## Glycated albumin: from biochemistry and laboratory medicine to clinical practice

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## Abstract

This review summarizes current knowledge about glycated albumin (GA). We review the changes induced by glycation on the properties of albumin, the pathological implications of high GA levels, GA quantification methods, and the use of GA as a complementary biomarker for diabetes mellitus diagnosis and monitoring and for dealing with long-term complications. The advantages and limits of this biomarker in different clinical settings are also discussed.

## 1. Introduction

The main functions of albumin include regulation of plasma oncotic pressure, maintenance of acid-base balance, action as a carrier molecule and as an antioxidant system. Adequate plasma levels of albumin (3.5-5.5 mg/dL)<sub>7</sub> as well as the maintenance of its structural integrity<sub>7</sub> are both essential if it is to function properly [1]. One of the main processes affecting the structure of albumin, hence its biochemical and physical properties, is glycation, a non-enzymatic-chemical reaction between reducing sugars and/or their degradation products and primary or secondary amine groups on proteins [2,3]. This results in the production of early and advanced glycation end-products (AGEs). Diabetes mellitus, a metabolic disease characterized by high blood glucose concentration, raising blood glucose, is considered the main pathology associated with high-a raise in the levels of glycation products which have been related just to the onset and progression of -There appears to be a relationship between glycation products and different detrimental diabetes complications, from cardiovascular pathologies to kidney diseases and blindness [4-6]. Among the different glycated proteins, some studies ascribe a potential pathogenic role only to glycated albumin (GA) [7]. Interest in GA has grown in the last few years, mainly in the field of diabetes monitoring, as a complementary biomarker to blood glucose and glycated hemoglobin (HbA1c) in specific clinical settings in

which these molecules do not work properly. In addition, due to the relationship between since-glycated

proteins play a key role in promoting and diabetes-related complications, monitoring GA may be an

additional and important toll in controlling the control of the risk of such complications. GA has emerged as a marker of diabetes complications.

In this review we summarize current knowledge about GA, discussing the biochemical properties of albumin, the effects of glycation on albumin, the pathological implications of high GA levels, the methods of GA quantification and the use of GA as a complementary biomarker for monitoring diabetes mellitus and dealing with <u>its</u> long-term complications of the disease.

### 2. Human serum albumin: biochemical and physiological aspects

Albumin is the most abundant circulating protein, accounting for 50-60% of total plasma proteins. In physiological conditions, its levels range from 3.5 to 5.5 g/dL. It is a multifunctional, small (66 kDa),

globular<u>and</u>, negatively charged protein. It comprises a single polypeptide chain of 585 amino acids with abundance of lysine (59) and arginine (24) residues, and containing 35 cysteine residues, 34 of which form 17 disulfide bridges, important for the overall tertiary structure of the protein [1,7]. Cysteine 34 (Cys 34), in contrast, is free and accounts for 80% of total plasma thiol. Cys 34 exists mainly in the reduced form, which gives albumin important antioxidant and scavenging properties Cysteine 34 (Cys-34), which mainly existsing in the reduced form, gives albumin important antioxidant and scavenging properties and accounts for 80% of total plasma thiol. Cys 34 exists mainly in the reduced form of total plasma thiol cys 34 exists mainly in the reduced form of total plasma thiol cys 34 exists mainly in the reduced form of total plasma thiol cys 34 exists mainly in the reduced form of total plasma thiol cys 34 exists mainly in the reduced form.

X-ray crystallographic studies indicated that the protein is organized in three domains (I, II, III), each containing two sub-domains, A and B. Two important ligand-binding sites,-Sudlow sites I and II are two important ligand-binding sites,-with different but not exclusive affinities. They, have been identified in respectively subdomains IIA and IIIA, respectively [9]. Site I binds mainly large heterocyclic compounds and endogenous substances, including bilirubin and porphyrins. Site II is smaller and less flexible, binds aromatic compounds, such as benzodiazepine and ibuprofen, and is more stereo-specific [10-12]. Also the N-terminal region and Cys-34 are two important binding sites to consider in discussing ligand binding properties of albumin.have binding properties. The N-terminal and Cys-34 also have binding capacity [13]. Drugs like cisplatin, D-penicillamine, and N-acetyl-cysteine, and some metal cations, mainly Au, Ag and Hg, interact with Cys-34. Other ions, such as Cu, Ni and Co, can bind to the N-terminal region. Also inIn this way albumin can, by limitinglimit the availability of these ions, their the interaction between these ions withand oxygen, thus preventing the production of -and-the generation of-reactive oxygen species, albumin exerts antioxidant properties [13].

Albumin is also considered the major Zn binding protein in plasma, although the nature and location of the binding is still not clear. The ability to bind iron is weaker [14,15]. The protein also contains seven binding sites for fatty acids [10,16].

In addition to these functions as an important antioxidant and binding protein, albumin is responsible for maintaining colloid osmotic pressure and is involved in acid-base homeostasis [1]. Any pathological condition that lowers circulating albumin levels, or any change in the protein structure - affecting its biochemical and physical properties - mightay result in a loss of functions [10].

#### 3. Glycation

Glycation is a non-enzymatic reaction between reducing sugars or their degradation products and free amino groups of proteins. The formation of glycated products is quite complex but, according to a more simplified view, it is often described as a chemical process that proceeds. The process proceeds through three broad main reaction-stages: initial, intermediate and late. In the initial stage, the amino-carbonyl interaction results in the formation of an intermediate aldimine (Schiff base). The intermediate stage involves chemical transformation of this early glycation product into a more stable ketoamine (Amadori product, e.g. fructosyl-lysine) and the last stage is the formation of irreversible AGEs, due to oxidation, degradation and other cross-linking reactions. It is still not known whether AGE products are primarily formed by oxidation of Amadori fructosyl or by free carbonyls (non-fructosyl glycation modifications).

To go deeply into the glycation process, it is possible to find a very detailed description of the chemistry of the glycation process in a review by Cho focused on the mechanistic profile of the glycation pathways [17]. Cho has provided a very detailed description of the chemistry of the glycation process in a review focused on the mechanistic profile of the glycation pathways <u>According to this</u>, <u>Seven are main stages utilized by Cho</u> for describing the formation of glycated products is divided into seven major stages. In tThis scenario-is clearly suggests evident not only the complexity of the glycation process but also the redundancy of some stages, the possibility of generating the same AGEs through different chemical pathways and the difficulty of conceiving any single strategy for the prevention and management of AGE formation, that is considered one leading cause of various health complications. Please refer to the review by Cho [17] to find additional information on the chemistry of each of the seven stages of the glycation process.

