

## **Hematocrit: The Neglected Variable of Extracorporeal CO<sub>2</sub> Removal**

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**Authors' contribution:** TL, AZ conceived the study; LV, MB, GF, ST, SMC, AZ, TL conducted experiments; LV, EC, FZ, AZ, TL analysed and interpreted the data; LV, EC, FZ,

AZ, TL prepared the first draft; GF, ST, SMC revised the article for important intellectual content. All authors revised and approved the final version to be published.

**Funding:** The study was funded by Italian Ministry of Health – Current research

**Running title:** Hematocrit and ECCO<sub>2</sub>R

**Total word count:** 1000 words

**Tables:** 1

**Figures:** 1

**References:** 10

**Keywords:** carbon dioxide; extracorporeal CO<sub>2</sub> removal; acid-base equilibrium; gas exchange; respiratory failure

## To the Editor:

### Introduction

Carbon dioxide (CO<sub>2</sub>), mainly carried in blood as bicarbonate, can be removed through the use of extracorporeal respiratory devices (1). The effects of several variables (*e.g.* gas flow and blood flow) on extracorporeal CO<sub>2</sub> removal (ECCO<sub>2</sub>R) have been extensively explored. Less attention has been dedicated to the potential influence of hematocrit.

### Methods

An *in-vitro* circuit primed with 3-L of swine blood was assembled with two membrane lungs (ML). The first ML, ventilated with CO<sub>2</sub>, guaranteed a *constant* pCO<sub>2</sub> of 50 (tolerance of  $\pm 2$  mmHg) at the inlet of the second ML, which removed CO<sub>2</sub> using a *fixed* oxygen flow of 5 L/min (**Figure 1A**). Blood temperature and flow through the second ML were constant (37°C and 250 ml/min). Blood was continuously dialyzed (MultiBic® 4mmol/L potassium, Fresenius) to keep lactate <2.5 mmol/L and to reach a baseline hematocrit of 35% (tolerance  $\pm 2\%$ ).

In addition, a filter was inserted before the second ML to *modulate* hematocrit by ultrafiltration. To *increase* hematocrit (by 10 and 20%) the ultrafiltrate was reinfused after the ML (Figure 1B); to *reduce* hematocrit (by 10 and 20%) the main blood flow was divided in a main branch, directed to the ML, and a secondary branch running through the filter. The obtained ultrafiltrate entered the main blood branch before the ML diluting blood. Blood of five pigs was tested twice (10 experiments). At each step (baseline, +10%, +20%, -10%, -20% of hematocrit), samples collected before (PRE) and after (POST) the second ML were analyzed.

The amount of CO<sub>2</sub> removed ( $V_M\text{CO}_2$ ) was measured, bicarbonate concentration ( $[\text{HCO}_3^-]$ ), PRE and POST ML blood CO<sub>2</sub> content and strong ion difference (SID) were calculated (2, 3).

### *Statistical analysis*

Data are presented as mean  $\pm$  SD. Effects of Hct variation, PRE and POST ML effect and their interaction on variables were evaluated by generalized linear mixed models (fixed and random effects). Pairwise comparisons were tested with Bonferroni's correction. A polynomial maximum likelihood multilevel linear regression model was applied to evaluate the relationship between  $V_M\text{CO}_2$  and hematocrit (2). A p-value $<0.05$  was considered statistically significant (SAS software v.9.4, SAS Institute, NC, USA and SigmaPlot, Systat Software, San Jose, CA).

### **Results**

During the experiments, hematocrit ranged from  $17\pm 2\%$  to  $54\pm 2\%$  (**Table**).  $V_M\text{CO}_2$  increased with hemoconcentration and was reduced with hemodilution (**Figure 1, Right**).

While PRE pCO<sub>2</sub> remained stable, POST pCO<sub>2</sub> was higher at steps with higher hematocrit (P $<0.0001$ ). Similarly, PRE  $[\text{HCO}_3^-]$  remained relatively constant, while a progressive reduction according to hematocrit was observed in POST samples, justifying the increased CO<sub>2</sub> extraction. Given that red blood cells (RBCs) have lower CO<sub>2</sub> content than plasma (3), PRE CO<sub>2</sub> content was lower during hemoconcentration. Despite that, output CO<sub>2</sub> content was markedly reduced, resulting in enhanced CO<sub>2</sub> removal.

