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Original article

BLOOD FIBRINGEN CONCENTRATIONS IN NEW ZEALAND WHITE RABBITS DURING THE FIRST YEAR OF LIFE

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Summary

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The aim of this study was to examine the reference range of blood plasma fibrinogen, which belongs to the group of moderate acute phase protein in rabbits. The experiment followed the dynamics of blood plasma fibrinogen (Fb) during the first year of life in 12 New Zealand White rabbits, 6 males and 6 females, starting at 1 month of age until the 7th month. It was shown that fibrinogen concentrations increased with the age. There was a statistically significant difference in Fb concentrations in rabbits at 3 (P<0.05), 6 (P<0.01) and 7 months of age (P<0.001) compared to levels obtained at 1 month of age in both genders. At ages of 1, 2 and 3 months, fibrinogen values were higher in male rabbits while in 7-month-old rabbits, plasma fibrinogen levels in females were significantly higher than those in male rabbits (P<0.01).

Key words: acute phase proteins, fibrinogen, rabbits

INTRODUCTION

The acute phase response (APR) refers to a nonspecific and complex reaction of an animal that occurs shortly after any tissue injury (Kushner *et al.*, 2006). During this response, there is an increased production and release of certain proteins known as acute phase proteins (APPs), which can be produced by hepatocytes and peripheral tissues (Schmidt & Eckersall, 2015). APPs production and secretion by the liver, mainly of glycoproteins, is one of

mechanisms involved in the response to mediators produced by leukocytes and macrophages during episodes of infection or inflammation (Conner *et al.*, 1988). The maximum serum concentrations of APPs are typically attained within 24 to 48 h after the challenge (Jain *et al.*, 2011).

One classification divides APP into "positive" and "negative". Plasma proteins, whose levels increase in response to challenge are in the group of positive,

while those exhibiting a decrease – in the group of negative APPs. The positive APPs are glycoproteins synthesised into the bloodstream and include haptoglobin, C-reactive protein, serum amyloid A, ceruloplasmin, fibrinogen and alpha 1-acid glycoprotein (Murata *et al.*, 2004). The negative APPs comprise albumin – the most abundant constitutive plasma protein, and transferrin.

Kushner *et al.* (2006) classified positive APPs into 3 groups: 1) APPs whose concentration increase by 50% (ceruloplasmin and C_3 complement); 2) APP exhibiting 2- to 3-fold increase (haptoglobin, fibrinogen and α -albumins with antiprotease activity); and 3) proteins, increasing extremely rapidly up to 1000 times (C-reactive protein and serum amyloid A). The changes in plasma APPs concentrations are often used for diagnostic purposes (Marshall, 1994; Andonova, 2002).

In most species, the fibrinogen (Fb) belongs to the group of positive and moderate APP. It is involved in haemostasis, providing a substrate for fibrin formation, and tissue repair providing a matrix for the migration of inflammatory related cells (Thomas, 2000). Fb specifically binds to CD 11/CD 18 integrins on the cell surface of migrated phagocytes, thereby triggering a cascade of intracellular signals that lead to enhancement of degranulation, phagocytosis, antibody-dependent cellular cytotoxicity and delay of apoptosis (Rubel *et al.*, 2001).

In order to assess the usefulness of Fb as a member of the group of moderate APPs and as a measure of the acute phase response in rabbits, the reference concentration range of this marker should be determined in male and female New Zealand White rabbits during the first year of their life. In previous experiments of ours we have established that fibrinogen in-

creased 2 or 3 times after experimental infection with *E. coli* and *St. aureus*. (Mircheva *et al.*, 2009; Dishlyanova *et al.*, 2013). Therefore, we belelive that Fb could serve as a good indicator for the incidence of infection or inflammation. Moreover, the methods used for its determination are very inexpensive (the nephelometric method of Podmore; heat precipitation method), as compared to the determination of the major APPs SAA and CRP for rabbits, which are species-specific and very costly.

To the best of our knowledge there is no available information about the concentration of blood plasma fibrinogen in healthy male and female New Zealand White rabbits at different ages, except the data of Dishlyanova *et al.* (2014) in male and female rabbits at age of 1, 2 and 3 months. According to some directives of the Society of Clinical Pathology for scientific purposes the normal ranges of fibrinogen in rabbits should be preferably known for comparison with the values accompanying disease states with inflammation, including infection.

Therefore the aim of this study was to study the age-related and gender-related changes in plasma concentration of fibrinogen in rabbits during the first year of their life.