[17]. According to this, the complex process of glycation can be divided into seven major stages. Stage (i) refers to the formation of the intermediate Schiff base and its rearrangement into the Amadori product. Stage (ii) deals with the transformation of the Amadori product into different AGEs through non-oxidative and oxidative reactions. Stage (iii) involves the degradation of the Schiff base to produce glyoxal and glyoxal-protein adducts which are further transformed into AGEs, like pentosidine and *N*<sup>e</sup> carboxymethyl lysine (CML), regardless of the Amadori pathway. Stage (iv) involves auto oxidative glycosylation leading to the formation of novel sugar molecules, like arabinoses, which have lost one atom in their carbon chain. In stage

(v) the arabinoses generated in stage (iv) are transformed into AGEs. Stage (vi) involves glucose In pathological conditions such as diabetes mellitus, the high glycemia promotes the formation of high levels of glycated proteins and AGEs [2,18,19]. Glycation is certainly detrimental to proteins because it affects both their tertiary structure and all the functions closely related to protein conformation [20]. Glycated proteins may also, *per se*, exert damaging effects, thus contributing to the onset and progression of different pathology related complications [21,22].

Among different glycated proteins, HbA1c is already used in clinical practice as a biomarker for both diabetes mellitus diagnosis and monitoring. Other specific glycated proteins may therefore be worth investigating as potential biomarkers in specific clinical settings, but to date GA is the only other glycated protein used as an indicator of glycemic control.

### 4. Glycated albumin: structural modification and pathological implications

Albumin is more sensitive to glycation than other proteins, mainly because of its high concentration and long half-life (about 21 days) and also due to the, besides the large number of lysine and arginine residues which may be involved in the formation of early and advanced glycation products [1,18,19]. Among the 59 lysines in albumin, only -a-few of them are real targets of the modifications found in GA. Lysine 525 is the main target site of glycation. Other important glycation sites include lysine 439, 199, 51, 378, 545, 12, 233, 276, 281, 317, 323 and also the N-terminus [20-25]. Probably the accessibility of these sites, the local acid-base catalysis effect and the pKa of each amine group are the main factors in the susceptibility of these residues.

In addition to early glycation products, AGEs have also been observed in albumin. These are the result of the oxidation/modification of adducts on glycated proteins to form alpha-oxaloaldehydes and their subsequent reaction with arginine and lysine residues in albumin. Some possible AGEs observed in albumin include methylglyoxal-derived hydroimidazolone isomer 1, glyoxal-derived hydroimidazolone isomer 1, tetrahydropyrimidine, N<sub>e</sub>-carboxymethyl-lysine, N<sub>e</sub>-carboxyethyl-lysine, imidazolone B and pyrraline [26–28]. The kinds of change and the extent of glycation on albumin strongly depend on the amounts of reducing sugars and/or their degradation products and the time available for reaction with albumin. The high reactivity of albumin towards these agents is one of the reasons making why-GA is-an interesting biomarker for

glycemic control. The pros and cons of using GA for monitoring glycemia and additional reasons why it might be a worthwhile biomarker are discussed in section 6.

Modifications induced by glycation significantly change the structure and ligand binding properties of albumin *in vitro*. Most of the changes in GA occur on several residues near the two Sudlow binding sites I and II, <u>thus</u> suggesting a <u>potential</u><sup>n</sup> overall alteration of albumin-binding properties. *In vitro* studies on different ligands, such as warfarin, L-tryptophan, sulfonylureas, salicylate, ketoprofen and fatty acids, and <u>those which usingutilized</u> *in vitro* modified albumin with glycation levels close to those seen in diabetes mellitus, confirmed that albumin's binding ability is affected by glycation. In some cases, the effect seems to be minimal, as for warfarin, whereas for other molecules, such as ketoprofen, bilirubin and L-tryptophan<sub>1</sub> the changes are more marked [29-34]. This means potential change(s) in the therapeutic effects and/or adverse effects for all drugs that bind to albumin, with a large change mainly for agents that work in a relatively narrow range.

Glycation <u>may also promote changes inhas also been associated with changes to</u> albumin's anti-oxidant and scavenging properties [8]. Although strongly backed by *in vitro* studies, the clinical significance of these findings has not been shown till now. The review by Anguizola et al. [35] provides further details on structural studies of GA, the rate of glycation and modification for human albumin and the effect of glycation on albumin's properties.

In the glycated form, albumin does not simply present changes in its physiological functions but <u>italso</u> acquires a pathological phenotype too. High GA levels may induce irreversible damage in the different organs and tissues that are the main targets of complications in diabetes mellitus (such as coronary arteries, cardiovascular system, kidney, eye and nervous system). For example, in the kidney, GA is transported across the glomerular capillaries and taken up by epithelial and mesangial cells where it increases the production of pro-oxidant molecules and contributes to the onset of nephropathy [4,36,37]. In the field of cardiovascular diseases, GA plays a role in the activation and aggregation of platelets, up-regulates the expression of adhesion molecules involved in the formation of atherosclerotic plaques, like ICAM-1 and VCAM-1, and promotes oxidation [38-40]. The activation of the cell-surface receptor for AGE (RAGE) is considered-the main mechanism through which GA exerts its damaging effects. In fact, engagement of RAGE leads to the activation of the nuclear transcription factor NF-κB, the production of pro-inflammatory

cytokines and growth factors, apoptosis, oxidative stress and prothrombotic activities, all events which have been associated to pathological consequences of increased levels AGE and GA [41-43]. The increased generation of intracellular reactive oxygen species is also considered one of the main mechanisms leading to the inhibition of glucose uptake both in adipocytes and muscle cells, which in turnthus promotesing insulin resistance [44].

## 5. Quantification of glycated albumin: methods

GA concentrations can be determined in several ways using colorimetric, enzymatic, chromatographic and mass-spectrometry methods. GA is always expressed as a percentage of albumin. Each test has its pros and cons, related to aspects like ease of use, skills, and instrument-hardware availability.

GA has beingen determined with a high-performance liquid chromatography (HPLC) \_-method [45,46] that can be used for clinical purposes despite its high cost and the need for skill and know-how. However, the HPLC method is not really useful for routine analysis because of its low throughput [47]. Mass spectrometry (MS) methods are also available but are not currently suitable for clinical purposes, for obvious reasons, mainly method standardization and its complexity.

In the last fifteen years a new<u>and</u>, accurate assay has been developed based on albumin-specific enzymatic protease and ketoamine-oxidase [47,48]. This assay, which allows the direct quantification of GA, is widely used in Japan, China, Taiwan and Korea for diabetes management, but has not yet been approved by the FDA for use in the U.S.A. and is not in use for clinical purposes in Europe either.

