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91	Abstract	This study aims to characterize myositis-specific antibodies in a well-defined cohort of patients with idiopathic inflammatory myopathy and to determine their association with cancer. Sera from 40 patients with polymyositis, dermatomyositis, and controls were tested by protein and RNA immunoprecipitation to detect autoantibodies, and immunoprecipitation-Western blot was used for anti-MJ/NXP-2, anti-MDA5, and anti-TIF1 γ / α identification. Medical records were re-evaluated with specific focus on cancer. Anti-MJ/NXP-2 and anti-TIF1 γ / α were the most common antibodies	

in dermatomyositis. In six dermatomyositis cases, we found five solid forms of cancer and one Hodgkin's lymphoma in long-term remission. Among patients with cancer-associated dermatomyositis, three were positive for anti-TIF1 γ/α , two for anti-Mi-2, and one for anti-MJ/NXP-2. The strongest positivity of anti-TIF1 γ was seen in two active forms of cancer, and this antibody was either negative or positive at low titers in the absence of cancer or in the 7-year remission Hodgkin's lymphoma. Four out of twenty (20 %) patients with polymyositis had solid cancer, but no specific association with autoantibodies was identified; further, none of the four cases of antisynthetase syndrome had a history of cancer. No serum myositis-associated autoantibody was observed in control sera, resulting in positive predictive value 75 %, negative predictive value 78.5 %, sensitivity 50 %, specificity 92 %, and area under the ROC curve 0.7083 for the risk of paraneoplastic DM in anti-TIF1 γ/α (+) patients. Myositis-specific autoantibodies can be identified thanks to the use of immunoprecipitation, and their association with cancer is particularly clear for anti-TIF1 γ/α in dermatomyositis. This association should be evaluated in a prospective study by immunoprecipitation in clinical practice.

92	Keywords separated by ' - '	Biomarkers - Cancer - Idiopathic inflammatory myositis - Immunoprecipitation
93	Foot note information	

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BRIEF REPORT

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Myositis-specific autoantibodies and their association with malignancy in Italian patients with polymyositis and dermatomyositis

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Abstract This study aims to characterize myositis-specific antibodies in a well-defined cohort of patients with idiopathic inflammatory myopathy and to determine their association with cancer. Sera from 40 patients with polymyositis, dermatomyositis, and controls were tested by protein and RNA immunoprecipitation to detect autoantibodies, and immunoprecipitation-Western blot was used for anti-MJ/NXP-2, anti-MDA5, and anti-TIF1γ/α identification. Medical records were re-evaluated with specific focus on cancer. Anti-MJ/NXP-2 and anti-TIF1γ/α were the most common antibodies in dermatomyositis. In six dermatomyositis cases, we found five solid forms of cancer and one Hodgkin's lymphoma in long-term remission. Among patients with cancer-associated dermatomyositis, three were positive for anti-TIF1γ/α, two for anti-Mi-2, and one for anti-MJ/NXP-2. The strongest positivity of anti-TIF1γ was seen in two active forms of cancer, and this antibody was either negative or positive at low titers in the absence of cancer or in the 7-year remission Hodgkin's lymphoma. Four out of twenty (20 %) patients with polymyositis had solid cancer, but no specific association with autoantibodies was identified;

further, none of the four cases of antisynthetase syndrome had a history of cancer. No serum myositis-associated autoantibody was observed in control sera, resulting in positive predictive value 75 %, negative predictive value 78.5 %, sensitivity 50 %, specificity 92 %, and area under the ROC curve 0.7083 for the risk of paraneoplastic DM in anti-TIF1γ/α (+) patients. Myositis-specific autoantibodies can be identified thanks to the use of immunoprecipitation, and their association with cancer is particularly clear for anti-TIF1γ/α in dermatomyositis. This association should be evaluated in a prospective study by immunoprecipitation in clinical practice.

