



Targeting Tumor Initiating Cells through Inhibition of Cancer Testis Antigens and Notch Signaling: A Hypothesis

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3 **Targeting Tumor Initiating Cells through Inhibition of Cancer Testis Antigens and**
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5 **Notch Signaling: A Hypothesis**
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43 **Running head:** Tumor initiating cells, CTA and Notch
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16 **Abstract**
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19 Tumor initiating cells (TICs) differ from normal stem cells (SCs) in their ability to
20 initiate tumorigenesis, invasive growth, metastasis and the acquisition of chemo and/or
21 radio-resistance. In the last years, several studies have indicated the potential role of the
22 Notch system as a key regulator of cellular stemness and tumor development.
23 Furthermore, the expression of cancer testis antigens (CTA) in TICs, and their role in SC
24 differentiation and biology, has become an important area of investigation. Here we
25 propose a model in which CTA expression and Notch signaling interacts to maintain the
26 sustainability of self-replicating tumor populations, ultimately leading to the development of
27 metastasis, drug resistance, and cancer progression. We hypothesize that Notch-CTA
28 interactions in TICs offer a novel opportunity for meaningful therapeutic interventions in
29 cancer.
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3 **Keywords:** Notch signaling, Cancer testis antigens, cancer, stem cells
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18 ***Tumor initiating cells: the dark side of stemness***
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20 Stem cells (SCs) are toti-potential cells capable of an unlimited number of
21 replication cycles and, under appropriate stimuli, differentiation into more specialized cells.
22 This basal cell compartment constitutes the internal repair and self-renewal system in
23 normal tissues. It is this balance between cellular self-renewal and differentiation what
24 maintains normal tissue architecture and function. Therefore, it is not surprising that
25 molecular derangements that affect this highly regulated process can lead to uncontrolled
26 cellular growth, aberrant differentiation and tumor development.
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32 The proposed existence of a population of tumor initiating cells (TIC) that mediate
33 tumorigenesis and cancer progression has forced us to revisit our understanding of the
34 fundamental processes governing cancer development. Unlike normal SCs, TICs have
35 lost their stringent regulation and display a tendency for tumorigenesis, cellular
36 invasiveness and metastatic potential [1]. The renewed interest in the study of TIC biology
37 stems from the fact that their identification and therapeutic targeting may result, not only
38 on improved tumor responses, but also in the potential eradication of neoplastic cells
39 associated with pharmacologic resistance and disease relapse.
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45 Several hypotheses have been proposed to account for the origin and development
46 of TICs. The stochastic model suggests that normally differentiated cells can acquire a
47 transformed phenotype capable of driving tumorigenesis through clonal evolution.
48 Alternatively, the hierarchic model proposes that normal SCs may undergo an oncogenic
49 event and subsequently serve as a continuous source of neoplastic cells, intrinsically
50 resistant to standard therapeutic interventions. These two views are not necessarily
51 mutually exclusive and a combined model may account for tumor development and
52 progression, more accurately [2]. For example, TICs may originate either from normal SCs
53 or more differentiated progenitors that acquire the capacity for self-renewal, invasive
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3 behavior and metastatic potential. Brabletz et al. has proposed the concept of “migrating
4 TICs” (mTICs) which possess both stemness and migratory capabilities associated with
5 the acquisition of epithelial-mesenchymal transition (EMT) phenotype [3]. This switch from
6 SCs to mTICs appears to relate, not only to genetic alterations, but also to micro-
7 environmental factors. These permissive micro-environmental niches may promote TIC
8 proliferation and differentiation into invasive and metastatic cell populations, resulting in
9 tumor progression and the emergence of resistance to standard treatments.
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12 TICs are usually characterized by the expression and/or deregulation of genes
13 involved in embryonic development, such as the Hedgehog, Notch and Wnt signaling
14 pathways. These three signaling pathways appear to be fundamental in TIC biology and
15 represent promising targets for therapeutic intervention [4]. In the next sections we will
16 discuss the role of the Notch signaling pathway in cellular stemness and tumorigenesis, as
17 well as its interaction with the family of proteins known as cancer testis antigens (CTAs),
18 recently found to be expressed on TICs [5].
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28 ***The Notch pathway***

