

https:/doi.org/10.1093/ckj/sfad076 Advance Access Publication Date: 6 April 2023 CKJ Review

CKJ REVIEW

Early aging and premature vascular aging in chronic kidney disease

Cem Tanriover¹, Sidar Copur¹, Ali Mutlu ¹, Ibrahim Batuhan Peltek¹, Andrea Galassi², Paola Ciceri², Mario Cozzolino ¹ and Mehmet Kanbay ³

¹Department of Medicine, Koc University School of Medicine, Istanbul, Turkey, ²Department of Health Sciences, Renal Division, University of Milan, Milan, Italy and ³Department of Medicine, Division of Nephrology, Koc University School of Medicine, Istanbul, Turkey

Correspondence to: Mario Cozzolino; E-mail: mario.cozzolino@unimi.it

ABSTRACT

Aging is the progressive decline of body functions and a number of chronic conditions can lead to premature aging characterized by frailty, a diseased vasculature, osteoporosis, and muscle wasting. One of the major conditions associated with premature and accelerated aging is chronic kidney disease (CKD), which can also result in early vascular aging and the stiffening of the arteries. Premature vascular aging in CKD patients has been considered as a marker of prognosis of mortality and cardiovascular morbidity and therefore requires further attention. Oxidative stress, inflammation, advanced glycation end products, fructose, and an aberrant gut microbiota can contribute to the development of early aging in CKD patients. There are several key molecular pathways and molecules which play a role in aging and vascular aging including nuclear factor erythroid 2-related factor 2 (Nrf-2), AMP-activated protein kinase (AMPK), sirtuin 1 (SIRT1), and klotho. Potential therapeutic strategies can target these pathways. Future studies are needed to better understand the importance of premature aging and early vascular aging and to develop therapeutic alternatives for these conditions.

LAY SUMMARY

CKD is an important cause of premature and accelerated aging. It results in early vascular aging together with arterial stiffness. Several cellular and molecular mechanisms can contribute to the development of early aging in CKD patients. Premature vascular aging in CKD patients has been considered as a prognostic marker of mortality and cardiovascular morbidity. Potential therapeutic strategies can target these pathways.

Keywords: CKD, CKD-MBD, inflammation, vascular calcification

INTRODUCTION

Chronic kidney disease (CKD) is characterized by having structurally and/or functionally abnormal kidneys present for more than >having one or more marker of kidney dysfunction such as albuminuria [1, 2]. It is believed that \sim 15% of the US general population has been impacted by CKD between 2013 and 2016 [2]. CKD is associated with a number of comorbidities and chronic conditions including premature aging [3, 4].

Received: 24.1.2023; Editorial decision: 5.4.2023

[©] The Author(s) 2023. Published by Oxford University Press on behalf of the ERA. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (https://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

The aging process can either be physiological or pathological, which is also termed premature aging. Physiological aging is the result of the functional decline of the body and it is influenced by genetic as well as environmental factors such as socioeconomic status, stress, sedentary lifestyle, diet, smoking, and consumption of alcohol and other drugs [5, 6]. On the other hand, premature aging is characterized by accelerated functional decline that results in aging earlier than expected for chronological age [7]. A number of chronic conditions are associated with premature aging characterized by frailty, a diseased vasculature, the development of osteoporosis, and muscle wasting [3, 8]. CKD, which is one of the major conditions associated with premature and accelerated aging, is also related to early vascular aging and the stiffening of the arteries [3, 9]. The premature stiffening of especially the central arteries in CKD patients has been considered as a marker of prognosis for mortality and cardiovascular morbidity and therefore requires further attention [9-13].

In this review, we first describe the underlying mechanisms of early aging in CKD patients. We then elucidate the characteristics of premature vascular aging in CKD and end-stage renal disease (ESRD) and delineate its clinical implications. We finally explain the key molecular pathways and molecules that are critical for the development of aging and vascular aging while discussing the potential therapeutic strategies that can target these molecular structures.

Early aging in chronic kidney disease

The definition of aging can be broadly described as the progressive loss of the functional ability and the physiological functions of the body together with declining fertility and higher mortality over time [8, 14]. It is influenced by genetic, epigenetic, and environmental factors [15]. Older age is associated with a higher prevalence of chronic diseases and many chronic conditions cause early aging [16–20]. One of the conditions associated with premature and accelerated aging is CKD which is characterized by progressive vascular disease and early vascular aging, muscle wasting, osteoporosis, frailty, and systemic inflammation [9, 21]. Oxidative stress, inflammation, an aberrant gut microbiota, advanced glycation end products, and fructose consumption and are all factors contributing to early aging in kidney disease patients (Fig. 1).

Oxidative stress

Oxidative stress is an important mechanism for accelerated aging and muscle wasting in CKD [22]. Oxidative stress in CKD results from intravenous iron treatment, the activation of the renin-angiotensin system (RAAS), decreased antioxidants, and features related with dialysis such as the incompatibility of membranes or fluids [23, 24]. Mitochondrial dysfunction also contributes to oxidative stress in CKD [25, 26]. Protein bound uremic toxins such as p-cresyl sulfate and indole-3-acetic acid are shown to inhibit mitochondrial oxidative phosphorylation in renal proximal tubule epithelial cells by inhibition of succinate dehydrogenase enzyme [27]. Oxidative stress also causes alterations in the molecular structure of proteins, carbohydrates and lipids with ensuing tissue and organ damage.

Cellular senescence, immunosenescence, and inflammaging

The concept of cellular senescence was discovered in the 1960s by demonstrating the loss of replicative potential in human cells

[28]. Senescence is characterized by cell cycle arrest in the G1 or G2 phase, apoptosis resistance, and altered gene expression [29]. Cellular senescence is a physiological process in embryonic development and wound healing but can be pathologic leading to aging and disease states [30]. Senescence has been suggested as a major cause of age-related diseases [31]. Cellular senescence can be induced by various stimuli, such as telomere shortening or dysfunction, mitochondrial dysfunction, epigenetic influences, DNA damage, oncogene activation, and inactivation of tumor suppressor genes [32]. Although senescent cells lose their replicative potential, they remain metabolically active. Senescent cells undergo several proinflammatory and pro-fibrotic changes in gene expression and cellular metabolism. This new phenotype is named senescence-associated secretory phenotype (SASP). SASP is characterized by increased expression and secretion of growth factors, cytokines, proteases, and chemokines [33]. These factors signal nearby cells in a paracrine fashion causing paracrine senescence and altering their surrounding environment [34]. SASP can also modulate the immune system with these factors. It can activate the immune system and increase the elimination of senescent cells or promote the persistence and accumulation of senescent cells [35]. With aging, several cells in the kidney, such as renal tubular epithelial cells, podocytes, mesangial cells, immune cells, and endothelial cells, undergo cellular senescence. However, senescence is most notably seen in renal tubular epithelial cells [36]. Renal tubular cell senescence is associated with the changes seen in aged kidneys, including tubular atrophy, interstitial fibrosis, and glomerulosclerosis. Although SASP might benefit tissue regeneration after an acute kidney injury, prolonged SASP exposure has detrimental effects on tissue function and repair, which eventually cause CKD [37]. Furthermore, SASP causes sterile inflammation and contributes progression of CKD by promoting fibrosis in the kidney [36].

Senescence can be associated with immune system dysfunction and dysregulation, changes collectively referred to as immunosenescence and inflammaging, respectively [38]. Immunosenescence is considered harmful because it is associated with low-grade sterile inflammation with decreased cellular responses against infections and vaccines [39]. Changes seen with immunosenescene are influenced by several factors such as genetics, nutrition, exercise, exposure to microorganisms, sex, and human cytomegalovirus status [38]. Inflammaging is characterized by high blood levels of proinflammatory cytokines in older individuals [40]. Inflammaging is associated with an increased risk of chronic diseases including cardiovascular diseases, CKD and dementia. Several factors are postulated as risk factors and causes of inflammaging, such as chronic infections, impaired autophagy and cellular degradation, visceral obesity, genetic susceptibility and altered microbiota, and increased gut permeability [41].

Inflammation and metaflammation

Inflammation results from hyperactive innate immunity, which is characterized by activated macrophages and increased proinflammatory cytokines such as interleukin 6, interleukin 1, and tumor necrosis factor [42]. There are also several changes in the adaptive immunity. Decreases in the number and function of naive T cells and increased numbers of memory T cells, especially proinflammatory CD4⁺CD28⁻ T cells are evidence of immunosenescence in CKD [43]. Visceral obesity, smoking, low-grade infection, and social and psychosocial stresses are associated with increased expression of inflammatory genes [44].

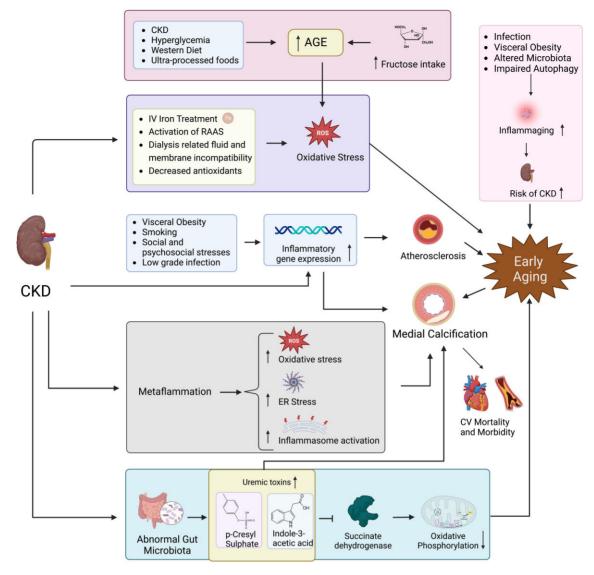


Figure 1: The factors contributing to early aging in CKD. AGE: Advanced glycation end products, CKD: Chronic kidney disease, CV: Cardiovascular, ER: Endoplasmic reticulum, RAAS: Renin-Angiotensin-Aldosterone system, ROS/Oxidative stress.

In addition, overhydration is a common complication in CKD, and it can also contribute to systemic inflammation via bacterial or endotoxin translocation due to severe gut edema [45]. Systemic inflammation leads to muscle wasting in CKD [46]. Proinflammatory cytokines produced by senescent cells with the SASP, activates proteolytic mechanisms, and impair muscle regeneration [29]. Inflammation accelerates atherosclerosis and is an independent risk factor for early medial calcification [47]. Accelerated medial calcification is common among patients with uremia and with other chronic inflammatory disorders [48-53]. Metaflammation is described as a long-term low-grade inflammatory state induced by metabolic variations causing increased reactive oxygen species (ROS), inflammasome activation, and endoplasmic reticulum stress, which is important for cell homeostasis and insulin signaling pathways. Therefore, metaflammation can cause cardiovascular morbidities due to induction of endothelial dysfunction and vascular calcifications [54].

