when pregnancy overlaps with lactation, a period of very high maternal nutritional demand (Adair 1993). In a recent systematic review, Dewey & Cohen (2007) reported that, in studies conducted in developing countries, longer birth interval has been associated with a lower risk of child malnutrition in some populations. Where such a significant relationship was shown, the reduction in stunting associated with a previous birth interval of 35 months ranged from ~10–50%, although considerable residual confounding variables existed in the studies. One study suggested a possible increased risk of maternal anaemia associated with short interpregnancy interval (Conde-Agudelo & Belizán 2000), but iron supplementation during pregnancy was not accounted for in the analysis. There was no clear evidence of a link between interpregnancy interval and maternal anthropometric status, perhaps due in part to changes in hormonal regulation of nutrient partitioning between the malnourished mother and the fetus (Dewey & Cohen 2007). Considering the methodological limitations apparent in the majority of current studies on birth interval and maternal and child nutritional status, there is a clear and urgent need for further research.

Maternal diet

Eating patterns

Eating habits (e.g. vegetarian diet, fast food frequency, breakfast skipping) impact the adequacy of nutrient intakes. Some studies showed an association between dietary patterns and pregnancy outcomes. A reduction in the risk of early delivery has been associated with a maternal mid-pregnancy Mediterranean-type diet rich in fruit and vegetables, that is characterized by high vitamin C, folate, α -tocopherol, magnesium, calcium, iron and vitamin D intake and low sugar and cholesterol intake (Mikkelsen *et al.* 2008). Vujkovic and colleagues (2007) found an increased risk of cleft lip or palate and high plasma total homocysteine levels with a maternal periconceptional Western diet that was high in meat, pizza, legumes and potatoes, and low in fruits.

Vegans may be at risk of vitamin B₁₂ deficiency as they do not consume any animal products (ADA Report 2009). A long-term ovo-lacto vegetarian diet has been shown to result in significantly lower serum vitamin B₁₂ and higher plasma total homocysteine concentrations during pregnancy and in an increased risk of vitamin B₁₂ deficiency with respect to a Western diet (Koebnick *et al.* 2004).

Similarly, meal patterns seem to be related to pregnancy outcomes. It is recommended that pregnant women 'eat small to moderate-sized meals at regular intervals, and eat nutritious snacks' in order to meet the increased nutritional needs. Prolonged periods of time without food can cause hypoglycemia, thus a physiological stress. In a prospective cohort study of risk factors for preterm birth, women were asked to indicate how many meals and snacks they usually ate per day and the time of consumption (Siega-Riz *et al.* 2001). Results showed that consuming food at a lower optimal frequency was associated to a slightly increased risk for delivering preterm mainly after premature rupture of the membranes.

Consumption behaviour in pregnancy is influenced by a complex range of psychological, socio-demographic and cultural factors. For any given community, an understanding of these variables is required when transferring recommendations into action. Social class may affect the quality of diet. On

1 the whole, low-income groups consume a poor-quality
2 diet, and diet-related diseases, such as obesity and
3 diabetes, have begun increasing among lower- and
4 middle-income groups (Popkin 2003). High palatability,
5 high convenience, and the low cost of energy-dense
6 foods in conjunction with large portions and
7 low satiating power may be the principal reasons for
8 overeating and weight gain (Drewnowski & Darmon
9 2005). In particular, a review undertaken by Darmon
10 & Drewnowski (2008) about the relationship between
11 socio-economic status and eating behaviour showed
12 that studies on the plasma biomarkers of dietary
13 exposure provide evidence that socio-economic
14 status affects vitamin intakes, and that low-income
15 pregnant or breastfeeding women are at greater risk
16 of insufficient vitamin and mineral intakes. For
17 instance, a dietary survey undertaken in UK showed
18 that diet of low-income pregnant women did not meet
19 the EAR for folate, calcium and iron (Mouratidou
20 *et al.* 2006). In addition, maternal education seems to
21 correlate to food choices. As demonstrated in a large
22 population-based birth cohort study in Finland, pregnant
23 women with higher education levels had higher
24 daily consumption of vegetables, fruits and berries,
25 leading to higher intakes of dietary fibres, and of some
26 vitamins (Arkkola *et al.* 2006). Similarly, the Pregnancy,
27 Infection and Nutrition Study in North Carolina,
28 involving 2063 pregnant women, showed that high school
29 graduates had significantly higher Diet Quality Index
30 for Pregnancy scores, and that higher percentages of
31 recommended vegetable servings were consumed by
32 better-educated women (Bodnar & Siega-Riz 2002).

35 *Geographic factors*

36 The micronutrient status of an entire community may
37 be influenced by region and seasonal variation
38 impacting the availability of micronutrients. Iodine
39 and selenium deficiencies tend to be geographically
40 specific because of deficiencies in the soil and therefore
41 the food chain (Ladipo 2000; WHO/FAO 2004).
42 The majority of vitamin D comes from sunlight exposure.
43 In most situations, during summer, approximately
44 30 min of skin exposure to sunlight in the middle
45 of the day can provide 50 000 IU (1.25 mg) of

46 vitamin D to people with white skin. Latitude and
47 season as well as skin pigmentation and ethnicity
48 influence the ability of the skin to provide the total
49 vitamin D needs of the individual (WHO/FAO 2004;
50 Yu *et al.* 2009). This means that in locations around
51 the equator, the most physiologically relevant and
52 efficient way of acquiring vitamin D is to synthesize it
53 endogenously in the skin (Hollis & Wagner 2004),
54 whereas during winter at latitudes higher than 42°,
55 vitamin D synthesis is virtually zero (WHO/FAO
56 2004). Taken together, these findings suggest that not
57 routinely sun-replete individuals or persons with
58 darker pigmentation should correct their vitamin D
59 status by consuming the amounts of vitamin D appropriate
60 for their population. Unfortunately, a recent review
61 by Hollis & Wagner (2004) indicated that, currently,
62 the appropriate dose of vitamin D during pregnancy
63 is unknown, although it appears to be greater
64 than the current dietary reference intake of
65 5–10 µg day⁻¹, and that further studies are necessary
66 to determine optimal vitamin D intakes for pregnancy
67 as a function of latitude and race. This concern was
68 confirmed by a prospective randomized controlled
69 study that took place in the UK comparing the effects
70 of a single dose of 200 000 IU vitamin D (calciferol)
71 and of a daily dose of 800 IU vitamin D (ergocalciferol)
72 from the 27th week to delivery on pregnant women
73 and their baby at delivery (Yu *et al.* 2009).
74 Results showed that despite supplementation
75 enhanced significantly the 25(OH)D levels within
76 supplemented groups with respect to the untreated
77 group, vitamin D sufficiency >50 nmol L⁻¹ was
78 achieved only in 30% of supplemented women, and
79 only 8% of babies were vitamin D sufficient in the
80 supplement group.

82 *Micronutrient bioavailability and diet*

83 Appropriate intake of micronutrients depends not
84 only on the quality of diet but also on their bioavailability.
85 Lack of accurate data on micronutrients' bioavailability
86 from natural food sources may be an ongoing concern
87 for policy-makers for setting dietary recommendations.
88 As an example, the recommended nutrient intakes
89 for dietary zinc (mg day⁻¹) to meet the normative
90 storage requirements from diets by the

Table 3. Recommended nutrient intakes for dietary zinc (mg day⁻¹) in pregnancy to meet the normative storage requirements from diets differing in zinc bioavailability and principal dietary characteristics for categorizing diets according to the potential bioavailability of their zinc[†] (adapted from WHO/FAO 2004)

Trimester	High bioavailability	Moderate bioavailability	Low bioavailability
	Refined diets low in cereal fibre and phytic acid content, with phytate–zinc molar ratio <5; adequate protein content principally from non-vegetable sources, such as meats and fish.	Mixed diets containing animal-fish protein. Lacto-ovo, ovo-vegetarian, or vegan diets not based primarily on unrefined cereal grains or high-extraction-rate flours. Phytate–zinc molar ratio of total diet = 5–15, or not >10 if more than 50% of the energy intake is accounted for by unfermented, unrefined cereal grains and flours and the diet is fortified with inorganic calcium salts. Availability of zinc improves when the diet includes animal protein or milks, or other protein sources or milks.	Diets high in unrefined, unfermented and ungerminated cereal grain, especially when fortified with inorganic calcium salts and intake of animal protein is negligible. Phytate–zinc molar ratio of total diet >15. High-phytate, soya–protein products as the primary protein source. Diets in which approximately 50% of the energy intake is accounted for by the following high-phytate foods: high-extraction-rate (≥90%) wheat, rice, maize, grains and flours, oatmeal and millet; chapatti flours and tanok; sorghum, cowpeas, pigeon peas, grams, kidney beans, black-eyed beans and groundnut flours. High intakes of inorganic calcium salts, either as supplements or as adventitious contaminants, potentiate the inhibitory effects, and low intakes of animal protein exacerbate these effects.
First	3.4	5.5	11.0
Second	4.2	7.0	14.0
Third	6.0	10.0	20.0

[†]At intakes adequate to meet the average normative requirements for absorbed zinc, the three availability levels correspond to 50%, 30% and 15% absorption.