The changes in  $[\text{HCO}_3^-]$  were closely mirrored by reductions in PRE-to-POST SID, attributed to increased chloride and decreased sodium concentrations in POST samples.

PRE pH was relatively constant, while POST pH changed markedly with hematocrit, with very alkaline values observed during hemodilution, indicating a poor non-carbonic buffer power of samples with low hemoglobin concentration.

## Discussion

Five variables regulating ECCO<sub>2</sub>R are classically mentioned: input pCO<sub>2</sub>, gas flow, blood flow, blood temperature and ML size. Less attention has been dedicated to hematocrit (4). A recent *in-vitro* study evaluated the role of hematocrit on ECCO<sub>2</sub>R (5). However, a relatively narrow range of hematocrit was explored and the mechanisms of enhanced CO<sub>2</sub> removal were not investigated.

Let us therefore analyze the physiological mechanisms likely driving the observed increased ECCO<sub>2</sub>R. Hypothesizing to use, in a setting akin to ours, a solution of water and electrolytes (Na<sup>+</sup>=140 mEq/L, Cl<sup>-</sup>=100 mEq/L, HCO<sub>3</sub><sup>-</sup>=40 mEq/L), the only factor influencing ECCO<sub>2</sub>R would be dissolved CO<sub>2</sub>, *i.e.* input pCO<sub>2</sub>. In this context, bicarbonate, *i.e.* dissociated CO<sub>2</sub>, can be considered as an “inert” form of CO<sub>2</sub> storage. Indeed, due to the absence of buffers, the reduction in [HCO<sub>3</sub><sup>-</sup>] induced by the pCO<sub>2</sub> decrease is negligible and, consequently, V<sub>M</sub>CO<sub>2</sub> is limited. If albumin is added, V<sub>M</sub>CO<sub>2</sub> will depend on the variation in pCO<sub>2</sub> (*as above*) and on the change in [HCO<sub>3</sub><sup>-</sup>]. In this context, the amount of undissociated weak-non carbonic acids (AH), able to dissociate (A<sup>-</sup>), is the driving factor allowing the [HCO<sub>3</sub><sup>-</sup>] reduction. If we add RBCs to the solution, these will allow greater electrolyte shifts and thus [HCO<sub>3</sub><sup>-</sup>] variations (2), effectively increasing V<sub>M</sub>CO<sub>2</sub>. In fact, RBCs act as internal acidifying elements, allowing the conversion of bicarbonate to pCO<sub>2</sub>, driven by SID decreases (6, 7). The same concept of “bicarbonate displacement” was exploited in several experimental settings using exogenous acids (8, 9). This, however, lead to unwanted effects, such as

persistent acid load or increased CO<sub>2</sub> production. In case of RBCs, the acid load results from charge redistribution, rather than its addition, and is therefore reversible.

The addition of RBCs (or their increased concentration) has another important consequence. As the CO<sub>2</sub> content of RBCs is lower than that of plasma (3), the net effect is a reduction of the amount of CO<sub>2</sub> reaching the ML. Interestingly, despite the reduced CO<sub>2</sub> content, the “exchangeable” CO<sub>2</sub> increases. We observed an increase in POST pCO<sub>2</sub> at the steps with higher hematocrit, suggesting that gas flow was insufficient for the achieved, higher POST pCO<sub>2</sub>, driven by the higher conversion of bicarbonate. A simple implication is that an increase in gas flow (kept intentionally constant) would have further augmented ECCO<sub>2</sub>R.

Should we therefore increase hematocrit in patients on ECCO<sub>2</sub>R? A higher hematocrit optimizes both the delivery of oxygen and CO<sub>2</sub> removal. However, the increased viscosity of blood and the risk of transfusions may well outweigh any benefit and caution is required to extrapolate these results to the clinical arena (10).

The major limitation of our study is the variable concentration of plasma proteins. During hemoconcentration, we likely obtained also an increased albumin concentration. The observed slightly lower PRE pH and [HCO<sub>3</sub><sup>-</sup>] can be attributed to albumin’s acidifying effect.

## **Conclusions**

Hematocrit is a key variable in determining ECCO<sub>2</sub>R efficiency. RBCs enhance CO<sub>2</sub> removal, increasing exchangeable CO<sub>2</sub> and facilitating bicarbonate conversion to dissolved CO<sub>2</sub>. Electrolyte shifts to and from RBCs guarantee electrical neutrality.

