

e Biotecnologie Molecolari

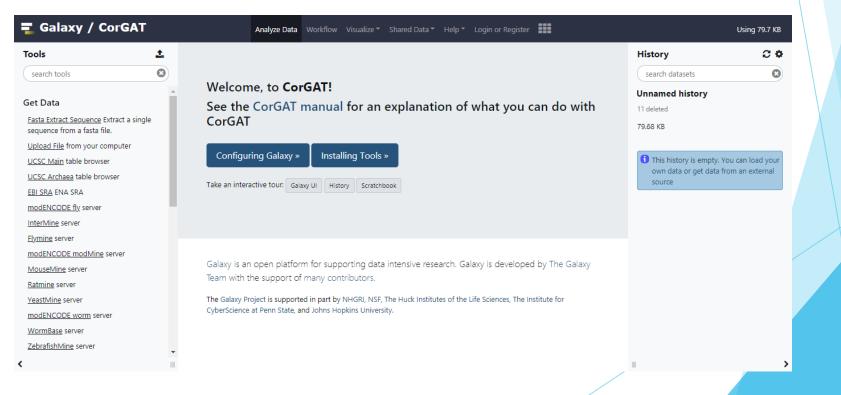
Dipartimento di Bioscienze

# **CorGAT** and **CorGAT**-tracker

## Functional annotation of SARS-CoV-2 genomes and tracking mutations and variants of concern

## CorGAT

- Coronavirus Genome Analysis Tool or CorGAT (DOI:10.1093/bioinformatics/btaa1047).
- Collection of Perl utilities and annotation files.
- Performs the functional annotation of SARS-CoV-2 genetic variants.



- Brief explanation of functioning:
  - 1. Alignment of complete assemblies of SARS-CoV-2 genomes to the reference sequence.

\Xi Galaxy / CorGAT

nucmer snp Align single fasta files to SARS-CoV-2 genome and call genetic

join nucmer Join multiple mumme output files in a phenetic matrix FunAnn Performs functional annotation of genetic variants multiFC Process multi-fasta files to derive a phenetic matrix of genetic

Tools/utilities for Haplogroup

Tools search tools Graph/Display Data Phenotype Association genome\_alignment **Coronavirus Genome Annotation** 

Tool

variants

variants.

assignment

Workflows All workflows

- 2. Obtain a list of polymorphic regions.
- 3. Functional annotation of the identified variants.