Joint Food and Agriculture Organization of the United Nations/World Health Organization (FAO/WHO) Expert Consultation on Human Vitamin and Mineral Requirements (WHO/FAO 2004) are stated according to different bioavailabilities (Table 3).

Dietary factors such as food matrix, chemical form, processing and cooking methods may modify micronutrient bioavailability (Hotz & Gibson 2007). They may limit absorption through nutritional interactions (e.g. fibre/phytate–minerals complexes), mineral–mineral interactions involved in the same metabolism, competition for a common transport site or transport ligand (e.g. zinc/copper, iron/manganese), and effects of drugs or chemicals on the metabolism of the nutrient (Keen *et al.* 2003). Alternatively, they may enhance absorption as in the case of iron if the diet contains abundant amounts of vitamin C and meat/fish (Gibson *et al.* 2006), or for carotenoids in the presence of dietary fats (van Het Hof *et al.* 2000), or for zinc by germination of cereals and legumes (Gibson *et al.* 2006). Interestingly, based on this assumption, the footnote to the original table in the

Nordic Country recommendation (NNR 2004) (see Table 1) states: ‘The composition of the meal influences the utilization of dietary iron. The availability increases if the diet contains abundant amounts of vitamin C and meat or fish daily, while it is decreased at simultaneous intake of, e.g. polyphenols or phytic acid; the utilization of zinc is negatively influenced by phytic acid and positively by animal protein. The recommended intakes are valid for a mixed animal/vegetable diet. For vegetarian cereal based diets, a 25–30% higher intake is recommended’.

Similarly, the form of micronutrient influences its bioavailability: i.e. haem-iron is absorbed better than non-haem iron. On the contrary, there is conflicting evidence as to whether the extent of conjugation of polyglutamyl folate is a limiting factor in folate bioavailability. Estimates of the extent of lower bioavailability of food folates compared with folic acid show great variation, depending on the methodological approach used (McNulty & Pentieva 2006). The EAR for vitamin A, expressed as mg retinol equivalents (mg RE), should account for the proportionate bio-

1 availability of preformed vitamin A (about 90%) and
2 provitamin A carotenoids from a diet that contains
3 sufficient fat (WHO/FAO 2004).

5 **Maternal age**

6 Maternal age represents a critical factor in micronu-
7 trient requirement. Accordingly, the dietary recom-
8 mended intake for micronutrients should take into
9 account maternal age; e.g. US recommendations for
10 vitamin K, vitamin C, calcium, phosphorus, magne-
11 sium and zinc in pregnancy are separated for age >18
12 or ≤18 years by the Institute of Medicine's Food and
13 ²⁶ Nutrition Board (<http://www.usda.gov>). Adolescent
14 pregnancy, as defined by WHO as pregnancy in those
15 ²⁷ aged 10–19 years (WHO 2004), appears to be a risk
16 factor for micronutrient deficiencies (Lenders *et al.*
17 ²⁸ 2000; Hall Moran 2007a). A systematic review of the
18 nutrient intakes of pregnant adolescents living in
19 industrialized countries suggested that, compared
20 with US dietary reference intake values, intake of
21 energy, iron, folate, calcium, vitamin E and magne-
22 sium were lower than those currently recommended
23 (Hall Moran 2007b).

24 In recent decades, adolescent pregnancy has
25 become an important public health issue because of
26 associated poor obstetric outcomes, particularly with
27 respect to fetal growth restriction and preterm deliv-
28 ery. Approximately a fifth of all births worldwide are
29 to adolescent mothers (Population Reference Bureau
30 2000) and, although the general trend over the past 20
31 years in Europe is that of declining adolescent preg-
32 nancy and birth rate, the distributions across Europe
33 are large, ranging from 42.69 live births per 1000
34 women aged 15–19 in Tajikistan to 5.39 live births per
35 1000 women in Switzerland (Avery & Lazdane 2008).
36 Despite this, relatively little is known about nutrient
37 intakes of adolescents during pregnancy, and few pro-
38 spective studies have been conducted in this popula-
39 tion. One recent prospective, observational study of
40 500 adolescents conducted in the UK highlighted the
41 extent of poor vitamin D status in pregnant adoles-
42 cents and suggested a clear relationship between
43 maternal folate and iron status and the incidence of
44 SGA birth and preterm delivery in this cohort (Baker
45 *et al.* 2009).

46 Despite the increasing prevalence of pregnancy in
47 women over the age of 40 years as a result of recent
48 advances in assisted reproductive technology, to our
49 knowledge, there are no studies related to nutritional
50 needs and reference values in this population of preg-
51 nant women. This lack of knowledge is reflected in the
52 European micronutrient recommendations for preg-
53 nancy, the vast majority of which do not differentiate
54 for maternal age.

55 **Conclusion**

56 Targeted recommendations must be given to guide
57 pregnant women in their food choice and dietary
58 supplement use so that they may obtain adequate
59 nutritional status and meet the increased need for
60 nutrients. The term 'vulnerability' represents a key
61 concept in assessing nutrient needs and defining rec-
62 ommended nutrient intakes for target populations at
63 risk of low intake. Several physiological and meta-
64 bolic factors characterize pregnant women such as
65 adaptation and timing of gestation, and determine
66 their nutritional requirements. In addition, environ-
67 mental and demographic variables seem to influence
68 the overall quality of diet and the adequacy of micro-
69 nutrient intake during pregnancy. Unfortunately, a
70 large number of European recommendations do not
71 consider these factors. Moreover, current research is
72 limited by sampling and measurement bias, and find-
73 ings are often inconclusive or contradictory. Thereby,
74 further studies and actions are urgently warranted to
75 address limitations and to:

- 76 • determine optimal biomarkers and concentrations
77 even with regards to non-classic actions of micronu-
78 trients on maternal and fetal outcomes;
- 79 • investigate of the most effective way to supply
80 micronutrients, including appropriate timing and
81 dosage. In this context, strategies of supplementation
82 and dietary intervention are currently under discus-
83 sion. Several studies are ongoing to evaluate the
84 effect of different timing in pregnancy (i.e. early or
85 late pregnancy) as well as the different frequencies of
86 supplementation (i.e. daily or weekly). Forms of
87 micronutrient supplement/intake are also of interest
88 as it is well acknowledged that micronutrient status is
89
90

1 influenced by both the content and the bioavailability
2 of the micronutrient in the diet;

3 • explore the influence of age and of role of socio-
4 economic factors on the nutrient requirements of
5 pregnant women.

6 Sources of funding

7
8 This research was undertaken as an activity of
9 the European Micronutrient Recommendations
10 Aligned (EURRECA) Network of Excellence
11 (<http://www.eurreca.org>) funded by the European
12 Commission Contract Number FP6 036196-2
13 (FOOD).