MATERIALS AND METHODS

The experimental procedure was approved by the Ethic Committee at the Faculty of Veterinary Medicine. The experiments were carried out on 12 New Zealand white rabbits divided in 2 groups of 6 rabbits in each: Ist group – 6 male and IInd group – 6 females, at the age of 1 month. All animals were born from healthy doe rabbits and kept in the rabbitry. They were fed

pelleted feed according to their age and had free access to tap water.

Blood samples for analysis of Fb were drawn with heparinised sterile tubes from *v. auricularis caudalis* from all rabbits at the age of 1, 2, 3, 6 and 7 months and immediately centrifuged (1,500 min⁻¹, 15 min, 4 °C) to obtain plasma.

Plasma Fb was determined immediately by the nephelometric method of Podmore with 10% Na₂SO₄ at a wavelength of 570 nm (Todorov, 1972).

The statistical analysis of the data was performed using SPSS 16.0 for Windows (SPSS Inc.). The concentration of Fb in male and female rabbits at different ages were compared with the concentrations at 1 month of age were assessed by paired t-test. The differences between genders were evaluated by the Student t-test. All data were expressed as mean \pm standard deviation (SD) and the differences were considered significant when P<0.05.

RESULTS

The time course of plasma Fb concentration is presented on Fig. 1. In both genders, plasma Fb values at 1 month of age were similar. Fb value in male rabbits at month 2 was 1.23 ± 0.26 g/L, at month 3 and 6 were 1.54 ± 0.39 ; 1.62 ± 0.58 g/L respectively and by the end of the experiment (age 7 months) -1.66 ± 0.39 g/L.

In female rabbits the mean Fb concentration at the first measurement was 0.46±0.13 g/L. At the age of 2 months, the Fb in female rabbits was 1.08±0.30 g/L and was further increased at month 3 (1.45±0.41 g/L). At the age of 6 and 7 months, the concentration of Fb in female rabbits were 1.68±0.23 and 2.34±0.65 g/L respectively.

Statistically significant difference was established at the age of 2 months (P<0.05) in male rabbits compared to 1 month of age. Substantial differences were present at 3 months of age (P<0.01) in both genders compared to the baseline measurement at month 1. The concentrations of Fb at the age of 6 and 7 months were statistically significantly (P<0.001) different compared to 1 month of age.

Statistically significant difference in Fb levels was established between genders at age of 7 month (P<0.01).

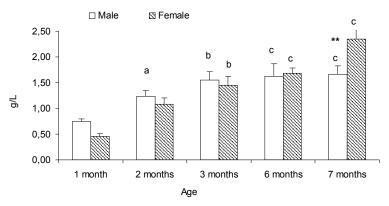


Fig. 1. Blood plasma fibrinogen concentrations in male and female New Zealand White rabbits at different ages. Data are presented as mean \pm SD (n=6). $^{a}P<0.05$, $^{b}P<0.01$, $^{c}P<0.001$ significant difference vs one month of age; ** P<0.01 significant difference between the genders.

DISCUSSION

The APPs are a group of blood proteins that contribute to restoring homeostasis and limiting microbial growth in an antibody-independent manner in animals with infection, inflammation, surgical trauma or stress (Murata *et al.*, 2004).

Fb and haptoglobin are affirmed to be "major" and "positive" APPs in rabbits (Petersen *et al.*, 2004), but the data about Fb indicating infection in rabbits are few (Kimura *et al.*, 1994; Mircheva *et al.*, 2009; Hana *et al.*, 2011; Dishlyanova *et al.*, 2013). Data about reference ranges of this parameter in the available literature are limited.

In several studies, a positive relationship was reported between the increase of the another acute phase protein: haptoglobin and aging in different animals species: pigs (Moya *et al.*, 2007); mice (Ding & Kopchik, 2011); rats (Ciftci *et al.*, 2012).

The study of Dishlyanova *et al.* (2014) provided proof for a positive relationship between blood Fb levels and aging in rabbits. The reported data, however, concerned only bunnies in the earliest period of their life – until 3 months of age.

The results from the present study provided the necessary and lacking information about blood plasma Fb concentrations in rabbits from both sexes according to their age until the 7th month of life.

