

Genotypic and allelic frequencies of progressive rod-cone degeneration and other main variants associated with progressive retinal atrophy in Italian dogs

Sara Ghilardi ¹ 💿 Mara	a Bagardi ¹ 💿 🕴 St	efano Frattini ²	Giulia E. Barbariga ¹
Paola G. Brambilla ¹ 🗅 🛛	Giulietta Minozzi ¹	🛯 💿 🕴 Michele Po	lli ¹ 💿

¹Department of Veterinary Medicine and Animal Sciences—DIVAS, University of Milan, Lodi, Italy

²Vetogene Laboratory—ENCI Servizi SRL, Milan, Italy

Correspondence

Giulietta Minozzi, Department of Veterinary Medicine and Animal Sciences—DIVAS, University of Milan, Lodi 26900, Italy. Email: giulietta.minozzi@unimi.it

Funding information APC funds of the University of Milan

Abstract

Background: Progressive retinal atrophy (PRA) is a group of canine inherited retinal disorders affecting up to 100 breeds. Genetic tests are available. The aim of this study was to retrospectively evaluate the genetic variants associated with PRA among dogs residing in Italy.

Methods: Genetic data of 20 variants associated with different forms of PRA were collected through DNA tests over a 10-year period for several dog breeds in the Italian canine population. Allelic and genotypic frequencies were calculated.

Results: A total of 1467 DNA tests were conducted for 1180 dogs. Progressive rod-cone degeneration (PRCD) was the most tested form of PRA, with 58.15% (n = 853) of the DNA tests. Among the widespread breeds in Italy, Labrador retrievers and toy poodles showed a prevalence of heterozygous carriers higher than 15%. Among the others, 175 DNA tests for golden retrievers (GR) showed a prevalence of heterozygous carriers of 13.04% (n = 12) for GR-PRA1 and 8.43% (n = 7) for GR-PRA2. The zwergschnauzer breed was tested for the type B and/or the type B1 forms of PRA with 25.32% (n = 20) heterozygous carriers and 0%, respectively.

Conclusion: The study offers an overview of the prevalence of PRCD and other PRA forms within some of the most popular breeds in Italy.

INTRODUCTION

Progressive retinal atrophy (PRA) is a heterogeneous group of inherited retinal disorders affecting up to 100 canine breeds and has been shown to be homologous with retinitis pigmentosa in humans.^{1–5} The disease is characterised by a primary degeneration of rod photoreceptors followed by the degeneration of cones. This initially causes night blindness, which then progresses to severe visual dysfunction under both dim and bright light; finally, the degeneration of cones can lead to total blindness.³ Some genetic variants responsible for PRA can lead to rod degeneration only, while others can involve cones as well; however, an earlier and more severe loss of rods represents the peculiar PRA phenotype.¹ Affected dogs show changes in the fundus in an ophthalmologic examination, such as hyperreflectivity of the tapetum, attenuation of retinal blood vessels and atrophy of the optic disk. At the early stage of the disease, electroretinography can be useful to distinguish PRA from other ocular diseases.^{1,6} Development of secondary cataracts in the late stage of the disease is often subsequently diagnosed.^{1,7}

While clinical signs are similar among various forms of PRA, the aetiology, age of onset and rate of progression vary between and within breeds.⁸ The early-onset form manifests during the postnatal retinal differentiation process and results from retinal degeneration or abnormal/interrupted retinal development; the progression rate of this form is fast and affected dogs usually manifest clinical signs at a young age. Late-onset PRA, on the contrary, is characterised by a retinal degeneration that starts after the maturation of the retina itself is complete; it shows a slow progression rate and clinical signs may not be manifest until later in life. Usually, it occurs because of defects in the correct maintenance of photoreceptors' normal function.¹

Various modes of inheritance have been described for PRA including autosomal recessive, autosomal dominant and X-linked; the most common mode of inheritance is recessive.^{2,3} Over 20 genes have been identified in PRA across different breeds with a wide range of genetic alleles reported.^{9–17} Genetic tests are available for the detection of various forms of PRA and are able to detect affected/at-risk dogs, heterozygous carriers and clear subjects.¹⁸

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

^{© 2023} The Authors. Veterinary Record Open published by John Wiley & Sons Ltd on behalf of British Veterinary Association.