Keywords Biomarkers · Cancer · Idiopathic inflammatory myositis · Immunoprecipitation

Introduction

Idiopathic inflammatory myopathy (IIM) is characterized by muscle inflammation, skin alterations, and internal organ involvement, resulting in muscle atrophy, skin microangiopathy, and tissue fibrosis [1]. IIMs are divided into several conditions with polymyositis (PM) and dermatomyositis (DM) as the most frequent forms despite being considered rare worldwide [2]. Beyond the clinical and histopathological differences, PM and DM can be further classified into subsets thanks to myositis-specific autoantibodies (MSA) which have diagnostic and prognostic roles [3, 4]. Some MSA have been known for decades, as for the anti-Jo-1 characterizing the antisynthetase syndrome or anti-Mi-2, peculiar for DM [5, 6]. Several MSA have been defined most recently by protein and RNA immunoprecipitation (IP). Paradigmatic MSA include DM-associated anti-MDA5, anti-MJ/NXP-2, and anti-TIF1γ/α which define specific clinical features and predict the

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64 association with cancer [2, 6, 7], sometimes without independent
65 confirmation [8].

66 To validate the proposed clinical associations, we utilized
67 IP for MSA in a well-characterized cohort of patients with PM
68 and DM from two clinical centers, with particular focus on the
69 specificities identified in recent years and their association
70 with cancer.

71 **Materials and methods**

72 **Patients**

73 The study included IIMs patients followed at the outpatient
74 clinic at Humanitas Research Hospital (Rozzano, Milan, Italy)
75 and Spedali Civili (Brescia, Italy) in the period 2013–2016.
76 We included sera from 20 patients with PM, 2 with
77 antisynthetase syndrome, 18 with DM, and controls represented
78 by healthy subjects (NHS; $n = 12$) and patients with systemic
79 sclerosis (SSc; $n = 79$), Behçet's disease (BD; $n = 45$),
80 and psoriatic arthritis (PsA; $n = 145$). We used established
81 criteria for the diagnosis of PM/DM, SSc, BD, and PsA and
82 collected clinical and laboratory data at enrollment.

83 The study was approved by the Institutional Review Board
84 of the hospitals and informed consent was obtained from all
85 subjects.

86 **Methods for autoantibody analysis**

87 Patients' sera were isolated from whole blood through centri-
88 fugation at 2000g for 15 min, and then stored in $-20\text{ }^{\circ}\text{C}$ freez-
89 er until use. MSA were first screened by protein-IP using ^{35}S -
90 methionine-labeled K562 cell extract followed by SDS-PAGE
91 and autoradiography, and by RNA-IP using unlabeled K562
92 cell extract followed by urea-PAGE and silver staining [9, 10].
93 MSA were determined using reference sera obtained from the
94 Autoantibody Standardization Committee (www.autoab.org)
95 and from internal controls.

96 Candidates for anti-MJ/NXP-2 and anti-MDA5 were
97 tested by IP-Western Blot (IP-WB) based on IP of a
98 140-kD protein, while candidates for anti-TIF1 γ/α were
99 selected based on bands at 155–140 kD by protein-IP. In
100 detail, 8 μl of candidate sera were cross-linked with
101 protein-A Sepharose beads and then immunoprecipitated
102 with cell extract from 10^7 K562 cells. Proteins were then
103 fractionated by 8 % SDS-PAGE and transferred to a nitro-
104 cellulose filter, probed with 1 $\mu\text{g/ml}$ of anti-MORC3
105 mouse polyclonal antibody (Abnova, Taipei City,
106 Taiwan) for MJ/NXP-2, followed by horseradish peroxi-
107 dase (HRP) goat anti-mouse IgG (1:5000 dilution)
108 (ThermoFisher, Waltham, MA, USA) and developed
109 using Immobilon Western Chemiluminescent HRP sub-
110 strate (Millipore, Darmstadt, Germany). The same

111 procedure was used for anti-MDA5 antibodies using
112 1:1000 rabbit anti-MDA5 antibody (Millipore,
113 Darmstadt, Germany) followed by 1:5000 HRP-
114 conjugated goat anti-rabbit Ig light chain antibody
115 (Jackson ImmunoResearch, West Grove, PA, USA), and
116 developed using Supersignal West Femto (ThermoFisher,
117 Waltham, MA, USA). For TIF1 γ IP-WB, we used 1:1000
118 mouse monoclonal anti-TIF1 γ antibody (Abcam,
119 Cambridge, UK), followed by 1:10.000 HRP goat anti-
120 mouse IgG (ThermoFisher, Waltham, MA, USA), and de-
121 veloped using Immobilon Western Chemiluminescent
122 HRP substrate (Millipore, Darmstadt, Germany).

