



Brain Calcifications: Genetic, Molecular, and Clinical Aspects

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Abstract: Many conditions can present with accumulation of calcium in the brain and manifest with a variety of neurological symptoms. Brain calcifications can be primary (idiopathic or genetic) or secondary to various pathological conditions (e.g., calcium–phosphate metabolism derangement, autoimmune disorders and infections, among others). A set of causative genes associated with primary familial brain calcification (PFBC) has now been identified, and include genes such as *SLC20A2*, *PDGFB*, *PDGFRB*, *XPR1*, *MYORG*, and *JAM2*. However, many more genes are known to be linked with complex syndromes characterized by brain calcifications and additional neurologic and systemic manifestations. Of note, many of these genes encode for proteins involved in cerebrovascular and blood–brain barrier functions, which both represent key anatomical structures related to these pathological phenomena. As a growing number of genes associated with brain calcifications is identified, pathways involved in these conditions are beginning to be understood. Our comprehensive review of the genetic, molecular, and clinical aspects of brain calcifications offers a framework for clinicians and researchers in the field.

Keywords: primary familial brain calcification (PFBC); SLC20A2; PDGFB; PDGFRB; XPR1; JAM2; MYORG

1. Introduction

Brain calcifications (BC) are intracranial calcium deposits localized in the brain parenchyma and its microvasculature [1,2]. Their prevalence ranges from 1% in young individuals up to 38% in elderly subjects [2–4]. Calcified areas are easily identified by clinicians as hyperdense alterations on brain CT. A certain degree of intracranial calcifications, particularly of the basal ganglia, pineal gland, choroid plexus, and habenula, can be considered a normal phenomenon associated with aging [2]. Indeed, BC are often incidental findings on neuroimaging of asymptomatic individuals; however, they can also be associated with many genetic and acquired disorders [5,6].

BC can be primary, as observed in several early- and late-onset genetic syndromes, or can be secondary to systemic alterations of phosphate–calcium metabolism (genetic and also acquired forms), intrauterine (e.g., TORCH) and post-natal infections (e.g., neurocysticercosis), hypoxic-ischemic injuries, toxic exposures (e.g., lead), brain tumors (e.g., oligodendrogliomas), and autoimmune disorders (e.g., systemic lupus erythematosus) [2,5].

Although large-scale epidemiological studies are lacking, the most common neurological disorder associated with late-onset BC is traditionally known as Fahr disease [5]. It is clinically defined by the variable presence of movement disorders, recurrent headaches, and psychiatric manifestations, in association with the presence of bilateral BC, most commonly in the basal ganglia, but also in the subcortical white matter, thalamus, and cerebellum [1]. Historically, different names have been used to refer to this neurological condition, including: idiopathic basal ganglia calcification (IBCG), bilateral striopallidodentate calcinosis



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). (BSPDC), and primary familial brain calcification (PFBC) [1]. PFBC is currently the most commonly used term and defines a genetically confirmed neurodegenerative disorder with BC, in absence of a secondary cause. The term "primary bilateral brain calcification" has been proposed to define both inherited and sporadic cases, facilitating the differentiation between primary and secondary forms, and including all possible anatomical distributions of calcium deposits [7].

Many other heterogeneous early-onset complex genetic syndromes manifest BC as a part of their clinical presentation, almost invariably in association with other signs and symptoms, including neurodevelopmental delay, intellectual disability, epilepsy, dystonia, dysmorphisms, and varied systemic involvement [1,5].

No disease-modifying therapies are available for primary brain calcifications, nor have definite pharmacological targets been identified. Therefore, it is important to increase awareness of the clinical and molecular aspects of this group of disorders in order to facilitate diagnosis and stimulate translational research in this often overlooked field. In this review, we describe the genetic, molecular, and clinical aspects of primary BC, with the main focus directed on a detailed characterization of PFBC. In addition, an overview of the early-onset genetic syndromes associated with BC is provided.

2. Genetics of Brain Calcification

In the past decades, genetic etiology of IBGC was suspected based on the identification of several familial clusters [8–11]. However, no genetic cause of IBGC was recognized before the identification in 2012 of *SLC20A2* as the first associated gene [12]. Since then, six genes have been definitively linked with PFBC: four inherited in an autosomal dominant manner (*SLC20A2*, *PDGFB*, *PDGFRB*, and *XPR1*) and two displaying an autosomal recessive inheritance (*MYORG* and *JAM2*) [13–17]. Very recently, a novel gene (i.e., *CMPK2*) has been proposed to be associated with autosomal recessive brain calcifications and this awaits confirmation in independent studies [18]. A review of data from 555 genetically diagnosed patients with PFBC revealed the following frequency of mutations: *SLC20A2*~60%, *MYORG*~13%, *PDGFB*~13%, *PDGFRB*~6%, *XPR1*~6%, *JAM2*~2% [19,20]. A brief genetic characterization of each these genes is provided below in association with the genetic evidence deriving from knock-out or hypomorphic animal models.

SLC20A2 (chr. 8p11.21, 11 exons) encodes the sodium-dependent phosphate transporter 2 (652 amino acids), which plays a fundamental role in cellular phosphate transport. Several pathogenic variants have been reported, including missense, nonsense, splice-disruptive, small indels, and gross deletions. Pathogenic variants have been shown to impair uptake of phosphate (loss-of-function mechanism of disease) [12]. Interestingly, a complete Slc20a2 knockout mouse showed extensive, bilateral calcifications in the thalamus, basal ganglia, and cortex, recapitulating the human disease [21].

MYORG (chr. 9p13.3, one coding exon) encodes the myogenesis regulating glycosidase (putative) (714 amino acids), a protein located in the endoplasmic reticulum and nuclear membranes whose functions remain poorly defined, but are probably linked to protein glycosylation [22]. Many pathogenic variants have been described, including missense, nonsense, and small indels. The presence of biallelic nonsense mutations of *MYORG* in PFBC patients strongly suggests that a functional loss of *MYORG* is associated with brain calcifications. Indeed, Myorg-KO mice and zebrafish develop progressive brain calcifications [14,23].

PDGFRB (chr. 5q32, 23 exons) encodes the platelet-derived growth factor receptor beta (PDGFR- β) (1106 amino acids), a cell surface tyrosine kinase receptor for mesenchymal cell mitogens. Several pathogenic variants are known, including missense and start-loss mutations. Pdgfrb-deficient mice lack pericytes, essential for blood–brain barrier (BBB) formation, and display increased vascular permeability [24].

PDGFB (chr. 22q13.1, six exons) encodes the platelet-derived growth factor subunit B (241 amino acids), which is a potent mitogen for cells of mesenchymal origin, by binding and activating PDGF receptor tyrosine kinases such as PDGFR- β . A number of pathogenic

variants have been reported in association with PFBC, including missense, nonsense, splicedisruptive, start loss, stop loss, and gross deletions. Hypomorphic Pdgfb mice develop brain calcifications that correlate with the degree of pericyte and BBB deficiency [17].

XPR1 (chr. 1q25.3, 15 exons) encodes the xenotropic and polytropic retrovirus receptor 1 (696 amino acids), which plays a role in phosphate homeostasis and mediates phosphate export from the cells. Few pathogenic variants have been reported, which include missense and splice disruptive. Remarkably, the majority of these are located in the SPX putative regulatory domain of the protein [15,25].

JAM2 (chr. 21q21.3, 10 exons) encodes the junctional adhesion molecule 2 (298 amino acids), which is localized at the tight junctions of both epithelial and endothelial cells and mediates heterotypic cell–cell interactions [26]. Being the most recently described gene, few pathogenic variants have been reported, including missense, start-loss, nonsense, and splice-disruptive. Pathogenic variants lead to reduction/absence of *JAM2* protein, consistent with a loss-of-function mechanism. The brain calcification phenotype is replicated in the Jam2 complete knockout mouse [13].

Genetics of Early-Onset Syndromes Associated with Brain Calcifications

Calcifications in the basal ganglia and other brain structures are also observed in several genetic diseases with normal calcium–phosphate metabolism. This clinically and genetically highly heterogeneous group of syndromes includes mitochondrial diseases (predominantly mtDNA mutations), Aicardi–Goutières syndrome (*ADAR, RNASEH2A, RNASEH2B, RNASEH2C, SAMHD1, TREX1, IFIH1*), Coats plus syndrome (*CTC1*), leukoencephalopathy with calcifications and cysts (*SNORD118*), Cockayne syndrome (*DDB2, ERCC1, ERCC2, ERCC3, ERCC4, ERCC5, ERCC6, ERCC8, GTF2H5, MPLKIP, POLH, XPA, XPC*) and many others. Deep clinical phenotyping is crucial before genetic testing in this context [27]. In the absence of specific diagnostic clues for a particular form (see below) the best genetic approach is next-generation sequencing (e.g., gene panels or exome sequencing) [27]. A detailed genetic description of every one of the many genes involved in these syndromes is beyond the scope of this review. A complete list is provided in Table 1.

Table 1. Genetic syndromes associated with brain calcifications—The table summarizes the genetic forms presenting brain calcifications. Demographic, clinical, and imaging traits for each syndrome are summarized in the table. MOI = mode of inheritance, AD = autosomal dominant, AR = autosomal recessive, mt = mitochondrial, XLR = X-linked recessive, IC = isolated cases. For the specific references of each condition, please refer to the main text.

| Gene (Age of Onset (Range, Years)/Penetrance (%)/MOI) | Main Neurological Symptoms | Other Symptoms | Brain Calcification |
|---|--|---|---|
| <i>SLC20A2</i> (1–84/60%/AD) | >20%: Parkinsonism, cognitive deficit, headache 10–20%: Tremor, dysarthria, dystonia <10%: Depression, psychosis, ataxia, anxiety, seizure, chorea | | Basal ganglia, subcortical white matter, and dentate nuclei |
| PDGFB (1–70/86%/AD) | >20%: Headache, cognitive deficit, depression 10–20%: Parkinsonism, psychosis, ataxia, tremor, chorea, anxiety <10%: Dystonia, dysarthria, seizure | Basal ganglia, subcortical white matter, and dentate nuclei | |
| PDGFBR (6–82/48%/AD) | >20%: Headache, cognitive deficit 10–20%: Depression, Parkinsonism <10%: Tremor, anxiety, dysarthria, seizure, psychosis, ataxia, tremor, chorea, dystonia | | Basal ganglia, subcortical white matter, and dentate nuclei |

Table 1. Cont.

