#### Check for updates

#### **OPEN ACCESS**

EDITED BY Pasquale Filannino, University of Bari Aldo Moro, Italy

REVIEWED BY Katharina Anne Scherf, Karlsruhe Institute of Technology (KIT), Germany Francesco Tovoli, University of Bologna, Italy

\*CORRESPONDENCE Luca Elli luca.elli@policlinico.mi.it

SPECIALTY SECTION This article was submitted to Nutritional Immunology, a section of the journal Frontiers in Nutrition

RECEIVED 29 July 2022 ACCEPTED 12 September 2022 PUBLISHED 06 October 2022

#### CITATION

Costantino A, Aversano GM, Lasagni G, Smania V, Doneda L, Vecchi M, Roncoroni L, Pastorello EA and Elli L (2022) Diagnostic management of patients reporting symptoms after wheat ingestion. *Front. Nutr.* 9:1007007. doi: 10.3389/fnut.2022.1007007

#### COPYRIGHT

© 2022 Costantino, Aversano, Lasagni, Smania, Doneda, Vecchi, Roncoroni, Pastorello and Elli. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Diagnostic management of patients reporting symptoms after wheat ingestion

Andrea Costantino<sup>1,2</sup>, Gloria Maria Aversano<sup>3</sup>, Giovanni Lasagni<sup>4</sup>, Veronica Smania<sup>2</sup>, Luisa Doneda<sup>5</sup>, Maurizio Vecchi<sup>1,2</sup>, Leda Roncoroni<sup>5</sup>, Elide Anna Pastorello<sup>6</sup> and Luca Elli<sup>1\*</sup>

<sup>1</sup>Gastroenterology and Endoscopy Unit, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy, <sup>2</sup>Department of Pathophysiology and Transplantation, University of Milan, Milan, Italy, <sup>3</sup>Department of Internal Medicine, ASST Grande Ospedale Metropolitano Niguarda, Milan, Italy, <sup>4</sup>Department of Allergology and Immunology, ASST Grande Ospedale Metropolitano Niguarda, Milan, Italy, <sup>5</sup>Department of Biomedical, Surgical and Dental Sciences, University of Milan, Milan, Italy, <sup>6</sup>Allergology, Casa di Cura Ambrosiana, Cesano Boscone, Italy

Many patients report symptoms after wheat ingestion experiencing a wide spectrum of clinical manifestations. Three possible diagnoses have been recognized: celiac disease (CD), wheat allergy (WA), and non-celiac (gluten) wheat sensitivity (NCGS/NCWS). CD is a chronic immune-mediated disease of the small bowel caused by exposure to dietary gluten in genetically predisposed individuals, with a prevalence of approximately 1%. It is characterized by mucosal inflammation and atrophy following exposure to gluten and improvement after gluten withdrawal. Food allergies are immunological responses to a food antigen. WA is the expression of an immunologically mediated process that can be immunoglobulin E (IgE) or non-IgE mediated; its many symptoms include urticaria/angioedema, asthma, rhinitis, and anaphylaxis. NCGS/NCWS is characterized by gastrointestinal and/or extra-intestinal symptoms after ingestion of gluten-containing food in subjects not affected by CD or WA. The aim of this review is to help physicians and nutritionists diagnose the cause of symptoms reported after wheat ingestion, thus avoiding patient frustration, inappropriate testing, and incorrect or missed diagnoses. An algorithm for the diagnostic approach in these patients is provided, to help to diagnose CD, WA, NCGS/NCWS or to identify possible functional disorders as the wheat-sensitive irritable bowel syndrome. A personalized approach, regular follow-up, and the help of a skilled healthcare professional are mandatory for patients with symptoms following wheat ingestion is provided. A gluten-free-diet is often recommended for patients with self-reported gluten/wheat-dependent symptoms; for patients with symptoms similar to those of functional diseases while there is evidence that a low-FODMAP diet could be the first option.

#### KEYWORDS

celiac disease, wheat allergy, gluten sensitivity, gluten-free-diet, food allergy, gluten, wheat-sensitive IBS

# Introduction

Over the last few decades, there has been much interest in sensitivity to or intolerance of wheat and its systemic and gastrointestinal clinical manifestations (1–3). However, confusion about the topic among clinicians and patients may lead to inappropriate testing and missed or incorrect diagnosis. The term food intolerance describes a large array of food-triggered complaints that do not involve the immune system and are experienced by up to one-fifth of the population (4). Lactose intolerance is common among the food intolerance (5, 6). Typical symptoms of food intolerance are mainly gastrointestinal and include gas, distention, bloating, nausea, and changes in stool consistency.

The global incidence of food allergies is rising, especially in industrialized countries. This increase is probably multifactorial, with changes in lifestyle and behavior playing a significant role. The hygiene hypothesis suggests that our environment has become "too clean" and the immune system may overreact unexpectedly to food proteins and create allergic responses (7). Many scientific studies are currently investigating the possible role of our gut microbiome (8, 9).

Many patients report symptoms after eating certain foods, but it is thought that the mechanism is not related to food intolerance or to allergy. Patient exposure to specific foods may also generate less specific and more diffuse symptoms, often described as food sensitivity. Patients frequently suspect the trigger is a gluten-containing food. The exact physiopathology is often unclear (3). In patients reporting symptoms following ingestion of gluten-containing food, it is essential to confirm or exclude celiac disease (CD), a common immune-mediated disease which affects approximately 1% of the population (10).

Celiac disease is a state of heightened immunological responsiveness to ingested gluten in some genetically susceptible individuals. The diagnosis is very important since lifelong adherence to a strict gluten-free diet (GFD) results in an optimal clinical outcome in the large majority of patients, although some report persistent symptoms despite following a GFD (11). If CD is excluded, the presence of certain symptoms (e.g., swelling, itching or irritation of the mouth/throat, hives or itchy rash of the skin, difficulty breathing) should lead clinicians to investigate a possible wheat allergy (WA). In case of exclusion of both CD and WA, non-celiac gluten (or wheat) sensitivity (NCGS/NCWS) should be considered in patients with reported symptoms only after gluten ingestion (12).

This review aims to present current evidence to help healthcare professionals—such as physicians or nutritionists follow the correct diagnostic approach and proper algorithm for managing patients presenting with symptoms after wheat ingestion.

# Possible diagnosis in patients reporting symptoms after wheat ingestion

### Celiac disease

Celiac disease is a chronic immune-mediated disease of the small bowel caused by exposure to dietary gluten which is composed by the related proteins glutelin and prolamin in genetically predisposed individuals. It is also known as glutensensitive enteropathy. The estimated global prevalence based on serological studies is approximately 1% (higher in Europe and North America) (13–15). It is diagnosed more frequently in females (F:M ratio of 2:1) and has a bimodal age at the diagnosis distribution, with an initial peak in the first 2 years of life and a second peak in the second or third decade (14).