Statistical analysis 123

124 All comparisons were performed by Mann-Whitney test and
125 Pearson Chi square test using Stata 13.1 for Macintosh
126 (StataCorp, 2013, CollegeStation, Texas, USA) and Prism
127 version 5.0 (GraphPad Software, Inc., La Jolla, CA, USA).
128 Statistical significance was accepted as $p < 0.05$.

Results 129

130 Through protein- and RNA-IP, we identified serum auto-
131 antibodies in IIMs as illustrated in Fig. 1a. None of our
132 control sera were positive for MSA. Before using IP and
133 IP-WB, only 6/18 (33 %) DM and 4/20 (20 %) PM cases
134 had autoantibodies detected by routine autoimmunity
135 tests, particularly anti-Ro/SSA, La/SSB, Mi-2, and -Jo-1.
136 Thanks to IP analysis, we confirmed this positivity and
137 identified additional MSA in patients with positive anti-
138 nuclear antibodies (ANA) by indirect immunofluores-
139 cence but negative autoantibody for extractable nuclear
140 antigens (ENA). We did not observe double autoantibody
141 positivity in our cohort, except for the association of Ro/
142 SSA, La/SSB, and Jo-1 as reported [11]. Protein- and
143 RNA-IP confirmed two less common antisynthetase anti-
144 bodies as anti-EJ (anti-glycyl tRNA synthetase) and anti-
145 PL-12 (anti-alanyl tRNA synthetase) in one PM and one
146 antisynthetase syndrome case, respectively (Fig. 1a).
147 Using RNA-IP, we identified the 7SL RNA band charac-
148 teristic of anti-SRP antibodies, in association with anti-
149 Ro/SSA in one patient with PM (Fig. 1b), and in both
150 cases necrotizing myositis was seen at muscle biopsy.
151 Eight cases (1 DM and 7 PM) remain seronegative by
152 IP, while in six DM cases, we identified bands at different
153 molecular weight by protein-IP but their antigenic signifi-
154 cance is still unknown (data not shown). Nine samples (5
155 IIMs and 4 SSc) had one band detectable around 140 kD
156 by protein IP, and were tested by IP-WB for anti-MJ/
157 NXP-2 and -MDA5 antibodies to identify the specificity
158 corresponding to this band. In three DM cases, we