The principal aim of this study was to retrospectively evaluate allele and genotype frequencies of different PRAassociated variants in Italian dogs over a 10-year period. This included assessing, for each dog, the genotypic heterozygosis or homozygosis, in order to provide a prevalence for the clear dogs, as well as of the affected/at-risk subjects and the heterozygous carriers.

MATERIALS AND METHODS

Ethical approval statement

All animals included in the study were owned by private owners or breeders, who provided written consent. Blood samples were collected by an authorised veterinarian.

Description of the study

This was a retrospective observational study. All DNA tests and associated results for the detection of PRA were provided by a commercial laboratory, Vetogene Laboratory (VL), between 2010 and 2021. The DNA tests were conducted on samples from dogs from all over Italy. The VL is one of the official reference laboratories for the execution of DNA tests for the Italian Kennel Club Ente Nazionale Cinofilia Italiana (ENCI). The VL received blood samples from dog owners, veterinarians and breeders who wanted to conduct the DNA test for PRA. Blood samples were officially collected by veterinarians into EDTA tubes and stored at +4°C for a maximum of 4–5 days prior to examination; each sample was accompanied by a certification from the veterinarian. Some of the dogs were tested for only one form of PRA, while others were tested for two or three forms of the condition. For each dog, breed and DNA test results (clear, heterozygous carrier, affected/at risk) were recorded.

DNA extraction and analysis

The DNA extraction was obtained from 100 to 200 μ L of the blood samples using the commercial Qiagen DNeasy Blood & Tissue kit (Qiagen, Hilden, Germany). Table 1 lists the 20 forms of PRA, with an associated publication reference that describes the procedures carried out by the VL to identify the PRA mutations for each specific gene. All samples were analysed by real-time PCR methods for each specific gene mutation, as listed in Table 1. Each genotype was generated with a single test, performing a single amplification for each mutation. The genotyping was validated directly on the sequence, looking at the mutation using the Sanger sequencing method.

Statistical analysis

Analyses were conducted using SAS 9.4 software (SAS Inc.). Initially, frequencies of clear, heterozygous carriers and affected/at-risk dogs were recorded according to the breed. Afterwards, only the PRA forms for groups of breeds that

counted more than 15 subjects were analysed. For these, allelic and genotypic frequencies were evaluated through the PROC FREQ and PROC ALLELE analyses offered with SAS.

RESULTS

A total of 1467 DNA tests for PRA were conducted for 1180 dogs: 81.78% (n = 965) were tested for one form of PRA, 12.12% (n = 143) for two forms and 6.10% (n = 72) for three forms. Absolute and relative frequencies of clear, heterozygous carriers and affected/at-risk dogs divided according to the PRA form are given in Table 2. Only frequencies for the seven forms with more than 45 DNA tests are reported for statistical relevance (further results are given in Table S1). In 85.07% of the tested dogs, the test result was clear for the gene (n = 1248), with 13.84% heterozygous carriers (n = 203) and 1.09% of dogs deemed affected/at risk (n = 16).

Progressive rod-cone degeneration (PRCD) was the most common test performed for PRA, with 58.15% (n = 853) of the DNA tests. The rod-cone dysplasia (RCD)1 form presented with the highest number of heterozygous carriers (34.78%) and affected/at-risk subjects (6.52%) among the PRA forms. The RCD1 was only tested in the Irish setter, given a previous study.²¹

Fifty-five breeds were tested and, in line with breeddistribution data reported for Italy by ENCI, the five most represented breeds in our population were golden retriever (19%), toy poodle (15%), zwergschnauzer (9%), Labrador retriever (9%) and the Australian shepherd dog (8%). The remaining dogs belonged to 50 different breeds, with each of them having a prevalence lower than 5%. A detailed analysis of the five breeds showed that PRCD was the PRA form with the highest number of heterozygous carriers (20% toy poodle, 15.79% Labrador retriever and 3.33% Australian shepherd dog). The samples from golden retriever dogs showed an absence of the PRCD risk allele, with all 104 tested subjects being clear. The zwergschnauzer-related samples were not tested for PRCD; all subjects in this breed were tested for the type B and/or the type B1 forms of PRA, with 25.32% heterozygous carriers for type B and no positive results for type B1.