| Gene (Age of Onset (Range, Years)/Penetrance (%)/MOI) | Main Neurological Symptoms | Other Symptoms | Brain Calcification | |
|---|--|--|---|--|
| XPR1 (16–79/70%/AD) | >20%: Cognitive deficit, Parkinsonism, dysarthria 10–20%: Tremor, ataxia <10%: Headache, anxiety, seizure, psychosis, depression, dystonia, chorea | | Basal ganglia, subcortical white matter, and dentate nuclei | |
| <i>MYORG</i> (8–87/91%/AR) | >20%: Dysarthria, cognitive deficit, ataxia, parkinsonism, psychosis, tremor 10–20%: Depression <10%: Headache, dystonia, chorea, seizure, anxiety | | Basal ganglia, subcortical white matter, and dentate nuclei | |
| <i>JAM2</i> (7–50/85%/AR) | >20%: Parkinsonism, ataxia, cognitive deficit, dysarthria, seizure <10%: Dystonia | | Basal ganglia, subcortical whi matter, and dentate nuclei, midbrain | |
| Mitochondrial conditions (MERRF, Leigh syndrome, KSS—mtDNA) (Childhood, adulthood/full/matrilinear) | Epilepsy, myoclonus, ataxia, spasticity Dysphagia (LS) | Epilepsy, myoclonus, ataxia, spasticity Short stature, optic atrophy, hearing loss, ptosis, diabetes mellitus, cardiomyopathy, neuropathy | | |
| Coats plus syndrome (<i>CTC1</i>) (Childhood/full/AR) | Developmental delay, epilepsy, spasticity, dystonia, ataxia Developmental delay, epilepsy, spasticity, dystonia, ataxia Metial telangiectasia and exudates, osteopenia and fractures, gastrointestinal and hepatic bleeding, portal hypertension, bone marrow suppression Growth retardation | | Calcification with leukoencephalopathy and brain cysts | |
| Aicardi-Goutières syndrome (ADAR, RNASEH2A, RNASEH2B, RNASEH2C, SAMHD1, TREX1, IFIH1) (Childhood (usually after a short period of normal development)/full/AR/AD) | Dystonia, hypotonia, developmental delay Increased startle reaction | Acquired microcephaly, Hepatosplenomegaly Sterile pyrexias Chilblain lesions (ears, hands, fee) Skin mottling Increased liver enzyme, thrombocytopenia (transitory), interferon signature | Calcifications of basal ganglia and white matter Leukoencephalopathy Progressive cerebral atrophy | |
| Leukoencephalopathy, cystic, without megalencephaly (<i>RNASET2</i>) (1 year/full/AR) | Seizure Neurological deterioration | Microcephaly | Brain calcification Cystic lesions (anterior, subcortical) Leukoencephalopathy | |
| Claudin-5 (<i>CLDN5</i>) (1 year/full/AR) | Seizure Developmental delay | | Atrophy of the pons Calcification of the vertical pontine white matter bundles Cortical and basal ganglia calcification | |
| Hoyeraal Hreidarsson syndrome (DKC1, TINF2) and Congenital Dyskeratosis (Zinsser-Engman-Cole syndrome) (TERC, TERT, NHP2, NOP10, WRAP53, RTEL1) (Childhood (first months)/full/AR X-linked/AD/AR) | TINF2) and Congenital tosis Engman-Cole ne) (TERC, TERT, NHP2, WRAP53, RTEL1) od (first /full/AR | | Cerebellar hypoplasia Basal ganglia and subcortical calcifications | |
| 3-hydroxyisobutyric aciduria (1st decade/NA/NA) | Hypotonia and impaired neurological development | Microcephaly Facial dysmorphism Episodes of ketoacidosis | Migration brain disorder Calcification in the frontal and subependymal region and periventricular | |
| Inborn error of folates Metabolism (FOLR1, SLC46A1, MTHFR, DHFR, MTFD1) (1st decade/full/AR) | Seizure Severe neurological deterioration | Leukopenia, megaloblastic bone marrow, hypoimmunoglobulinemia. | Demyelination Calcification of basal ganglia and white matter | |
| Carbonic anhydrase II deficiency syndrome (CA2) (Congenital/full/AR) | Possible mental retardation | Osteopetrosis Renal tubular acidosis | Calcification of the basal ganglia, thalamus, subcortical white matter | |

Table 1. Cont.

| Gene (Age of Onset (Range, Years)/Penetrance (%)/MOI) | Main Neurological Symptoms | Other Symptoms | Brain Calcification Microhemorrhage or and deep intracerebral hemorrhages Lacunar infarcts Dilated perivascular spaces Leukoencephalopathy Porencephaly Intracranial calcifications | |
|---|--|--|--|--|
| Collagen IV disorders (<i>COL4A1</i> , <i>COL4A2</i>) (Childhood–adult/full/AD) | Seizure Ischemic or hemorrhagic events Migraine with aura (also isolated) Severe cases with intellectual disability and infantile motor symptoms Neuropathy, muscle cramps | Systemic vascular involvement (aneurysms) and symptoms (renal, ocular, muscular) | | |
| Cockayne syndrome (DDB2, ERCC1, ERCC2, ERCC3, ERCC4, ERCC5, ERCC6, ERCC8, GTF2H5, MPLKIP, POLH, XPA, XPC) (1st decade/Full/AR) Nasu Hakola | Spectrum of conditions Neurodevelopmental and growth delay Impaired vision, hearing and severe neurological deficits | Cataract Scoliosis, contractures Arthrogryposis, microphthalmia | Calcification in the basal ganglia (possible) | |
| Disease—Polycystic lipomembranous osteodysplasia with sclerosing leukoencephalopathy 1 and 2 (<i>TYROBP</i> , <i>TREM2</i>) (3rd decade (bone changes) 4th decade (neurological symptoms)/full/AR) | Presenile dementia Parkinsonism Seizure | Cyst-like bone lesions | Brain atrophy Leukodystrophy Basal ganglia calcification | |
| Nakajo Nishimura syndrome (<i>PSMB8</i>) (Childhood (Japanese)/full/AR) | | Nodular-erythema like changes Lipodystrophy Finger Joint deformities Specific inflammatory signature and hyperpyrexia Possible cardiac alteration | Basal ganglia calcification Vessel calcifications (brain and systemic) | |
| Adams Oliver syndrome 2 (DOCK6) (Congenital/full/AR) | Epilepsy | Facial dysmorphism Ophthalmological abnormalities (congenital cataract, rod dystrophy, vitreoretinal abnormalities, optic atrophy) Aplasia cutis congenita of the scalp Terminal transverse limb defects Systemic abnormalities (cardiac, renal, hepatic) | Periventricular calcifications and ventricular enlargement | |
| Raine Syndrome (FAM20C) (77) (Congenital/full/AR) | Neurodevelopmental delay Hydrocephalus Self-mutilating behavior | Microcephaly Facial dysmorphism Exophthalmos, gum hyperplasia, osteosclerosis | Punctiform periventricular and white matter calcifications | |
| Keutel Syndrome (MGP) (Childhood/full/AR) | | | | |
| Fried Syndrome (<i>AP1S2</i>) (Childhood/full/X-linked recessive) | Neurodevelopmental delay hydrocephalus Hypotonia, mental retardation, stereotypies | Microcephaly Short stature Facial dysmorphism Optic nerve atrophy | Basal ganglia calcification | |
| RAB39B (<40 years/full/X-linked) | Parkinsonism | Juvenile Parkinsonism Intellectual disability | Basal ganglia calcifications reported in some cases | |
| Hemorrhagic destruction of the brain, subependymal calcification, and cataracts (JAM3) | Seizure Severe neurological deterioration Early death | Congenital cataract | Brain hemorrhagic destruction Subependymal calcification | |
| Pseudo-TORCH syndrome 1 (<i>OCLN</i>) (Congenital/full/AR) | Seizure Pyramidal signs Hypotonia | Microcephaly Possible dysmorphic features | Calcification of the white matter, basal ganglia, cerebellum, brain stem Cortical dysgenesis | |
| Pseudo-TORCH syndrome 2 (<i>USP18</i>) (Congenital/full/AR) | Seizure Neurological deterioration | Microcephaly Possible dysmorphic features Immune abnormalities Intravascular coagulopathy | Calcifications Cerebral hemorrhage Cortical dysgenesis | |

| Gene (Age of Onset (Range, Years)/Penetrance (%)/MOI) | Main Neurological Symptoms | Other Symptoms | Brain Calcification | |
|--|--|---|--|--|
| Immunodeficiency 38 with basal ganglia calcification (ISG15) (1st-2nd decade/full/AR) | Seizure | Recurrent infections | Basal ganglia and cortical calcification | |
| Spondyloenchondrodysplasia with immune dysregulation (<i>ACP5</i>) (1st decade/full/AR) | Developmental delay Spasticity | Spondyloenchondrodysplasia Short statue Skeletal dysplasia Immune dysregulation (multiple infections and autoimmune diseases) | Late-onset cerebral calcification | |
| Rajab interstitial lung disease with brain calcifications (FARSA, FARSB) (Infancy–early childhood/full/AR) | Delayed motor development Cognitive development can be preserved | Delayed growth Interstitial lung disease Systemic abnormalities (hepatic, renal, skeletal) Dysmorphic features (possible immune abnormalities) | Calcifications of the basal ganglia and cortex Possible leukoencephalopathy | |
| Seizures, early onset, with neurodegeneration and brain calcification (<i>NRROS</i>) (1st decade/full/AR) | Seizure Developmental regression Pyramidal signs | | Brain atrophy White matter abnormalities Calcifications Possible corpus callosum hypoplasia Possible cystic degeneration of the white matter | |
| Spastic paraplegia 56, autosomal recessive (CYP2U1) (1st decade/full/AR) | Spastic paraplegia (and also upper limbs involvement) Dystonia Cognitive impairment | Macular dystrophy Pseudoxanthoma elasticum | White matter lesion Atrophy of the corpus callosum Calcifications of the globus pallidus | |
| Down syndrome (Trisomy 21) (Congenital/full/IC) | Mental retardation Decreased muscle tone | Dysmorphic features, palm crease, large tongue, umbilical hernia, intestinal blockage, congenital heart disease | Basal ganglia (mostly globus pallidus) | |
| Pseudohypoparathyroidism (GNAS, PRKAR1A, PDE4D, PDE3A) (1st–5th decade/full/AD) | Hypocalcemia tetani Seizure Mental retardation | Short stature Subcutaneous calcifications Brachydactyly Hypocalcemia, hyperphosphatemia, and increased serum concentration of PTH (PTH resistance) Normal renal function | Calcification of the basal ganglia, thalamus, subcortical white matter | |

Table 1. Cont.

3. Molecular Mechanisms of Brain Calcifications

The pathophysiology of PFBC is profoundly linked to the loss of integrity of the BBB and should be distinguished from the calcification processes that take place in secondary brain calcifications, in which calcium deposition develops via two main pathways: dystrophic calcification resulting from membrane disruption and uncontrolled calcium entry (hypoxic ischemic injury, intrauterine infections), and calcium–phosphate metabolism alterations (pseudohypoparathyroidism and pseudopseudohypoparathyroidism, mainly linked to loss of function mutations of GNAS, involved in the intracellular transmittal pathway of PTH) [28].

Considering the functions of the known causative genes, the pathology of PFBC can be ascribed to altered phosphate transportation or impaired permeability through the BBB, which both result in increased inorganic phosphate (Pi) levels in CSF and brain interstitial fluid. The excess of Pi induces its deposition in the form of calcium hydroxyapatite $(Ca_{10}[PO_4]_6[OH]_2)$ in the vascular extracellular matrix (Figure 1).

