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## REFERENCES

1. Lee JK, Kang S, Wang X, et al. HAP1 loss confers L-asparaginase resistance in ALL by downregulating the calpain-1-Bid-caspase-3/12 pathway. *Blood*. 2019;133(20):2222-2232.
2. Dobson SM, Waanders E, McLeod J, et al. Defining functional heterogeneity in acute lymphoblastic leukemia [abstract]. *Blood*. 2013;122(21). Abstract 1365.
3. Cooper SL, Brown PA. Treatment of pediatric acute lymphoblastic leukemia. *Pediatr Clin North Am*. 2015;62(1):61-73.
4. Conter V, Valsecchi MG, Parasole R, et al. Childhood high-risk acute lymphoblastic leukemia in first remission: results after chemotherapy or transplant from the AIEOP ALL 2000 study. *Blood*. 2014;123(10):1470-1478.
5. Leoni V, Biondi A. Tyrosine kinase inhibitors in BCR-ABL positive acute lymphoblastic leukemia. *Haematologica*. 2015;100(3):295-299.
6. Pieters R, Hunger SP, Boos J, et al. L-asparaginase treatment in acute lymphoblastic leukemia: a focus on Erwinia asparaginase. *Cancer*. 2011;117(2):238-249.
7. Su N, Pan YX, Zhou M, Harvey RC, Hunger SP, Kilberg MS. Correlation between asparaginase sensitivity and asparagine synthetase protein content, but not mRNA, in acute lymphoblastic leukemia cell lines. *Pediatr Blood Cancer*. 2008;50(2):274-279.
8. Zhu L, Song X, Tang J, et al. Huntingtin-associated protein 1: a potential biomarker of breast cancer. *Oncol Rep*. 2013;29(5):1881-1887.
9. Peng S, Gerasimenko JV, Tsigorka T, et al. Calcium and adenosine triphosphate control of cellular pathology: asparaginase-induced pancreatitis elicited via protease-activated receptor 2. *Philos Trans R Soc Lond B Biol Sci*. 2016;371(1700):20150423.

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## TRANSPLANTATION

Comment on Gagelmann et al, page 2233

# Stem cell transplant in MF: it's time to personalize

Francesco Passamonti | University of Insubria

**In this issue of *Blood*, Gagelmann et al<sup>1</sup> describe an integrated clinical-molecular prognostic model (Myelofibrosis Transplant Scoring System [MTSS]) to predict outcome post stem cell transplant (SCT) in myelofibrosis (MF). MF is a clonal stem cell neoplasm with heterogeneous clinical phenotypes and well-defined driver mutations found in ~90% of the cases.<sup>2</sup> Despite the introduction of JAK inhibitors,<sup>3</sup> MF still remains an incurable disease with a median survival of 4 to 5 years. SCT is an option for MF patients,<sup>4</sup> but, because of the very high risk of mortality, the patient selection is very critical.**

A next-generation sequencing (NGS)-based 18-gene panel was available in patients of the study. The transplant regimen was mainly reduced intensity in the training cohort and myeloablative in the validation cohort. Variables included in the model are age  $\geq 57$  years, Kamofsky performance status  $< 90\%$ , platelet count  $< 150 \times 10^9/L$ , leukocyte count  $> 25 \times 10^9/L$ , HLA-mismatched unrelated donor, presence of the ASXL1 mutation, and a non-CALR/MPL genotype (see figure). The resulting 5-year overall survival (OS) was 90% for low risk, 77% for intermediate risk,

50% for high risk, and 34% for very high risk.