	ualize ▼ Shared Data ▼ Help ▼ Login or R						Using 150.3 KB				
multiFC Process multi-fasta files to derive a phe	enetic matrix of genetic variants. (Galaxy Version	1)	•	Options		earch datasets	2 <b>4</b>				
multifasta						named history					
🗅 🙆 🗅 15: Test.fa				•	~	own, 14 deleted					
Multifasta file of SARS-CoV-2 genomes					150	.26 KB					
✓ Execute						GCA_009858895.3_A9 89v3_genomic.fna	M9 🕑 🖋 🗙				
What it does?					15:	Test.fa	• / ×				
This tool is used to align SARS-CoV-2 genes, in mul using nurmer. The output will consist in a single tat as many rows as the number of variants observed in present, 0=absent will be used to indicate whether FunAnn tool to obtain the functional annotation of	oular file with as may columns as the number of n the genomes. For every genome assembly and that genome carries a specific variant. This table	genom variant	es provideo a simple b	d in input. inary code	And						
📮 Galaxy / CorGAT	Analyze Data Workflow	v Visu	alize 🍷 S	hared Data	r∓ Help∓	Login or Register		ι.	lsing 151.7 KB		
Tools 📩								History	<b>C O</b>		
search tools	FunAnn Performs functional annotation	of gene	tic variants	(Galaxy Ve	rsion 1)			search datasets	8		
Graph/Display Data	merged show-snps tabular ouput							Unnamed history			
Phenotype Association								4 shown, 14 deleted			
genome_alignment	18: Test_GenVar.tsv						• 🖻				
Coronavirus Genome Annotation	annotates SARS-CoV-2 variants							151.71 KB			
Tool	✓ Execute							18: Test_GenVar.tsv	⊛ / ×		
nucmer snp Align single fasta files to								17: multiFC on data 15: log f	@ # X		
SARS-CoV-2 genome and call genetic variants	What it does?							ile	. , ,		
j <u>oin nucmer</u> Join multiple mummer output files in a phenetic matrix	This program reads a tabular formatted file, in pseudo vcf format, as obtained fron annotation of SARS-CoV-2 variants. Please notice that the program performs mini					error checks, and that it	is designed to work	16: GCA_009858895.3_ASM9 85889v3_genomic.fna	• / ×		
<u>FunAnn</u> Performs functional annotation of genetic variants	exclusively with the reference annotation of the SARS-CoV-2 genome as available format can be found also in this Galaxy, under Shared Data -> Data Libraries -> SA						genome in fasta	15: Test.fa	⊛ # ×		
<u>multiFC</u> Process multi-fasta files to derive a phenetic matrix of genetic variants.	The output file is again, a tabular file delinea the input file. A more detailed description of .										
Tools/utilities for Haplogroup assignment	🚍 Galaxy / CorGAT				Analyze Dat	a Workflow Visuali	ze 🍨 Shared Data 👻	Help * Login or Register			Using 159.
Workflows	Tools	<u>*</u>	POS	REF	ALT	annot				History	0
All workflows	search tools	3	POS	REF	ALT	annot				search datasets	
*	Graph/Display Data		241 733	C T	T C	5'UTR:nc.C241T,NA,		:c.468T>C,p.D156D,synonymous;		Unnamed history	
`	Phenotype Association		1926	c	т			:c.4681>C,p.D156D,synonymous; 121C>T,p.T374I,missense;		6 shown, 14 deleted	
genome_alignment			2035	G	T			230G>T,p.L410F,missense;		-	
	Coronavirus Genome Annotatio	n	2749	С	т			3:c.30C>T,p.D10D,synonymous;		159.24 KB	
	Tool		3037		т			c.318C>T,p.F106F,synonymous;		20: FunAnn on data 18: lo	ogf 🕑 🖋
	nucmer snp Align single fasta files to		3798					3:c.1079T>.,p.E374*,frameshiftDel;		ile	
	SARS-CoV-2 genome and call genetic		3828	C	T			.1109C>T,p.S370L,missense;		19: Functional annotatio	no 👁 🌶
	variants		4178 5096	A	Т			.1459A>T,p.K487*,stopGain; sp3:c.2377AA>,p.S794*,frameshift	Del	f SARS-CoV-2 genomes	
	join nucmer Join multiple mummer		5096	AA				sp3:c.2392AA>p.L803*,frameshift		abular format	
	output files in a phenetic matrix		5224	т	c			p3:c.2505T>C,p.T835T,synonymous;		18: Test GenVar.tsv	۲
	<u>FunAnn</u> Performs functional annotation		5367	G	т			2648G>T,p.R883I,missense;			

6319 A

6613 A

8017 G

11291 G

11296 T

11483 G

11653 C

12778 C

11098 TTTACC

orf1ab:c.6054A>G,p.P2018P,synonymous:nsp3:c.3600A>G,p.P1200P,synonymous

031T>G,p.F3677L,missense;nsp6:c.324T>G,p.F108L,missense

orf1ab:c.11218G>.,p.G3746\*,frameshiftDel;nsp6:c.511G>.,p.G177\*,frameshiftDel

orf1ab:c.11388C>A,p.L3796L,synonymous;nsp6:c.681C>A,p.L227L,synonymous;

orf1ab:c.12513C>T,p.Y4171Y,synonymous;nsp9:c.93C>T,p.Y31Y,synonymous;

orf1ab:c.11026G>A,p.G3676S,missense:nsp6:c.319G>A,p.G107S,missens

nymousinsn3rc 5208G x Tin &1766& synonymous

orf1abrc 7752G x T n A2584A svnc

of genetic variants

variants.

assignment

Workflows

All workflow

multiFC Process multi-fasta files to

derive a phenetic matrix of genetic

Tools/utilities for Haplogroup

👁 🥖 🗙

17: multiFC on data 15: log f 🛛 🔿 🗶

16: GCA\_009858895.3\_ASM9 💿 🖋 🗙

85889v3\_genomic.fna

15: Test.fa

...p.FLPF42F.inf

- The output is a simple table containing:

  - 3. Alternative allele.
  - 4. Functional annotation. 8. MFE annotation.
  - 5. Allele frequency

- 1. Genomic position. 6. Epitopes annotation.
- 2. Reference allele. 7. Annotation of sites under selective pressure.