Nutrition and digestion: gut microbiota, advanced glycation end products, and fructose consumption

Patients with CKD and ESRD are at risk of having an abnormal intestinal microbiota, characterized by alterations in saccharolytic bacteria to increases in proteolytic bacteria [55–57]. This change in the microbiota results in the production of several uremic toxin precursors in the intestines such as p-cresol and indole, both subsequently metabolized by the colonic mucosa and the liver to indoxyl sulfate and p-cresyl sulfate resulting in a high toxicity specifically targeting the cardiovascular system [55, 57, 58]. The intestines are also known to modulate the immune system both locally and systemically via complex interactions between the gut microbiota and immune system. Short chain fatty acids, which are metabolites produced in the colon by bacterial fermentation, have modulatory functions on the immune system [59, 60]. In addition, with an impaired gut barrier, intestinal bacteria and endotoxins infiltrate the mucosa and translocate to the blood stream, circulating to different organs and tissues including the kidney contributing to inflammation [61].

Advanced glycation end products (AGEs) are a group of compounds that are formed by the nonenzymatic glycation of lipids, proteins and DNA. AGEs are not only formed during hyperglycemia, but also in states with high oxidative stress such as in CKD [62]. Furthermore, AGEs can be found in ultra-processed foods. The western diet, which is low in fruits and vegetables and high in animal proteins and processed foods, is a risk factor for CKD [63]. During food processing, high temperatures, dehydration, decompression, salt, irradiation, and preservatives significantly alter the lipids, proteins, and carbohydrates and lead to the formation of AGEs within foods [64]. The kidneys are the primary site of AGE excretion and in chronically diseased kidneys the circulating levels of AGEs increase [65]. Excessive AGEs can contribute to the progression of CKD. AGEs and their receptors including advanced glycation end productspecific receptor (RAGE) trigger oxidative stress and inflammation, in turn potentially contributing to aging [66]. Thus, AGEs are related to cardiovascular complications and progression of renal dysfunction as well as early aging in CKD. Excessive intake of fructose is known to be associated with hypertension, diabetes, and metabolic syndrome, which are risk factors for the development of CKD [67, 68]. In animal studies, fructose induces tubular cell proliferation with low-grade tubulointerstitial injury by the induction of chemoattractant proteins such as monocyte chemoattractant protein-1 from tubular cells and intercellular adhesion molecule-1 in renal microvascular endothelial cells [69, 70]. Long-term fructose consumption was shown to increase AGEs and accelerated aging in an animal study [71].

Premature vascular aging in CKD patients

In addition to accelerated aging, CKD and ESRD have been associated with vascular calcification, premature vascular aging and the stiffening of the arteries [9, 72]. Early vascular aging refers to accelerated age-related changes in arterial structure and function [73]. Early vascular aging is characterized by profound medial vascular calcification, which is primarily driven by vascular smooth muscle cells [47]. Early vascular aging is an intermediate cardiovascular endpoint and independent predictor of cardiovascular disease and cardiovascular mortality [74]. Arterial stiffness, which can be measured with carotid-femoral pulse wave velocity, is a hallmark of early vascular aging and pulse wave velocity has been proposed as a marker of early vascular aging [75]. Characteristically, arterial stiffness is much more evident in the aorta as well as the other central arteries compared to those located in the periphery in this patient group [9, 76]. An increased aortic stiffness can be considered to be a contributor to left ventricular hypertrophy and a fall in the perfusion of the coronary arteries as well as a marker of prognosis for mortality and cardiovascular morbidity with the pulse wave velocity of the aorta being an independent predictor for all-cause and cardiovascular mortality [9-12, 77, 78]. Furthermore, small vessel disease in the cerebral structures has also impacted the development of cognitive impairment in CKD patients [79].

The stiffening of the arteries represents the overall aging of the arterial network [9]. Arterial remodeling and enlargement together with a greater arterial stiffness and early vascular aging are seen in the earlier stages of CKD, in line with the fall in renal function [9, 80]. In addition, one study reported that the stiffening of the aorta was independently related to the rate of change of kidney function in individuals with CKD stages 3 and 4 [81]. The independent predictors of \geq 25% decrease in kidney func-

tion or initiation of renal replacement therapy were the pulse wave velocity of the aorta, systolic blood pressure, and the urine protein-to-creatinine ratio [81].

There is an important relationship in regard to the pulse wave velocity of the aorta and age, in comparison to the general population: A gradual fall in the difference in the mortality rate occurs with increasing age in patients with ESRD [9]. The pulse wave velocity of the aorta has been reported to predict the cardiovascular and all-cause mortality significantly in younger individuals with ESRD [9]. One study reported that a pulse wave velocity >12 m/s was able to present prognostic value in patients with ESRD younger than 60 years of age, however in older individuals this prognostic information was no longer relevant [82]. A 10 to 30 times greater cardiovascular mortality exists in individuals with ESRD in comparison to the general population and among young patients mortality rates are up to 500 times greater [9, 83].

Overall, early aging of the vasculature and stiffening of the arteries are in close relation to CKD and ESRD. A higher aortic stiffness has been suggested as a prognostic marker and aortic pulse wave velocity has been considered an independent predictor for all-cause and cardiovascular mortality. Given that vascular problems could potentially occur in the initial stages of CKD, clinicians should consider earlier testing of the cardiovascular system in younger or early-stage kidney disease patients to gain information to better understand the prognosis and to direct the treatment strategies of these patients [9]. Furthermore, future large-scale studies are needed to improve our understanding regarding the clinical implications of early aging, premature vascular stiffening and vascular calcification in general in CKD and ESRD patients.

The major molecular pathways associated with aging and vascular aging

Several key molecular pathways and molecules exist that play a role in the development of aging and vascular aging. Potential therapeutic strategies can target these pathways (Fig. 2).

Nuclear factor erythroid 2-related factor 2 (nrf2)

Nuclear factor erythroid 2-related factor 2 (Nrf-2), a basicleucine-zipper-like transcription factor, is a key regulator of the balance between pro-oxidative or antioxidative defense mechanisms [84]. The major functions of Nrf-2 include the upregulation of the genes encoding for antioxidant or phase II detoxifying enzymes such as NAD(P)H (nicotinamide adenine dinucleotide phosphate) dehydrogenase-1 (NQO1), heme oxygenase-1/2, tryptophan hydroxylase-1 or glutathionetransferase [85]. At basal conditions, the activity of Nrf-2 located at cytosol is suppressed via Kelch-like ECH-associated protein1 (Keap1), which is involved in the ubiquitination and proteasomal degradation of Nrf-2 while oxidative signals result in the nuclear translocation of Nrf-2 [86-88]. Additionally, few Keap-1 independent regulatory mechanisms for Nrf-2 activity have been identified including the activity of glycogen synthase kinase 3β (GSK- 3β) and endoplasmic reticulum stress [89, 90]. In vitro and in vivo studies have demonstrated that over-expression or activation of Nrf-2 result in decline in the expression of proinflammatory cytokines such as the association between over-expression of Nrf-2 at endothelial cells and decreased expression of vascular cell adhesion molecule-1 (VCAM-1), monocyte chemoattractant protein-1 (MCP-1), and tumor necrosis factor-alpha

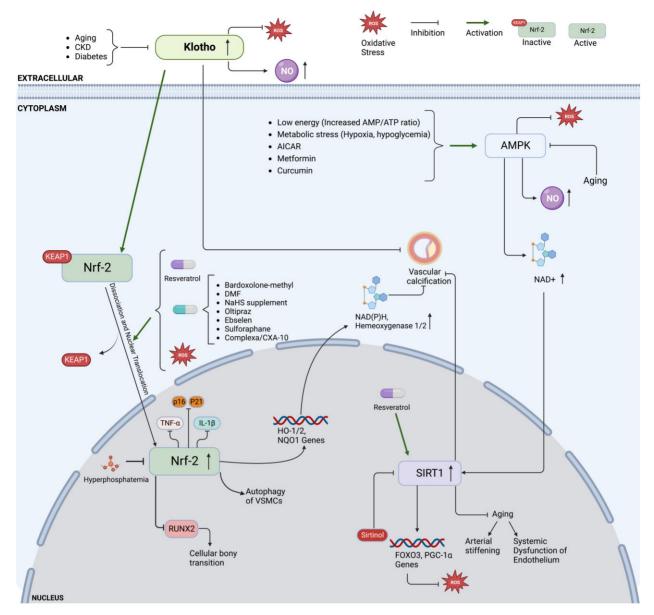


Figure 2: The major molecular pathways associated with aging and vascular aging. KEAP1 maintains Nrf-2 in the inactivated state. Oxidative stress and several medications such as resveratrol activate and translocate Nrf-2 via the dissociation of KEAP1. Activated Nrf-2 upregulates the genes encoding antioxidants such as NQO1 and HO-1/2 that, in turn, inhibit vascular calcification. Hyperphosphatemia suppresses the activity of Nrf-2. Klotho activates Nrf-2 while also increasing NO, decreasing oxidative stress and potentially improving vascular dysfunction. AMPK can be activated via metabolic stress, low energy states, AICAR, metformin, and curcumin. The activity of AMPK decreases in aging resulting in arterial stiffening and endothelial dysfunction. Activated AMPK decreases oxidative stress and elevates NO. AMPK also increases, via NAD⁺, SIRT1 activity that plays a role in inhibiting aging and vascular calcification. SIRT1 also has an antioxidant function through the transcription of FOXO3 and PGC-1*a*. Furthermore, SIRT1 inhibits arterial stiffening and endothelial dysfunction that result from aging. SIRT1 can be activated by resveratrol. AICAR: Aminoimidazole carboxamide ribonucleotide, AMP: Adenosine monophosphate, AMPK: AMP-activated protein kinase, ATP: Adenosine triphosphate, CKD: Chronic Kidney Disease, DMF: Dimethyl fumarate, FOXO3: Forkhead Box O3, HO-1/2: heme oxygenase-1/2, IL-1: Interleukin 1, KEAP1: Kelch-like ECH-associated protein 1, NAD+: Nicotinamide adenine dinucleotide, NAHS: Sodium hydrosulfide, NO: Nitric oxide, NQO1: NAD phosphate) dehydrogenase-1, Nrf-2: Nuclear factor erythroid 2-related factor 2, PGC-1*a*: Peroxisome proliferator-activated receptor-gamma coactivator-1- alpha, p16(CDKN2A): Cyclin-dependent kinase inhibitor 2A, p21 (CDKN1A): Cyclin-dependent kinase inhibitor 1, ROS/Oxidative stress, RUNX2: Runt-related transcription factor 2, SIRT1: Sirtuin 1, TNF-*a*: Tumor necrosis factor-*a*, VSMC: Vascular smooth muscle cell.