Celiac disease is characterized by mucosal inflammation, villous atrophy, and crypt hyperplasia, which occur after exposure to dietary gluten and improve after withdrawal of gluten from the diet (11). Transglutaminase 2, the celiac autoantigen expressed in the intestinal mucosa, modifies immunogenic gluten peptides through deamidation of some charge-neutral glutamine residues, yielding negatively charged glutamic acid residues. This modification promotes gluten peptide presentation by human leukocyte antigen (HLA) DQ2 or DQ8 molecules on mucosal antigen-presenting cells and enables the activation and expansion of gluten peptide-specific CD4+ type 1 helper T cells and the secretion of proinflammatory cytokines. This process leads to villous atrophy and crypt hyperplasia and to B-cell differentiation and the production of transglutaminase 2 IgA (16).

It is crucial to differentiate CD from NCGS/NCWS because of the risks of nutritional deficiency and complications associated with CD. It is also important to differentiate CD from NCGS/NCWS since only for CD patients the adherence to a strict life-long GFD is necessary, considering the difficulty and the costs maintaining it (12, 17).

#### Diagnosis of celiac disease

While thereal benefit of general population screening to detect asymptomatic CD has not been demonstrated (18), appropriate tests to diagnose or exclude CD in patients reporting typical symptoms is essential.

Diagnosis is based on serological testing in adults with suggestive gastrointestinal symptoms including chronic or recurrent diarrhea, signs of malabsorption, unexpected weight loss, abdominal pain, and distension or bloating, or laboratory evidence suggestive of CD such as iron deficiency anemia, folate or vitamin B12 deficiency, and persistent elevation in serum aminotransferases. CD should also be investigated if patients report extra-intestinal signs or symptoms such as fatigue, recurrent headaches, recurrent fetal loss, low birthweight offspring, persistent aphthous stomatitis, dermatological manifestations, metabolic bone disease, or premature osteoporosis (19).

Patients with a very high probability of having CD (e.g., those with diarrhea with features of malabsorption, gastrointestinal signs, and chronic symptoms with a family history of CD) should undergo serological testing and duodenal biopsy to determine whether they have a rare form of seronegative CD or whether their symptoms have another cause (e.g., autoimmune enteropathy, giardiasis, Whipple's disease). All testing for CD should ideally be performed while the patient is on a gluten-containing diet (19). The diagnostic approach for adults is shown in **Figure 1**.

Transglutaminase 2 (tTG)-IgA antibody is the single preferred test for the detection of CD in adults due to its high sensitivity and specificity (20–22). In addition, total IgA levels should be measured since CD is associated with IgA deficiency in 2–3% of patients (23). In patients with IgA deficiency, tTG IgG-based testing, should be performed. The tTG-IgA and endomysial (EMA)-IgA antibody tests have similar sensitivity and both tests have high specificity (19). Deamidated gliadin peptide (DGP)-IgG may be performed for children <3 years old with a good diagnostic accuracy in this setting.

Adult patients with positive serological testing should undergo an upper endoscopy with small bowel biopsy to confirm the diagnosis. Multiple biopsies of the duodenum (at least four oriented biopsies of the post-bulbar duodenum) are recommended.

Histological features of CD range from mild alteration characterized only by increased intraepithelial lymphocytes to severe atrophic mucosa with complete loss of villi and crypt hyperplasia (24). The histological severity of intestinal lesions in CD is graded using the Marsh-Oberhuber classification (25). Subtotal or total atrophy (Marsh type 3 lesion), while not pathognomonic for CD, is diagnostic in association with positive serology.

Endoscopic features suggestive of CD have low sensitivity (59–94%), but the reported specificity ranges from 92 to 100%. These findings may be seen with other disorders such as giardiasis, autoimmune enteropathy, and HIV infection (26).

Celiac disease is strongly associated with the HLA class II genes HLA-DQ2 and HLA-DQ8 located on chromosome 6p21. Approximately 95% of individuals with CD express DQ2 heterodimer, while the remainder express HLA-DQ8, with a perfect negative predictive value for CD close to 100% in the absence of these genes. However, they are necessary but not sufficient for the diagnosis. Genetic testing should not be used routinely for the initial diagnosis of CD, but can be used to rule out CD in some clinical situations including Marsh 1–2

histology in seronegative patients, for the evaluation of patients not tested for CD before starting a GFD, and when the results of celiac-specific serology and histology are discrepant (19).

A food challenge should be considered if patients are already on a GFD and carry genetic susceptibility to CD: it is important to carefully evaluate the clinical response and patients should undergo serological tests and duodenal biopsy if necessary (19).

Only for children, diagnosis without an intestinal biopsy is an option for those young patients who have all three following characteristics: (a) tTG-IgA > 10 times the upper limit of normal; (b) positive EMA-IgA results; and (c) symptoms compatible with CD, as outlined in the guidelines of the European Society for Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) and the British Society for Paediatric Gastroenterology, Hepatology and Nutrition (BSPGHAN) (27, 28). On contrast, duodenum biopsy is necessary for adults also because a baseline histology can be useful in case of a CD refractory to the GFD.

Screening of asymptomatic first-degree family members of CD patients is recommended. HLA-genotype testing may be carried out as the first-line test; if negative, no further investigation is needed, but if positive, periodic screening with celiac-specific serology should be performed.

#### Management of celiac disease

It is very important to correctly diagnose CD given its excellent response to a GFD (11).

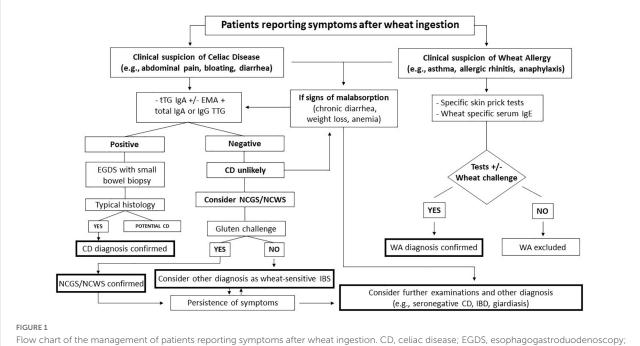
Celiac patients should be educated to avoid food products that contains gluten derived from wheat, barley or rye. On the other side, soybean or tapioca flours, rice, maize and buckwheat are inherently gluten-free and safe. Contamination needs to be avoided, paying also attention to labels on prepared foods and condiments that might contain additives (e.g., stabilizers, emulsifiers) containing gluten.

Separate speech must be made for oats, that should be introduced into the diet with caution, monitoring possible patients' adverse reactions. Oat consumption should be limited because very likely to be contaminated with wheat/rye/barley: patients can only consume certified gluten-free oats, with crossed grain symbol (19).

Newly diagnosed patients should be referred to a dietitian for discussion of dietary management, evaluation of potential nutrient deficiency, and provision of information on maintaining a strict GFD containing healthy alternatives to gluten (29, 30).

There is evidence that compliance with a GFD is better in those with more knowledge about the diet and CD. Also, the support of health providers and families has a strong positive impact (31).

In recent years, it has been observed that remote monitoring by nutritionists and physicians and point-of-care testing for gluten detection may be helpful in managing patients with CD and checking adherence to a GFD (32).



Flow chart of the management of patients reporting symptoms after wheat ingestion. CD, celiac disease; EGDS, esophagogastroduodenoscopy; EMA, endomysial antibody; NCGS/NCWS, non-celiac gluten-wheat sensitivity; tTG; transglutaminase; WA, wheat allergy; IBS, irritable bowel syndrome.