**Conflict of interest statement:** Prof. Grasselli reports personal fees from Maquet, Draeger, Pfizer, Thermo Fisher, MSD and Gilead outside the submitted work. Prof. Alberto Zanella is inventor of patents related to the topic. The remaining authors have disclosed that they do not have any potential conflict of interest.

**Acknowledgments:** We thank Marina Leonardelli (Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico) and Patrizia Minunno (Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico) for their valuable support.

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**TABLE 1: physiological variables**

		TARGET Hct variation					
		-20	-10	0 (Baseline)	+10	+20	<i>P-value</i>
<b>Hct (%)</b>	<b>PRE</b>	17.0±2.3	26.4±3.1	35.4±1.9	44.7±1.8	53.9±2.4	<.000 1
		†‡§#	‡§#	§#	#		
<b>V<sub>M</sub>CO<sub>2</sub> (mL/min)</b>		31.0±4.8	37.3±5.7	42.5±6.3	44.9±6.7	48.7±7.6	<.000 1
		†‡§#	‡§#	#	#		
<b>pH</b>	<b>PRE</b>	7.428±0.03	7.428±0.04	7.392±0.04	7.408±0.02	7.407±0.01	0.006 4
		7	0	1	5	7	
	<b>POST</b>	8.012±0.04	7.924±0.05	7.810±0.06	7.761±0.05	7.721±0.03	<.000 1
		3	1	6	7	7	
<b>pCO<sub>2</sub> (mmHg)</b>	<b>PRE</b>	50.5±1.4	49.5±1.8	50.3±1.0	49.6±0.7	50.2±1.0	0.118 8
		*	*	*	*	*	
	<b>POST</b>	13.8±1.0	14.6±1.2	15.7±1.6	16.9±2.5	17.5±1.8	<.000 1
		‡§#	§#	#			
<b>Plasma [HCO<sub>3</sub><sup>-</sup>] (mmol/L)</b>	<b>PRE</b>	33.4±2.4	32.8±2.2	30.7±3	31.3±1.6	31.6±1.3	0.001 5
		*‡§	*‡	*	*	*	
	<b>POST</b>	34.8±2.0	30.2±1.8	25.0±2.6	23.9±1.1	22.6±0.6	<.000 1
		†‡§#	‡§#	#			
<b>Plasma [TCO<sub>2</sub>] (mmol/L)</b>	<b>PRE</b>	35.0±2.3	34.3±2.2	32.3±3.0	32.9±1.6	33.1±1.3	0.001 5
		‡§	*‡	*	*	*	
	<b>POST</b>	35.3±2.0	30.6±1.8	25.5±2.6	24.4±1.2	23.1±0.6	<.000 1
		†‡§#	‡§#	#			
<b>Blood [TCO<sub>2</sub>] (mmol/L)</b>	<b>PRE</b>	32.7±1.9	30.7±1.6	27.8±2.3	27.1±1.1	26.1±0.9	<.000 1
		†‡§#	*‡§#	*	*	*	
	<b>POST</b>	32.8±1.8	27.3±1.5	21.7±2.1	19.8±0.9	17.8±0.6	<.000 1
		†‡§#	‡§#	§#	#		
<b>[Cl<sup>-</sup>] (mEq/L)</b>	<b>PRE</b>	109.1±2.5	109.4±2.8	109.1±3.2	109.3±2.8	109.1±3.0	0.815 1
		*	*	*	*	*	
	<b>POST</b>	110.1±2.1	111.1±3.0	111.3±3.2	112±3.1	112.4±3.4	<.000 1
		†‡§#	#	#			
<b>[Na<sup>+</sup>] (mEq/L)</b>	<b>PRE</b>	141.8±1.8	141.7±1.8	141.2±2.0	141.5±1.8	142.5±1.4	0.136 5
		*	*	*	*	*	
	<b>POST</b>	140.6±2.0	139.8±2.0	138.8±1.8	139.2±1.9	139.7±1.3	0.012 5
		‡					
<b>SID (mEq/L)</b>	<b>PRE</b>	38.2±1.9	37.8±2.3	37±2.5	37.3±1.8	38.5±2.4	0.132 9
		*	*	*	*	*	
	<b>POST</b>	35.8±1.6	34±2.3	32.2±2.3	32.1±1.9	32.3±2.8	<.000 1
		‡§#	§				