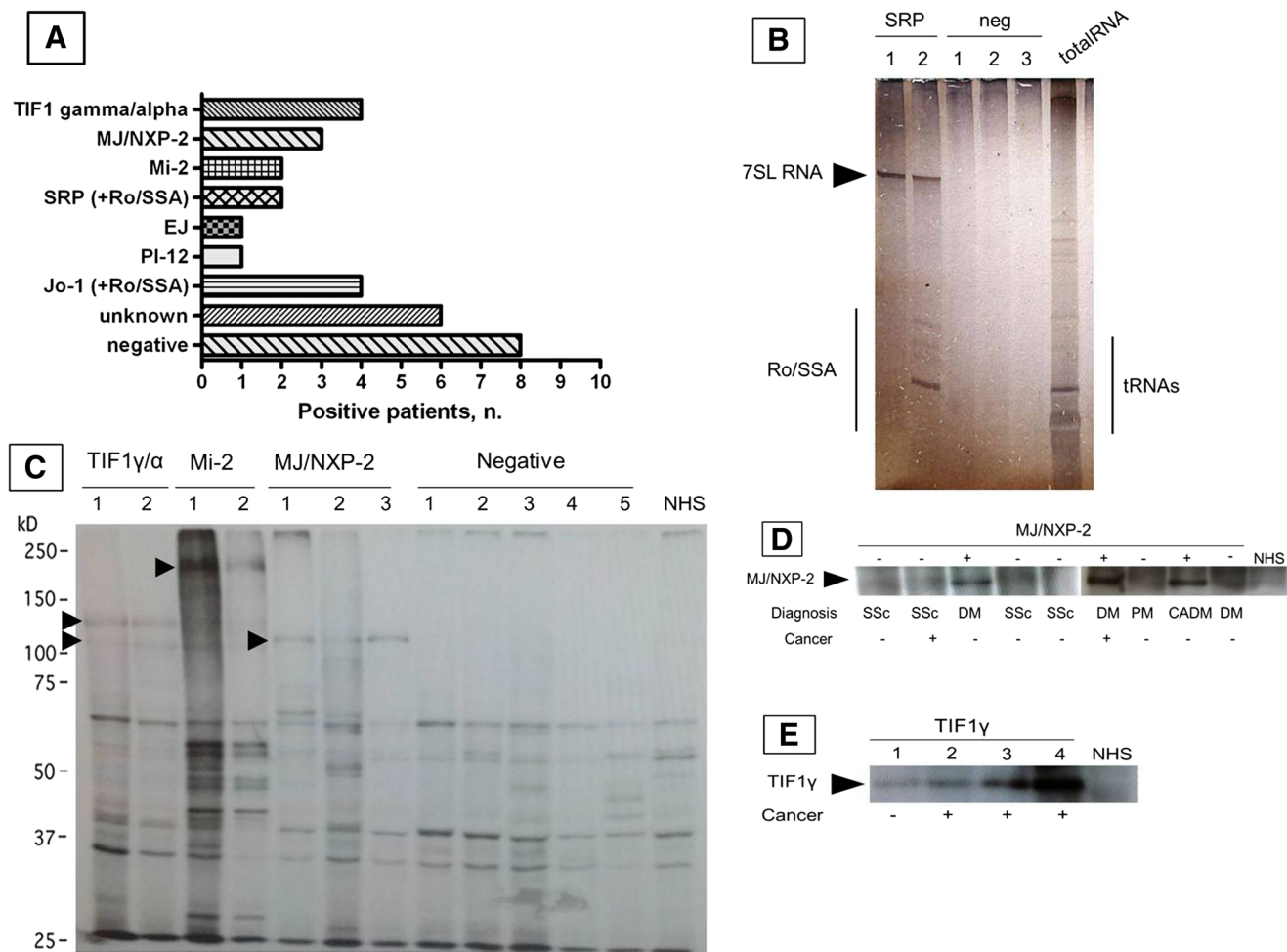


Fig. 1 MSA identified in our cohort of Italian PM/DM through the use of protein-IP, RNA-IP, and IP-WB. **a** Bar graph showing the specific autoantibodies and the number of corresponding cases identified in our cohort of PM/DM patients. **b** RNA-IP of two positive SRP samples, recognized by the band corresponding to 7SL RNA (*black arrow*). In one case, association with anti-Ro/SSA antibodies was identified (*black vertical line*) and supported by protein-IP (data not shown). The three negative RNA-IP samples shown are the anti-MJ/NXP-2 (+) patients reported in **c** and **d**, that typically do not show reactivity by RNA-IP. Total RNA was used as positive control. **c** Protein-IP (8 % SDS-PAGE gel) of representative PM/DM patients and corresponding autoantibodies: two anti-TIF1γ/α (+) cases shown by the bands at 155/140 kD (*black arrows*); two additional cases not shown in this protein-IP gel were identified based on the mobility of the same bands), two anti-Mi-2 (+) cases shown by the 240, 150, 72, 65, 63, 50, and 34 kD bands (*black arrow* for the 240 kD band), and the three anti-MJ/NXP-2 (+) cases

identified by the 140 kD band (*black arrow*). Five DM cases negative for MSA are also shown, and one NHS (normal human serum) is present in the last lane. **d** IP-WB for anti-MJ/NXP-2 positive cases. The three anti-MJ/NXP-2 (+) cases shown in the protein-IP gel in **c** were tested together with other samples (myositis and SSc) that had 140 kD band at protein-IP; no SSc sample had positivity for MJ/NXP-2. This panel also represents in which cases an association with cancer was present, as described in Table 2. **e** IP-WB for anti-TIF1γ positive cases. The four anti-TIF1γ (+) cases identified by protein-IP through the detection of the 155/140 kD bands were positive by IP-WB as shown in this panel. The weakest sample (#1) was the only one not associated with cancer until the moment of evaluation of the patient, sample #2 had a diagnosis of Hodgkin's Lymphoma 7 years before DM onset and it is now considered in remission, sample #3 has active lung cancer, and sample #4 has advanced ovary cancer. One normal human serum (NHS) is represented in the last lane

159 confirmed the anti-MJ/NXP-2 positivity (Fig. 1c, d),
160 while no MSA was detected in our control population.