(TNF- α) [91–93]. A study in which the effects of T-cell specific augmentation of Nrf-2 on mice are being reviewed has demonstrated that upregulation of Nrf-2 is associated with higher levels of CD25(+) and FOXP3(+) regulatory T cells and lower levels of proinflammatory cytokines such as TNF- α , interferon-gamma (IFN- γ), and interleukin-17, thus, leading to protection from ischemia-reperfusion injury-induced acute kidney injury (AKI) [94]. A study conducted on 2155 patients with stage 1–5 CKD has identified five metabolites for which serum levels are highly linked to the markers of kidney injury. Among those five metabolites, supplementation of cultured human kidney cells with 5-methoxytryptophan (5-MTP) results in the inhibition of NF- κ B signaling and amelioration of renal interstitial fibrosis in response to ischemia-reperfusion injury that is mediated via the upregulation of Nrf-2 signaling pathway [95]. Moreover, studies conducted on rats have illustrated that supplementation with sulforaphane, a Nrf-2 agonist, leads to decline in arsenic-induced nephrotoxicity mediated via decline in the formation of renal ROS and lipid peroxidation products, DNA damage, and increased formation of phase II antioxidant enzymes [96].

Early vascular aging has been linked to vascular stiffening, higher risk for cardiovascular diseases, and cardiovascular mortality while the exact underlying pathophysiological mechanisms are unclear [97]. Along with chronic low-grade inflammation, increased oxidative stress and formation of ROS result in endothelial dysfunction that has been associated with vascular calcification and cellular senescence [98]. In vitro and animal studies have shown that supplementation with sodium hydrosulfide, an activator of Nrf-2/Keap-1 signaling system, results in the upregulation of HO-1/2 and NQO1, both of which are antioxidant enzymes, causing the amelioration of vascular calcification [99, 100]. Moreover, Nrf-2 activation leads to the suppression of cellular bony transition mediated via runt-related transcription factor 2 (Runx2) [101]. Hyperphosphatemia among CKD patients have shown to suppress Nrf-2 activity both at transcriptional, translational and post-translational levels while administration of resveratrol, an agonist for Nrf-2, has been linked to decline in the deposition of mineralized matrix at vasculature, decline in mitochondrial damage, and intracellular calcium deposition, therefore reversing the hyperphosphatemia-related vascular alterations [102, 103]. Furthermore, treatment of mice or rat subjects with dimethyl fumarate (DMF), another activator of Nrf-2, has led to significant decline in vascular calcification at aorta and carotid artery even under hypercalcemic and hyperphosphatemic environments [104, 105]. Interestingly, activation of Nrf-2 results in the autophagy of vascular smooth muscle cells to ameliorate vascular calcification through undiscovered pathophysiological mechanisms [106].

The role of Nrf-2 signaling in cellular senescence, a state characterized by cellular growth arrest without losing metabolic activity, has been investigating various cell types including cardiac muscle cells, vascular endothelial cells, and epithelial tissues [107-109]. Studies conducted on Nrf-2 knockout mice have illustrated that Nrf-2 depletion results in the upregulation of cellular senescence markers such as cyclin-dependent kinase inhibitor 2A (p16INK4a, CDKN2A) and cyclin-dependent kinase inhibitor 1 (p21, CDKN1A), increased production of proinflammatory cytokines involved in the process such as interleukin-1beta (IL-1 β) and TNF [110]. The underlying mechanism is not clear, nevertheless, one hypothesis includes the downregulation of Nrf-2 via miRNAs derived from senescent cells such as miR-126, miR-21, and miR-100 [111]. Moreover, a study conducted by Stenvinkel et al. on patients with living donor kidney transplantation showed that expression of cyclin-dependent kinase inhibitor 2A (p16INK4a, CDKN2A) is also related to the severity of vascular calcification in ESRD [112].

Despite its central role in the physiology of cellular senescence, vascular calcification, and vascular aging, the efficiency of treatment strategies targeting Nrf-2 is not well established. A phase I human clinical trial investigating the role of once daily administration of bardoxolone-methyl, a Nrf-2 agonist, on 47 patients with advanced stage solid organ tumors or lymphoma demonstrated potential beneficial effects with few dose-limiting adverse effects, mainly being hepatotoxicity, while upregulation of Nrf-2 is mediated via upregulation of NQO1 mRNA levels [113]. Moreover, another Nrf-2 agonist referred as DMF is currently at phase III for the treatment of multiple sclerosis [114]. Furthermore, there are multiple ongoing clinical trials investigating the clinical utility of various Nrf-2 activators such as DMF (NCT02784834, NCT02546440, NCT00810836), bardoxolonemethyl (NCT00550849, NCT00811889, NCT01351675), oltipraz (NCT00006457, NCT02068339), sulforaphane (NCT01008826, NCT02880462, NCT02801448, NCT03220542), sulforadex (NCT01228084), ebselen (NCT03013400), and complexa/CXA-10 (NCT02248051, NCT03449524, NCT03422510). Despite promising initial results from early phase clinical trials, there is clear need for future studies investigating the effects of Nrf-2 agonists on vascular calcification, malignancies, and vascular aging.

AMP-activated protein kinase (AMPK)-sirtuin 1 (SIRT1)

AMP-activated protein kinase (AMPK) and Sirtuin 1 (SIRT1) are both associated with vascular aging and age-related kidney damage [115–118]. AMPK and SIRT1 are closely linked, characterized by AMPK-deficient states causing an improper activation of SIRT1 and its downstream pathways in reduced energy states [116–118]. AMPK elevates SIRT1 activity by rising NAD⁺ levels in the cell and therefore regulates the downstream SIRT1 signaling molecules such as forkhead box O3 (FOXO3) and peroxisome proliferator-activated receptor- γ coactivator 1 (PGC1 or PPARGC1A), which also act as potential substrates for AMPK [119–122].

AMPK is a serine-threonine kinase that, through the serinethreonine liver kinase B1 (LKB1)-AMPK pathway, is activated via metabolic stresses that block ATP generation such as hypoxia and hypoglycemia or increase ATP use such as the contraction of muscles [123]. AMPK can also be activated by metformin and aminoimidazole carboxamide ribonucleotide (AICAR) [118, 124]. AMPK, which is crucial in energy-sensing, also plays an important role in energy balance, stress resistance, and metabolism [118, 123].

Lesniewski et al. showed that exercise decreased oxidative stress and elevated nitric oxide bioavailability leading to the restoration of endothelium-dependent dilation (EDD) in old mice and that AMPK once active showed similar effects to exercise [125]. In comparison to younger controls of 3-6 months of age, older mice (28-30 months) had decreased arterial AMPK levels and suppression of EDD by superoxides [125]. AMPK activated by AICAR resulted in an elevation of arterial AMPK and reversal of the impaired EDD [125]. Similarly, one study showed that AMPK activity was reported to be decreased in the cerebral arteries of aged rodents and the administration of curcumin ameliorated aging associated cerebrovascular dysfunction through the AMPK/uncoupling protein 2 (UCP2) pathway [126]. Furthermore, activating AMPK by using metformin has been suggested to increase endothelial function in rodents with type 1 and 2 diabetes [118, 127, 128]. In addition, one clinical study reported that short-term metformin use ameliorated arterial stiffness and endothelial function in young women with PCOS, suggesting the activation of AMPK via metformin a key underlying mechanism explaining these results [129]. Several trials (NCT03309007, NCT01765946) have investigated the effect of metformin on anti-aging through AMPK signaling. Kreutzenberg et al. (NCT01765946) have reported that prediabetic individuals had improved effector pathways known regulate longevity in animal models following metformin therapy [130].

These findings suggest that AMPK signaling is a crucial pathway related to aging and vascular aging. Future preclinical and large-scale clinical studies are needed to better understand the role of AMPK and to guide therapeutic strategies through this pathway.

SIRT1 plays an important role in longevity and is responsible for the deacetylation of histone and non-histone proteins for the modification of transcription factors, coregulators, and proteins to adjust gene expression in accordance with the current energy state of the cell and to resist stress by tempering proinflammatory and oxidative stress pathways [118, 131-134]. SIRT1 activity has been shown to decrease oxidative stress, proapoptotic pathways, and inflammation, and improve telomere stabilization, DNA repair pathways, and insulin sensitivity [118, 135]. SIRT1 is known to inhibit renal inflammation, fibrosis, and renal cell apoptosis [136]. Decreased levels of SIRT1 aggregate renal fibrosis, a characteristic feature in CKD [137]. Several animal studies investigated mechanisms of SIRT1-related renal fibrosis, including decreased matrix metalloproteinase 14 expression and disinhibition of profibrotic TGF- β 1 [138, 139]. The role of SIRTs in the pathogenesis of kidney diseases is nicely summarized previously in a comprehensive review [140].

Studies have postulated that decreased SIRT1 may be critical in the development of a dysfunctional vascular endothelium associated with aging [141-144]. An increased production of SIRT1 in endothelial cells contribute to the prevention of systemic dysfunction of the endothelium and a heightened stiffening of large arteries, both changes related to aging [145-147]. Furthermore, a study with endothelial SIRT1-deleted mouse found that SIRT1 deletion is associated with accelerated senescence of endothelial cells with impaired endothelial dependent vasodilation [148]. Endothelial senescence can explain endothelial dysfunction. Although endothelial senescence shows similar features to other types of senescence, it also shows some unique features [149]. A unique feature of endothelial SASP is its role in arterial dysfunction, including increased levels of ROS and reduced nitric oxide levels [116]. In one study, Donato et al. showed that in older (30 months) mice aortic protein expression of SIRT1 was significantly decreased compared to younger mice (5-7 months) and acetylcholine induced peak EDD was significantly inferior in isolated femoral arteries with aging [141]. The application of SIRT1 inhibitor sirtinol led to a decreased EDD in both young and old mice [141]. Furthermore, SIRT1 is shown to be a potential inhibitor of vascular calcification with reduced SIRT1 being associated with vascular calcification development and activated SIRT1 resulting in decreased vascular calcification [136, 150-153]. These studies suggest that SIRT1 is an important molecule in vascular dysfunction and aging and could potentially give rise to therapeutic strategies to combat this condition.

Several compounds have been investigated to evaluate the effect of SIRT1 activation on vascular dysfunction, and among these the best studied molecule is resveratrol. This molecule acts as an activator of SIRT1 as well as functions through \leq 15 other pathways including as an agonist of Nrf-2 together with antioxidant and phytoestrogen effects [102, 118, 154–157]. One study has shown that resveratrol administered to older aged mice had a significant decrease in markers of aging such as lower albuminuria, inflammation, apoptosis in the endothelium of vessels and an elevated elasticity in the aorta, improved motor coordination, decreased cataracts, and a preservation of bone density [158]. However, this study could not detect a prolonged lifespan in mice treated with resveratrol [158].

As of January 2022, there were a total of 194 listed trials on resveratrol on https://clinicaltrials.gov. Among these, 18 studies have investigated the effects of resveratrol on vascular conditions (NCT01842399, NCT01668836, NCT03597568, NCT01564381, NCT02246660, NCT02690064, NCT03436992, NCT04633551, NCT03743636, NCT04449198, NCT01881347. NCT03762096, NCT02998918, NCT01185067, NCT02137421, NCT03253913, NCT05093244, NCT04117022). One study assessed the role of caloric restriction and resveratrol on the sirtuin system in women and men between 55 and 65 years of age (NCT01668836) [159]. This study was conducted on 48 healthy subjects randomized to 30 days of resveratrol (500 mg/day) or caloric restriction (1000 cal/day) [159]. Both resveratrol and caloric restriction led to an elevated plasma levels of SIRT with no difference between the two groups, however, plasma levels for an endogenous secretory receptor for an advanced glycation end product (esRAGE) were not changed and was similar for both groups [159]. SIRT1 and esRAGE are associated with the protection of the vasculature [159]. These results indicate that an increase in both molecules occur following resveratrol administration, which can potentially have a protective effect on vascular dysfunction; future studies are needed to investigate this condition.