## Wheat allergy

#### Wheat allergy definition

Wheat allergy (WA) is the clinical expression of an immunologically mediated process that can be immunoglobulin E (IgE) or non-IgE mediated (33). IgE-mediated presentations include: (a) food allergy, (b) wheat-dependent exercise-induced anaphylaxis (WDEIA), (c) baker's asthma, and (d) contact dermatitis. The non-IgE-mediated, among which CD has been already analyzed, forms include also (a) eosinophil esophagitis, (b) eosinophil gastritis, and (c) dermatitis herpetiformis, where the eosinophilic pathologies are mediated by activation of innate immunity through IL-C2 activity, in contrast to CD and dermatitis herpetiformis which are caused by an immunemediated mechanism.

Among the different species of wheat, most allergology studies have focused on bread wheat (*Triticum aestivum*) as the marker of wheat sensitivity. Approximately 90% of the wheat cultivated worldwide is bread wheat, while the remaining 10% is durum wheat (*Triticum durum*) which grows well only in the dry climate of Southern Europe and Northern Africa. Recent studies have shown that the allergenic proteins in *T. durum* and *Triticum dicoccum* (spelt) are similar to those in *T. aestivum* and have similar allergenicity in adults and children (34).

#### Epidemiology of wheat allergy

The true prevalence of WA is unknown as self-reported food hypersensitivity overestimates prevalence, but few studies

have followed the gold standard method of diagnosis using a double-blind, placebo-controlled food challenge (DBPCFC). Recent studies by Gupta et al. in children and adults estimated prevalence was around 0.5% in children, and around 0.8% in adults, with 52% of cases developing in adulthood. In the USA, wheat is the most common adult-onset food allergy after shellfish (35). Interestingly, prevalence in children can depend upon the timing of wheat introduction, with higher prevalence if wheat is introduced after the age of 6 months (36). However, a high proportion of children tend to grow out of their WA (37, 38). Adult-onset WA seems to have a favorable prognosis, as recently described (39).

#### Clinical aspects

IgE-mediated reactions to wheat can manifest as different phenotypes. Food allergy to wheat can result from gastrointestinal or cutaneous contact, with different clinical presentations, while baker's asthma is due to sensitization through inhalation.

There are four types of wheat allergy:

(a) Classic wheat food allergy. WA manifests with a variety of symptoms that include urticaria/angioedema, asthma, allergic rhinitis, abdominal pain, vomiting, and anaphylaxis, and is diagnosed following the criteria established by the European Academy of Allergy and Clinical Immunology (EAACI) food allergy and anaphylaxis guidelines (40, 41). Rarely, WA can be a risk factor for severe anaphylaxis and death (42, 43). In children, it can worsen pre-existing atopic eczema.

- (b) Wheat-dependent exercise-induced anaphylaxis. WDEIA is a severe type of food allergy (44) that presents during or after intense exercise following wheat ingestion in wheat allergic subjects. Wheat is the most common food causing food-dependent exercise-induced anaphylaxis (FDEIA). Other factors can enhance anaphylaxis such as consumption of non-steroidal anti-inflammatory drugs (NSAIDs) or alcohol. Omega 5-gliadins (also called Tri a 19) are the major wheat allergen responsible for WDEIA. In a limited number of cases, high-molecular-weight glutenin subunits (HMW-GS) also plays a role in WDEIA (45), as well as the wheat lipid transfer proteins (LTP) Tri a 14, especially in Southern Europe. This is a particularly severe form and is frequently associated with anaphylaxis to maize or rice (46).
- (c) Baker's asthma. Baker's asthma and rhinitis are respiratory occupational diseases caused by sensitization to wheat flours, and are more frequent in atopic subjects (47). Baker's asthma is one of the most common forms of occupational asthma globally. Regulatory frameworks and workplace control measures should reduce flour dust exposures and related risks. Interestingly, affected subjects can usually safely eat wheat.
- (d) Contact dermatitis. Some cases of WDEIA reported between 2009 and 2012 in Japan were caused by mucocutaneous sensitization to hydrolyzed wheat protein added to soap (Glupearl 19) (48). A nationwide investigation revealed that over 2,000 individuals were affected. Interestingly, after the withdrawal of the soap, more than 50% of patients were in remission at 60 months (49).
- (e,f) Eosinophilic esophagitis and gastritis. Wheat plays an important role in non-IgE-mediated food allergy, especially in eosinophilic esophagitis and gastritis. These are considered T2 antigen-driven diseases characterized histologically by eosinophilic inflammation and symptoms related to esophageal dysfunction, such as dysphagia, food impaction, and vomiting due to eosinophilic infiltration. Even though atopy is a predisposing condition, IgE antibodies do not seem to play a role in the etiopathogenesis, which is actually driven by innate immunity stimulation through alteration of the mucosal barrier inducing the activation of IL-C2 cells that produce IL-13 and IL-5 responsible for eosinophil recruitment (50).

#### Pathogenesis

The pathogenesis of IgE-mediated WA likely shares the same mechanisms as other food allergies, beginning with a sensitization phase during which wheat allergens are processed by dendritic cells (DC). By interacting with the intestinal epithelial barrier, in the presence of a virus or other irritant stimulus, they cause the release of alarmins, such as IL-33, IL-25, and especially thymic stromal lymphopoietin (TSLP), which induces the expression of OX40 on dendritic cells type 2 (cDC2 103+). These induce the switch from Th0 to Th2 that stimulate B-lymphocytes to produce wheat allergen-specific IgE. The same cDC2 103+ in the absence of damaging stimuli, induce the response of T regs which is the basis of oral tolerance (51). The symptomatic phase is due to the interaction of IgE with mast cells that induces the release of preformed histamine and the production of inflammatory mediators such as prostaglandins, leukotrienes (52), PAF, and cytokines responsible for symptoms.

A recent study focused on the specific mechanism of sensitization to wheat. In a mouse model, skin was sensitized to wheat by repeated skin application of salt-soluble protein extract from durum wheat (53). After 9 weeks, the researchers elicited robust increases in wheat-specific IgE (sIgE) levels and anaphylaxis following oral administration of wheat. This research clearly demonstrates the power of wheat as a food allergen (54). It also supports the dual-allergen exposure hypothesis for the development of food allergies in a mouse model. The authors also demonstrated selective elevation of IL-6 upon oral allergen challenge, suggesting it has a key role in eliciting anaphylaxis *via* the oral route. A similar human model may be represented by the hydrolyzed wheat protein anaphylaxis associated with Glupearl 19 soap as a result of its repeated use, demonstrating sensitization through the skin (55).

#### Wheat allergens

Wheat proteins can be classified into two fractions based on solubility in salt. The salt soluble fraction includes albumins and globulin and represents 15–20% of total proteins. The salt insoluble fraction includes gliadin and gluten and accounts for approximately 80% of wheat protein content (56). Wheat allergens are present in both fractions. A total of 28 wheat allergens have been recognized by the International Union of Immunological Societies (IUIS), several of which are relevant to food allergy and are listed in **Table 1**.

Profilins (Tri a 12) are one of the allergens underlying crossreactivity with grass pollen. Recently peroxidase 1 (35 kDa) and beta-glucosidase (60 kDa) were identified as specific IgE-binding wheat proteins cross-reacting with grass pollen allergens (57, 58).