Progressive rod-cone degeneration was tested in 34 different breeds. Table 3 summarises the number of clear, heterozygous carriers and PRCD-affected/at-risk dogs for each breed, considering only the breeds in which more than 15 dogs were tested (n = 9). Six of these breeds presented with heterozygous carrier percentages higher than 15%. The Australian cattle dog had a high number of PRCD heterozygous carriers (33.90%) and affected/at-risk subjects (3.39%) compared with other breeds. A further 20 breeds were tested for PRCD (n = 160) with none positive for heterozygous carrier status or affected/at-risk dogs; they are not reported in Table 3.

The DNA tests for golden retriever-(GR)-PRA1 and GR-PRA2 were only conducted on samples from golden retrievers because the disease is only reported in this breed.^{19,20} Ninetytwo dogs were tested for GR-PRA1 and 13.04% (n = 12) were heterozygous carriers of the *SLC4A3* gene mutation. For GR-PRA2, 83 dogs were tested and 8.43% (n = 7) were heterozygous carriers of the *TTC8* risk allele. None of the tested ~~~~

TABLE 1 Identified genes for progressive retinal atrophy (PRA) in dogs, description of the disease and list of affected/at-risk breeds.

variant ID	OMIA phenotype ID	canine chromosome)	Tested variant ^a	Disease and transmission	Affected/at-risk breed(s)	Publication reference
76	OMIA001298-9615	PRCD (CFA9)	g.4188663C>T	PRCD (Progressive rod-cone degeneration) ^b	Various number of breed and mixed breeds	14
575	OMIA001572-9615	SLC4A3 (CFA37)	g.26145752_26145753insC	Golden retriever PRA1 ^b	Golden retriever	19
949	OMIA001984-9615	TTC8 (CFA8)	g.60090186del	Golden retriever PRA2 ^b	Golden retriever	20
282	OMIA000882-9615	PDE6B (CFA3)	g.91747713C>T	RCD1 (Rod-cone dysplasia) ^b	Irish setter	21
528	OMIA001674-9615	PDE6B (CFA3)	g.91747713C>T	RCD1b ^b	American Staffordshire terrier	22
710	OMIA:001260-9615	RD3 (CFA7)	Insertion_22 bp	RCD2 ^b	Collie	11
475	OMIA001314-9615	PDE6A (CFA4)	g.59145362del	RCD3 ^b	Cardigan Welsh corgi	12
583	OMIA:001575-9615	C2orf71 (CFA37)	c.3149_3150insC	RCD4 ^b	Gordon setter, Irish setter	23
359	OMIA001876-9615	SAG (CFA25)	g.44843440T>C	PRA ^b	Basenji	24
547	OMIA001977-9615	CNGA1 (CFA13)	g.43831897_43831900del	PRA ^b	Shetland sheepdog	25
918	OMIA000830-9615	CNGB1 (CFA2)	g.58622673_58622675- delinsCTAGCTAC	PRA ^b	Papillon, phalène dog	26
699	OMIA001432-9615	RPGRIP1 (CFA15)	g.18332036_18332037ins	CORD1 (Cone-rod dystrophy) ^b	Miniature longhaired dachshund	27
634	OMIA001455-9615	NPHP4 (CFA5)	g.59912991_59913168del	CORD2 ^b	Standard wirehaired dachshund	28
574	OMIA001521-9615	CCDC66 (CFA20)	g.33745452_33745453insT	Generalised PRA ^b	Schapendoes, Portuguese water dog	29,30
29	OMIA001346-9615	RHO (CFA20)	g.5637394G>C	Autosomal dominant PRA ^c	English mastiff, bullmastiff	13
480	OMIA000831-9615	RPGR (CFAX)	g.33126490_33126494del	X-linked PRA	Siberian husky, samoyed	17
641	OMIA001523-9615	<i>COL9A2</i> (CFA15)	Deletion_1267 bp	Oculoskeletal dysplasia	Labrador retriever, samoyed	31
		PDC (CFA7)	c.244C>G p.Arg82Gly	PRA type A	Miniature schnauzer	32
1068	OMIA:001311-9615	PPT1 (CFA15)	g.2874661_2875048- con2877563_2877607inv	PRA type B	Miniature schnauzer	33
1170	OMIA:001311-9615	HIVEP3 (CFA15)	g.1432293G>A	PRA type B1	Miniature schnauzer	33

Note: Only PRA forms evaluated in the present study are reported. Online Mendelian Inheritance in Animals (OMIA) identification numbers are provided for each phenotype and variant where available.