14 References

15
16 ADA Report (2009) Position of the American Dietetic
17 Association: vegetarian diets. *Journal of the American*
18 *Dietetic Association* **109**, 1266–1282.
19 Adair S-R. (1993) Biological determinants of pregnancy
20 weight gain: a longitudinal study of Filipino women.
21 *American Journal of Clinical Nutrition* **57**, 365–372.
22 Agarwal A., Gupta S. & Sharma R.K. (2005) Role of oxi-
23 dative stress in female reproduction. *Reproductive*
24 *Biology and Endocrinology* **3**, 28. doi:10.1186/1477-7827-
25 3-28.
26 Aggett P.J., Bresson J., Haschke F., Hernell O., Koletzko B.,
27 Lafeber H.N. *et al.* (1997) Recommended dietary allow-
28 ances (RDAs), recommended dietary intakes (RDIs),
29 recommended nutrient intakes (RNIs), and population
30 reference intakes (PRIs) are not 'recommended intakes'.
31 *Journal of Pediatric Gastroenterology and Nutrition* **25**,
32 236–241.
33 Ahn Y., Kim Y., Park H., Park B. & Lee H. (2007) Prenatal
34 vitamin C status is associated with placental apopto-
35 sis in normal-term human pregnancies. *Placenta* **28**,
36 31–38.
37 Allen L.H. (2005) Multiple micronutrients in pregnancy
38 and lactation: an overview. *American Journal of Clinical*
39 *Nutrition* **81**, 1206S–1212S.
40 Alvarez-Pedrerol M., Guxens M., Mendez M., Canet Y.,
41 Martorell R., Espada M. *et al.* (2009) Iodine levels and
42 thyroid hormones in healthy pregnant women and birth
43 weight of their offspring. *European Journal of Endocri-*
44 *nology* **160**, 423–429.
45 Antonangeli L., Maccherini D., Cavaliere R., Di Giulio C.,
46 Reinhardt B., Pinchera P. *et al.* (2002) Comparison of
47 two different doses of iodide in the prevention of gesta-
48 tional goiter in marginal iodine deficiency: a longitudinal
49 study. *European Journal of Endocrinology* **147**, 29–34.

50 Arkkola T., Uusitalo U., Pietikäinen M., Metsälä J.,
51 Kronberg-Kippilä C., Erkkola M. *et al.* (2006) Dietary
52 intake and use of dietary supplements in relation to
53 demographic variables among pregnant Finnish women.
54 *British Journal of Nutrition* **96**, 913–920.
55 Arrigoni O. & De Tullio M.C. (2002) Ascorbic acid: much
56 more than just an antioxidant. *Biochimica et Biophysica*
57 *Acta* **1569**, 1–9.
58 Ashwell M., Lambert J.P., Alles M.S., Branca F., Bucchini
59 L., Brzozowska A. *et al.* (2008) How we will produce the
60 evidence-based EURRECA toolkit to support nutrition
61 and food. *European Journal of Nutrition* **47**, S2–S16. [20]
62 Avery L. & Lazdane G. (2008) What do we know about
63 sexual and reproductive health of adolescents in
64 Europe? *The European Journal of Contraception and*
65 *Reproductive Health Care* **13**, 58–70.
66 Baker P.N., Wheeler S.J., Sanders T.A., Thomas J.E.,
67 Hutchinson C.J., Clarke K. *et al.* (2009) A prospective
68 study of micronutrient status in adolescent pregnancy.
69 *American Journal of Clinical Nutrition* **89**, 1114–1124.
70 Beard J. (2003) Iron deficiency alters brain development
71 and functioning. *Journal of Nutrition* **133**, 1468S–1472S.
72 Beard J.L. (2008) Why iron deficiency is important in
73 infant development. *Journal of Nutrition* **138**, 2534–2536.
74 Beaton G.H. (2000) Iron needs during pregnancy: do we
75 need to rethink our targets? *American Journal of Clinical*
76 *Nutrition* **72**, 265S–271S.
77 Bendich A. (2001) Micronutrients in women's health and
78 immune function. *Nutrition* **17**, 858–867.
79 Bo S., Menato G., Villois P., Gambino R., Cassader M.,
80 Cotrino I. *et al.* (2009) Iron supplementation and gesta-
81 tional diabetes in midpregnancy. *American Journal of*
82 *Obstetrics and Gynecology* **201**, 158.e1–158.e6.
83 Bodnar L.M. & Siega-Riz A.M. (2002) A diet quality
84 index for pregnancy detects variation in diet and differ-
85 ences by sociodemographic factors. *Public Health Nutri-*
86 *tion* **5**, 801–809.
87 Bodnar L.M., Catov J.M., Simhan H.N., Holick M.F.,
88 Powers R.W. & Roberts J.M. (2007) Maternal vitamin D
89 deficiency increases the risk of preeclampsia. *Journal of*
90 *Clinical Endocrinology Metabolism* **92**, 3517–3522.
91 Braekke K., Ueland P.M., Harsem N.K., Karlsen A.,
92 Blomhoff R. & Staff A.C. (2007) Homocysteine, cys-
93 teine, and related metabolites in maternal and fetal
94 plasma in preeclampsia. *Pediatric Research* **62**, 319–324.
95 de Bree A., van Dusseldorp M., Brouwer I.A., van het Hof
96 K.H. & Steegers-Theunissen R.P.M. (1997) Folate intake
97 in Europe: recommended, actual and desired intake.
98 *European Journal of Nutrition* **51**, 643–660.
99 Breyman C. (2002) Iron supplementation during preg-
100 nancy. *Fetal and Maternal Medicine Review* **13**, 1–29.
101 Brown A.S. & Susser E.S. (2008) Prenatal nutritional defi-
102 ciency and risk of adult schizophrenia. *Schizophrenia*
103 *Bulletin* **34**, 1054–1063.