161 The main clinical and laboratory features of patients are
162 described in Table 1. The diagnosis of myositis was confirmed
163 by muscle biopsy and/or electromyography only in 13/18
164 (72 %) of DM patients, coined clinically amyopathic DM.
165 No significant difference was detected for organ involvement,
166 laboratory tests abnormalities, and ongoing therapies in DM
167 and PM patients, while the expression of anti-TIF1γ/α

168 antibodies was significantly associated to DM patients
169 ($p = 0.04$) as shown in Table 1. The ANA pattern reported
170 by routine autoimmunity tests was very variable for titer and
171 pattern, and in some cases also defined as "negative" (Tables 1
172 and 2), thus it was necessary to proceed with further testing by
173 IP for the identification of MSA. Two anti-MJ/NXP-2 neces-
174 sary to proceed with further testing by IP for the identification
175 of MSA. Two anti-MJ/NXP-2 (+) DM patients had severe
176 diffuse calcinosis that required surgical removal in one case,
177

		DM (n = 18)	PM (n = 20)	p
t1.1	Table 1 Main demographic and			
t1.2	clinical features of our cohort of			
	DM and PM patients, for which			
	we performed serum IP analysis.			
t1.3	Two anti-synthetase cases are not			
t1.4	included			
t1.5				
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ANA anti-nuclear antibodies, CK creatine kinase, ENA extractable nuclear antigen, GI gastro-intestinal, IP immunoprecipitation, ns not significant

*Confirmed by electromyography and/or muscle biopsy

177 and ongoing therapy with pamidronate infusions in one case
 178 of clinically amyopathic DM [12]. The association with
 179 cancer was present only in one DM case positive for this autoantibody (Fig. 1d). No serum was positive for anti-MDA5 antibodies, and in fact, no patient in our PM/DM cohort had symptoms such as rapidly progressive interstitial lung disease that are usually associated with this autoantibody [6]. The four anti-TIF1γ (+) sera have history of cancer in the three strongest positive cases (Fig. 1e), while the weakest positive case is the only one without cancer history until the moment of our clinical evaluation.

188 Cumulatively, we calculated the positive predictive value
 189 (75 %), negative predictive value (78.5 %), sensitivity (50 %),
 190 specificity (91.6 %), and area under the ROC curve (0.7083)
 191 for the risk of paraneoplastic DM in anti-TIF1γ/α (+) patients
 192 and these were compared to previous reports.

Discussion

193
 194 The routine use of protein- and RNA-IP may increase the
 195 detection rate of rare autoantibodies in clinical practice, particularly in rare conditions such as PM and DM, thus maximizing the diagnostic and prognostic power of these biomarkers. In fact and despite the low incidence and prevalence worldwide, IIMs are characterized by wide clinical phenotype variability, mirrored by a significant number of MSA. We thus utilized the sensitive and specific IP to identify autoantibody prevalence and clinical significance in a well-defined cohort of Italian patients affected by PM/DM, with particular focus on cancer associations.

205 Our most relevant findings include that anti-MJ/NXP-2
 206 and -TIF1γ/α antibodies are the two most frequent MSA in
 207 DM cases previously anti-ENA negative at routine tests.