GA definition varies with the assay methods and their target molecules, thus providing different information. HPLC defines GA as the ratio of glycated albumin molecules to total albumin molecules, <u>whereas</u> enzymatic assays and MS measure the concentration as the ratio of glycated albumin amino acids to total albumin [48]. To avoid misinterpretation of results and their clinical meaning, an equation has been developed to match enzymatic assay GA percentages to the HPLC findings [49]. Each of the GA quantification methods previously indicated will indicate how much albumin is glyctaed

Other tests can calculate and/or mathematically derive indices of glycation, like the glycated protein ratio [50], aimed at achieving the same elinical utility as GA. However, the GA quantification method is specific for GA and is different from "fructosamine" or total glycated protein methods. These quantify all the

glycated serum proteins, depending closely on total protein levels, and can be affected by a high serum uric "total glycated protein" which quantify. These last, in fact, quantify all serum glycated proteins and they also protein levels, high serum uric acid concentration [50] and unspecific serum reducing activities [51]. There are also-other tests which that can calculate or mathematically derive indices of glycation, like the glycated ratio [52]. Although these tests aim to achieve the same clinical utility as GA they are totally different from methods performing the specific quantification of GA.

Unlike the ketoamine oxidase method for GA quantification, the assay for total glycated serum protein is approved for use in the U.S.A. but, as already mentioned, is not interchangeable with specific measurement of GA. As a result, although there are FDA approved tests for fructosamine and total glycated serum proteins, and some commercial laboratories do HPLC assays for GA, there are no FDA approved commercial assays for GA for use by clinical laboratories in the U.S.A.

When we qua. ntify GA we give a specific information on how much albumin is glycated. This kind of information is thus different from what we can obtain with other tests like "fructosamina" or "total glycated protein" which quantify. These last, in fact, quantify all serum glycated proteins and they also be but the results may be strongly affected by total protein levels, high serum uric acid concentration [50] and unspecific serum reducing activities [51]. There are also-other tests which that can calculate or mathematically derive indices of glycation, like the glycated protein ratio [52]. Although these tests aim to achieve the same clinical utility as GA they are totally different from methods performing the specific quantification of GA.

In the U.S.A, although some commercial laboratories do HPLC assays for GA, there are no FDA approved commercial assays for GA quantification. This means that also the ketoamine oxidase method for GA quantification is not yet approved for clinical purpose. At now, the only FDA-approved tests include total glycated serum protein and fructosamine assays that, as already mentioned, are not interchangeable with specific measurement of GA.

6. Glycated albumin as a biomarker in different clinical settings

6.1 GA for screening and monitoring diabetes mellitus

**Formattato:** Tipo di carattere:

GA is considered useful for assessing the degree of protein glycation directly dependent on glucose exposure. GA is already considered reliable in Asian countries such as Japan, China and Korea where it is measured as an intermediate glycemic control marker. GA is mainly trusted as a marker of glycation in diabetes mellitus management. It is currently used in Asian countries, such as Japan, China and Korea, for diabetes screening, population stratification and classification for the risk of developing diabetes, and for driving therapies [53-55]. Table 1 summarizes the latest studies exploring the -GA cut-offs for diabetes diagnosis and monitoring.

The conventional biomarkers employed for screening and monitoring diabetes mellitus include fasting glucose, postprandial glucose and HbA1c. Although these markers provide useful information, in certain clinical conditions they are inadequate. HbA1c is a glycation index that reflects the glycemic status over a period of 120 days, corresponding to the mean lifespan of erythrocytes. Because hemoglobin is found inside these cells, its glycation may be affected by any condition affecting their lifespan (hemolytic anemia, hemorrhage, folate and vitamin B12 deficiency anemia, aplastic anemia, nephropathy) and hemoglobin metabolism (variant hemoglobin, thalassemia) [56-59]. In fact, HbA1c is lower than plasma glucose in all those conditions of shortened erythrocyte lifespan\_r-and is higher when their lifespan is increased. Changes in red blood cell survival are <u>in fact</u> an important determinant of the discordance between HbA1c levels and other measures of glycemic control, <u>both</u> in hematologically normal individuals as well as in people with hematological disorders [60,61]. HbA1c is the result of glycation occurring both on older and new erythrocytes; but the reaction is slower on newly formed cells [59,62]. For these reasons, in conditions involving reduced erythrocyte lifespan the ratio of new to old cells will be higher and, as consequence, the HbA1c level will be lower. Unlike HbA1c, GA is not inside the erythrocytes so it is not influenced by their lifespan.

GA is also independent of iron deficiency [58,61,63]. In individuals with iron deficiency, HbA1c levels are higher than plasma glucose. This is very likely because cessation of erythropoiesis results in no new unglycated hemoglobin being produced. As a result, the existing hemoglobin becomes older, raising the mean age of circulating erythrocytes, hence also the percentage of glycated hemoglobin in the circulation. Pregnancy is aA physiological condition in which HbA1c suffers some limits as an indicator of glycemic pregnancy. During pregnancy HbA1c undergoes a biphasic change: it drops from the first to the second trimester, due to the decrease in plasma glucose levels, and rises again from the second to the third trimester, probably due to iron deficiency [64]. Unlike HbA1c, GA is not affected by iron deficiency [65,66] and, it is an intermediate-term glycemic marker (this <u>will beis</u> discussed in the next section), it enables pregnant with diabetes to maintain stricter glycemic control, important to lower the risk of fetal and maternal complications. GA is also a useful biomarker for monitoring diabetes mellitus in newborns, where HbA1c does not properly reflect the glycemic status on account of the high levels of fetal hemoglobin [67]. Patients with diabetes and end-stage renal diseases under dialysis also cannot be efficiently managed with HbA1c, because of <u>the lower hemoglobin and erythropoietin concentrations and the reduced persistence of red blood cells due to mechanical disruption</u> the reduced persistence of red blood cells due to mechanical disruption and erythropoietin dosage [68,69]. GA may be a better indicator of their glycemic status.

Recent studies have indicated suggested GA as a useful diagnostic tool for diabetes screening both in the general population as well asnd in individuals with a pre-diabetic condition. The KOPS study [54] suggested the utility of GA in diabetes mellitus diagnosis in the Japanese population. A strong correlation has been found between GA, HbA1c and fasting glucose. GA 15.5% was an optimal cut-off for predicting diabetes (sensitivity 83.3%; specificity 83.3%) (Table 1). In a large cross-sectional Japanese community-based population study [55] GA appeared useful as a screening tool in individuals with fasting glucose between 5.5-6.9 mmol/L and HbA1c <6.5%, in whom the OGTT test was used to diagnose diabetes. Optimal thresholds for diabetes screening were 15.2% for GA and 5.9% for HbA1c in this population. Using these cut-offs, GA and HbA1c had the same sensitivity (62.1%) for detecting diabetes; the specificity was 61.9 % for GA and 66.7% for HbA1c. Sumner et al. [70] suggested that in U.S.-based Africans with pre-diabetes mellitus, diagnosed by -a 2h-OGTT test, HbA1c and GA used as single tests detected ≤50% of individuals with pre-diabetes. However, combining them made it possible to identify nearly 80%.