Fb is used in cattle and sheep as a reliable indicator of the presence of inflammation, bacterial infection or surgical trauma (Pfeffer *et al.*, 1993; Hirvonen *et al.*, 1996; Cheryk *et al.*, 1998; Hirvonen & Pyorala, 1998). In horses, Fb has been used for the diagnosis of infections (Falcon *et al.*, 1985; Johnstone & Crane, 1986; Morris *et al.*, 1988; Held *et al.*, 1990). The increase of Fb associated with surgical castration of bulls seems to be independent of plasma cortisol concentra-

tions (Fisher et al., 1997). Hyperfibrinogenaemia has been induced by intramuscular injection of Freund's complete adjuvant (Mills et al., 1998). The increased Fb levels observed in pregnant dogs three weeks following implantation suggest that Fb may be useful for early pregnancy diagnosis in dogs (Concannon et al., 1996). Zapryanova et al. (2013) reported that the Fb can be useful for diagnosis and follow up of a bacterial infection in dogs. Fb is used as an early marker of acute inflammation of the mammary gland in goats caused by Staphyloccocus aureus (Fasulkov et al., 2014).

CONCLUSIONS

The results showed that plasma Fb concentration increased with age. There was a statistically significant difference in Fb concentrations in rabbits at 3 (P<0.05), 6 (P<0.01) and 7 months of age (P<0.001) compared to levels obtained at 1 month of age in both genders. During first 6 months of their life fibrinogen concentration in male and female rabbits were similar but there was a statistically significant difference in Fb concentration between genders at 7 months of age (P<0.01).

The present data could be useful to the researchers using rabbits as experimental animals as reference values for comparative purposes in the interpretation of the changes of this APP in different inflammatory conditions in rabbits.

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REFERENCES

- Andonova, M., 2002. Role of innate defense mechanisms in acute phase response against gram-negative agents. *Bulgarian Journal of Veterinary Medicine*, 5, 77–92.
- Cheryk, L. A., K. E. Hooper-Mcgrevy & P. A. Gentry, 1998. Alterations in bovine platelet function and acute phase proteins induced by *Pasteurella haemolytica* A1. Canadian Journal of Veterinary Research, 62, 1–8.
- Ciftci, G., G. F. Yarim, S. Cenesiz & A. Findik, 2012. Age-related changes in haptoglobin phenotypes, some non-enzymatic antioxidants and electrophoretic profiles of serum proteins in rats. *Turkish Journal of Biochemistry*, 37, 457–462.
- Concannon, P. W., T. Gimpel, L. Newton & V. D. Castracane, 1996. Postimplantation increase in plasma fibrinogen concentration with increase in relaxin concentration in pregnant dogs. American Journal of Veterinary Research, 57, 1382–1385.
- Conner, J. G. & P. D. Eckersall, 1988. Bovine and canine acute phase proteins. *Veterina*ry Research Communication, 12, 169–178.
- Ding, J. & J. J. Kopchick, 2011. Plasma biomarkers of mouse aging. Age (Dordrecht, Netherlands), 33, 291–307.
- Dishlyanova, E., T. Georgieva, T. Vlaykova & I. Georgiev, 2013. Effect of *Staphylococcus aureus* infection on fibrinogen and plasma protein concentrations in experimentally obese rabbits. Farm animal proteomics. Proceeding of 4th Management Committee Meeting and 3rd Meeting of Working Groups 1, 2 & 3 of COST Action FA 1002 Kosice, Slovakia, pp. 128–131.
- Dishlyanova, E., T. Mircheva, I. Penchev, V. Petrov, T. Vlaykova, F. Ceciliani & R. Vasileva, 2014. Blood fibrinogen concentration in New Zealand White rabbits during first three months of their life. Farm animal proteomics. Proceeding of the 5th Management Committee Meeting and 4th Meeting working Groups 1, 2 & 3 of

- COST Action FA 1002 Milano, Italy, pp. 217–219.
- Fasulkov, I., M. Karadaev, V. Urumova & T. Mirvheva, 2014. Determination of plasma fibrinogen and haptoglobin, hematological and blood biochemical changes in Bulgarian local goats with experimentally induced Staphyloccocus aureus mastitis. Turkish Journal of Veterinary and Animal Sciences, 38, 439–444.
- Falcon, J., B. P. Smith, T. R. O'Brien, G. P. Carlson & E. Biberstein, 1985. Clinical and radiographic findings in Coryne-bacterium equi pneumonia of foals. Journal of the American Veterinary Medical Association, 186, 593–599.
- Fisher, A. D., M. A. Crowe, E. M. O'Nuallain, M. L. Monaghan, J. A. Larkin, P. O'Kiely & W. J. Enright, 1997. Effects of cortisol on *in vitro* interferon-gamma production, acute-phase proteins, growth, and feed intake in a calf castration model. *Journal of Animal Science*, 75, 1041–1047.
- Hana, A., S. I. O. Salasia, S. Mangkoewidjojo & D. L. Kusindarto, 2011. Blood profile of rabbits infected with *Eimeria magna*. *Animal Production*, 13, 185–190.
- Held, J. P., S. Adair, M. D. McGavin, W. H. Adams, R. Toal & J. Henton, 1990. Bacterial epididymitis in two stallions. *Journal of the American Veterinary Medical Association*, 197, 602–604.
- Hirvonen, J. & S. Pyorala, 1998. Acute-phase response in dairy cows with surgicallytreated abdominal disorders. *The Veteri*nary Journal, 155, 53–61.
- Hirvonen, J., S. Pyorala & H. Jousimies-Somer, 1996. Acute phase response in heifers with experimentally induced mastitis. *The Journal of Dairy Research*, **63**, 351–360
- Jain, S., V. Gautam & S. Naseem, 2011. Acute-phase proteins: As diagnostic tool. Journal of Pharmacy And Bioallied Sciences, 3, 118–127.
- Johnstone, I. B. & S. Crane, 1986. Hemostatic abnormalities in equine colic. *American*