Non-specific lipid transfer proteins such as Tri a 14 from bread wheat and Tri tu 14 from durum wheat are allergens involved in IgE-mediated food allergy, WDEIA, and baker's asthma, especially in Southern Europe. These forms are also associated to non-IgE-mediated food allergy as demonstrated in a subject with baker's asthma and CD (59, 60).

Omega-5 gliadin (Tri a 19) have been identified as major allergens in WDEIA. Tri a 25, peroxidase, and serine proteinase inhibitor all have a role in baker's asthma. Alphaamylase/trypsin inhibitors are the major allergens in both baker's

Allergen	Biochemical name	MW (SDS-PAGE) (kDa)	Route of allergen exposure
Tri a 12	Profilin	14	Food
Tri a 14	Non-specific lipid transfer protein 1	9	Food
Tri a 19	Omega-5 gliadin	65	Food
Tri a 25	Thioredoxin	25	Food
Tri a 26	High-molecular-weight glutenin	88	Food
Tri a 27	Thiol reductase homolog	27	Airway
Tri a 28	Dimeric alpha-amylase inhibitor 0.19	13	Airway
Tri a 29	Tetrameric alpha-amylase inhibitor CM1/CM2	13	Airway
Tri a 32	1-cys-peroxiredoxin		Airway
Tri a Bd36 kd	Peroxidase	36	Food
Tri a 37	Alpha purothionin	12	Food
Tri a 39	Serine protease inhibitor-like protein		Airway

#### TABLE 1 Triticum aestivum (bread wheat) allergens.

MW, molecular weight; SDS-PAGE, sodium dodecyl sulfate-polyacrylamide gel electrophoresis.

asthma and food allergy (56, 61). They are heat resistant and lack significant cross-reactivity to grass pollen allergens. Tri a 37 is a plant defense protein. It is highly stable, resistant to heat and digestion, and a potent allergen (62).

High-molecular-weight glutenins (Tri a 26) are major allergen in WDEIA but have been described in only a few cases (63, 64).

#### Diagnosis of wheat allergy

The diagnosis of WA relies on the demonstration of IgE sensitization and clinical reactivity. Allergic sensitization must be confirmed by in vivo (skin prick tests) and/or in vitro (specific serum IgE) tests. If severe anaphylaxis has not been documented, it is important to verify symptoms with DBPCFC following the schedules and doses already published (65, 66). The diagnosis of WDEIA is challenging as the usual approach of performing exercise after a wheat-based meal (44) can lead to false-negative results. Brockow et al. analyzed patients with a clinical history of WDEIA and positive omega 5-gliadin-specific IgE levels. In all patients, including some with previous negative wheat challenge results, WDEIA was confirmed by challenge with up to 80 g of gluten alone or with gluten plus cofactors. The authors concluded that higher doses of gluten can elicit a reaction even in the absence of enhancing factors (65). This study seems to demonstrate that IgE positivity to Tri a 19 can be diagnostic for WDEIA, suggesting the importance of component-resolved diagnosis (CRD) in the diagnosis of WA. However, sIgE (and also the skin prick test) cannot distinguish between true WA and sensitization when sIgE to grass pollen is also present, due to the high rate of cross-reactivity between the two allergens (67). In addition to omega-5 gliadin (Tri a 19), which is strictly associated with WDEIA, positivity to Tri a 14, the wheat non-specific lipid transfer protein, is also associated with WDEIA, especially in patients with multiple LTP-containing vegetable food allergies. Given the high rate of cross-reactivity in these cases, a wheat challenge should always be performed (68). Other relevant allergens useful for diagnosis are alpha-amylase/trypsin inhibitors, which are associated in particular with baker's asthma.

Recent data suggest also a diagnostic role for the basophil activation test (69).

#### Management of wheat allergy

The management of WA varies according to the clinical manifestations and sensitization profile to various allergens. Depending on symptom severity, the diet should be based on either total wheat elimination or the maximum tolerated dose. Although a few patients with food allergy have severe reactions to very small amounts of allergen, most patients can tolerate low doses of the culprit food and may never experience a severe reaction (70). In these cases, consumption of low doses could help them reach tolerance more quickly. An epinephrine autoinjector should be provided to all patients with a history of systemic reactions (44).

Regarding WDEIA, the management of patients with omega-5 gliadin allergy depends on the severity of the reaction (71). Christensen et al. showed that regular intake of wheat not related to exercise can be recommended. The clinical threshold in WDEIA seems to be lowered in patients on a wheat-free diet, while the opposite is seen in patients with regular wheat intake long before exercise (63).

Interestingly, oral immunotherapy has been proposed as a new treatment for WA in children, showing promising results (71, 72).

## Non-celiac (gluten)-wheat sensitivity

The term NCGS was first used in a case report describing the resolution of persistent gastrointestinal symptoms with the adoption of a GFD in a patient in whom CD had been excluded (73).

Non-celiac (gluten) wheat sensitivity is characterized by gastrointestinal and/or extra-intestinal symptoms reported following gluten consumption in subjects not affected by CD or WA (74). Prevalence is difficult to estimate but, in the absence of identified risk factors, NCWS seems to be associated with female gender and young/middle age (75).

The clinical presentation of NCGS/NCWS includes gastrointestinal symptoms (abdominal pain, bloating, and altered bowel habit) and extra-intestinal symptoms, such as fatigue, headache, bone or joint pain, mood disorders, and skin manifestations (e.g., eczema or rash). Reported symptoms usually follow the consumption of gluten and improve after gluten withdrawal (75).

Non-celiac (gluten) wheat sensitivity is a diagnosis of exclusion; the gold standard for diagnosis is a doublestep approach as defined by the Salerno Experts Criteria (76). After exclusion of CD and WA, patients start a 6week gluten-containing diet followed by a GFD for at least 6 weeks. A decrease of at least 30% in the baseline score is considered a positive response. Step 2 includes a 1-week challenge (GFD and gluten or placebo) followed by a 1-week washout period following a strict GFD, and then crossover to the second 1-week challenge. A variation in symptoms of at least 30% between gluten and placebo challenge discriminates a positive from a negative result (76).

# Management of non-celiac (gluten) wheat sensitivity

For diagnosing NCGS/NCWS is essential that testing for CD and WA must be negative, symptoms must improve with a GFD, and diagnosis must be confirmed by the gluten challenge.

However, those who present with self-reported glutenrelated symptoms and may have both NCGS/NCWS and secondly those who present with IBS-type symptoms and could have gluten or wheat sensitive IBS. Those patients report that food plays an important role in their IBS-type symptoms with estimates of up to 80% of patients having postprandial symptomology, and up to 40% reporting specific "food intolerances." The fundamental difference between NCGS/NCWS and IBS is that patients with NCGS/NCWS self-report symptoms when consuming gluten and have identified or perceive gluten as the culprit. Conversely IBS patients do not report gluten as a specific stimulus for their symptoms. However, previously previous studies have demonstrated that wheat is a commonly reported "food intolerance" when IBS patients are specifically questioned (12).