## 6.3 Intermediate-term glycemic status

Compared to HbA1c, GA is considered an intermediate-term glycemic indicator, because the turnover of albumin is shorter than the lifespan of erythrocytes (20 *vs.* 120 days). GA also rises sooner than HbA1c when glycemic status worsens. This means that GA is more useful as an indicator of glycemic status in all those

conditions requiring short-term control of changes in glycemia, such as after the start or modifications of diabetes treatments [53]. The potential beneficial effects of a dietary strategy and lifestyle changes, often applied as first steps to improve glycemia before any drug therapy, could be monitored by GA too [71,72]. GA seems useful for early monitoring of the worsening of glycemic control in patients after discharge from hospital education programs. Because it rises faster than HbA1c, GA means that countermeasures can be taken promptly [73].

A large rise in plasma glucose in a very short time is sometimes seen in patients with type 1 diabetes. As a result, GA rises more than HbA1c at diagnosis, and the GA/HbA1c ratio may be higher than in patients with untreated type 2 diabetes. A GA/HbA1c ratio  $\geq$ 3.2 (sensitivity 97%, specificity 98%) was therefore proposed as a cut-off for identifying patients with type 1 diabetes, in which hyperglycemia comes on rapidly, from those with untreated type 2 diabetes [74].

The reason why GA levels rise faster than HbA1c depends closely on albumin's biochemistry, its high glycation speed and its half-life in serum. The glycation speed of albumin is around 4.5 times that of Hb [75].

#### 6.4 Postprandial glycemia

In terms of cardiovascular risk, postprandial glycemia may be a stronger risk factor than fasting glucose, so it needs to be strictly controlled [76-78]. In patients with diabetes, mainly type 1, post-prandial glucose may fluctuate over a wide range. Comparing HbA1c and GA in terms of their ability to distinguish postprandial glucose fluctuations, GA reflects them better [79.80].[79.80]: patients with postprandial hyperglycemia have higher GA than HbA1c. Thus, unlike HbA1c, which is mainly reflects thean indicator of the mean plasma glucose concentration, GA strongly reflects postprandial glucose and seems a more appropriate indicator biomarker of glycemic control in diabetespostprandial glucose levels [53,79,80].

Marked hyperglycemia is also often seen in gastrectomy patients after loading. Their GA/HbA1c ratio is usually was higher than controls, suggesting that GA rises more than HbA1c and seems a better marker for glycemic excursions in these individuals [81].

It is still not clear why GA reflects postprandial glycemia better than HbA1c, but the different glycation rates of the two molecules may be involved.

#### 6.5 GA as a biomarker of complications of diabetes mellitus and other diseases

Most studies focusing on algorithms for patient's follow-up usually compare GA with HbA1c, fasting glucose, OGTT and fructosamine, seeking the best parameter combination with the highest predictive value for diabetes-related complications. As we discussed earlier, GA may be directly implicated in the development of different complications related to diabetes, playing a role as a pathogenic molecule. This has been observed in the onset of nephropathy and in the development of atheromatous plaques. The Atherosclerosis Risk Communities (ARIC) study suggested there might be independent relationships between GA and retinopathy, chronic kidney disease and the incidence of diabetes mellitus. GA was similar to HbA1c in the prediction of retinopathy and chronic kidney disease, but lower than HbA1c for diabetes [82]. In a case-cohort subpopulation of the DCCT/EDIC study GA and HbA1c had similar associations with nephropathy and retinopathy which were strengthened when both measures were considered together, but only HbA1c was associated with the risk of cardiovascular diseases [83]. On the other hand, GA, similar to HbA1c, was further associated with intima media thickness, an indicator of subclinical atherosclerosis, in individuals with and without diabetes, suggesting that it might be <u>also</u> a helpful marker for estimating atherosclerosis risk [84,85].

These data suggest that GA may be an additional aid not only for monitoring and screening diabetes mellitus but also in risk stratification.

#### 6.6 What are the limits of GA?

Undoubtedly in certain specific clinical conditions GA offers some advantages over the classical markers for monitoring glycemic status. In some specific disorders GA levels are either lower or higher than the mean plasma glucose concentration, mainly because of changes in the albumin metabolism. In all conditions with increased albumin catabolism, such as nephrotic syndrome and hyperthyroidism, GA would be lower than blood glucose [86,87]. In obesity too, GA levels would be lower than glycemia because of the increased albumin catabolism promoted by the chronic micro-inflammation [88,89]. A similar alteration has been seen in smokers, patients with non-alcoholic fatty liver disease, hypertriglyceridemia and hyperuricemia [90-92]. In all conditions with reduced albumin metabolism, such as liver cirrhosis and hypothyroidism, GA is higher than blood glucose. In chronic liver diseases (CLD), both GA and HbA1c have some limits for monitoring

the glycemic status. GA is higher than blood glucose because of the reduced albumin synthesis and HbA1c is lower because of the increased erythrocyte catabolism. Simultaneous measurements of HbA1c and GA, and the calculation of CLD-HbA1c (mean of HbA1c and GA/3) wasere proposed as a useful indicator of glycemic control in CLD patients [93-95].

#### 7. Conclusions

GA is an interesting biomarker for diabetes mellitus: 1) as it is an intermediate-term marker of the glycemic status it gives more information than the short-term (glycemia) and long-term (HbA1c) biomarkers currently employed in clinical practice; 2) in specific clinical conditions (altered erythrocyte lifespan, pregnancy and end-stage renal disease) it should be preferred to HbA1c; 3) it is probably also useful for diabetes mellitus screening and risk stratification of diabetes-related complications. Table 2 summarizes the pros and cons of GA use.

In conclusion, the introduction of this biomarker in clinical practice could help clinicians in the early diagnosis of diabetes mellitus, and in planning measures to prevent long-term diabetes complications.