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30 The Notch family of proteins includes a total of four highly homologous receptors
31 (Notch-1, Notch-2, Notch-3, Notch-4) with important roles in the regulation of various
32 processes, including neurogenesis, gliogenesis, myogenesis, vasculogenesis,
33 hematopoiesis and epidermal development [6]. The Notch system is involved in the
34 homeostasis of adult tissues by promoting self-renewal of SCs, determining cell fate (such
35 as commitment towards T or B cell lineage) and regulating the differentiation of many cell
36 types. Because of its extensive involvement in all these processes, mutations and/or
37 deregulation of Notch receptors and/or ligands has been associated with the genesis of
38 various malignancies [7]. Notch signaling depends on its interaction with specific ligands,
39 proteolytic cleavage, release of Notch’s cytoplasmic portion and subsequent translocation
40 to the nucleus [8]. In vertebrates, two closely related families of ligands are capable of
41 interacting with Notch receptors: the Delta-like (DLL-1, -3, -4 and DLL-2, described only in
42 Xenopus) and the Serrate-like (Jagged1 and 2) ligands [9]. Notch signaling is dependent
43 on cell-to-cell communication and is activated when Notch ligands, present on the
44 “sending cell”, bind to Notch receptor on the “receiving cell”. Ligand binding triggers
45 conformational changes in Notch receptors that allow access to the ADAM/TACE (A
46 disintegrin and metalloprotease/tumor-necrosis-factor α converting enzyme)
47 metalloprotease. The resulting proteolytic cleavage occurs within the extracellular domain
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3 (approximately 12 amino acids proximal to the transmembrane domain), at a site referred
4 to as the S2 cleavage site [8]. ADAM/TACE leaves a short-lived fragment anchored to the
5 plasma membrane, known as Notch extracellular truncation (NEXT), which is then
6 recognized by the inactive aminopeptidase domain of Nicastrin (NCT) and transferred to
7 the active site of the enzyme γ -secretase. γ -secretase operates within the transmembrane
8 domain (between site S3, near the inner plasma membrane, and site S4, near the middle
9 of the transmembrane domain) and releases the Notch intracellular domain (NICD) which
10 then translocates to the nucleus (or endoplasmic reticulum), and interacts with the DNA-
11 binding protein complex CBF-1/RBP-Jk (Cp-binding factor-1/(Recombination signal
12 sequence-binding protein Jk) to regulate transcription of Notch-dependent genes [9, 10].
13 NICD transcriptional targets include Hairy/Enhancer of Split (HES) genes, HES-related
14 repressor protein (HERP), class C bHLH (basic-helix-loop-helix) proteins acting as
15 transcriptional repressors of tissue-specific genes (such as MASH-1 in neurogenesis,
16 MyoD in myogenesis and E2A in B lymphopoiesis), the pre-TCR alpha (which
17 participates in regulating the T cell progenitors differentiation), cell cycle regulatory genes
18 such as cyclin D2, cyclin D1, p21 and p27, and genes from different chemokine systems
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Notch signaling is involved in the regulation of SC fate, in both embryonic and adult
tissues. Activation of the Notch pathway has been shown to regulate the transit of nervous
SCs from quiescence to a proliferative state and, together with Wnt signaling, also
coordinates proliferation and differentiation of intestinal SCs [14]. Since deregulation of
the Notch system has been shown to be involved in the patho-biology of many neoplasms,
it is not surprising this signaling pathway is of fundamental importance in the molecular
processes involved in TIC development.