^aAll variant positions are referred to the CanFam3.1 genome build, except for X-linked PRA, which refers to the ROS_Cfam 1.0 genome build.

^bAutosomal recessive.

^cAutosomal dominant.

golden retrievers were considered to be affected by/at risk of developing either disease.

The allelic frequencies for the most widespread forms of PRA are reported in Table S2.

DISCUSSION

To the best of the authors' knowledge, this is the first study of the allelic and genotypic frequencies of PRA for dogs drawn from the Italian canine population over a 10-year period. An interesting result is the large number of DNA tests conducted, indicating that genetic testing is considered a useful tool by veterinarians and breeders to detect heterozygous carriers and affected/at-risk subjects. The results of our study detected a high percentage of PRA risk allele carriers in the Italian population (13.84% of heterozygous carriers), a percentage that increased when analysing only the Irish setter for the RCD1 form (34.78% of heterozygous carriers). These frequencies represent a great concern, especially because most of the DNA tests were conducted on dogs, which are used as breeding animals. The RCD1 is a late-onset form of PRA; however, it should be suggested to breeders to conduct a DNA test on Irish setters in the first years of life because dogs are used for reproduction from a young age (2 years old), so the mating of untested pairs represents a great risk.

The results showed that only 1.09% of the tested dogs were affected by/at risk of manifesting PRA. This result may be because veterinarians and breeders do not require a DNA test in a dog with a clinical diagnosis of PRA. The Italian Kennel Club demands a clinical certification of PRA exemption in some breeds to allow the dog into specific competitions; consequently, these dogs are presented for an ophthalmological evaluation from a young age (6–8 months) and could possibly be diagnosed with early-onset forms of PRA. However, an early ophthalmological examination does not exclude the late-onset forms of PRA, such as PRCD, which appears at an adult-old age. Therefore, DNA testing in these cases is imperative to identify heterozygous carriers and affected/atrisk dogs. Thus, the limited use of DNA tests in late-onset

TABLE 2	Absolute and relative frequencies of clear, heterozygous carriers and affected/at-risk Italian dogs of the forms of progressive retinal atrophy
(PRA) for which	ch more than 45 DNA test results were collected.

Gene	Clear	Heterozygous/carrier	Affected/at risk	Total
Progressive Rod-Cone	717	128	8	853
Degeneration	84.06%	15.01%	0.94%	
Golden retriever PRA1	80	12	0	92
	86.96%	13.04%	0%	
Golden retriever PRA2	76	7	0	83
	91.57%	8.43%	0%	
Rod-cone dysplasia - RCD1	27	16	3	46
	58.7%	34.78%	6.52%	
Cone-rod dystrophy - CORD1	37	10	2	49
	75.51%	20.41%	4.08%	
PRA type B	58	20	1	79
	73.42%	25.32%	1.27%	
RCD4	90	5	1	96
	93.75%	5.21%	1.04%	
Other ^a	163	5	1	169
	96.45%	2.95%	0.60%	
Total	1248	203	16	1467
	85.07%	13.84%	1.09%	

^aForms of PRA with less than 45 DNA tests included PRA type A (*n* = 31), CORD2 (*n* = 30), RCD3 (*n* = 23), X-linked PRA (*n* = 19), PRA (*CNGA1*) (*n* = 14), RCD2 (*n* = 14), PRA type B1 (*n* = 12), oculoskeletal dysplasia (*n* = 9), PRA (*CNGB1*) (*n* = 6), RCD1b (*n* = 5), PRA (*SAG*) (*n* = 2), generalised PRA (*n* = 2) and autosomal dominant PRA (*n* = 2) (see Table S1).

TABLE 3	Absolute and	relative f	requencies of	f clear, l	neterozygous	carriers and	l progressive r	od-cone	degeneration	(PRCD)	-affected/	at-risk It	alian dogs,
divided accord	ling to breed.												