Overall SIRT1 is a crucial molecule in the prevention of vascular dysfunction that can be targeted as a therapeutic strategy and resveratrol is a promising agent which functions by acting on SIRT1. Future large-scale clinical studies with long follow-up times as well as preclinical studies to understand the pathophysiologic mechanisms underlying this molecule are needed.

Phosphate and klotho

Hyperphosphatemia has a crucial role in early aging in patients with CKD. As clearance decreases in CKD, increased levels of inorganic phosphate can cause vascular aging and inflammation [160]. In addition, hyperphosphatemia could precipitate oxidative stress [161]. The activation of osteogenic genes, the production of hydroxyapatite, and vascular calcifications have all been linked to high phosphate levels. Owing to their sensitivity to inorganic phosphate concentrations, vascular smooth muscle cells can alter and adjust some of their functions. These modifications in response to changes in inorganic phosphate trigger calcification-promoting processes [162].

Klotho exists as a membrane-bound and soluble form. The soluble form can act as a hormone and regulate glycosidase and transporter actions whereas the membrane-bound klotho plays a role in FGFR signaling [163]. Low klotho levels are correlated with kidney dysfunction, increased risk of atherosclerosis, and accelerated aging [164]. Klotho deficiency is one of the markers of early aging and an important contributor of vascular calcification leading to the hyperplasia of the intimal layer, calcification of the media, endothelial dysfunction, an increased stiffness within the arteries, hypertension, and impairments in vasculogenesis [165–168].

In one of the earliest studies investigating the role of klotho in the aging process, Kuro-o *et al.* showed that a defective expression of the klotho gene led to a phenotype similar to human aging in mice, which consisted of a decreased lifespan, arteriosclerosis, osteoporosis, infertility, emphysema, and atrophied skin [169]. Later studies reported that the over-expression of klotho led to a prolonged life span, provided protective cardiac effects, and decreased oxidative stress in mice [118, 170–172]. Futhermore, klotho concentrations are reported to fall with increasing age in humans with a decrease in klotho levels by 2-fold from 40 to 70 years of age [173, 174]. Klotho levels are also known to decline in several frequently seen diseases such as CKD, diabetes, and neurodegenerative conditions [175].

Table 1: The: current knowns and unknowns surrounding early aging and premature vascular aging.

Knowns

Oxidative stress due to uremic toxins, activated RAAS, and decreased antioxidants cause muscle wasting and early aging.

Visceral obesity, smoking, low-grade infection, and social and psychosocial stresses are associated with increased expression of inflammatory genes.

Altered gut microbiota and impaired gut barrier contribute to systemic inflammation.

Early vascular aging in CKD causes arterial remodeling, increased arterial stiffness, cardiovascular morbidity, and mortality.

Nrf-2 reduces oxidative stress, renal interstitial fibrosis, and vascular calcification in CKD through several mechanisms.

Klotho is a signaling protein with two forms: the free form in the cytosol and the membrane-bound form. Klotho levels correlate with kidney function, and its deficiency significantly contributes to vascular calcification in CKD.

Unknowns

Nrf-2 depletion results in the upregulation of cellular senescence markers such as p16INK4a (CDKN2A) and p21 (CDKN1A). However, the underlying mechanisms are not clear.

Future preclinical and large-scale clinical studies are needed to understand the role of AMPK to better guide therapeutic strategies through this pathway.

Recombinant klotho and gene therapy strategies are promising approaches. However, further studies are required to evaluate these therapeutic mechanisms better and transition their use toward the clinics. Cellular senescence, immunosenescence, and inflammaging cause sterile inflammation, aggravating kidney damage.

Metainflammation contributes to cardiovascular morbidities through endothelial dysfunction and vascular calcifications.

AGEs and high fructose diet are both risk factors and drivers of the progression of CKD and early aging.

Aortic stiffness is suggested as a prognostic marker, and aortic pulse wave velocity has been considered an independent predictor for all-cause and cardiovascular mortality.

AMPK and SIRT1 are closely linked signaling proteins protective against vascular calcification and are related to longevity.

Nrf-2 stimulates the autophagy of vascular smooth muscle cells to ameliorate vascular calcification through undiscovered pathophysiological mechanisms.

Resveratrol is a promising agent acting on SIRT1, Nrf-2 as well as several other pathways. Future large-scale clinical studies with long follow-up times and preclinical studies are needed to understand the pathophysiologic mechanisms underlying this molecule.

AGE: Advanced glycation end products, AMPK: AMP-activated protein kinase

CKD: Chronic kidney disease, Nrf-2: Nuclear factor erythroid 2-related factor 2, p16INK4a (CDKN2A): Cyclin-dependent kinase inhibitor 2A, p21 (CDKN1A): Cyclin-dependent kinase inhibitor 1, RAAS: Renin-Angiotensin-Aldosterone system, SIRT1: Sirtuin 1

Klotho has also been shown to play a role in vascular changes. One study reported that serum klotho concentrations were lowered by ${\sim}45\%$ in individuals with arterial stiffness and hypertension [176]. Furthermore, klothohaplodeficient (Klotho+/-) mice demonstrated significant elevations in in pulse wave velocity and blood pressure suggesting worsening arterial stiffness and hypertension [176]. Klotho-haplodeficiency (Klotho+/-) was also shown to decrease endothelial nitric oxide synthase (eNOS) expression in the aorta as well as lead to impairments in the endothelial functions in the resistance arteries and the aorta in mice [118, 176, 177]. Increasing klotho has been associated with improving the antioxidant protective defensive mechanisms in rats and mice, thus potentially contributing to the ameliorations in endothelial function [118, 178]. In addition, in Otsuka Long-Evans Tokshuma fatty rats that embark important risk factors associated with atherosclerosis such as obesity, hyperglycemia, hypertriglyceridemia, and hypertension, adenovirus-mediated klotho gene delivery was reported to improve the dysfunction of the vascular endothelium, elevate the production of nitric oxide, decrease heightened blood pressures, and contribute to the prevention of medial hypertrophy and perivascular fibrosis [179]. Of note, the blood pressure effects of klotho gene delivery have not been reliable and consistent throughout the literature [118, 178].

A few reviews have also emphasized the interactions and the interdependency between klotho and the mTOR, AMPK, and SIRT1 pathways as they all play a part in vascular aging [116, 118]. Furthermore, Klotho was reported to be an activator of Nrf2 in several preclinical studies [175, 180–183]. This activation potentially contributes to the prevention of kidney and vascular diseases [175].

Given the potential detrimental effects associated with low klotho, an important therapeutic goal would be to reverse this situation and increase its levels. Several studies have shown that a safe target would be to revert klotho back within or near normal values instead of increasing its levels more than the normal range [175]. It is also worth mentioning that the majority of information regarding the therapies surrounding klotho were derived from rodent-based studies [175].

In a recent review, Prud'homme *et al.* summarized the current clinical drugs and those under development as well as the supplements and several other therapeutic mechanisms that increase klotho levels [175]. Among the currently available medications, RAAS inhibitors (losartan, valsartan), statins (atorvastatin, pitavastatin, simvastatin fluvastatin), peroxisome proliferator-activated receptor- γ (PPAR- γ) agonists (rosiglitazone, ciglitazone, pioglitazone), mechanistic target of rapamycin (mTOR) inhibitors (rapamycin, everolimus), vitamin D, glucagon-like peptide-1 (GLP-1) receptor agonist

(exendin-4) and dipeptidyl peptidase-4 (DPP-4) inhibitors (linagliptin, sitagliptin, vildagliptin), metformin, pentoxifylline, antiplasmodial (dihydroartemisinin), and endothelin-1 receptor antagonist (Atrasentan) have all been shown to elevate klotho in a variety of conditions in different studies [175]. Moreover, a recent study investigated the cardiorenal protective role of sodium glucose co-transporter-2 inhibitors (SGLT2i) in patients with diabetic kidney disease. The group receiving SGLT2i had statistically increased serum klotho levels. The same study also found that SGLT2i prevented klotho decrease due to high glucose concentrations in cultured proximal tubular cells [184].

Future large-scale studies are needed to better understand the role of these drugs on klotho and how the potential increase of this molecule contributes to the clinical effects of these drugs. Among the available experimental therapies, recombinant klotho and gene therapy strategies are promising approaches [175]. However further studies are required to better evaluate these therapeutic mechanisms and transition their use toward the clinics.

CONCLUSION

CKD is an important cause of premature, accelerated aging and can result in early vascular aging together with the stiffening of the arteries. The current knowns and unknowns surrounding early aging and premature vascular aging are summarized in Table 1.

Several underlying mechanisms such as oxidative stress, inflammation, advanced glycation end products, fructose, and an aberrant gut microbiota can contribute to the development of early aging in CKD patients. Premature vascular aging in CKD patients has been considered as a prognostic marker of mortality and cardiovascular morbidity. There are several key molecular pathways and molecules that play a role aging and vascular aging. Potential therapeutic strategies can target these pathways. Future studies are needed to better understand the importance of premature aging and early vascular aging to develop therapeutic alternatives for these conditions.

ACKNOWLEDGEMENTS

Figures in this review are made with the use of app.biorender.com.

DATA AVAILABILITY STATEMENT

No new data were generated or analyzed in support of this research.

FUNDING

This study was not funded by any grant.

CONFLICT OF INTEREST STATEMENT

M.C. is the Ad Interim Editor-in-Chief of CKJ. M.K. is member of the CKJ editorial board.

ETHICAL APPROVAL

This article does not contain any studies with human participants or animals performed by any of the authors.

AUTHORS' CONTRIBUTIONS

• Contributed substantially to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work: Cem Tanriover, Sidar Copur, Mehmet Kanbay, Ali Mutlu, Ibrahim Batuhan Peltek.

• Drafted the work or revised it critically for important intellectual content: Mehmet Kanbay, Mario Cozzolino, Andrea Galassi, Paola Ciceri.

