



Chronic myeloproliferative neoplasms

Philadelphia chromosome-negative classical myeloproliferative neoplasms: revised management recommendations from European LeukemiaNet

Tiziano Barbui¹ · Ayalew Tefferi² · Alessandro M. Vannucchi³ · Francesco Passamonti⁴ · Richard T. Silver⁵ · Ronald Hoffman⁶ · Srdan Verstovsek⁷ · Ruben Mesa⁸ · Jean-Jacques Kiladjian⁹ · Rüdiger Hehlmann¹⁰ · Andreas Reiter¹⁰ · Francisco Cervantes¹¹ · Claire Harrison¹² · Mary Frances Mc Mullin¹³ · Hans Carl Hasselbalch¹⁴ · Steffen Koschmieder¹⁵ · Monia Marchetti¹⁶ · Andrea Bacigalupo¹⁷ · Guido Finazzi¹ · Nicolaus Kroeger¹⁸ · Martin Griesshammer¹⁹ · Gunnar Birgegard²⁰ · Giovanni Barosi²¹

Received: 27 November 2017 / Revised: 16 January 2018 / Accepted: 26 January 2018 / Published online: 27 February 2018
© Macmillan Publishers Limited, part of Springer Nature 2018

Abstract

This document updates the recommendations on the management of Philadelphia chromosome-negative myeloproliferative neoplasms (Ph-neg MPNs) published in 2011 by the European LeukemiaNet (ELN) consortium. Recommendations were produced by multiple-step formalized procedures of group discussion. A critical appraisal of evidence by using Grades of Recommendation, Assessment, Development and Evaluation (GRADE) methodology was performed in the areas where at least one randomized clinical trial was published. Seven randomized controlled trials provided the evidence base; earlier phase trials also informed recommendation development. Key differences from the 2011 diagnostic recommendations included: lower threshold values for hemoglobin and hematocrit and bone marrow examination for diagnosis of polycythemia vera (PV), according to the revised WHO criteria; the search for complementary clonal markers, such as *ASXL1*, *EZH2*, *IDH1/IDH2*, and *SRSF2* for the diagnosis of myelofibrosis (MF) in patients who test negative for *JAK2V617*, *CALR* or *MPL* driver mutations. Regarding key differences of therapy recommendations, both recombinant interferon alpha and the JAK1/JAK2 inhibitor ruxolitinib are recommended as second-line therapies for PV patients who are intolerant or have inadequate response to hydroxyurea. Ruxolitinib is recommended as first-line approach for MF-associated splenomegaly in patients with intermediate-2 or high-risk disease; in case of intermediate-1 disease, ruxolitinib is recommended in highly symptomatic splenomegaly. Allogeneic stem cell transplantation is recommended for transplant-eligible MF patients with high or intermediate-2 risk score. Allogeneic stem cell transplantation is also recommended for transplant-eligible MF patients with intermediate-1 risk score who present with either refractory, transfusion-dependent anemia, blasts in peripheral blood > 2%, adverse cytogenetics, or high-risk mutations. In these situations, the transplant procedure should be performed in a controlled setting.

Introduction

Over the last few years, significant progress has been made in a number of areas of Philadelphia chromosome-negative myeloproliferative neoplasms (Ph-neg MPNs), i.e., polycythemia vera (PV), essential thrombocythemia (ET), and myelofibrosis (MF). The observation that specific mutations of the calreticulin (*CALR*) gene are present in a significant proportion of patients with ET and MF has further expanded the knowledge of driver mutation profile providing new diagnostic and prognostic opportunities [1, 2]. Additional, non-driver, myeloid gene mutations with prognostic relevance in MF have been identified [3, 4]. Moreover, a

These authors contributed equally: Tiziano Barbui, Ayalew Tefferi.

Electronic supplementary material The online version of this article (<https://doi.org/10.1038/s41375-018-0077-1>) contains supplementary material, which is available to authorized users.

✉ Giovanni Barosi
barosig@smatteo.pv.it

Extended author information available on the last page of the article

considerable number of randomized controlled trials have examined pharmacological and non-pharmacological interventions [5–14].

The new strides in the knowledge of Ph-neg MPNs have been matched by unprecedented efforts toward providing physicians with guidance principles and practice recommendations to appropriately translate these novelties into clinical use [15–23]. However, the majority of the published recommendations are based on subjective judgments without rigorous appraisal of existing evidence. Therefore, the decisions on the best management of patients with Ph-neg MPNs remains a challenge requiring a high degree of professional expertise.

This project aims to update the 2011 European LeukemiaNet (ELN) recommendations on the management of Ph-neg MPNs [24]. The goal was to produce consensus-based recommendations in the areas where no scientific evidence is available, and evidence-based recommendations when controlled clinical trials were available.

Methods

An Expert Panel (hereafter referred to as the Panel) of 23 experts was selected for their expertise in research and clinical practice of Ph-neg MPNs. A clinician with expertise in clinical epidemiology (G. Ba.) coordinated the process. During an initial meeting, the subset of key-questions deserving revision among those exploited in the previous guidelines document was chosen. A structured literature search for English-language publications using electronic data-bases such as MEDLINE, 2011–2017 (March) and EMBASE, 2011–2017 (March) was subsequently performed. Publications which measured efficacy of drugs or procedures in individuals with Ph-neg MPNs with a comparison group were critically appraised to rate confidence in estimates of effect for each outcome. For this last objective, we used the methodology of Grades of Recommendation, Assessment, Development and Evaluation (GRADE) [25]. For each trial, we first analyzed the internal validity with use of the Cochrane Collaboration's risk of bias assessment tool [26]. We then evaluated the external validity by assessing four dimensions of trials quality: precision, consistency, directness, and publication bias.

Multiple-step procedures were utilized for achieving recommendations. Two panelists (TB and AT) drafted the updated statements addressing the identified key questions. Subsequently, each panelist scored his/her agreement with those statements and provided suggestions for rephrasing. In order to exploit this phase of the process, a five-round Delphi process was conducted [27].

Results

Diagnostic procedures for Ph-neg MPNs

In patients with clinical or hematologic phenotype of PV, the 2016 revised World Health Organization (WHO) classification uses lower threshold values for hemoglobin and hematocrit [28] (Table 1). These lower thresholds require bone marrow (BM) examination for correct diagnosis [29].

In ET, *CALR* mutations were added to the *JAK2* and *MPL* driver mutations as a major diagnostic criteria [30]. In MF, the 2016-WHO classification distinguishes pre-fibrotic (pre-PMF) from ET and from overtly fibrotic primary MF (PMF) [31]. The diagnosis of post-PV or post-ET MF should be consistent with the criteria published by the International Working Group for MPN Research and Treatment (IWG-MRT) [32].

In addition to the three driver mutations (*JAK2*, *CALR*, and *MPL*), PV, ET, and MF are all characterized by the optional presence of additional somatic mutations that include *LNK*, *CBL*, *TET2*, *ASXL1*, *IDH*, *IKZF1*, *EZH2*, *DNMT3A*, *TP53*, *NFE2*, *SF3B1*, *SRSF2*, *U2AF1* as well as atypical *MPL* or *JAK2* mutations [4, 33, 34]. In the absence of any of the three driver clonal mutations, the search for the most frequent additional mutations is of help in determining the clonal nature of the disease.

Recommendations

In all three categories of Ph-neg MPNs, i.e., PV, ET, and MF, strict adherence to the 2016 revised WHO diagnostic criteria is recommended.

BM biopsy is a necessary diagnostic test in any patient suspected of a Ph-neg MPN, with the exception of patients with PV with a hemoglobin greater than 18.5 g/dL in males and greater than 16.5 g/dl in females.

Peripheral blood or BM screening for driver mutations, i.e., *JAK2V617F*, *CALR*, and *MPL*, is recommended in any patient who may have a Ph-neg MPN.

JAK2V617F should be the first test in patients suspected of any of the three diseases; in the case of *JAK2V617F* negativity, *CALR* and *MPL*, in that order, should then be tested in ET and MF, while *JAK2* exon 12 should be tested in those *JAK2V617F* negative patients suspected of PV.

Search for complementary clonal markers, such as *ASXL1*, *EZH2*, *IDH1/IDH2*, and *SRSF2* is recommended in patients who tested negative for the three driver mutations and have BM features and a clinical phenotype consistent with MF.