There is insufficient evidence to determine whether NCGS/NCWS is a chronic relapsing/remitting disease or selflimiting disease and how strict the diet needs to be. A trial of gluten reintroduction can be attempted in individuals who remain asymptomatic for a period of time; however, symptoms with reintroduction are likely to be extremely variable among individuals (28). Therefore, once the diagnosis of NCGS/NCWS is reached, the management and follow-up of patients are not standardized, because of the unclear pathogenesis and variable clinical presentation (30, 76). The authors suggest that a logical approach could be to undertake a GFD for a limited period, followed by the gradual reintroduction of gluten. After some period on a GFT, the reintroduction of gluten could start with cereals of low gluten content (e.g., oats). Chronically, periods of GFD could be suggested during symptomatic relapses as a sort of "on-demand" therapy, also considering other possible treatments when there is the suspicion of a wheat sensitive IBS.

# Discussion

The diagnosis of CD and WA is objective, albeit difficult. nonetheless the Patient with symptoms can find a pathological explanation for them, and a specialist to whom they could refer for their disease (the gastroenterologist with the nutritionist or the allergologist), and the most effective therapy, even in the case of refractory CD or severe WA. However, there are no clear serological or histopathological criteria to confirm a diagnosis of NCGS/NCWS since antibodies (tTG-IgA and EMA-IgA) indicating CD are negative in NCGS/NCWS and histology does not show signs of atrophy.

In addition, patients with NCGS/NCWS may believe they do not have disease or conversely, that clinicians think they are imaging it. NCGS/NCWS has often been compared to functional diseases and described as an irritable bowel syndrome (IBS)like entity (12, 77). Indeed, NCGS/NCWS has been reported as wheat-sensitive irritable bowel syndrome when fulfilling the criteria for IBS according to the Rome Foundation (12).

Furthermore, symptoms sometimes improve in patients with either NCGS/NCWS or IBS after a reduction in FODMAP intake (fermentable, oligo-, di-, monosaccharides, and polyols) (78). FODMAPs enter the colon where they are fermented, causing the production of gas and since they are osmotically active, they can lead to increased water content in the intestinal lumen. This process is thought to be amplified in the presence of intestinal dysbiosis and may cause symptoms such as abdominal pain, diarrhea, flatulence, and bloating (79). A GFD is often recommended for patients with self-reported gluten/wheat-dependent symptoms, but a low-FODMAP diet should be the first option for patients with symptoms similar to those of functional diseases. In these patients, evidence may justify the prescription of a low-FODMAP diet because of its demonstrated beneficial effects In particular, three meta-analyses have demonstrated that a low FODMAP diet improves global symptoms in IBS patients compared with other dietary interventions (80–82). A recent randomized controlled study in patients with IBS with diarrhea (IBS-D) evaluated the efficacy and acceptability of a short-term strict low FODMAP diet and of a long-term modified FODMAP diet compared with traditional dietary advice, and showed that both the strict and modified low FODMAP diets are acceptable and lead to significant improvements in symptoms and quality of life (83). United European Gastroenterology (UEG) guidelines on functional bowel disorders with diarrhea recommend the shortterm use of a low FODMAP diet in patients with IBS-D when other measures have failed (84).

The complexity of the diet requires the involvement of experienced nutritionists (83, 85–87).

Nutritionists and physicians should clearly explain to patients referring all his symptoms to gluten ingestion that it is likely that in addition to gluten, other non-gluten wheat components, e.g., fructans, yeasts (which are scientifically known as FODMAPs) may also be the responsible for triggering symptoms.

It is likely that in some patients the ingestion of glutencontaining foods may have a nocebo effect, while in others the avoidance of gluten-containing foods could have a placebo effect (88). When wheat-sensitive IBS is suspected, also non-diet specific treatments should be considered, such as neuromodulators, probiotics, and physical activity (84, 89, 90).

In conclusion, a personalized approach, regular follow-up, and the help of a skilled nutritionist are mandatory for patients reporting symptoms after wheat ingestion.

# References

1. Aziz I, Hadjivassiliou M, Sanders DS. Does gluten sensitivity in the absence of coeliac disease exist? *BMJ*. (2012) 345:e7907. doi: 10.1136/bmj.e7907

2. Biesiekierski JR, Newnham ED, Irving PM, Barrett JS, Haines M, Doecke JD, et al. Gluten causes gastrointestinal symptoms in subjects without celiac disease: a double-blind randomized placebo-controlled trial. *Am J Gastroenterol.* (2011) 106:508–14;quiz515. doi: 10.1038/ajg.2010.487

3. Sapone A, Bai JC, Ciacci C, Dolinsek J, Green PH, Hadjivassiliou M, et al. Spectrum of gluten-related disorders: consensus on new nomenclature and classification. *BMC Med.* (2012) 10:13. doi: 10.1186/1741-7015-10-13

4. Young E, Stoneham MD, Petruckevitch A, Barton J, Rona R. A population study of food intolerance. *Lancet*. (1994) 343:1127–30. doi: 10.1016/s0140-6736(94) 90234-8

5. Lomer MC, Parkes GC, Sanderson JD. Review article: lactose intolerance in clinical practice-myths and realities. *Aliment Pharmacol Ther.* (2008) 27:93–103. doi: 10.1111/j.1365-2036.2007.03557

6. Shaukat A, Levitt MD, Taylor BC, MacDonald R, Shamliyan TA, Kane RL, et al. Systematic review: effective management strategies for lactose intolerance. *Ann Intern Med.* (2010) 152:797–803. doi: 10.7326/0003-4819-152-12-201006150-00241

7. Yazdanbakhsh M, Kremsner PG, van Ree R. Allergy, parasites, and the hygiene hypothesis. *Science*. (2002) 296:490–4. doi: 10.1126/science.296.5567.490

8. Huffnagle GB. The microbiota and allergies/asthma. *PLoS Pathog.* (2010) 6:e1000549. doi: 10.1371/journal.ppat.1000549

# Author contributions

AC, GA, GL, VS, and EP wrote the manuscript. LD reviewed the manuscript. LE, EP, LR, and MV supervised the study and critically reviewed the manuscript. All authors contributed to the article and approved the submitted version.

# Funding

This study was partially funded by Italian Ministry of Health - Current research IRCCS.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

9. Lambrecht BN, Hammad H. The immunology of the allergy epidemic and the hygiene hypothesis. *Nat Immunol.* (2017) 18:1076–83. doi: 10.1038/ni.3829

10. Singh P, Arora A, Strand TA, Leffler DA, Catassi C, Green PH, et al. Global prevalence of celiac disease: systematic review and meta-analysis. *Clin Gastroenterol Hepatol.* (2018) 16:823.e-36.e. doi: 10.1016/j.cgh.2017.0 6.037

11. Biagi F, Corazza GR. Mortality in celiac disease. Nat Rev Gastroenterol Hepatol. (2010) 7:158–62. doi: 10.1038/nrgastro.2010.2

12. Catassi C, Alaedini A, Bojarski C, Bonaz B, Bouma G, Carroccio A, et al. The overlapping area of non-celiac gluten sensitivity (NCGS) and wheat-sensitive irritable bowel syndrome (IBS): an update. *Nutrients.* (2017) 9:1268. doi: 10.3390/ nu9111268

13. Rubio-Tapia A, Ludvigsson JF, Brantner TL, Murray JA, Everhart JE. The prevalence of celiac disease in the United States. *Am J Gastroenterol.* (2012) 107:1538–44;quiz1537,1545. doi: 10.1038/ajg.2012.219