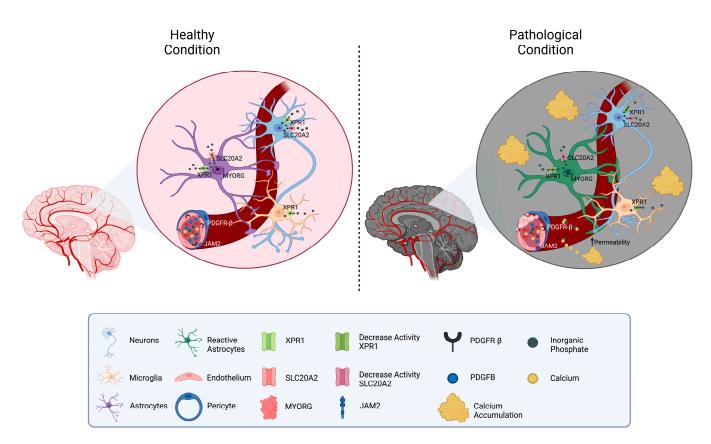


Figure 1. Schematic representation of the molecular mechanisms related to PFBC-genes. A neurovascular unit is depicted. The localization of the genes related to PFBC (*SLC20A2, PDGFB, PDGFRB, XPR1, MYORG*, and *JAM2*) is shown in the figure. On the left side, physiological Pi homeostatic processes are illustrated (mostly modulated by SLC20A2 (import) and XPR1 (export)). On the right side, pathological conditions due to gene mutations' protein down-regulation are depicted. Reduced levels of MYORG result in impaired function and confirmation of astrocytes. Disfunction of PDGFB, PDGFRB, and JAM2 causes increased permeability of the vessels and calcium leakage. Interstitial accumulation of Pi and calcium is responsible for the formation of brain calcifications.

The reasons for the greater basal ganglia vulnerability to calcium deposition are not completely understood but may depend on the higher expression of the causative genes in the basal ganglia neurovascular system, as well as on the peculiar vascularization of these areas [29].

SLC20A2 and *XPR1* operate directly in inorganic phosphate transportation: the former, mainly expressed by neurons, astrocytes, and vascular smooth muscle cells (VSMC), imports extracellular Pi into the cell mediating its transportation from CSF to blood; the latter exports intracellular Pi out of cells such as neurons, astrocytes, and microglia [15]. Mutations of these transporters lead to the accumulation of extracellular Pi, due to reduced Pi internalization in *SLC20A2* mutants, and to intracellular accumulation with subsequent down-regulation of uptake in *XPR1* mutants. It has been hypothesized that persistent elevation in extracellular Pi may induce intracranial calcification, possibly via trans-differentiation of vascular smooth muscle cells (VSMCs) into osteoblast-like cells [30].

Conversely, *PDGFRB*, *PDGFB*, *MYORG*, and *JAM2* operate in the brain neurovascular unit (made of neurons, astrocytes, endothelial cells, pericytes, and VSMCs) and indirectly impair the BBB permeability [31]. PDGFR- β is expressed mainly by pericytes and VSMCs, while PDGFB is predominantly released by the endothelium, enabling the recruitment of PDGFR- β -expressing cells and thus promoting the wrapping of the cerebral vessel [32]. Deficiency of PDGF-B/PDGFR- β signaling causes alteration in pericyte recruitment and endothelial cell morphology, leading to increased vascular permeability. MYORG is predominantly expressed in astrocytes and localized in the endoplasmic reticulum; though presumed to act as an α -glucosidase, its molecular function remains largely unknown [22]. Astrocytic endfeet connect the endothelial cells to the extracellular matrix and neurons, thus exerting an important function in the maintenance of the BBB. Speculatively, intracellular ER alterations may impair astrocytes' properties and conformation, causing BBB disruption. JAM2 is a key component of the tight junctions between endothelial cells in the neuro-vascular unit. *JAM2* loss-of-function causes disarrangement of tight junctions, leading to high Pi leakage from the blood to the brain tissue [13].

Concerning genetic brain calcifications other than PFBC, at least three additional pathogenetic mechanisms haver been identified:

- Cerebral microangiopathies and gliosis, resulting from the malfunctioning of cellular systems fundamental in vessels' stability and/or angiogenesis, such as small nucleolar RNA involved in ribosome biogenesis in leukoencephalopathy with calcification and cysts (LCC) [33], telomeres in Coats plus syndrome, nucleases in Aicardi– Goutières syndrome and type IV collagen in *COL4A1*-related disease [33–36].
- 2. **Reaction to microgliopathy and neuronal loss**, often seen in more complex encephalopathies such as Cockayne syndrome where microscopic examination has highlighted the presence of both iron and calcium deposits, or the NRROS-related disorders, Adams–Oliver syndrome and Nasu–Hakola disease [37].
- 3. **Mitochondriopathies**, where brain calcification may be attributed to co-present endocrine alterations (hypoparathyroidism) and/or to progressive cell degeneration caused by mitochondrial dysfunction in the brain [38].

The underlying pathogenetic mechanisms of ectopic/dystrophic calcifications of noncerebral soft tissues are extremely different and are mainly associated with systemic mineral imbalance (i.e., hyperparathyroidism) or tissue alteration and necrosis (i.e., myositis ossificans) [39].

4. Clinical Aspects

4.1. Primary Familial Brain Calcifications: Clinical Aspects

Significant overlap is found in the clinical presentation of patients with PFBC caused by mutations of the monogenic forms identified so far (including *SLC20A2*, *PDGF*, *PDGFRB*, *XPR1*, *MYORG*, *JAM2*). Characteristic features of PFBC include motor symptoms such as hypokinetic (i.e., Parkinsonism) more than hyperkinetic (chorea, dystonia, or ataxia) movement disorders, seizures, pyramidal signs, cognitive decline, and psychiatric symptoms (such as behavioral changes, depression, anxiety, ADHD, psychosis, or bipolar disorders) [19]. Dysarthria and dysphagia have also been reported [26,40–43]. Headache (migraine with and without aura) and dizziness are often present and may be the presenting features prompting imaging study and thus diagnosis in subjects who are otherwise asymptomatic [41,44].

According to a recent extensive review of all the reported cases of PFBC, headache is very frequent (about 30–40% of cases) in *PDGFB*, *SLC20A2*, and *PDGFRB* mutation carriers [19]. Cognitive deficits are found in up to 50% of subjects with biallelic *JAM2* variants, followed by *MYORG*, and in more than one-third of cases of *XPR1*, *PDGFB*, *SLC20A2*. Parkinsonism is present in up to 80% of subjects with *JAM2*, in about 30% of cases of *XPR1*, *SLC20A2*, *MYORG*, and less frequently in *PDGFB*, *PDGFRB* mutation carriers. Other movement disorders, including chorea and dystonia, display lower frequency (less than 15% of cases). Ataxia is recurrent in *JAM2* cases (in up to 60% of subjects) and MYORG, and less frequent in *PDGFB*, *XPR1*, or *SLC20A2*. Psychiatric manifestations mainly include depression, psychosis, or personality changes. Depression is more frequent and is found especially in *PDGFB*, *MYORG*, and *PDGFRB* carriers, and less in other forms. Headaches are very common (about 30–40% of cases) in *PDGFB*, *SLC20A2*, and *PDGFRB* carriers. Finally, dysarthria is a frequent finding in subjects with biallelic variants of *MYORG* (more than 70% of cases), as well as in *JAM2* and *XPR1* mutation carriers (Table 1) [19].

In carriers of pathogenic variants of *SLC20A2*, additional features have been reported including forms of tremor (head tremor, intention tremor of the upper limbs),

blepharospasm, torticollis, facial palsy, apraxia, palilalia, myoclonus (described as mostly cortical), cramps, active denervation at electromyographic (EMG) recording, polyneuropathy, syncope, as well as ischemic episodes (both transitory and stroke) [40,41,44–51]. Seizures have been described as both grand mal generalized or focal. Interestingly, one subject was described presenting a classical form of paroxysmal kinesigenic choreoathetosis with response to carbamazepine, as classically seen in subjects with PRRT2-related paroxysmal kinesigenic dyskinesia (PKD) [42]. PKD with response to carbamazepine was also reported in one subject with mutations of the *PDGFRB* gene [52]. Carriers of pathogenic variants of this gene also had episodes of urinary incontinence, neuropathy, cramps, and restless leg syndrome [46,49,53]. In subjects with PDGFB mutations, spasmodic adductor dysphonia has been reported, migraine is usually not associated with aura, and chorea and Parkinsonism have so far been found with similar frequency [17,54–56]. Neuropathy has also been described in subjects with *JAM2* mutations [26].

In these subjects, Parkinsonism is often levodopa responsive, with subjects developing motor fluctuations and dyskinesia [16,41,43,50,57]. Freezing of gait has also been observed in subjects with *SLC20A2* mutations [41]. A phenotype resembling supranuclear gaze palsy (PSP) with poor response to levodopa was reported in one patient with *PDGFRB* mutation who presented symptoms at age 79 [58].

For the autosomal forms of PFBC, age of onset of clinical manifestation spans from childhood (first decade) to the 80s, with more frequent presentation between the fourth and sixth decades [19]. Among these forms, *PDGFB* seems to be related to an earlier age of onset (first or second decade), while XPR1 has been found to manifest only after the third decade [19]. Considering autosomal recessive forms of PFBC, age of onset of *JAM2*-manifesting carriers spanned between 8 and 38 years of age. *MYORG* has been reported in subjects with age of onset spanning from the first to the eighth decade [19]. Gender is equally represented in all the genetic forms of PFBC [19].

Two-thirds of carriers usually manifest symptoms, with higher penetrance for the recessive forms (*JAM2* and *MYORG*) and *PDGFB*, followed by *XPR1*, *SLC20A2*, and less than 50% for *PDGFRB* [19]. Intrafamilial variability can be observed for all these genes, both in terms of age of onset as well as clinical presentation, arguing against a strong genotype–phenotype correlation [40]. However, gene variants, clinical traits, and/or age of onset or both showed consistency among family members in certain families [16,44,48,50,56,59].

Interestingly, two identical female twins with mutations of the *PDGFRB* gene showed a similar clinical phenotype (characterized mostly by paresthesia, cramps, and congenital atrial septal defect) and only five-year difference in the age of onset (49 and 54 years) [59]. Both twins presented brain calcification in the white matter, but one presented more extensive calcification in the basal ganglia (with later age of onset) [59].

In PFBC, brain calcifications are reported in the subcortical white matter as well as in the basal ganglia and cerebellum. Interestingly, affected *PDGFB* carriers presented more severe calcification in the thalamus, cerebellum, and white matter, as well as calcifications outside the basal ganglia, compared with non-affected carriers [19]. In these forms, calcification is a progressive process, so that the extent of brain calcification has a tendency to increase with age [19]. Of note, about 75% of the heterozygous carriers of *MYORG* were found to present brain calcifications despite remaining clinically asymptomatic, suggesting a dosage effect related to this gene [60,61]. A genotype-to-phenotype correlation has been observed more in regards to the extent of brain calcifications then in terms of severity of clinical presentation [19]. Of note, pontine calcifications are typically overserved in subjects with mutations or *MYORG* [62]. In subjects with PFBC, serum levels of calcium, parathormone (PRH), and vitamin D were found to be within range [26,41,42,50,51,55,57,63–67].

