

Clonal Hematopoiesis of Indeterminate Potential in Patients with Immunoglobulin Light Chain AL Amyloidosis

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Abstract:

Immunoglobulin light chain (AL) amyloidosis is characterized by the deposition of misfolded monoclonal free light chains, with cardiac complications accounting for patients' mortality. Clonal hematopoiesis of indeterminate potential (CHIP) has been associated with worse cardiovascular outcomes in the general population. Its significance in AL amyloidosis remains unclear. We collected clinical information and outcome data on 76 patients with a diagnosis of AL amyloidosis who underwent deep-targeted sequencing for myeloid neoplasia-associated mutations between April 2018 and August 2023. Variant allele fraction was set at 2% to call CHIP-associated mutations. CHIP mutations were present in AL amyloidosis patients at a higher frequency than age-matched control individuals. Sixteen patients (21%) had at least 1 CHIP mutation. DNMT3A was the most frequent mutation (7/16, 44%). Compared to patients without CHIP, patients with CHIP were enriched for the presence of t(11;14) (69% vs 25%, respectively, $p = 0.004$) and, for patients with renal involvement, a lower Palladini renal stage ($p = 0.001$). At a median follow-up of 32.5 months, the presence of CHIP was not associated with worse overall survival or major organ dysfunction progression-free survival. Larger studies and longer follow-up are needed to better define the impact of CHIP in patients with AL amyloidosis.

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1 **Clonal Hematopoiesis of Indeterminate Potential in Patients with Immunoglobulin Light**
2 **Chain AL Amyloidosis**

3 **CHIP in AL Amyloidosis**

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30 **Key Points**

- 31 • CHIP is more prevalent in AL amyloidosis patients than in the general population
- 32 • CHIP presence was associated with the presence of t(11;14), and a lower Palladini renal
33 stage in patients with renal involvement

34

35 **Abstract**

36 Immunoglobulin light chain (AL) amyloidosis is characterized by the deposition of misfolded
37 monoclonal free light chains, with cardiac complications accounting for patients' mortality.
38 Clonal hematopoiesis of indeterminate potential (CHIP) has been associated with worse
39 cardiovascular outcomes in the general population. Its significance in AL amyloidosis remains
40 unclear. We collected clinical information and outcome data on 76 patients with a diagnosis of
41 AL amyloidosis who underwent deep-targeted sequencing for myeloid neoplasia-associated
42 mutations between April 2018 and August 2023. Variant allele fraction was set at 2% to call
43 CHIP-associated mutations. CHIP mutations were present in AL amyloidosis patients at a higher
44 frequency than age-matched control individuals. Sixteen patients (21%) had at least 1 CHIP
45 mutation. DNMT3A was the most frequent mutation (7/16, 44%). Compared to patients without
46 CHIP, patients with CHIP were enriched for the presence of t(11;14) (69% vs 25%, respectively,
47 $p = 0.004$) and, for patients with renal involvement, a lower Palladini renal stage ($p = 0.001$). At
48 a median follow-up of 32.5 months, the presence of CHIP was not associated with worse overall

49 survival or major organ dysfunction progression-free survival. Larger studies and longer follow-
50 up are needed to better define the impact of CHIP in patients with AL amyloidosis.

51 **Introduction**

52 Immunoglobulin light chain (AL) amyloidosis is a plasma cell (PC) dyscrasia whose hallmark is
53 the production and secretion of misfolded, monoclonal immunoglobulin free light chains
54 (FLC).^{1,2} The circulating FLC drives pathogenesis causing direct cytotoxicity in its soluble form
55 and disruption of target organ architecture upon deposition as insoluble fibrillary aggregates. It is
56 thought that any organ/system in the body can be involved by AL amyloidosis deposition with
57 the heart and kidneys being the most commonly affected organs.^{3,4} The severity of cardiac
58 involvement is the single most important prognostic factor in AL amyloidosis as cardiac events
59 represent the major cause of mortality.^{5,6} Renal involvement occurs in up to 70% of cases and is
60 a key determinant of patients morbidity, decreased quality of life, and ineligibility for clinical
61 trials.^{7,8} To aid in patients' prognostication and management, several staging systems have been
62 developed to evaluate cardiac involvement⁹⁻¹², while Palladini and colleagues have introduced a
63 score predicting the risk of progression to renal replacement therapy.⁷ Cytogenetic alterations
64 have also prognostic implications and are an important predictor of response to anti-neoplastic
65 therapies.¹³⁻¹⁶ Among those, t(11;14) is the most prevalent.¹⁶ It portends a lower response rate to
66 bortezomib-based therapies¹³ and is a negative prognostic factor, being associated with decreased
67 progression-free survival (PFS) compared to patients without any alteration.¹⁵ Based on the
68 recently published Andromeda study, daratumumab appears to overcome the negative prognostic
69 impact of t(11;14).⁸ Further, extrapolating from multiple myeloma, the presence of t(11;14) is a
70 biomarker for response to Bcl2 inhibitor venetoclax.^{4,8,17}

71 Clonal hematopoiesis of indeterminate potential (CHIP) refers to the presence of clonal, somatic
72 mutations of myeloid-related genes in the absence of overt myeloid neoplasia or cytopenia. The
73 most commonly CHIP-associated mutations involve DNMT3A, TET2, and/or ASXL1 genes,