There was no consensus concerning the search of additional clonal markers such as *TP53*, *TET2*, *DNMT3A*, and *CBL* in MF, and no consensus concerning the need for

Table 1 World Health Organization (WHO) 2016 diagnostic criteria in chronic myeloproliferative neoplasms

	Polycythemia vera (PV) ^a	Essential thrombocythemia (ET) ^b	Primary myelofibrosis (PMF) ^c	Overt PMF ^f
Major criteria				
1	Hemoglobin > 16.5 g/dl (men) > 16 g/dl (women) or hematocrit > 49% (men) > 48% (women) or increased red cell mass (RCM)	Platelet count $\geq 450 \times 10^9/l$	Pre-PMF Megakaryocytic proliferation and atypia, ^d without reticulin fibrosis > grade 1, ^e accompanied by increased age-adjusted bone marrow cellularity, granulocytic proliferation, and often decreased erythropoiesis	Megakaryocyte proliferation and atypia ^d accompanied by either reticulin and/or collagen fibrosis (grade 2 or 3)
2	BM with age-adjusted hypercellularity and trilineage myeloproliferation with pleomorphic, mature megakaryocytes (differences in size)	BM with megakaryocyte proliferation with large and mature morphology. No significant left-shift of neutrophil granulopoiesis or erythropoiesis and very rarely minor (grade 1) increase in reticulin fibers	Not meeting WHO criteria for BCR-ABL1 + CML, PV, ET, MDS, or other myeloid neoplasm	Not meeting WHO criteria for BCR-ABL1 + CML, PV, ET, MDS, or other myeloid neoplasm
3	Presence of JAK2 mutation	Not meeting WHO criteria for BCR-ABL1 + CML, PV, PMF, MDS, or other myeloid neoplasm	Presence of JAK2, CALR, or MPL mutation or in the absence of these mutations, presence of another clonal marker ^f or absence of minor reactive bone marrow reticulin fibrosis ^g	Presence of JAK2, CALR, or MPL mutation or in the absence, the presence of another clonal marker ^f or absence of evidence for reactive bone marrow fibrosis ^h
4		Presence of JAK2, CALR, or MPL mutation		
Minor criteria				
1	Subnormal serum erythropoietin level	Presence of a clonal marker (e.g., abnormal karyotype) or absence of evidence for reactive thrombocytosis	Presence of one or more of the following: Anemia not attributed to a comorbid condition Palpable splenomegaly Leukocytosis $\geq 11 \times 10^9/l$ Elevated LDH ⁱ	Presence of one or more of the following: Anemia not attributed to a comorbid condition Palpable splenomegaly Leukocytosis $\geq 11 \times 10^9/L$ Elevated LDH ⁱ Leukoerythroblastosis

BM bone marrow, CML chronic myeloid leukemia, MDS myelodysplastic syndrome

^a PV diagnosis requires meeting either all three major criteria or the first two major criteria and one minor criterion

^b ET diagnosis requires meeting all four major criteria or first three major criteria and one minor criterion

^c prePMF diagnosis requires all three major criteria and at least one minor criterion

^d Small-to-large megakaryocytes with aberrant nuclear/cytoplasmic ratio and hyperchromatic and irregularly folded nuclei and dense clustering

^e In cases with grade 1 reticulin fibrosis, the megakaryocyte changes must be accompanied by increased marrow cellularity, granulocytic proliferation and often decreased erythropoiesis (that is, prePMF)

^f In the absence of any of the 3 major clonal mutations, the search for the most frequent accompanying mutations (ASXL1, EZH2, TET2, SRSF2, SF3B1) are of help in determining the clonal nature of the disease.

^g Minor (grade I) reticulin fibrosis secondary to infection, autoimmune disorder or another chronic inflammatory condition.

^h Bone marrow fibrosis secondary to infection, autoimmune disorder or another chronic inflammatory condition

ⁱ Confirmed in two consecutive determinations.

^j Above the upper limit of the institutional reference range.

searching for complementary clonal markers in ET. Thus, this decision should follow individual institutional preference.

Risk stratification

Risk stratification in PV and ET

The conventional prognostic systems in PV and ET are based upon age and history of thrombosis, which separate patients into low-risk (age < 60 years and no history of thrombosis) or high-risk (age ≥ 60 years or prior thrombosis) categories [35–37].

In PV, evidence is accumulating that the leukocyte count is implicated in the process of thrombogenesis [38, 39]. However, an intermediate-risk category in PV that includes younger patients with coexisting cardiovascular risk factors or leukocytosis in the absence of previous thrombosis, has not yet been formally validated.

In ET, a new scoring system that includes *JAK2V617F* mutation and general cardiovascular risk factors as predictors of thrombosis (IPSET-thrombosis), has been developed, revised, and validated [40–43] (Table 2).

Recommendations

In PV, age and previous thrombosis history are the prognostic parameters used to classify patients in low-risk and high-risk categories.

In ET, the IPSET-thrombosis system that includes age, previous thrombosis, cardiovascular risk factors, and *JAK2V617F* mutation, is the recommended prognostic system and it should be scored in all patients at diagnosis.

This implies that general risk factors for thrombosis, including smoking habit, diabetes mellitus, arterial hypertension, and hypercholesterolemia, should also be considered.

Risk stratification in MF

Robust prognostic modeling in primary myelofibrosis (PMF) started with the development of the International

Table 2 The IPSET criteria for evaluating the thrombotic risk of essential thrombocythemia (ET)

Risk factor	HR	Score
Age > 60 years	1.50	1
Cardiovascular risk factors	1.56	1
Previous thrombosis	1.93	2
<i>JAK2V617F</i>	2.04	2

Low risk implies a score = 0–1; intermediate risk, score = 2; and high risk, score ≥ 3

Prognostic Scoring System (IPSS) in 2009 [44]. The IWG-MRT subsequently developed a dynamic prognostic model (DIPSS) that can be applied at any time during the disease course [45]. IPSS- and DIPSS-independent risk factors for overall survival (OS) in PMF were subsequently identified: they included unfavorable karyotype [46, 47], the need for red cell transfusion [48, 49], and a platelet count < 100 × 10⁹/l [50]. Accordingly, DIPSS was modified into DIPSS-plus [51] (Table 3).

Recently, the prognostic relevance of driver and additional non-driver mutations was carefully investigated [3, 52]. Authors reported the longest OS in *CALR*⁺*ASXL1*⁻

Table 3 Risk stratification of primary myelofibrosis (PMF)

Prognostic model	Risk groups
IPSS	
Risk factors (weight)	Low risk: 0 point
Age > 65 years (1 point)	Intermediate-1 risk: 1 point
Constitutional symptoms (1 point)	Intermediate-2 risk: 2 points
Hemoglobin < 10 g/dl (1 point)	High risk: ≥ 3 points
WBC count > 25 × 10 ⁹ /l (1 point)	
Circulating blasts ≥ 1% (1 point)	
DIPSS	
Risk factors (weight)	Low risk: 0 point
Age > 65 years (1 point)	Intermediate-1 risk: 1–2 point
Constitutional symptoms (1 point)	Intermediate-2 risk: 3–4 points
Hemoglobin < 10 g/dl (2 points)	High risk: 5–6 points
WBC count > 25 × 10 ⁹ /l (1 point)	
Circulating blasts ≥ 1% (1 point)	
DIPSS-plus	
Risk factors (weight)	Low risk: 0 point
DIPSS score low (0 point)	Intermediate-1 risk: 1
DIPSS-int 1 (1 point)	Intermediate-2 risk: 2–3 points
DIPSS-int 2 (2 points)	High risk: 4–6 points
DIPSS high (3 points)	
RBC transfusion need (1 point)	
PLT count < 100 × 10 ⁹ /l (1 point)	
Unfavorable karyotype ^a (1 point)	

RBC red blood cell

^a (+ 8, -7/7q, i(17)q, -5/5q, 12p-, inv(3), 11q23 rearrangements)

patients (median, 10.4 years) and shortest in *CALR*⁻*ASXL1*⁺ patients (median, 2.3 years). *CALR*⁺*ASXL1*⁺ and *CALR*⁻*ASXL1*⁻ patients had similar survival and were grouped together in an intermediate-risk category (median survival, 5.8 years) [53, 54]. Furthermore, targeted studies with next generation sequencing have identified *ASXL1*, *SRSF2*, *CBL*, *KIT*, *RUNX1*, and *SH2B3* mutations as having an adverse effect on OS or leukemia-free survival (LFS), but these analyses did not include a multivariate assessment of all factors [55, 56].

It has been claimed that genetic information has the potential to integrate and improve the current prognostic systems [57]. However, the Panel cast doubts on its general clinical utility when used for making therapeutic decisions in MF, except in the setting of allogeneic stem cell transplantation (allo-HSCT) [58].