Notch and TICs

Activation of the Notch system promotes self-renewal of TICs in several malignancies and participates in the interactions between tumor cells and the microenvironment, in both primary and metastatic neoplasms [15]. The Notch signaling pathway regulates not only the formation of TICs, but also their acquisition of the EMT phenotype which promotes metastasis and drug resistance [16]. Notch signaling has been demonstrated to be crucial in neural SCs biology based on its ability to inhibit neuronal differentiation [17]. The observation that Notch activation up-regulates AKT and MCL-1 expression in glioma SCs after radiation, indicates its importance in mediating resistance

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3 to DNA damage-induced apoptosis [18]. Glioblastoma multiforme (GBM) SCs appear to
4 be sensitive to Notch-inhibitors, reflecting the importance of Notch and HES gene family
5 activation in their biology. A study by Shiras et al. demonstrated that Notch blockade can
6 reduce the CD133-positive SC fraction and abolished the non SC population in a human
7 model of GBM, suggesting this pathway is involved in maintenance of the TIC
8 compartment and tumorigenesis [19]. Notch and Wnt pathways also play crucial roles in
9 coordinating proliferation and differentiation of intestinal stem cells in mice [14], and their
10 activation leads to amplification of the intestinal SC/progenitor cell pool and inhibition of
11 cell differentiation [20]. The importance of Notch signaling in gastrointestinal tissues is
12 further supported by its association with chemotherapy resistance in gastric tumors [21]
13 and up-regulation of Notch1 and HES1 in colon adenocarcinomas, compared to normally
14 differentiated colonic cells [22]. Since maintenance of undifferentiated, proliferative cells in
15 intestinal crypts and adenomas requires activation of the Notch pathway, inhibition of
16 Notch signaling by γ -secretase inhibitors has been proposed as a treatment for colorectal
17 cancer [23].

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19 The Notch pathway is also deregulated in breast cancer (BC) and its activation may
20 protect BCs from drug-induced apoptosis [24, 25]. Indeed, Notch signaling has been
21 suggested as one of the conserved developmental pathways leading to TIC maintenance
22 and resistance to conventional chemotherapy in this disease [26]. In ovarian cancer (OC)
23 Notch3 activation has been associated with chemo-resistance and poor prognosis [27,28].
24 McAuliffe et al. have demonstrated over-expression of Notch3 in OC is associated with an
25 increased TIC population and development of resistance to cisplatin, while Notch3
26 inhibition results in reduction of TICs and restoration of sensitivity to platinum agents [29].
27 Aberrations in Notch signaling have also been linked to several hematological
28 malignancies, including T-cell acute lymphoblastic leukemia (T-ALL), acute myeloid
29 leukemia (AML), lymphoma and multiple myeloma (MM) [6, 12]. The role of Notch
30 signaling in hematopoiesis and in hematopoietic SC (HSC) development and maintenance
31 has been extensively reviewed by Bigas and Espinosa [30], the fundamental finding being
32 that activation of the Notch pathway is necessary for HSCs maintenance, and its inhibition
33 results in HSCs differentiation and depletion [31]. The role of Notch signaling in the
34 biology of leukemic SCs (LSCs), however, appears to be context-dependent and requires
35 further study before its importance is completely understood in these diseases [31].
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58 ***Cancer Testis Antigens***