74 commonly known under the acronym DTA.¹⁸ CHIP incidence increases with age, being most
75 frequent in the elderly population and nearly absent before the age of 40.^{19–23} Its occurrence has
76 been associated with a higher incidence of hematological malignancies^{19,20,22} and cardiovascular
77 disease, as well as worse cardiovascular outcomes^{22,24–27}. In the context of hematological
78 neoplasms, CHIP has been detected in 9.7 – 21.6% of patients with PC dyscrasia and 14 – 29%
79 with lymphoma.^{23,28–32} In patients with multiple myeloma (MM) and lymphoma receiving
80 autologous stem cell transplant (ASCT), CHIP presence was identified as an adverse prognostic
81 factor.^{28,31} Thari and colleagues also noted a higher risk of progression to Waldenström
82 macroglobulinemia (WM) in patients with IgM monoclonal gammopathy of undeterminate
83 significance or smoldering WM carrying DTA mutations.²⁹ Two studies previously reported on
84 the incidence of CHIP in patients with AL amyloidosis. The incidence of CHIP was 15% (4 out
85 of 27 patients) in one study and 21% (10 out of 47 patients) in the other and did not correlate
86 with any specific clinical features. It is important to note that while the presence of CHIP was not
87 found to have prognostic significance in these studies, the association between CHIP and
88 cytogenetic alterations or cardiac involvement was not investigated.^{32,33}

89 We were interested in exploring whether an association exists between CHIP and cardiac
90 outcome, based on prior studies. We were also interested in understanding the co-existence of
91 relevant disease characteristics with CHIP.

92 Hence, we performed a single-center, retrospective cohort study including 76 consecutive
93 patients with AL amyloidosis who were seen at BWH/DFCI between April 2018 and August
94 2023 and had a bone marrow biopsy performed with a targeted myeloid mutation panel assessed.

95 This is the largest study to date looking broadly at the prevalence and impact of the presence of
96 CHIP in AL amyloidosis patients.

97

98 **Materials and Methods**

99 Patients

100 We identified patients seen at BWH/DFCI for a diagnosis of AL amyloidosis who underwent
101 deep-targeted sequencing for myeloid neoplasia-associated mutations between April 2018 and
102 August 2023. We retrospectively collected clinical information, including age, gender, ethnicity,
103 smoking status, FLC subtype, European modification of 2004 Mayo stage, organ involvement,
104 cytogenetics alterations detected by fluorescent in-situ hybridization (FISH), left ventricular
105 ejection fraction, Palladini Renal stage for patients with renal involvement.^{7,9} Additionally, we
106 looked at the type of anti-neoplastic treatment, depth of hematological response, and whether or
107 not patients received an ASCT. We chose as a primary outcome the major-organ dysfunction
108 event-free survival (MOD-PFS) as defined by Kastiris and colleagues⁸ and the overall survival
109 (OS). We selected as a secondary outcome the cardiac-specific disease response and PFS
110 (assessed at 6 months and 12 months post-commencement of therapy), as defined by Palladini
111 and colleagues.⁷ Next-generation sequencing on bone marrow samples was performed using our
112 custom-validated assay, Rapid Heme Panel (RHP).³⁴ Genes assessed for CHIP attribution
113 included: JAK1, JAK3, PDGFRA, SFA3A1, DNMT3A, GNB1, CEBPA, SBDS, FLT3, KRAS,
114 BCORL1, PIGA, SF3B1, ASXL1, CTCF, CSF3R, CUX1, NOTCH3, PPM1D, ZRSR2, ATM,
115 CCND1, KMT2A, EP300, EZH2, SETD2, SH2B3, GNAS, GATA1, IKZF3, PRPF8, KIT,
116 NOTCH2, WT1, TET2, PIK3CA, PTPN11, CREBBP, NOTCH1, BRCC3, DDX41, TP53,
117 CALR, LUC7L2. The median coverage obtained was 759 reads per base (IQR 1131).

118 CHIP status was assigned to patient samples when putative driver lesions in genes associated
119 with myeloid neoplasms were observed at Variant Allele Frequency (VAF) higher than 2%.
120 Reported variants were then analyzed and filtered according to common practice standards
121 through a semi-automatic pipeline.³⁵⁻³⁷ The age-specific CHIP rates reported by Jaiswal et al.
122 have been used to calculate the standardized incidence rate.¹⁹

123 This study was approved by the BWH/DFCI Institutional Review Board with the approval
124 number 2023P001501 and was conducted in accordance with the Declaration of Helsinki.

125

126 Statistical Analysis

127 Statistical analyses were performed using Stata statistical software release 17 (StataCorp LLC,
128 College Station, TX) and R version 4.2.3 (*Shortstop Beagle*). Normal distribution was visually
129 assessed for all continuous variables. Data dispersion was assessed with standard deviation (SD)
130 for normally distributed variables and with interquartile range (IQR) for non-normally
131 distributed variables. Baseline demographics and disease characteristics were compared between
132 patients with and without CHIP. Comparison for normally-distributed variables was performed
133 with the independent samples t-test, while, for non-normally distributed, with the Wilcoxon rank
134 sum (Mann Whitney U) test. Comparison for categorical variables was performed with the Chi-
135 square test and the Fisher exact test. MOD-PFS and cardiac-specific PFS were measured from
136 the time of diagnosis to the time of the first MOD-PFS/cardiac-progression defining event or
137 were censored at the last follow-up. All reported p values were two-sided, with a statistical
138 significance set at <0.05. We used the Kaplan-Meier method to estimate the survival curves for
139 the OS and MOD-PFS, and the Log-rank test to assess the difference between survival curves.