Recommendations

In MF, the IPSS, based on hematological and clinical variables, is the recommended prognostic system and should be scored in all patients at diagnosis.

There is increasing evidence that integration of IPSS with additional genetic information, i.e., cytogenetics and molecular parameters, allows a more detailed individualized prognostic classification.

For this reason, cytogenetic studies, classification of *CALR* mutations into type 1/like and type 2/like, and screening for non-driver additional mutations including at least *ASXL1* and *SRSF2*, has become current practice in research centers.

The Panel agreed that a complete genetic assessment should be encouraged in all patients for the prognostic assessment at diagnosis. However, the Panel also claimed that failure to perform a full genetic characterization at the time of diagnosis is acceptable in clinical practice.

DIPSS, based on hematological and clinical variables, or DIPSS-plus, based on hematological, clinical and cytogenetic variables, are the recommended systems for prognostic re-assessment during the disease course.

Molecular assessment during the course of the disease (at least *ASXL1* mutation) is recommended for therapeutic decisions in selected MF patients, such as to decide a transplant in those who have an intermediate-1 risk category according to the DIPSS/DIPSS-plus score.

Management of PV

First-line therapy

The ELN-2011 recommendations for first-line therapy of patients with PV include phlebotomy and low-dose acetyl salicylic acid. In high-risk patients either hydroxyurea or

recombinant interferon alpha (rIFN α) are the recommended first-line cytoreductive therapies at any age [24].

Recently, in the randomized phase 3 CYTO-PV trial, 365 PV patients were randomly assigned to receive either more intensive phlebotomy treatment (target hematocrit, <45%) or less intensive phlebotomy treatment (target hematocrit, 45 to 50%) [10]. After a median follow-up of 31 months, the primary endpoint, i.e., time until death from cardiovascular cause or major thrombotic events, was recorded in 2.7% patients in the low-hematocrit group and in 9.8% patients in the high-hematocrit group (HR in the high-hematocrit group, 3.91; 95% confidence interval (CI) = 1.45 to 10.53). According to GRADE methodology, we used CI as the primary tool for judging the precision of the results [59]. The low CI boundary of the HR closest to no effect (HR = 1) provided only 0.45-fold greater rate of cardiovascular events in patients with higher-hematocrit target. Thus, the quality of evidence on the precision of benefit was deemed to be low, and the core quality of evidence of the trial was deemed moderate. This indicates that additional research is likely to have important impact on our confidence in the estimate of effect. Notwithstanding, the Panel reasoned that, in a risk-benefit ratio perspective, this result reinforces the previous empirically determined recommendation to maintain the hematocrit lower than 45% [24].

In the dearth of experimental evidence on the effects of iron supplementation in PV patients under TP, the Panel reasoned by focusing on the balance between the discomfort of iron deficiency and the risk of increased hematocrit with iron supplementation.

Phase 2 trials have reported that pegylated interferon alpha-2a can induce durable hematological and molecular responses in patients with PV [60–62].

Recommendations

The Panel strongly recommended that all patients with PV should be managed with phlebotomy to maintain the hematocrit below 45%, together with daily low-dose acetyl salicylic acid.

In the induction phase, phlebotomy regimen should vary from 300 to 450 ml of blood withdrawn weekly or twice weekly until the hematocrit target is reached. Maintenance phase should have the same amount of blood volume removed per phlebotomy as in the induction phase, while the phlebotomy intervals should be determined by the levels of hematocrit.

Cytoreduction is strongly recommended in high-risk cases, i.e., patients with an age older than 60 years, or those with a previous thrombotic event.

The Panel convened that poor tolerance to phlebotomy is an additional indication to cytoreductive therapy.

In the case of documented severe tissue iron deficiency associated with detrimental symptoms (pica, mouth paresthesia, esophagitis, rest-less legs), iron supplementation is indicated. Occurrence of undesired hematocrit worsening due to iron therapy, indicates the need of cytoreduction.

Symptomatic or progressive splenomegaly, severe disease-related symptoms, platelet counts greater than $1500 \times 10^9/l$ or leukocyte count higher than $15 \times 10^9/l$, are further indications to start cytoreductive therapy.

Either hydroxyurea or rIFN α is the first-line cytoreductive therapy at any age. However, the Panel agreed it is wise to adopt a cautionary principle and carefully consider the use of hydroxyurea in young patients.

All patients should be managed aggressively for their cardiovascular risk factors.

Therapy change and second-line therapy

Conventional second-line therapy in PV includes hydroxyurea as a substitute for rIFN α and rIFN α as a substitute of hydroxyurea [24]. Recently, one phase 2 and two phase 3 randomized trials (RESPONSE trials) have reported efficacy of the JAK1/JAK2 inhibitor ruxolitinib in individuals with PV who were resistant or intolerant to hydroxyurea [7, 8, 63]. Following these results, ruxolitinib received approval in 2014 from the US Food and Drug Administration (FDA) and in 2015 from the European Medicine Agency (EMA) for the treatment of patients with PV who have an inadequate response to or cannot tolerate hydroxyurea.

We downgraded the overall quality of evidence for the critical outcomes of RESPONSE trials because we found risk of performance bias, and indirectness in the comparator. On the contrary, we rated the hematocrit response and primary response as having a high level of evidence. Overall, the judgment of the quality of evidence was deemed to be moderate (Appendix 1 for critical appraisal of evidence of the RESPONSE trials).

In a randomized, double-blind, double-dummy, phase 3b trial (RELIEF), PV patients who were well-controlled with a stable dose of hydroxyurea but who reported PV-related symptoms, were randomized to ruxolitinib or hydroxyurea to measure the frequency of patients reaching $\geq 50\%$ improvement from baseline in total symptom score [11]. Ruxolitinib was associated with a non-significant trend towards improved PV-related symptoms vs. hydroxyurea.

Recommendations

PV patients classified as low-risk at diagnosis require the introduction of cytoreductive therapy once they reach the age of 60, or once they develop major thrombotic complications.

The Panel agreed that both rIFN α and ruxolitinib are appropriate second-line drug therapies for PV patients who are intolerant or have inadequate response to hydroxyurea. In this setting, the recommendation of use of ruxolitinib was judged by the Panel as strong, even though the confidence in the outcome measures was moderate. In the absence of a direct comparison of the two agents, the choice should be based on the patient's age and drug availability. rIFN α should be preferred in young patients in need of long-term treatment.

Intermittent doses of busulphan may be considered in very elderly patients.

Management of ET

First-line therapy

Current guidelines favor hydroxyurea as first-line therapy of ET patients in need of cytoreduction [21]. The recommendation is based on the superiority of hydroxyurea combined with acetyl salicylic acid vs. anagrelide combined with acetyl salicylic acid in the UK-PT1 trial in a cohort of high-risk patients diagnosed on the basis of Polycythemia Vera Study Group criteria [63, 64]. More recent phase 2 trials have reported that pegylated interferon alpha-2a can induce durable hematological and molecular responses in patients with ET [61, 65]. Moreover, in patients with ET strictly diagnosed according to the WHO system and thus excluding early-PMF cases, a phase 3 non-inferiority trial (ANAHYDRET) documented that anagrelide proved non-inferior when compared to hydroxyurea in the prevention of thrombotic complications [9]. The critical appraisal of the trial recognized non-blinded therapy assignment, and indirectness of the population, since patients enrolled in the trial were defined at high-risk according to non-standard criteria (platelet count $\geq 1000 \times 10^9/l$ and increase of platelet count $\geq 300 \times 10^9/l$ within 3 months). However, the precision of the results was considered high. Finally, the evidence that anagrelide is comparable with hydroxyurea was judged moderate.

The 2011-ELN guidelines indicate that all patients with ET should be managed with low-dose acetyl salicylic acid if microvascular disturbances are present [24]. New observations have delineated the appropriate use of acetyl salicylic acid as a prevention of vascular events in ET patients. In a retrospective study, the rate of major thrombosis was significantly lower in the subgroup of patients ≥ 60 years when used in association with cytoreductive therapy, and compensates the increased risk of bleeding [66]. In a retrospective analysis, antiplatelet therapy in CALR-mutated patients did not affect the risk of thrombosis but was associated with higher incidence of bleeding, while in JAK2V617F-mutated patients, low-dose acetyl salicylic

acid was associated with a reduced incidence of venous thrombosis with no effect on the risk of bleeding [67]. Recently, Barbui and co-workers [68] delineated a category of “very-low risk” ET, i.e., patients without cardiovascular risk factors, age \leq 60 years and *JAK2*-unmutated.