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3 Cancer Testis Antigens (CTA) are a sub-class of tumor associated antigens first cloned by
4 van der Bruggen et al. in 1991 [32]. CTA expression is mainly restricted to the testis and
5 neoplastic cells of various histological origins, with the vast majority of these molecules
6 having negligible or no expression in normal somatic cells [33-43]. This restricted
7 expression pattern depends on methylation of upstream control elements and can be
8 altered by DNA methyl-transferase-1 inhibitors, such as 5-aza-2-deoxycytidine, and
9 histone deacetylase (HDAC) inhibitors [44]. Since cancer is characterized by global
10 genomic hypomethylation, CpG island hypomethylation is likely to represent a major
11 mechanism for re-activation of CTA transcriptional expression in cancer. Simpson et al.
12 have recently suggested that CTA expression in cancer cells may be the result of re-
13 activation of a gametogenic program as part of the process of neoplastic transformation
14 [45]. Indeed, a number of CTAs normally expressed at various stages of gametogenesis
15 have also been found in cancer cells, suggesting the triggering event is re-activation of a
16 germ cell developmental program [45]. This theory supports the recent observation that
17 some CTAs are expressed in TICs, cells characterized by self-renewal abilities similar to
18 germ cells.
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21 The possibility that CTAs may be expressed by TICs suggests these molecules may
22 also serve as therapeutic targets. TIC-specific interventions, such as vaccines and/or
23 other immunotherapeutic approaches targeting CTAs, may result in eradication of this self-
24 renewal cell population and increased cure rates in specific malignancies. In fact, their
25 restricted tissue expression, high immunogenicity and presence of HLA-I-restricted
26 epitopes, make CTAs ideal candidates for the immunotherapeutic targeting of cancer [36,
27 46, 47]. So far, most CTA-specific vaccines have targeted CTAs such as MAGE-A, NY-
28 ESO-1, and SSX, with variable success [48, 49]. More recently, a novel group of CTAs
29 belonging to the sub-group of scaffolding sperm proteins, namely SP17, AKAP-4 and
30 Ropporin-1, have shown promise as targets for immunotherapeutic interventions in certain
31 malignancies [33, 36, 40, 41, 50-52]. Among these, only SP17 has been evaluated in the
32 clinical setting [53]. In this study, SP17-loaded autologous dendritic cells (DCs) were used
33 to treat a MM patient, who had developed disease progression, and resulted in SP17-
34 specific immunity and reduction of serum paraprotein levels [53]. Based on these
35 preliminary results, evaluation of this CTA sub-group of scaffolding sperm proteins is
36 clearly warranted in a larger group of MM patients. More importantly, further studies to
37 unravel the link between CTA expression and TIC biology are necessary in order to
38 develop effective TIC-targeted therapies in the future.
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CTAs as markers of tumor stemness

The expression of CTAs by TICs, and the functional relation between CTAs and cell stemness, has become an active area of research. It has been suggested that CTA expression may play a role in SC differentiation pathways and embryonic development. For example, during embryonic development the epiblast gives rise to mesenchymal SCs (MSCs) and primordial germ cells with active CTA expression programs [54]. Differentiation of these primordial cells results in down-regulation of CTA expression and may explain why CTAs are not detected in spermatocytes or oocytes, but are present in gametagonia [54]. In addition to other processes associated with SCs, such as the level of histone acetylation and lack of cellular differentiation, CTAs may serve as markers of stemness in specific neoplasms. The role of CTAs as SC markers is supported by the detection of CTA expression in bone marrow MSCs. Expression of CTAs, such as NY-ESO-1, MAGE1, SSX and N-RAGE, has been shown to be down-regulated in adipocytes and osteocytes after their complete differentiation from MSCs [55]. CTA expression may also confer a functional advantage to cancer cells by modulating their invasive and metastatic potential. For example, SSX down-regulation in melanoma cells results in decreased expression of matrix metalloprotease-2 (MMP2) and migration potential [56]. Since the process of malignant transformation may be defined by mutational deregulation of SC programs for self-renewal and differentiation, CTA expression may serve, not only as a potential marker of tumor cell stemness, but as an attractive target for treatment of TIC-dependent metastatic, recurrent and/or resistant disease [57].