14. Lebwohl B, Rubio-Tapia A. Epidemiology, presentation, and diagnosis of celiac disease. *Gastroenterology.* (2021) 160:63–75. doi: 10.1053/j.gastro.2020.06. 098

15. Lebwohl B, Sanders DS, Green PHR. Coeliac disease. Lancet. (2018) 391:70-81. doi: 10.1016/S0140-6736(17)31796-8

16. Dieterich W, Ehnis T, Bauer M, Donner P, Volta U, Riecken EO, et al. Identification of tissue transglutaminase as the autoantigen of celiac disease. *Nat Med.* (1997) 3:797–801. doi: 10.1038/nm0797-797

17. Syage JA, Kelly CP, Dickason MA, Ramirez AC, Leon F, Dominguez R, et al. Determination of gluten consumption in celiac disease patients on a gluten-free diet. *Am J Clin Nutr.* (2018) 107:201–7. doi: 10.1093/ajcn/nqx049

18. Chou R, Bougatsos C, Blazina I, Mackey K, Grusing S, Selph S. Screening for celiac disease: evidence report and systematic review for the US preventive services task force. *JAMA*. (2017) 317:1258–68. doi: 10.1001/jama.2016.10395

19. Al-Toma A, Volta U, Auricchio R, Castillejo G, Sanders DS, Cellier C, et al. European society for the study of coeliac disease (ESsCD) guideline for coeliac disease and other gluten-related disorders. *United European Gastroenterol J.* (2019) 7:583–613. doi: 10.1177/2050640619844125

20. Dieterich W, Laag E, Schöpper H, Volta U, Ferguson A, Gillett H, et al. Autoantibodies to tissue transglutaminase as predictors of celiac disease. *Gastroenterology.* (1998) 115:1317–21. doi: 10.1016/s0016-5085(98)70 007-1

21. Li M, Yu L, Tiberti C, Bonamico M, Taki I, Miao D, et al. A report on the International transglutaminase autoantibody workshop for celiac disease. *Am J Gastroenterol.* (2009) 104:154–63. doi: 10.1038/ajg.2008.8

22. Molberg O, Mcadam SN, Körner R, Quarsten H, Kristiansen C, Madsen L, et al. Tissue transglutaminase selectively modifies gliadin peptides that are recognized by gut-derived T cells in celiac disease. *Nat Med.* (1998) 4:713–7. doi: 10.1038/nm0698-713

23. McGowan KE, Lyon ME, Butzner JD. Celiac disease and IgA deficiency: complications of serological testing approaches encountered in the clinic. *Clin Chem.* (2008) 54:1203–9. doi: 10.1373/clinchem.2008.103606

24. Ferguson A, Arranz E, O'Mahony S. Clinical and pathological spectrum of coeliac disease-active, silent, latent, potential. *Gut.* (1993) 34:150–1. doi: 10.1136/gut.34.2.150

25. Oberhuber G, Granditsch G, Vogelsang H. The histopathology of coeliac disease: time for a standardized report scheme for pathologists. *Eur J Gastroenterol Hepatol.* (1999) 11:1185–94. doi: 10.1097/00042737-199910000-00019

26. Shah VH, Rotterdam H, Kotler DP, Fasano A, Green PH. All that scallops is not celiac disease. *Gastrointest Endosc.* (2000) 51:717–20. doi: 10.1067/mge.2000. 104977

27. Husby S, Koletzko S, Korponay-Szabó I, Kurppa K, Mearin ML, Ribes-Koninckx C, et al. European society paediatric gastroenterology, Hepatology and Nutrition guidelines for diagnosing coeliac disease 2020. *J Pediatr Gastroenterol Nutr.* (2020) 70:141–56. doi: 10.1097/MPG.000000000002497

28. Murch S, Jenkins H, Auth M, Bremner R, Butt A, France S, et al. Joint BSPGHAN and coeliac UK guidelines for the diagnosis and management of coeliac disease in children. *Arch Dis Child*. (2013) 98:806–11. doi: 10.1136/archdischild-2013-303996

29. Hopper AD, Hadjivassiliou M, Butt S, Sanders DS. Adult coeliac disease. *BMJ*. (2007) 335:558–62. doi: 10.1136/bmj.39316.442338.AD

30. Elli L, Ferretti F, Orlando S, Vecchi M, Monguzzi E, Roncoroni L. Management of celiac disease in daily clinical practice. *Eur J Intern Med.* (2019) 61:15–24. doi: 10.1016/j.ejim.2018.11.012

31. Ludvigsson JF, Card T, Ciclitira PJ, Swift GL, Nasr I, Sanders DS, et al. Support for patients with celiac disease: a literature review. *United European Gastroenterol* J. (2015) 3:146–59. doi: 10.1177/2050640614562599

32. Costantino A, Roncoroni L, Noviello D, Nandi N, Lombardo V, Scricciolo A, et al. Nutritional and gastroenterological monitoring of patients with celiac disease during COVID-19 pandemic: the emerging role of telemedicine and point-of-care gluten detection tests. *Front Nutr.* (2021) 8:622514. doi: 10.3389/fnut.2021.622514

33. Sievers S, Rohrbach A, Beyer K. Wheat-induced food allergy in childhood: ancient grains seem no way out. *Eur J Nutr.* (2020) 59:2693–707. doi: 10.1007/s00394-019-02116-z

34. Pastorello EA, Farioli L, Robino AM, Trambaioli C, Conti A, Pravettoni V. A lipid transfer protein involved in occupational sensitization to spelt. *J Allergy Clin Immunol.* (2001) 108:145–6. doi: 10.1067/mai.2001.114988

35. Gupta RS, Warren CM, Smith BM, Jiang J, Blumenstock JA, Davis MM, et al. Prevalence and severity of food allergies among US adults. *JAMA Netw Open.* (2019) 2:e185630. doi: 10.1001/jamanetw

36. Nishimura T, Fukazawa M, Fukuoka K, Okasora T, Yamada S, Kyo S, et al. Early introduction of very small amounts of multiple foods to infants: a randomized trial. *Allergol Int.* (2022) 71:345–53. doi: 10.1016/j.alit.2022.03.0015

37. Sicherer SH, Warren CM, Dant C, Gupta RS, Nadeau KC. Food allergy from infancy through adulthood. *J Allergy Clin Immunol.* (2020) 8:1854–64. doi: 10.1016/j.jaip.2020.02.010

38. Poole JA, Barriga K, Leung DY, Hoffman M, Eisenbarth GS, Rewers M, et al. Timing of initial exposure to cereal grains and the risk of wheat allergy. *Pediatrics.* (2006) 117:2175–82. doi: 10.1542/peds.2005-1803 39. Scibilia J, Rossi CM, Losappio LM, Mirone C, Farioli L, Pravettoni V, et al. Favorable prognosis of wheat allergy in adults. J Invest Allergol Clin Immunol. (2019) 29:118–23. doi: 10.18176/jiaci.0296

40. Salcedo G, Quirce S, Diaz-Perales A. Wheat allergens associated with baker's asthma. J Invest Allergol Clin Immunol. (2011) 21:81–94.