140 We used a Cox regression and a stratified Cox regression model to assess the time-to-event
141 outcome and calculate hazard ratios (HR) with 95% confidence intervals (CI). The stratified
142 multivariate Cox regression model was built using a forward selection principle and following a
143 10:1 events to covariate ratio.

144 This study was approved by the BWH/DFCI Institutional Review Board with the approval
145 number 2023P001501.

146

147 **Results**

148 CHIP is present at a higher prevalence in AL amyloidosis patients as compared to a healthy
149 population

150 We identified a total of 76 patients. Sixteen patients (21%) had at least 1 CHIP mutation. Figure
151 1 shows the detected mutational profile. DNMT3A was the most frequently involved gene (7/16,
152 44%), followed by TET2, GNB1, ATM (each 2/16, 12.5%), and SF3B1, TP53, ZRSR2, EZH2,
153 BRCC3, PPM1D, ASXL1 (6%). DNMT3A variants included 6 missense (2 of which were in the
154 same patient) at the R882, R736, Y735, I780, and F755 residues, and 3 nonsense lesions. TET2
155 lesions included 3 stop codons and 1 variant in the catalytic domain (residues 1843-2002).
156 ASXL1, GNB1, and ATM-reported variants included known missense hotspots. Of note, 5
157 samples carried more than one CHIP-defining lesion. The median VAF of CHIP-associated
158 mutations was 0.036. A subset of 4 cases carried CHIP variants with an allele frequency equal to
159 or higher than 10%, suggesting the presence of a larger clone. Based on the age distribution of
160 our patients, the age-standardized incidence rate of CHIP would be 6%.

161 Ten patients (13%) had more than one sample available. Eight out of ten patients with more than
162 one bone marrow biopsy available had no evidence of CHIP on either biopsy. In contrast, the
163 other 2 patients (20%) were initially negative and subsequently had a biopsy positive for CHIP
164 (DNMT3A, ZRSR2). There were no patients with CHIP on a first biopsy who were CHIP
165 negative on a subsequent biopsy (Figure 2).

166

167 Clinical characteristics

168 Baseline demographics are shown in Table 1. The mean age of our cohort was 63 years (range 44
169 – 85). Thirty-two patients were females (42%), 6 patients (8%) were black, 2 (3%) were Asian, 1
170 was Middle-Eastern (1%), 2 (3%) self-reported as other, and 65 (85%) were white. No
171 significant difference was noted in epidemiologic characteristics between patients harboring a
172 CHIP mutation and those without ($p>0.05$). Sixty-nine percent of patients with CHIP and 67% of
173 patients without CHIP ($p = 1$) were treatment naïve at the time the RHP was obtained. Among
174 patients with CHIP, 2 (12.5%), 5 (31%), 7 (44%), and 2 (12.5%) had a Mayo stage of I, II, IIIA,
175 and IIIB, respectively. Among those without CHIP, 19 (32%), 13 (22%), 14 (24%), and 13
176 (24%), had a Mayo stage of I, II, IIIA, and IIIB, respectively. There was no statistically
177 significant difference in the partition of the Mayo stage between the two groups. When focusing
178 on patients with histopathology-proven or clinically determined renal involvement, patients with
179 CHIP were more likely to have a lower Palladini renal stage⁷ ($p = 0.001$). No significant
180 difference was noted regarding the frequency of organ involvement (i.e., heart, kidneys, liver,
181 lung, autonomic nervous system, peripheral nervous system, gastrointestinal tract, and soft
182 tissue) and the total number of organs affected. The left ventricular ejection fraction (LVEF) and
183 the presence of anginal symptoms were also evaluated: the mean LVEF was 55% (SD 6.4%) for

184 patients with CHIP and 53% (SD 10.2%) for those without. None of the CHIP patients had an
185 LVEF \leq 40% as opposed to 7 (12%) non-CHIP patients, but this difference was not statistically
186 significant ($p = 0.34$). Two patients with CHIP (13%) reported anginal symptoms, as opposed to
187 4 patients without CHIP (7%) but this difference was not statistically significant ($p = 0.60$). We
188 then assessed the cytogenetic profile of our cohort (Table 2). A total of 54 patients could be
189 evaluated with FISH, 13 with CHIP (81%), and 41 without CHIP (68%). Among those with
190 CHIP, 11/13 (85%) were found to harbor the t(11;14) as opposed to 15/41 (37%) without CHIP
191 ($p = 0.004$). To exclude a possible confounding effect of age on the association between CHIP
192 and the t(11;14) we performed a multivariate logistic regression to assess this association when
193 keeping age constant. Even after adjusting for age, the association between CHIP and t(11;14)
194 remained statistically significant (adjusted odds ratio 10.92, CI 1.91 – 62.31, $p = 0.007$). No
195 other cytogenic abnormality was found to be significantly associated with CHIP. In regards to the
196 treatment received, 35 (46%) patients received the combination cyclophosphamide, bortezomib,
197 dexamethasone (CyBorD), 28 (37%) CyBorD in association with Daratumumab (Dara-CyBorD),
198 while 12 (16%) other regimens (bortezomib – dexamethasone [VD], lenalidomide – bortezomib
199 – dexamethasone [RVD], Dara-VD, and melphalan - dexamethasone). Ten (13%) patients
200 received ASCT during their disease course, including 2 (12.5%) patients with CHIP and 8 (13%)
201 without.