REFERENCES

- Andrassy KM. Comments on 'KDIGO 2012 Clinical Practice Guideline for the evaluation and management of chronic kidney disease'. *Kidney Int* 2013;84:622–3. https://doi.org/10. 1038/ki.2013.243
- Charles C, Ferris AH. Chronic kidney disease. Prim Care 2020;47:585–95. https://doi.org/10.1016/j.pop.2020.08.001
- Kooman JP, Kotanko P, Schols AM et al. Chronic kidney disease and premature ageing. Nat Rev Nephrol 2014;10:732–42. https://doi.org/10.1038/nrneph.2014.185
- MacRae C, Mercer SW, Guthrie B et al. Comorbidity in chronic kidney disease: a large cross-sectional study of prevalence in Scottish primary care. Br J Gen Pract 2021;71:e243–9. https://doi.org/10.3399/bjgp20X714125
- Gadecka A, Bielak-Zmijewska. A Slowing down ageing: the role of nutrients and microbiota in modulation of the epigenome. Nutrients 2019;11:1251. https://doi.org/10.3390/ nu11061251
- Gutierrez M, Tomas JM, Calatayud P. Contributions of psychosocial factors and physical activity to successful aging. Span J Psychol 2018;21:E26. https://doi.org/10.1017/sjp.2018. 27
- Hamczyk MR, Nevado RM, Barettino A et al. Biological versus chronological aging: JACC focus seminar. J Am Coll Cardiol 2020;75:919–30. https://doi.org/10.1016/j.jacc.2019. 11.062
- Green S, Hillersdal L. Aging biomarkers and the measurement of health and risk. Hist Philos Life Sci 2021;43:28. https://doi.org/10.1007/s40656-021-00367-w
- London GM. Arterial stiffness in chronic kidney disease and end-stage renal disease. Blood Purif 2018;45:154–8. https://doi.org/10.1159/000485146
- Blacher J, Guerin AP, Pannier B et al. Impact of aortic stiffness on survival in end-stage renal disease. Circulation 1999;99:2434–9. https://doi.org/10.1161/01.CIR.99.18. 2434
- Laurent S, Boutouyrie P, Asmar R et al. Aortic stiffness is an independent predictor of all-cause and cardiovascular mortality in hypertensive patients. *Hypertension* 2001;37:1236–41. https://doi.org/10.1161/01.HYP.37.5.1236
- Cruickshank K, Riste L, Anderson SG et al. Aortic pulsewave velocity and its relationship to mortality in diabetes and glucose intolerance: an integrated index of vascular function? Circulation 2002;106:2085–90. https://doi.org/10. 1161/01.CIR.0000033824.02722.F7
- Kanbay M, Solak Y, Siriopol D et al. Sclerostin, cardiovascular disease and mortality: a systematic review and metaanalysis. Int Urol Nephrol 2016;48:2029–42. https://doi.org/ 10.1007/s11255-016-1387-8
- Libertini G. Aging definition. In: Gu D, Dupre ME, (eds), Encyclopedia of Gerontology and Population Aging. Chamonix: Springer International Publishing; 2019, 1–10.

- 15. Pal S, Tyler JK. Epigenetics and aging. Sci Adv 2016;2:e1600584. https://doi.org/10.1126/sciadv.1600584
- Crowson CS, Liang KP, Therneau TM et al. Could accelerated aging explain the excess mortality in patients with seropositive rheumatoid arthritis? Arthritis Rheum 2010;62:378–82.
- Pathai S, Lawn SD, Gilbert CE et al. Accelerated biological ageing in HIV-infected individuals in South Africa: a case-control study. AIDS 2013;27:2375–84. https://doi.org/ 10.1097/QAD.0b013e328363bf7f
- von Haehling S, Steinbeck L, Doehner W et al. Muscle wasting in heart failure: an overview. Int J Biochem Cell Biol 2013;45:2257–65. https://doi.org/10.1016/j.biocel.2013. 04.025
- Langen RC, Gosker HR, Remels AH et al. Triggers and mechanisms of skeletal muscle wasting in chronic obstructive pulmonary disease. Int J Biochem Cell Biol 2013;45:2245–56. https://doi.org/10.1016/j.biocel.2013.06.015
- 20. Kooman JP, Broers NJ, Usvyat L et al. Out of control: accelerated aging in uremia. Nephrol Dial Transplant 2013;**28**:48–54. https://doi.org/10.1093/ndt/gfs451
- 21. Dai L, Qureshi AR, Witasp A *et al*. Early vascular ageing and cellular senescence in chronic kidney disease. *Comput Struct Biotechnol J* 2019;17:721–9. https://doi.org/10.1016/ j.csbj.2019.06.015
- 22. Marquez-Exposito L, Tejedor-Santamaria L, Valentijn FA et al. Oxidative stress and cellular senescence are involved in the aging kidney. Antioxidants (Basel) 2022;**11**.
- 23. Stevenson KS, Radhakrishnan K, Patterson CS et al. Breath ethane peaks during a single haemodialysis session and is associated with time on dialysis. J Breath Res 2008;2:026004. https://doi.org/10.1088/1752-7155/2/2/026004
- 24. Himmelfarb J, Stenvinkel P, Ikizler TA et al. The elephant in uremia: oxidant stress as a unifying concept of cardiovascular disease in uremia. *Kidney Int* 2002;**62**:1524–38. https://doi.org/10.1046/j.1523-1755.2002.00600.x
- Gomez LA, Hagen TM. Age-related decline in mitochondrial bioenergetics: does supercomplex destabilization determine lower oxidative capacity and higher superoxide production? Semin Cell Dev Biol 2012;23:758–67. https://doi. org/10.1016/j.semcdb.2012.04.002
- Ciceri P, Cozzolino M. The emerging role of iron in heart failure and vascular calcification in CKD. Clin Kidney J 2021;14:739–45. https://doi.org/10.1093/ckj/sfaa135
- Mutsaers HA, Wilmer MJ, Reijnders D et al. Uremic toxins inhibit renal metabolic capacity through interference with glucuronidation and mitochondrial respiration. Biochim Biophys Acta 2013;1832:142–50. https://doi.org/10. 1016/j.bbadis.2012.09.006
- Hayflick L. The limited in vitro lifetime of human diploid cell strains. Exp Cell Res 1965;37:614–36. https://doi.org/10. 1016/0014-4827(65)90211-9
- Campisi J, d'Adda di Fagagna F. Cellular senescence: when bad things happen to good cells. Nat Rev Mol Cell Biol 2007;8:729–40. https://doi.org/10.1038/nrm2233
- Huang W, Hickson LJ, Eirin A et al. Cellular senescence: the good, the bad and the unknown. Nat *Rev* Nephrol 2022;18:611–27. https://doi.org/10.1038/ s41581-022-00601-z
- McHugh D, Gil J. Senescence and aging: causes, consequences, and therapeutic avenues. J Cell Biol 2018;217:65– 77. https://doi.org/10.1083/jcb.201708092
- Kudlova N, De Sanctis JB, Hajduch M. Cellular senescence: molecular targets, biomarkers, and senolytic drugs. Int J Mol Sci 2022;23:4168. https://doi.org/10.3390/ijms23084168

- Coppe JP, Patil CK, Rodier F et al. Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. PLoS Biol 2008;6:2853–68. https://doi.org/10.1371/journal. pbio.0060301
- 34. Jang JH, Chand HS, Bruse S et al. Connective tissue growth factor promotes pulmonary epithelial cell senescence and is associated with COPD severity. COPD 2017;14:228–37. https://doi.org/10.1080/15412555.2016.1262340
- Burton DGA, Stolzing A. Cellular senescence: immunosurveillance and future immunotherapy. Ageing Res Rev 2018;43:17–25. https://doi.org/10.1016/j.arr.2018.02. 001
- Wang WJ, Cai GY, Chen XM. Cellular senescence, senescence-associated secretory phenotype, and chronic kidney disease. Oncotarget 2017;8:64520–33. https://doi.org/10.18632/oncotarget.17327
- Sturmlechner I, Durik M, Sieben CJ et al. Cellular senescence in renal ageing and disease. Nat Rev Nephrol 2017;13:77–89. https://doi.org/10.1038/nrneph.2016.183
- Santoro A, Bientinesi E, Monti D. Immunosenescence and inflammaging in the aging process: age-related diseases or longevity? Ageing Res Rev 2021;71:101422. https://doi.org/ 10.1016/j.arr.2021.101422
- Pawelec G. Age and immunity: what is "immunosenescence"? Exp Gerontol 2018;105:4–9. https://doi.org/10.1016/ j.exger.2017.10.024
- Franceschi C, Bonafe M, Valensin S et al. Inflamm-aging. An evolutionary perspective on immunosenescence. Ann N Y Acad Sci 2000;908:244–54. https://doi.org/10.1111/j. 1749-6632.2000.tb06651.x
- Ferrucci L, Fabbri E. Inflammageing: chronic inflammation in ageing, cardiovascular disease, and frailty. Nat Rev Cardiol 2018;15:505–22. https://doi.org/10.1038/ s41569-018-0064-2
- 42. Kato S, Chmielewski M, Honda H et al. Aspects of immune dysfunction in end-stage renal disease. *Clin J Am Soc Nephrol* 2008;**3**:1526–33. https://doi.org/10.2215/CJN.00950208
- Betjes MG, Meijers RW, Litjens NH Loss of renal function causes premature aging of the immune system. Blood Purif 2013;36:173–8. https://doi.org/10.1159/000356084
- 44. McGuinness D, McGlynn LM, Johnson PC et al. Socioeconomic status is associated with epigenetic differences in the pSoBid cohort. Int J Epidemiol 2012;41:151–60. https://doi.org/10.1093/ije/dyr215
- Carrero JJ, Stenvinkel P. Inflammation in end-stage renal disease—what have we learned in 10 years? Semin Dial 2010;23:498–509. https://doi.org/10.1111/j.1525-139X.2010. 00784.x
- Meuwese CL, Carrero JJ, Stenvinkel P. Recent insights in inflammation-associated wasting in patients with chronic kidney disease. Contrib Nephrol 2011;171:120–6. https://doi. org/10.1159/000327228
- Shanahan CM. Mechanisms of vascular calcification in CKD-evidence for premature ageing? Nat Rev Nephrol 2013;9:661–70. https://doi.org/10.1038/nrneph.2013.176
- Matsuoka S, Yamashiro T, Diaz A et al. The relationship between small pulmonary vascular alteration and aortic atherosclerosis in chronic obstructive pulmonary disease: quantitative CT analysis. Acad Radiol 2011;18:40–46. https://doi.org/10.1016/j.acra.2010.08.013
- Paccou J, Brazier M, Mentaverri R et al. Vascular calcification in rheumatoid arthritis: prevalence, pathophysiological aspects and potential targets. Atherosclerosis 2012;224:283–90. https://doi.org/10.1016/j.atherosclerosis.2012.04.008