Expression of CTAs in tumor SCs or TICs has been evaluated in several tumor models. GBM is a primary glial malignancy believed to have a defined SC population [58]. CD133 expression has been suggested as a marker of stemness in this malignancy based on the observation that CD133 positive cells can give rise to tumorigenic cells [58,59]. Interestingly, CTAs such as LAGE-1, NY-ESO-1 and MAGE family members are also expressed in GBM CD133 positive cells, compared with differentiated or parent cells [59]. Moreover, the expression of MAGE family members and CD133 is down-regulated as cells become more differentiated, indicating a correlation between expression of these molecules and the undifferentiated cellular state [59]. Since CD133 negative GBM cells can also be tumorigenic [58], CD133 expression can't be considered a specific marker for GBM stemness. Therefore, determination of expression of certain CTA expression patterns could be helpful in determining cellular stemness among CD133 negative GBM

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3 cell populations. Studies with MM clonogenic precursors (CD138 negative B cells thought
4 to be MM SCs) have also revealed expression of MAGE A3/A6 and MAGE-C1/C7, as well
5 as the protective role these CTAs confer against apoptosis [60]. MSCs can also give rise
6 to TICs with specific neoplastic characteristics and it has been suggested these TICs
7 comprise the main cellular compartment expressing CTAs within a given tumor [56, 57].
8 Gjerstorff et al. have found high levels of expression of several CTAs, such as CAGE,
9 MAGE-A and XAGE-1, in tumorigenic MSCs, as opposed to primary human MSCs [57].
10 These observations support the notion that CTA expression may be associated with
11 neoplastic transformation of normal SCs. Since promoter hypomethylation is one of the
12 main regulatory mechanisms involved in CTA expression, changes on CTA-specific
13 epigenetic control mechanisms may help differentiate neoplastic (i.e., TIC) from normal
14 SCs. This distinction is important since it could have both therapeutic and prognostic
15 implications.
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27 **Interactions between Notch signaling and CTA expression in TICs: Lessons from** 28 **MM.**