	Clear	Heterozygous carrier	Affected/at risk	Total
Toy poodle ^a	137	35	3	175
	78.29%	20.00%	1.71%	
Labrador retriever ^a	111	21	1	133
	83.46%	15.79%	0.75%	
Australian shepherd dog ^a	116	4	0	120
	96.67%	3.33%	0%	
English cocker spaniel	56	20	1	77
	72.73%	25.97%	1.30%	
Australian cattle dog	37	20	2	59
	62.71%	33.90%	3.39%	
Standard poodle	46	6	0	52
	88.46%	11.54%	0%	
Bolognese dog	15	1	0	16
	93.75%	6.25%	0%	
Miniature poodle	12	4	0	16
	75.00%	25.00%	0%	
Nova Scotia duck tolling retriever	9	7	0	16
	56.25%	43.75%	0%	
Other breeds ^b	18	10	1	29
	62.07%	34.48%	3.45%	
Total	557	128	8	693
	80.36%	18.48%	1.16%	

Note: Only breeds that counted more than 15 subjects are reported. In bold, the breeds that presented with a percentage of heterozygous carriers higher than 15%.

^aDog breeds that are among the five most popular breeds reported in Italy.

^b Breeds that counted fewer than 15 subjects include Portuguese water dog (n = 11), Entlebucher mountain dog (n = 8), Karelian bear dog (n = 6), Kai (n = 3) and Lapponian herder (n = 1).

forms of PRA could be the reason why PRCD is the most widespread PRA form in our population, along with GR-PRA1, GR-PRA2 and CORD1. Progressive rod-cone degeneration is the most widespread form of PRA mainly because it affects several different breeds,^{14,15} while another explanation of the diffusion of GR-PRA1 and GR-PRA2 lies in the large numbers of golden retrievers present and tested in Italy.

In recent years, there have been few published studies on the genotypic and allelic frequencies of canine PRA in other countries.^{34–37} One genetic study,³⁸ with sample sizes consisting of millions of dogs from 150 countries, reported results on 250 genetic disease-associated variants in the general dog population. It included PRA-related statistics that varied between purebred and mixed breed dogs. The allele frequency for the PRCD allele in 16,825 Labrador retrievers was 7.2%, which is in line with the reported 15.79% carrier frequency in our samples from Labrador retrievers. The allelic frequency for PRCD in Australian shepherd dogs was 0.38% (tested for 2293 dogs), while our study identified a lower incidence of that risk allele (0.02%) although the population size in our study was only 120 dogs (Table S2). The allelic frequency in 94 toy poodles was estimated to be 2.1%,38 compared with 11.7% in our sample of 175 dogs from the Italian population. In other breeds, when the number of animals tested was higher, as in the case of the standard poodle, the frequency of the risk allele was more comparable. The PRCD was 0.5% in 4197 standard poodles,³⁸ compared with 5.8% in 52 dogs from the Italian population. The frequency of the PRCD allele in the English cocker spaniel was estimated to be 9.5% based on 579 dogs,³⁸ compared with our result of 14.3% for 77 dogs in the Italian cohort.

In conclusion, our study offers an overview of the genotypic and allelic frequencies of various forms of PRA for a cohort of dogs in Italy. The results suggest that the PRA genetic test may be considered a useful tool by veterinarians and breeders to identify affected/at risk and heterozygous carriers among the canine population. Performing the DNA test in young subjects before the dogs are used in breeding programmes is desirable in order to improve dogs' health and welfare through the implementation of specific breeding schemes.

AUTHOR CONTRIBUTIONS

Conceptualisation: Michele Polli, Sara Ghilardi and Paola G. Brambilla. *Data curation*: Sara Ghilardi. *Formal analysis*: Giulia E. Barbariga. *Funding acquisition*: Michele Polli. *Sampling*: Michele Polli and Stefano Frattini. *Methodology*: Michele Polli and Giulietta Minozzi. *Writing original draft*: Michele Polli, Sara Ghilardi and Mara Bagardi. *Writing, review and editing*: Paola G. Brambilla, Sara Ghilardi, Giulietta Minozzi and Michele Polli. All authors read and agreed to the published version of the manuscript.

ACKNOWLEDGEMENTS

The authors are grateful to the Vetogene Laboratory for access to the test data and the information that led to the compilation of Table 1. This work was supported by the APC funds of the University of Milan (for the publication fee).