Whether the brain calcifications are the cause of these conditions or an epiphenomenon is still not clear. Interestingly, in PFBC the extent of brain calcification does not necessarily correlate with the severity of clinical symptoms. Indeed, these forms present a reduced penetrance and extensive brain calcifications can be found in asymptomatic subjects. The current knowledge on the contribution of additional genetic variants in adjunct to the classical PFBC genes in terms of disease penetrance and phenotypic presentation is very limited. Cases were reported of two sisters presenting with basal ganglia calcifications and generalized epilepsy as main phenotype carrying mutations in the *SLC20A2* and in the *CHRNB2* genes (known to be associated with AD frontal lobe epilepsy) and a young boy with basal ganglia calcification and refractory epilepsy carrying mutations in the *SLC20A2* and in the *SLC20A2* and in the *SCN2A* genes [68,69]. In addition, a small deletion on chromosome 8 including the *SLC20A2* and *THAP1* genes (associated with dystonia type 6) was considered responsible for a predominant dystonic phenotype with earlier age of onset compared with carriers of *SLC20A2* alone [70–73].

Compared with classical forms of PFBC, a patient with a mutation in the *CARS* gene and a variant of uncertain significance in the *PDGFRB* gene showed reduced levels of calcium in the peripheral blood [74]. Mutations of the *CASR* gene are associated with hypoparathyroidism, hypocalcemia, and relative hypercalciuria consistent with the low levels of calcium observed in this subject [75].

Finally, one rare case of digenic mutation of both *SLC20A2* and *PDGFRB* genes presented with an early onset (5 years) and severe basal ganglia and frontal calcification, while the heterozygous parents showed only mild brain calcification and no symptoms [76]. A role for digenic variants in determining the penetrance of the disease and the phenotypic presentation has been suggested by the abovementioned studies and warrants further investigation.

4.2. Other Forms of Brain Calcifications: Clinical Aspects

Brain calcifications can be one of the features of (mostly) pediatric encephalopathy due to a number of different genes (summarized in Table 1). These are usually complex conditions that may present with developmental delay and complex neurological presentations, including combinations of pyramidal signs, hypo or hyperkinetic movement disorders, ataxia, epilepsy, and cognitive impairment (such as speech delay/absence). From a radiological perspective, calcification of the basal ganglia, cerebellum, and white matter calcifications, as well as leukoencephalopathy are usually present.

In certain instances, specific radiological characteristics are detrimental to guide the differential diagnosis. The presence of subcortical cysts is suggestive of a diagnosis of LCC due to compound heterozygous variants in the *SNORD118* gene, or of Coats plus syndrome (cerebroretinal microangiopathy) due to mutations in the conserved telomere maintenance component 1 (*CTC1*) gene [77,78]. In Coats plus syndrome, LCC is associated with systemic microvascular manifestation, such as cutaneous or retinal telangiectasia, as well as vascular abnormalities causing gastrointestinal, hepatic, and bone marrow hemorrhage, and osteosclerotic lesions [78]. Cystic degeneration of the white matter, associated with calcification and leukodystrophy, was found in autoptic brains of patients with the NRROS-mutation associated syndrome, Rajab interstitial lung disease with brain calcifications-2, and *RNASET2*-related leukoencephalopathy [79–81].

Another difference between PFBC and other encephalopathies with brain calcification is the natural history of these conditions. While PFBCs usually manifest after the second or third decade, or earlier in life but after a normal development, other encephalopathies with brain calcification can present at birth with progressive developmental delay, such as in Cockayne syndrome [82]. Some forms present a biphasic progression (such as Aicardi– Goutières syndrome) while others are static encephalopathies (such as Adams–Oliver syndrome 2) [77,83].

Certain mitochondrial conditions, including the myoclonus epilepsy with ragged-red fibers (MERRF), Kearns–Sayre disease, and Leigh syndrome, can present brain calcifications [84–89]. Of note, since these conditions are caused by mutations in the mitochondrial DNA, patients also present classic mitochondrial features (such as short stature, optic atrophy, hearing loss, ptosis, diabetes mellitus, cardiomyopathy, neuropathy, myopathy, as well as lactic acidosis) and a matrilinear heritability that can help in the differential diagnosis [84–89].

Characteristic patterns of calcifications are found in patients harboring *CLDN5* mutations who present involvement of the vertical pontine white matter bundles, other than cortical and basal ganglia calcifications [90].

Other conditions associated with brain calcifications can present in association with cutaneous abnormalities, as well as dysmorphism, often involving the extremities, and specific inflammatory signatures. Onset of neurological symptoms after initial normal developmental, post-natal microcephaly, sterile pyrexia, chilblain lesions, and characteristic interferon signature in peripheral blood are suggestive of one of the forms of Aicardi–Goutières syndrome [91]. Persistent pancytopenia and dyskeratosis are consistent with mutation of genes involved in controlling telomere length and causing Hoyeraal Hreidarsson syndrome and family of diseases called dyskeratosis congenita that can present with severe childhood onset of progressive neurological deterioration and brain calcifications [92]. Erythematous nodular-like lesions, lipodystrophy, extremity joint deformity, specific inflammatory signature, and hyperpyrexia are found in Nakajo Nishimura syndrome, so far described only in Japanese patients [93]. Frequent association with autoimmune conditions is found in spondyloenchondrodysplasia with immune dysregulation syndrome, while frequent infection and susceptibility to TBC is found in immunodeficiency 38 with basal ganglia calcification syndrome [94,95]. Interstitial lung symptoms are, instead, characteristic of Rajab interstitial lung disease with brain calcifications 1 and 2 [80,96].

Cartilage abnormalities with dysmorphism, such as facial dysmorphism and brachytelephalangism are characteristics of Keutel Syndrome [97]. Dysmorphic features and terminal transverse limb defects can also be found in Adams–Oliver syndrome 2 where brain MRI shows characteristic periventricular calcifications and ventricular enlargement [83,98]. Hydrocephalus and basal ganglia calcifications are also found in male subjects with Fried syndrome [99–101].

Some conditions present a more catastrophic course, as in the case of the *JAM3*-related condition where the vascular brain disease that causes cerebral calcifications is also responsible for lethal hemorrhages [102]. Prominent early onset spastic paraplegia with brain calcifications, white matter abnormalities, and corpus callosum abnormalities are present in spastic paraplegia 56, with characteristic pseudoxanthoma elasticum due to calcium accumulation in the tendons (usually in the neck) [103].

Early onset Parkinsonism and intellectual disability with mild calcifications of the basal ganglia can be found in patients with sporadic or hereditary (X-linked) juvenile Parkinsonism due to mutations of the *RAB39B* gene [104,105]. Adult-onset neurodegeneration with early onset dementia and Parkinsonism is also found in Nasu–Hakola disease where brain atrophy and white matter abnormalities are associated with basal ganglia calcifications and cystic lesions of the bones [106].

Interestingly, genetic mutations have been identified in patients that resemble either congenital infection (pseudo-TORCH 1 and 2) or pseudohypoparathyroidism [107]. Finally, in some cases, brain calcification is due to genetic metabolic conditions that can benefit from prompt recognition and treatment, such as in inborn error of folate metabolism or 3-hydroxyisobutyric aciduria, for alleviating or preventing some of the symptoms [108,109]. Adult brain calcification findings presenting with vascular events (ischemic or hemorrhagic) or migraine are also characteristic of the spectrum of disorders associated with collagen *COL4A1* and 2 mutations [110–112].

5. Conclusions

Brain calcification can represent the epiphenomenon of a large number of systemic and genetic conditions. Associated clinical presentation (such as dysmorphism, cutaneous abnormalities, or immunological traits) can help in the process of differential diagnosis (Table 2). Systemic conditions, such as impaired calcium metabolism, infectious, and autoimmune/inflammatory conditions should always be considered in the diagnostic process, as they are curable. **Table 2. Demographics and common comorbidities associated with brain calcifications syndrome.** The table shows the frequency of certain comorbidities that are found in brain calcifications syndrome that can help in the differential diagnosis. For specific references to each condition, please refer to the main text.

| | Adult Onset | Pediatric | Skin | Skeletal | Autoimmune/ Hematological | Pyrexia |
|---|-----------------------|---------------------------------------|---------------|---------------|------------------------------|--------------|
| | | Onset | Abnormalities | Abnormalities | Abnormalities | |
| Calcium metabolism disorder | \checkmark | \checkmark | | ✓ | | |
| SLE PFBC | ✓ | | | | ✓ | |
| | ✓ | <u> </u> | | | | |
| Aicardi- Goutières syndrome | | <u> </u> | √ | ✓ | \checkmark | ✓ |
| Coats plus syndrome Leukoencephalopathy, cystic, without | | \checkmark | V | V | V | |
| megalencephaly | | \checkmark | | | | |
| Claudin-5 | | \checkmark | | | | |
| Hoyeraal Hreidarsson syndrom | | | \checkmark | \checkmark | \checkmark | |
| Congenital Dyskeratosis | | | | | | |
| (Zinsser-Engman-Cole syndrome) | | \checkmark | \checkmark | \checkmark | ✓ | |
| 3-hydroxyisobutyric aciduria | | \checkmark | | | | |
| Inborn error of folates Metabolism | | · · · · · · · · · · · · · · · · · · · | | | \checkmark | |
| Carbonic anhydrase II deficiency | | | | , | | |
| syndrome | | \checkmark | | \checkmark | | |
| Collagen IV disorders | | \checkmark | | | | |
| Cockayne syndrome | | · · · · · · · · · · · · · · · · · · · | | \checkmark | | |
| Polycystic lipomembranous | | | | | | |
| osteodysplasia with sclerosing | | \checkmark | | √ | | |
| leukoencephalopathy 1 & 2 (Nasu Hakola Disease) | | | | | | |
| Nakajo Nishimura syndrome x | | \checkmark | √ | \checkmark | \checkmark | |
| Adams Oliver syndrome 2 | | \checkmark | \checkmark | | \checkmark | |
| Raine Syndrome | | ✓ | | \checkmark | | |
| Keutel Syndrome | | \checkmark | | \checkmark | | |
| Fried Syndrome | | \checkmark | | | | |
| Hemorrhagic destruction of the brain, subependymal calcification, and cataracts | | \checkmark | | | | |
| Pseudo-TORCH syndrome 1 | | | | | | |
| Pseudo-TORCH syndrome 2 | | √ | | | \checkmark | |
| Immunodeficiency 38 with basal | | | | | v | |
| ganglia calcification | | \checkmark | | | √ | |
| Spondyloenchondrodysplasia with | | | | | | |
| immune dysregulation | | \checkmark | | \checkmark | √ | |
| Rajab interstitial lung disease with | | | | | | |
| brain calcifications | | \checkmark | | | | |
| Seizures, early-onset, with | | | | | | |
| neurodegeneration and brain | | \checkmark | | | | |
| calcification | | | | | | |
| Spastic paraplegia 56, autosomal | | 1 | 1 | | | |
| recessive | | \checkmark | \checkmark | | | |
| Pseudohypoparathyroidism | | \checkmark | | | | |
| Calcium metabolism disorder | \checkmark | \checkmark | | \checkmark | | |
| SLE | \checkmark | | | | \checkmark | |
| PFBC | \checkmark | \checkmark | | | | |
| Aicardi- Goutières syndrome | | \checkmark | √ | √ | \checkmark | \checkmark |
| Coats plus syndrome | | \checkmark | \checkmark | \checkmark | \checkmark | |
| Leukoencephalopathy, cystic, without | | \checkmark | | | | |
| megalencephaly | | | | | | |
| Claudin-5 | | ✓ | | | | |
| Hoyeraal Hreidarsson syndrom | | \checkmark | ✓ | ✓ | \checkmark | |
| Congenital Dyskeratosis | | \checkmark | \checkmark | \checkmark | ✓ | |
| (Zinsser-Engman-Cole syndrome) | | | · · | | | |
| 3-hydroxyisobutyric aciduria | | ✓ | | | | |
| Inborn error of folates Metabolism | | \checkmark | | | √ | ļ |
| Carbonic anhydrase II deficiency syndrome | | \checkmark | | \checkmark | | |
| Collagen IV disorders | | \checkmark | 1 | | | + |
| Cockayne syndrome | | ✓ | | \checkmark | | |
| | | • | | • | 1 | <u> </u> |