Recommendations

In high-risk ET, three drugs have provided evidence of being able to produce hematological response, i.e., hydroxyurea, rINF α , and anagrelide.

The Panel agreed on recommending hydroxyurea and rINF α as first-line therapy agents.

Even though the majority of the experts indicated anagrelide as an appropriate choice for first-line therapy in ET, the Panel did not reach a consensus on recommending the agent in this setting, arguing that the evidence of non-inferiority with hydroxyurea was of insufficient quality, and the risk-benefit ratio unfavorable.

The use of cytoreductive drugs in otherwise low- or intermediate-risk patients carrying well-controlled cardiovascular risk factors is not generally recommended.

From the results of retrospective observational studies, all patients with IPSET-thrombosis high-risk disease should receive low-dose acetyl salicylic acid. Patients with low- or intermediate-risk ET should receive low-dose acetyl salicylic acid when age is \geq 60 years, or when uncontrolled cardiovascular risk factors or *JAK2*V617F mutation are present.

Therapy change and second-line therapy

The 2011-ELN guidelines claimed that in case of refractoriness or intolerance to first-line therapy with hydroxyurea [69], non leukemogenic drugs such as anagrelide or rINF α should be recommended [24].

Recommendations

Patients with ET should start a cytoreductive treatment as soon as they shift to the high-risk category by reaching the age of 60 years or by the occurrence of a major thrombotic or hemorrhagic event. Increasing platelet count above $1500 \times 10^9/l$ is an additional indication to cytoreductive therapy.

Cytoreduction may be also required for progressive myeloproliferation (e.g., increasing splenomegaly) or uncontrolled ET-related systemic symptoms.

Anagrelide or rINF α are the recommended second-line therapy of ET after hydroxyurea.

Management of MF

Since the recommendations on the management of MF published in 2011 [24], a number of new therapeutic agents

have been proposed and some of them have been experimentally tested [5, 6, 12–14, 70–74]. Among these, ruxolitinib has been approved both by FDA and EMA. The Panel was thus questioned whether the new targeted therapy proved to favorably modify the natural history of the disease, or significantly prolong survival. By applying GRADE methodology, we derived pooled estimates across the two randomized controlled trials (COMFORT trials) and we calculated pooled HRs and 95% CIs using random-effect models for all-cause mortality [75]. We documented a significant benefit associated with ruxolitinib treatment at late follow-up. Our meta-analysis, however, failed to meet the “optimal information size” criteria: thus, we downgraded the confidence in the estimates of survival advantage of ruxolitinib for imprecision (too few events) [59]. A similar analysis was published by the Cochrane collaboration [76], and later on by Cervantes and co-workers [77]. These results suggested that the evidence from the two COMFORT trials on improved survival in MF patients treated with ruxolitinib is moderate to poor. Therefore, the Panel agreed that moving to a disease-oriented therapeutic strategy with ruxolitinib for MF was not justified, and the revised recommendations were issued across the different phenotypes/problems of the disease.

Asymptomatic patients with low- or intermediate-1 risk disease

There is no evidence to support the value of disease-modifying therapy in patients with IPSS/DIPSS/DIPSS-plus low or intermediate-1 risk disease. However, some of these patients might require palliative therapy for anemia, splenomegaly, or constitutional symptoms, as recommended in the previous guidelines [24].

Recommendations

We recommend observation alone for IPSS/DIPSS/DIPSS-plus low- or intermediate-1 MF risk patients who lack significant symptoms, and who do not display significant anemia (hemoglobin $<$ 10 g/dl), splenomegaly (palpable spleen size $>$ 10 cm), leukocytosis (leukocyte count $>$ $25 \times 10^9/l$), or marked thrombocytosis (platelet count $>$ $1000 \times 10^9/l$).

If cytoreductive treatment for the reduction of leukocytosis or thrombocytosis is indicated, the first-line drug of choice is hydroxyurea.

Treatment of MF-associated anemia

According to the 2011-ELN recommendations, treatment of MF-associated anemia includes androgens, prednisone, epoetins, danazol, thalidomide, or lenalidomide [24].

Recently, pomalidomide, a thalidomide analog, has been tested in a number of phase 2 and one phase 3 trial [12,78–84]. In a phase 3 randomized trial, 252 transfusion-dependent subjects were randomly assigned (2:1) to pomalidomide or placebo [12]. Trialists and subjects were blinded to treatment allocation. Pomalidomide and placebo achieved similar RBC-transfusion-independence response rates (16%).

At the end, no evidence-based recommendations for the best strategy of treating MF-associated anemia can be issued. The Panel discussed the limitations of conventional drug therapy on the basis of case reports [85, 86], and clinical experience.

Recommendations

The choice of a specific drug for MF-associated anemia should be based on overall toxicity profile and its expected risk in the individual patient.

Androgen preparations should be avoided in patients with prostate disease or concomitant liver disease.

Thalidomide and its analogs should be avoided in patients with documented peripheral neuropathy grade 2, and its use should be strictly monitored in those with grade 1 neuropathy or who are at risk for such complication such as diabetics.

In patients with transfusion dependency, a low rate of response to epoetins is expected, therefore the risk/benefit of a therapeutic use with epoetins is questionable.

Lenalidomide use is justified in cases with the presence of del(5q31).

There is currently not enough evidence to recommend combination therapy for MF-associated anemia, other than the addition of a short course of prednisone therapy to treatment with thalidomide.

Treatment of MF-associated splenomegaly

Conventionally, first-line therapy for MF-associated splenomegaly is hydroxyurea [24]. Since the major benefits of ruxolitinib therapy in MF were the reduction in splenomegaly and amelioration of MF-related symptoms [6, 87, 88], these results challenged the primary use of hydroxyurea in this indication. A GRADE analysis of COMFORT trials documented that the overall quality of evidence supporting the benefit of ruxolitinib in reducing splenomegaly in patients with intermediate-2 or high-risk disease was high, due to the randomized design, a clear demonstration of a dose–response relationship between ruxolitinib dose and spleen volume reduction. However, the quality of evidence was reduced due to unblindness and serious indirectness of the comparator [75]. For these limitations, the Panel argued for a limited use of the drug in the therapy of splenomegaly

in MF. For low-intermediate-1 patients, the Panel reasoned that the extent of the benefit identifying the threshold for treatment is a function of the prognosis of patients and the toxicity/inconveniences of therapy. Thus, patients with low-risk disease were judged to have no indication of use of ruxolitinib, while intermediate-1 disease are candidates to therapy only when symptoms in need of treatment are present.

Recommendations

Ruxolitinib is recommended as first-line approach for MF-associated splenomegaly in patients with intermediate-2 or high-risk disease.

In patients with intermediate-1 risk disease and highly symptomatic splenomegaly, i.e., with the presence of local symptoms, or impairment of food intake, first-line therapy is ruxolitinib.

In other patients with intermediate-1 risk disease, and in those with low-risk disease in need of therapy for MF-associated splenomegaly, hydroxyurea is recommended as first-line therapy.

Ruxolitinib is also recommended for reducing splenomegaly in patients with splenomegaly not responding or intolerant to hydroxyurea.

Allogeneic stem cell transplantation

Current guidelines candidate to allo-SCT those patients with intermediate-2 or high-risk disease [24]. The emergence of new prognostic predictors, such as MF-associated mutations [3, 4, 33, 34], adverse cytogenetics [46, 47], high need of transfusions [48, 49], or high number of circulating blasts [89], drew the Panel to discuss the value of new detrimental markers in widening the transplant indication, particularly in those patients who have an intermediate-1 risk disease. In a recent study on the impact of molecular genetics on the outcome after allo-SCT [90], patients with *ASXL1* mutations had a lower transplant success rate than all comers. The Panel judged this result as preliminary due to the small number of patients analyzed. Moreover, the Panel argued that there is no evidence that allo-SCT cannot overcome the risks associated with the *ASXL1* mutational pattern. Thus, the Panel favored considering allo-SCT in these patients.

Recommendations

We recommend considering allogeneic stem cell transplant for all transplant-eligible patients with IPSS/DIPSS/DIPSS-plus high or intermediate-2 risk.

The Panel also recommended consideration of allogeneic stem cell transplantation for transplant-eligible patients with

IPSS/DIPSS/DIPSS-Plus intermediate-1 risk score who present with either refractory, transfusion-dependent anemia, a percentage of blasts in peripheral blood >2% in at least two repeated manual measurements, adverse cytogenetics, or high-risk mutations,

In this situation, the transplant procedure should be performed in a controlled setting (registries, clinical trial).