- Cozzolino M, Ciceri P. Ectopic calcification in Uremia: where do we stand? Blood Purif 2020;49:641–2. https://doi.org/10. 1159/000506178
- Ciceri P, Artioli L, Magagnoli L et al. The role of uremic retention solutes in the MIA syndrome in hemodialysis subjects. Blood Purif 2022;1–13.
- Ciceri P, Tettamanti G, Galassi A et al. Pro-calcifying analysis of uraemic serum from patients treated with medium cut-off membrane in a prospective, cross-over study. Clin Kidney J 2021;14:1798–807. https://doi.org/10. 1093/ckj/sfaa216
- Ejaz AA, Nakagawa T, Kanbay M et al. Hyperuricemia in kidney disease: a major risk factor for cardiovascular events, vascular calcification, and renal damage. Semin Nephrol 2020;40:574–85. https://doi.org/10.1016/j.semnephrol.2020. 12.004
- Kanbay M, Yerlikaya A, Sag AA et al. A journey from microenvironment to macroenvironment: the role of metaflammation and epigenetic changes in cardiorenal disease. Clin Kidney J 2019;12:861–70. https://doi.org/10. 1093/ckj/sfz106
- Cupisti A, Bolasco P, D'Alessandro C et al. Protection of residual renal function and nutritional treatment: first step strategy for reduction of uremic toxins in end-stage kidney disease patients. Toxins (Basel) 2021;13:289. https://doi.org/ 10.3390/toxins13040289
- Sabatino A, Regolisti G, Brusasco I et al. Alterations of intestinal barrier and microbiota in chronic kidney disease. Nephrol Dial Transplant 2015;30:924–33. https://doi.org/10. 1093/ndt/gfu287
- Tanriover C, Ucku D, Basile C et al. On the importance of the interplay of residual renal function with clinical outcomes in end-stage kidney disease. J Nephrol 2022;35:2191– 204. https://doi.org/10.1007/s40620-022-01388-9
- Vanholder R, Schepers E, Pletinck A et al. The uremic toxicity of indoxyl sulfate and p-cresyl sulfate: a systematic review. J Am Soc Nephrol 2014;25:1897–907. https://doi.org/ 10.1681/ASN.2013101062
- Louis P, Flint HJ. Formation of propionate and butyrate by the human colonic microbiota. Environ Microbiol 2017;19:29–41. https://doi.org/10.1111/1462-2920.13589
- Arpaia N, Campbell C, Fan X et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. Nature 2013;504:451–5. https://doi.org/10.1038/ nature12726
- Meijers B, Farre R, Dejongh S et al. Intestinal barrier function in chronic kidney disease. Toxins (Basel) 2018;10:298. https://doi.org/10.3390/toxins10070298
- Stinghen AE, Massy ZA, Vlassara H et al. Uremic toxicity of advanced glycation end products in CKD. J Am Soc Nephrol 2016;27:354–70. https://doi.org/10.1681/ASN.2014101047
- 63. Hariharan D, Vellanki K, Kramer H. The western diet and chronic kidney disease. Curr Hypertens Rep 2015;17:16. https://doi.org/10.1007/s11906-014-0529-6
- 64. Fotheringham AK, Gallo LA, Borg DJ et al. Advanced glycation end products (AGEs) and chronic kidney disease: does the modern diet AGE the kidney? Nutrients 2022;14:2675. https://doi.org/10.3390/nu14132675
- Rabbani N, Thornalley PJ. Advanced glycation end products in the pathogenesis of chronic kidney disease. *Kidney Int* 2018;93:803–13. https://doi.org/10.1016/j.kint.2017.11.034
- 66. Linden E, Cai W, He JC *et al*. Endothelial dysfunction in patients with chronic kidney disease results from advanced glycation end products (AGE)-mediated inhibition of en-

dothelial nitric oxide synthase through RAGE activation. Clin J Am Soc Nephrol 2008;3:691–8. https://doi.org/10.2215/ CJN.04291007

- Chen J, Muntner P, Hamm LL et al. The metabolic syndrome and chronic kidney disease in U.S. adults. Ann Intern Med 2004;140:167–74. https://doi.org/10.7326/ 0003-4819-140-3-200402030-00007
- Khitan Z, Kim DH. Fructose: a key factor in the development of metabolic syndrome and hypertension. J Nutr Metab 2013;2013:682673. https://doi.org/10.1155/ 2013/682673
- Nakayama T, Kosugi T, Gersch M et al. Dietary fructose causes tubulointerstitial injury in the normal rat kidney. *Am J Physiol Renal Physiol* 2010;298:F712–720. https://doi.org/ 10.1152/ajprenal.00433.2009
- Cirillo P, Gersch MS, Mu W et al. Ketohexokinasedependent metabolism of fructose induces proinflammatory mediators in proximal tubular cells. J Am Soc Nephrol 2009;20:545–53. https://doi.org/10.1681/ASN.2008060576
- Levi B, Werman MJ. Long-term fructose consumption accelerates glycation and several age-related variables in male rats. J Nutr 1998;128:1442–9. https://doi.org/10.1093/jn/128.9.1442
- Kanbay M, Copur S, Tanriover C et al. The pathophysiology and management of vascular calcification in chronic kidney disease patients. Expert Rev Cardiovasc Ther 2023;21:75– 85. https://doi.org/10.1080/14779072.2023.2174525
- Nilsson PM, Boutouyrie P, Laurent S. Vascular aging: a tale of EVA and ADAM in cardiovascular risk assessment and prevention. Hypertension 2009;54:3–10. https://doi.org/ 10.1161/HYPERTENSIONAHA.109.129114
- 74. Cunha PG, Boutouyrie P, Nilsson PM et al. Early Vascular ageing (EVA): definitions and clinical applicability. Curr Hypertens Rev 2017;13:8–15. https://doi.org/10.2174/ 1573402113666170413094319
- Boutouyrie P, Bruno RM. The clinical significance and application of vascular stiffness measurements. Am J Hypertens 2019;32:4–11. https://doi.org/10.1093/ajh/hpy145
- Pannier B, Guerin AP, Marchais SJ et al. Stiffness of capacitive and conduit arteries: prognostic significance for end-stage renal disease patients. *Hypertension* 2005;45:592–6. https://doi.org/10.1161/01.HYP.0000159190.71253.c3
- Copur S, Ucku D, Cozzolino M et al. Hypoxia-inducible factor signaling in vascular calcification in chronic kidney disease patients. J Nephrol 2022;35:2205–13. https://doi.org/10. 1007/s40620-022-01432-8
- Sag AA, Covic A, London G et al. Clinical imaging of vascular disease in chronic kidney disease. Int Urol Nephrol 2016;48:827–37. https://doi.org/10.1007/s11255-016-1240-0
- 79. Bronas UG, Puzantian H, Hannan M. Cognitive impairment in chronic kidney disease: vascular milieu and the potential therapeutic role of exercise. Biomed Res Int 2017;2017:2726369. https://doi.org/10.1155/2017/2726369
- Briet M, Bozec E, Laurent S et al. Arterial stiffness and enlargement in mild-to-moderate chronic kidney disease. Kidney Int 2006;69:350–7. https://doi.org/10.1038/sj.ki. 5000047
- Ford ML, Tomlinson LA, Chapman TP et al. Aortic stiffness is independently associated with rate of renal function decline in chronic kidney disease stages 3 and 4. Hypertension 2010;55:1110–5. https://doi.org/10.1161/HYPERTENSIONAHA.109.143024
- 82. Ferreira JP, Girerd N, Pannier B et al. High pulse-wave velocity defines a very High cardiovascular risk cohort of

dialysis patients under age 60. Am J Nephrol 2017;45:72–81. https://doi.org/10.1159/000453338

- Foley RN, Parfrey PS, Sarnak MJ. Clinical epidemiology of cardiovascular disease in chronic renal disease. Am J Kidney Dis 1998;32:S112–119.https://doi.org/10.1053/ajkd.1998. v32.pm9820470
- Hayes JD, Dinkova-Kostova AT. The Nrf2 regulatory network provides an interface between redox and intermediary metabolism. Trends Biochem Sci 2014;39:199–218. https://doi.org/10.1016/j.tibs.2014.02.002
- Yu M, Xu M, Liu Y et al. Nrf2/ARE is the potential pathway to protect Sprague–Dawley rats against oxidative stress induced by quinocetone. *Regul Toxicol Pharm* 2013;66:279–85. https://doi.org/10.1016/j.yrtph.2013.04.005
- Cullinan SB, Gordan JD, Jin J et al. The Keap1-BTB protein is an adaptor that bridges Nrf2 to a Cul3-based E3 ligase: oxidative stress sensing by a Cul3-Keap1 ligase. Mol Cell Biol 2004;24:8477–86. https://doi.org/10.1128/MCB. 24.19.8477-8486.2004
- Kobayashi A, Kang M-I, Okawa H et al. Oxidative stress sensor Keap1 functions as an adaptor for Cul3-based E3 ligase to regulate proteasomal degradation of Nrf2. Mol Cell Biol 2004;24:7130–9. https://doi.org/10.1128/MCB.24.16. 7130-7139.2004
- Dinkova-Kostova AT, Holtzclaw WD, Kensler TW. The role of Keap1 in cellular protective responses. Chem Res Toxicol 2005;18:1779–91. https://doi.org/10.1021/tx050217c
- Granatiero V, Konrad C, Bredvik K et al. Nrf2 signaling links ER oxidative protein folding and calcium homeostasis in health and disease. Life Sci. Alliance 2019;2:e201900563. https://doi.org/10.26508/lsa.201900563
- 90. Rada P, Rojo AI, Evrard-Todeschi N et al. Structural and functional characterization of Nrf2 degradation by the glycogen synthase kinase 3/β-TrCP axis. Mol Cell Biol 2012;32:3486–99. https://doi.org/10.1128/MCB. 00180-12
- Kobayashi EH, Suzuki T, Funayama R et al. Nrf2 suppresses macrophage inflammatory response by blocking proinflammatory cytokine transcription. Nat Commun 2016;7:1– 14. https://doi.org/10.1038/ncomms11624
- 92. Teasdale JE, Hazell GG, Peachey AM et al. Cigarette smoke extract profoundly suppresses $TNF\alpha$ -mediated proinflammatory gene expression through upregulation of ATF3 in human coronary artery endothelial cells. Sci Rep 2017;7:1– 10. https://doi.org/10.1038/srep39945
- Calvert JW, Jha S, Gundewar S et al. Hydrogen sulfide mediates cardioprotection through Nrf2 signaling. Circ Res 2009;105:365–74. https://doi.org/10.1161/CIRCRESAHA.109. 199919
- Noel S, Martina MN, Bandapalle S et al. T lymphocytespecific activation of Nrf2 protects from AKI. J Am Soc Nephrol 2015;26:2989–3000. https://doi.org/10.1681/ASN. 2014100978
- 95. Chen DQ, Cao G, Chen H et al. Identification of serum metabolites associating with chronic kidney disease progression and anti-fibrotic effect of 5-methoxytryptophan. Nat Commun 2019;10:1476. https://doi.org/10.1038/s41467-019-09329-0
- 96. Thangapandiyan S, Ramesh M, Miltonprabu S et al. Sulforaphane potentially attenuates arsenic-induced nephrotoxicity via the PI3K/akt/Nrf2 pathway in albino Wistar rats. Environ Sci Pollut Res 2019;26:12247–63. https://doi.org/10.1007/s11356-019-04502-w