| | Adult Onset | Pediatric Onset | Skin Abnormalities | Skeletal Abnormalities | Autoimmune/ Hematological Abnormalities | Pyrexia |
|---|-------------|---------------------------------------|---------------------------------------|---------------------------|---|---------|
| Polycystic lipomembranous osteodysplasia with sclerosing leukoencephalopathy 1 & 2 (Nasu Hakola Disease) | | \checkmark | | \checkmark | | |
| Nakajo Nishimura syndrome x | | \checkmark | | | | |
| Adams Oliver syndrome 2 | | · · · · · · · · · · · · · · · · · · · | · · · · · · · · · · · · · · · · · · · | | · · · · · · · · · · · · · · · · · · · | |
| Raine Syndrome | | \checkmark | | \checkmark | | |
| Keutel Syndrome | | \checkmark | | \checkmark | | |
| Fried Syndrome | | \checkmark | | | | |
| Hemorrhagic destruction of the brain, subependymal calcification, and cataracts | | \checkmark | | | | |
| Pseudo-TORCH syndrome 1 | | \checkmark | | | | |
| Pseudo-TORCH syndrome 2 | | \checkmark | | | \checkmark | |
| Immunodeficiency 38 with basal ganglia calcification | | \checkmark | | | \checkmark | |
| Spondyloenchondrodysplasia with immune dysregulation | | \checkmark | | \checkmark | \checkmark | |
| Rajab interstitial lung disease with brain calcifications | | \checkmark | | | | |
| Seizures, early-onset, with neurodegeneration and brain calcification | | \checkmark | | | | |
| Spastic paraplegia 56, autosomal recessive | | \checkmark | \checkmark | | | |
| Pseudohypoparathyroidism | | \checkmark | | | | |

Table 2. Cont.

There is no strong correlation between the genetic mutations and specific patterns of calcium deposition in the brain. However, additional findings such as brain cysts or prominent white matter abnormalities can be suggestive of specific forms.

Interestingly, many of the genes responsible for these forms are involved in mechanisms of angiogenesis and in the BBB, suggesting common pathways that could be tackled by therapeutic approaches to these conditions. A deepening in the understanding of the genetic architecture and the identification of new genetic forms of brain calcification will be crucial in this direction.

The most common genetic forms of brain calcifications (PFBC) can present with very mild and non-specific symptoms (such as headache or vertigo) and this diagnosis is likely to be overlooked in a large number of subjects. A role for digenic variants in determining the penetrance of the disease and the phenotypic presentation has been suggested by a few works in the literature and it warrants further investigation.

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References

- 1. Manyam, B.V. What Is and What Is Not "Fahr's Disease". Park. Relat. Disord. 2005, 11, 73–80. [CrossRef] [PubMed]
- Saade, C.; Najem, E.; Asmar, K.; Salman, R.; El Achkar, B.; Naffaa, L. Intracranial Calcifications on CT: An Updated Review. J. Radiol. Case Rep. 2019, 13, 1–18. [CrossRef]
- Yamada, M.; Asano, T.; Okamoto, K.; Hayashi, Y.; Kanematsu, M.; Hoshi, H.; Akaiwa, Y.; Shimohata, T.; Nishizawa, M.; Inuzuka, T.; et al. High Frequency of Calcification in Basal Ganglia on Brain Computed Tomography Images in Japanese Older Adults. *Geriatr. Gerontol. Int.* 2013, 13, 706–710. [CrossRef] [PubMed]
- 4. Simoni, M.; Pantoni, L.; Pracucci, G.; Palmertz, B.; Guo, X.; Gustafson, D.; Skoog, I. Prevalence of CT-Detected Cerebral Abnormalities in an Elderly Swedish Population Sample. *Acta Neurol. Scand.* **2008**, *118*, 260–267. [CrossRef] [PubMed]
- 5. Donzuso, G.; Mostile, G.; Nicoletti, A.; Zappia, M. Basal Ganglia Calcifications (Fahr's Syndrome): Related Conditions and Clinical Features. *Neurol. Sci.* 2019, 40, 2251–2263. [CrossRef] [PubMed]
- Westenberger, A.; Balck, A.; Klein, C. Primary familial brain calcifications: Genetic and clinical updates. *Curr. Opin. Neurol.* 2019, 32, 571–578. [CrossRef]
- Ferreira, L.D.; Mendes de Oliveira, J.R. The Need for Consensus on Primary Familial Brain Calcification Nomenclature. J. Neuropsychiatry Clin. Neurosci. 2018, 30, 291–293. [CrossRef]
- 8. Foley, J. Calcification of the Corpus Stiatum and Dentate Nuclei Occurring in a Family. J. Neurol. Neurosurg. Psychiatry 1951, 14, 253–261. [CrossRef]
- 9. Ellie, E.; Julien, J.; Ferrer, X. Familial Idiopathic Striopallidodentate Calcifications. *Neurology* **1989**, *39*, 381–385. [CrossRef] [PubMed]
- 10. Boller, F.; Boller, M.; Gilbert, J. Familial Idiopathic Cerebral Calcifications. J. Neurol. Neurosurg. Psychiatry 1977, 40, 280–285. [CrossRef]
- 11. Moskowitz, M.A.; Winickoff, R.N.; Heinz, E.R. Familial Calcification of the Basal Ganglions: A Metabolic and Genetic Study. *N. Engl. J. Med.* **1971**, *285*, 72–77. [CrossRef] [PubMed]
- Wang, C.; Li, Y.; Shi, L.; Ren, J.; Patti, M.; Wang, T.; de Oliveira, J.R.M.; Sobrido, M.-J.; Quintáns, B.; Baquero, M.; et al. Mutations in SLC20A2 Link Familial Idiopathic Basal Ganglia Calcification with Phosphate Homeostasis. *Nat. Genet.* 2012, 44, 254–256. [CrossRef] [PubMed]
- Schottlaender, L.V.; Abeti, R.; Jaunmuktane, Z.; Macmillan, C.; Chelban, V.; O'Callaghan, B.; McKinley, J.; Maroofian, R.; Efthymiou, S.; Athanasiou-Fragkouli, A.; et al. Bi-Allelic JAM2 Variants Lead to Early-Onset Recessive Primary Familial Brain Calcification. Am. J. Hum. Genet. 2020, 106, 412–421. [CrossRef]
- 14. Yao, X.-P.; Cheng, X.; Wang, C.; Zhao, M.; Guo, X.-X.; Su, H.-Z.; Lai, L.-L.; Zou, X.-H.; Chen, X.-J.; Zhao, Y.; et al. Biallelic Mutations in MYORG Cause Autosomal Recessive Primary Familial Brain Calcification. *Neuron* **2018**, *98*, 1116–1123.e5. [CrossRef]
- Legati, A.; Giovannini, D.; Nicolas, G.; López-Sánchez, U.; Quintáns, B.; Oliveira, J.R.M.; Sears, R.L.; Ramos, E.M.; Spiteri, E.; Sobrido, M.-J.; et al. Mutations in XPR1 Cause Primary Familial Brain Calcification Associated with Altered Phosphate Export. *Nat. Genet.* 2015, 47, 579–581. [CrossRef]
- Nicolas, G.; Pottier, C.; Maltête, D.; Coutant, S.; Rovelet-Lecrux, A.; Legallic, S.; Rousseau, S.; Vaschalde, Y.; Guyant-Maréchal, L.; Augustin, J.; et al. Mutation of the PDGFRB Gene as a Cause of Idiopathic Basal Ganglia Calcification. *Neurology* 2013, *80*, 181–187. [CrossRef] [PubMed]
- Keller, A.; Westenberger, A.; Sobrido, M.J.; García-Murias, M.; Domingo, A.; Sears, R.L.; Lemos, R.R.; Ordoñez-Ugalde, A.; Nicolas, G.; da Cunha, J.E.G.; et al. Mutations in the Gene Encoding PDGF-B Cause Brain Calcifications in Humans and Mice. *Nat. Genet.* 2013, 45, 1077–1082. [CrossRef]
- Zhao, M.; Su, H.-Z.; Zeng, Y.-H.; Sun, Y.; Guo, X.-X.; Li, Y.-L.; Wang, C.; Zhao, Z.-Y.; Huang, X.-J.; Lin, K.-J.; et al. Loss of Function of CMPK2 Causes Mitochondria Deficiency and Brain Calcification. *Cell. Discov.* 2022, *8*, 128. [CrossRef]
- Balck, A.; Schaake, S.; Kuhnke, N.S.; Domingo, A.; Madoev, H.; Margolesky, J.; Dobricic, V.; Alvarez-Fischer, D.; Laabs, B.-H.; Kasten, M.; et al. Genotype-Phenotype Relations in Primary Familial Brain Calcification: Systematic MDSGene Review. *Mov. Disord.* 2021, 36, 2468–2480. [CrossRef] [PubMed]
- Xu, X.; Sun, H.; Luo, J.; Cheng, X.; Lv, W.; Luo, W.; Chen, W.-J.; Xiong, Z.-Q.; Liu, J.-Y. The Pathology of Primary Familial Brain Calcification: Implications for Treatment. *Neurosci. Bull.* 2022, *39*, 659–674. [CrossRef]
- Jensen, N.; Schrøder, H.D.; Hejbøl, E.K.; Füchtbauer, E.-M.; de Oliveira, J.R.M.; Pedersen, L. Loss of Function of Slc20a2 Associated with Familial Idiopathic Basal Ganglia Calcification in Humans Causes Brain Calcifications in Mice. *J. Mol. Neurosci.* 2013, 51, 994–999. [CrossRef] [PubMed]
- Meek, R.W.; Brockerman, J.; Fordwour, O.B.; Zandberg, W.F.; Davies, G.J.; Vocadlo, D.J. The Primary Familial Brain Calcification-Associated Protein MYORG Is an α-Galactosidase with Restricted Substrate Specificity. *PLoS Biol.* 2022, 20, e3001764. [CrossRef]
- Zhao, M.; Lin, X.-H.; Zeng, Y.-H.; Su, H.-Z.; Wang, C.; Yang, K.; Chen, Y.-K.; Lin, B.-W.; Yao, X.-P.; Chen, W.-J. Knockdown of Myorg Leads to Brain Calcification in Zebrafish. *Mol. Brain* 2022, 15, 65. [CrossRef]
- 24. Daneman, R.; Zhou, L.; Kebede, A.A.; Barres, B.A. Pericytes Are Required for Blood-Brain Barrier Integrity during Embryogenesis. *Nature* 2010, *468*, 562–566. [CrossRef] [PubMed]
- Anheim, M.; López-Sánchez, U.; Giovannini, D.; Richard, A.-C.; Touhami, J.; N'Guyen, L.; Rudolf, G.; Thibault-Stoll, A.; Frebourg, T.; Hannequin, D.; et al. XPR1 Mutations Are a Rare Cause of Primary Familial Brain Calcification. J. Neurol. 2016, 263, 1559–1564. [CrossRef] [PubMed]