Splenectomy

In patients with symptomatic and progressive splenomegaly, which proved to be resistant to medical therapy, in particular to hydroxyurea, ruxolitinib, or experimental therapies, splenectomy necessarily remains an option [91]. Recent reports on splenectomy in MF [90, 92] have confirmed the seminal results derived from the largest series of cases [93, 94].

Recommendations

Splenectomy remains a viable palliative treatment option for drug-refractory symptomatic splenomegaly and may be considered when drug-induced anemia hampers the effective use of hydroxyurea or ruxolitinib.

Discussion

Consensus techniques and careful revision of new evidences were used to update recommendations for the management of Ph-neg MPNs published in 2011 [24]. The GRADE method, allowed the explicit use of factors that can increase or decrease the quality of the evidence. Although most recommendations remained unchanged, some key differences emerged. Changes included the criteria for the diagnosis, refinement of risk stratification, and the recommendation for therapy of PV and MF due to the emergence of the new anti-JAK1/JAK2 drug, ruxolitinib.

With the rapid development of scientific knowledge, new evidence is emerged between the time we collected the relevant literature for this project and the time we wrote the manuscript. Indeed, a clinical-molecular prognostic model (the MYSEC-PM) to predict OS in patients with post-PV and post-ET MF has been proposed that could improve the prognostic classification of these patients [95]. New predictive factors for anemia response to erythropoiesis-stimulating agents in MF have been analyzed [96]. A randomized controlled phase 3 trial compared a new PEG-proline-IFN-alpha-2b to hydroxyurea (PROUD-PV trial) resulting in a non-inferior efficacy and higher tolerability of PEG-proline-IFN [97, 98]. Finally, a randomized phase 2 trial comparing ruxolitinib to best available therapy in patients with ET resistant or intolerant to hydroxyurea

(MAJIC trial) documented that ruxolitinib is not superior to current second-line treatments for ET [99].

As such, these recommendations, though currently accurate, will likely need to be updated in the future.

Acknowledgements AMV, FP, and G Ba were supported by a grant from Associazione Italiana per la Ricerca sul Cancro (AIRC; Milan, Italy), Special Program Molecular Clinical Oncology 5 × 1000 to AIRC-Gruppo Italiano Malattie Mieloproliferative (AGIMM) project #1005.

Author contributions TB, AT, and GBa developed the study design. GBa did the critical appraisal of evidence of the randomized trials. TB and AT wrote the first draft of the recommendations. All the authors participated to the consensus meetings. GBa wrote the manuscript. All authors critically revised, reviewed, and approved the final version of this study.

Compliance with ethical standards

Conflict of interest TB received advisory board fee from Novartis; FC received advisory board fee from Novartis, honoraria from Novartis, AOP Orphan Disease and Shire, and speaker bureau fee from Novartis, AOP and Shire; AMV received advisory board fee from Novartis, and Speaker bureau fee from Novartis and Shire; MM received speaker bureau fee from Gilead and Amgen; SK received research funding from Novartis, Bristol-Myers Squibb, and Janssen, honoraria and advisory board fees from Novartis, Incyte/Ariad, Pfizer, Bristol-Myers Squibb, Janssen, AOP, CTI, and travel support from Shire; AR received advisory board fee and honoraria from Novartis; MG received honoraria from Shire, Novartis, Baxalta, AOP Orphan Disease, Gilead and Janssen; JJK received advisory board fee and funding from Novartis and AOP Orphan; AB received honoraria from Novartis, Therakos, Sanofi and Adienne, and advisory board fee from Novartis; MFMM received honoraria and consulting fee from Novartis and Gilead; SV received support for research from: Incyte Corporation, Roche, Astra Zeneca, Lilly Oncology, NS Pharma, Bristol Mayers Squibb, Celgene, Gilead, Seattle Genetics, Promedior, CTI BioPharma Corp., Galena BioPharma, Pfizer, Genentech, Blueprint Medicines Corp.; RM received consulting fee from Novartis, AOP, Shire, and research funding from Incyte, Gilead, Pharmessential, Celgene and Promedior; R Ho received research funding from Incyte and Janssen; FP served on advisory boards for Sanofi, Gilead, Janssen Pharmaceutical, Celgene, Novartis, Bristol-Myers Squibb and Roche; GB received advisory board fee from Novartis. GF and G Bi have no conflict of interest.

References

1. Klampfl T, Gisslinger H, Harutyunyan AS, Nivarthi H, Rumi E, Milosevic JD, et al. Somatic mutations of calreticulin in myeloproliferative neoplasms. *N Engl J Med*. 2013;369:2379–90.
2. Nangalia J, Massie CE, Baxter EJ, Nice FL, Gundem G, Wedge DC, et al. Somatic CALR mutations in myeloproliferative neoplasms with nonmutated JAK2. *N Engl J Med*. 2013;369:2391–405.
3. Guglielmelli P, Lasho TL, Rotunno G, Score J, Mannarelli C, Pancrazzi A, et al. The number of prognostically detrimental mutations and prognosis in primary myelofibrosis: an international study of 797 patients. *Leukemia*. 2014;28:1804–10.
4. Tefferi A. Novel mutations and their functional and clinical relevance in myeloproliferative neoplasms: JAK2, MPL, TET2, ASXL1, CBL, IDH and IKZF1. *Leukemia*. 2010;24:1128–38.