Table 2 Main characteristics of the anti-MJ/NXP-2 (+) and TIF1 γ/α (+) cases identified in our cohort of PM/DM patients. The cases described in this table are shown in Fig. 1d, e

	Anti-MJ/NXP-2 case 1	Anti-MJ/NXP-2 case 2	Anti-MJ/NXP-2 case 3	Anti-TIF1 γ/α case 1	Anti-TIF1 γ/α case 2	Anti-TIF1 γ/α case 3	Anti-TIF1 γ/α case 4
t2.3	Demographic data						
t2.4	Female	Female	Male	Male	Female	Female	Female
t2.5	33	21	21	72	40	59	54
t2.6	DM	CADM	DM	DM	DM	CADM	DM
t2.7	19	15	18	67	22	58	52
t2.8	Clinical data						
t2.9	+	+	+	+	+	+	+
	(Gottron's papules, erythematous rash)	(Gottron's papules, erythematous rash)	(V-neck erythema)	(Gottron's papules, erythematous rash)	(Gottron's papules, heliotrope rash)	(Gottron's papules, erythematous rash)	(Gottron's papules, erythematous rash)
t2.10	+++	+++	-	-	-	-	-
t2.11	+	-	+	+	+	-	+
t2.12	-	-	-	-	-	-	-
t2.13	-	-	-	-	-	-	+
t2.14	-	-	-	-	-	-	-
t2.15	+	-	-	-	+	+	+
t2.16	Thyroid	-	-	-	Hodgkin's lymphoma	Lung adenocarcinoma	Ovary
t2.17	31	-	-	-	15	56	52
t2.18	+	+	+	+	+	-	+
	(PDN, HCQ, MTX, CsA, IV Ig, AZA)	(PLQ)	(PDN, PLQ, MTX, IV Ig, AZA, CsA)	(PDN, MMF)	(PDN, HCQ, CTX, AZA)		(PDN, MTX, IV Ig)
t2.19	Laboratory data						
t2.20	+	-	+	+	+	-	+
t2.21	+	Normal	Normal	Normal	Normal	Normal	+
t2.22	1:640 nuclear dots	1:160 speckled	1:80 speckled	Negative	1:160 speckled	Negative	>1:640 speckled

ANA anti-nuclear antibodies, AZA azathioprine, CADM clinically amyopathic DM, CK creatine kinase, CsA cyclosporine, CTX cyclophosphamide, DM dermatomyositis, HCQ hydroxychloroquine, IV Ig intravenous immunoglobulins, MMF mycophenolate mofetil, MTX methotrexate, PDN prednisone

*The order of these therapies corresponds to the chronological order they were used by the patients

208 Accordingly, to what reported in the literature, our three anti-
 209 MJ/NXP-2 (+) DM cases have juvenile onset DM with typical
 210 skin DM features, no internal organ involvement, and the
 211 worst clinical manifestation is severe calcinosis [13]. All these
 212 cases required immunosuppressive therapy beyond steroids to
 213 control muscle inflammation, but in one case, DM was not
 214 completely controlled and this unresponsive patient had a di-
 215 agnosis of papillary thyroid cancer 12 years after the onset of
 216 DM. In fact, cancer has been reported in adult anti-MJ/NXP-
 217 2(+) DM patients despite not being confirmed in our previous
 218 publication on a different Italian cohort [10, 14]. The identifi-
 219 cation of anti-MJ/NXP-2 antibodies was based on the first
 220 observation of a common band of 140 kD molecular weight

221 by protein-IP, but it was then necessary to perform IP-WB to
 222 have a positive result for MJ/NXP-2. Anti-MDA5 antibodies
 223 also migrate in the same molecular weight range, but no sam-
 224 ple tested positive by IP-WB and it was concordant with the
 225 clinical observation that these samples did not show the sug-
 226 gestive clinical features (i.e., rapidly progressive interstitial
 227 lung disease) that are commonly referred to anti-MDA5
 228 positivity.

229 We detected serum anti-TIF1 γ/α antibodies in four DM
 230 cases, only in one case there was no history of cancer despite
 231 extensive screening exams and it was the weakest positive
 232 case. All the other three cases have cancer history. In one case,
 233 this autoantibody was present in a DM patient with Hodgkin's

234 lymphoma diagnosed and treated 7 years prior to the onset of
 235 DM features, and considered in remission at the time of the
 236 blood draw. This is in contrast with previous reports of anti-
 237 TIF1 γ / α antibodies not found in juvenile DM cases associat-
 238 ed with cancer and of the highest associated risk of malignan-
 239 cy during the year prior to and the year after IIMs diagnosis
 240 [15]. The two strongest samples positive for anti-TIF1 γ / α
 241 antibodies have active cancer unresponsive to treatment at
 242 the time of blood drawn. It is also important to highlight the
 243 fact that both our two anti-Mi-2(+) DM cases had a history of
 244 breast cancer prior to DM onset but no higher risk of cancer
 245 has been reported in association with this autoantibody [4].
 246 Our estimate of the positive and negative predictive values,
 247 sensitivity, specificity, and area under the ROC curve for the
 248 risk of paraneoplastic DM in anti-TIF1 γ / α (+) patients were
 249 concordant to previous reports [16–21].