CONFLICTS OF INTEREST STATEMENT

Stefano Frattini is an employee of the Vetogene Laboratory. The laboratory had no role in the design of the study, analysis and interpretation of the data, or the preparation of the article. The other authors declare they have no conflicts of interest.

ETHICS STATEMENT

The authors confirm that the ethical policies of the journal, as noted on the journal's author guidelines page, have been adhered to. No ethical approval was required because the study analysed test data provided by the laboratory, using samples derived from general clinical practice, subject to permission to release the test results from the dog owners.

DATA AVAILABILITY STATEMENT

Data analysed and described in this study are available from the corresponding authors upon reasonable request.

ORCID

Sara Ghilardi b https://orcid.org/0000-0001-7738-3204 Mara Bagardi b https://orcid.org/0000-0002-5247-3796 Paola G. Brambilla b https://orcid.org/0000-0003-4255-555X

Giulietta Minozzi b https://orcid.org/0000-0002-0020-8798 Michele Polli b https://orcid.org/0000-0003-0129-0161

REFERENCES

- Miyadera K, Acland GM, Aguirre GD. Genetic and phenotypic variations of inherited retinal diseases in dogs: the power of within- and across-breed studies. Mamm Genome. 2012;23:40–61. https://doi.org/10. 1007/s00335-011-9361-3
- Bunel M, Chaudieu G, Hamel C, Lagoutte L, Manes G, Botherel N, et al. Natural models for retinitis pigmentosa: progressive retinal atrophy in dog breeds. Hum Genet. 2019;138:441–53. https://doi.org/10.1007/ s00439-019-01999-6
- Urkasemsin G, Pongpanich M, Sariya L, Kongcharoen A, Buddhirongawatr R, Rungarunlert S, et al. Whole genome sequencing identifies a homozygous nonsense mutation in the JPH2 gene in Shih Tzu dogs with progressive retinal atrophy. Anim Genet. 2021;52:714–19. https://doi.org/10.1111/age.13118
- Petersen-Jones SM. Animal models of human retinal dystrophies. Eye. 1998;12:566–70. https://doi.org/10.1038/eye.1998.146
- Petersen-Jones SM. Advances in the molecular understanding of canine retinal diseases. J Small Anim Pract. 2005;46:371–80. https://doi.org/10. 1111/j.1748-5827.2005.tb00333.x
- Pasmanter N, Petersen-Jones SM. A review of electroretinography waveforms and models and their application in the dog. Vet Ophthalmol. 2020;23:418–35. https://doi.org/10.1111/vop.12759
- Trecenti-Santana AS, Gumiero Guiraldelli G, Garrido Albertino L, Franco Ferreira J, Michelsen Andrade F, Secorun Borges A, et al. Allele frequency of SLC 4 A 3 (PRA1), TTC 8 (PRA2), and PRA–PRCD mutations in golden retrievers in Brazil. Front Vet Sci. 2022;9:973854. https:// doi.org/10.3389/fvets.2022.973854
- Downs LM, Hitti R, Pregnolato S, Mellersh CS. Genetic screening for PRA-associated mutations in multiple dog breeds shows that PRA is heterogeneous within and between breeds. Vet Ophthalmol. 2014;17:126–30. https://doi.org/10.1111/vop.12122
- Clements PJ, Gregory CY, Peterson-Jones SM, Sargan DR, Bhattacharya SS. Confirmation of the rod cGMP phosphodiesterase beta subunit (PDE beta) nonsense mutation in affected RCD-1 Irish setters in the UK and development of a diagnostic test. Curr Eye Res. 1993;12:861–66. https:// doi.org/10.3109/02713689309020391
- Dekomien G, Runte M, Godde R, Epplen JT. Generalized progressive retinal atrophy of Sloughi dogs is due to an 8-bp insertion in exon 21 of the PDE6B gene. Cytogenet Cell Genet. 2000;90:261–67. https://doi.org/ 10.1159/000056785
- Kukekova AV, Goldstein O, Johnson JL, Richardson MA, Pearce-Kelling SE, Swaroop A, et al. Canine RD3 mutation establishes rod-cone dysplasia type 2 (rcd2) as ortholog of human and murine rd3. Mamm Genome. 2009;20:109–23. https://doi.org/10.1007/s00335-008-9163-4