5. Verstovsek S, Mesa RA, Gotlib J, Levy RS, Gupta V, DiPersio JF, et al. A double-blind, placebo-controlled trial of ruxolitinib for myelofibrosis. *N Engl J Med*. 2012;366:799–807.
6. Harrison C, Kiladjan JJ, Al-Ali HK, Gisslinger H, Waltzman R, Stalbovska V, et al. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis. *N Engl J Med*. 2012;366:787–98.
7. Vannucchi AM, Kiladjan JJ, Grieshammer M, Masszi T, Durrant S, Passamonti F, et al. Ruxolitinib versus standard therapy for the treatment of polycythemia vera. *N Engl J Med*. 2015;372:426–35.
8. Passamonti F, Grieshammer M, Palandri F, Egyed M, Benevolo G, Devos T, et al. Ruxolitinib for the treatment of inadequately controlled polycythemia vera without splenomegaly (RESPONSE-2): a randomised, open-label, phase 3b study. *Lancet Oncol*. 2017;18:88–99.
9. Gisslinger H, Gotic M, Holowiecki J, Penka M, Thiele J, Kvasnicka HM, et al. Anagrelide compared with hydroxyurea in WHO-classified essential thrombocythemia: the ANAHYDRET Study, a randomized controlled trial. *Blood*. 2013;121:1720–28.
10. Marchioli R, Finazzi G, Specchia G, Cacciola R, Cavazzina R, Cilloni D, et al. Cardiovascular events and intensity of treatment in polycythemia vera. *N Engl J Med*. 2013;368:22–33.
11. Mesa R, Vannucchi AM, Yacoub A, Zachee P, Garg M, Lyons R, et al. The efficacy and safety of continued hydroxycarbamide therapy versus switching to ruxolitinib in patients with polycythemia vera: a randomized, double-blind, double-dummy, symptom study (RELIEF). *Br J Haematol*. 2017;176:76–85.
12. Tefferi A, Al-Ali HK, Barosi G, Devos T, Gisslinger H, Jiang Q, et al. A randomized study of pomalidomide vs placebo in persons with myeloproliferative neoplasm-associated myelofibrosis and RBC-transfusion dependence. *Leukemia*. 2017;31:1252.
13. Pardanani A, Harrison C, Cortes JE, Cervantes F, Mesa RA, Milligan D, et al. Safety and efficacy of fedratinib in patients with primary or secondary myelofibrosis: a randomized clinical trial. *JAMA Oncol*. 2015;1:643–51.
14. Mesa RA, Kiladjan JJ, Catalano JV, Devos T, Egyed M, Hellmann A, et al. SIMPLIFY-1: A phase iii randomized trial of momelotinib versus ruxolitinib in janus kinase inhibitor-naïve patients with myelofibrosis. *J Clin Oncol*. 2017;35:3844–50 [epub ahead of print].
15. Mesa RA. NCCN debuts new guidelines for myeloproliferative neoplasms. *J Natl Compr Canc Netw*. 2017;15:720–22.
16. Choi CW, Bang SM, Jang S, Jung CW, Kim HJ, Kim HY, Choi CW, Bang SM, Jang S, et al. Guidelines for the management of myeloproliferative neoplasms. *Korean J Intern Med*. 2015;30:771–88.
17. Reilly JT, McMullin MF, Beer PA, Butt N, Conneally E, Duncombe AS, et al. Use of JAK inhibitors in the management of myelofibrosis: a revision of the British Committee for Standards in Haematology Guidelines for Investigation and Management of Myelofibrosis 2012. *Br J Haematol*. 2014;167:418–20.
18. Vannucchi AM, Barbui T, Cervantes F, Harrison C, Kiladjan JJ, Kröger N, et al. ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2015;26:v85–99.
19. Marchetti M, Barosi G, Cervantes F, Birgegård G, Grieshammer M, Harrison C, et al. Which patients with myelofibrosis should receive ruxolitinib therapy? ELN-SIE evidence-based recommendations. *Leukemia*. 2017;31:882–88.
20. Sliwa T, Beham-Schmid C, Burgstaller S, Buxhofer-Ausch V, Gastl G, Geissler K, et al. Austrian recommendations for the management of primary myelofibrosis, post-polycythemia vera myelofibrosis and post-essential thrombocythemia myelofibrosis: an expert statement. *Wien Klin Wochenschr*. 2017;129:293–02.
21. Ho PJ, Marlton P, Tam C, Stevenson W, Ritchie D, Bird R, et al. Practical management of myelofibrosis with ruxolitinib. *Intern Med J*. 2015;45:1221–30.
22. Guglielmelli P, Pietra D, Pane F, Pancrazzi A, Cazzola M, Vannucchi AM, et al. Recommendations for molecular testing in classical Ph1-neg myeloproliferative disorders- A consensus project of the Italian Society of Hematology. *Leuk Res*. 2017;58:63–72.
23. Kröger NM, Deeg JH, Olavarria E, Niederwieser D, Bacigalupo A, Barbui T, et al. Indication and management of allogeneic stem cell transplantation in primary myelofibrosis: a consensus process by an EBMT/ELN international working group. *Leukemia*. 2015;29:2126–33.
24. Barbui T, Barosi G, Birgegård G, Cervantes F, Finazzi G, Grieshammer M, et al. European LeukemiaNet. Philadelphia-negative classical myeloproliferative neoplasms: critical concepts and management recommendations from European LeukemiaNet. *J Clin Oncol*. 2011;29:761–70.
25. Atkins D, Best D, Briss PA, Eccles M, Falck-Ytter Y, Flottorp S, et al. Grading quality of evidence and strength of recommendations. *BMJ*. 2004;328:1490.
26. Higgins JPT, Green S, editors. *Cochrane handbook for systematic reviews of interventions version 5.1.0 [updated March 2011]*. The Cochrane Collaboration, 2011. Available from www.cochrane-handbook.org.
27. William PL, Webb C. The Delphi technique: a methodological discussion. *J Adv Nurs*. 1994;19:180–6.
28. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood*. 2016;127:2391–405.
29. Kvasnicka HM, Orazi A, Thiele J, Barosi G, Bueso-Ramos CE, Vannucchi AM, et al. European LeukemiaNet study on the reproducibility of bone marrow features in masked polycythemia vera and differentiation from essential thrombocythemia. *Am J Hematol*. 2017;92:1062–67 [epub ahead of print].
30. Tefferi A, Pardanani A. Genetics: CALR mutations and a new diagnostic algorithm for MPN. *Nat Rev Clin Oncol*. 2014;11:125–26.
31. Kvasnicka HM, Thiele J. Prodromal myeloproliferative neoplasms: the 2008 WHO classification. *Am J Hematol*. 2010;85:62–9.
32. Barosi G, Mesa RA, Thiele J, Cervantes F, Campbell PJ, Verstovsek S, et al. International Working Group for Myelofibrosis Research and Treatment (IWG-MRT). Proposed criteria for the diagnosis of post-polycythemia vera and post-essential thrombocythemia myelofibrosis: a consensus statement from the International Working Group for Myelofibrosis Research and Treatment. *Leukemia*. 2008;22:437–38.
33. Tefferi A, Thiele J, Vannucchi AM, Barbui T. An overview on CALR and CSF3R mutations and a proposal for revision of WHO diagnostic criteria for myeloproliferative neoplasms. *Leukemia*. 2014;28:1407–13.
34. Tefferi A, Finke CM, Lasho TL, Wassie EA, Knudson R, Ketterling RP, et al. U2AF1 mutations in primary myelofibrosis are strongly associated with anemia and thrombocytopenia despite clustering with JAK2V617F and normal karyotype. *Leukemia*. 2014;28:431–33.
35. Tefferi A, Barbui T. Essential thrombocythemia and polycythemia vera: Focus on clinical practice. *Mayo Clin Proc*. 2015;90:1283–93.
36. Barbui T, Carobbio A, Rumi E, Finazzi G, Gisslinger H, Rodeghiero F, et al. In contemporary patients with polycythemia vera, rates of thrombosis and risk factors delineate a new clinical epidemiology. *Blood*. 2014;124:3021–23.
37. Tefferi A, Rumi E, Finazzi G, Gisslinger H, Vannucchi AM, Rodeghiero F, et al. Survival and prognosis among 1545 patients with contemporary polycythemia vera: an international study. *Leukemia*. 2013;27:1874–81.