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30 The discovery of embryonic, germinal and tumor-restricted expression of CTAs has
31 advanced the hypothesis of a functional correlation between embryogenesis,
32 gametogenesis, and tumorigenesis [61,62]. These observations have partially resurrected
33 John Beard's "Trophoblastic Theory of Cancer" proposing that all tumors carry a
34 population of cells with germ-like features originating from germinal cells which have failed
35 to migrate to the gonads [63]. Thus, aberrant CTA expression in tumors may reflect re-
36 activation of a gametogenic program involved in maintenance of stemness in specific
37 cellular compartments, such as TICs [62]. As previously discussed, there is evidence
38 supporting the hypothesis that the Notch pathway may also be associated with reactivation
39 of a germinal cell transcription program in TICs. Since Notch signaling is an important
40 regulator of germ cell development, it is likely that the common expression pattern shared
41 by Notch1 and CTAs in germ cells [14-18] reflects a biologically relevant interaction
42 between these two pathways.
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52 Several CTAs are expressed in MM cells and their expression appears to play an
53 important role in cell proliferation and resistance to apoptosis [60, 64-67]. Our group was
54 the first to identify expression of two novel CTAs, AKAP-4 and SP17, in more than 75% of
55 MM cell lines and patients [37]. Jungbluth et al. have reported expression of MAGE-C1
56 and MAGE-A3/6 in MM, their expression correlating with MM cell proliferation [68]. CTA
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3 expression in MM appears to occur early in the evolution of this disease and maintained
4 thorough clinical progression, suggesting CTA expression may confer a selective
5 advantage to MM neoplastic cells [69]. CTA expression in malignant MM cells has also
6 been correlated with the activation of Notch signaling through the Jagged-2 ligand [70].
7 Moreover, activation of Notch signaling has been associated with drug resistance, as well
8 as cross-talk between MM cells and bone marrow microenvironment [71,72], the latter
9 being a critical requirement for maintenance of MM SC population [67].
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11 *In vitro* and *in vivo* studies support a “compartmental model” for MM. In this model
12 two cellular compartments exist: 1. a small, proliferating fraction of self-renewing
13 progenitor cells (the so called MM SC); 2. a non-proliferating compartment exhibiting
14 plasma cell differentiation and resistance to apoptosis [73]. Here, we propose a different
15 model in which Notch signaling and CTA expression interact to maintain a self-replicating
16 MM cell population. Although CTA expression has been described in a broad range of
17 solid and hematologic malignancies [33,35,36,39,40,41,43], MM is distinct in that a
18 significant majority of patients express AKAP-4 and/or SP17 in their tumor cells [34,37,38,
19 42]. No other type of cancer has such a close association with specific CTA expression
20 pattern. To date, the molecular and biological actions of SP17 and AKAP4 are
21 incompletely understood. Nonetheless, several reports have implicated SP17 in cell-to-cell
22 adhesion and/or cell migration of lympho-hematopoietic cells, possibly through its heparan
23 sulphate binding domain [74]. SP17 has two additional functional domains, an N-terminal
24 domain highly conserved among mammals and 45% identical to the type II alpha
25 regulatory subunit of protein kinase A (RII-PKA) and a C-terminal calmodulin binding
26 domain [75,76]. AKAPs represent a family of proteins characterized by their ability to bind
27 and regulate PKA function by localizing this critical enzyme to specific sub-cellular
28 compartments. Its structural homology to a PKA regulatory subunit suggests a potential
29 role for SP17 in signal transduction. In fact, non-PKA proteins containing an RII-PKA-like
30 domain at their N-terminal, such as Sp17, can interact with AKAPs via hydrophobic
31 residues [75]. For example, Lea et al. have demonstrated SP17 can bind the RII-PKA
32 binding site of AKAP3, the most abundant AKAPs in the fibrous sheath of mouse flagella
33 of spermatozoa [77]. This finding suggests a mechanism by which SP17 may regulate
34 PKA-dependent processes. By physically interacting with AKAPs RII-PKA binding site,
35 SP17 may disrupt the primary interaction of between AKAPs and PKA, resulting in mis-
36 localization of PKA and interference with its activity at specific subcellular compartments,
37 as well as modulation of other critical signal transduction pathways related to PKA
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3 activation. Alternatively, and by virtue of AKAP-specific anchoring to subcellular locations,
4 AKAP-bound SP17 may be translocated to specific compartments where it may exert, as
5 of yet, unknown biological functions.
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8 Activation of AKAP proteins can also occur under a variety of stimuli leading to
9 increase intracellular cAMP levels and consequent PKA activation [78]. Interestingly, a
10 role for PKA signaling in Notch activation has been clearly demonstrated by Weber and
11 colleagues [79]. The investigators showed that treatment of osteoclasts with parathyroid
12 hormone (PTH) induces a cAMP/PKA-dependent 8-fold increase in Jagged1 expression.
13 This finding is particularly relevant since Jagged1-mediated Notch activation has been
14 shown to drive proliferation of MM cells [70]. Another clue pointing to a role of PKA
15 signaling in Notch activation has been provided by a study by Yurugi-Kobayashi and
16 colleagues showing that cAMP-dependent PKA activation leads to suppression of COUP-
17 TFII expression, up-regulation of NRP1, and consequent Notch activation, in vascular
18 progenitor cells [80]. These findings indicate a possible role for SP17 and/or AKAP-4-
19 dependent PKA activation in the regulation of Notch ligand expression (Figure 1).
20 Therefore, coordination of SP17/AKAP4 expression and Notch signaling may result in
21 enhanced self-renewal and survival abilities of MM TIC compartment. Taken together,
22 these data suggest SP17 may play a regulatory role in the activation of important signal
23 transduction pathways by modulating the interaction between PKA and AKAPs. SP17
24 could also participate, through its RII-like N-terminal domain and the central heparan
25 sulphate binding domain, in key developmental processes such as cell migration and
26 Hedgehog signal transduction [81-83].
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41 **Future perspectives: immunotherapeutic implications**