- Cen, Z.; Chen, Y.; Chen, S.; Wang, H.; Yang, D.; Zhang, H.; Wu, H.; Wang, L.; Tang, S.; Ye, J.; et al. Biallelic Loss-of-Function Mutations in JAM2 Cause Primary Familial Brain Calcification. *Brain* 2020, 143, 491–502. [CrossRef]
- Tonduti, D.; Panteghini, C.; Pichiecchio, A.; Decio, A.; Carecchio, M.; Reale, C.; Moroni, I.; Nardocci, N.; Campistol, J.; Garcia-Cazorla, A.; et al. Encephalopathies with Intracranial Calcification in Children: Clinical and Genetic Characterization. *Orphanet J. Rare Dis.* 2018, 13, 135. [CrossRef]
- Deng, H.; Zheng, W.; Jankovic, J. Genetics and Molecular Biology of Brain Calcification. *Ageing Res. Rev.* 2015, 22, 20–38. [CrossRef]
- Lagrue, E.; Abe, H.; Lavanya, M.; Touhami, J.; Bodard, S.; Chalon, S.; Battini, J.-L.; Sitbon, M.; Castelnau, P. Regional Characterization of Energy Metabolism in the Brain of Normal and MPTP-Intoxicated Mice Using New Markers of Glucose and Phosphate Transport. J. Biomed. Sci. 2010, 17, 91. [CrossRef]
- Steitz, S.A.; Speer, M.Y.; Curinga, G.; Yang, H.Y.; Haynes, P.; Aebersold, R.; Schinke, T.; Karsenty, G.; Giachelli, C.M. Smooth Muscle Cell Phenotypic Transition Associated with Calcification: Upregulation of Cbfa1 and Downregulation of Smooth Muscle Lineage Markers. *Circ. Res.* 2001, *89*, 1147–1154. [CrossRef]
- 31. Schaeffer, S.; Iadecola, C. Revisiting the Neurovascular Unit. Nat. Neurosci. 2021, 24, 1198–1209. [CrossRef] [PubMed]
- 32. Betsholtz, C.; Keller, A. PDGF, Pericytes and the Pathogenesis of Idiopathic Basal Ganglia Calcification (IBGC). *Brain Pathol.* **2014**, 24, 387–395. [CrossRef] [PubMed]
- Jenkinson, E.M.; Rodero, M.P.; Kasher, P.R.; Uggenti, C.; Oojageer, A.; Goosey, L.C.; Rose, Y.; Kershaw, C.J.; Urquhart, J.E.; Williams, S.G.; et al. Mutations in SNORD118 Cause the Cerebral Microangiopathy Leukoencephalopathy with Calcifications and Cysts. *Nat. Genet.* 2016, 48, 1185–1192. [CrossRef] [PubMed]
- Meuwissen, M.E.C.; Schot, R.; Buta, S.; Oudesluijs, G.; Tinschert, S.; Speer, S.D.; Li, Z.; van Unen, L.; Heijsman, D.; Goldmann, T.; et al. Human USP18 Deficiency Underlies Type 1 Interferonopathy Leading to Severe Pseudo-TORCH Syndrome. *J. Exp. Med.* 2016, 213, 1163–1174. [CrossRef] [PubMed]
- 35. Barth, P.G. The Neuropathology of Aicardi-Goutières Syndrome. *Eur. J. Paediatr. Neurol.* 2002, 6 (Suppl. SA), A27–A31; discussion A37–A39, A77–A86. [CrossRef]
- Livingston, J.; Doherty, D.; Orcesi, S.; Tonduti, D.; Piechiecchio, A.; La Piana, R.; Tournier-Lasserve, E.; Majumdar, A.; Tomkins, S.; Rice, G.; et al. COL4A1 Mutations Associated with a Characteristic Pattern of Intracranial Calcification. *Neuropediatrics* 2011, 42, 227–233. [CrossRef] [PubMed]
- Wilson, B.T.; Stark, Z.; Sutton, R.E.; Danda, S.; Ekbote, A.V.; Elsayed, S.M.; Gibson, L.; Goodship, J.A.; Jackson, A.P.; Keng, W.T.; et al. The Cockayne Syndrome Natural History (CoSyNH) Study: Clinical Findings in 102 Individuals and Recommendations for Care. *Genet. Med.* 2016, *18*, 483–493. [CrossRef]
- 38. Finsterer, J.; Kopsa, W. Basal Ganglia Calcification in Mitochondrial Disorders. Metab. Brain Dis. 2005, 20, 219–226. [CrossRef]
- 39. Giachelli, C.M. Ectopic calcification: Gathering hard facts about soft tissue mineralization. *Am. J. Pathol.* **1999**, *154*, 671–675. [CrossRef]
- David, S.; Ferreira, J.; Quenez, O.; Rovelet-Lecrux, A.; Richard, A.-C.; Vérin, M.; Jurici, S.; Le Ber, I.; Boland, A.; Deleuze, J.-F.; et al. Identification of Partial SLC20A2 Deletions in Primary Brain Calcification Using Whole-Exome Sequencing. *Eur. J. Hum. Genet.* 2016, 24, 1630–1634. [CrossRef]
- Hsu, S.C.; Sears, R.L.; Lemos, R.R.; Quintáns, B.; Huang, A.; Spiteri, E.; Nevarez, L.; Mamah, C.; Zatz, M.; Pierce, K.D.; et al. Mutations in SLC20A2 Are a Major Cause of Familial Idiopathic Basal Ganglia Calcification. *Neurogenetics* 2013, 14, 11–22. [CrossRef] [PubMed]
- Yamada, M.; Tanaka, M.; Takagi, M.; Kobayashi, S.; Taguchi, Y.; Takashima, S.; Tanaka, K.; Touge, T.; Hatsuta, H.; Murayama, S.; et al. Evaluation of SLC20A2 Mutations That Cause Idiopathic Basal Ganglia Calcification in Japan. *Neurology* 2014, *82*, 705–712. [CrossRef] [PubMed]
- Lenglez, S.; Sablon, A.; Fénelon, G.; Boland, A.; Deleuze, J.-F.; Boutoleau-Bretonnière, C.; Nicolas, G.; Demoulin, J.-B. Distinct Functional Classes of PDGFRB Pathogenic Variants in Primary Familial Brain Calcification. *Hum. Mol. Genet.* 2022, *31*, 399–409. [CrossRef] [PubMed]
- Chen, W.-J.; Yao, X.-P.; Zhang, Q.-J.; Ni, W.; He, J.; Li, H.-F.; Liu, X.-Y.; Zhao, G.-X.; Murong, S.-X.; Wang, N.; et al. Novel SLC20A2 Mutations Identified in Southern Chinese Patients with Idiopathic Basal Ganglia Calcification. *Gene* 2013, 529, 159–162. [CrossRef] [PubMed]
- Brighina, L.; Saracchi, E.; Ferri, F.; Gagliardi, M.; Tarantino, P.; Morzenti, S.; Musarra, M.; Patassini, M.; Annesi, G.; Ferrarese, C. Fahr's Disease Linked to a Novel SLC20A2 Gene Mutation Manifesting with Dynamic Aphasia. *Neurodegener. Dis.* 2014, 14, 133–138. [CrossRef]
- Pasanen, P.; Mäkinen, J.; Myllykangas, L.; Guerreiro, R.; Bras, J.; Valori, M.; Viitanen, M.; Baumann, M.; Tienari, P.J.; Pöyhönen, M.; et al. Primary Familial Brain Calcification Linked to Deletion of 5' Noncoding Region of SLC20A2. *Acta Neurol. Scand.* 2017, 136, 59–63. [CrossRef]
- Nan, H.; Takaki, R.; Ichinose, Y.; Tsuchiya, M.; Koh, K.; Hanyu, S.; Shindo, K.; Takiyama, Y. Novel SLC20A2 Mutation in Primary Familial Brain Calcification with Disturbance of Sustained Phonation and Orofacial Apraxia. *J. Neurol. Sci.* 2018, 390, 1–3. [CrossRef]
- Coppola, A.; Hernandez-Hernandez, L.; Balestrini, S.; Krithika, S.; Moran, N.; Hale, B.; Cordivari, C.; Sisodiya, S.M. Cortical Myoclonus and Epilepsy in a Family with a New SLC20A2 Mutation. *J. Neurol.* 2020, 267, 2221–2227. [CrossRef]