38. Barbui T, Carobbio A, Rambaldi A, Finazzi G, Barbui T, Carobbio A, et al. Perspectives on thrombosis in essential thrombocythemia and polycythemia vera: is leukocytosis a causative factor? *Blood*. 2009;114:759–63.
39. Barbui T, Masciulli A, Marfisi MR, Tognoni G, Finazzi G, Rambaldi A, et al. White blood cell counts and thrombosis in polycythemia vera: a subanalysis of the CYTO-PV study. *Blood*. 2015;126:560–61.
40. Barbui T, Finazzi G, Carobbio A, Thiele J, Passamonti F, Rumi E, et al. Development and validation of an International Prognostic Score of thrombosis in World Health Organization-essential thrombocythemia (IPSET-thrombosis). *Blood*. 2012;120:5128–33.
41. Barbui T, Vannucchi AM, Buxhofer-Ausch V, De Stefano V, Betti S, Rambaldi A, et al. Practice-relevant revision of IPSET-thrombosis based on 1019 patients with WHO-defined essential thrombocythemia. *Blood Cancer J*. 2015;5:e369.
42. Haider M, Gangat N, Lasho T, Abou Hussein AK, Elala YC, Hanson C, et al. Validation of the revised International Prognostic Score of Thrombosis for Essential Thrombocythemia (IPSET-thrombosis) in 585 Mayo Clinic patients. *Am J Hematol*. 2016;91:390–94.
43. Finazzi G, Carobbio A, Guglielmelli P, Cavalloni C, Salmoiraghi S, Vannucchi AM. Calreticulin mutation does not modify the IPSET score for predicting the risk of thrombosis among 1150 patients with essential thrombocythemia. *Blood*. 2014;124:2611–12.
44. Cervantes F, Dupriez B, Pereira A, Passamonti F, Reilly JT, Morra E, et al. New prognostic scoring system for primary myelofibrosis based on a study of the International Working Group for Myelofibrosis Research and Treatment. *Blood*. 2009;113:2895–901.
45. Passamonti F, Cervantes F, Vannucchi AM, Morra E, Rumi E, Cazzola M, et al. A dynamic prognostic model to predict survival in primary myelofibrosis: a study by the IWG-MRT (International Working Group for Myeloproliferative Neoplasms Research and Treatment). *Blood*. 2010;115:1703–08.
46. Hussein K, Pardanani AD, Van Dyke DL, Hanson CA, Tefferi A. International Prognostic Scoring System-independent cytogenetic risk categorization in primary myelofibrosis. *Blood*. 2010;115:496–99.
47. Caramazza D, Begna KH, Gangat N, Vaidya R, Siragusa S, Van Dyke DL, et al. Refined cytogenetic-risk categorization for overall and leukemia-free survival in primary myelofibrosis: a single center study of 433 patients. *Leukemia*. 2011;25:82–8.
48. Tefferi A, Siragusa S, Hussein K, Schwager SM, Hanson CA, Pardanani A, et al. Transfusion-dependency at presentation and its acquisition in the first year of diagnosis are both equally detrimental for survival in primary myelofibrosis—prognostic relevance is independent of IPSS or karyotype. *Am J Hematol*. 2010;85:14–7.
49. Elena C, Passamonti F, Rumi E, Malcovati L, Arcaini L, Boveri E, et al. Red blood cell transfusion-dependency implies a poor survival in primary myelofibrosis irrespective of IPSS and DIPSS. *Haematologica*. 2011;96:167–70.
50. Patnaik MM, Caramazza D, Gangat N, Hanson CA, Pardanani A, Tefferi A. Age and platelet count are IPSS-independent prognostic factors in young patients with primary myelofibrosis and complement IPSS in predicting very long or very short survival. *Eur J Haematol*. 2010;84:105–8.
51. Gangat N, Caramazza D, Vaidya R, George G, Begna K, Schwager S, et al. DIPSS plus: a refined Dynamic International Prognostic Scoring System for primary myelofibrosis that incorporates prognostic information from karyotype, platelet count, and transfusion status. *J Clin Oncol*. 2011;29:392–7.
52. Tefferi A, Guglielmelli P, Lasho TL, Rotunno G, Finke C, Mannarelli C, et al. CALR and ASXL1 mutations-based molecular prognostication in primary myelofibrosis: an international study of 570 patients. *Leukemia*. 2014;28:1494–1500.
53. Tefferi A, Lasho TL, Tischer A, Wassie EA, Finke CM, Belachew AA, et al. The prognostic advantage of calreticulin mutations in myelofibrosis might be confined to type 1 or type 1-like CALR variants. *Blood*. 2014;124:2465–66.
54. Guglielmelli P, Rotunno G, Fanelli T, Pacilli A, Brogi G, Calabresi L, et al. Validation of the differential prognostic impact of type 1/type 1-like versus type 2/type 2-like CALR mutations in myelofibrosis. *Blood Cancer J*. 2015;5:e360.
55. Tefferi A, Jimma T, Sulai NH, Lasho TL, Finke CM, Knudson RA, et al. IDH mutations in primary myelofibrosis predict leukemic transformation and shortened survival: clinical evidence for leukemogenic collaboration with JAK2V617F. *Leukemia*. 2012;26:475–80.
56. Lasho TL, Jimma T, Finke CM, Patnaik M, Hanson CA, Ketterling RP, et al. SRSF2 mutations in primary myelofibrosis: significant clustering with IDH mutations and independent association with inferior overall and leukemia-free survival. *Blood*. 2012;120:4168–71.
57. Tefferi A, Vannucchi AM. Genetic risk assessment in myeloproliferative neoplasms. *Mayo Clin Proc*. 2017;92:1283–90.
58. Guglielmelli P, Lasho TL, Rotunno G, Mudireddy M, Mannarelli C, Nicolosi M, et al. MIPSS70: Mutation-Enhanced International Prognostic Score System for transplantation-age patients with primary myelofibrosis. *J Clin Oncol*. 2018;36:310–8.
59. Guyatt GH, Oxman AD, Kunz R, Brozek J, Alonso-Coello P, Rind D, et al. GRADE guidelines 6. Rating the quality of evidence—imprecision. *J Clin Epidemiol*. 2011;64:1283–93.
60. Kiladjian JJ, Cassinat B, Chevret S, Turlure P, Cambier N, Roussel M, et al. Pegylated interferon- α -2a induces complete hematologic and molecular responses with low toxicity in polycythemia vera. *Blood*. 2008;112:3065–72.
61. Quintás-Cardama A, Abdel-Wahab O, Manshouri T, Kilpivaara O, Cortes J, Roupie AL, et al. Molecular analysis of patients with polycythemia vera or essential thrombocythemia receiving pegylated interferon α -2a. *Blood*. 2013;122:893–901.
62. Gowin K, Jain T, Kosiorek H, Tibes R, Camoriano J, Palmer J, et al. Pegylated interferon alpha-2a is clinically effective and tolerable in myeloproliferative neoplasm patients treated off clinical trial. *Leuk Res*. 2017;54:73–7.
63. Verstovsek S, Passamonti F, Rambaldi A, Barosi G, Rosen PJ, Rumi E, et al. A phase 2 study of ruxolitinib, an oral JAK1 and JAK2 inhibitor, in patients with advanced polycythemia vera who are refractory or intolerant to hydroxyurea. *Cancer*. 2014;120:513–20.
64. Harrison CN, Campbell PJ, Buck G, Wheatley K, East CL, Bareford D, et al. Primary thrombocythemia 1 study. Hydroxyurea compared with anagrelide in high-risk essential thrombocythemia. *N Engl J Med*. 2005;353:33–45.
65. Cassinat B, Verger E, Kiladjian JJ. Interferon alfa therapy in CALR-mutated essential thrombocythemia. *N Engl J Med*. 2014;371:188–9.
66. Alvarez-Larrán A, Pereira A, Arellano-Rodrigo E, Hernández-Boluda JC, Cervantes F, Besses C. Cytoreduction plus low-dose aspirin versus cytoreduction alone as primary prophylaxis of thrombosis in patients with high-risk essential thrombocythemia: an observational study. *Br J Haematol*. 2013;161:865–71.
67. Alvarez-Larrán A, Pereira A, Guglielmelli P, Hernández-Boluda JC, Arellano-Rodrigo E, Ferrer-Marín F, et al. Antiplatelet therapy versus observation in low-risk essential thrombocythemia with a CALR mutation. *Haematologica*. 2016;101:926–31.