202

203 The presence of CHIP does not impact MOD-PFS or OS

204 After a median follow-up from diagnosis of 32.5 months (range 0.5 – 168), 11 patients (14%)
205 died and 30 (39%) had a MOD-PFS defining event including death. The presence of CHIP was
206 not associated with lower OS ($p = 0.483$) (Figure 2A) or with lower MOD-PFS ($p = 0.815$)

207 (Figure 2B). In the univariate Cox proportional hazard model, variables associated with an
208 increased hazard of mortality were age (HR 1.12, 95% CI, 1.04 – 1.21, p = 0.003), a Mayo stage
209 >2 (HR 5.74, 95% CI 1.22 – 26.93, p= 0.027), LVEF ≤40% (4.11, 95% CI 1.09 – 15.53, p =
210 0.037) and a co-occurring diagnosis of coronary artery disease (CAD) (HR 4.94, 95% CI 1.31 –
211 18.63, p = 0.018). In this cohort, CHIP was not associated with an increased hazard for mortality.

212 We then looked at MOD-PFS, and the variables associated with an increased hazard for MOD-
213 defining events were Mayo stage 3a or 3b (HR 4.68, 95% CI 1.94 – 11.29, p = 0.001), an LVEF
214 ≤40% (HR 3.09, 95% CI 1.15 – 8.34, p = 0.026), a diagnosis of CAD (HR 5.43, 95% CI 2.13 –
215 13.88, p < 0.001), the presence of orthostatic hypotension requiring treatment (HR 2.35, 95% CI
216 1.08 – 5.10, p = 0.030). CHIP was not associated with an increased hazard for MOD events.

217 A multivariate Cox regression model stratified for age (44-54, 55-64, 65-71, >71) for MOD-PFS
218 was constructed, which included CAD and Mayo stage 3a or 3b, in addition to CHIP status.
219 Mayo stage 3a or 3b (HR 4.10, 95% CI 1.55 – 10.81, p = 0.004) and the presence of CAD (HR
220 4.89, 95% CI 1.58 – 15.16, p = 0.006) were associated with an increased hazard for MOD-
221 defining event (Figure 3). CHIP was not associated with an increased hazard for MOD-defining
222 events in the multivariate models.

223 We then focused solely on cardiac-specific outcomes assessed at 6-months and 12-months. In
224 patients with cardiac involvement at diagnosis (n = 58), patients harboring CHIP had a lower rate
225 of cardiac organ response compared to those without CHIP, but this difference was not
226 statistically significant (7/14, 50%, versus 28/44, 64%, respectively, p = 0.532). Cardiac-specific
227 disease progression was observed in a total of 29 patients (38%), including 8 patients with CHIP
228 (50%) and 21 patients without CHIP (35%). CHIP presence was not associated with a higher

229 hazard for cardiac-specific disease progression in a univariate Cox regression analysis (HR 1.66,
230 95% CI 0.70 – 3.94, p = 0.250).

231

232 **Discussion**

233 Hereby, we describe the prevalence, clinical characteristics, and outcome implications of CHIP
234 presence in a cohort of 76 consecutive AL amyloidosis patients seen at our center. We noted that
235 CHIP occurs at a higher frequency (21%) than expected for an age-matched healthy population
236 (5-10% depending on the studies).^{19,20} The median VAF of CHIP-associated mutations was
237 0.036, lower than what was previously reported in AL amyloidosis patients, but comparable to
238 MM patients.^{31,32} We also report on an association between CHIP and the presence of
239 prognostically adverse, t(11;14). Harboring CHIP was not associated with a decreased OS,
240 MOD-PFS, or cardiac-PFS, and, although a lower rate of cardiac response was observed, this
241 difference was not statistically significant. The presence of orthostatic hypotension requiring
242 treatment with midodrine was found to be associated with worse MOD-PFS in a univariate
243 analysis but was not included in the multivariate model. Conversely, an association between an
244 underlying diagnosis of CAD and a Mayo stage 3a or 3b and decreased MOD-PFS was detected
245 in a multivariate model.

246 The prevalence of CHIP we reported (21%) is consistent with the available literature for patients
247 with hematological malignancies, where CHIP driven by DTA genes was identified in 15 – 21%
248 of patients.^{32,33} In our cohort, DNMT3A was the most frequently mutated gene, followed by
249 TET2, which appears in line with previous studies on patients with plasma cell dyscrasia.^{32,33}

250 Consistent with prior data in AL patients, CHIP status did not correlate with age or smoking
251 status.³² This is different from what others have observed in the general population and in the
252 context of MM.^{20,31,38}

253 Furthermore, our study included longitudinal data for a subset of patients. Interestingly, two
254 patients (59 and 80 years old at the time of diagnosis) with no CHIP detected at the initial RHP
255 evaluation were subsequently found to have a CHIP (DNMT3A and ZRSR2, respectively) after
256 only a few cycles of therapy.