POS	REF	ALT	annot
POS	REF	ALT	annot
241	С	Т	5'UTR:nc.C241T,NA,NA;
733	т	С	nsp1:c.468T>C,p.D156D,synonymous;orf1ab:c.468T>C,p.D156D,synonymous;
1926	С	Т	orf1ab:c.1661C>T,p.T554I,missense;nsp2:c.1121C>T,p.T374I,missense;
2035	G	Т	orf1ab:c.1770G>T,p.L590F,missense;nsp2:c.1230G>T,p.L410F,missense;
2749	С	Т	orf1ab:c.2484C>T,p.D828D,synonymous;nsp3:c.30C>T,p.D10D,synonymous;
3037	С	Т	orf1ab:c.2772C>T,p.F924F,synonymous;nsp3:c.318C>T,p.F106F,synonymous;
3798	т		orf1ab:c.3533T>.,p.E1192*,frameshiftDel;nsp3:c.1079T>.,p.E374*,frameshiftDel;

AF	Epitopes		Hyphy	MFE
AF			Hyphy	MFE
94.382501			NA	NA
0.213728			NA	NA
0.031701			fel:true;meme:false;kind:negative;	NA
0.110145		410:LATNNLVVM,7,HLA-B*35:01;HLA-B*46:01;HLA-C*01:0.	fel:true;meme:true;kind:positive;	NA
0.214636		7:FGDDTVIEV,1,HLA-C*08:01;5	NA	NA
96.462537		98:LASHMYCSF,5,HLA-B*15	fel:true;meme:false;kind:negative;	NA
0			NA	NA

## **CorGAT-tracker**

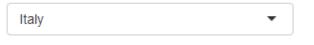
- Shiny based dashboard for the visualization of the prevalence of SARS-CoV-2 lineages and mutations of concern.
- Based on CorGAT derived annotations.
- A Galaxy release is under development.
- Data are represented in an interactive way.
- Users can personalize data visualization through a series of widgets that allow to modify as many parameters, among which:
  - 1. The country of origin of the data.
  - 2. The interval of time to be displayed.
  - 3. The minimum number of sequenced genomes. 5. A lineage of interest to be

represented.

visualized.

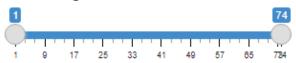
4. A mutation of interest to be

#### Country



Visualize data for the selected country

#### Weeks range



Time lapse of interest (number of weeks from a fixed date)