- 1 Buckley A.J., Jaquiere A.L. & Harding J.E. (2005) Nutri-
2 tional programming of adult disease. *Cell and Tissue*
3 **322**, 73–79. [30]
- 4 Camargo C.A. Jr, Rifas-Shiman S.L., Litonjua A.A., Liton-
5 jua A.A., Rich-Edwards J.W., Weiss S.T. *et al.* (2007)
6 Maternal intake of vitamin D during pregnancy and risk
7 of recurrent wheeze in children at 3 y of age. *American*
8 *Journal of Clinical Nutrition* **85**, 788–795.
- 9 Carlin A. & Alfirevic Z. (2008) Physiological changes of
10 pregnancy and monitoring. *Best Practice & Research*
11 *Clinical Obstetrics and Gynaecology* **22**, 801–823.
- 12 Catalano P.M., Nizielski S.E., Shao J., Preston L., Qiao L.
13 & Friedman J.E. (2002) Downregulated IRS-1 and
14 PPAR γ in obese women with gestational diabetes: rela-
15 tionship to FFA during pregnancy. *American Journal of*
16 *Physiology. Endocrinology and Metabolism* **282**, E522–
17 E533.
- 18 Cetin I., Alvino G., Radaelli T. & Pardi G. (2005) Fetal
19 nutrition: a review. *Acta Paediatrica* **94**, 7S–13S.
- 20 Cetin I., Berti C. & Calabrese S. (2009) Role of micronu-
21 trients in the periconceptual period. *Human Repro-*
22 *duction Update* **••**, ••–••. doi:10.1093/humupd/dmp025. [31]
- 23 Clifton-Bligh R.J., McElduff P. & McElduff A. (2008)
24 Maternal vitamin D deficiency, ethnicity and gestational
25 diabetes. *Diabetic Medicine* **25**, 678–684.
- 26 Cogswell M.E., Parvanta I., Ickes L., Yip R. & Brittenham
27 G.M. (2003) Iron supplementation during pregnancy,
28 anemia, and birth weight: a randomized controlled
29 trial. *American Journal of Clinical Nutrition* **78**,
30 773–781.
- 31 COMA (1991) Panel on DRVs of the Committee on
32 Medical Aspects of Food Policy. *Dietary reference values*
33 *(DRVs) for food energy and nutrients for the UK*,
34 Report on Health and Social Subjects 41.
- 35 Conde-Agudelo A. & Belizán J.M. (2000) Maternal mor-
36 bidity and mortality associated with interpregnancy
37 interval: cross sectional study. *British Medical Journal*
38 **321**, 1255–1259.
- 39 Coutinho R., David R.J. & Collins J.W. (1997) Relation of
40 prenatal birth weights to infant birth weight among
41 African Americans and whites in Illinois: a transgenera-
42 tional study. *American Journal of Epidemiology* **146**,
43 804–809. [32]
- 44 Czeizel A.E. & Dudás I. (1992) Prevention of the first
45 occurrence of neural-tube defects by periconceptual
46 vitamin supplementation. *The New England Journal of*
47 *Medicine* **327**, 1832–1835.
- 48 Czeizel A.E., Tímár L. & Sárközi A. (1999) Dose-
49 dependent effect of folic acid on the prevention of
50 orofacial clefts. *Pediatrics* **104**, e66.
- 51 D-A-CH (2000) German Nutrition Society, Austrian Nutri-
52 tion Society, Swiss Society for Nutrition Research. *Refer-*
53 *ence Values for Nutrient Intake (D-A-CH)*. Frankfurt am
54 Main.
- Darmon N. & Drewnowski A. (2008) Does social class
55 predict diet quality? *American Journal of Clinical Nutri-*
56 *tion* **87**, 1107–1117. 57
- De Bo H. & Harding J.E. (2006) The development origins
58 of adult disease (Barker) hypothesis. *Australian New*
59 *Zealand Journal of Obstetrics and Gynaecology* **46**, 4–14. 60
- Desoye G. & Hauguel-De Mouzon S. (2007) The human
61 placenta in gestational diabetes mellitus. The insulin and
62 cytokine network. *Diabetes Care* **30**, S120–S126. 63
- Devereux G., Litonjua A.A., Turner S.W., Craig L.C.A.,
64 McNeill G., Martindale S. *et al.* (2007) Maternal vitamin
65 D intake during pregnancy and early childhood wheez-
66 ing. *American Journal of Clinical Nutrition* **85**, 853–859. 67
- Dewey K.G. & Cohen R.J. (2007) Does birth spacing
68 affect maternal or child nutritional status? A systematic
69 literature review. *Maternal & Child Nutrition* **3**, 151–173. 70
- Díaz J.R., de las Cagigas A. & Rodríguez R. (2003) Micro-
71 nutrient deficiencies in developing and affluent coun-
72 tries. *European Journal of Nutrition* **57**, S70–S72. [33] 73
- Dodds L., Fell D.B., Dooley K.C., Armson B.A., Allen
74 A.C., Nassar B.A. *et al.* (2008) Effect of homocysteine
75 concentration in early pregnancy on gestational hyper-
76 tensive disorders and other pregnancy outcomes.
77 *Clinical Chemistry* **54**, 326–334. 78
- Doets E.L., de Wit L.S., Dhonukshe-Rutten R.A.M.,
79 Cavelaars A.E.J.M., Raats M.R., Timotijevic L. *et al.*
80 (2008) Current micronutrient recommendations in
81 Europe: towards understanding their differences and
82 similarities. *European Journal of Nutrition* **47**, S17–S40. 83
- Drewnowski A. & Darmon N. (2005) Food choices and
84 diet costs: an economic analysis. *Journal of Nutrition*
85 **135**, 900–904. 86
- Eilander A., Hundscheid D.C., Osendarp S.J., Transler C.
87 & Zock P.L. (2007) Effects of n-3 long chain polyunsatu-
88 rated fatty acid supplementation on visual and cognitive
89 development throughout childhood: A review of human
90 studies. *Prostaglandins, Leukotrienes and Essential Fatty*
91 *Acids* **76**, 189–203. 92
- Fleming T.P., Kwong W.Y., Porter R., Ursell E., Fesenko I.,
93 Wilkins A. *et al.* (2004) The embryo and its future.
94 *Biology of Reproduction* 1046–1054. 95
- Forges T., Monnier-Barbarino P., Alberto J.M., Guéant-
96 Rodriguez R.M., Daval J.L. & Guéant J.L. (2007)
97 Impact of folate and homocysteine metabolism on
98 human reproductive health. *Human Reproduction*
99 *Update* **••**, ••–••. [34] 100
- Gardiner H.M. (2007) Early environmental influences on
101 vascular development. *Early Human Development* **83**,
102 819–823. 103
- Gibson R., Perlas L. & Hotz C. (2006) Improving the bio-
104 availability of nutrients in plant foods at the household
105 level. *Proceedings of the Nutrition Society* **65**, 160–168. 106
- Gluckman P.D. & Pinal C.S. (2002) Maternal-placental-
107 fetal interactions in the endocrine regulation of fetal
108

- 1 growth. Role of Somatotrophic Axes. *Endocrine* **19**,
2 81–89. 55
- 3 35 Gluckman P.D., Hanson M.A. & Pinal C.S. (2005) The
4 developmental origins of adult disease. *Maternal &*
5 *Child Nutrition* 130–141. 56
- 6 Gluckman P.D., Hanson M.A., Cooper C. & Thornburg
7 K.L. (2008) Effect of in utero and early-life conditions
8 on adult health and disease. *The New England Journal*
9 *of Medicine* 61–73. 57
- 10 Goddijn-Wessel T.A.W., Wouters M.G.A.J., vd Molen E.F.,
11 Spuijbroek M.D.E.H., Steegers-Theunissen R.P.M., Blom
12 H.J. *et al.* (1996) Hyperhomocysteinemia: a risk factor
13 for placental abruption or infarction. *European Journal*
14 *of Obstetrics & Gynecology and Reproductive Biology*
15 **66**, 23–29. 58
- 16 Guéant J-L., Guéant-Rodriguez R-M., Anello G., Bosco P.,
17 Brunaud L., Romano C. *et al.* (2003) Genetic determi-
18 nants of folate and vitamin B12 metabolism: a common
19 pathway in neural tube defect and down syndrome?
20 *Clinical Chemistry and Laboratory Medicine* **41**, 1473–
21 1477. 59
- 22 36 Hall Moran V. (2007a) Nutritional status in pregnant ado-
23 lescents: a systematic review of biochemical markers.
24 *Maternal & Child Nutrition* **3**, 74–93. 60
- 25 Hall Moran V. (2007b) A systematic review of dietary
26 assessments of pregnant adolescents in industrialised
27 countries. *British Journal of Nutrition* **97**, 411–425. 61
- 28 Hauguel-de Mouzon S., Lepercq J. & Catalano P. (2006)
29 The known and unknown of leptin in pregnancy.
30 *American Journal of Obstetrics and Gynecology* **194**,
31 1537–1545. 62
- 32 Herrera E. (2000) Metabolic adaptations in pregnancy and
33 their implications for the availability of substrates to the
34 fetus. *European Journal of Clinical Nutrition* **54**, S47–
35 S51. 63
- 36 Hindmarsh P.C., Geary M.P.P., Rodeck C.H., Jackson M.R.
37 & Kingdom J.C.P. (2000) Effect of early maternal iron
38 stores on placental weight and structure. *Lancet* **356**,
39 719–723. 64
- 40 Hoey L., Strain J.J. & McNulty H. (2009) Studies of
41 biomarker responses to intervention with vitamin B-12:
42 a systematic review of randomized controlled trials.
43 *American Journal of Clinical Nutrition* **89**,
44 1981S–1996S. 65
- 45 van het Hof K.H., West C.E., Weststrate J.A. & Hautvast
46 J.G.A. (2000) Dietary factors that affect the bioavailabil-
47 ity of carotenoids. *Journal of Nutrition* **130**, 503–506. 66
- 48 Hollis B.W. & Wagner C.L. (2004) Assessment of dietary
49 vitamin D requirements during pregnancy and lactation.
50 *American Journal of Clinical Nutrition* **79**, 717–726. 67
- 51 Horvath A., Koletzko B. & Szajewska H. (2007) Effect of
52 supplementation of women in high-risk pregnancies with
53 long-chain polyunsaturated fatty acids on pregnancy
54 outcomes and growth measures at birth: a meta-analysis
of randomized controlled trials. *British Journal of Nutri-
tion* **98**, 253–259. 68
- Hotz C. & Gibson R.S. (2007) Traditional food-processing
and preparation practices to enhance the bioavailability
of micronutrients in plant-based diets. *Journal of Nutri-
tion* **137**, 1097–1100. 69
- Innis S.M. (2003) Perinatal biochemistry and physiology of
long-chain polyunsaturated fatty acids. *Journal of Pediat-
rics* **143**, S1–S8. 70
- Innis S.M. (2007) Fatty acids and early human develop-
ment. *Early Human Development* **83**, 761–766. 71
- Innis S.M. & Friesen R.W. (2008) Essential n-3 fatty acids
in pregnant women and early visual acuity maturation in
term infants. *American Journal of Clinical Nutrition* **87**,
548–557. 72
- Insel B.J., Schaefer C.A., McKeague I.W., Susser E.S. &
Brown A.S. (2008) Maternal iron deficiency and the risk
of schizophrenia in offspring. *Archives of General Psy-
chiatry* **65**, 1136–1144. 73
- Jauniaux E., Poston L. & Burton G.J. (2006) Placenta-
related diseases of pregnancy: involvement of oxidative
stress and implication in human evolution. *Human*
Reproduction Update **12**, 747–755. 74
- Javaid M.K., Crozier S.R., Harvey N.C., Taylor P., Inskip
H.M., Godfrey K.M. *et al.* (2006) Maternal vitamin D
status during pregnancy and childhood bone mass
at age 9 years: a longitudinal study. *Lancet* **367**,
36–43. 75
- Judge M.P., Harel O. & Lammi-Keefe C.J. (2007) Maternal
consumption of a docosahexaenoic acid-containing func-
tional food during pregnancy: benefit for infant perfor-
mance on problem-solving but not on recognition
memory tasks at age 9 mo. *American Journal of Clinical*
Nutrition **85**, 1572–1577. 76
- Kalhan S.C. (2000) Protein metabolism in pregnancy.
American Journal of Clinical Nutrition **71**,
S1249–S1255. 77
- Keen C.L., Clegg M.S., Hanna L.A., Lanoue L., Rogers
J.M., Daston G.P. *et al.* (2003) The plausibility of micro-
nutrient deficiencies being a significant contributing
factor to the occurrence of pregnancy complications.
Journal of Nutrition **133**, 1597S–1605S. 78
- King J.C. (2000) Physiology of pregnancy and nutrient
metabolism. *American Journal of Clinical Nutrition* **71**,
S1218–S1225. 79
- King J.C. (2003) The risk of maternal nutritional depletion
and poor outcomes increases in early or closely
spaced pregnancies. *Journal of Nutrition* **133**, 1732S–
1736S. 80
- Koebnick C., Hoffmann I., Dagnelie P.C., Heins U.A.,
Wickramasinghe S.N., Ratnayaka I.D. *et al.* (2004) Long-
term ovo-lacto. vegetarian diet impairs vitamin B-12
status in pregnant women. *Journal of Nutrition* **134**,
3319–3326. 81
- 37 82