Conflict of Interest: The authors declare that they have no conflict of interest

#### References

- 1. Theodore Peters, J.: All About Albumin. Academic Press, (1995)
- 2. Thornalley, P.J., Langborg, A., Minhas, H.S.: Formation of glyoxal, methylglyoxal and 3-deoxyglucosone

in the glycation of proteins by glucose. Biochem J 344 Pt 1, 109-116 (1999)

3. Kim, K.J., Lee, B.W.: The roles of glycated albumin as intermediate glycation index and pathogenic

protein. Diabetes Metab J 36(2), 98-107 (2012). doi:10.4093/dmj.2012.36.2.98

4. Ziyadeh, F.N., Han, D.C., Cohen, J.A., Guo, J., Cohen, M.P.: Glycated albumin stimulates fibronectin gene expression in glomerular mesangial cells: involvement of the transforming growth factor-beta system.
Kidney Int 53(3), 631-638 (1998)

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5. Jeffcoate, S.L.: Diabetes control and complications: the role of glycated haemoglobin, 25 years on. Diabet Med **21**(7), 657-665 (2004)

6. Hartog, J.W., Voors, A.A., Bakker, S.J., Smit, A.J., van Veldhuisen, D.J.: Advanced glycation endproducts (AGEs) and heart failure: pathophysiology and clinical implications. Eur J Heart Fail **9**(12), 1146-1155 (2007)

7. He, X.M., Carter, D.C.: Atomic structure and chemistry of human serum albumin. Nature **358**(6383), 209-215 (1992)

8. Oettl, K., Stauber, R.E.: Physiological and pathological changes in the redox state of human serum

albumin critically influence its binding properties. Br J Pharmacol 151(5), 580-590 (2007)

9. Sudlow, G., Birkett, D.J., Wade, D.N.: The characterization of two specific drug binding sites on human serum albumin. Mol Pharmacol **11**(6), 824-832 (1975)

 Kragh-Hansen, U., Chuang, V.T., Otagiri, M.: Practical aspects of the ligand-binding and enzymatic properties of human serum albumin. Biol Pharm Bull 25(6), 695-704 (2002)

11. Otagiri, M.: A molecular functional study on the interactions of drugs with plasma proteins. Drug Metab Pharmacokinet **20**(5), 309-323 (2005).

12. Simard, J.R., Zunszain, P.A., Ha, C.E., Yang, J.S., Bhagavan, N.V., Petitpas, I., Curry, S., Hamilton,

J.A.: Locating high-affinity fatty acid-binding sites on albumin by x-ray crystallography and NMR

spectroscopy. Proc Natl Acad Sci U S A **102**(50), 17958-17963 (2005)

13. Eom, J.E., Lee, E., Jeon, K.H., Sim, J., Suh, M., Jhon, G.J., Kwon, Y.: Development of an albumin

copper binding (ACuB) assay to detect ischemia modified albumin. Anal Sci 30(10), 985-990 (2014)

14. Masuoka, J., Saltman, P.: Zinc(II) and copper(II) binding to serum albumin. A comparative study of dog,

bovine, and human albumin. J Biol Chem **269**(41), 25557-25561 (1994)

Loban, A., Kime, R., Powers, H.: Iron-binding antioxidant potential of plasma albumin. Clin Sci (Lond)
 93(5), 445-451 (1997)

Curry, S., Mandelkow, H., Brick, P., Franks, N.: Crystal structure of human serum albumin complexed with fatty acid reveals an asymmetric distribution of binding sites. Nat Struct Biol 5(9), 827-835 (1998)
 Cho, S.J., Roman, G., Yeboah, F., Konishi, Y.: The road to advanced glycation end products: a

mechanistic perspective. Curr Med Chem 14(15), 1653-1671 (2007)

Monacelli, F., Storace, D., D'Arrigo, C., Sanguineti, R., Borghi, R., Pacini, D., Furfaro, A.L., Pronzato, M.A., Odetti, P., Traverso, N.: Structural alterations of human serum albumin caused by glycative and oxidative stressors revealed by circular dichroism analysis. Int J Mol Sci 14(6), 10694-10709 (2013)
 Baraka-Vidot, J., Planesse, C., Meilhac, O., Militello, V., van den Elsen, J., Bourdon, E., Rondeau, P.: Glycation alters ligand binding, enzymatic, and pharmacological properties of human albumin. Biochemistry 54(19), 3051-3062 (2015)

20. Day, J.F., Thorpe, S.R., Baynes, J.W.: Nonenzymatically glucosylated albumin. In vitro preparation and isolation from normal human serum. J Biol Chem **254**(3), 595-597 (1979)

21. Iberg, N., Fluckiger, R.: Nonenzymatic glycosylation of albumin in vivo. Identification of multiple

glycosylated sites. J Biol Chem 261(29), 13542-13545 (1986)

22. Zoellner, H., Hou, J.Y., Hochgrebe, T., Poljak, A., Duncan, M.W., Golding, J., Henderson, T., Lynch,

G.: Fluorometric and mass spectrometric analysis of nonenzymatic glycosylated albumin. Biochem Biophys Res Commun **284**(1), 83-89 (2001)

23. Frolov, A., Hoffmann, R.: Identification and relative quantification of specific glycation sites in human serum albumin. Anal Bioanal Chem **397**(6), 2349-2356 (2010)

24. Barnaby, O.S., Cerny, R.L., Clarke, W., Hage, D.S.: Quantitative analysis of glycation patterns in human serum albumin using 16O/18O-labeling and MALDI-TOF MS. Clin Chim Acta **412**(17-18), 1606-1615 (2011)

25. Awasthi, S., Murugan, N.A., Saraswathi, N.T.: Advanced Glycation End Products Modulate Structure and Drug Binding Properties of Albumin. Mol Pharm **12**(9), 3312-3322 (2015)

26. Brancia, F.L., Bereszczak, J.Z., Lapolla, A., Fedele, D., Baccarin, L., Seraglia, R., Traldi, P.:

Comprehensive analysis of glycated human serum albumin tryptic peptides by off-line liquid

chromatography followed by MALDI analysis on a time-of-flight/curved field reflectron tandem mass

spectrometer. J Mass Spectrom 41(9), 1179-1185 (2006)

27. Wa, C., Cerny, R.L., Clarke, W.A., Hage, D.S.: Characterization of glycation adducts on human serum albumin by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry. Clin Chim Acta **385**(1-2), 48-60 (2007)