257 The association between lower Palladini renal stage⁷ and the presence of CHIP was unexpected,
258 as it was the association between CHIP and the presence of the t(11;14). Age was found not to be
259 a significant confounder in the association between t(11;14) and CHIP, making this association
260 even more intriguing. However, in this cohort of patients, it is impossible to ascertain whether
261 CHIP followed or preceded a diagnosis of t(11;14) plasma cell disorder, and the biological
262 significance of this association still needs to be elucidated. No difference in regards to the
263 kidney-specific outcome was noted between patients with or without CHIP (data not shown), but
264 given the low number of patients, no adjustment could be performed for the renal involvement at
265 diagnosis or the Palladini renal stage⁷ in patients with known renal involvement. Larger
266 prospective studies are needed to clarify the significance of these preliminary findings and
267 potential cause-effect relationship.

268 In our study, the presence of CHIP was not associated with all-cause mortality or MOD-PFS.
269 Although this finding appears in contrast with previous studies on MM and lymphomas
270 undergoing ASCT^{28,31}, they are consistent with what was observed in AL amyloidosis patients in
271 other studies.^{32,33} However, other possible explanations for the absence of an association between
272 CHIP and worse cardiac outcomes may be the small sample size and the limited follow-up of our

273 series. Furthermore, there may have been a bias in patients enrolled in our study as critically ill
274 patients who could not travel to our center for care or who died prior to commencing plasma
275 cell-directed therapy were not included, potentially confounding the association with CHIP.

276 Interestingly, we identified an association between the presence of orthostatic hypotension
277 requiring treatment and reduced MOD-PFS. Although we chose not to include this variable in
278 the final multivariate analysis to avoid overfitting the model, the impact between orthostatic
279 hypotension and lower outcome persisted when included in the multivariate analysis (data not
280 shown). This has not been previously reported and warrants further evaluation.

281 There are several limitations to our study. First, given the limited sample size, the power of our
282 analysis is significantly reduced, with wide confidence intervals. Second, because of the short
283 duration of follow-up, a low number of events was detected, reducing our capacity to fit more
284 variables into our regression model. Lastly, in our series, $t(11;14)$ did not emerge as a negative
285 prognostic factor and was therefore not included in the multivariate model. This could be due to
286 the use of DaraCyBorD in over a third of the patients or possibly to the short duration of follow-
287 up and low incidence of events.

288 In conclusion, we showed that CHIP mutations are frequent in the largest cohort of patients with
289 AL amyloidosis analyzed to date. We demonstrated an association between CHIP and a lower
290 Palladini renal stage⁷ at diagnosis and the presence of $t(11;14)$. No impact on OS or MOD-PFS
291 was observed for patients with CHIP as compared to patients without CHIP. Larger prospective
292 studies and a more prolonged follow-up are needed to better elucidate the impact of CHIP in
293 patients with AL amyloidosis and to establish a cause-effect relationship with the observed
294 associations.

295

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302

303 **Authorship**

304 Contribution: G.B. and N.B. designed the study. P.L. and G.B. wrote the manuscript. P.L. and
305 A.M. performed the analysis. E.B., T.C., M.M., and S.B. collected the data. G.B. and N.B.
306 secured funding for research. All the authors critically revised the manuscript and agreed on the
307 final version.

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418 ASXL1 mutations are strongly associated with smoking. *Leukemia*. 2020;34(10):2660-2672.
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- 420

421 **Table 1.** Demographics of AL amyloidosis patients with and without CHIP.

Demographics	Total (n=76)	With CHIP (n=16)	Without CHIP (n=60)	P (CHIP versus no CHIP)
Age (mean ±SD)	63 ±9.9	66 ±10.1	63 ±9.8	0.28*
Gender				
Male	44 (57.9%)	7 (43.7%)	37 (61.7%)	0.26†
Female	32 (42.1%)	9 (56.3%)	23 (38.3)	
Ethnicity				
White	65 (85%)	15 (94%)	50 (84%)	0.83†
Black	6 (8%)	1 (6%)	5 (8%)	
Asian	2 (3%)	0	2 (3%)	
Middle-Eastern	1 (1%)	0	1 (2%)	
Other	2 (3%)	0	2 (3%)	
Smoking status				
Smoker	43 (58%)	7 (44%)	24 (41%)	0.54†
Non-Smoker	31 (42%)	9 (56%)	34 (59%)	
Other Neoplasm				
Yes	13 (17%)	3 (19%)	10 (17%)	0.55†
MM	9 (12%)	3 (19%)	6 (10%)	
SMM	2 (3%)	0	2 (3%)	
WM	1 (1%)	0	1 (2%)	
CML	1 (1%)	0	1 (2%)	
No	63 (83%)	13 (81%)	50 (83%)	
Type of FLC				
Lambda	63 (82.9%)	12 (75%)	51 (85%)	0.45†
Kappa	13 (17.1%)	4 (25%)	9 (15%)	
Treatment status at CHIP evaluation				
Newly Diagnosed	51 (67%)	11 (69%)	40 (67%)	1†