- 1 Koletzko B., Aggett P.J., Bindels J.G., Bung P., Ferré P., Gif
2 A. *et al.* (1998) Growth, development and differentia-
3 tion: a functional food science approach. *British Journal*
4 *of Nutrition* **80**, S5–S45.
- 5 Koletzko B., Lien E., Agostoni C., Böhles H., Campoy C.,
6 Cetin I. *et al.* (2008) The roles of long-chain polyunsatu-
7 rated fatty acids in pregnancy, lactation and infancy:
8 review of current knowledge and consensus recommen-
9 dations. *Journal of Perinatal Medicine* **36**, 5–14.
- 10 Kovacs C.S. (2008) Vitamin D in pregnancy and lactation:
11 maternal, fetal, and neonatal outcomes from human and
12 animal studies. *American Journal of Clinical Nutrition*
13 **88**, S520–S528.
- 14 Ladipo O.A. (2000) Nutrition in pregnancy: mineral and
15 vitamin supplements. *American Journal of Clinical*
16 *Nutrition* **72**, 280S–290S.
- 17 Lain K.Y. & Catalano P.M. (2007) Metabolic changes in
18 pregnancy. *Clinical Obstetrics and Gynecology* **50**, 938–
19 948.
- 20 LARN (1996) *Livelli di Assunzione Raccomandati di*
21 *Energia e Nutrienti per la Popolazione Italiana. Revision.*
22 *Società Italiana di Nutrizione Umana (SINU).*
- 23 Lenders C.M., McElrath T.F. & Scholl M.T. (2000) Nutri-
24 tion in adolescent. *Current Opinion in Pediatrics* **12**,
25 291–296.
- 26 Locksmith G.J. & Duff P. (1998) Preventing Neural Tube
27 Defects: the importance of periconceptional folic acid
28 supplements. *Obstetrics and Gynecology* **91**, 1027–1034.
- 29 Lowe N.M., Fekete K. & Decsi T. (2009) ••. *American*
30 *Journal of Clinical Nutrition* **89**, 2040S–2051S. [38]
- 31 Lukas W.D. & Campbell B.C. (2000) Evolutionary and
32 ecological aspects of early brain malnutrition in humans.
33 *Human Nature* **11**, 1–26.
- 34 Lumey L.H. & Stein A.D. (1997) Offspring birth weights
35 after maternal intrauterine undernutrition: a comparison
36 within sibships. *American Journal of Epidemiology* **146**,
37 810–819. [39]
- 38 Maghbooli Z., Hossein-Nezhad A., Karimi F., Shafaei A.R.
39 & Larijani B. (2008) Correlation between vitamin D3
40 deficiency and insulin resistance in pregnancy.
41 *Diabetes/Metabolism Research and Reviews* **24**, 27–32.
- 42 Makrides M., Crowther C.A., Gibso R.A., Gibson R.S. &
43 Skeaff C.M. (2003) Efficacy and tolerability of low-dose
44 iron supplements during pregnancy: a randomized con-
45 trolled trial. *American Journal of Clinical Nutrition* **78**,
46 145–153.
- 47 McCaffery P.J., Adams J., Maden M. & Rosa-Molinar E.
48 (2003) Too much of a good thing: retinoic acid as an
49 endogenous regulator of neural differentiation and
50 exogenous teratogen. *European Journal of Neuroscience*
51 **18**, 457–472.
- 52 McCall K.A., Huang C.-C. & Fierke C.A. (2000) Function
53 and mechanism of zinc metalloenzymes. *Journal of*
54 *Nutrition* **130**, 1437S–1446S.
- 55 McMillen C. & Robinson J.S. (2005) Developmental
56 origins of the metabolic syndrome: prediction, plasticity,
57 and programming. *Physiological Reviews* **85**,
58 571–633.
- 59 McNulty H. & Pentieva K. (2006) Folate bioavailability.
60 *Proceedings of the Nutrition Society* **63**, 529–536.
- 61 McNulty H. & Scott J.M. (2008) Intake and status of folate
62 and related B-vitamins: considerations and challenges in
63 achieving optimal status. Intake and status of folate and
64 related B-vitamins: considerations and challenges in
65 achieving optimal status. *British Journal of Nutrition* **99**,
66 S48–S54.
- 67 Merialdi M., Caulfield L.E., Zavaleta N., Figueroa A.,
68 Costigan K.A., Dominici F. *et al.* (2004) Randomized
69 controlled trial of prenatal zinc supplementation and
70 fetal bone growth. *American Journal of Clinical Nutri-*
71 *tion* **79**, 826–830. [40]
- 72 Mikkelsen T.B., Østerdal M.L., Knudsen V.K., Haugen M.,
73 Meltzer H.M., Bakketeig L. *et al.* (2008) Association
74 between a Mediterranean-type diet and risk of preterm
75 birth among Danish women: a prospective cohort study.
76 *Acta Obstetrica et Gynecologica Scandinavica* **87**, 325–
77 330.
- 78 Milman N. (2006) Iron and pregnancy – a delicate balance.
79 *Annals of Hematology* **85**, 559–565.
- 80 Milman N., Bergholt T., Eriksen L., Byg K.-E., Graudal N.,
81 Pedersen P. *et al.* (2005) Iron prophylaxis during preg-
82 nancy – How much iron is needed? A randomized dose-
83 response study of 20–80 mg ferrous iron daily in
84 pregnant women. *Acta Obstetrica et Gynecologica Scan-*
85 *dinavica* **84**, 238–247.
- 86 Moreiras O., Carbajal A., Cabrera L. & Cuadrado C.
87 (2007) Ingestas recomendadas de energía y nutrientes
88 para la población española. In: *Tablas de composición de*
89 *alimentos, 11ª edición revisada y ampliada* (eds •• •• &
90 •• ••), pp. 227–230. Ediciones Pirámide/Grupo Anaya:
91 SA. [41]
- 92 Mouratidou T., Ford F., Prountzou F. & Fraser R. (2006)
93 Dietary assessment of a population of pregnant women
94 in Sheffield, UK. *The British Journal of Nutrition* 929–
95 935.
- 96 MRC Vitamin Study Research Group (1991) Prevention of
97 neural tube defects: results of the Medical Research
98 Council Vitamin Study. *Lancet* **338**, 131–137.
- 99 Mullis P.-E. & Tonella P. (2008) Regulation of fetal growth:
100 consequences and impact of being born small. *Best Prac-*
101 *tice & Research Clinical Endocrinology & Metabolism*
102 **22**, 173–190.
- 103 Munro N.B. & Eckerman K.F. (1998) Impacts of physi-
104 ological changes during pregnancy on maternal bioki-
105 netic modelling. *Radiation Protection Dosimetry* **79**,
106 327–333.
- 107 Newnham J.P., Moss T.J.M., Nitsos I., Sloboda D.M. &
108 Challis J.R.G. (2002) Nutrition and the early origins of