- 97. Stenvinkel P, Larsson TE. Chronic kidney disease: a clinical model of premature aging. Am J Kidney Dis 2013;62:339–51. https://doi.org/10.1053/j.ajkd.2012.11.051
- Liu C, Gidlund E-K, Witasp A et al. Reduced skeletal muscle expression of mitochondrial-derived peptides humanin and MOTS-C and Nrf2 in chronic kidney disease. Am J Physiol Renal Physiol 2019;317:F1122–31. https://doi.org/10.1152/ ajprenal.00202.2019
- Aghagolzadeh P, Radpour R, Bachtler M et al. Hydrogen sulfide attenuates calcification of vascular smooth muscle cells via KEAP1/NRF2/NQO1 activation. Atherosclerosis 2017;265:78–86. https://doi.org/10.1016/j.atherosclerosis. 2017.08.012
- 100. Ganster F, Burban M, De La Bourdonnaye M et al. Effects of hydrogen sulfide on hemodynamics, inflammatory response and oxidative stress during resuscitated hemorrhagic shock in rats. Crit Care 2010;14:1–11. https://doi.org/ 10.1186/cc9257
- 101. Hinoi E, Fujimori S, Wang L et al. Nrf2 negatively regulates osteoblast differentiation via interfering with Runx2-dependent transcriptional activation. J Biol Chem 2006;281:18015–24. https://doi.org/10.1074/jbc. M600603200
- Zhang P, Li Y, Du Y et al. Resveratrol ameliorated vascular calcification by regulating sirt-1 and Nrf2. Transplant Proc 2016;48:3378–86. https://doi.org/10.1016/j.transproceed. 2016.10.023
- 103. Yoo KD, Kang S, Choi Y et al. Sex, age, and the association of serum phosphorus with all-cause mortality in adults with normal kidney function. Am J Kidney Dis 2016;67:79– 88. https://doi.org/10.1053/j.ajkd.2015.06.027
- 104. Ha C-M, Park S, Choi Y-K et al. Activation of Nrf2 by dimethyl fumarate improves vascular calcification. Vasc Pharmacol 2014;63:29–36. https://doi.org/10.1016/j.vph.2014.06.007
- 105. Wei R, Enaka M, Muragaki Y. Activation of KEAP1/NRF2/P62 signaling alleviates high phosphate-induced calcification of vascular smooth muscle cells by suppressing reactive oxygen species production. Sci Rep 2019;9:1–13. https://doi. org/10.1038/s41598-019-46824-2
- 106. Yao L, Wang J, Tian BY et al. Retracted: Activation of the Nrf2-ARE Signaling Pathway Prevents Hyperphosphatemia-Induced Vascular Calcification by Inducing Autophagy in Renal Vascular Smooth Muscle Cells. Wiley Online Library; 2017;
- 107. Zhou L, Zhang H, Davies KJA et al. Aging-related decline in the induction of Nrf2-regulated antioxidant genes in human bronchial epithelial cells. Redox Biol 2018;14:35–40. https://doi.org/10.1016/j.redox.2017.08.014
- 108. Ungvari Z, Bailey-Downs L, Sosnowska D et al. Vascular oxidative stress in aging: a homeostatic failure due to dysregulation of NRF2-mediated antioxidant response. Am J Physiol Heart Circ Physiol 2011;301:H363–372. https://doi.org/10. 1152/ajpheart.01134.2010
- 109. Gounder SS, Kannan S, Devadoss D et al. Impaired transcriptional activity of Nrf2 in age-related myocardial oxidative stress is reversible by moderate exercise training. PLoS ONE 2012;7:e45697. https://doi.org/10.1371/journal. pone.0045697
- 110. Fulop GA, Kiss T, Tarantini S et al. Nrf2 deficiency in aged mice exacerbates cellular senescence promoting cerebrovascular inflammation. *Geroscience* 2018;40:513–21. https://doi.org/10.1007/s11357-018-0047-6
- 111. Kuosmanen SM, Sihvola V, Kansanen E et al. MicroRNAs mediate the senescence-associated decline of NRF2 in

endothelial cells. Redox Biol 2018;18:77–83. https://doi.org/ 10.1016/j.redox.2018.06.007

- 112. Stenvinkel P, Luttropp K, McGuinness D et al. CDKN2A/p16INK4(a) expression is associated with vascular progeria in chronic kidney disease. Aging (Albany NY) 2017;**9**:494–507. https://doi.org/10.18632/aging.101173
- 113. Hong DS, Kurzrock R, Supko JG et al. A phase I firstin-human trial of bardoxolone methyl in patients with advanced solid tumors and lymphomas. Clin Cancer Res 2012;**18**:3396–406. https://doi.org/10.1158/1078-0432. CCR-11-2703
- 114. Gold R, Arnold DL, Bar-Or A et al. Long-term effects of delayed-release dimethyl fumarate in multiple sclerosis: interim analysis of ENDORSE, a randomized extension study. Mult Scler 2017;23:253–65. https://doi.org/10.1177/ 1352458516649037
- 115. Zeng L, Chen R, Liang F et al. Silent information regulator, Sirtuin 1, and age-related diseases. Geriatr Gerontol Int 2009;9:7–15. https://doi.org/10.1111/j.1447-0594.2008. 00504.x
- Donato AJ, Morgan RG, Walker AE et al. Cellular and molecular biology of aging endothelial cells. J Mol Cell Cardiol 2015;89:122–35. https://doi.org/10.1016/j.yjmcc.2015.01.021
- 117. Canto C, Jiang LQ, Deshmukh AS et al. Interdependence of AMPK and SIRT1 for metabolic adaptation to fasting and exercise in skeletal muscle. *Cell Metab* 2010;**11**:213–9. https://doi.org/10.1016/j.cmet.2010.02.006
- 118. Donato AJ, Machin DR, Lesniewski LA. Mechanisms of dysfunction in the aging vasculature and role in age-related disease. Circ Res 2018;123:825–48. https://doi.org/10.1161/ CIRCRESAHA.118.312563
- 119. Canto C, Gerhart-Hines Z, Feige JN et al. AMPK regulates energy expenditure by modulating NAD+ metabolism and SIRT1 activity. Nature 2009;458:1056–60. https://doi.org/10. 1038/nature07813
- 120. Shackelford DB, Shaw RJ. The LKB1-AMPK pathway: metabolism and growth control in tumour suppression. Nat Rev Cancer 2009;9:563–75. https://doi.org/10.1038/ nrc2676
- 121. Greer EL, Oskoui PR, Banko MR et al. The energy sensor AMP-activated protein kinase directly regulates the mammalian FOXO3 transcription factor. J Biol Chem 2007;282:30107–19. https://doi.org/10.1074/jbc. M705325200
- 122. Jager S, Handschin C, St-Pierre J et al. AMP-activated protein kinase (AMPK) action in skeletal muscle via direct phosphorylation of PGC-1alpha. Proc Natl Acad Sci U S A 2007;104:12017–22. https://doi.org/10.1073/pnas. 0705070104
- 123. Hardie DG, Hawley SA, Scott JW. AMP-activated protein kinase—development of the energy sensor concept. J Physiol 2006;574:7–15. https://doi.org/10.1113/jphysiol. 2006.108944
- 124. Fisslthaler B, Fleming I. Activation and signaling by the AMP-activated protein kinase in endothelial cells. Circ Res 2009;105:114–27. https://doi.org/10.1161/CIRCRESAHA.109. 201590
- 125. Lesniewski LA, Zigler MC, Durrant JR et al. Sustained activation of AMPK ameliorates age-associated vascular endothelial dysfunction via a nitric oxide-independent mechanism. Mech Ageing Dev 2012;133:368–71. https://doi. org/10.1016/j.mad.2012.03.011

- 126. Pu Y, Zhang H, Wang P et al. Dietary curcumin ameliorates aging-related cerebrovascular dysfunction through the AMPK/uncoupling protein 2 pathway. Cell Physiol Biochem 2013;32:1167–77. https://doi.org/10.1159/000354516
- 127. Sartoretto JL, Melo GA, Carvalho MH et al. Metformin treatment restores the altered microvascular reactivity in neonatal streptozotocin-induced diabetic rats increasing NOS activity, but not NOS expression. Life Sci 2005;77:2676– 89. https://doi.org/10.1016/j.lfs.2005.05.022
- Katakam PV, Ujhelyi MR, Hoenig M et al. Metformin improves vascular function in insulin-resistant rats. Hypertension 2000;35:108–12. https://doi.org/10.1161/01.HYP.35.1.
- 129. Agarwal N, Rice SP, Bolusani H et al. Metformin reduces arterial stiffness and improves endothelial function in young women with polycystic ovary syndrome: a randomized, placebo-controlled, crossover trial. J Clin Endocrinol Metab 2010;95:722–30. https://doi.org/10.1210/jc.2009-1985
- 130. de Kreutzenberg SV, Ceolotto G, Cattelan A et al. Metformin improves putative longevity effectors in peripheral mononuclear cells from subjects with prediabetes. A randomized controlled trial. Nutr Metab Cardiovasc Dis 2015;25:686–93. https://doi.org/10.1016/j.numecd.2015. 03.007
- Boily G, Seifert EL, Bevilacqua L et al. SirT1 regulates energy metabolism and response to caloric restriction in mice. PLoS ONE 2008;3:e1759. https://doi.org/10.1371/ journal.pone.0001759
- 132. Bordone L, Cohen D, Robinson A et al. SIRT1 transgenic mice show phenotypes resembling calorie restriction. Aging Cell 2007;6:759–67. https://doi.org/10.1111/j.1474-9726. 2007.00335.x
- Pillarisetti S. A review of Sirt1 and Sirt1 modulators in cardiovascular and metabolic diseases. Recent Pat Cardiovasc Drug Discov 2008;3:156–64. https://doi.org/10.2174/ 157489008786263989
- 134. Yeung F, Hoberg JE, Ramsey CS et al. Modulation of NFkappaB-dependent transcription and cell survival by the SIRT1 deacetylase. EMBO J 2004;23:2369–80. https://doi.org/ 10.1038/sj.emboj.7600244
- 135. Rahman S, Islam R. Mammalian Sirt1: insights on its biological functions. Cell Commun Signal 2011;9:11. https://doi. org/10.1186/1478-811X-9-11
- 136. Yan J, Wang J, He JC et al. Sirtuin 1 in chronic kidney disease and therapeutic potential of targeting Sirtuin 1. Front Endocrinol (Lausanne) 2022;13:917773. https://doi.org/10.3389/ fendo.2022.917773
- 137. Li P, Liu Y, Qin X et al. SIRT1 attenuates renal fibrosis by repressing HIF-2alpha. Cell Death Discov 2021;7:59. https:// doi.org/10.1038/s41420-021-00443-x
- 138. Vasko R, Xavier S, Chen J et al. Endothelial sirtuin 1 deficiency perpetrates nephrosclerosis through downregulation of matrix metalloproteinase-14: relevance to fibrosis of vascular senescence. J Am Soc Nephrol 2014;25:276–91. https://doi.org/10.1681/ASN.2013010069
- 139. Huang XZ, Wen D, Zhang M et al. Sirt1 activation ameliorates renal fibrosis by inhibiting the TGF-beta/Smad3 pathway. J Cell Biochem 2014;115:996–1005. https://doi.org/ 10.1002/jcb.24748
- 140. Hong YA, Kim JE, Jo M et al. The role of sirtuins in kidney diseases. Int J Mol Sci 2020;21:6686. https://doi.org/10.3390/ ijms21186686