- 28. Barnaby, O.S., Cerny, R.L., Clarke, W., Hage, D.S.: Comparison of modification sites formed on human serum albumin at various stages of glycation. Clin Chim Acta **412**(3-4), 277-285 (2011)
- 29. Barzegar, A., Moosavi-Movahedi, A.A., Sattarahmady, N., Hosseinpour-Faizi, M.A., Aminbakhsh, M.,
- Ahmad, F., Saboury, A.A., Ganjali, M.R., Norouzi, P.: Spectroscopic studies of the effects of glycation of
- human serum albumin on L-Trp binding. Protein Pept Lett 14(1), 13-18 (2007)
- 30. Kisugi, R., Kouzuma, T., Yamamoto, T., Akizuki, S., Miyamoto, H., Someya, Y., Yokoyama, J., Abe, I.,
- Hirai, N., Ohnishi, A.: Structural and glycation site changes of albumin in diabetic patient with very high
- glycated albumin. Clin Chim Acta 382(1-2), 59-64 (2007)
- 31. Joseph, K.S., Moser, A.C., Basiaga, S.B., Schiel, J.E., Hage, D.S.: Evaluation of alternatives to warfarin
- as probes for Sudlow site I of human serum albumin: characterization by high-performance affinity
- chromatography. J Chromatogr A 1216(16), 3492-3500 (2009)
- 32. Joseph, K.S., Hage, D.S.: The effects of glycation on the binding of human serum albumin to warfarin and L-tryptophan. J Pharm Biomed Anal **53**(3), 811-818 (2010)
- 33. Baraka-Vidot, J., Guerin-Dubourg, A., Bourdon, E., Rondeau, P.: Impaired drug-binding capacities of in vitro and in vivo glycated albumin. Biochimie **94**(9), 1960-1967 (2012)
- 34. Matsuda, R., Anguizola, J., Joseph, K.S., Hage, D.S.: Analysis of drug interactions with modified
- proteins by high-performance affinity chromatography: binding of glibenclamide to normal and glycated
- human serum albumin. J Chromatogr A 1265, 114-122 (2012)
- 35. Anguizola, J., Matsuda, R., Barnaby, O.S., Hoy, K.S., Wa, C., DeBolt, E., Koke, M., Hage, D.S.:
- Review: Glycation of human serum albumin. Clin Chim Acta 425, 64-76 (2013)
- 36. Ziyadeh, F.N., Cohen, M.P.: Effects of glycated albumin on mesangial cells: evidence for a role in
- diabetic nephropathy. Mol Cell Biochem 125(1), 19-25 (1993)
- 37. Chen, S., Cohen, M.P., Ziyadeh, F.N.: Amadori-glycated albumin in diabetic nephropathy:
- pathophysiologic connections. Kidney Int Suppl 77, S40-44 (2000)
- 38. Jin, C., Lu, L., Zhang, R.Y., Zhang, Q., Ding, F.H., Chen, Q.J., Shen, W.F.: Association of serum
- glycated albumin, C-reactive protein and ICAM-1 levels with diffuse coronary artery disease in patients with
- type 2 diabetes mellitus. Clin Chim Acta 408(1-2), 45-49 (2009)

 Rubenstein, D.A., Maria, Z., Yin, W.: Glycated albumin modulates endothelial cell thrombogenic and inflammatory responses. J Diabetes Sci Technol 5(3), 703-713 (2011)

40. Blache, D., Bourdon, E., Salloignon, P., Lucchi, G., Ducoroy, P., Petit, J.M., Verges, B., Lagrost, L.:
Glycated albumin with loss of fatty acid binding capacity contributes to enhanced arachidonate oxygenation and platelet hyperactivity: relevance in patients with type 2 diabetes. Diabetes 64(3), 960-972 (2015)
41. Bierhaus, A., Humpert, P.M., Morcos, M., Wendt, T., Chavakis, T., Arnold, B., Stern, D.M., Nawroth, P.P.: Understanding RAGE, the receptor for advanced glycation end products. J Mol Med (Berl) 83(11), 876-886 (2005)

42. Stirban, A., Gawlowski, T., Roden, M.: Vascular effects of advanced glycation endproducts: Clinical effects and molecular mechanisms. Mol Metab **3**(2), 94-108 (2014)

43. Shim, E., Babu, J.P.: Glycated albumin produced in diabetic hyperglycemia promotes monocyte secretion of inflammatory cytokines and bacterial adherence to epithelial cells. J Periodontal Res 50(2), 197-204 (2015)

44. Miele, C., Riboulet, A., Maitan, M.A., Oriente, F., Romano, C., Formisano, P., Giudicelli, J., Beguinot,
F., Van Obberghen, E.: Human glycated albumin affects glucose metabolism in L6 skeletal muscle cells by impairing insulin-induced insulin receptor substrate (IRS) signaling through a protein kinase C alpha-mediated mechanism. J Biol Chem 278(48), 47376-47387 (2003)

45. Shima, K., Ito, N., Abe, F., Hirota, M., Yano, M., Yamamoto, Y., Uchida, T., Noguchi, K.: High-performance liquid chromatographic assay of serum glycated albumin. Diabetologia **31**(8), 627-631 (1988).
46. Yasukawa, K., Abe, F., Shida, N., Koizumi, Y., Uchida, T., Noguchi, K., Shima, K.: High-performance affinity chromatography system for the rapid, efficient assay of glycated albumin. J Chromatogr **597**(1-2), 271-275 (1992)

47. Kouzuma, T., Usami, T., Yamakoshi, M., Takahashi, M., Imamura, S.: An enzymatic method for the measurement of glycated albumin in biological samples. Clin Chim Acta 324(1-2), 61-71 (2002)
48. Kohzuma, T., Yamamoto, T., Uematsu, Y., Shihabi, Z.K., Freedman, B.I.: Basic performance of an enzymatic method for glycated albumin and reference range determination. J Diabetes Sci Technol 5(6), 1455-1462 (2011).

49. Takei, I., Hoshino, T., Tominaga, M., Ishibashi, M., Kuwa, K., Umemoto, M., Tani, W., Okahashi, M., Yasukawa, K., Kohzuma, T., Sato, A.: Committee on Diabetes Mellitus Indices of the Japan Society of Clinical Chemistry-recommended reference measurement procedure and reference materials for glycated albumin determination. Ann Clin Biochem (2015)

50. Shafi, T., Sozio, S.M., Plantinga, L.C., Jaar, B.G., Kim, E.T., Parekh, R.S., Steffes, M.W., Powe, N.R., Coresh, J., Selvin, E.: Serum fructosamine and glycated albumin and risk of mortality and clinical outcomes in hemodialysis patients. Diabetes Care **36**(6), 1522-1533 (2013)

51. Schleicher, E.D., Mayer, R., Wagner, E.M., Gerbitz, K.D.: Is serum fructosamine assay specific for determination of glycated serum protein? Clin Chem **34**(2), 320-323 (1988)

52. Rodriguez-Capote, K., Tovell, K., Holmes, D., Dayton, J., Higgins, T.N.: Analytical evaluation of the Diazyme glycated serum protein assay on the siemens ADVIA 1800: comparison of results against HbA1c for diagnosis and management of diabetes. J Diabetes Sci Technol **9**(2), 192-199 (2015)

53. Takahashi, S., Uchino, H., Shimizu, T., Kanazawa, A., Tamura, Y., Sakai, K., Watada, H., Hirose, T.,

Kawamori, R., Tanaka, Y.: Comparison of Glycated Albumin (GA) and Glycated Hemoglobin (HbA1c) in

Type 2 Diabetic Patients: Usefulness of GA for Evaluation of Short-term Changes in Glycemic Control. Endocrine Journal **54**(1), 139-144 (2007)

54. Furusyo, N., Koga, T., Ai, M., Otokozawa, S., Kohzuma, T., Ikezaki, H., Schaefer, E.J., Hayashi, J.:

Utility of glycated albumin for the diagnosis of diabetes mellitus in a Japanese population study: results from

the Kyushu and Okinawa Population Study (KOPS). Diabetologia 54(12), 3028-3036 (2011)