- 1 adult disease. *Asia Pacific Journal of Clinical Nutrition*
2 **11**, S537–S542.
- 3 NNR (Nordic Nutrition Recommendations) (2004) *Inte-*
4 *grating nutrition and physical activity*, 4th edn. Nordic
5 Council of Ministers: Copenhagen.
- 6 Oken E., Ning Y., Rifas-Shiman S.L., Rich-Edwards J.W.,
7 Olsen S.F. & Gillman M.W. (2007) Diet during preg-
8 nancy and risk of preeclampsia or gestational hyperten-
9 sion. *Annals of Epidemiology* **17**, 663–668.
- 10 Ounsted M., Scott A. & Ounsted C. (2008) Transmission
11 through the female line of a mechanism constraining
12 human fetal growth. *International Journal of Epidemiol-*
13 **42** *ogy* **37**, 245–250.
- 14 Pardi G. & Cetin I. (2006) Human fetal growth and organ
15 development: 50 years of discoveries. *American Journal*
16 *of Obstetrics and Gynecology* **194**, 1088–1099.
- 17 Pelto G. (1987) Cultural issues in maternal and child
18 health and nutrition. *Social Science and Medicine* **25**,
19 553–559.
- 20 Picciano M.F. (2003) Pregnancy and lactation: physiologi-
21 cal adjustments, nutritional requirements and the role of
22 dietary supplements. *Journal of Nutrition* **133**, 1997S–
23 2002S.
- 24 Pijls L., Ashwell M. & Lambert J. (2009) EURRECA – a
25 Network of Excellence to align European micronutrient
26 recommendations. *Food Chemistry* **113**, 748–753.
- 27 Pitkin R.M. (2007) Folate and neural tube defects.
28 **43** *American Journal of Clinical Nutrition* **85**, 285S–288S.
- 29 Popkin B.M. (2003) The nutrition transition in developing
30 world. *Development Policy Review* **21**, 581–597.
- 31 Population Reference Bureau (2000) *The World's Youth*
32 *2000*. Population Reference Bureau: Washington DC.
33 Available at: [http://www.phishare.org/files/](http://www.phishare.org/files/249_WorldsYouth_Eng.pdf#xml=http://www.phishare.org/cgi-bin/textis/webinator/elibsearch/xml.txt?query=world+youth&pr=phishare&order=r&cq=&id=4446a9d06)
34 [249_WorldsYouth_Eng.pdf#xml=http://www.](http://www.phishare.org/cgi-bin/textis/webinator/elibsearch/xml.txt?query=world+youth&pr=phishare&order=r&cq=&id=4446a9d06)
35 [phishare.org/cgi-bin/textis/webinator/elibsearch/](http://www.phishare.org/cgi-bin/textis/webinator/elibsearch/xml.txt?query=world+youth&pr=phishare&order=r&cq=&id=4446a9d06)
36 [xml.txt?query=world+youth&pr=phishare&order=r&](http://www.phishare.org/cgi-bin/textis/webinator/elibsearch/xml.txt?query=world+youth&pr=phishare&order=r&cq=&id=4446a9d06)
37 **44** [cq=&id=4446a9d06](http://www.phishare.org/cgi-bin/textis/webinator/elibsearch/xml.txt?query=world+youth&pr=phishare&order=r&cq=&id=4446a9d06)
- 38 Prentice A. (1994) Maternal calcium requirements during
39 pregnancy and lactation. *American Journal of Clinical*
40 *Nutrition* **59**, 477S–483S.
- 41 Ravelli A.C.J., van der Meulen J.H.P., Michels R.P.J.,
42 Osmond C., Barker D.J.P., Hales C.N. *et al.* (1998)
43 Glucose tolerance in adults after prenatal exposure to
44 famine. *Lancet* **351**, 173–177.
- 45 Ravelli A.C.J., van der Meulen J.H.P., Osmond C. &
46 Barker D.J.P. & Bleker O.P. (1999) Obesity at the age of
47 50 y in men and women exposed to famine prenatally.
48 *American Journal of Clinical Nutrition* **70**, 811–816.
- 49 Ray J.G. & Laskin C.A. (1999) Folic acid and homocys-
50 t(e)ine metabolic defects and the risk of placental
51 abruption, pre-eclampsia and spontaneous pregnancy
52 loss: a systematic review. *Placenta* **20**, 519–529.
- 53 Rayman M.P., Bode P. & Redman C.W.G. (2003) Low sele-
54 nium status is associated with the occurrence of the
pregnancy disease preeclampsia in women from the
United Kingdom. *American Journal of Obstetrics and*
Gynecology **189**, 1343–1349.
- Redman C.W.G., Sacks G.P. & Sargent I.L. (1999) Preec-
lampsia: an excessive maternal inflammatory response to
pregnancy. *American Journal of Obstetrics and Gynecol-*
ogy **180**, 499–506.
- Ristic-Medic D., Piskackova Z., Hooper L., Ruprich J.,
Casgrain A., Ashton K. *et al.* (2009) Methods of assess-
ment of iodine status in humans: a systematic review.
American Journal of Clinical Nutrition **89**, 2052S–2069S.
- Roberts J.M., Balk J.L., Bodnar L.M., Belizán J.M., Bergely
E. & Martinez A. (2003) Nutrient involvement in preec-
lampsia. *Journal of Nutrition* **133**, 1684S–1692S.
- Ronnenberg A.G., Wood R.J., Wang X., Xing H., Chen C.,
Chen D. *et al.* (2004) Preconception hemoglobin and
ferritin concentrations are associated with pregnancy
outcome in a prospective cohort of Chinese women.
Journal of Nutrition **134**, 2586–2591.
- Roseboom T.J., van der Meulen J.H.P., Osmond C., Barker
D.J.P., Ravelli A.C.J., Schroeder-Tanka J.M. *et al.* (2000)
Coronary heart disease after prenatal exposure to the
Dutch famine, 1944–45. *Heart* **84**, 595–598.
- Roseboom T.J., van der Meulen J.H.P., Ravelli A.C.J.,
Osmond C., Barker D.J.P. & Bleker O.P. (2001) Effects
of prenatal exposure to the Dutch famine on adult
disease in later life: an overview. *Molecular and Cellular*
Endocrinology **185**, 93–98.
- Ross A.C. (2006) Vitamin A and carotenoids. In: *Modern*
Nutrition in Health and Disease (eds M.E. Shils, M.
Shike, A.C. Ross, B. Caballero & R.J. Cousins), 10th
edn, pp. 351–375. Lippincott Williams & Wilkins: ••, ••. **45**
- Ryan-Harshman M. & Aldoori W. (2008) Vitamin B12 and
health. *Canadian Family Physician* **54**, 536–541.
- Salle B.L., Delvin E.E., Lapillonne A., Bishop N.J. & Glo-
rioux F.H. (2000) Perinatal metabolism of vitamin D.
American Journal of Clinical Nutrition **71**, S1317–S1324.
- Schoen C. & Rosen T. (2009) Maternal and perinatal risks
for women over 44 – a review. *Maturitas* **64**, 109–113. **46**
- Scholl T.O. (2005) Iron status during pregnancy: setting the
stage for mother and infant. *American Journal of Clinical*
Nutrition **81**, 1218S–1222S.
- Scholl T.O. & Hediger M.L. (1994) Anemia and iron-
deficiency anemia: compilation of data on pregnancy
outcome. *American Journal of Clinical Nutrition* **59**,
492S–501S.
- Siega-Riz A.M., Bodnar L.M. & Savitz D.A. (2002) What
are pregnant women eating? Nutrient and food group
differences by race. *American Journal of Obstetrics and*
Gynecology **186**, 480–486. **47**
- Siega-Riz A.M., Herrmann T.S., Savitz D.A. & Thorp J.M.
(2001) Frequency of eating during pregnancy and its
effect on preterm delivery. *American Journal of Epide-*
miology **153**, 647–652.