- Magistrelli, L.; Croce, R.; De Marchi, F.; Basagni, C.; Carecchio, M.; Nasuelli, N.; Cantello, R.; Invernizzi, F.; Garavaglia, B.; Comi, C.; et al. Expanding the Genetic Spectrum of Primary Familial Brain Calcification Due to SLC2OA2 Mutations: A Case Series. *Neurogenetics* 2021, 22, 65–70. [CrossRef]
- Gagliardi, M.; Morelli, M.; Annesi, G.; Nicoletti, G.; Perrotta, P.; Pustorino, G.; Iannello, G.; Tarantino, P.; Gambardella, A.; Quattrone, A. A New SLC20A2 Mutation Identified in Southern Italy Family with Primary Familial Brain Calcification. *Gene* 2015, 568, 109–111. [CrossRef]
- Zhang, X.; Ma, G.; Zhao, Z.; Zhu, M. SCL20A2 Mutation Presenting with Acute Ischemic Stroke: A Case Report. *BMC Neurol.* 2018, 18, 11. [CrossRef]
- 52. Ramos, E.M.; Carecchio, M.; Lemos, R.; Ferreira, J.; Legati, A.; Sears, R.L.; Hsu, S.C.; Panteghini, C.; Magistrelli, L.; Salsano, E.; et al. Primary Brain Calcification: An International Study Reporting Novel Variants and Associated Phenotypes. *Eur. J. Hum. Genet.* **2018**, *26*, 1462–1477. [CrossRef]
- Yao, X.-P.; Wang, C.-; Su, H.-Z.; Guo, X.-X.; Lu, Y.-Q.; Zhao, M.; Liu, Y.-B.; Lai, J.-H.; Chen, H.-T.; Wang, N.; et al. Mutation Screening of PDGFB Gene in Chinese Population with Primary Familial Brain Calcification. *Gene* 2017, 597, 17–22. [CrossRef] [PubMed]
- Nicolas, G.; Jacquin, A.; Thauvin-Robinet, C.; Rovelet-Lecrux, A.; Rouaud, O.; Pottier, C.; Aubriot-Lorton, M.-H.; Rousseau, S.; Wallon, D.; Duvillard, C.; et al. A de Novo Nonsense PDGFB Mutation Causing Idiopathic Basal Ganglia Calcification with Laryngeal Dystonia. *Eur. J. Hum. Genet.* 2014, 22, 1236–1238. [CrossRef] [PubMed]
- 55. Hayashi, T.; Legati, A.; Nishikawa, T.; Coppola, G. First Japanese Family with Primary Familial Brain Calcification Due to a Mutation in the PDGFB Gene: An Exome Analysis Study. *Psychiatry Clin. Neurosci.* **2015**, *69*, 77–83. [CrossRef]
- 56. Biancheri, R.; Severino, M.; Robbiano, A.; Iacomino, M.; Del Sette, M.; Minetti, C.; Cervasio, M.; Caro, M.D.B.D.; Striano, P.; Zara, F. White Matter Involvement in a Family with a Novel PDGFB Mutation. *Neurol. Genet.* **2016**, *2*, e77. [CrossRef] [PubMed]
- 57. Rohani, M.; Poon, Y.-Y.; Naranian, T.; Fasano, A. SCL20A2 Mutation Mimicking Fluctuating Parkinson's Disease. *Park. Relat. Disord.* 2017, 39, 93–94. [CrossRef] [PubMed]
- Sanchez-Contreras, M.; Baker, M.C.; Finch, N.A.; Nicholson, A.; Wojtas, A.; Wszolek, Z.K.; Ross, O.A.; Dickson, D.W.; Rademakers, R. Genetic Screening and Functional Characterization of PDGFRB Mutations Associated with Basal Ganglia Calcification of Unknown Etiology. *Hum. Mutat.* 2014, 35, 964–971. [CrossRef]
- Mathorne, S.W.; Sørensen, K.; Fagerberg, C.; Bode, M.; Hertz, J.M. A Novel PDGFRB Sequence Variant in a Family with a Mild Form of Primary Familial Brain Calcification: A Case Report and a Review of the Literature. *BMC Neurol.* 2019, 19, 60. [CrossRef]
- 60. Chen, Y.; Cen, Z.; Chen, X.; Wang, H.; Chen, S.; Yang, D.; Fu, F.; Wang, L.; Liu, P.; Wu, H.; et al. MYORG Mutation Heterozygosity Is Associated With Brain Calcification. *Mov. Disord.* **2020**, *35*, 679–686. [CrossRef]
- Arkadir, D.; Lossos, A.; Rahat, D.; Abu Snineh, M.; Schueler-Furman, O.; Nitschke, S.; Minassian, B.A.; Sadaka, Y.; Lerer, I.; Tabach, Y.; et al. MYORG Is Associated with Recessive Primary Familial Brain Calcification. *Ann. Clin. Transl. Neurol.* 2019, 6, 106–113. [CrossRef] [PubMed]
- Grangeon, L.; Wallon, D.; Charbonnier, C.; Quenez, O.; Richard, A.-C.; Rousseau, S.; Budowski, C.; Lebouvier, T.; Corbille, A.-G.; Vidailhet, M.; et al. Biallelic MYORG Mutation Carriers Exhibit Primary Brain Calcification with a Distinct Phenotype. *Brain* 2019, 142, 1573–1586. [CrossRef] [PubMed]
- Uno, A.; Tamune, H.; Kurita, H.; Hozumi, I.; Yamamoto, N. SLC20A2-Associated Idiopathic Basal Ganglia Calcification-Related Recurrent Psychosis Response to Low-Dose Antipsychotics: A Case Report and Literature Review. *Cureus* 2020, 12, e12407. [CrossRef] [PubMed]
- 64. Bu, W.; Hou, L.; Zhu, M.; Zhang, R.; Zhang, X.; Zhang, X.; Tang, J.; Liu, X. SLC20A2-Related Primary Familial Brain Calcification with Purely Acute Psychiatric Symptoms: A Case Report. *BMC Neurol.* **2022**, *22*, 265. [CrossRef]
- Li, M.; Fu, Q.; Xiang, L.; Zheng, Y.; Ping, W.; Cao, Y. SLC20A2-Associated Idiopathic Basal Ganglia Calcification (Fahr Disease): A Case Family Report. BMC Neurol. 2022, 22, 438. [CrossRef]
- 66. Soh, D.; Lang, A. Diffuse Brain Calcification, a Novel SLC20A2 Variant, Vertical Supranuclear Gaze Palsy, and Systemic Lupus Erythematosus. *Mov. Disord. Clin. Pract.* 2019, *6*, 403–405. [CrossRef]
- 67. Kuroi, Y.; Akagawa, H.; Yoneyama, T.; Kikuchi, A.; Maegawa, T.; Onda, H.; Kasuya, H. Novel SLC20A2 Mutation in a Japanese Pedigree with Primary Familial Brain Calcification. *J. Neurol. Sci.* **2019**, *399*, 183–185. [CrossRef]
- Fjaer, R.; Brodtkorb, E.; Øye, A.-M.; Sheng, Y.; Vigeland, M.D.; Kvistad, K.A.; Backe, P.H.; Selmer, K.K. Generalized Epilepsy in a Family with Basal Ganglia Calcifications and Mutations in SLC20A2 and CHRNB2. *Eur. J. Med. Genet.* 2015, *58*, 624–628. [CrossRef]
- 69. Knowles, J.K.; Santoro, J.D.; Porter, B.E.; Baumer, F.M. Refractory Focal Epilepsy in a Paediatric Patient with Primary Familial Brain Calcification. *Seizure* **2018**, *56*, 50–52. [CrossRef]
- Mu, W.; Tochen, L.; Bertsch, C.; Singer, H.S.; Barañano, K.W. Intracranial Calcifications and Dystonia Associated with a Novel Deletion of Chromosome 8p11.2 Encompassing SLC20A2 and THAP1. *BMJ Case Rep.* 2019, 12, e228782. [CrossRef]
- Wszolek, Z.K.; Baba, Y.; Mackenzie, I.R.; Uitti, R.J.; Strongosky, A.J.; Broderick, D.F.; Baker, M.C.; Melquist, S.; Hutton, M.L.; Tsuboi, Y.; et al. Autosomal Dominant Dystonia-plus with Cerebral Calcifications. *Neurology* 2006, 67, 620–625. [CrossRef] [PubMed]