250 In our PM cohort, four cases of cancer were reported, in
 251 three being diagnosed several years prior to and in one con-
 252 comitant to the onset of PM; in all cases, solid forms affecting
 253 the thyroid, colon, breast, skin, and only two of them had a
 254 known autoantibody signature at routine tests represented by
 255 anti-Ro/SSA antibodies. No tumor was reported in the four
 256 cases affected by antisynthetase syndrome, and no MSA was
 257 identified in SSc, BD, PsA cases with or without a history of
 258 cancer. No PM case showed positivity for anti-3-hydroxy-3-
 259 methylglutaryl-coenzyme A reductase (HMGCR) antibodies
 260 despite the onset of necrotizing myopathy after the use of
 261 statin [3]. Other autoantibodies that were reported by routine
 262 autoimmunity laboratories through techniques such as immu-
 263 noblotting were not confirmed by IP, as for anti-PM/Scl
 264 (PM100) and PL-7.

265 We collected data relative to indirect immunofluorescence
 266 ANA patterns reported by routine laboratory tests, and we
 267 observed that the most frequent ANA pattern reported in our
 268 anti-MJ/NXP-2(+) and anti-TIF1 γ / α (+) cases is speckled, and
 269 in one anti-MJ/NXP-2(+) case, the presence of nuclear dots
 270 suggestive for promyelocytic leukemia nuclear bodies was
 271 reported, thus needing further evaluation [10]. Despite the
 272 use of the most sensitive techniques, eight patients with
 273 IIMs remained negative for both ANA and ENA, while in
 274 six additional cases, we could identify bands by protein-IP,
 275 but no clear specificity [22]. These gaps underline the existing
 276 limitations in the identification of autoantibodies in rheumatic
 277 diseases such as PM/DM which are mainly due to lack of
 278 standardization for ANA and ENA, low number of positive
 279 cases studied for autoantibodies clinical association, identifi-
 280 cation of rare and new autoantibodies through time and labor-
 281 consuming techniques such as IP, and the lack of commercial-
 282 ly available techniques that may help in the identification of
 283 rare autoantibodies in a clinical setting [23] and shed light on
 284 PM/DM pathogenesis [2]. We acknowledge that the efforts of
 285 international registries such as Euromyositis or the
 286 Autoantibody Standardization Committee are expected to

minimize the frequency of seronegative cases and to provide 287
 a clear estimate of the prevalence of rare autoantibodies [24]. 288

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Compliance with ethical standards 291
 292

Disclosures None. 293

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References 295

1. Lundberg IE et al (2016) Diagnosis and classification of idiopathic 297
 inflammatory myopathies. *J Intern Med* 280(1):39–51 298

2. Ceribelli A. et al. (2016) *The Immune Response and the* 299
Pathogenesis of Idiopathic Inflammatory Myositis: a Critical
Review. *Clin Rev Allergy Immunol* 300
 301

3. Gunawardena H. (2015) *The Clinical Features of Myositis-* 302
Associated Autoantibodies: a Review. *Clin Rev Allergy Immunol* 303

4. Satoh M. et al. (2015) *A Comprehensive Overview on Myositis-* 304
Specific Antibodies: New and Old Biomarkers in Idiopathic
Inflammatory Myopathy. *Clin Rev Allergy Immunol* 305
 306

5. Targoff IN (1992) Autoantibodies in polymyositis. *Rheum Dis Clin* 307
N Am 18(2):455–482 308

6. Ceribelli A et al (2014) Prevalence and clinical significance of anti- 309
 MDA5 antibodies in European patients with polymyositis/derma- 310
 tomyositis. *Clin Exp Rheumatol* 32(6):891–897 311