- Petersen-Jones SM, Entz DD, Sargan DR. cGMP phosphodiesterasealpha mutation causes progressive retinal atrophy in the Cardigan Welsh Corgi dog. Invest Ophthalmol Vis Sci. 1999;40:1637–44. PMID: 10393029
- Kijas JW, Miller BJ, Pearce-Kelling SE, Aguirre GD, Acland GM. Canine models of ocular disease: outcross breedings define a dominant disorder present in the English mastiff and bull mastiff dog breeds. J Hered. 2003;94:27–30. https://doi.org/10.1093/jhered/esg007
- Zangerl B, Goldstein O, Philip AR, Lindauer SJP, Pearce-Kelling SE, Mullins RF, et al. Identical mutation in a novel retinal gene causes progressive rod-cone degeneration in dogs and retinitis pigmentosa in humans. Genomics. 2006;88:551–63. https://doi.org/10.1016/j.ygeno. 2006.07.007
- Dostal J, Hrdlicova A, Horak P. Progressive rod-cone degeneration (PRCD) in selected dog breeds and variability in its phenotypic expression. Vet Med. 2011;56:243–47. https://doi.org/10.17221/1564-VETMED
- Acland GM, Aguirre GD. Retinal degenerations in the dog: IV. Early retinal degeneration (ERD) in Norwegian elkhounds. Exp Eye Res. 1987;44:491–521. https://doi.org/10.1016/s0014-4835(87)80160-4
- Zhang Q, Acland GM, Wu WX, Johnson JL, Pearce-Kelling S, Tulloch B, et al. Different RPGR exon ORF15 mutations in Canids provide insights into photoreceptor cell degeneration. Hum Mol Genet. 2002;11:993–1003. https://doi.org/10.1093/hmg/11.9.993
- Palanova A. The genetics of inherited retinal disorders in dogs: implications for diagnosis and management. Vet Med. 2016;7:41–51. https://doi. org/10.2147/VMRR.S63537
- Downs LM, Wallin-Håkansson B, Boursnell M, Marklund S, Hedhammar A, Truvé K, et al. A frameshift mutation in golden retriever dogs with progressive retinal atrophy endorses SLC4A3 as a candidate gene for human retinal degenerations. PLoS One. 2011;6:e21452. https://doi.org/10.1371/journal.pone.0021452
- Downs LM, Wallin-Håkansson B, Bergström T, Mellersh CS. A novel mutation in TTC8 is associated with progressive retinal atrophy in the golden retriever. Canine Genet Epidemiol. 2014;1:4. https://doi.org/10. 1186/2052-6687-1-4
- Suber ML, Pittler SJ, Qin N, Wright GC, Holcombe V, Lee RH, et al. Irish setter dogs affected with rod/cone dysplasia contain a nonsense mutation in the rod cGMP phosphodiesterase beta-subunit gene. Proc Natl Acad Sci U S A. 1993;90:3968–72. https://doi.org/10.1073/pnas.90.9.3968
- Goldstein O, Mezey JG, Schweitzer PA, Boyko AR, Gao C, Bustamante CD, et al. IQCB1 and PDE6B mutations cause similar early onset retinal degenerations in two closely related terrier dog breeds. Invest Ophthalmol Vis Sci. 2013;54:7005–19. https://doi.org/10.1167/iovs.13-12915
- Downs LM, Bell JS, Freeman J, Hartley C, Hayward LJ, Mellersh CS. Late-onset progressive retinal atrophy in the Gordon and Irish Setter breeds is associated with a frameshift mutation in C2orf71. Anim Genet. 2013;44:169–77. https://doi.org/10.1111/j.1365-2052.2012.02379.x
- Goldstein O, Jordan JA, Aguirre GD, Acland GM. A non-stop Santigen gene mutation is associated with late onset hereditary retinal degeneration in dogs. Mol Vis. 2013;19:1871–84
- Wiik AC, Ropstad EO, Ekesten B, Karlstam L, Wade CM, Lingaas F. Progressive retinal atrophy in Shetland sheepdog is associated with a mutation in the CNGA1 gene. Anim Genet. 2015;46:515–21. https://doi. org/10.1111/age.12323
- Ahonen SJ, Arumilli M, Lohi H. A CNGB1 frameshift mutation in Papillon and Phalène dogs with progressive retinal atrophy. PLoS One. 2013;8:e72122. https://doi.org/10.1371/journal.pone.0072122
- Mellersh CS, Boursnell MEG, Pettitt L, Ryder EJ, Holmes NG, Grafham D, et al. Canine RPGRIP1 mutation establishes cone-rod dystrophy in miniature longhaired dachshunds as a homologue of human Leber congenital amaurosis. Genomics. 2006;88:293–301. https://doi.org/10.1016/j. ygeno.2006.05.004