41. Muraro A, Werfel T, Hoffmann-Sommergruber K, Roberts G, Beyer K, Bindslev-Jensen C, et al. EAACI food allergy and anaphylaxis guidelines: diagnosis and management of food allergy. *Allergy*. (2014) 69:1008–25. doi: 10.1111/all.12429

42. Simons FE. Anaphylaxis. J Allergy Clin Immunol. (2010) 125(Suppl. 2):S161-81. doi: 10.1016/j.jaci.2009.12.981

43. Dorris S. Fatal food anaphylaxis: registering a rare outcome. Ann Allergy Asthma Immunol. (2020) 124:445-6. doi: 10.1016/j.anai.2020.02.001

44. Scherf KA, Brockow K, Biedermann T, Koehler P, Wieser H. Wheatdependent exercise-induced anaphylaxis. *Clin Exp Allergy.* (2016) 46:10–20. doi: 10.1111/cea.12640

45. Feldweg AM. Food-dependent, exercise-induced anaphylaxis: diagnosis and management in the outpatient setting. *J Allergy Clin Immunol Pract.* (2017) 5:283–8. doi: 10.1016/j.jaip.2016.11.022

46. Pastorello EA, Farioli L, Stafylaraki C, Scibilia J, Mirone C, Pravettoni V, et al. Wheat-dependent exercise-induced anaphylaxis caused by a lipid transfer protein and not by  $\omega$ -5 gliadin. Ann Allergy Asthma Immunol. (2014) 112:386.e–7.e. doi: 10.1016/j.anai.2014.01.012

47. Quirce S, Diaz-Perales A. Diagnosis and management of grain-induced asthma. *Allergy Asthma Immunol Res.* (2013) 5:348–56. doi: 10.4168/aair.2013.5. 6.348

48. Chinuki Y, Morita E. Wheat-dependent exercise-induced anaphylaxis sensitized with hydrolyzed wheat protein in soap. *Allergol Int.* (2012) 61:529–37. doi: 10.2332/a

49. Hiragun M, Ishii K, Yanase Y, Hiragun T, Hide M. Remission rate of patients with wheat allergy sensitized to hydrolyzed wheat protein in facial soap. *Allergol Int.* (2016) 65:109–11. doi: 10.1016/j.alit.2015.07.004

50. Gonsalves NP, Aceves SS. Diagnosis and treatment of eosinophilic esophagitis. J Allergy Clin Immunol. (2020) 145:1–7. doi: 10.1016/j.jaci.2019.11.011

51. Sampath V, Tupa D, Graham MT, Chatila TA, Spergel JM, Nadeau KC. Deciphering the black box of food allergy mechanisms. *Ann Allergy Asthma Immunol.* (2017) 118:21–7. doi: 10.1016/j.anai.2016.10.017

52. Gould HJ, Sutton BJ. IgE in allergy and asthma today. *Nat Rev Immunol.* (2008) 8:205–17. doi: 10.1038/nri2273

53. Lozano-Ojalvo D, Berin C, Tordesillas L. Immune basis of allergic reactions to food. J Invest Allergol Clin Immunol. (2019) 29:1–14. doi: 10.18176/jiaci.0355

54. Gao H, Jorgensen R, Raghunath R, Ng PKW, Gangur V. An adjuvantfree mouse model using skin sensitization without tape-stripping followed by oral elicitation of anaphylaxis: a novel pre-clinical tool for testing intrinsic wheat allergenicity. *Front Allergy.* (2022) 3:926576. doi: 10.3389/falgy.2022.926576

55. Laurière M, Pecquet C, Bouchez-Mahiout I, Snégaroff J, Bayrou O, Raison-Peyron N, et al. Hydrolysed wheat proteins present in cosmetics can induce immediate hypersensitivities. *Contact Derm.* (2006) 54:283–9. doi: 10.1111/j.0105-1873.2006.00830.x

56. Pastorello EA, Farioli L, Conti A, Pravettoni V, Bonomi S, Iametti S, et al. Wheat IgE-mediated food allergy in European patients: alpha-amylase inhibitors, lipid transfer proteins and low-molecular-weight glutenins. Allergenic molecules recognized by double-blind, placebo-controlled food challenge. *Int Arch Allergy Immunol.* (2007) 144:10–22. doi: 10.1159/000102609

57. Nilsson N, Nilsson C, Ekoff H, Wieser-Pahr S, Borres MP, Valenta R, et al. Grass-allergic children frequently show asymptomatic low-level IgE cosensitization and cross-reactivity to wheat. *Int Arch Allergy Immunol.* (2018) 177:135–44. doi: 10.1159/000489610

58. Ogino R, Chinuki Y, Yokooji T, Takizawa D, Matsuo H, Morita E. Identification of peroxidase-1 and beta-glucosidase as cross-reactive wheat allergens in grass pollen-related wheat allergy. *Allergol Int.* (2021) 70:215–22. doi: 10.1016/j.alit.2020.09.005

59. Pastorello EA, Aversano MG, Mascheri A, Farioli L, Losappio LM, Mirone C, et al. Celiac disease in a patient with baker's asthma and wheat allergy due to Tri a 14. *Case Rep Clin Med.* (2015) 4:253–6.

60. Missaoui K, Gonzalez-Klein Z, Jemli S, Garrido-Arandia M, Diaz-Perales A, Tome-Amat J, et al. Identification and molecular characterization of a novel non-specific lipid transfer protein (TdLTP2) from durum wheat. *PLoS One.* (2022) 17:e0266971. doi: 10.1371/journal.pone.0266971

61. Geisslitz S, Weegels P, Shewry P, Zevallos V, Masci S, Sorrells M, et al. Wheat amylase/trypsin inhibitors (ATIs): occurrence, function and health aspects. *Eur J Nutr.* (2022) 61:2873–80. doi: 10.1007/s00394-022-02841-y

62. Cianferoni A. Wheat allergy: diagnosis and management. J Asthma Allergy. (2016) 9:13–25. doi: 10.2147/JAA.S81550

63. Christensen MJ, Eller E, Mortz CG, Brockow K, Bindslev-Jensen C. Clinical and serological follow-up of patients with WDEIA. *Clin Trans Allergy*. (2019) 9:26. doi: 10.1186/s13601-019-0265-8

64. Morita E, Matsuo H, Chinuki Y, Takahashi H, Dahlström J, Tanaka A. Food-dependent exercise-induced anaphylaxis - importance of omega-5 gliadin and HMW-glutenin as causative antigens for wheat-dependent exercise-induced anaphylaxis. *Allergol Int.* (2009) 58:493–8. doi: 10.2332/allergolint.09-RAI-0125

65. Brockow K, Kneissl D, Valentini L, Zelger O, Grosber M, Kugler C, et al. Using a gluten oral food challenge protocol to improve diagnosis of wheat-dependent exercise-induced anaphylaxis. *J Allergy Clin Immunol.* (2015) 135:977.e–84.e. doi: 10.1016/j.jaci.2014.08.024

66. Scibilia J, Pastorello EA, Zisa G, Ottolenghi A, Bindslev-Jensen C, Pravettoni V, et al. Wheat allergy: a double-blind, placebo-controlled study in adults. *J Allergy Clin Immunol.* (2006) 117:433–9. doi: 10.1016/j.jaci.2005.10.014