- 1 Siega-Riz A.M., Hartzema A.G., Turnbull C., Thorp J.,
2 McDonald T. & Cogswell M.E. (2006) The effects of
3 prophylactic iron given in prenatal supplements on iron
4 status and birth outcomes: a randomized controlled trial.
5 *American Journal of Obstetrics and Gynecology* **194**,
6 512–519.
- 7 Sørensen H.J., Nielsen P.R., Pedersen C.B. & Mortensen
8 P.B. (2010) Association between prepartum maternal
9 iron deficiency and offspring risk of schizophrenia:
10 population-based cohort study with linkage of Danish
11 national registers. *Schizophrenia Bulletin* **●●**, ●●–●●.
12 **48** doi:10.1093/schbul/sbp167.
- 13 Steegers E.A.P. (2005) Begin at the beginning: some reflec-
14 tions on future periconceptional and obstetric care and
15 research in the Netherlands. *European Clinics in Obstet-
16 rics and Gynaecology* **1**, 203–214. **49**
- 17 Steen M.T., Boddie A.M., Fisher A.J., Macmahon W., Saxe
18 D., Sullivan K.M. *et al.* (1998) Neural-tube defects are
19 associated with low concentrations of cobalamin
20 (Vitamin B₁₂) in amniotic fluid. *Prenatal Diagnosis* **18**,
21 545–555.
- 22 Strobel M., Tinz J. & Biesalski H-K. (2007) The impor-
23 tance of b-carotene as a source of vitamin A with special
24 regard to pregnant and breastfeeding women. *European
25 Journal of Nutrition* **46**, I/1–I/20.
- 26 Sunde R.A., Paterson E., Evenson J.K., Barnes K.M.,
27 Lovegrove J.A. & Gordon M.H. (2008) Longitudinal
28 selenium status in healthy British adults: assessment
29 using biochemical and molecular biomarkers. *British
30 Journal of Nutrition* **99**, S37–S47.
- 31 Szajewska H., Horvath A. & Koletzko B. (2006) Effect of
32 n–3 long-chain polyunsaturated fatty acid supplementa-
33 tion of women with low-risk pregnancies on pregnancy
34 outcomes and growth measures at birth: a meta-analysis
35 of randomized controlled trials. *American Journal of
36 Clinical Nutrition* **83**, 1337–1344.
- 37 Tamura T. & Picciano M.F. (2006) Folate and human
38 reproduction. *American Journal of Clinical Nutrition* **83**,
39 993–1016.
- 40 Thomas M. & Weisman S.M. (2006) Calcium supplementa-
41 tion during pregnancy and lactation: effects on the
42 mother and the fetus. *American Journal of Obstetrics
43 and Gynecology* **194**, 937–945.
- 44 Vanderlelie J., Venardos K., Clifton V.L., Gude N.M.,
45 Clarke F.M. & Perkins A.V. (2005) Increased biological
46 oxidation and reduced anti-oxidant enzyme activity in
47 pre-eclamptic placentae. *Placenta* **26**, 53–58.
- Vujkovic M., Ocke M.C., van der Spek P.J., Yazdanpanah
N., Steegers E.A. & Steegers-Theunissen R.P.M. (2007)
Maternal Western dietary patterns and the risk of devel-
oping a cleft lip with or without a cleft palate. *Obstetrics
and Gynecology* **110**, 378–384.
- de Weerd S., Steegers E.A.P., Heinen M.M., van den
Eertwegh S., Vehof R.M.E.J. & Steegers-Theunissen
R.P.M. (2003b) Preconception nutritional intake and
lifestyle factors: first results of an explorative study.
*European Journal of Obstetrics & Gynecology and
Reproductive Biology* **111**, 167–172. **50**
- Weissgerber T.L. & Wolfe L.A. (2006) Physiological adap-
tation in early human pregnancy: adaptation to balance
maternal–fetal demands. *Applied Physiology, Nutrition,
and Metabolism* **31**, 1–11.
- West J.K.P., Katz J., Khattry S.K., LeClerq S.C., Pradhan
E.K., Shrestha S.R. *et al.* (1999) Double blind, cluster
randomised trial of low dose supplementation with
vitamin A or β -carotene on mortality related to preg-
nancy in Nepal. *British Medical Journal* **318**, 570–575.
- Williamson C.S. (2006) Nutrition in pregnancy. *Nutrition
Bulletin* **31**, 28–59.
- Wood R.J. & Ronnenberg A.G. (2006) Iron. In: *Modern
Nutrition in Health and Disease* (eds M.E. Shils,
M. Shike, A.C. Ross, B. Caballero & R.J. Cousins),
10th edn, pp. 248–270. Lippincott Williams & Wilkins:
●●, ●●. **51**
- World Health Organization (2004) Adolescent pregnancy.
Issues in adolescent health and development. *WHO Dis-
cussion Papers on Adolescence*. Available at: [http://
www.eurreca.org/folders/Research](http://www.eurreca.org/folders/Research) **52**
- World Health Organization and Food and Agriculture
Organization of the United Nations (2004) Folate and
folic acid. In: *Vitamin and Mineral Requirements in
Human Nutrition (FAO/WHO Report)*, 2nd edn, pp 289–
302. World Health Organization and Food and Agricul-
ture Organization of the United Nations: ●●. **53**
- Yu C.K.H., Sykes L., Sethi M., Teoh T.G. & Robinson S.
(2009) Vitamin D deficiency and supplementation
during pregnancy. *Clinical Endocrinology* **70**,
685–690.
- Zimmerman M.B. (2008) Methods to assess iron and
iodine status. *British Journal of Nutrition* **99**, S2–S9.
- Zimmermann M.B. (2009) Iodine deficiency in pregnancy
and the effects of maternal iodine supplementation on
the offspring: a review. *American Journal of Clinical
Nutrition* **89**, S668–S672.

Toppan Best-set Premedia Limited	
Journal Code: MCN	Proofreader: Mony
Article No: 269	Delivery date: 15 July 2010
Page Extent: 18	Copyeditor:

AUTHOR QUERY FORM

Dear Author,

During the preparation of your manuscript for publication, the questions listed below have arisen. Please attend to these matters and return this form with your proof.

Many thanks for your assistance.