- Wiik AC, Wade C, Biagi T, Ropstad EO, Bjerkas E, Lindblad-Toh K, et al. A deletion in nephronophthisis 4 (NPHP4) is associated with recessive cone-rod dystrophy in standard wire-haired dachshund. Genome Res. 2008;18:1415–21. https://doi.org/10.1101/gr.074302.107
- Dekomien G, Vollrath C, Petrasch-Parwez E, Boevé MH, Akkad DA, Gerding WM, et al. Progressive retinal atrophy in Schapendoes dogs: mutation of the newly identified CCDC66 gene. Neurogenetics. 2010;11:163–74. https://doi.org/10.1007/s10048-009-0223-z
- Murgiano L, Becker D, Spector C, Carlin K, Santana E, Niggel JK, et al. CCDC66 frameshift variant associated with a new form of earlyonset progressive retinal atrophy in Portuguese water dogs. Sci Rep. 2020;10:21162. https://doi.org/10.1038/s41598-020-77980-5
- Goldstein O, Guyon R, Kukekova A, Kuznetsova TN, Pearce-Kelling SE, Johnson J, et al. COL9A2 and COL9A3 mutations in canine autosomal recessive oculoskeletal dysplasia. Mamm Genome. 2010;21:398–408. https://doi.org/10.1007/s00335-010-9276-4
- 32. Zhang Q, Acland GM, Parshall CJ, Haskell J, Ray K, Aguirre GD. Characterization of canine photoreceptor phosducin cDNA and identification of a sequence variant in dogs with photoreceptor dysplasia. Gene. 1998;215:231–39. https://doi.org/10.1016/s0378-1119(98)00310-2
- Murgiano L, Becker D, Torjman D, Niggel JK, Milano A, Cullen C, et al. Complex structural PPT1 variant associated with non-syndromic canine retinal degeneration. G3 Genes Genomes Genetics. 2019;9:425–37. https://doi.org/10.1534/g3.118.200859
- Takanosu M. Different allelic frequency of progressive rod-cone degeneration in two populations of Labrador retrievers in Japan. J Vet Med Sci. 2017;79:1746–48. https://doi.org/10.1292/jvms.17-0243
- 35. Kohyama M, Tada N, Mitsui H, Tomioka H, Tsutsui T, Yabuki A, et al. Real-time PCR genotyping assay for canine progressive rod-cone degeneration and mutant allele frequency in toy poodles, chihuahuas and miniature dachshunds in Japan. J Vet Med Sci. 2016;78:481–84. https:// doi.org/10.1292/jvms.15-0279
- 36. Andrade LR, Caceres AM, Trecenti AS, Brandao CVS, Gandolfi MG, Aguiar EV, et al. Allele frequency of the C.5G>A mutation in the PRCD gene responsible for progressive retinal atrophy in English cocker spaniel dogs. Animals. 2019;9:844. https://doi.org/10.3390/ani9100844
- 37. Freitas HM, Somma AT, Moore BA, Montiani-Ferreira F. Retrospective and prospective study of progressive retinal atrophy in dogs presented to the veterinary hospital of the Federal University of Parana, Brazil. Open Vet J. 2021;11:370–78. https://doi.org/10.5455/OVJ.2021.v11.i3.6
- Donner J, Freyer J, Davison S, Anderson H, Blades M, Honkanen L, et al. Genetic prevalence and clinical relevance of canine Mendelian disease variants in over one million dogs. PLoS Genet. 2023;19:e1010651. https:// doi.org/10.1371/journal.pgen.1010651

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Ghilardi S, Bagardi M, Frattini S, Barbariga GE, Brambilla PG, Minozzi G, et al. Genotypic and allelic frequencies of progressive rod-cone degeneration and other main variants associated with progressive retinal atrophy in Italian dogs. Vet Rec Open. 2023;10:e77. https://doi.org/10.1002/vro2.77