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43 Cancer vaccines represent the most advanced immunotherapeutic approach for
44 oncologic patients. Classically, cancer vaccines are designed to exploit tumor targets
45 derived from cancer-inducing infectious agents and/or tumor-associated antigens. We
46 believe CTAs represent ideal cancer antigenic targets based on their selective expression
47 in immune privileged sites (i.e., testes) and neoplasms, but not in normal cells. Moreover,
48 the expression of CTAs by TICs, represent a powerful target for immunotherapeutic
49 interventions that may eradicate this fundamental and aberrant cellular compartment
50 responsible for initiation and maintenance of the neoplastic clone. With the discovery of
51 TIC-associated CTA expression the possibility of improving disease-free survival and cure
52 rates in cancer patients may become a reality. The efficacy of TIC-targeting vaccines has
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3 been explore with some success in acute myeloid leukemia (AML) [84]. However, the lack
4 of AML-TIC-restricted targets raises concerns about potential toxicities, which in principle
5 could be minimized by targeting TIC-associated CTAs [85].
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8 It is evident that any strategy designed to increase expression of cancer-restricted
9 antigens would result in better responses to cancer vaccine strategies [86]. Regulation of
10 CTA expression in tumors of different histological origin is based on epigenetic tuning of
11 promoter availability and chromatin remodeling [87-90]. Notch signaling has been shown
12 to down-regulate the expression of specific targets by increasing promoter methylation in
13 genes such as pRb and vascular endothelial growth factor receptor (VEGFR)-2 [91].
14 Therefore, the possibility exists that Notch signaling may contribute to mitigate the
15 expression of CTAs in selected tumoral sub-populations, such as TICs. In this scenario,
16 Notch-inhibiting agents, in combination with CTA-targeting vaccines, would improve
17 therapeutic efficacy by increasing the probability of TIC eradication. Additionally, inhibition
18 of Notch signaling by itself is expected to reduce the self-renewal ability of TICs, which
19 would then make these cells more susceptible to CTA-specific cytotoxic lymphocyte killing.
20 The potential drawback of this approach, however, would be the lack of tumor specificity of
21 Notch targeting, compared with CTA targeting. Currently, the only strategy tested in
22 patients to inhibit Notch signaling is the use γ -secretase inhibitors, which results in serious
23 gastrointestinal toxicity due to induction of intestinal metaplasia, a consequence of
24 deregulation of goblet cell differentiation in the intestine [92,93]. Intestinal metaplasia is
25 caused by the concurrent inhibition of both Notch1 and 2, indicating independent targeting
26 of Notch1 or Notch2 specific antibodies may eliminate the observed toxicity, as
27 demonstrated animal models of acute leukemias [94]. The use of antibodies that
28 specifically block tumor-associated Notch isoforms, and/or their ligands, represent a
29 potential alternative to toxic demethylating agents, such as 5-aza-2'-deoxycytidine, and γ -
30 secretase inhibitors [95]. As discussed above, selective Notch blockade would result in
31 increased CTA expression by TICs making these more susceptible to the cytotoxic effects
32 of therapeutic cancer vaccines. To date, several clinical trials are evaluating the safety
33 and efficacy of Notch inhibition in both solid and hematological malignancies, including MM
34 (www.clinicaltrials.gov). These studies should provide the necessary information to design
35 future clinical trials combining Notch inhibition and CTA overexpression and immuno-
36 targeting for the treatment of selective cancers [86,95,96].
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Declaration of Interest Statement

The authors have no conflict of interest to disclose.

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23 Figure legends

24 **Figure 1. Possible role of CTA signaling in Notch activation in MM.** CTA (i.e. SP17, AKAP4)
25 could directly enhance Notch activation by stabilizing the Notch intracellular domain (1), or they
26 could activate PKA through cAMP increase, which may lead to Notch ligand Jagged1-
27 2overexpression, eventually resulting in Notch activation and consequent MM TIC self-renewal.
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