Query References	Query	Remark
q1	AUTHOR: A running head short title was not supplied; please check if this one is suitable and, if not, please supply a short title of up to 55 characters that can be used instead.	
q2	AUTHOR: Please confirm that the corresponding author address is correct.	
q3	AUTHOR: The abbreviation list has been deleted because abbreviations are already defined in the text; please confirm that this is OK.	
q4	AUTHOR: A change has been made in the article title; please confirm that this is OK.	
q5	Please provide the city location of affiliation addresses †, ‡, § and **, and please provide the postal/zip codes for all affiliation addresses.	
q6	AUTHOR: European Union: Is this the correct definition of EU? Please change if incorrect.	
q7	AUTHOR: Please provide a Key Message section for this article.	
q8	AUTHOR: Hall Moran & Dykes 2009 has not been included in the Reference List, please supply full publication details.	
q9	AUTHOR: Please check this website address throughout the article and confirm that it is correct. (Please note that it is the responsibility of the author(s) to ensure that all URLs given in this article are correct and useable.)	
q10	AUTHOR: 'Publications were searched. . .': The meaning of this sentence is not clear; please confirm that this is correct.	
q11	AUTHOR: Goddijn-Wessel <i>et al.</i> 1996 has been changed to Goddijn-Wessel <i>et al.</i> 1996 so that this citation matches the Reference List. Please confirm that this is correct.	
q12	AUTHOR: Stroebel <i>et al.</i> 2007 has been changed to Strobel <i>et al.</i> 2007 throughout so that this citation matches the Reference List. Please confirm that this is correct.	
q13	AUTHOR: Shah and Rajalakshmi 1987 has not been included in the Reference List, please supply full publication details.	
q14	AUTHOR: Argawal <i>et al.</i> 2005 has been changed to Agarwal <i>et al.</i> 2005 so that this citation matches the Reference List. Please confirm that this is correct.	
q15	AUTHOR: 'Dietary antioxidants seem. . .': A change has been made in this sentence; please confirm that this is correct.	

q16	AUTHOR: Salle 2000 has been changed to Salle <i>et al.</i> 2000 so that this citation matches the Reference List. Please confirm that this is correct.	
q17	AUTHOR: Ronnengerg <i>et al.</i> 2004 has been changed to Ronnenberg <i>et al.</i> 2004 so that this citation matches the Reference List. Please confirm that this is correct.	
q18	AUTHOR: Beard <i>et al.</i> 2003 has been changed to Beard 2003 so that this citation matches the Reference List. Please confirm that this is correct.	
q19	AUTHOR: Cosgwell <i>et al.</i> 2003 has been changed to Cogswell <i>et al.</i> 2003 so that this citation matches the Reference List. Please confirm that this is correct.	
q20	AUTHOR: Innis 2008 has been changed to Innis & Friesen 2008 so that this citation matches the Reference List. Please confirm that this is correct.	
q21	AUTHOR: Mulismatun et al. 2001 has not been included in the Reference List, please supply full publication details.	
q22	AUTHOR: Ortega et al. 1999 has not been included in the Reference List, please supply full publication details.	
q23	AUTHOR: de Bo & Harding 2006 has been changed to De Bo & Harding 2006 so that this citation matches the Reference List. Please confirm that this is correct.	
q24	AUTHOR: Food and Agriculture Organization of the United Nations/World Health Organization: Is this the correct definition of FAO/WHO? Please change if incorrect.	
q25	AUTHOR: McNulty & Pentieva 2004 has been changed to McNulty & Pentieva 2006 so that this citation matches the Reference List. Please confirm that this is correct.	
q26	AUTHOR: Please check this website address and confirm that it is correct. (Please note that it is the responsibility of the author(s) to ensure that all URLs given in this article are correct and useable.)	
q27	AUTHOR: WHO 2007 has been changed to WHO 2004 so that this citation matches the Reference List. Please confirm that this is correct.	
q28	AUTHOR: Hall Moran <i>et al.</i> 2007a has been changed to Hall Moran 2007a so that this citation matches the Reference	
q29	AUTHOR: Ashwell, Lambert, Alles, Branca, Bucchini, Brzozowska <i>et al.</i> , 2008 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q30	AUTHOR: Please confirm that the article title is correct for Buckley <i>et al.</i> 2005.	
q31	AUTHOR: Please confirm that the year of publication is correct for Cetin <i>et al.</i> 2009. Also, If this reference has now been published in print, please add relevant volume/page information and update the year information both in the citation and in the Reference List, and remove the DOI information.	
q32	AUTHOR: Coutinho, David, Collins, 1997 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	

q33	AUTHOR: Díaz, de las Cagigas, Rodríguez, 2003 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q34	AUTHOR: Please provide the volume number and page range for Forges <i>et al.</i> 2007.	
q35	AUTHOR: Gluckman, Pinal, 2002 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q36	AUTHOR: Guéant J-L, Anello, Bosco, Brunaud, Romano <i>et al.</i> , 2003 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q37	AUTHOR: King, 2000 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q38	AUTHOR: Please provide the article title for Lowe <i>et al.</i> 2009.	
q39	AUTHOR: Lumey, Stein, 1997 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q40	AUTHOR: Merialdi, Caulfield, Zavaleta, Figueroa, Costigan, Dominici, Dipietro, 2004 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q41	AUTHOR: Please provide the Editors of the cited chapter for Moreiras <i>et al.</i> 2007.	
q42	AUTHOR: Ounsted, Scott, Ounsted, 2008 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q43	AUTHOR: Pitkin, 2007 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q44	AUTHOR: Please check this website address and confirm that it is correct. (Please note that it is the responsibility of the author(s) to ensure that all URLs given in this article are correct and useable.)	
q45	AUTHOR: Please provide the city location of the publisher for Ryan-Harshman & Aldoori 2008.	
q46	AUTHOR: Schoen, Rosen, 2009 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q47	AUTHOR: Siega-Riz, Bodnar, Savitz, 2002 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q48	AUTHOR: Please provide the volume number and page range for Sørensen <i>et al.</i> 2010 and remove the DOI information if it has been published in print now.	
q49	AUTHOR: Steegers, 2005 has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q50	AUTHOR: de Weerd, Steegers, Heinen, van den Eertwegh, Vehof, Steegers-Theunissen, 2003b has not been cited in the text. Please indicate where it should be cited; or delete from the Reference List.	
q51	AUTHOR: Please provide the city location of the publisher for Wood & Ronnenberg 2006.	

q52	AUTHOR: Please check this website address and confirm that it is correct. (Please note that it is the responsibility of the author(s) to ensure that all URLs given in this article are correct and useable.)	
q53	AUTHOR: Please provide the location of the publisher for World Health Organization and Food and Agriculture Organization of the United Nations 2004.	
q54	AUTHOR: Please spell out IUGR.	
q55	AUTHOR: Figure 1 is of poor quality labels and lines are blurry. Please check required artwork specifications at http://authorservices.wiley.com/submit_illust.asp?site=1	
q56	AUTHOR: Sunde <i>et al.</i> 2009 has been changed to Sunde <i>et al.</i> 2008 so that this citation matches the Reference List. Please confirm that this is correct.	
q57	AUTHOR: Fairweather-Tait 2008 has not been included in the Reference List, please supply full publication details.	
q58	AUTHOR: Figure 1 is of poor quality (Any labels and lines is blurry). Please check required artwork specifications at http://authorservices.wiley.com/submit_illust.asp?site=1	

MARKED PROOF

Please correct and return this set

Please use the proof correction marks shown below for all alterations and corrections. If you wish to return your proof by fax you should ensure that all amendments are written clearly in dark ink and are made well within the page margins.

<i>Instruction to printer</i>	<i>Textual mark</i>	<i>Marginal mark</i>
Leave unchanged	... under matter to remain	Ⓟ
Insert in text the matter indicated in the margin	∧	New matter followed by ∧ or ∧ [Ⓢ]
Delete	/ through single character, rule or underline or ┌───┐ through all characters to be deleted	Ⓞ or Ⓞ [Ⓢ]
Substitute character or substitute part of one or more word(s)	/ through letter or ┌───┐ through characters	new character / or new characters /
Change to italics	— under matter to be changed	↙
Change to capitals	≡ under matter to be changed	≡
Change to small capitals	≡ under matter to be changed	≡
Change to bold type	~ under matter to be changed	~
Change to bold italic	≈ under matter to be changed	≈
Change to lower case	Encircle matter to be changed	≡
Change italic to upright type	(As above)	⊕
Change bold to non-bold type	(As above)	⊖
Insert 'superior' character	/ through character or ∧ where required	Υ or Υ under character e.g. Υ or Υ
Insert 'inferior' character	(As above)	∧ over character e.g. ∧
Insert full stop	(As above)	⊙
Insert comma	(As above)	,
Insert single quotation marks	(As above)	ʹ or ʸ and/or ʹ or ʸ
Insert double quotation marks	(As above)	“ or ” and/or ” or ”
Insert hyphen	(As above)	⊥
Start new paragraph	┌	┌
No new paragraph	┐	┐
Transpose	┌┐	┌┐
Close up	linking ○ characters	○
Insert or substitute space between characters or words	/ through character or ∧ where required	Υ
Reduce space between characters or words		↑