67. Sander I, Raulf-Heimsoth M, Düser M, Flagge A, Czuppon AB, Baur X. Differentiation between cosensitization and cross-reactivity in wheat flour and grass pollen-sensitized subjects. *Int Arch Allergy Immunol.* (1997) 112:378–85. doi: 10.1159/000237483

68. Pastorello EA, Toscano A, Scibilia G, Stafylaraki C, Rossi CM, Borgonovo L, et al. Clinical features of wheat allergy are significantly different between Tri a 14 sensitized patients and Tri a 19 sensitized ones. *Int Arch Allergy Immunol.* (2022) 183:591–9. doi: 10.1159/000520936

69. Gabler AM, Gebhard J, Norwig MC, Eberlein B, Biedermann T, Brockow K, et al. Basophil activation to gluten and non-gluten proteins in wheat-dependent exercise-induced anaphylaxis. *Front Allergy.* (2022) 3:822554. doi: 10.3389/falgy. 2022.822554

70. Sicherer SH, Abrams EM, Nowak-Wegrzyn A, Hourihane JO. Managing food allergy when the patient is not highly allergic. *J Allergy Clin Immunol Pract.* (2022) 10:46–55. doi: 10.1016/j.jaip.2021.05

71. Nowak-Węgrzyn A, Wood RA, Nadeau KC, Pongracic JA, Henning AK, Lindblad RW, et al. Multicenter, randomized, double-blind, placebo-controlled clinical trial of vital wheat gluten oral immunotherapy. *J Allergy Clin Immunol.* (2019) 143:651.e–61.e. doi: 10.1016/j.jaci.2018.08.041

72. Babaie D, Ebisawa M, Soheili H, Ghasemi R, Zandieh F, Sahragard M, et al. Oral wheat immunotherapy: long-term follow-up in children with wheat anaphylaxis. *Int Arch Allergy Immunol.* (2022) 183:306–14. doi: 10.1159/00051 9692

73. Ellis A, Linaker BD. Non-coeliac gluten sensitivity? *Lancet.* (1978) 1:1358–9. doi: 10.1016/s0140-6736(78)92427-3

74. Fasano A, Sapone A, Zevallos V, Schuppan D. Nonceliac gluten sensitivity. *Gastroenterology.* (2015) 148:1195–204. doi: 10.1053/j.gastro.2014.12.049

75. Aziz I, Lewis NR, Hadjivassiliou M, Winfield SN, Rugg N, Kelsall A, et al. A UK study assessing the population prevalence of self-reported gluten sensitivity and referral characteristics to secondary care. *Eur J Gastroenterol Hepatol.* (2014) 26:33–9. doi: 10.1097/01.meg.0000435546. 87251.f7

76. Catassi C, Elli L, Bonaz B, Bouma G, Carroccio A, Castillejo G, et al. Diagnosis of non-celiac gluten sensitivity (NCGS): the salerno experts' criteria. *Nutrients.* (2015) 7:4966–77. doi: 10.3390/nu7064966

77. Volta U, Caio G, Karunaratne TB, Alaedini A, De Giorgio R. Non-coeliac gluten/wheat sensitivity: advances in knowledge and relevant questions. *Expert Rev Gastroenterol Hepatol.* (2017) 11:9–18. doi: 10.1080/17474124.2017.1260003

78. Roncoroni L, Bascuñán KA, Doneda L, Scricciolo A, Lombardo V, Branchi F, et al. A low FODMAP gluten-free diet improves functional gastrointestinal disorders and overall mental health of celiac disease patients: a randomized controlled trial. *Nutrients*. (2018) 10:1023. doi: 10.3390/nu10081023

79. Böhn L, Störsrud S, Törnblom H, Bengtsson U, Simren M. Self-reported food-related gastrointestinal symptoms in IBS are common and associated with more severe symptoms and reduced quality of life. *Am J Gastroenterol.* (2013) 108:634–41. doi: 10.1038/ajg.2013.105

80. Schumann D, Klose P, Lauche R, Dobos G, Langhorst J, Cramer H. Low fermentable, oligo-di-mono-saccharides and polyol diet in the treatment of irritable bowel syndrome: a systematic review and meta-analysis. *Nutrition.* (2018) 45:24–31. doi: 10.1016/j.nut.2017.07.004

81. Dionne J, Ford AC, Yuan Y, Chey WD, Lacy BE, Saito YA, et al. A systematic review and meta-analysis evaluating the efficacy of a gluten-free diet and a low FODMAPs diet in treating symptoms of irritable bowel syndrome. *Am J Gastroenterol.* (2018) 113:1290–300. doi: 10.1038/s41395-018-0195-4

82. Lanenvan AS, Bree de A, Greyling A. Efficacy of a low-FODMAP diet in adult irritable bowel syndrome: a systematic review and meta-analysis. *Eur J Nutr.* (2021) 60:3505–22. doi: 10.1007/s00394-020-02473-0

83. Goyal O, Batta S, Nohria S, Kishore H, Goyal P, Sehgal R, et al. Low fermentable oligosaccharide, disaccharide, monosaccharide, and polyol diet in patients with diarrhea-predominant irritable bowel syndrome: a prospective, randomized trial. *J Gastroenterol Hepatol.* (2021) 36:2107–15. doi: 10.1111/jgh. 15410

84. Savarino E, Zingone F, Barberio B, Marasco G, Akyuz F, Akpinar H, et al. Functional bowel disorders with diarrhoea: clinical guidelines of the United European gastroenterology and European society for neurogastroenterology and motility. *United European Gastroenterol J*. (2022) 10:556–84. doi: 10.1002/ueg2. 12259

85. Whelan K, Martin LD, Staudacher HM, Lomer MCE. The low FODMAP diet in the management of irritable bowel syndrome: an evidence-based review of FODMAP restriction, reintroduction and personalisation in clinical practice. *J Hum Nutr Diet.* (2018) 31:239–55. doi: 10.1111/jhn.12530

86. Rej A, Sanders DS, Buckle R, Buckle RL, Aziz I, Shaw C. What is the optimal FODMAP threshold in IBS? *J Gastroenterol Hepatol.* (2021) 36:1723-5. doi: 10.1111/jgh.15470

87. Rej A, Aziz I, Tornblom H, Sanders DS, Simren M. The role of diet in irritable bowel syndrome: implications for dietary advice. *J Intern Med.* (2019) 286:490–502. doi: 10.1111/joim.12966

88. Aziz I, Hadjivassiliou M, Sanders DS. The spectrum of noncoeliac gluten sensitivity. *Nat Rev Gastroenterol Hepatol.* (2015) 12:516–26. doi: 10.1038/nrgastro. 2015.107

89. Vasant DH, Paine PA, Black CJ, Houghton LA, Everitt HA, Corsetti M, et al. British society of gastroenterology guidelines on the management of irritable bowel syndrome. *Gut.* (2021) 70:1214–40. doi: 10.1136/gutjnl-2021-324598

90. Costantino, A, Pessarelli P, Vecchiato M, Vecchi M, Basilisco G, Ermolao A. A practical guide to the proper prescription of physical activity in patients with irritable bowel syndrome. *Digest Liver Dis.* (2022). doi: 10.1016/j.dld.2022.08.034 [Epub ahead of print].