55. Ikezaki, H., Furusyo, N., Ihara, T., Hayashi, T., Ura, K., Hiramine, S., Mitsumoto, F., Takayama, K.,

Murata, M., Kohzuma, T., Ai, M., Schaefer, E.J., Hayashi, J.: Glycated albumin as a diagnostic tool for

diabetes in a general Japanese population. Metabolism **64**(6), 698-705 (2015)

56. Gram-Hansen, P., Eriksen, J., Mourits-Andersen, T., Olesen, L.: Glycosylated haemoglobin (HbA1c) in

iron- and vitamin B12 deficiency. J Intern Med 227(2), 133-136 (1990)

57. Coban, E., Ozdogan, M., Timuragaoglu, A.: Effect of iron deficiency anemia on the levels of hemoglobinA1c in nondiabetic patients. Acta Haematol 112(3), 126-128 (2004)

58. Kim, C., Bullard, K.M., Herman, W.H., Beckles, G.L.: Association between iron deficiency and A1C Levels among adults without diabetes in the National Health and Nutrition Examination Survey, 1999-2006. Diabetes Care 33(4), 780-785 (2010)

59. Church, D., Simmons, D.: More evidence of the problems of using HbA1c for diagnosing diabetes? The known knowns, the known unknowns and the unknown unknowns. J Intern Med 276(2), 171-173 (2014)
60. Cohen, R.M., Franco, R.S., Khera, P.K., Smith, E.P., Lindsell, C.J., Ciraolo, P.J., Palascak, M.B., Joiner, C.H.: Red cell life span heterogeneity in hematologically normal people is sufficient to alter HbA1c. Blood 112(10), 4284-4291 (2008)

61. Koga, M., Saito, H., Mukai, M., Matsumoto, S., Kasayama, S.: Influence of iron metabolism indices on glycated haemoglobin but not glycated albumin levels in premenopausal women. Acta Diabetol **47 Suppl 1**, 65-69 (2010)

 Krzisnik, C., Lukac-Bajalo, J.: Glycosylated hemoglobin in fractions of erythrocytes of different ages. J Endocrinol Invest 16(7), 495-498 (1993)

63. Radin, M.S.: Pitfalls in hemoglobin A1c measurement: when results may be misleading. J Gen Intern Med **29**(2), 388-394 (2014)

64. Phelps, R.L., Honig, G.R., Green, D., Metzger, B.E., Frederiksen, M.C., Freinkel, N.: Biphasic changes in hemoglobin A1c concentrations during normal human pregnancy. Am J Obstet Gynecol **147**(6), 651-653 (1983)

65. Hashimoto, K., Noguchi, S., Morimoto, Y., Hamada, S., Wasada, K., Imai, S., Murata, Y., Kasayama, S., Koga, M.: A1C but not serum glycated albumin is elevated in late pregnancy owing to iron deficiency.
Diabetes Care **31**(10), 1945-1948 (2008)

66. Hashimoto, K., Osugi, T., Noguchi, S., Morimoto, Y., Wasada, K., Imai, S., Waguri, M., Toyoda, R.,Fujita, T., Kasayama, S., Koga, M.: A1C but not serum glycated albumin is elevated because of iron

deficiency in late pregnancy in diabetic women. Diabetes Care 33(3), 509-511 (2010)

67. Suzuki, S., Koga, M., Amamiya, S., Nakao, A., Wada, K., Okuhara, K., Hayano, S., Sarhat, A.R.,

Takahashi, H., Matsuo, K., Tanahashi, Y., Fujieda, K.: Glycated albumin but not HbA1c reflects glycaemic control in patients with neonatal diabetes mellitus. Diabetologia **54**(9), 2247-2253 (2011)

68. Freedman, B.I., Shenoy, R.N., Planer, J.A., Clay, K.D., Shihabi, Z.K., Burkart, J.M., Cardona, C.Y., Andries, L., Peacock, T.P., Sabio, H., Byers, J.R., Russell, G.B., Bleyer, A.J.: Comparison of glycated albumin and hemoglobin A1c concentrations in diabetic subjects on peritoneal and hemodialysis. Perit Dial Int **30**(1), 72-79 (2010)

69. Freedman, B.I., Shihabi, Z.K., Andries, L., Cardona, C.Y., Peacock, T.P., Byers, J.R., Russell, G.B., Stratta, R.J., Bleyer, A.J.: Relationship between assays of glycemia in diabetic subjects with advanced chronic kidney disease. Am J Nephrol **31**(5), 375-379 (2010)

70. Sumner, A.E., Duong, M.T., Aldana, P.C., Ricks, M., Tulloch-Reid, M.K., Lozier, J.N., Chung, S.T.,

Sacks, D.B.: A1C Combined With Glycated Albumin Improves Detection of Prediabetes in Africans: The Africans in America Study. Diabetes Care **39**(2), 271-277 (2016)

71. Alssema, M., Boers, H.M., Ceriello, A., Kilpatrick, E.S., Mela, D.J., Priebe, M.G., Schrauwen, P.,

Wolffenbuttel, B.H., Pfeiffer, A.F.: Diet and glycaemia: the markers and their meaning. A report of the Unilever Nutrition Workshop. Br J Nutr **113**(2), 239-248 (2015)

Noda, K., Zhang, B., Iwata, A., Nishikawa, H., Ogawa, M., Nomiyama, T., Miura, S., Sako, H., Matsuo, K., Yahiro, E., Yanase, T., Saku, K.: Lifestyle changes through the use of delivered meals and dietary counseling in a single-blind study. The STYLIST study. Circ J 76(6), 1335-1344 (2015)

73. Murai, J., Soga, S., Saito, H., Koga, M.: Usefulness of glycated albumin for early detection of deterioration of glycemic control state after discharge from educational admission. Endocr J 60(4), 409-413 (2013)

74. Koga, M., Murai, J., Saito, H., Kasayama, S., Imagawa, A., Hanafusa, T., Kobayashi, T.: Serum glycated albumin to haemoglobin A(1C) ratio can distinguish fulminant type 1 diabetes mellitus from type 2 diabetes mellitus. Ann Clin Biochem **47**(Pt 4), 313-317 (2010)

 Ueda, Y., Matsumoto, H.: Recent topics in chemical and clinical research on glycated albumin. J Diabetes Sci Technol 9(2), 177-182 (2015)

76. Tominaga, M., Eguchi, H., Manaka, H., Igarashi, K., Kato, T., Sekikawa, A.: Impaired glucose tolerance is a risk factor for cardiovascular disease, but not impaired fasting glucose. The Funagata Diabetes Study. Diabetes Care 22(6), 920-924 (1999) 77. Glucose tolerance and mortality: comparison of WHO and American Diabetes Association diagnostic criteria. The DECODE study group. European Diabetes Epidemiology Group. Diabetes Epidemiology:
Collaborative analysis Of Diagnostic criteria in Europe. Lancet **354**(9179), 617-621 (1999)