68. Barbui T, Vannucchi AM, Buxhofer-Ausch V, De Stefano V, Betti S, Rambaldi A, et al. Practice-relevant revision of IPSET-thrombosis based on 1019 patients with WHO-defined essential thrombocythemia. *Blood. Cancer J.* 2015;5:e369.
69. Barosi G, Besses C, Birgegard G, Briere J, Cervantes F, Finazzi G, et al. A unified definition of clinical resistance/intolerance to hydroxyurea in essential thrombocythemia: results of a consensus process by an international working group. *Leukemia.* 2007;21:277–80.
70. Komrokji RS, Seymour JF, Roberts AW, Wadleigh M, To LB, Scherber R, et al. Results of a phase 2 study of pacritinib (SB1518), a JAK2/JAK2 (V617F) inhibitor, in patients with myelofibrosis. *Blood.* 2015;125:2649–56.
71. Pardanani A, Laborde RR, Lasho TL, Finke C, Begna K, Al-Kali A, et al. Safety and efficacy of CYT387, a JAK1 and JAK2 inhibitor, in myelofibrosis. *Leukemia.* 2013;27:1322–27.
72. Mascarenhas JO, Talpaz M, Gupta V, Foltz LM, Savona MR, Paquette R, et al. Primary analysis of a phase II open-label trial of INCB039110, a selective JAK1 inhibitor, in patients with myelofibrosis. *Haematologica.* 2017;102:327–35.
73. Tefferi A, Lasho TL, Begna KH, Patnaik MM, Zblewski DL, Finke CM, et al. A pilot study of the telomerase inhibitor imetelstat for myelofibrosis. *N Engl J Med.* 2015;373:908–19.
74. Duffield JS, Lupher ML Jr. PRM-151 (recombinant human serum amyloid P/pentaxin 2) for the treatment of fibrosis. *Drug News Perspect.* 2010;23:305–15.
75. Barosi G, Rosti V, Gale RP. Critical appraisal of the role of ruxolitinib in myeloproliferative neoplasm-associated myelofibrosis. *Onco Targets Ther.* 2015;8:1091–102.
76. Martí-Carvajal AJ, Anand V, Solà I. Janus kinase-1 and Janus kinase-2 inhibitors for treating myelofibrosis. *Cochrane Database Syst Rev.* 2015;4:CD010298.
77. Cervantes F, Pereira A. Does ruxolitinib prolong the survival of patients with myelofibrosis? *Blood.* 2017;129:832–37.
78. Ross DM. Responses to pomalidomide and placebo in myelofibrosis-related anaemia. *Leukemia.* 2017;31:532–33.
79. Schlenk RF, Stegelmann F, Reiter A, Jost E, Gattermann N, Hebart H, et al. Pomalidomide in myeloproliferative neoplasm-associated myelofibrosis. *Leukemia.* 2017;31:889–95.
80. Daver N, Shastri A, Kadia T, Newberry K, Pemmaraju N, Jabbour E, et al. Phase II study of pomalidomide in combination with prednisone in patients with myelofibrosis and significant anemia. *Leuk Res.* 2014;38:1126–29.
81. Daver N, Shastri A, Kadia T, Newberry K, Pemmaraju N, Jabbour E, et al. Modest activity of pomalidomide in patients with myelofibrosis and significant anemia. *Leuk Res.* 2013;37:1440–44.
82. Begna KH, Pardanani A, Mesa R, Litzow MR, Hogan WJ, Hanson CA, et al. Long-term outcome of pomalidomide therapy in myelofibrosis. *Am J Hematol.* 2012;87:66–8.
83. Begna KH, Mesa RA, Pardanani A, Hogan WJ, Litzow MR, McClure RF, et al. A phase-2 trial of low-dose pomalidomide in myelofibrosis. *Leukemia.* 2011;25:301–4.
84. Mesa RA, Pardanani AD, Hussein K, Wu W, Schwager S, Litzow MR, et al. Phase 1/2 study of pomalidomide in myelofibrosis. *Am J Hematol.* 2010;85:129–30.
85. Huang J, Tefferi A. Erythropoiesis stimulating agents have limited therapeutic activity in transfusion-dependent patients with primary myelofibrosis regardless of serum erythropoietin level. *Eur J Haematol.* 2009;83:154–55.
86. Tefferi A, Lasho TL, Mesa RA, Pardanani A, Ketterling RP, Hanson CA. Lenalidomide therapy in del(5)(q31)-associated myelofibrosis: cytogenetic and JAK2V617F molecular remissions. *Leukemia.* 2007;21:1827–28.
87. Mead AJ, Milojkovic D, Knapper S, Garg M, Chacko J, Farquharson M, et al. Response to ruxolitinib in patients with intermediate-1-, intermediate-2-, and high-risk myelofibrosis: results of the UK ROBUST Trial. *Br J Haematol.* 2015;170:29–39.
88. Al-Ali HK, Griesshammer M, le Coutre P, Waller CF, Liberati AM, Schafhausen P, et al. Safety and efficacy of ruxolitinib in an open-label, multicenter, single-arm phase 3b expanded-access study in patients with myelofibrosis: a snapshot of 1144 patients in the JUMP trial. *Haematologica.* 2016;101:1065–73.
89. Rago A, Latagliata R, Montanaro M, Montefusco E, Andriani A, Crescenzi SL, et al. Hemoglobin levels and circulating blasts are two easily evaluable diagnostic parameters highly predictive of leukemic transformation in primary myelofibrosis. *Leuk Res.* 2015;39:314–17.
90. Kröger N, Panagiota V, Badbaran A, Zabelina T, Trivai I, Araujo Cruz MM, et al. Impact of molecular genetics on outcome in myelofibrosis patients after allogeneic stem cell transplantation. *Biol Blood Marrow Transplant.* 2017;23:1095–101.
91. Barosi G, Vannucchi AM, De Stefano V, Pane F, Passamonti F, Rambaldi A, et al. Identifying and addressing unmet clinical needs in Ph-neg classical myeloproliferative neoplasms: a consensus-based SIE, SIES, GITMO position paper. *Leuk Res.* 2014;38:155–160.
92. Santos FP, Tam CS, Kantarjian H, Cortes J, Thomas D, Pollock R, et al. Splenectomy in patients with myeloproliferative neoplasms: efficacy, complications and impact on survival and transformation. *Leuk Lymphoma.* 2014;55:121–7.
93. Mesa RA, Nagorney DS, Schwager S, Allred J, Tefferi A. Palliative goals, patient selection, and perioperative platelet management: outcomes and lessons from 3 decades of splenectomy for myelofibrosis with myeloid metaplasia at the Mayo Clinic. *Cancer.* 2006;107:361–70.
94. Barosi G, Ambrosetti A, Buratti A, Finelli C, Liberato NL, Quaglini S, et al. Splenectomy for patients with myelofibrosis with myeloid metaplasia: pretreatment variables and outcome prediction. *Leukemia.* 1993;7:200–206.
95. Passamonti F, Giorgino T, Mora B, Guglielmelli P, Rumi E, Maffioli M, et al. A clinical-molecular prognostic model to predict survival in patients with post polycythemia vera and post essential thrombocythemia myelofibrosis. *Leukemia.* 2017;31:2726–31 [pub ahead of print].
96. Hernández-Boluda JC, Correa JG, García-Delgado R, Martínez-López J, Alvarez-Larrán A, Fox ML, et al. Predictive factors for anemia response to erythropoiesis-stimulating agents in myelofibrosis. *Eur J Haematol.* 2017;98:407–14.
97. Gisslinger H, Zagrijtschuk O, Buxhofer-Ausch V, Thaler J, Schloegl E, Gastl GA, et al. Ropeninterferon alfa-2b, a novel IFN α -2b, induces high response rates with low toxicity in patients with polycythemia vera. *Blood.* 2015;126:1762–69.
98. Gisslinger H, Klade C, Georgiev P, Skotnicki A, Gercheva-Kyuchukova L, Egyed M, et al. Final results from PROUD-PV a randomized controlled phase 3 trial comparing ropeginterferon alfa-2b to hydroxyurea in polycythemia vera patients. Presented at the American Society of Hematology 58th Annual Meeting; December 3–6, San Diego, CA; *Blood* 2016; 128: Abstract 475.
99. Harrison CN, Mead AJ, Panchal A, Fox S, Yap C, Gbandi E, et al. Ruxolitinib vs best available therapy for ET intolerant or resistant to hydroxycarbamide. *Blood.* 2017;130:1889–97.

Affiliations

Tiziano Barbui¹ · Ayalew Tefferi² · Alessandro M. Vannucchi³ · Francesco Passamonti⁴ · Richard T. Silver⁵ · Ronald Hoffman⁶ · Srđan Verstovsek⁷ · Ruben Mesa⁸ · Jean-Jacques Kiladjian⁹ · Rüdiger Hehlmann¹⁰ · Andreas Reiter¹⁰ · Francisco Cervantes¹¹ · Claire Harrison¹² · Mary Frances Mc Mullin¹³ · Hans Carl Hasselbalch¹⁴ · Steffen Koschmieder¹⁵ · Monia Marchetti¹⁶ · Andrea Bacigalupo¹⁷ · Guido Finazzi¹ · Nicolaus Kroeger¹⁸ · Martin Griesshammer¹⁹ · Gunnar Birgegard²⁰ · Giovanni Barosi²¹

¹ Division of Hematology, Ospedale Giovanni XXIII, Bergamo, Italy

² Division of Hematology, Department of Medicine, Mayo Clinic, Rochester, MN, USA

³ CRIMM, Center Research and Innovation of Myeloproliferative Neoplasms, University of Florence, AOU Careggi, Florence, Italy

⁴ Division of Hematology, Department of Medicine and Surgery, Ospedale di Circolo, University of Insubria, ASST Sette Laghi, Varese, Italy

⁵ Division of Hematology-Oncology, Weill Cornell Medicine, New York, NY, USA

⁶ Tisch Cancer Institute, Department of Medicine, Mount Sinai School of Medicine, New York, NY, USA

⁷ Department of Leukemia, M.D. Anderson Cancer Center, University of Texas, Houston, TX, USA

⁸ UT Health San Antonio Cancer Center, San Antonio, TX, USA

⁹ Centre d'Investigations Cliniques (CIC 1427), Assistance Publique—Hôpitaux de Paris, Hôpital Saint-Louis, INSERM, Université Paris 7, Paris, France

¹⁰ Department of Hematology and Oncology, University Hospital Mannheim, Heidelberg University, Mannheim, Germany

¹¹ Hospital Clínic, IDIBAPS, University of Barcelona,

Barcelona, Spain

¹² Department of Hematology, Guy's and St. Thomas' NHS Foundation Trust, London, UK

¹³ Centre for Medical Education, Queen's University, Belfast, UK

¹⁴ Department of Hematology, Zealand University Hospital, Roskilde, Denmark

¹⁵ Department of Hematology, Oncology, Hemostaseology and Stem Cell Transplantation, Faculty of Medicine, RWTH Aachen University, Aachen, Germany

¹⁶ Hematology Day Service, Oncology SOC, Hospital Cardinal Massaia, Asti, Italy

¹⁷ Istituto di Ematologia, Università Cattolica del Sacro Cuore, Fondazione Policlinico Universitario Gemelli, Rome, Italy

¹⁸ Department of Stem Cell Transplantation, University Hospital Hamburg-Eppendorf, Hamburg, Germany

¹⁹ Department of Hematology and Oncology, Johannes Wesling Medical Centre Minden, Academic Hospital of the University of Hannover, Minden, Germany

²⁰ Department of Hematology, Uppsala University, Uppsala, Sweden

²¹ Center for the Study of Myelofibrosis, IRCCS Policlinico S. Matteo Foundation, Pavia, Italy