Rel/Ref	25 (33%)	5 (31%)	20 (33%)	
Median time in months from diagnosis	1 (0 – 159)	1 (0 – 41)	1 (0 – 159)	0.73‡
Median number of lines of therapy (range)	1 (1 – 3)	1 (1 – 3)	1.5 (1 – 2)	0.64‡
Mayo Stage				
I	21 (28%)	2 (12.5%)	19 (32%)	0.44‡
II	18 (24%)	5 (31%)	13 (22%)	
IIIA	21 (28%)	7 (44%)	14 (24%)	
IIIB	15 (20%)	2 (12.5%)	13 (22%)	
Mayo stage IIIA - IIIB	36 (48%)	9 (56%)	27 (46%)	0.58
Palladini Kidney Stage (if kidney involvement)				0.001‡
I	9 (28%)	5 (100%)	4 (15%)	
II	14 (44%)	0	14 (52%)	
III	9 (28%)	0	9 (33%)	
Number of Organs involved (median, range)	3 (1 – 7)	3 (1 – 5)	3 (1 – 7)	0.43‡
Sites				
Heart	60 (79%)	15 (94%)	45 (75%)	N.S.
Renal	32 (42%)	5 (31%)	27 (45%)	
ANS	23 (30%)	8 (50%)	15 (25%)	
PNS	19 (25%)	3 (19%)	16 (27%)	
GI	36 (47%)	8 (50%)	28 (47%)	
Soft tissue/Skin	32 (42%)	7 (44%)	25 (42%)	
Liver	3 (4%)	1 (6%)	2 (3%)	N.As.
Lungs/Pleura	4 (5%)	0	4 (7%)	
Lymph node	2 (3%)	0	2 (3%)	
LVEF (mean ±SD)	53 ±9.5	55 ±6.4	53 ±10.2	0.28*

≤40%	9.5%	0	12%	0.34†
≤50%	28%	13%	33%	0.13†
Coronary artery disease				
Yes	6 (8%)	2 (12%)	4 (7%)	0.6†
No	70 (92%)	14 (88%)	56 (93%)	
Del 17p				
Yes	36 (13%)	0 (13%)	1 (13%)	1†
No	18 (58%)	13 (69%)	40 (55%)	
N.A.	22 (29%)	3 (19%)	19 (32%)	
t(4;14)				
Yes	1 (1%)	0	1 (2%)	1†
No	53 (70%)	13 (81%)	40 (67%)	
N.A.	22 (29%)	3 (19%)	19 (32%)	
Monosomy 13				
Yes	5 (7%)	0	5 (8%)	0.32†
No	49 (64%)	13 (81%)	36 (60%)	
N.A.	22 (29%)	3 (19%)	19 (32%)	
Gain 1q				
Yes	6 (8%)	2 (12%)	4 (7%)	0.62†
No	48 (63%)	11 (69%)	37 (61%)	
N.A.	22 (29%)	3 (19%)	19 (32%)	
≥2 trisomies				
Yes	3 (4%)	0	3 (5%)	1†
No	51 (67%)	13 (81%)	38 (63%)	
N.A.	22 (29%)	3 (19%)	19 (32%)	
t(11;14)				
Yes	26 (34%)	11 (69%)	15 (25%)	0.004†
No	28 (37%)	2 (12%)	26 (43%)	

N.A.	22 (29%)	3 (19%)	19 (32%)	
BM PC percentage (median, IQR)	15% (7% – 20%)	17% (10% – 22%)	15% (7% – 20%)	0.08‡
First line treatment				
CyBorD	35 (46%)	5 (31%)	30 (50%)	0.33†
Dara-CyBorD	28 (37%)	8 (50%)	20 (33%)	
Other§	12 (16%)	3 (19%)	9 (15%)	
None	1 (1%)	0	1 (2%)	
ASCT				
Yes	10 (13%)	2 (12.5%)	8 (13%)	1†
No	66 (87%)	14 (87.5%)	52 (87%)	

422
423 SD, standard deviation; rel/ref, relapsed/refractory; SMM, smoldering multiple myeloma; CML, chronic
424 myeloid leukemia; N.S., non-statistically significant; N.As., statistical significance not assessed; N.A., not
425 available; BM, bone marrow; PC, plasma cells; IQR, interquartile range.

426 * Using two-samples independent t-test.

427 † Using Fisher’s exact test.

428 ‡ Using Wilcoxon rank-sum test.

429 §Other regimens were bortezomib – dexamethasone, lenalidomide – bortezomib – dexamethasone,
430 daratumumab – bortezomib – dexamethasone, and melphalan – dexamethasone.

431

432 **Figure 1. Oncoplot displaying CHIP results.** Oncoplot showing the relative genetic
433 contribution to CHIP in the cohort (76 patients, of which 16 carrying CHIP-defining lesions).
434 Mutation types are color-coded according to legend. The right barchart shows the number of
435 samples carrying a specific genetic variant while the top barchart recapitulates the number of
436 variants per sample. DTA lesions are shown in the first three lines followed by other genes
437 involved in hematologic neoplasms.

438
439 **Figure 2. Longitudinal data.** Plot showing patients with longitudinal samples available. White
440 circles represent RHP not showing any CHIP, while black squares represent RHP where CHIP
441 was detected. Two patients (PT 14 and PT16) who had no CHIP at the initial RHP, were later
442 found to harbor one CHIP (DNMT3A and ZRSR2, respectively). In contrast, no CHIP was found
443 in the repeated RHP of the remaining eight patients.