78. Heine, R.J., Balkau, B., Ceriello, A., Del Prato, S., Horton, E.S., Taskinen, M.R.: What does postprandial hyperglycaemia mean? Diabet Med **21**(3), 208-213 (2004)

79. Yoshiuchi, K., Matsuhisa, M., Katakami, N., Nakatani, Y., Sakamoto, K., Matsuoka, T., Umayahara, Y., Kosugi, K., Kaneto, H., Yamasaki, Y., Hori, M.: Glycated albumin is a better indicator for glucose excursion than glycated hemoglobin in type 1 and type 2 diabetes. Endocr J 55(3), 503-507 (2008)
80. Koga, M., Murai, J., Saito, H., Kasayama, S.: Glycated albumin and glycated hemoglobin are influenced

differently by endogenous insulin secretion in patients with type 2 diabetes. Diabetes Care **33**(2), 270-272 (2010)

 Koga, M., Murai, J., Saito, H., Mukai, M., Matsumoto, S., Kasayama, S.: Glycated albumin levels are higher relative to glycated haemoglobin levels in gastrectomized subjects. Ann Clin Biochem 47(Pt 1), 39-43 (2010)

82. Selvin, E., Rawlings, A.M., Grams, M., Klein, R., Sharrett, A.R., Steffes, M., Coresh, J.: Fructosamine and glycated albumin for risk stratification and prediction of incident diabetes and microvascular complications: a prospective cohort analysis of the Atherosclerosis Risk in Communities (ARIC) study. Lancet Diabetes Endocrinol 2(4), 279-288 (2014)

83. Nathan, D.M., McGee, P., Steffes, M.W., Lachin, J.M., Group, D.E.R.: Relationship of glycated albumin to blood glucose and HbA1c values and to retinopathy, nephropathy, and cardiovascular outcomes in the DCCT/EDIC study. Diabetes **63**(1), 282-290 (2014)

 Furusyo, N., Koga, T., Ai, M., Otokozawa, S., Kohzuma, T., Ikezaki, H., Schaefer, E.J., Hayashi, J.:
 Plasma glycated albumin level and atherosclerosis: results from the Kyushu and Okinawa Population Study (KOPS). Int J Cardiol 167(5), 2066-2072 (2013)

85. Ma, X., Shen, Y., Hu, X., Hao, Y., Luo, Y., Tang, J., Zhou, J., Bao, Y., Jia, W.: Associations of glycated haemoglobin A1c and glycated albumin with subclinical atherosclerosis in middle-aged and elderly Chinese population with impaired glucose regulation. Clin Exp Pharmacol Physiol **42**(6), 582-587 (2015)

86. Inaba, M., Okuno, S., Kumeda, Y., Yamada, S., Imanishi, Y., Tabata, T., Okamura, M., Okada, S., Yamakawa, T., Ishimura, E., Nishizawa, Y., Osaka, C.K.D.E.R.G.: Glycated albumin is a better glycemic indicator than glycated hemoglobin values in hemodialysis patients with diabetes: effect of anemia and erythropoietin injection. J Am Soc Nephrol **18**(3), 896-903 (2007)

87. Koga, M., Murai, J., Saito, H., Matsumoto, S., Kasayama, S.: Effects of thyroid hormone on serum glycated albumin levels: study on non-diabetic subjects. Diabetes Res Clin Pract 84(2), 163-167 (2009)
88. Nishimura, R., Kanda, A., Sano, H., Matsudaira, T., Miyashita, Y., Morimoto, A., Shirasawa, T., Kawaguchi, T., Tajima, N.: Glycated albumin is low in obese, non-diabetic children. Diabetes Res Clin Pract 71(3), 334-338 (2006)

89. Koga, M., Otsuki, M., Matsumoto, S., Saito, H., Mukai, M., Kasayama, S.: Negative association of obesity and its related chronic inflammation with serum glycated albumin but not glycated hemoglobin levels. Clin Chim Acta **378**(1-2), 48-52 (2007)

90. Koga, M., Murai, J., Saito, H., Mukai, M., Kasayama, S.: Serum glycated albumin, but not glycated haemoglobin, is low in relation to glycemia in hyperuricemic men. Acta Diabetol 47(2), 173-177 (2010)
91. Koga, M., Murai, J., Saito, H., Mukai, M., Kasayama, S.: Serum glycated albumin, but not glycated hemoglobin, is low in relation to glycemia in men with hypertriglyceridemia. J Diabetes Investig 1(5), 202-207 (2010)

92. Koga, M., Murai, J., Saito, H., Mukai, M., Kasayama, S.: Serum glycated albumin levels, but not glycated hemoglobin, is low in relation to glycemia in non-diabetic men with nonalcoholic fatty liver disease with high alanine aminotransferase levels. Clin Biochem **43**(12), 1023-1025 (2010)

93. Trenti, T., Cristani, A., Cioni, G., Pentore, R., Mussini, C., Ventura, E.: Fructosamine and glycated hemoglobin as indices of glycemic control in patients with liver cirrhosis. Ric Clin Lab **20**(4), 261-267 (1990)

94. Koga, M., Kasayama, S., Kanehara, H., Bando, Y.: CLD (chronic liver diseases)-HbA1C as a suitable indicator for estimation of mean plasma glucose in patients with chronic liver diseases. Diabetes Res Clin Pract **81**(2), 258-262 (2008)

95. Bando, Y., Kanehara, H., Toya, D., Tanaka, N., Kasayama, S., Koga, M.: Association of serum glycated albumin to haemoglobin A1C ratio with hepatic function tests in patients with chronic liver disease. Ann Clin Biochem **46**(Pt 5), 368-372 (2009)

96. Araki, T., Ishikawa, Y., Okazaki, H., Tani, Y., Toyooka, S., Satake, M., Miwa, U., Tadokoro, K.:

Introduction of glycated albumin measurement for all blood donors and the prevalence of a high glycated albumin level in Japan. J. Diabetes Investig. **3**(6), 492-497 (2012)

97. Ai, M., Otokozawa, S., Schaefer, E.J., Asztalos, B.F., Nakajima, K., Shrader, P., Kathiresan, S., Meigs,

J.B., Williams, G., Nathan, D.M.: Glycated albumin and direct low density lipoprotein cholesterol levels in type 2 diabetes mellitus. Clin. Chim. Acta **406**(1-2), 71-74 (2009)

98. ADA: Standards of medical care in diabetes--2015: summary of revisions. Diabetes Care **38 Suppl**, S4 (2015)

99. American Diabetes, A.: Standards of medical care in diabetes-2015 abridged for primary care providers.Clin. Diabetes 33(2), 97-111 (2015)