- Journal of Veterinary Research, 47, 356–358
- Kimura, M., L. A. Toth, H. Agostini, A. B. Cady, J. A. Majde & J. M. Krueger, 1994. Comparison of acute phase responses induced in rabbits by lipopolysaccharide and double-stranded RNA. *American Journal* of Physiology, 267, 1569–1605.
- Kushner, I., D. Rzewincki & D. Samols, 2006. What does minor elevation of C-reactive protein signify? The American Journal of Medicine, 119, 17–28.
- Marshall, W., 1994. Plasma proteins. In: *An Illustrated Textbook of Clinical Chemistry*, 2nd edn, ed W. Marshall, Mosby-Year Book Europe, London, UK, pp. 210–221.
- Mills, P. C., D. E. Auer, H. Kramer, D. Barry & J. C. Ng, 1998. Effects of inflammationassociated acute-phase response on hepatic and renal indices in the horse. *Australian Veterinary Journal*, 76, 187–194.
- Mircheva, T., I. Penchev, S. Tanev, A. Vachkov, V. Petrov, P. D. Eckersall, L. Sotirov, L. Lazarov, Ts. Christov & J. Nikolov, 2009. Variations of acute phase protein (haptoglobin, fibrinogen and ceruloplasmn) concentration in weaning rabbits after experimental infection with E. coli. Revue de Médecine Vétérinaire, 160, 133–139.
- Morris, D. D., J. Messick, R. H. Whitlock, J. Palmer, M. V. Ward & B. F. Feldman, 1988. Effect of equine ehrlichial colitis on the hemostatic system in ponies. *American Journal of Veterinary Research*, 49, 1030–1036.
- Moya, L., L. A. Boyle, P. B. Lynch & S. Arkins, 2007. Age-related changes in proinflammatory cytokines, acute-phase proteins and cortisol concentrations in neonatal piglets. *Neonatology*, 91, 44–48.
- Murata, H., N. Shimada & M. Yoshioka, 2004. Current research on acute phase proteins in veterinary diagnosis: An overview. *The Veterinary Journal*, 168, 28–40.
- Petersen, H. H., J. P. Nielsen & P. M. Heequard, 2004. Application of acute phase protein measurements in veterinary clini-

- cal chemistry. *Veterinary Research*, **35**, 163–187.
- Pfeffer, A., K. M. Rogers, L. O'Keeffe & P. J. Osborn, 1993. Acute phase protein response, food intake, liveweight change and lesions following intrathoracic injection of yeast in sheep. Research in Veterinary Science, 55, 360–366.
- Rubel, C., G. C. Fernandez, G. Dran, M. B. Bompadre, M. A. Isturiz, & M. S. Palermo, 2001. Fibrinogen promotes neutrophil activation and delays apoptosis. *Jour*nal of *Immunology*, 166, 2002–2010.
- Schmidt, E. & P. D. Eckersall, 2015. Acute phase proteins as markers of infectious diseases in small animals. *Acta Veterinaria*, 65, 149–161.
- Thomas, J. S., 2000. Overview of plasma proteins and protein electrophoresis. In: Schalm's Veterinary Hematology, 5th edn, eds B. F. Feldman, J. G. Zinkl & N. C. Jain, Lippincott Williams & Wilkins, Philadelphia, pp. 891–898.
- Todorov, J., 1972. Nephelometric determination of fibrinogen (method of Podmore). In: *Clinical Laboratory Technics*, Sofia, Medizina and Fizkultura, p. 250 (BG).
- Zapryanova, D., T. Mircheva & S. A. Denev, 2013. Plasma protein profiles and fibrinogen concentrations in dogs with experimentally induced *Staphylococcus aureus* infection. *Revue de Médecine Vétérinaire*, 164, 150–155.

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