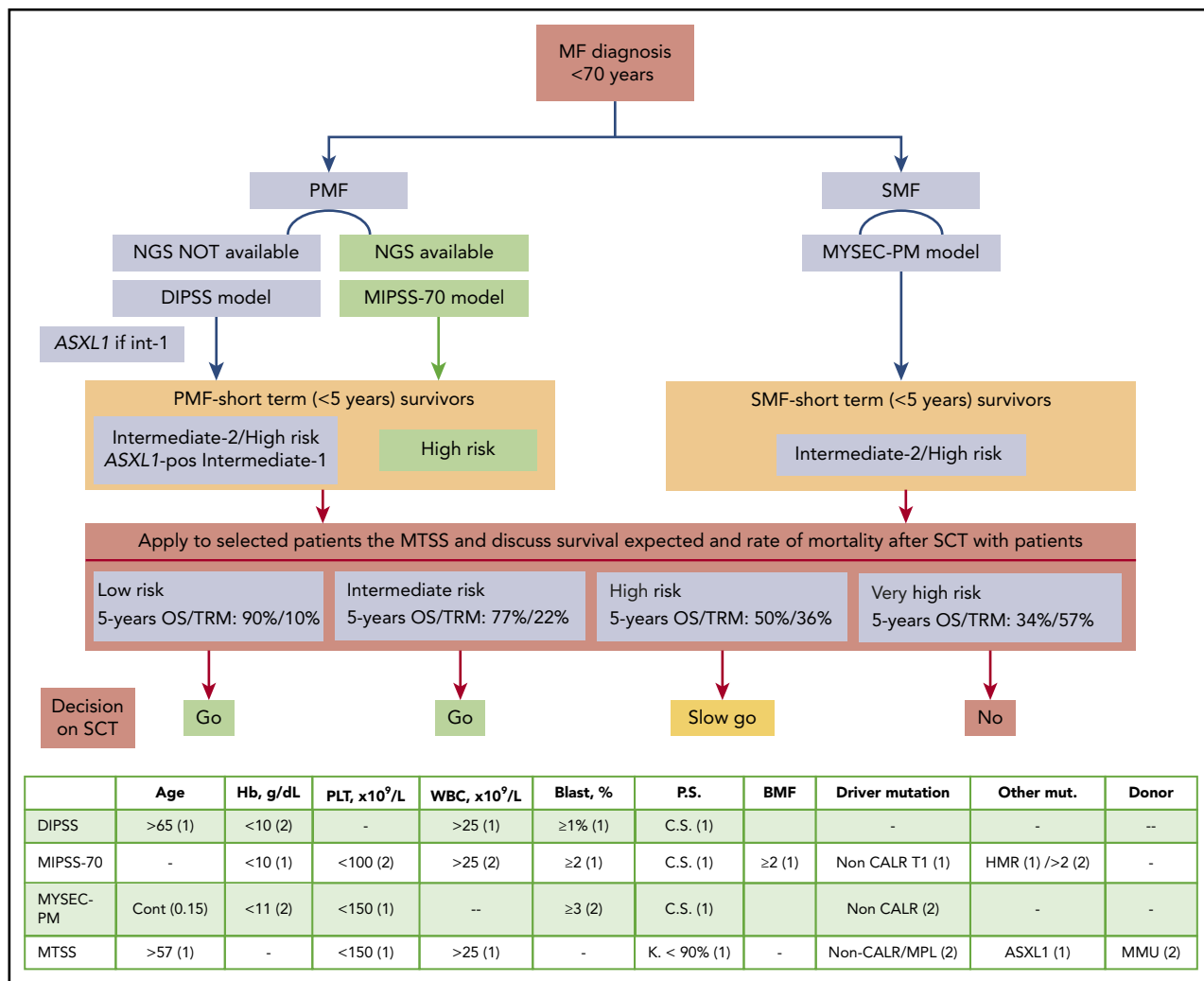
The 5-year survival of the whole cohort was 62%, higher than the 47% reported at the beginning in 1999 by Guardiola et al<sup>5</sup> from the European Bone Marrow Transplantation. The results of the study reported here may be impacted by the 9% of patients transplanted with low-risk disease, who would currently not be transplant candidates.<sup>6</sup> However, low-risk patients have been included in all SCT reports. Notably, the model is applicable in primary MF (PMF)

or in secondary MF (SMF), such as post-polycythemia vera and postessential thrombocythemia MF.

The MTSS will implement current risk models (see figure) with the aim of improving personalization of therapy in MF patients suitable for SCT (age  $< 70$  years). Some models include only clinical variables; other models include clinical and molecular, and others are based on karyotypes. Many MF patients have no analyzable metaphases, and about two-thirds of the cases have a normal karyotype,<sup>7,8</sup> making karyotype-based models vastly unpracticable. Each patient must receive a prognostic assessment at the time of diagnosis of MF (see figure), according to the MF type: PMF or SMF. For PMF, patients evaluated by NGS can be accessed using the MIPSS-70 model.<sup>9</sup> However, if NGS is not available, the DIPSS model still remains a robust and validated tool. In situations of limited NGS availability, at least ASXL1 mutation in intermediate-1 DIPSS-scored young patients should be determined.<sup>4</sup> In SMF, the MYSEC-PM<sup>10</sup> is recommended because it was specifically developed in SMF. Among MF patients, those with a median expectation of life  $< 5$  years are potential candidates for SCT, according to latest European LeukemiaNet recommendations.<sup>4</sup> This corresponds to ASXL1 mutated-intermediate-1/intermediate-2/high-risk DIPSS, high-risk MIPSS-70, and intermediate-2/high-risk MYSEC-PM patients (see figure).

At this juncture, MTSS becomes helpful. Low- and intermediate-risk MTSS has a clear indication for SCT, as the 5-year OS of 90% and 77%, respectively, is superior to other therapies, and this result balances the rate of nonrelapse mortality (NRM) of 10% to 22%. The following cases illustrate the difficulties in the management of high-risk and very-high-risk MTSS situations.