444
445 **Figure 3. Overall survival and MOD-PFS.** Kaplan-Mayer curves showing overall survival (A)
446 and MOD-PFS (B) expressed in months among AL amyloidosis patients with (red line) and
447 without (black line) CHIP.

448
449 **Figure 4. Multivariate Cox regression model.** Forest plot recapitulating MOD-PFS
450 determinants in terms of Hazard Ratios. Mayo stage >2, and the presence of CAD were found to
451 be statistically significant negative MOD-PFS predictors ($p=0.004$ and $p=0.006$, respectively).

Figure 1

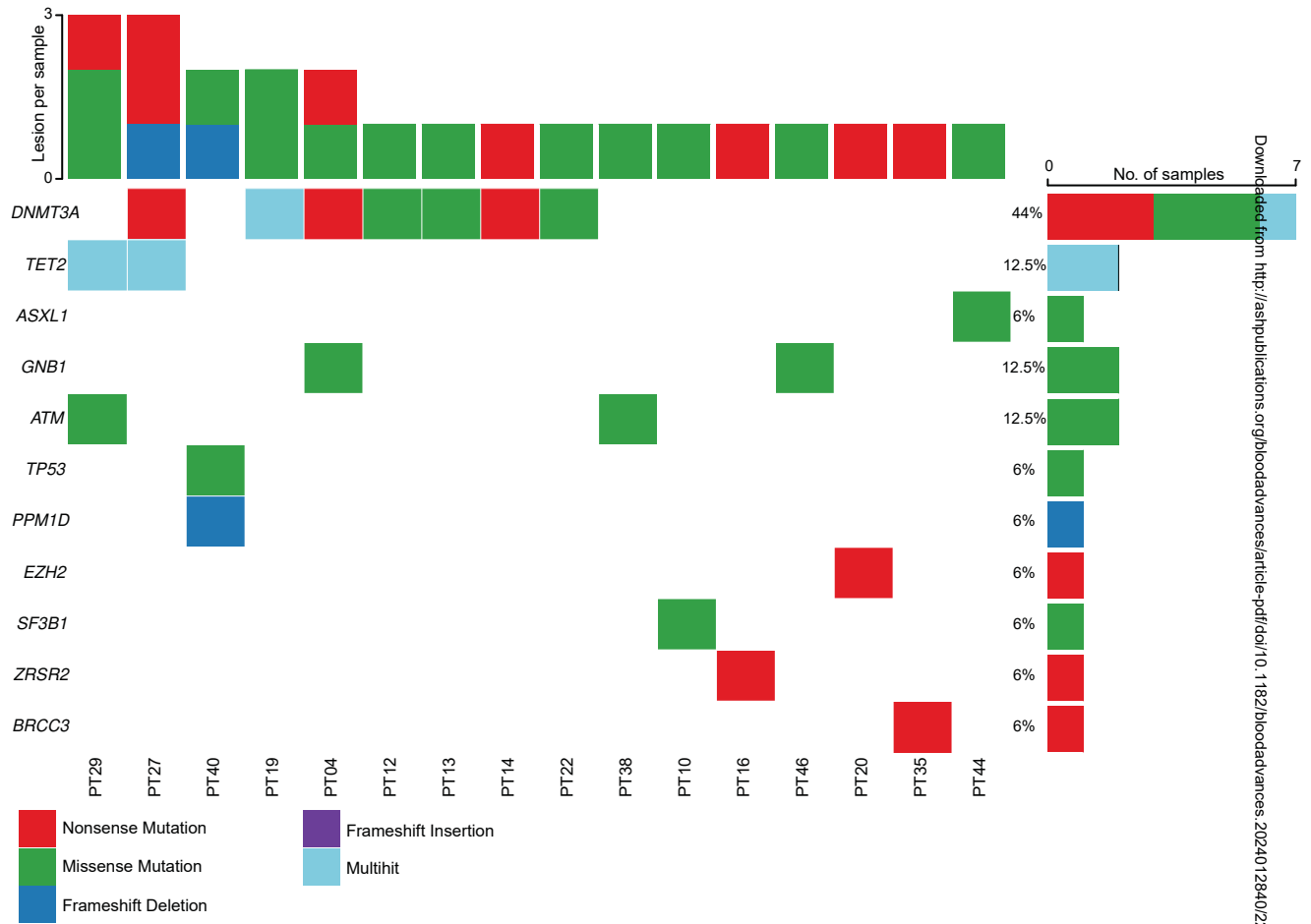


Figure 2

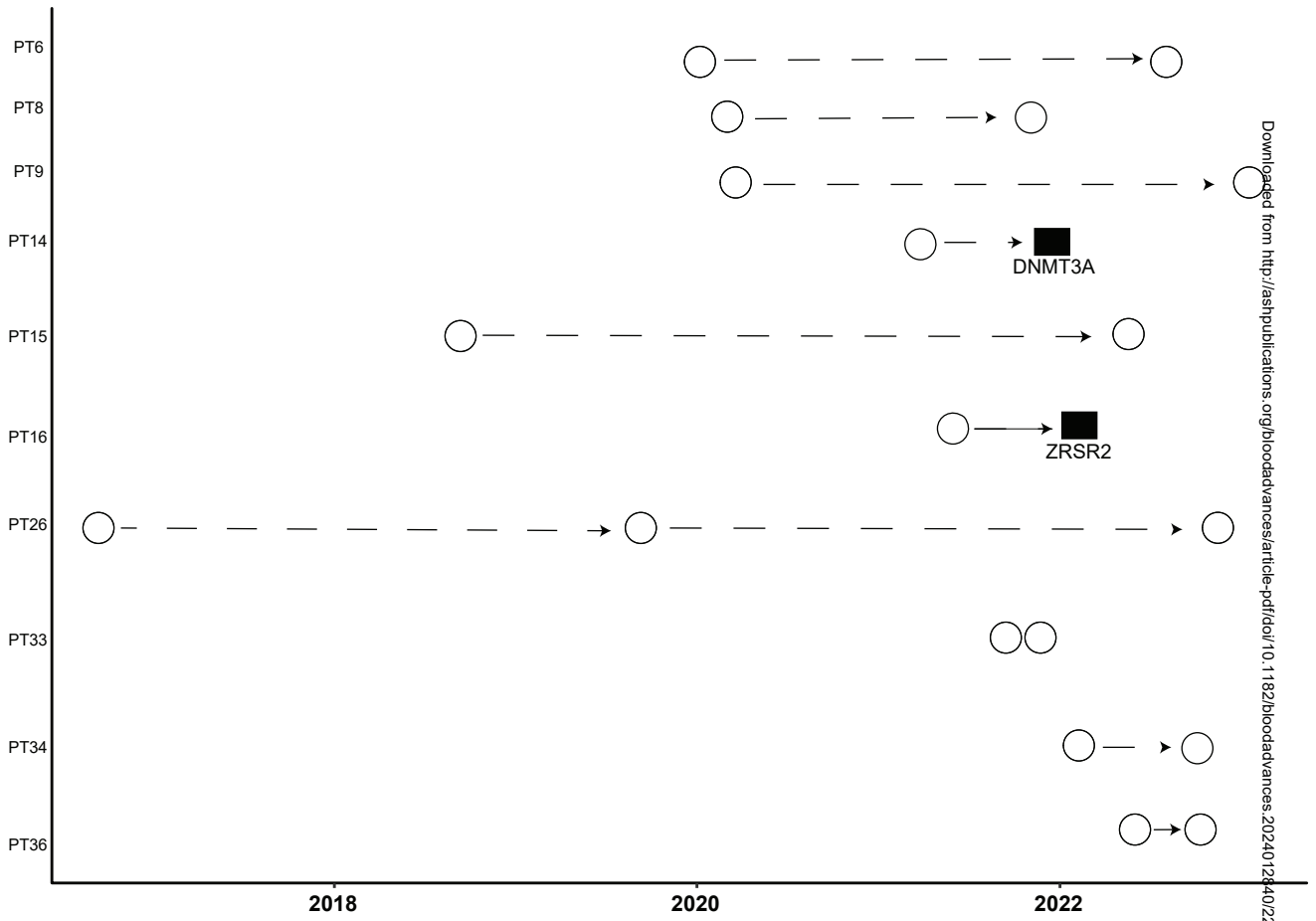
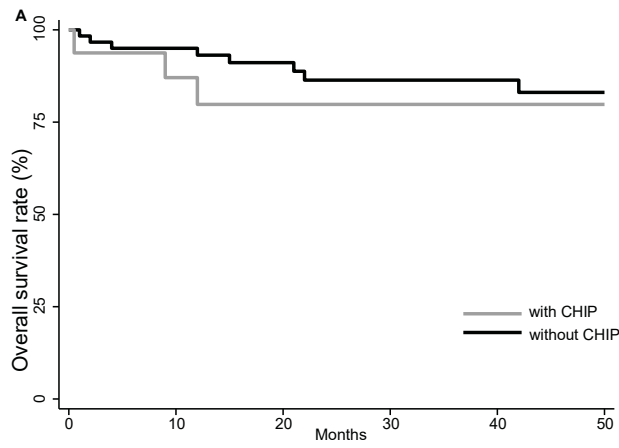
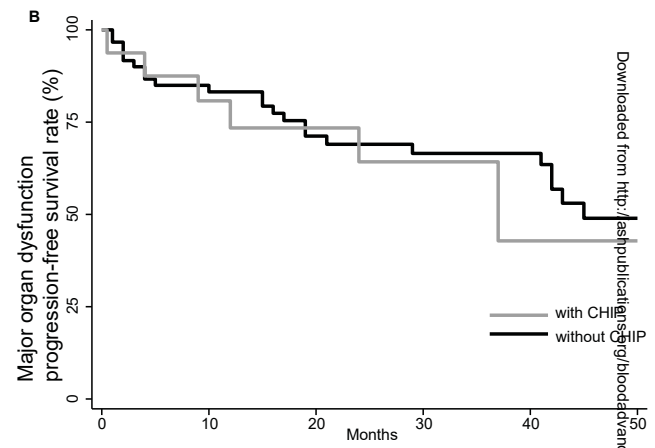


Figure 3



	Number at risk					
With CHIP	16	13	9	7	3	3
Without CHIP	60	53	41	34	28	12
Months	0	10	20	30	40	50



	Number at risk					
With CHIP	16	12	8	7	2	2
Without CHIP	60	48	34	27	22	8
Months	0	10	20	30	40	50

Figure 4