7. Satoh M et al (2012) Autoantibodies to transcription intermediary 312
 factor TIF1beta associated with dermatomyositis. *Arthritis Res* 313
Ther 14(2):R79 314

8. Fiorentino DF et al (2016) PUF60: a prominent new target of the 315
 autoimmune response in dermatomyositis and Sjogren's syndrome. 316
Ann Rheum Dis 75(6):1145–1151 317

9. Ceribelli A et al (2010) Anti-Th/To are common antinucleolar au- 318
 toantibodies in Italian patients with scleroderma. *J Rheumatol* 319
 37(10):2071–2075 320

10. Ceribelli A et al (2012) Anti-MJ/NXP-2 autoantibody specificity in 321
 a cohort of adult Italian patients with polymyositis/dermatomyosi- 322
 tis. *Arthritis Res Ther* 14(2):R97 323

11. Yamasaki Y et al (2016) Clinical subsets associated with different 324
 anti-aminoacyl transfer RNA synthetase antibodies and their asso- 325
 ciation with coexisting anti-Ro52. *Mod Rheumatol* 26(3):403–409 326

12. Palaniappan P, Lionel AP, Kumar S (2014) Successful treatment of 327
 juvenile cutis in juvenile dermatomyositis with pamidronate. *J* 328
Clin Rheumatol 20(8):454–455 329

13. Rider LG et al (2013) The myositis autoantibody phenotypes of the 330
 juvenile idiopathic inflammatory myopathies. *Medicine* 331
 (Baltimore) 92(4):223–243 332

14. Ichimura Y et al (2012) Anti-NXP2 autoantibodies in adult patients 333
 with idiopathic inflammatory myopathies: possible association with 334
 malignancy. *Ann Rheum Dis* 71(5):710–713 335

15. Tiniakou, E. and A.L. Mammen, *Idiopathic Inflammatory* 336
Myopathies and Malignancy: a Comprehensive Review. *Clin Rev* 337
Allergy Immunol, 2015. 338

16. Selva-O'Callaghan A et al (2010) Malignancy and myositis: novel 339
 autoantibodies and new insights. *Curr Opin Rheumatol* 22(6):627– 340
 632 341

17. Targoff IN et al (2006) A novel autoantibody to a 155-kd protein is 342
 associated with dermatomyositis. *Arthritis Rheum* 54(11):3682– 343
 3689 344

345	18.	Gunawardena H et al (2008) Clinical associations of autoantibodies to a p155/140 kDa doublet protein in juvenile dermatomyositis. <i>Rheumatology (Oxford)</i> 47(3):324–328	358
346			359
347			360
348	19.	Kaji K et al (2007) Identification of a novel autoantibody reactive with 155 and 140 kDa nuclear proteins in patients with dermatomyositis: an association with malignancy. <i>Rheumatology (Oxford)</i> 46(1):25–28	361
349			362
350			363
351			364
352	20.	Chinoy H et al (2007) The diagnostic utility of myositis autoantibody testing for predicting the risk of cancer-associated myositis. <i>Ann Rheum Dis</i> 66(10):1345–1349	365
353			366
354			367
355	21.	Fujikawa K et al (2009) Association of distinct clinical subsets with myositis-specific autoantibodies towards anti-155/140-kDa polypeptides, anti-140-kDa polypeptides, and anti-aminoacyl tRNA synthetases in Japanese patients with dermatomyositis: a single-centre, cross-sectional study. <i>Scand J Rheumatol</i> 38(4):263–267	368
356			369
357			369
370			369
	22.	Selmi C et al (2016) Serum antinuclear and extractable nuclear antigen antibody prevalence and associated morbidity and mortality in the general population over 15 years. <i>Autoimmun Rev</i> 15(2): 162–166	369
			370
	23.	Cavazzana I et al (2016) Testing for myositis specific autoantibodies: comparison between line blot and immunoprecipitation assays in 57 myositis sera. <i>J Immunol Methods</i> 433:1–5	370
			371
	24.	Chan EK et al (2016) Report on the second International Consensus on ANA Pattern (ICAP) workshop in Dresden 2015. <i>Lupus</i> 25(8): 797–804	371
			372

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