1. **Case 1:** A 68-year-old man with post polycythemia vera–MF (intermediate-2 MYSEC-PM). The projected life expectancy is 4.5 years (MYSEC-PM); hence, he is an SCT candidate. However, his MTSS category is high risk, resulting in a 5-year median survival of 50% and a transplant related mortality (TRM) of 36%. In this case, survival is equivalent with standard therapy or proceeding to SCT. It seems reasonable not to proceed to SCT. However,



Critical information to personalize management of patients with MF. (Top) Decision flowchart for each patient with MF suitable for SCT. (Bottom) All prognostic models in use with variables and score in parentheses for assessing risk category to each patient. Dynamic International Prognostic Scoring System (DIPSS) categories: low (0); intermediate-1 (1-2), intermediate-2 (3-4), high risk (5-6); Mutation-Enhanced International Prognostic Score System (MIPSS-70) categories: low (0-1); intermediate (2-4), high risk (≥5); Myelofibrosis Secondary Prognostic model (MYSEC-PM) categories: low (<11); intermediate-1 (11-13), intermediate-2 (14-15), high risk (≥16), <http://www.mysec-pm.eu>; MTSS categories: low (0-2); intermediate (3-4), high (5), very-high risk (6-9). BM, bone marrow; BMF, bone marrow fibrosis; Cont, continuous; C.S., constitutional symptoms; Hb, hemoglobin value; HMR, high-molecular risk; K., Karnofsky; MMU, mismatched unrelated; mut., mutation; PLT, platelet count; P.S., performance status; WBC, white blood cell count.

if the patient were younger (aged 40-60 years), it would be reasonable to proceed to SCT as the only therapy, with a survival plateau after 5 years.

- Case 2: A 60-year-old woman with PMF (NGS data available) with high-risk MIPSS-70 and an estimated 5-year survival of 29%. This patient has an MTSS score of 7, a very-high risk MTSS category. Her 5-year survival after SCT is 34%, with NRM of 57%. As survival is poor with either option, a clinical trial of novel therapy should be pursued.

In conclusion, prognostic models are of critical importance to personalize management

of MF. Currently, assessment of patients should include molecular evaluation with NGS or, at least, evaluation of ASXL1 mutation. SCT still remains a high-risk procedure in MF, but, after 5 years, 40% of the patients can be considered cured. The MTSS helps in individualizing the SCT decision in MF patients.

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#### REFERENCES

- Gagelmann N, Ditschkowski M, Bogdanov R, et al. Comprehensive clinical-molecular transplant scoring system for myelofibrosis undergoing stem cell transplantation. *Blood*. 2019;133(20):2233-2242.

- Passamonti F, Mora B, Maffioli M. New molecular genetics in the diagnosis and treatment of myeloproliferative neoplasms. *Curr Opin Hematol*. 2016;23(2):137-143.
- Passamonti F, Maffioli M. The role of JAK2 inhibitors in MPNs 7 years after approval. *Blood*. 2018;131(22):2426-2435.
- Barbui T, Tefferi A, Vannucchi AM, et al. Philadelphia chromosome-negative classical myeloproliferative neoplasms: revised management recommendations from European LeukemiaNet. *Leukemia*. 2018;32(5):1057-1069.
- Guardiola P, Anderson JE, Bandini G, et al. Allogeneic stem cell transplantation for agnogenic myeloid metaplasia: a European Group for Blood and Marrow Transplantation, Société Française de

- Greffe de Moelle, Gruppo Italiano per il Trapianto del Midollo Osseo, and Fred Hutchinson Cancer Research Center Collaborative Study. *Blood*. 1999;93(9):2831-2838.
6. Kröger N, Giorgino T, Scott BL, et al. Impact of allogeneic stem cell transplantation on survival of patients less than 65 years of age with primary myelofibrosis. *Blood*. 2015;125(21):3347-3350, quiz 3364.
7. Passamonti F, Cervantes F, Vannucchi AM, et al. A dynamic prognostic model to predict survival in primary myelofibrosis: a study by the IWG-MRT (International Working Group for Myeloproliferative Neoplasms Research and Treatment). *Blood*. 2010;115(9):1703-1708.
8. Mora B, Giorgino T, Guglielmelli P, et al. Value of cytogenetic abnormalities in post-polycythemia vera and post-essential thrombocythemia myelofibrosis: a study of the MYSEC project. *Haematologica*. 2018;103(9):e392-e394.
9. Guglielmelli P, Lasho TL, Rotunno G, et al. MIPSS70: mutation-enhanced international prognostic score system for transplantation-age patients with primary myelofibrosis. *J Clin Oncol*. 2018;36(4):310-318.
10. Passamonti F, Giorgino T, Mora B, et al. A clinical-molecular prognostic model to predict survival in patients with post polycythemia vera and post essential thrombocythemia myelofibrosis. *Leukemia*. 2017;31(12):2726-2731.

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