- 141. Donato AJ, Magerko KA, Lawson BR et al. SIRT-1 and vascular endothelial dysfunction with ageing in mice and humans. J Physiol 2011;**589**:4545–54. https://doi.org/10.1113/ jphysiol.2011.211219
- 142. Donato AJ, Walker AE, Magerko KA et al. Life-long caloric restriction reduces oxidative stress and preserves nitric oxide bioavailability and function in arteries of old mice. Aging Cell 2013;12:772–83. https://doi.org/10.1111/acel.12103
- 143. Rippe C, Lesniewski L, Connell M et al. Short-term calorie restriction reverses vascular endothelial dysfunction in old mice by increasing nitric oxide and reducing oxidative stress. Aging Cell 2010;9:304–12. https://doi.org/10.1111/ j.1474-9726.2010.00557.x
- 144. Gano LB, Donato AJ, Pasha HM et al. The SIRT1 activator SRT1720 reverses vascular endothelial dysfunction, excessive superoxide production, and inflammation with aging in mice. Am J Physiol Heart Circ Physiol 2014;**307**:H1754–1763. https://doi.org/10.1152/ajpheart.00377.2014
- 145. Guo Y, Xu A, Wang Y. SIRT1 in endothelial cells as a novel target for the prevention of early vascular aging. J Cardiovasc Pharmacol 2016;**67**:465–73. https://doi.org/10.1097/FJC. 000000000000344
- 146. Bai B, Vanhoutte PM, Wang Y. Loss-of-SIRT1 function during vascular ageing: hyperphosphorylation mediated by cyclin-dependent kinase 5. Trends Cardiovasc Med 2014;24:81–84. https://doi.org/10.1016/j.tcm.2013.07. 001
- 147. Begum MK, Konja D, Singh S et al. Endothelial SIRT1 as a target for the prevention of arterial aging: promises and challenges. J Cardiovasc Pharmacol 2021;78:S63–77. https://doi.org/10.1097/FJC.00000000001154
- 148. Chen J, Xavier S, Moskowitz-Kassai E et al. Cathepsin cleavage of sirtuin 1 in endothelial progenitor cells mediates stress-induced premature senescence. Am J Pathol 2012;180:973–83. https://doi.org/10.1016/j.ajpath.2011.11. 033
- 149. Han Y, Kim SY. Endothelial senescence in vascular diseases: current understanding and future opportunities in senotherapeutics. Exp Mol Med 2023;55:1–12. https://doi. org/10.1038/s12276-022-00906-w
- 150. Takemura A, Iijima K, Ota H et al. Sirtuin 1 retards hyperphosphatemia-induced calcification of vascular smooth muscle cells. Arterioscler Thromb Vasc Biol 2011;31:2054–62. https://doi.org/10.1161/ATVBAHA.110. 216739
- 151. Akiyoshi T, Ota H, Iijima K et al. A novel organ culture model of aorta for vascular calcification. Atherosclerosis 2016;244:51–58. https://doi.org/10.1016/j.atherosclerosis. 2015.11.005
- 152. Bartoli-Leonard F, Wilkinson FL, Schiro A et al. Suppression of SIRT1 in diabetic conditions induces osteogenic differentiation of human vascular smooth muscle cells via RUNX2 signalling. Sci Rep 2019;9:878. https://doi.org/10. 1038/s41598-018-37027-2
- 153. Badi I, Mancinelli L, Polizzotto A et al. miR-34a promotes vascular smooth muscle cell calcification by downregulating SIRT1 (Sirtuin 1) and Axl (AXL Receptor Tyrosine Kinase). Arterioscler Thromb Vasc Biol 2018;38:2079–90. https: //doi.org/10.1161/ATVBAHA.118.311298
- 154. Ciccone L, Piragine E, Brogi S et al. Resveratrol-like compounds as SIRT1 activators. Int J Mol Sci 2022;**23**:15105. https://doi.org/10.3390/ijms232315105
- 155. Das DK, Maulik N. Resveratrol in cardioprotection: a therapeutic promise of alternative medicine. Mol Interv 2006;6:36–47. https://doi.org/10.1124/mi.6.1.7

- 156. Stivala LA, Savio M, Carafoli F et al. Specific structural determinants are responsible for the antioxidant activity and the cell cycle effects of resveratrol. J Biol Chem 2001;**276**:22586–94. https://doi.org/10.1074/jbc.M101846200
- 157. Labinskyy N, Csiszar A, Veress G et al. Vascular dysfunction in aging: potential effects of resveratrol, an antiinflammatory phytoestrogen. Curr Med Chem 2006;13:989– 96. https://doi.org/10.2174/092986706776360987
- 158. Pearson KJ, Baur JA, Lewis KN et al. Resveratrol delays agerelated deterioration and mimics transcriptional aspects of dietary restriction without extending life span. Cell Metab 2008;8:157–68. https://doi.org/10.1016/j.cmet.2008.06.011
- 159. Roggerio A, Strunz CMC, Pacanaro AP et al. Gene expression of sirtuin-1 and endogenous secretory receptor for advanced glycation end products in healthy and slightly overweight subjects after caloric restriction and resveratrol administration. Nutrients 2018;10:937. https://doi.org/10.3390/nu10070937
- Ebert T, Pawelzik SC, Witasp A et al. Inflammation and premature ageing in chronic kidney disease. Toxins (Basel) 2020;12:227. https://doi.org/10.3390/toxins12040227
- 161. Figuer A, Bodega G, Tato P et al. Premature aging in chronic kidney disease: the outcome of persistent inflammation beyond the bounds. Int J Environ Res Public Health 2021;18:8044. https://doi.org/10.3390/ijerph18158044
- Azpiazu D, Gonzalo S, Gonzalez-Parra E et al. Role of pyrophosphate in vascular calcification in chronic kidney disease. Nefrologia (Engl Ed) 2018;38:250–7. https://doi.org/10. 1016/j.nefroe.2018.03.003
- 163. Yamada S, Giachelli CM. Vascular calcification in CKD-MBD: roles for phosphate, FGF23, and klotho. Bone 2017;100:87–93. https://doi.org/10.1016/j.bone.2016.11.012
- 164. Donate-Correa J, Ferri CM, Martin-Nunez E et al. Klotho as a biomarker of subclinical atherosclerosis in patients with moderate to severe chronic kidney disease. Sci Rep 2021;11:15877. https://doi.org/10.1038/s41598-021-95488-4
- 165. Kuro-o M. Klotho and aging. Biochim Biophys Acta 2009;**1790**:1049–58. https://doi.org/10.1016/j.bbagen.2009. 02.005
- 166. Hu MC, Shi M, Zhang J et al. Klotho deficiency causes vascular calcification in chronic kidney disease. J Am Soc Nephrol 2011;22:124–36. https://doi.org/10.1681/ASN.2009121311
- 167. Mencke R, Hillebrands JL., consortium N. The role of the anti-ageing protein Klotho in vascular physiology and pathophysiology. Ageing Res Rev 2017;35:124–46. https://doi. org/10.1016/j.arr.2016.09.001
- Zhou H, Pu S, Zhou H et al. Klotho as potential autophagy regulator and therapeutic target. Front Pharmacol 2021;12:755366. https://doi.org/10.3389/fphar.2021.755366
- 169. Kuro-o M, Matsumura Y, Aizawa H et al. Mutation of the mouse klotho gene leads to a syndrome resembling ageing. Nature 1997;390:45–51. https://doi.org/10.1038/ 36285
- 170. Kurosu H, Yamamoto M, Clark JD et al. Suppression of aging in mice by the hormone Klotho. Science 2005;309:1829–33. https://doi.org/10.1126/science.1112766
- 171. Xie J, Cha SK, An SW et al. Cardioprotection by Klotho through downregulation of TRPC6 channels in the mouse heart. Nat Commun 2012;3:1238. https://doi.org/10.1038/ ncomms2240
- 172. Hsieh CC, Kuro-o M, Rosenblatt KP et al. The ASK1signalosome regulates p38 MAPK activity in response to levels of endogenous oxidative stress in the Klotho mouse models of aging. Aging (Albany NY) 2010;2:597–611. https://doi.org/10.18632/aging.100194

- 173. Semba RD, Cappola AR, Sun K et al. Plasma klotho and cardiovascular disease in adults. J Am Geriatr Soc 2011;**59**:1596–601. https://doi.org/10.1111/j.1532-5415.2011. 03558.x
- 174. Xiao NM, Zhang YM, Zheng Q et al. Klotho is a serum factor related to human aging. Chin Med J (Engl) 2004;117: 742–7.
- 175. Prud'homme GJ, Kurt M, Wang Q. Pathobiology of the Klotho antiaging protein and therapeutic considerations. Front Aging 2022;3:931331. https://doi.org/10.3389/ fragi.2022.931331
- 176. Gao D, Zuo Z, Tian J et al. Activation of SIRT1 attenuates Klotho deficiency-induced arterial stiffness and hypertension by enhancing AMP-activated protein kinase activity. Hypertension 2016;68:1191–9. https://doi.org/10.1161/ HYPERTENSIONAHA.116.07709
- 177. Saito Y, Yamagishi T, Nakamura T et al. Klotho protein protects against endothelial dysfunction. Biochem Biophys Res Commun 1998;248:324–9. https://doi.org/10.1006/bbrc.1998.
 8943
- 178. Ohta J, Rakugi H, Ishikawa K et al. Klotho gene delivery suppresses oxidative stress in vivo. *Geriatr Gerontol Int* 2007;7:293–9.
- 179. Saito Y, Nakamura T, Ohyama Y et al. In vivo klotho gene delivery protects against endothelial dysfunction in multiple risk factor syndrome. Biochem Biophys Res Com-

mun 2000;**276**:767–72. https://doi.org/10.1006/bbrc.2000. 3470

- 180. Maltese G, Psefteli PM, Rizzo B et al. The anti-ageing hormone klotho induces Nrf2-mediated antioxidant defences in human aortic smooth muscle cells. J Cell Mol Med 2017;21:621–7. https://doi.org/10.1111/jcmm.12996
- 181. Zhu H, Gao Y, Zhu S et al. Klotho improves cardiac function by suppressing reactive oxygen species (ROS) mediated apoptosis by modulating mapks/Nrf2 signaling in doxorubicin-induced cardiotoxicity. Med Sci Monit 2017;23:5283–93. https://doi.org/10.12659/MSM. 907449
- 182. Xiang T, Luo X, Ye L et al. Klotho alleviates NLRP3 inflammasome-mediated neuroinflammation in a temporal lobe epilepsy rat model by activating the Nrf2 signaling pathway. Epilepsy Behav 2022;128:108509. https://doi.org/10. 1016/j.yebeh.2021.108509
- 183. Xing L, Guo H, Meng S et al. Klotho ameliorates diabetic nephropathy by activating Nrf2 signaling pathway in podocytes. Biochem Biophys Res Commun 2021;534:450–6. https://doi.org/10.1016/j.bbrc.2020.11.061
- 184. Mora-Fernández C, Sánchez-Niño MD, Donate-Correa J et al. Sodium-glucose co-transporter-2 inhibitors increase Klotho in patients with diabetic kidney disease: a clinical and experimental study. Biomed Pharmacother 2022;154:113677.