Mean  $\pm$  SD of the physiological variables measured before (PRE) and after (POST) passing through the membrane lung at different target hematocrit variations. **PRE and POST ML effect** and **interaction** P-values (when applicable) were always  $<.0001$ ; **target Hct variation** P-values were  $<.0001$  except for  $[Cl^-]$  ( $P=0.0023$ ),  $[Na^+]$  ( $P=0.0838$ ) and SID ( $P=0.0013$ ). P-values reported in Table represent: **target Hct variation effect** for “Hct” and “ $V_MCO_2$ ”, while, for all other variables **target Hct variation effect sliced for PRE-POST** values. Post-hoc analysis: \*  $p<0.05$  PRE vs POST; †  $p<0.05$  vs Hct -10; ‡  $p<0.05$  vs Hct 0; §  $p<0.05$  vs Hct +10; #  $p<0.05$  vs Hct +20. Hct = hematocrit;  $V_MCO_2$  = The amount of  $CO_2$  removed by the membrane lung;  $pCO_2$  = partial pressure of carbon dioxide;  $[HCO_3^-]$  = bicarbonate concentration;  $[TCO_2]$  =  $CO_2$  content;  $[Cl^-]$  = chloride concentration;  $[Na^+]$  = sodium concentration; SID = strong ion difference.

## Figure Legends

**Figure 1: Left:** Scheme of the closed-loop circuit used for the experiments (A), describing both the system applied to achieve *hemoconcentration* (B) and *hemodilution* (C). The circuit was initially heparinized with 15000 Units of heparin. **Right:** The relationship between hematocrit and the amount of CO<sub>2</sub> removed through the membrane lung (V<sub>M</sub>CO<sub>2</sub>). Different symbols identify the 5 target Hct steps: baseline (black triangles), 10% Hct increase (white triangles), 20% Hct increase (black squares), 10% Hct decrease (white circles), 20% Hct decrease (black circles). Each experiment is identified by a different line. The polynomial maximum likelihood multilevel model results are represented by the blue line with 95% confidence interval (grey area). V<sub>M</sub>CO<sub>2</sub> increased with square absolute Hct values according to the following model:  $V_MCO_2 = 17.2 + 0.91 \times Hct - 0.0062 \times Hct^2$ .

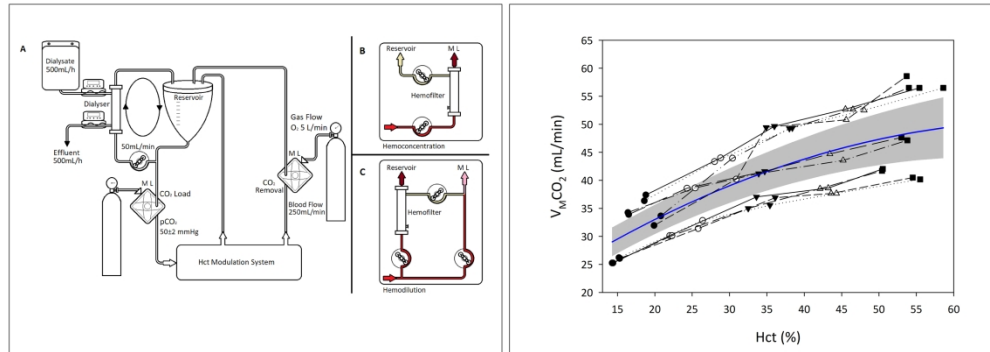


Figure 1: Left: Scheme of the closed-loop circuit used for the experiments (A), describing both the system applied to achieve hemoconcentration (B) and hemodilution (C). The circuit was initially heparinized with 15000 Units of heparin. Right: The relationship between hematocrit and the amount of CO<sub>2</sub> removed through the membrane lung (VMCO<sub>2</sub>). Different symbols identify the 5 target Hct steps: baseline (black triangles), 10% Hct increase (white triangles), 20% Hct increase (black squares), 10% Hct decrease (white circles), 20% Hct decrease (black circles). Each experiment is identified by a different line. The polynomial maximum likelihood multilevel model results are represented by the blue line with 95% confidence interval (grey area). VMCO<sub>2</sub> increased with square absolute Hct values according to the following model:  $VMCO_2 = 17.2 + 0.91 \times Hct - 0.0062 \times Hct^2$ .

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