- Baker, M.; Strongosky, A.J.; Sanchez-Contreras, M.Y.; Yang, S.; Ferguson, W.; Calne, D.B.; Calne, S.; Stoessl, A.J.; Allanson, J.E.; Broderick, D.F.; et al. SLC20A2 and THAP1 Deletion in Familial Basal Ganglia Calcification with Dystonia. *Neurogenetics* 2014, 15, 23–30. [CrossRef]
- Fujioka, S.; Strongosky, A.J.; Hassan, A.; Rademakers, R.; Dickson, D.W.; Wszolek, Z.K. Clinical Presentation of a Patient with SLC20A2 and THAP1 Deletions: Differential Diagnosis of Oromandibular Dystonia. *Parkinsonism Relat. Disord.* 2015, 21, 329–331. [CrossRef] [PubMed]
- DeMeo, N.N.; Burgess, J.D.; Blackburn, P.R.; Gass, J.M.; Richter, J.; Atwal, H.K.; van Gerpen, J.A.; Atwal, P.S. Co-Occurrence of a Novel PDGFRB Variant and Likely Pathogenic Variant in CASR in an Individual with Extensive Intracranial Calcifications and Hypocalcaemia. *Clin. Case Rep.* 2018, 6, 8–13. [CrossRef] [PubMed]
- Schouten, B.J.; Raizis, A.M.; Soule, S.G.; Cole, D.R.; Frengley, P.A.; George, P.M.; Florkowski, C.M. Four Cases of Autosomal Dominant Hypocalcaemia with Hypercalciuria Including Two with Novel Mutations in the Calcium-Sensing Receptor Gene. *Ann. Clin. Biochem.* 2011, 48, 286–290. [CrossRef] [PubMed]
- Sun, H.; Cao, Z.; Gao, R.; Li, Y.; Chen, R.; Du, S.; Ma, T.; Wang, J.; Xu, X.; Liu, J.Y. Severe Brain Calcification and Migraine Headache Caused by SLC20A2 and PDGFRB Heterozygous Mutations in a Five-Year-Old Chinese Girl. *Mol. Genet. Genom. Med.* 2021, 9, e1670. [CrossRef]
- Crow, Y.J.; Marshall, H.; Rice, G.I.; Seabra, L.; Jenkinson, E.M.; Baranano, K.; Battini, R.; Berger, A.; Blair, E.; Blauwblomme, T.; et al. Leukoencephalopathy with Calcifications and Cysts: Genetic and Phenotypic Spectrum. *Am. J. Med. Genet. A* 2021, 185, 15–25. [CrossRef]
- Anderson, B.H.; Kasher, P.R.; Mayer, J.; Szynkiewicz, M.; Jenkinson, E.M.; Bhaskar, S.S.; Urquhart, J.E.; Daly, S.B.; Dickerson, J.E.; O'Sullivan, J.; et al. Mutations in CTC1, Encoding Conserved Telomere Maintenance Component 1, Cause Coats Plus. *Nat. Genet.* 2012, 44, 338–342. [CrossRef]
- Smith, C.; McColl, B.W.; Patir, A.; Barrington, J.; Armishaw, J.; Clarke, A.; Eaton, J.; Hobbs, V.; Mansour, S.; Nolan, M.; et al. Biallelic Mutations in NRROS Cause an Early Onset Lethal Microgliopathy. *Acta Neuropathol.* 2020, 139, 947–951. [CrossRef]
- 80. Krenke, K.; Szczałuba, K.; Bielecka, T.; Rydzanicz, M.; Lange, J.; Koppolu, A.; Płoski, R. FARSA Mutations Mimic Phenylalanyl-TRNA Synthetase Deficiency Caused by FARSB Defects. *Clin. Genet.* **2019**, *96*, 468–472. [CrossRef]
- Tonduti, D.; Orcesi, S.; Jenkinson, E.M.; Dorboz, I.; Renaldo, F.; Panteghini, C.; Rice, G.I.; Henneke, M.; Livingston, J.H.; Elmaleh, M.; et al. Clinical, Radiological and Possible Pathological Overlap of Cystic Leukoencephalopathy without Megalencephaly and Aicardi-Goutières Syndrome. *Eur. J. Paediatr. Neurol.* 2016, 20, 604–610. [CrossRef] [PubMed]
- 82. Laugel, V. Cockayne Syndrome. In *GeneReviews®*; Adam, M.P., Everman, D.B., Mirzaa, G.M., Pagon, R.A., Wallace, S.E., Bean, L.J., Gripp, K.W., Amemiya, A., Eds.; University of Washington, Seattle: Seattle, WA, USA, 1993.
- Sukalo, M.; Tilsen, F.; Kayserili, H.; Müller, D.; Tüysüz, B.; Ruddy, D.M.; Wakeling, E.; Ørstavik, K.H.; Snape, K.M.; Trembath, R.; et al. DOCK6 Mutations Are Responsible for a Distinct Autosomal-Recessive Variant of Adams-Oliver Syndrome Associated with Brain and Eye Anomalies. *Hum. Mutat.* 2015, *36*, 593–598. [CrossRef] [PubMed]
- Federico, A.; Cornelio, F.; Di Donato, S.; Ederli, E.; Fabrizi, G.M.; Manneschi, L.; Guazzi, G.C. Mitochondrial Encephalo-Neuro-Myopathy with Myoclonus Epilepsy, Basal Nuclei Calcification and Hyperlactacidemia. *Ital. J. Neurol. Sci.* 1988, 9, 65–71. [CrossRef]
- 85. Fukuhara, N.; Tokiguchi, S.; Shirakawa, K.; Tsubaki, T. Myoclonus Epilepsy Associated with Ragged-Red Fibres (Mitochondrial Abnormalities): Disease Entity or a Syndrome? Light-and Electron-Microscopic Studies of Two Cases and Review of Literature. *J. Neurol. Sci.* **1980**, *47*, 117–133. [CrossRef] [PubMed]
- 86. Hart, Z.H.; Chang, C.H.; Perrin, E.V.; Neerunjun, J.S.; Ayyar, R. Familial Poliodystrophy, Mitochondrial Myopathy, and Lactate Acidemia. *Arch. Neurol.* **1977**, *34*, 180–185. [CrossRef]
- 87. Kuriyama, M.; Umezaki, H.; Fukuda, Y.; Osame, M.; Koike, K.; Tateishi, J.; Igata, A. Mitochondrial Encephalomyopathy with Lactate-Pyruvate Elevation and Brain Infarctions. *Neurology* **1984**, *34*, 72–77. [CrossRef]
- Markesbery, W.R. Lactic Acidemia, Mitochondrial Myopathy, and Basal Ganglia Calcification. *Neurology* 1979, 29, 1057–1060. [CrossRef]
- 89. Morgan-Hughes, J.A.; Hayes, D.J.; Clark, J.B.; Landon, D.N.; Swash, M.; Stark, R.J.; Rudge, P. Mitochondrial Encephalomyopathies: Biochemical Studies in Two Cases Revealing Defects in the Respiratory Chain. *Brain* **1982**, *105 Pt 3*, 553–582. [CrossRef]
- Deshwar, A.R.; Cytrynbaum, C.; Murthy, H.; Zon, J.; Chitayat, D.; Volpatti, J.; Newbury-Ecob, R.; Ellard, S.; Lango Allen, H.; Yu, E.P.; et al. Variants in CLDN5 Cause a Syndrome Characterized by Seizures, Microcephaly and Brain Calcifications. *Brain* 2022, awac461. [CrossRef]
- 91. Crow, Y.J. Aicardi-Goutières Syndrome. In *GeneReviews*[®]; Adam, M.P., Everman, D.B., Mirzaa, G.M., Pagon, R.A., Wallace, S.E., Bean, L.J., Gripp, K.W., Amemiya, A., Eds.; University of Washington, Seattle: Seattle, WA, USA, 1993.
- 92. Zhang, M.-J.; Cao, Y.-X.; Wu, H.-Y.; Li, H.-H. Brain Imaging Features of Children with Hoyeraal-Hreidarsson Syndrome. *Brain Behav.* 2021, *11*, e02079. [CrossRef]
- Ohmura, K. Nakajo-Nishimura Syndrome and Related Proteasome-Associated Autoinflammatory Syndromes. J. Inflamm. Res. 2019, 12, 259–265. [CrossRef] [PubMed]
- Briggs, T.A.; Rice, G.I.; Adib, N.; Ades, L.; Barete, S.; Baskar, K.; Baudouin, V.; Cebeci, A.N.; Clapuyt, P.; Coman, D.; et al. Spondyloenchondrodysplasia Due to Mutations in ACP5: A Comprehensive Survey. J. Clin. Immunol. 2016, 36, 220–234. [CrossRef] [PubMed]

- 95. Zhang, X.; Bogunovic, D.; Payelle-Brogard, B.; Francois-Newton, V.; Speer, S.D.; Yuan, C.; Volpi, S.; Li, Z.; Sanal, O.; Mansouri, D.; et al. Human Intracellular ISG15 Prevents Interferon-α/β over-Amplification and Auto-Inflammation. *Nature* 2015, 517, 89–93. [CrossRef] [PubMed]
- Zadjali, F.; Al-Yahyaee, A.; Al-Nabhani, M.; Al-Mubaihsi, S.; Gujjar, A.; Raniga, S.; Al-Maawali, A. Homozygosity for FARSB Mutation Leads to Phe-TRNA Synthetase-Related Disease of Growth Restriction, Brain Calcification, and Interstitial Lung Disease. *Hum. Mutat.* 2018, 39, 1355–1359. [CrossRef]
- 97. Cancela, M.L.; Laizé, V.; Conceição, N.; Kempf, H.; Murshed, M. Keutel Syndrome, a Review of 50 Years of Literature. *Front. Cell. Dev. Biol.* **2021**, *9*, 642136. [CrossRef] [PubMed]
- 98. Dehdashtian, A.; Dehdashtian, M. Adams-Oliver Syndrome: A Case with Full Expression. *Pediatr. Rep.* **2016**, *8*, 6517. [CrossRef] [PubMed]
- 99. Fried, K. X-Linked Mental Retardation and-or Hydrocephalus. Clin. Genet. 1972, 3, 258–263. [CrossRef]
- 100. Fried, K.; Sanger, R. Possible Linkage between Xg and the Locus for a Gene Causing Mental Retardation with or without Hydrocephalus. *J. Med. Genet.* **1973**, *10*, 17–18. [CrossRef]
- 101. Saillour, Y.; Zanni, G.; Des Portes, V.; Heron, D.; Guibaud, L.; Iba-Zizen, M.T.; Pedespan, J.L.; Poirier, K.; Castelnau, L.; Julien, C.; et al. Mutations in the AP1S2 Gene Encoding the Sigma 2 Subunit of the Adaptor Protein 1 Complex Are Associated with Syndromic X-Linked Mental Retardation with Hydrocephalus and Calcifications in Basal Ganglia. *J. Med. Genet.* 2007, 44, 739–744. [CrossRef]
- 102. Akawi, N.A.; Canpolat, F.E.; White, S.M.; Quilis-Esquerra, J.; Morales Sanchez, M.; Gamundi, M.J.; Mochida, G.H.; Walsh, C.A.; Ali, B.R.; Al-Gazali, L. Delineation of the Clinical, Molecular and Cellular Aspects of Novel JAM3 Mutations Underlying the Autosomal Recessive Hemorrhagic Destruction of the Brain, Subependymal Calcification, and Congenital Cataracts. *Hum. Mutat.* 2013, 34, 498–505. [CrossRef]
- 103. Durand, C.M.; Dhers, L.; Tesson, C.; Tessa, A.; Fouillen, L.; Jacqueré, S.; Raymond, L.; Coupry, I.; Benard, G.; Darios, F.; et al. CYP2U1 Activity Is Altered by Missense Mutations in Hereditary Spastic Paraplegia 56. *Hum. Mutat.* 2018, 39, 140–151. [CrossRef]
- 104. Shi, C.-H.; Zhang, S.-Y.; Yang, Z.-H.; Yang, J.; Shang, D.-D.; Mao, C.-Y.; Liu, H.; Hou, H.-M.; Shi, M.-M.; Wu, J.; et al. A Novel RAB39B Gene Mutation in X-Linked Juvenile Parkinsonism with Basal Ganglia Calcification. *Mov. Disord.* 2016, 31, 1905–1909. [CrossRef]
- 105. Ciammola, A.; Carrera, P.; Di Fonzo, A.; Sassone, J.; Villa, R.; Poletti, B.; Ferrari, M.; Girotti, F.; Monfrini, E.; Buongarzone, G.; et al. X-Linked Parkinsonism with Intellectual Disability Caused by Novel Mutations and Somatic Mosaicism in RAB39B Gene. *Park. Relat. Disord.* 2017, 44, 142–146. [CrossRef]
- 106. Bianchin, M.M.; Martin, K.C.; de Souza, A.C.; de Oliveira, M.A.; de Mello Rieder, C.R. Nasu-Hakola Disease and Primary Microglial Dysfunction. *Nat. Rev. Neurol.* **2010**, *6*, 523. [CrossRef]
- 107. Gonçalves, F.G.; Caschera, L.; Teixeira, S.R.; Viaene, A.N.; Pinelli, L.; Mankad, K.; Alves, C.A.P.F.; Ortiz-Gonzalez, X.R.; Andronikou, S.; Vossough, A. Intracranial Calcifications in Childhood: Part 2. *Pediatr. Radiol.* **2020**, *50*, 1448–1475. [CrossRef]
- Pope, S.; Artuch, R.; Heales, S.; Rahman, S. Cerebral Folate Deficiency: Analytical Tests and Differential Diagnosis. J. Inherit. Metab. Dis. 2019, 42, 655–672. [CrossRef]
- Chitayat, D.; Meagher-Villemure, K.; Mamer, O.A.; O'Gorman, A.; Hoar, D.I.; Silver, K.; Scriver, C.R. Brain Dysgenesis and Congenital Intracerebral Calcification Associated with 3-Hydroxyisobutyric Aciduria. J. Pediatr. 1992, 121, 86–89. [CrossRef]
- 110. Plaisier, E.; Ronco, P. COL4A1-Related Disorders. In *GeneReviews[®]*; Adam, M.P., Everman, D.B., Mirzaa, G.M., Pagon, R.A., Wallace, S.E., Bean, L.J., Gripp, K.W., Amemiya, A., Eds.; University of Washington, Seattle: Seattle, WA, USA, 1993.
- 111. Verbeek, E.; Meuwissen, M.E.C.; Verheijen, F.W.; Govaert, P.P.; Licht, D.J.; Kuo, D.S.; Poulton, C.J.; Schot, R.; Lequin, M.H.; Dudink, J.; et al. COL4A2 Mutation Associated with Familial Porencephaly and Small-Vessel Disease. *Eur. J. Hum. Genet.* 2012, 20, 844–851. [CrossRef]
- 112. Guey, S.; Hervé, D. Main Features of COL4A1–COL4A2 Related Cerebral Microangiopathies. *Cereb. Circ. Cogn. Behav.* 2022, 3, 100140. [CrossRef]

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