#### Min number of genomes (Lineages)

1
25
50
100
500
1000

Minimum number of sequenced genomes required to display a Lineage

#### Min number of genomes (Lineages+)

$\bigcirc$	1			
$\bigcirc$	5			
$\bigcirc$	10			
۲	15			
0	25			
0	50			

Minimum number of sequenced genomes required to display a Lineage+

#### Lineage

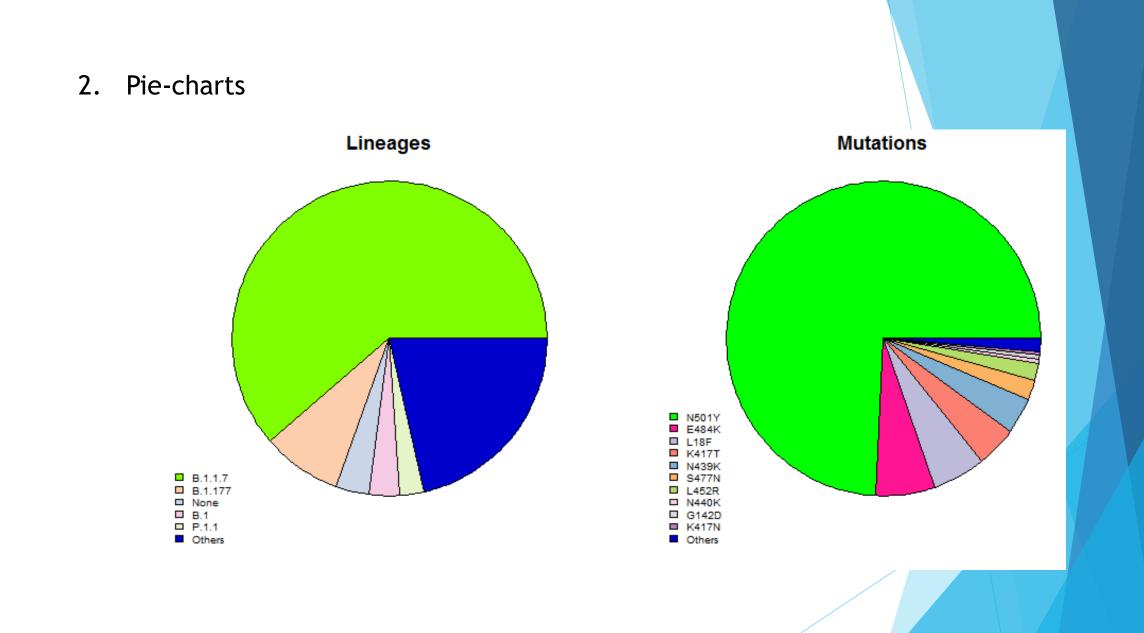
B.1.1.7	•
Produce a scatterplot for the selected Linea	ge

#### Mutation

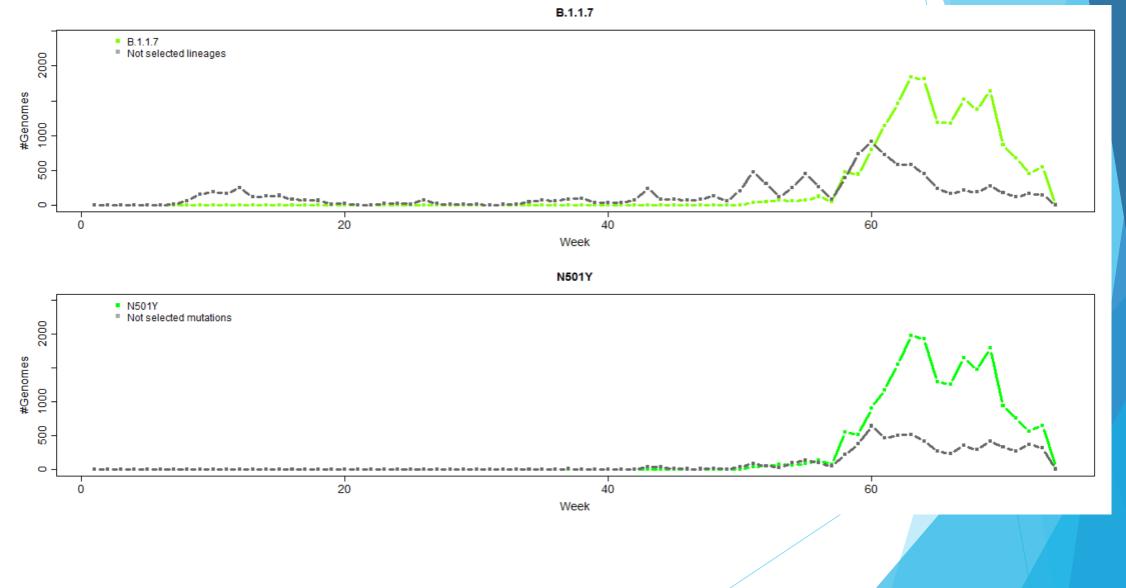
N501Y	-	,
-------	---	---

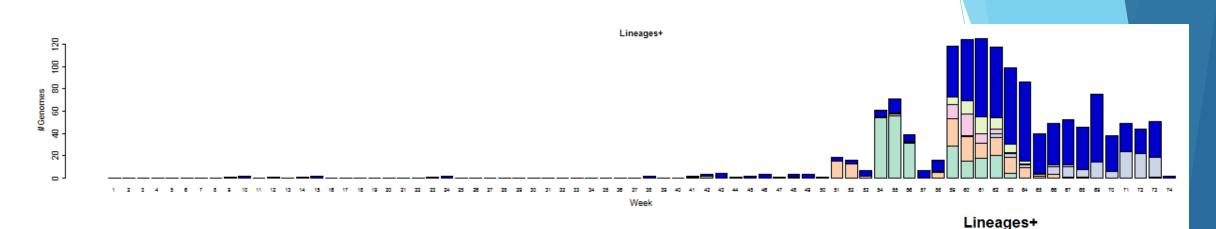
Produce a scatterplot for the selected Mutation

• CorGAT-tracker produces 3 different kinds of plot: 1. Barplots Lineages 2000 #Genomes 1000 1500 200 ۲ م Week Mutations 2000 #Genomes 1000 1500 <mark>8</mark>. 62 70 ....

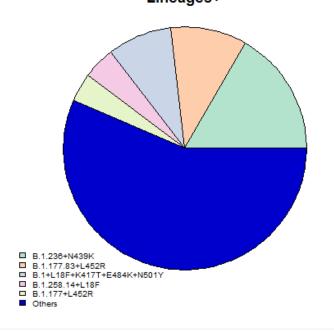


### 3. Scatterplots





- In CorGAT-tracker lineage annotations can be "augmented" by reporting the list of MOC that are observed in a genome, but are not specific to its assigned lineage.
- Augmented annotations are called Lineages+ in-app.
- Lineages+ prevalence in time is represented using a barplot and a pie-chart

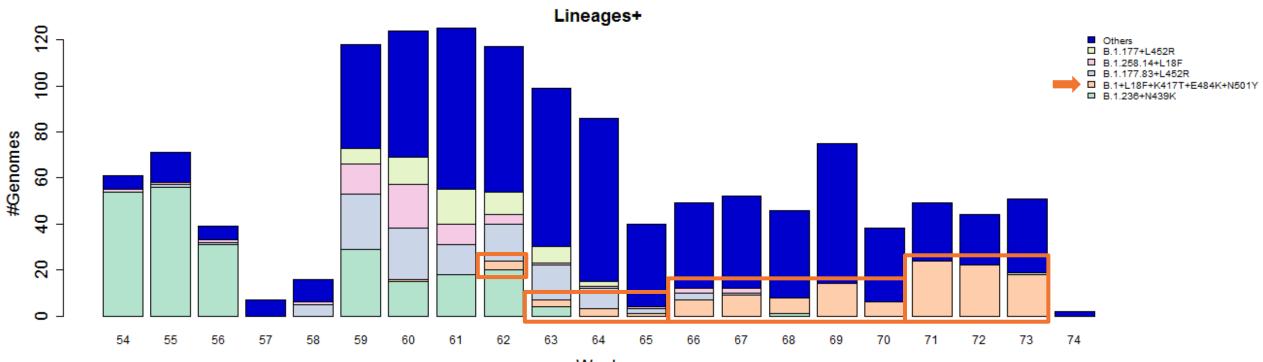


Min number of genomes (Lineages)

## **Final considerations**

- CorGAT and CorGAT-tracker will provide a useful addition to the currently available "arsenal" of bioinformatics methods for the genomic surveillance of SARS-CoV-2.
- CorGAT has a sensitivity comparable to other similar tools, but also provides additional layers of annotation.
- Example: Identification of misclassified SARS-CoV-2 genomes in Italy.

• Genomes classified as B.1 but presented additional mutations on the spike protein.



Week



- In depth studies highlighted that:
  - 1. The majority of the additional spike mutations in the misclassified B.1 were in common with P.1.
  - 2. A "group specific" mutation, P681H, can be identified in the spike protein of the misclassified genomes.
- It is possible to speculate that these genomes represent a newly emerged lineage, however further investigations are required.
- P.1+P681H was recently added to the ECDC list of Variants Under Monitoring

## **Availability**

- CorGAT is already available through Galaxy at the following link:
   http://corgat.cloud.ba.infn.it/galaxy
- A Galaxy release for CorGAT-tracker is under development.
- Further information about the tools can be found in the respective GitHub repositories:
  - https://github.com/matteo14c/CorGAT (CorGAT)
  - https://github.com/F3rika/CorGAT-tracker (CorGAT-tracker)





**UNIVERSITÀ DEGLI STUDI DI MILANO** 

# Thank you!

Special thanks to:

- Matteo Chiara
- Federico Zambelli
- Marco Antonio Tangaro Graziano Pesole •
- Pietro Mandreoli
- David S. Horner